



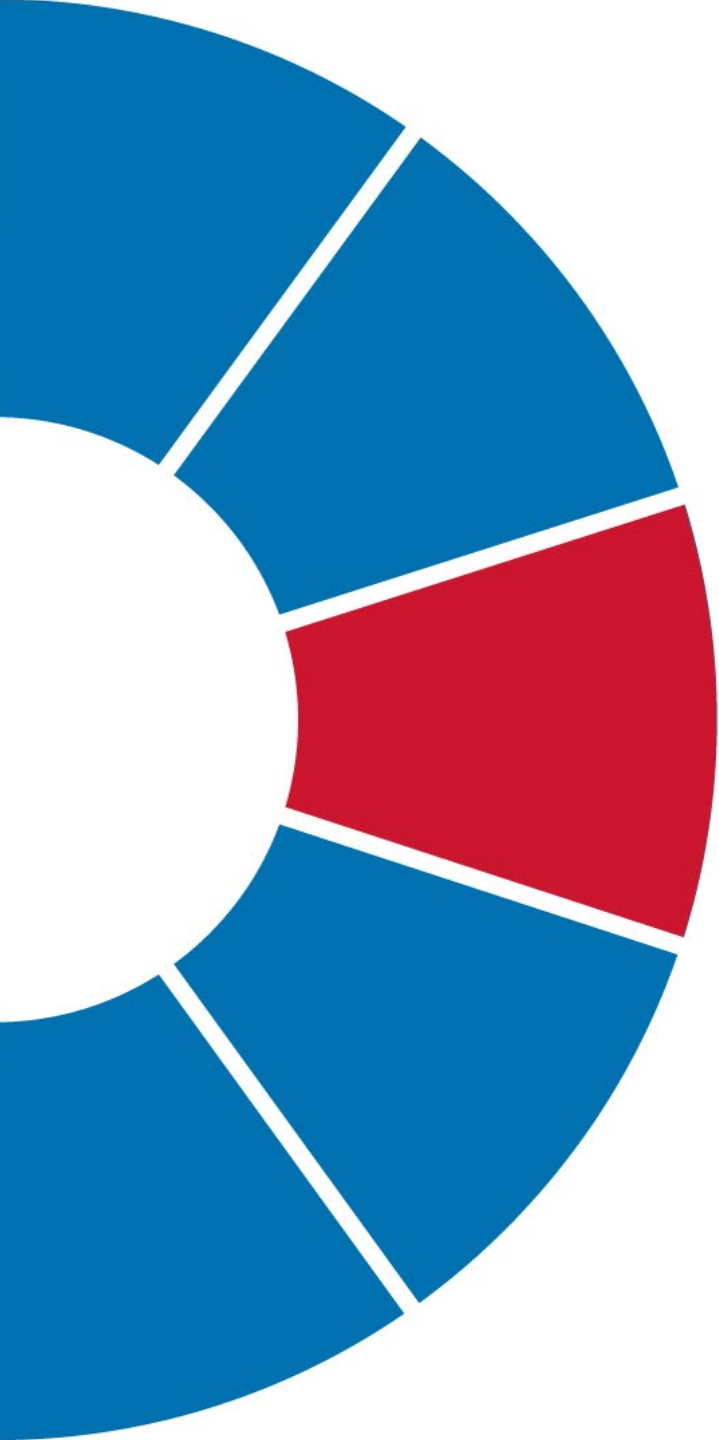
QUANTIFICATION OF MR SPECTRA

Cristina Cudalbu

CIBM MRI EPFL AIT

cristina.cudalbu@epfl.ch

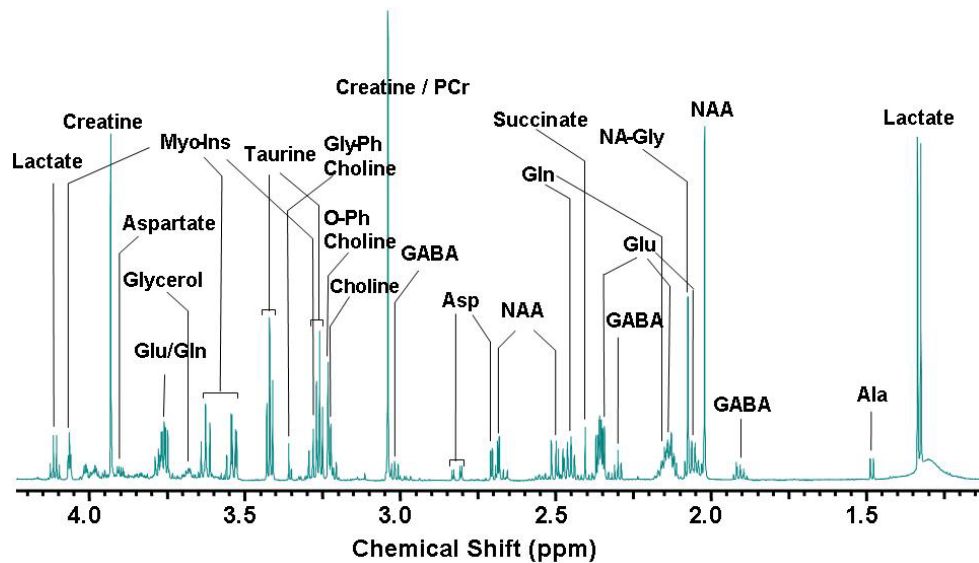




QUANTIFICATION

MAGNETIC RESONANCE SPECTROSCOPY

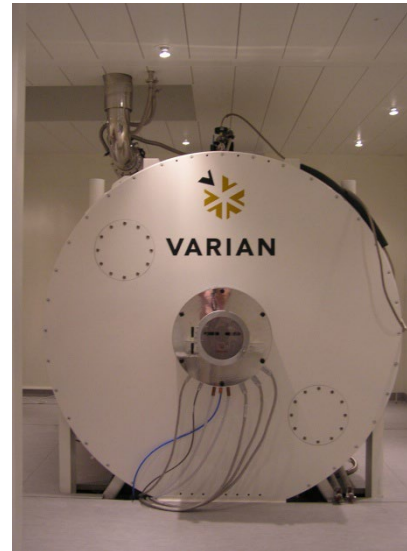
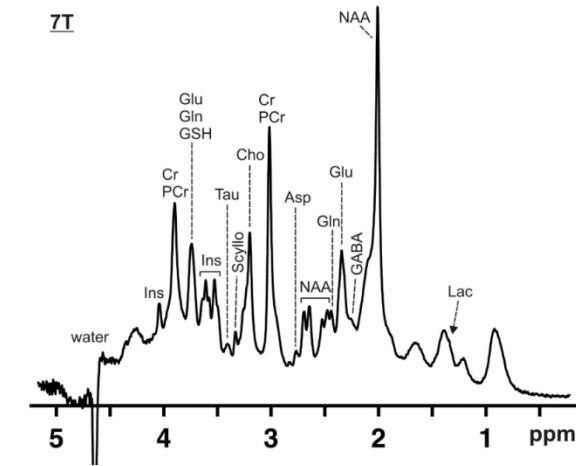
■ NMR Spectroscopy



MAGNETIC RESONANCE SPECTROSCOPY

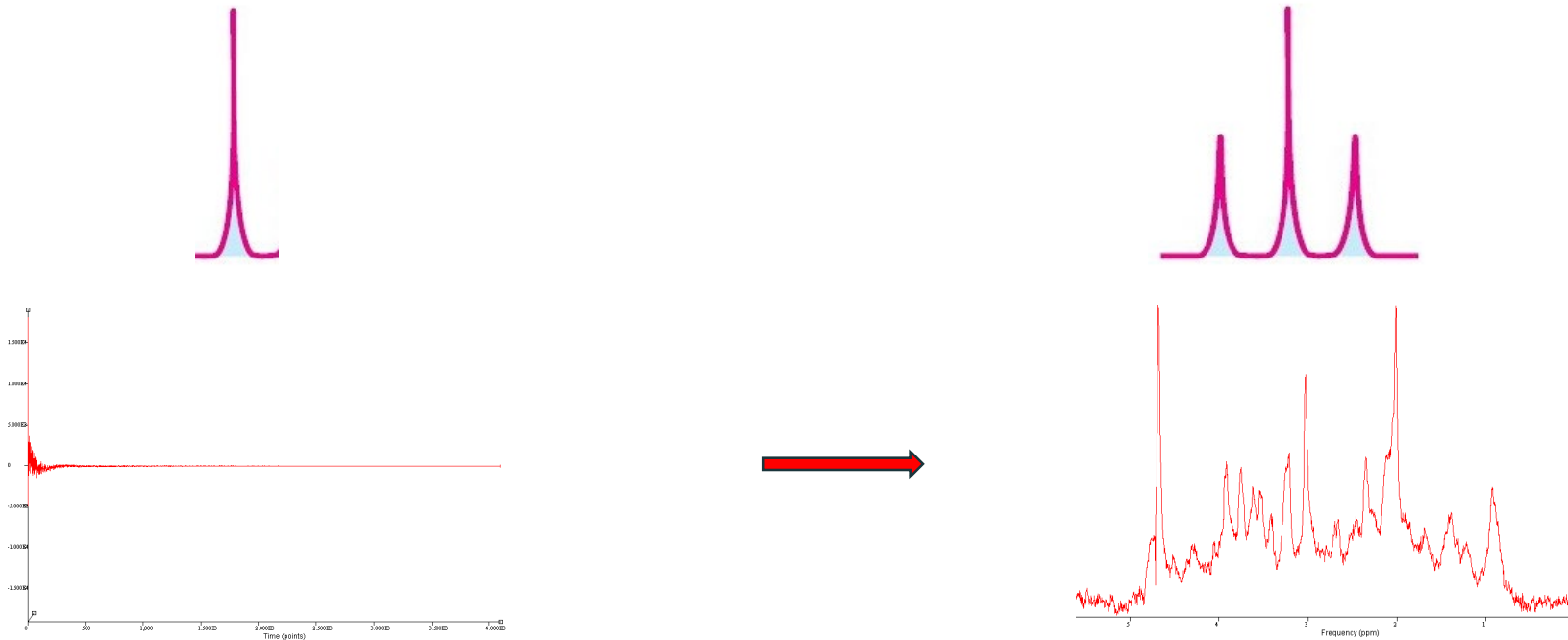
■ In Vivo MR spectroscopy (MRS)

- Measurement of different metabolites- metabolism
- Different organs
- Different nuclei: ^1H , ^{13}C , ^{31}P , ^{15}N , ..
- Different magnetic fields
- Different acquisition parameters



QUANTIFICATION

- MRS – principal goal – quantification of changes in concentration of known metabolites



- Frequencies
- Amplitudes
- Damping factor
- Phase
- Beginning time

- Frequencies
- Surface
- Linewidths
- Zero order phase.
- first order phase

WHY IN VIVO ¹H MR SPECTROSCOPY ?

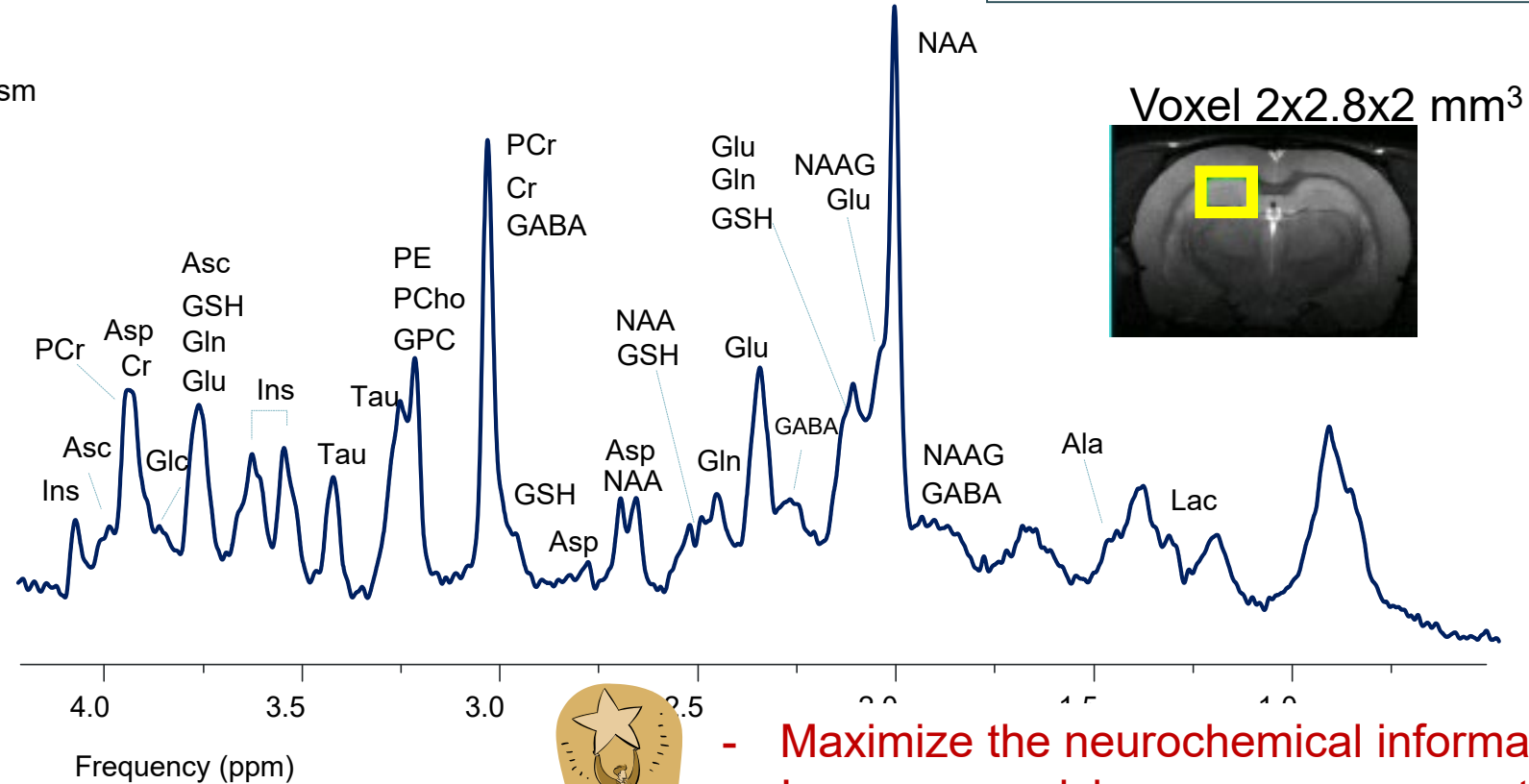
Neurochemical Profile at 9.4T

>18 Markers of :

Myelination/Cell proliferation
Energy metabolism
Osmoregulation
Neurotransmitter metabolism
Antioxidants

- at high magnetic field (9.4T)
- ultra short TE (2.8ms)

- *in vivo*
- non invasively
- localized in hippocampus



- Maximize the neurochemical information
- Increase precision, accuracy – quantification
- Increase the reliability of obtained concentrations





SPECIAL ISSUE REVIEW ARTICLE

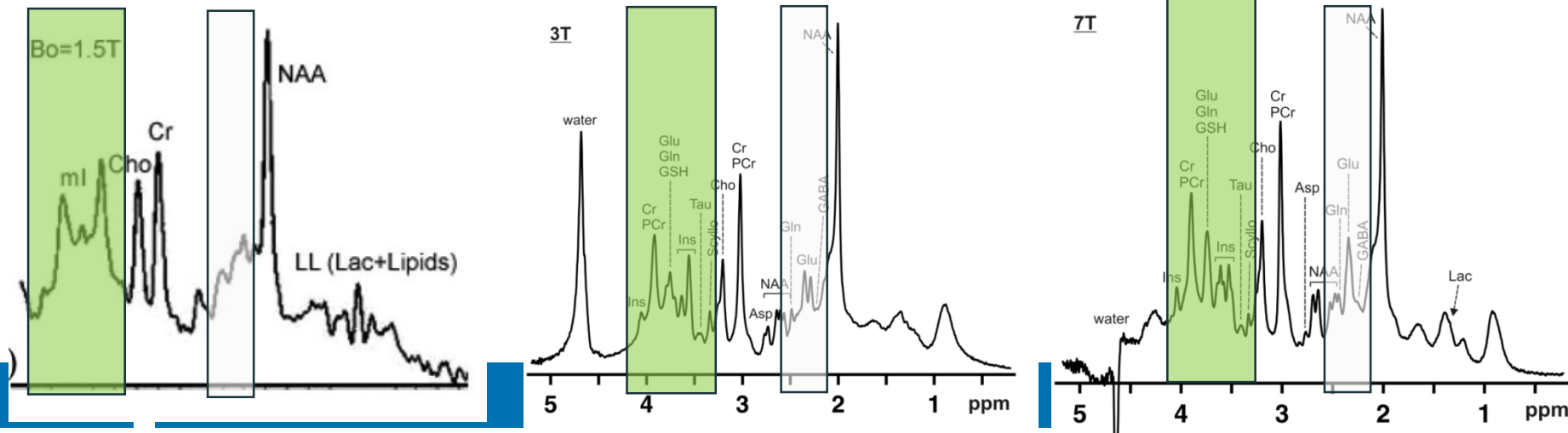
NMR
IN BIOMEDICINE **WILEY**

Terminology and concepts for the characterization of in vivo MR spectroscopy methods and MR spectra: Background and experts' consensus recommendations

Roland Kreis¹  | Vincent Boer²  | In-Young Choi³ | Cristina Cudalbu⁴  |
Robin A. de Graaf⁵  | Charles Gasparovic⁶  | Arend Heerschap⁷ |
Martin Krššák⁸  | Bernard Lanz^{9,10}  | Andrew A. Maudsley¹¹  |
Martin Meyerspeer^{12,13}  | Jamie Near¹⁴  | Gülin Öz¹⁵  | Stefan Posse¹⁶  |
Johannes Slotboom¹⁷  | Melissa Terpstra¹⁵ | Ivan Tkáč¹⁵  | Martin Wilson¹⁸ |
Wolfgang Bogner¹⁹  | Experts' Working Group on Terminology for MR Spectroscopy

WHY HIGH MAGNETIC FIELD ?

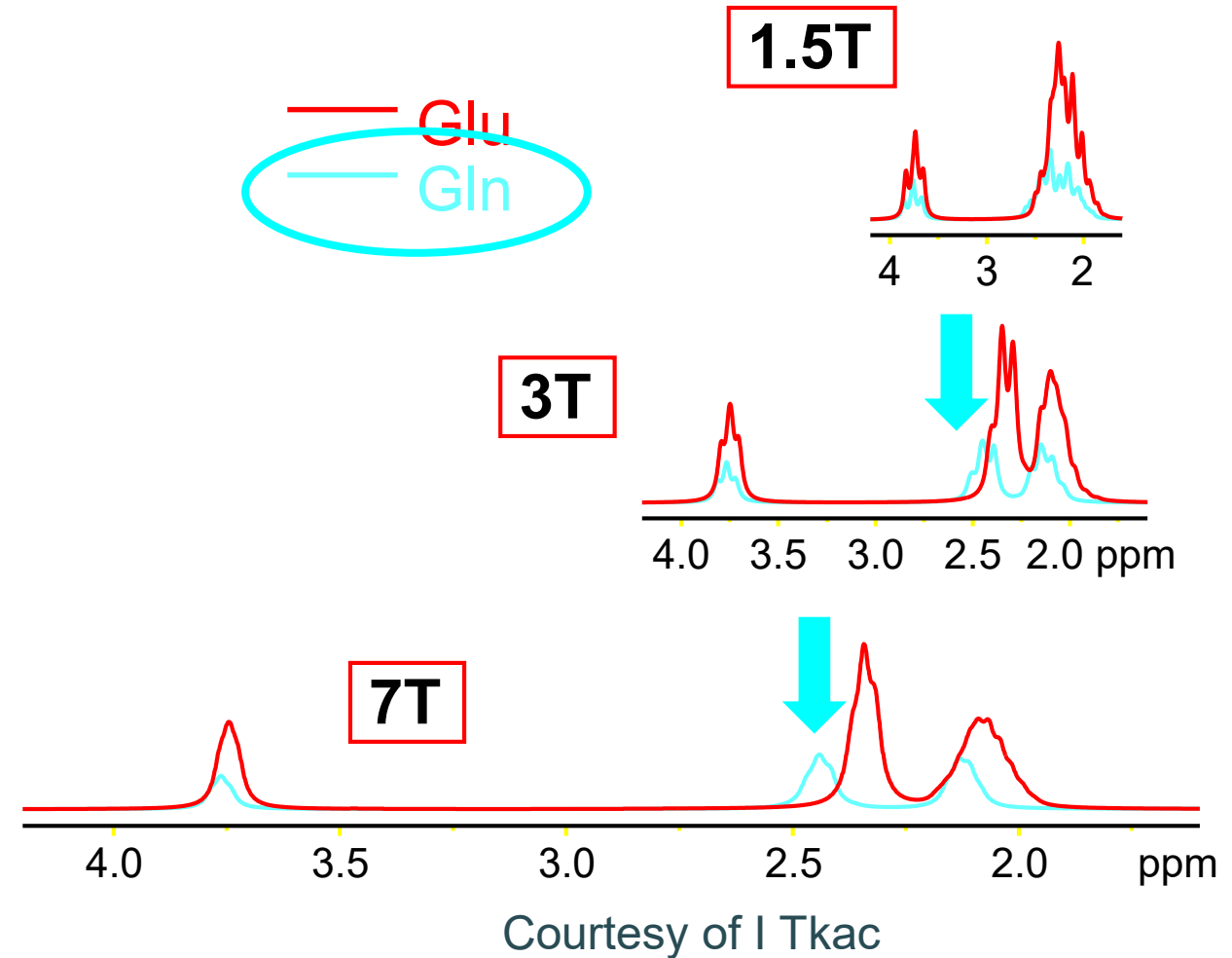
- More signal
- More spectral resolution
- And more sensitivity



WHY HIGH B_0 ?

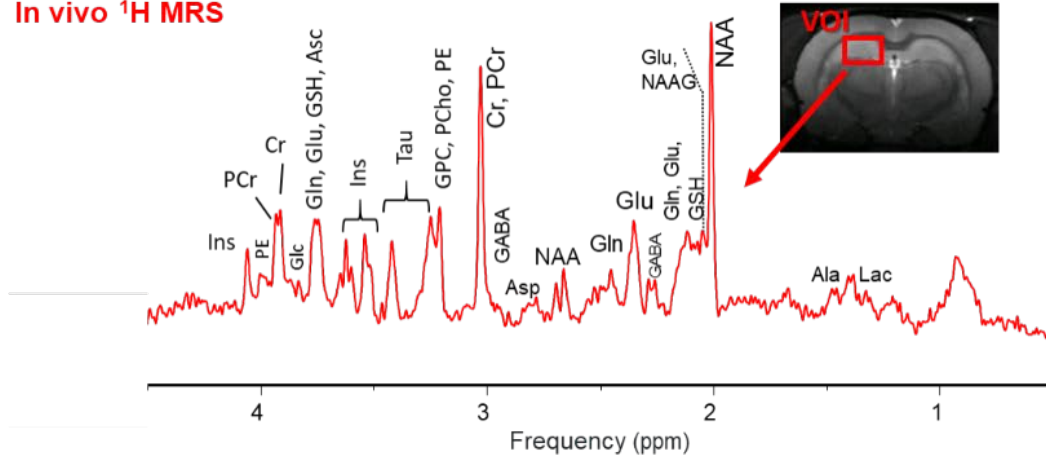
■ Enormous progress

- $\uparrow$ SNR 😊
- $\uparrow$ chemical shift dispersion – $\uparrow$ resolution 😊
- decreased strong J-coupling effects
- Improve quantification precision and accuracy
- $\downarrow T_2^*$ - $\uparrow$ spectral lw in Hz 😞

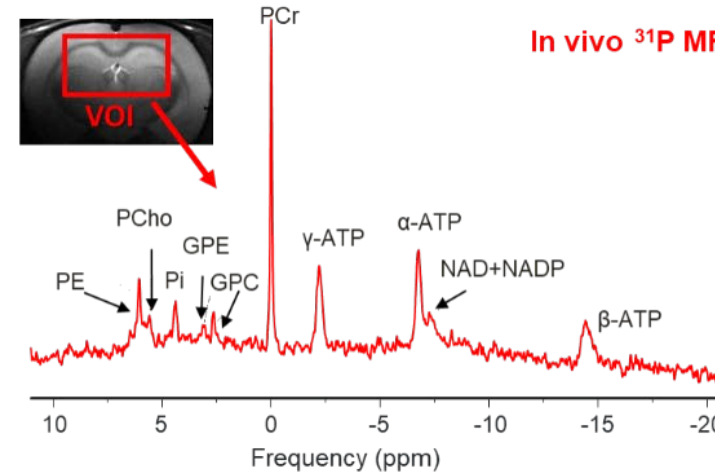


X NUCLEI MRS – 9.4T

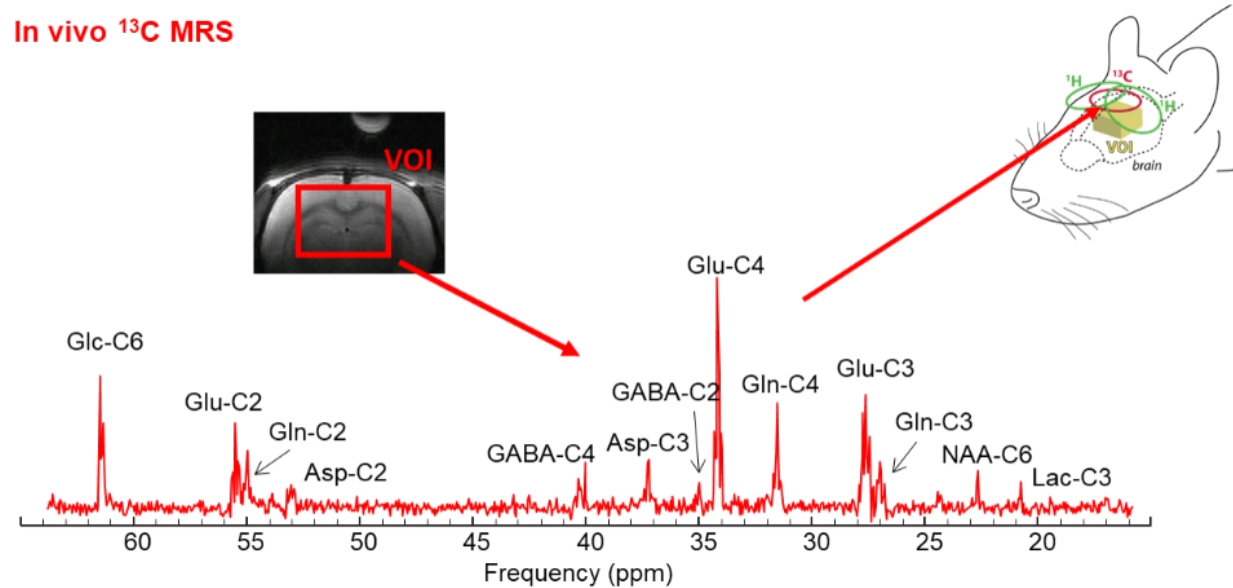
In vivo ^1H MRS



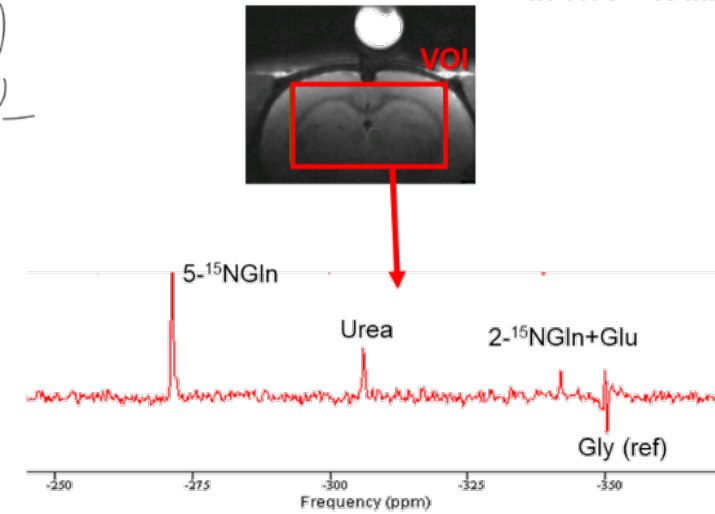
In vivo ^{31}P MRS



In vivo ^{13}C MRS



In vivo ^{15}N MRS



CLINICAL VS PRECLINICAL DATA

	Preclinical	Clinical
Subjects	Animal (rat/mouse) - anesthesia	Human – no anesthesia
Motion	😊	? 😞
Time for scanning	😊 Shim, WS, OVS, ...	😞
@scanner	MRS experts	Not necessary MRS experts
Amount of data	+++	+++++++



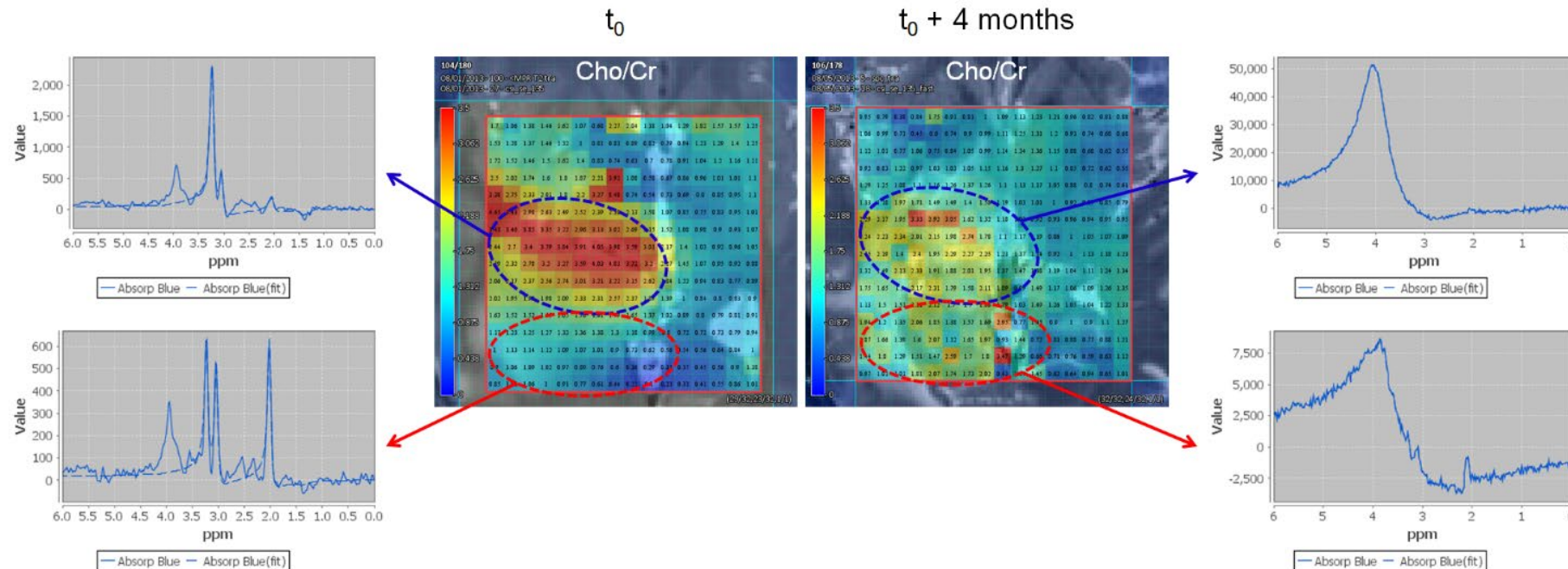
(Lots) Data with sometimes artifacts

- Need to be identified
- Corrected or data discarded
- Preprocessed
- Quantified

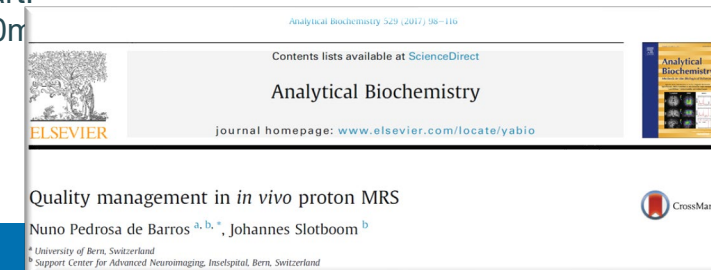


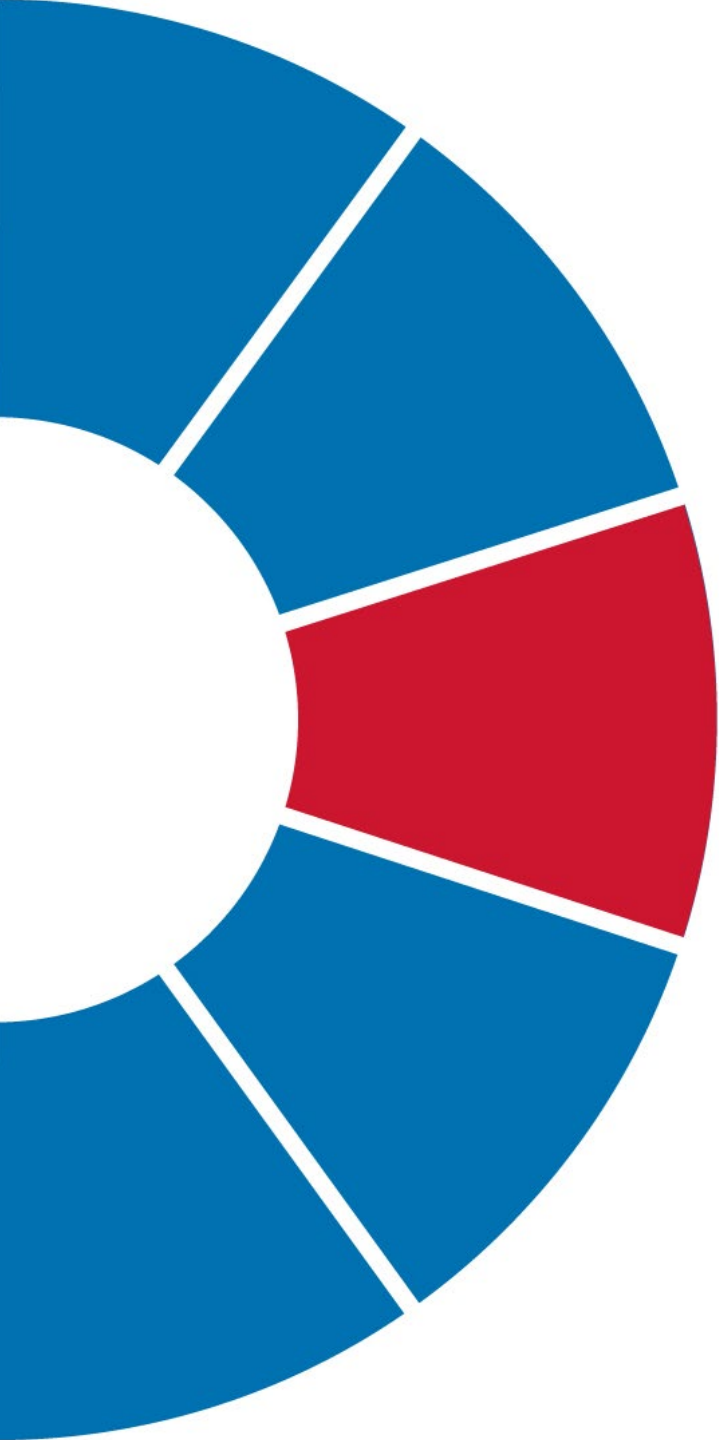
- 😞 Draw incorrect decisions
- 😞 Standard quantification software – NO quality check
- 😞 CRLB

CLINICAL VS PRECLINICAL DATA

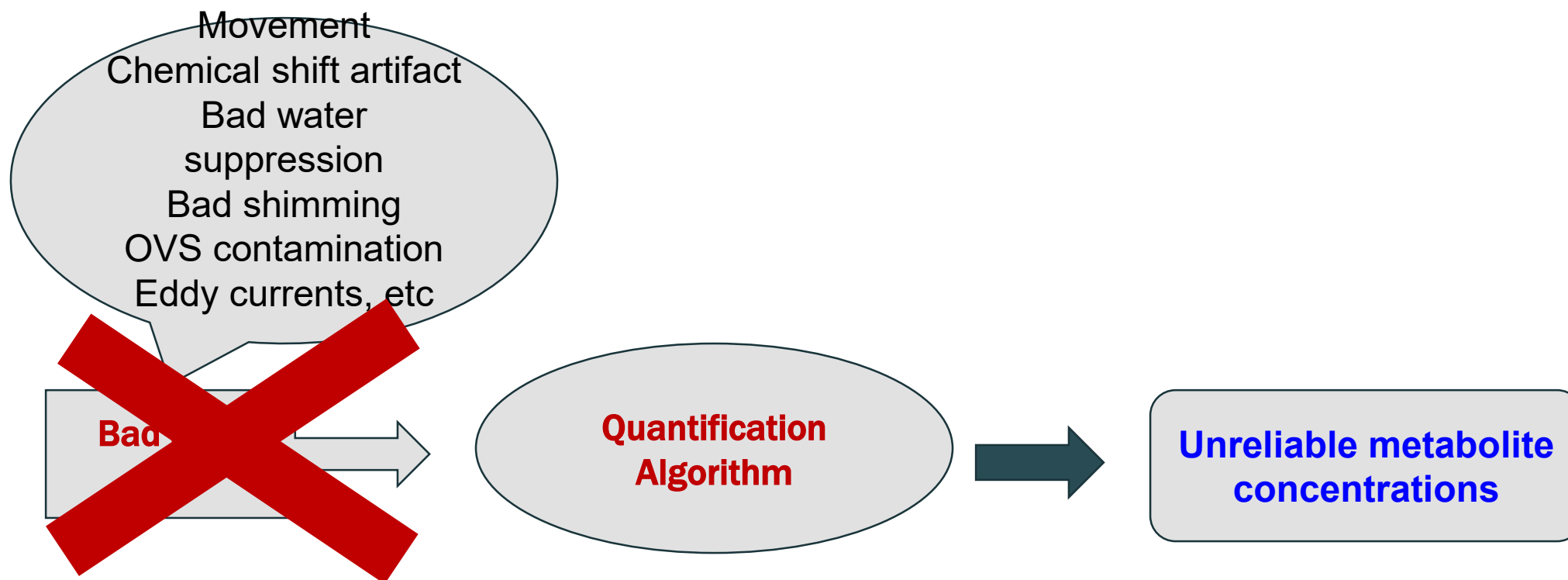


Example showing two Cho/Cr maps of a Glioma patient acquired 4 months apart.
Siemens Aera 1.5 T, 2D-PRESS, 12x12 interpolated to 32x32, TE/TR 135/1500m
From N Barros et al, Analytical Biochemistry, 2017





ARTIFACTS IN 1H MRS

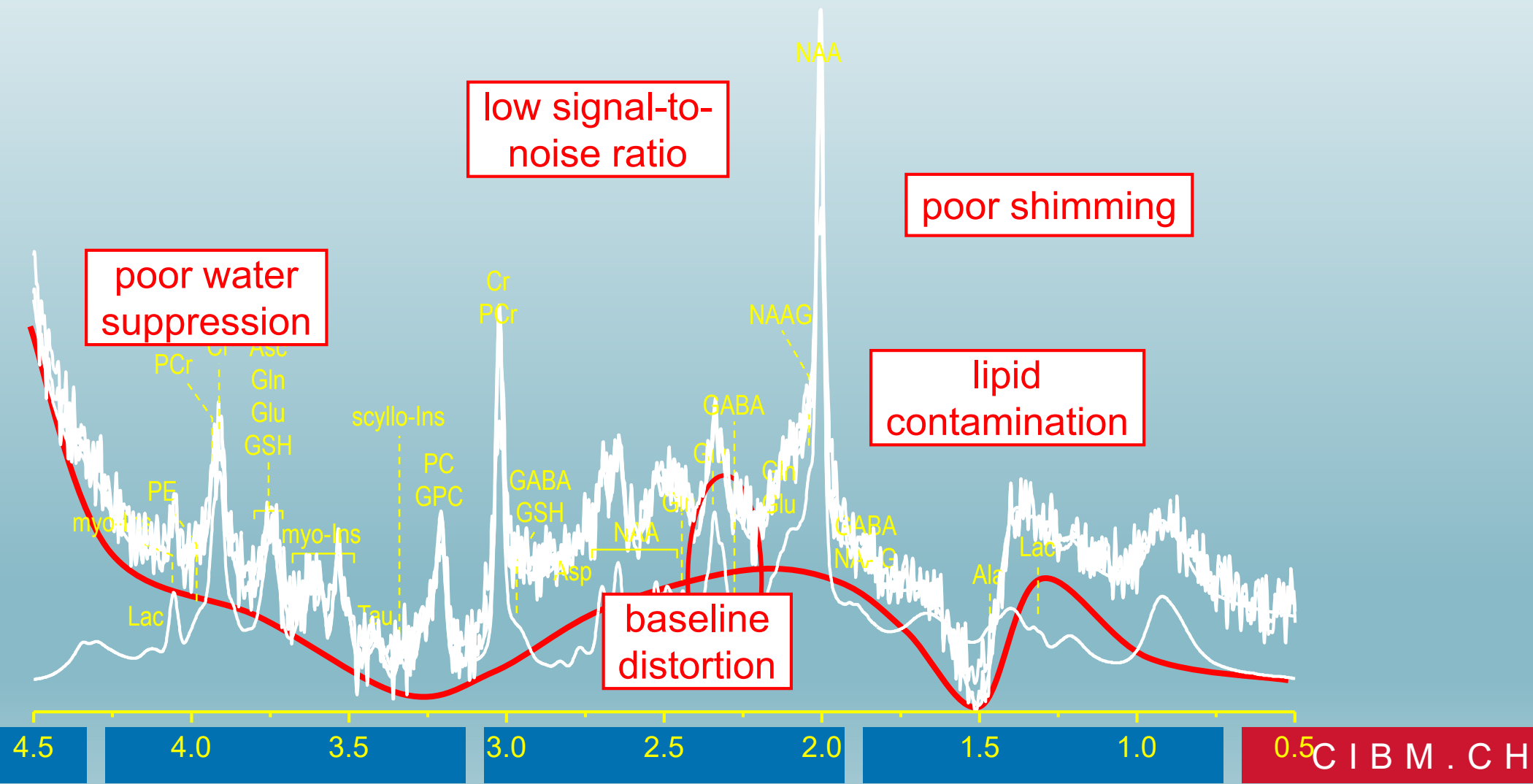


ISMRM 2010, Potentials and Challenges at High Field MRS

R Kreis, NMR Biomed 2004, 17:361

Tkáč I, et al, Appl Magn Reson. 2005 29:139.

DISTORTIONS OF 1H NMR SPECTRUM OF THE HUMAN BRAIN AT 7T



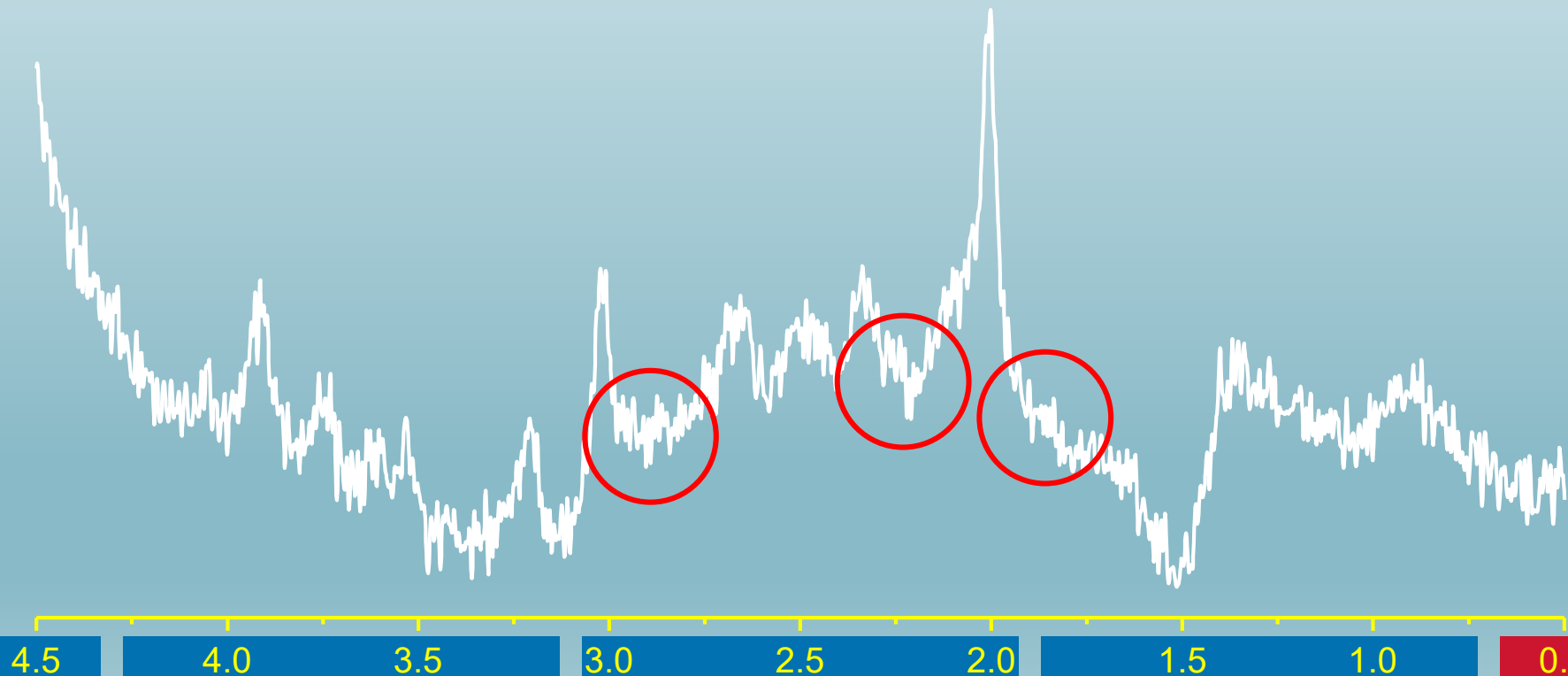
IS RELIABLE QUANTIFICATION OF GABA AND GLU POSSIBLE?

GLUTAMATE?

DIFFICULT, LIMITED PRECISION

GABA?

IMPOSSIBLE FROM THIS TYPE OF SPECTRA!!



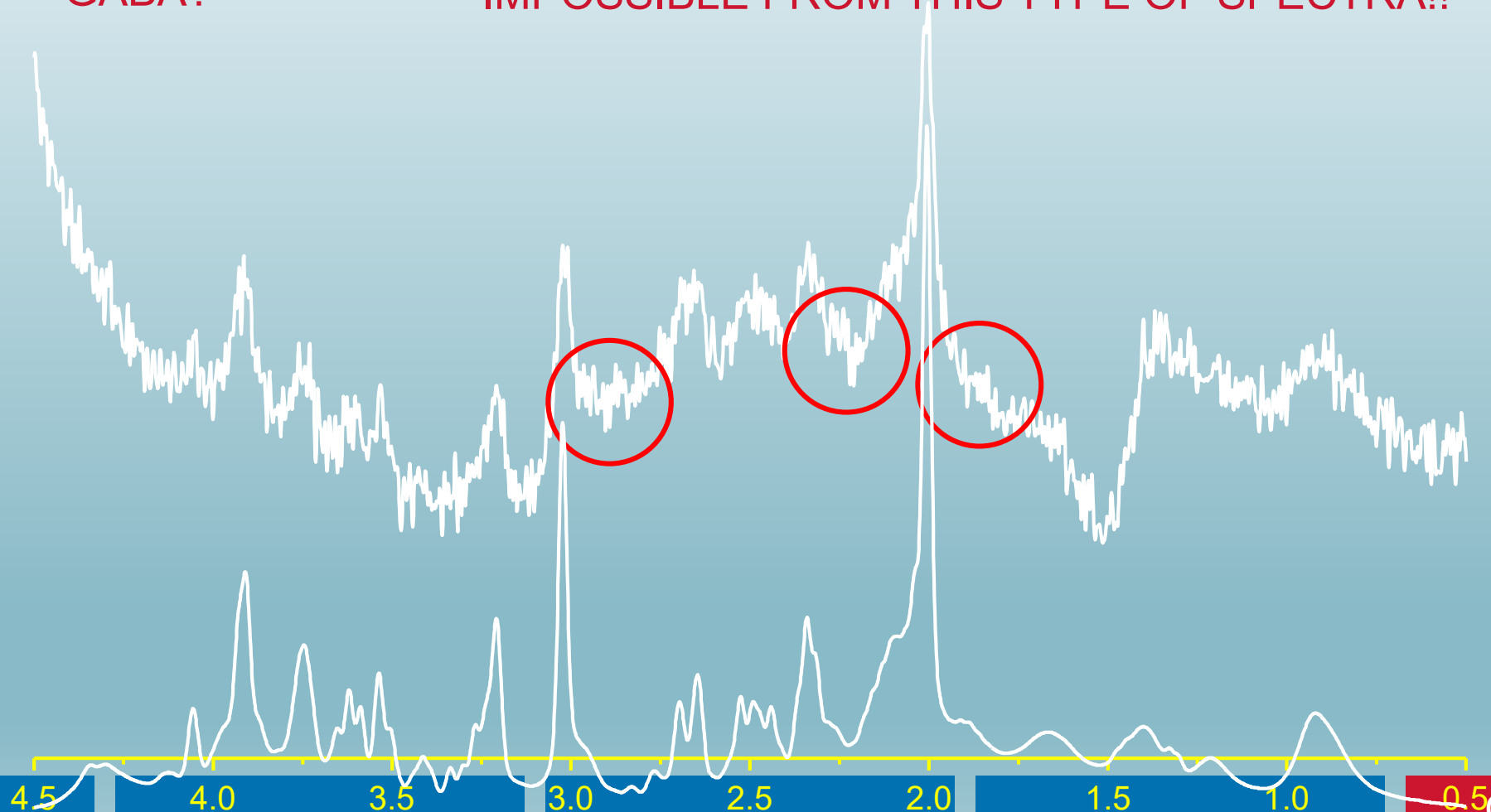
IS RELIABLE QUANTIFICATION OF GABA AND GLU POSSIBLE?

GLUTAMATE?

DIFFICULT, LIMITED PRECISION

GABA?

IMPOSSIBLE FROM THIS TYPE OF SPECTRA!!



CIBM.CH

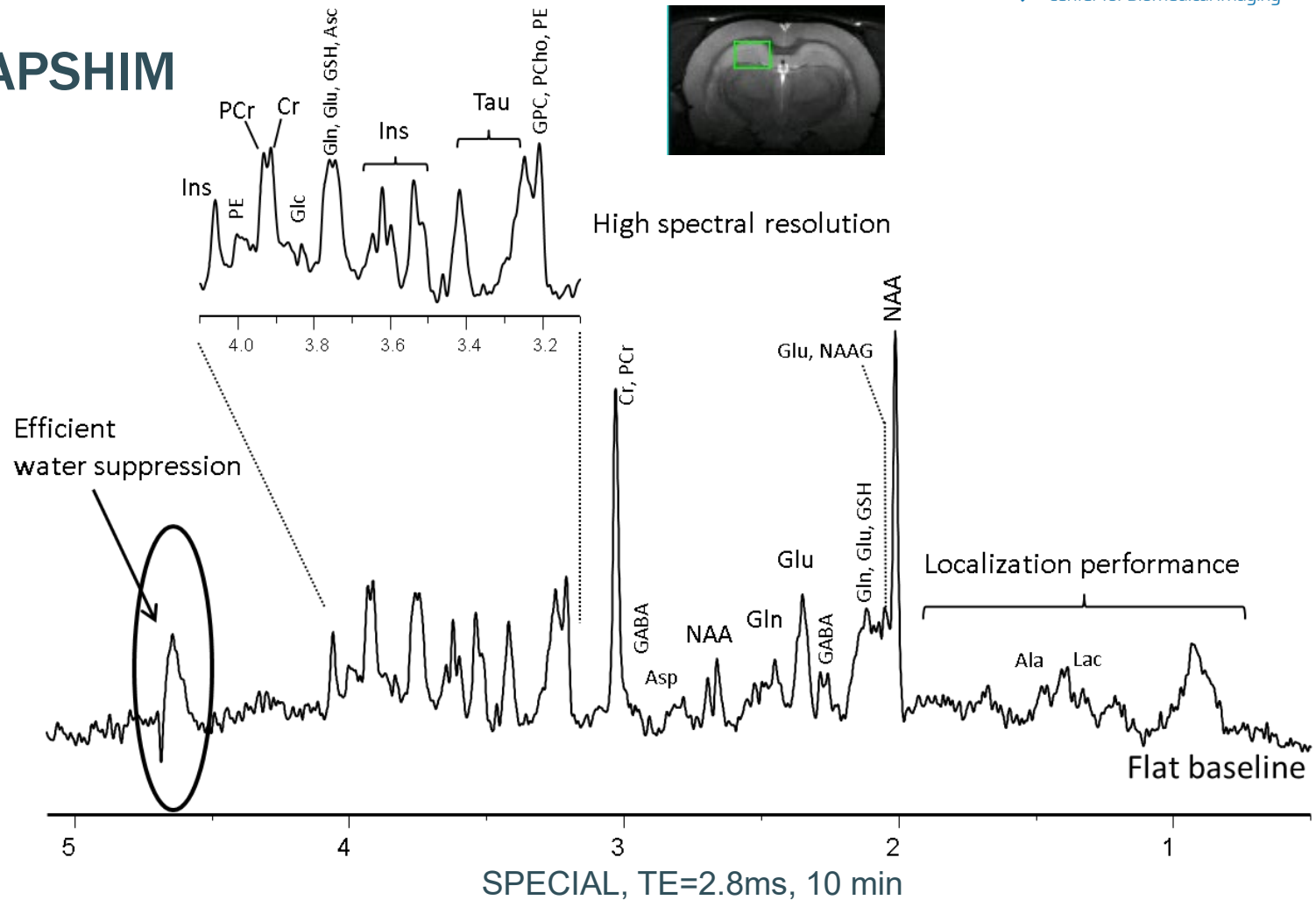
GOOD QUALITY DATA AT SHORT TE

- Shimming: e.g. FASTMAP, MAPSHIM
- Pulse sequence at short TE

Provide undistorted multiplets and no T_2 weighting

Increased no of metabolites

Improved quantification



ACQUISITION SEQUENCES

- Reduced CSDE
- Good localization :
 - double : OVS+Seq or LASER
- Strong crusher gradients
- Good WS

Received: 23 March 2020 | Accepted: 23 November 2020
DOI: 10.1002/nbm.4459

SPECIAL ISSUE REVIEW ARTICLE

NMR
IN BIOMEDICINE WILEY

Water and lipid suppression techniques for advanced ^1H MRS and MRSI of the human brain: Experts' consensus recommendations

Ivan Tkáč¹ | Dinesh Deelchand¹ | Wolfgang Dreher² | Hoby Hetherington³ | Roland Kreis⁴ | Chathura Kumaragamage⁵ | Michal Považan⁶ | Daniel M. Spielman⁷ | Bernhard Strasser⁸ | Robin A. de Graaf⁵

Received: 11 November 2019 | Revised: 29 March 2020 | Accepted: 30 April 2020
DOI: 10.1002/nbm.4325

SPECIAL ISSUE REVIEW ARTICLE

NMR
IN BIOMEDICINE WILEY

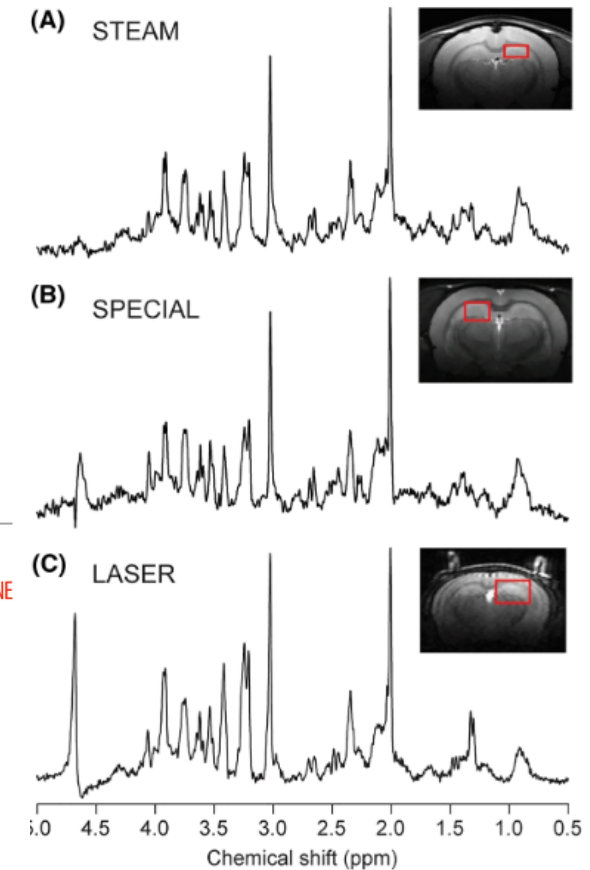


Magnetic resonance spectroscopy in the rodent brain: Experts' consensus recommendations

LANZ ET AL.

NMR
IN BIOMEDICINE WILEY 9 of 20

FIGURE 3 Example ^1H MR spectra obtained in rodent brains at 9.4 T with STEAM (A), SPECIAL (B) and LASER (C) sequences. A, STEAM spectrum: rat brain, $2.3 \times 1.3 \times 2.5 \text{ mm}^3$ voxel placed in the hippocampus, TR = 5 s, TE = 2 ms, TM = 20 ms, number of averages = 448. Spectrum is shown with Gaussian factor = 0.15. B, SPECIAL spectrum: rat brain, $2 \times 2.8 \times 2 \text{ mm}^3$ voxel placed in the hippocampus, TR = 4, TE = 2.8 ms, number of averages = 160. C, LASER spectrum: mouse brain, $1.7 \times 2.25 \times 2.25 \text{ mm}^3$ voxel placed in hippocampus, TR = 4 s, TE = 27 ms, number of averages = 384. The STEAM spectrum was provided by Ivan Tkáč



Received: 15 March 2019 | Revised: 29 October 2019 | Accepted: 7 November 2019
DOI: 10.1002/nbm.4236

SPECIAL ISSUE REVIEW ARTICLE

WILEY NMR
IN BIOMEDICINE

Advanced single voxel ^1H magnetic resonance spectroscopy techniques in humans: Experts' consensus recommendations

Gülen Öz¹ | Dinesh K. Deelchand¹ | Jannie P. Wijnen² | Vladimír Mlynárik³ | Lijing Xin⁴ | Ralf Mekte⁵ | Ralph Noeske⁶ | Tom W.J. Scheenen^{7,8} | Ivan Tkáč¹ | the Experts' Working Group on Advanced Single Voxel ^1H MRS

C I B M . C H

RF COILS

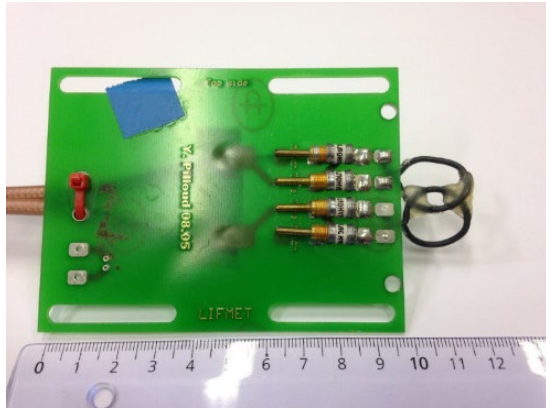
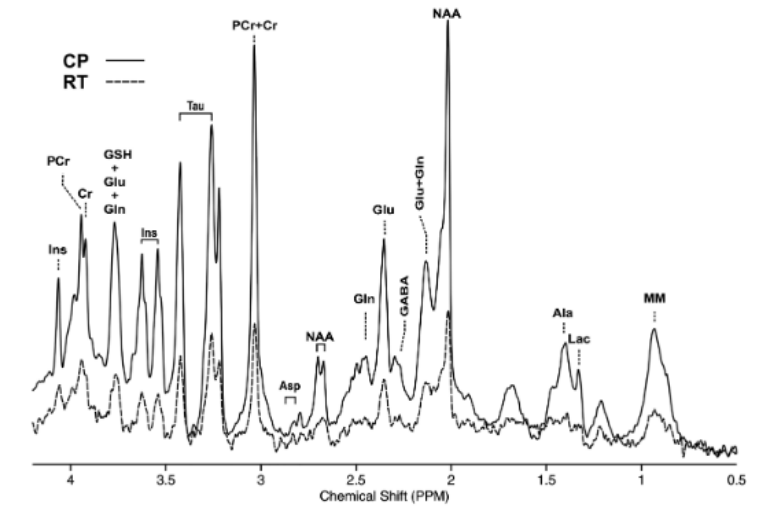


FIGURE 1 Spectra acquired with the STEAM sequence (TR/TE = 5000/3.5 ms, 384 averages) in a $2.0 \times 1.1 \times 2.0 \text{ mm}^3$ VOI located in the mouse frontal cortex. A cryogenically cooled ^1H two-element phased-array transmit/receive coil was employed for excitation and signal reception (solid line). As a comparison, a 72 mm diameter birdcage quadrature volume resonator was used for excitation and a ^1H receive-only 2×2 surface array coil was used for signal reception (dotted line). A 5.2-fold higher SNR was obtained with the cryoprobe (CP) compared with the room-temperature probe (RT)



Received: 11 November 2019 | Revised: 29 March 2020 | Accepted: 30 April 2020
DOI: 10.1002/nbm.4325

SPECIAL ISSUE REVIEW ARTICLE

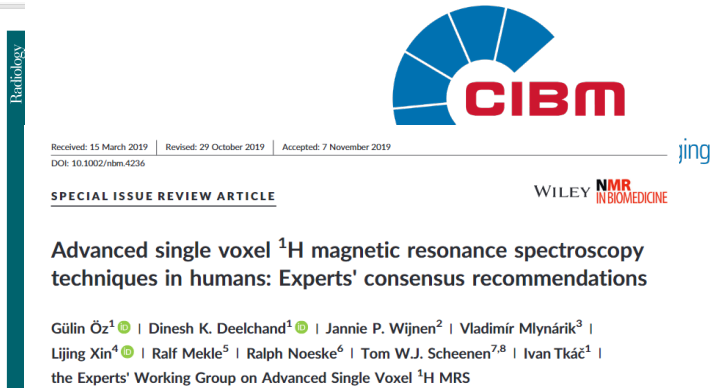
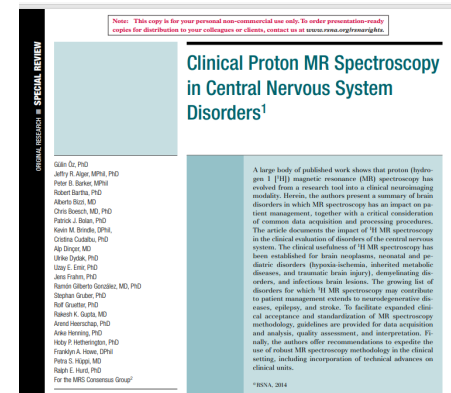
NMR
IN BIOMEDICINE WILEY

Magnetic resonance spectroscopy in the rodent brain: Experts' consensus recommendations

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MINIMUM QUALITY CRITERIA

- SNR >3 for major resonances
- Linewidth - shimming
- Lineshape – symmetric
- WS
- No lipid contamination
- No other visible artifacts
 - Not so obvious artifacts (Chemical shift artifact, localization inaccuracies, signal cancellation, etc)
- CRBs
- Residuals with unexplained features



Signal

Quantification

CIBM.CH

ISMRM 2010, Potentials and Challenges at High Field MRS

R Kreis, NMR Biomed 2004, 17:361

Tkáč I, et al, Appl Magn Reson. 2005 29:139.

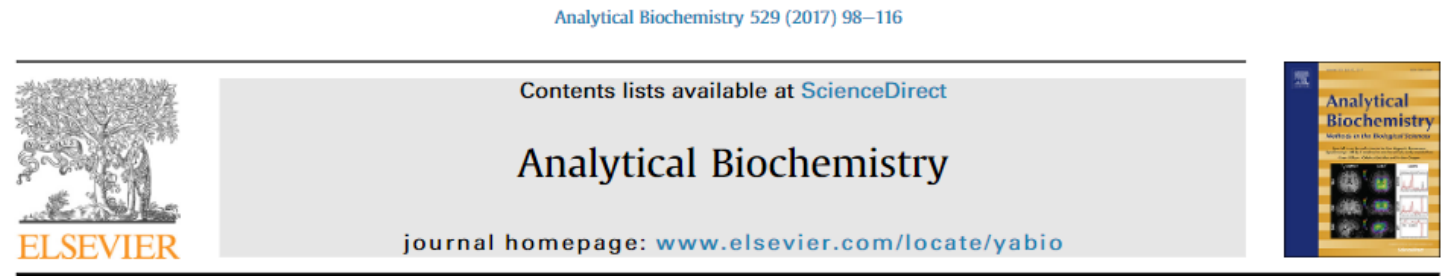
<http://www.ismrm.org/workshops/Spectroscopy16/program.htm>

2nd TRANSACT Workshop, University Bern: Quality Issues in Clinical MRS 2014

The MRS Consensus Group. (2014), Radiology, 270(3):658-79.

QUALITY CONTROL

- **Quality Control** – detection of artifacts (rejection or correction)
 - Signal Quality Control
 - Quantification Quality Control
 - Automatic (semi-automatic)



Quality management in *in vivo* proton MRS

Nuno Pedrosa de Barros^{a, b, *}, Johannes Slotboom^b

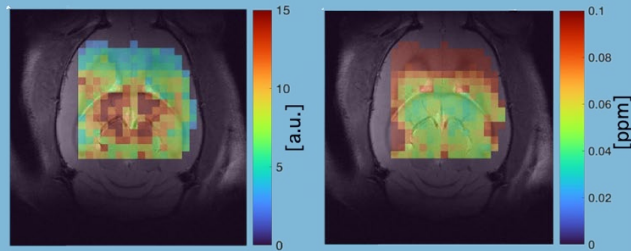
^a University of Bern, Switzerland

^b Support Center for Advanced Neuroimaging, Inselspital, Bern, Switzerland



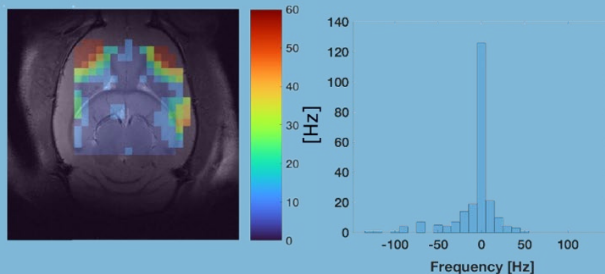
MRS4BRAIN TOOLBOX : MRSI (QUALITY CONTROL)

Quality check (*before fitting*)



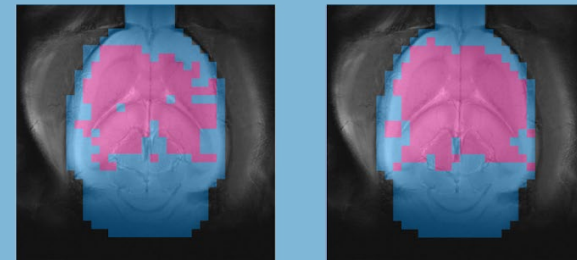
SNR

Linewidth



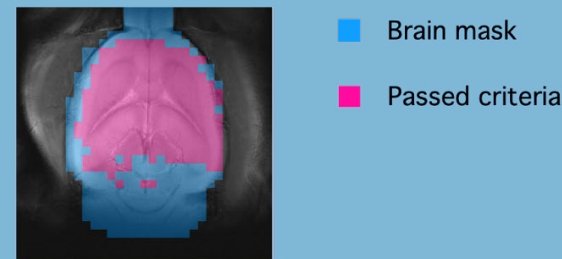
B0 map and histogram

Quality control (*after fitting*)



$SNR > 0.75 \times \overline{SNR}$

$FWHM < 1.25 \times \overline{FWHM}$



$CRLB < 30 \%$

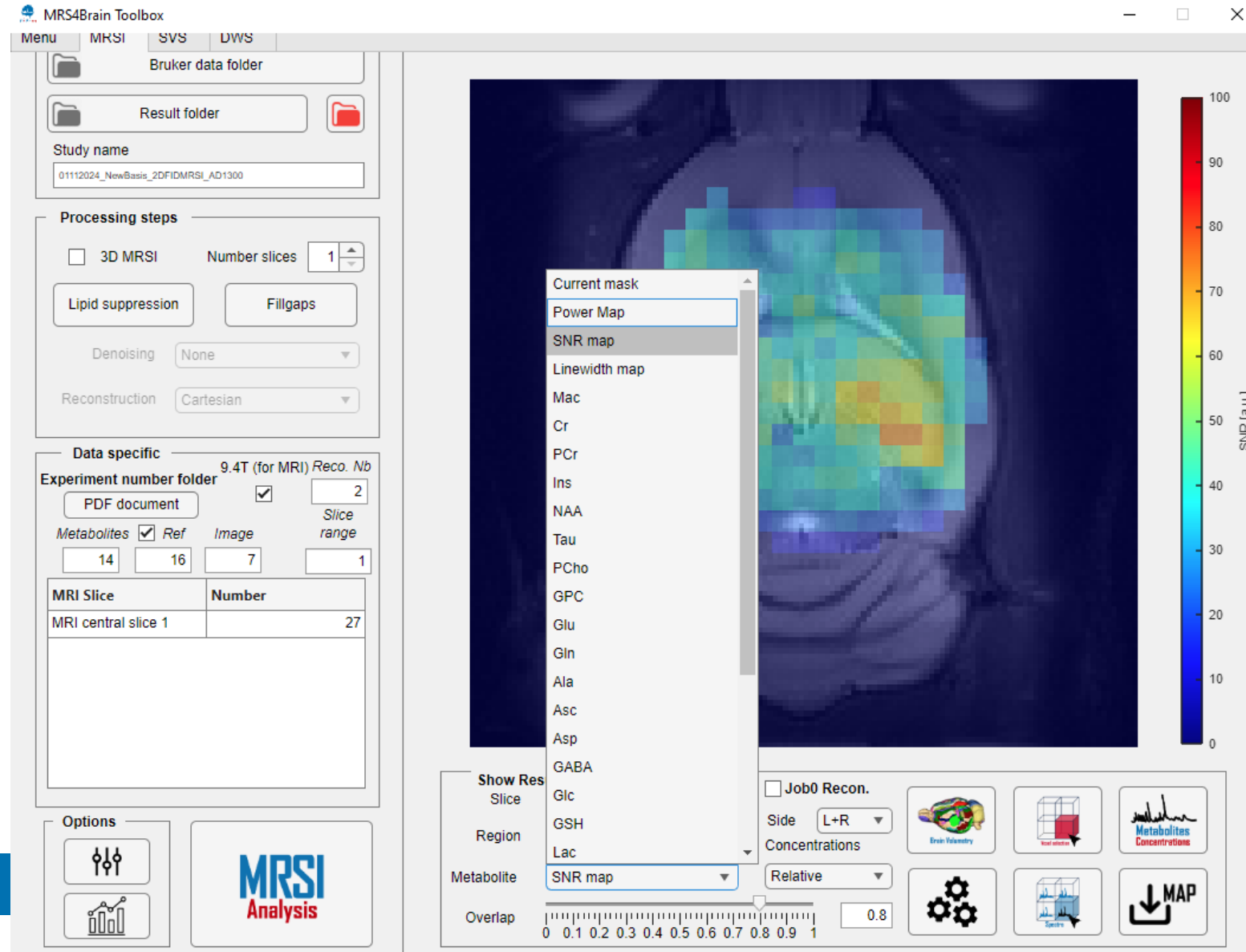
- 4 Quality Metrics reported :
Signal-to-noise ratio (SNR),
linewidth (lw or FWHM), B_0
shift & Cramer Rao lower
bound (CRLB)
- Quality check before fitting
(no masking)
- Quality control after fitting
(masking of unwanted data)

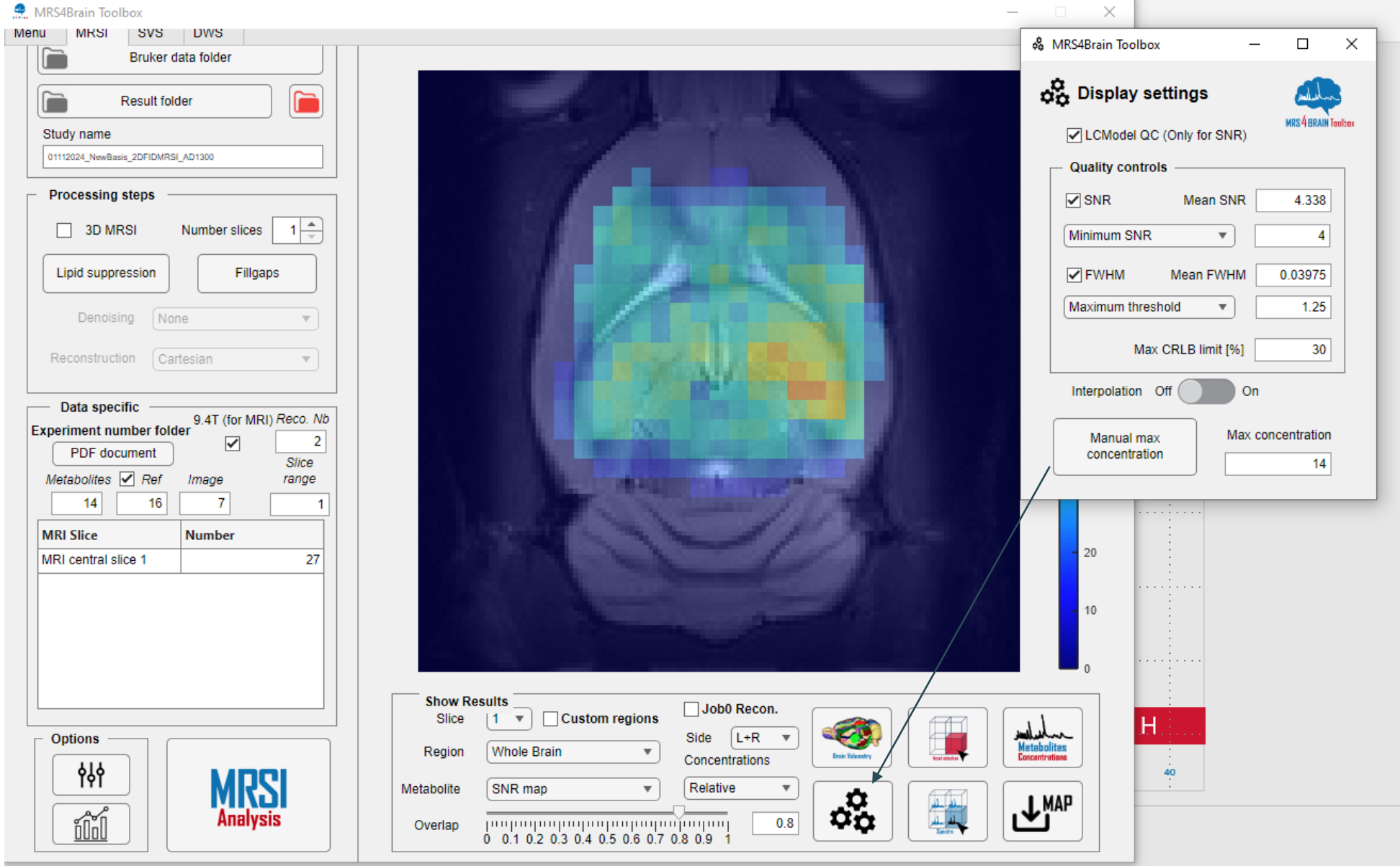


See all video recordings available on gdocs

QUALITY CONTROL IN MRS4BRAIN TOOLBOX

See all video recordings available on gdocs





See all video recordings available on gdocs

Choose a directory

← → ↕

This PC > cudalbu_biblio (\\cibmaitsv1.epfl.ch) (Z:) > DATA > CSI > 9.4T-FIDCSI-Data > 20241101_091603_011124_Fem_MRSI2D3D_SVSMM_011124_Fem_MRSI2D3D_SVSMM_1_1

Search 20241101_091603_0111...

Organize New folder

R

Jessie_DWSdata

MM_DIR-IR_2018

jMRUI_v7.0_build3

scannerdata

DWMRS_Workshop_2023

Outlook

Projects

protocols

Creative Cloud Files

OneDrive - epfl.ch

This PC

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Documents

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MRS 2022 Workshop

Music

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Local Disk (C:)

Data2024 (E:)

PhysioSheets (\\sb1files\LIFMET) (L:)

Kasia (\\cibmaitsv1.epfl.ch) (Q:)

Animal Facility (\\cibmaitsv1) (T:)

lifmet (\\sb1files) (U:)

Volumetry (\\cibmaitsv1.epfl.ch) (V:)

Volumetry (\\cibmaitsv1) (W:)

BDL (\\cibmaitsv1.epfl.ch) (X:)

glioma (\\cibmaitsv1.epfl.ch) (Y:)

cudalbu_biblio (\\cibmaitsv1.epfl.ch) (Z:)

Network

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12	01/11/2024 11:04	File folder	
13	01/11/2024 11:04	File folder	
14	01/11/2024 11:48	File folder	
15	01/11/2024 11:05	File folder	
16	01/11/2024 11:05	File folder	
17	01/11/2024 13:03	File folder	
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20	01/11/2024 13:03	File folder	
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23	01/11/2024 14:47	File folder	
24	01/11/2024 14:31	File folder	
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AdjResult	01/11/2024 17:59	File folder	
Linewidths	22/11/2024 14:47	File folder	
Masks	01/11/2024 18:17	File folder	
PowerMaps	01/11/2024 18:07	File folder	
registration	01/11/2024 12:00	File folder	
shims	01/11/2024 17:59	File folder	
SNR_maps	22/11/2024 14:42	File folder	

Folder: Linewidths

Select FolderCancel

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PREPROCESSING STEPS

- Phase offsets (0 and 1st order)
- Subject motion
- Scanner drift
- Eddy currents
- Alignment of spectra that need to be added/subtracted
- Water contamination
-

PREPROCESSING STEPS

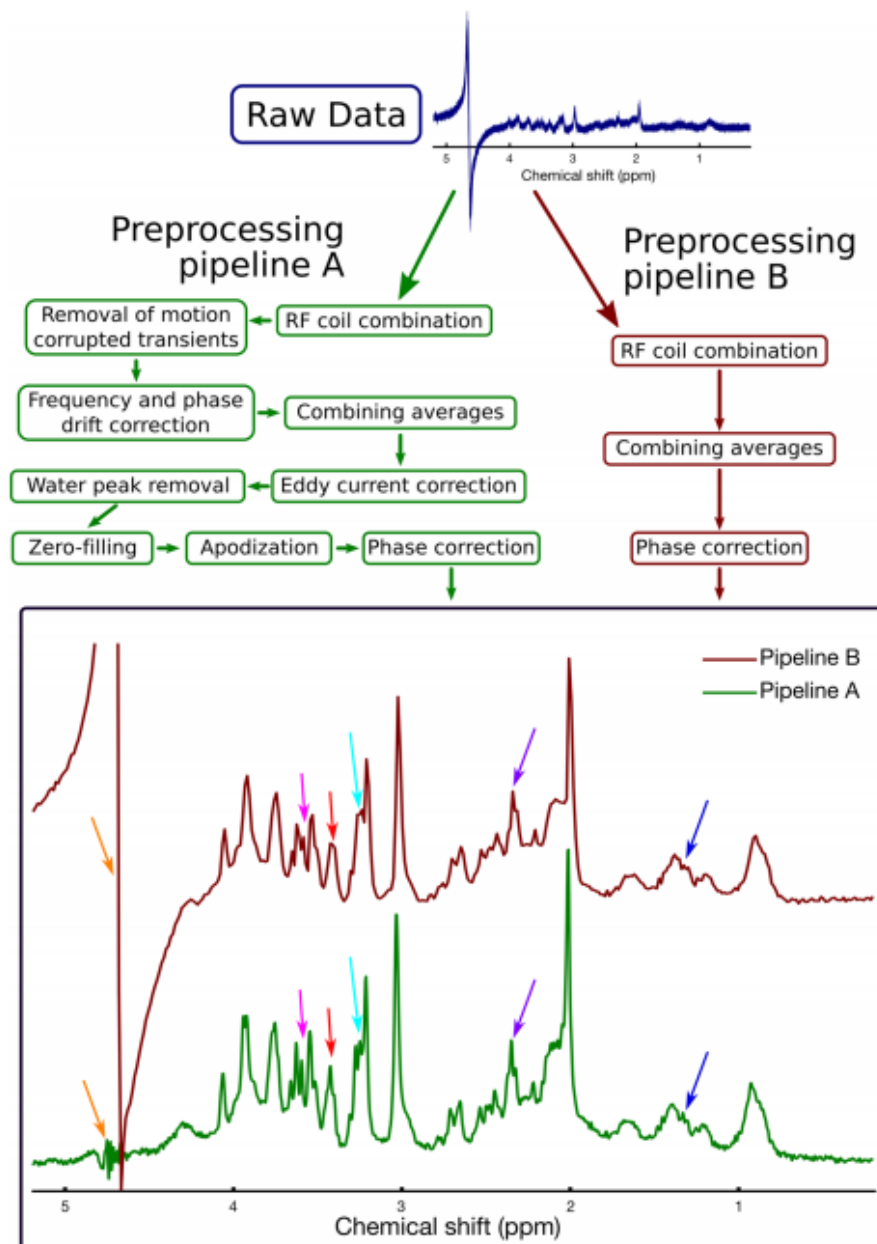
■ Why ?

- quantification algorithms do not account for all of them
- Lead to errors in the quantification process

■ Requirements:

- Automated methods
- Included in the quantification software

FIGURE 4 Illustration of two example processing pipelines, applied to the same raw data. The dataset was obtained from a rat brain using the PRESS sequence at 7 T with $T_E = 11$ ms. Processing pipeline B (dark red boxes, right-hand side) includes only basic steps to combine the coils and transients (similar to the standard processing pipeline provided by clinical scanner vendors). Processing pipeline A (green boxes, left-hand side) involves additional steps to remove motion-corrupted averages, to retrospectively correct frequency and phase drift, and to remove eddy current artefacts. Pipeline A resulted in several noticeable improvements in spectral quality, including reduced water contamination (orange arrows), and improved visual definition of most spectral peaks, including lactate (1.3 ppm, dark blue arrows), glutamate-H4 (2.3 ppm, purple arrows), tCho (3.2 ppm, light blue arrows), taurine (3.4 ppm, red arrows) and myo-inositol (3.5 ppm, pink arrows). These improvements highlight the importance of using an appropriate processing pipeline. Note that, as stated in the recommendation tables, zero-filling and apodization may be used to improve the visual appearance of the spectrum, but should not be performed prior to spectral analysis



MRS4Brain Toolbox

Menu MRSI SVS DWS

CIBM Center for Biomedical Imaging **MRS4BRAIN** Toolbox

Data management

Bruker data folder

Result folder

Study name

Processing steps

☐ 3D MRSI Number slices 1

Lipid suppression Fillgaps

Denoising None

Reconstruction Cartesian

Data specific

9.4T (for MRI) Reco. Nb

Experiment number folder PDF document 1

Metabolites ☒ Ref Image Slice range

20 21 12 1

MRI Slice	Number
MRI central slice 1	8

Options

MRSI Analysis

Show Results

Slice 1 ☐ Custom regions ☐ Job0 Recon.

Region Whole Brain Side L+R

Concentrations NAA Relative

Overlap 0.8

0 0.1 0.2 0.3 0.4 0.5 0.6 0.7 0.8 0.9 1

Brain Volume

Metabolite Concentrations

MAP

Concentration C+PCr

1 0.9 0.8 0.7 0.6 0.5 0.4 0.3 0.2 0.1 0

See all video recordings available on gdocs

SVS/DWS preferences

Preferences SVS/DWS

☒ NRATIO

LCModel path Basis set

Configs STEAM_Brayan_Tests_13112024 Original

☐ NSIMUL PPM start 4.3

☐ VITRO PPM end 0.2

NRATIO 12 DKNTMN 0.25

WCONC 4.444e+0.

Relative metabolite Cr+PCr

Relative concentration 8

Combination

NCOMB 4

Index	Combination
1	NAA+NAAG
2	Glu+Gln
3	GPC+PCho
4	Cr+PCr

Omission

NOMIT 15

Index	Omitted
1	-CrCH2
2	Gua
3	Ser
4	Lip13a
5	Lip13b
6	Lip09
7	MM09
8	Lip20
9	MM20
10	MM12
11	MM14

Use

NUSE 7

Index	Used
1	NAA
2	Gln
3	PCr
4	Cr
5	Ins
6	Tau
7	PCho

Pre processing

Fid-A steps : ☒ Align averages ☐ Outliers removal ☒ Small Voxel

Line Broadening 12

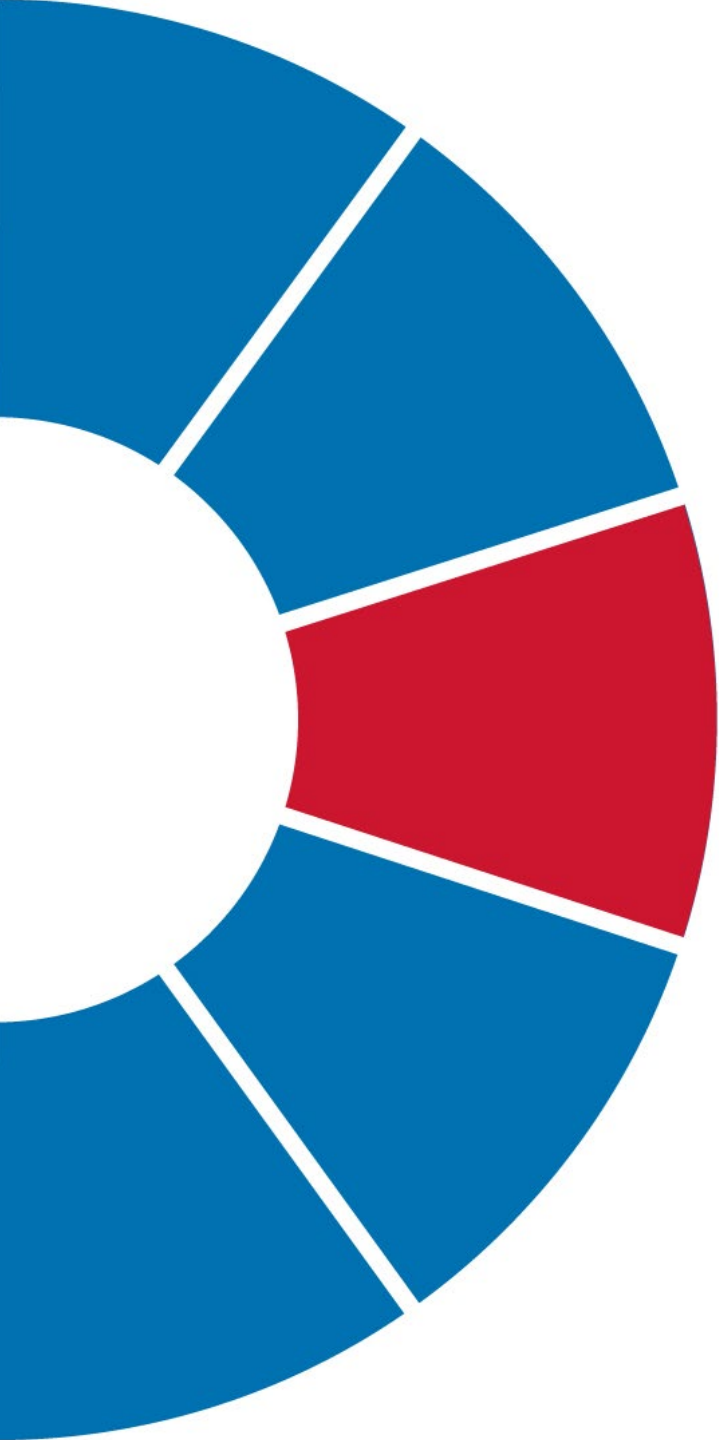
Frequency range [ppm] : min 7 max 8

Rejection threshold 1.5

Maximum time 0.5

Save Delete ISIS

See all video
recordings
available on gdocs

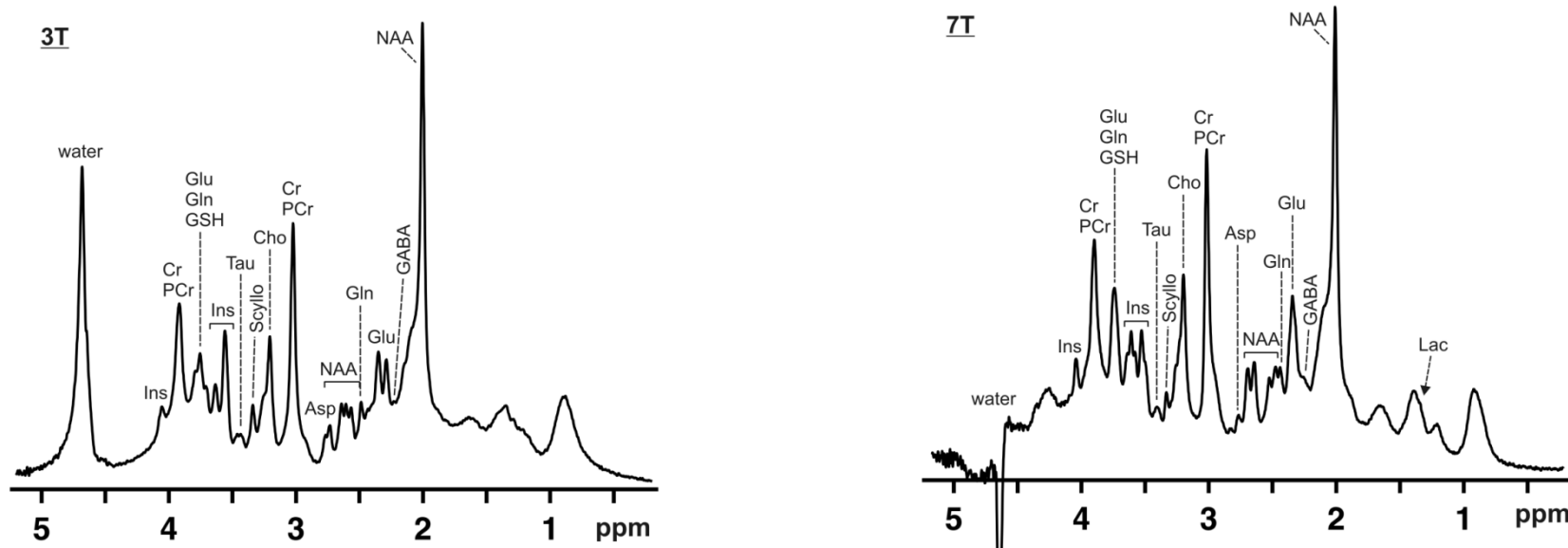


QUANTIFICATION SOFTWARE

QUANTIFICATION

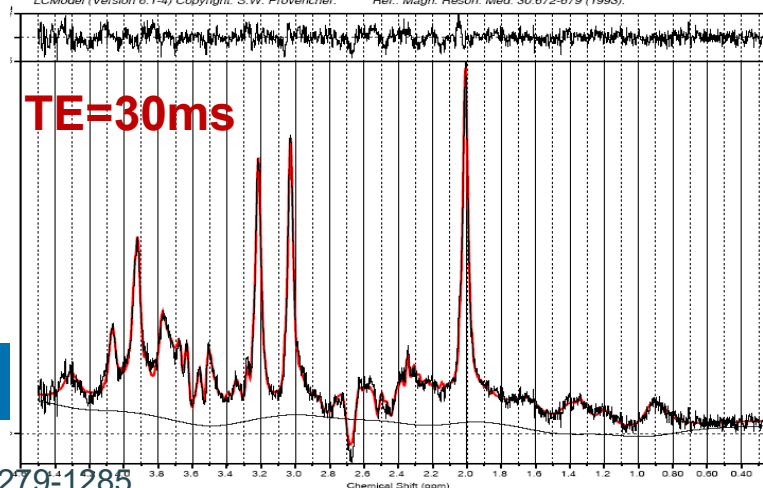
- MRS – principal goal – **quantification of changes in concentration of known metabolites**
- Accurate and precise quantification:
 - Signal quality
 - Hardware performance
 - Pulse sequence design & adjustment
 - Data (pre)processing (estimate the signal amplitude or peak area)
 - Quantification strategies (→ tissue content)
- Final Goal:
 - Maximize the neurochemical information
 - Increase the precision and accuracy of quantification
 - Maximize the reliability of neurochemical data

QUANTIFICATION



Gambarota_Giulio (07.03.07-16:42:34-STD-1.3.12.2.1107.5.2.34.18931) Series/Acq=31/1 (2007.11.05 18:04
TR/TE/NS=4000/30/64, 8.0mL (M 042Y, 80kg) RALF Spectroscopy
Data of: Center for Biomedical Imaging, Lausanne

LCModel (Version 6.1-4) Copyright: S.W. Provencher. Ref.: Magn. Reson. Med. 30:672-679 (1993).



Signals are different (acq param, Bo, nucleus, etc)

– quantification is also different

THE ALGORITHMS ARE AUTOMATIC
BUT NOT FULLY PUSH-BUTTON
NEED TO BE ADAPTED

QUANTIFICATION

In vivo spectra - high B_0
- short TE



Increased - sensitivity
- spectral resolution

is difficult:

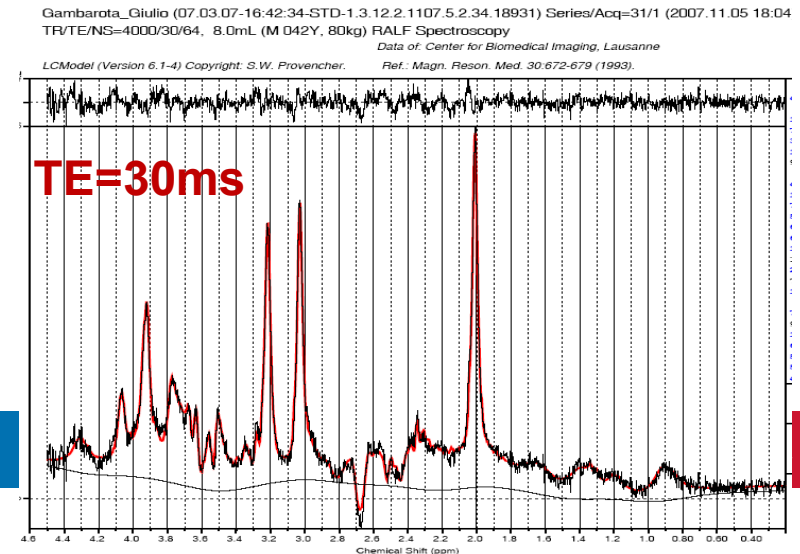
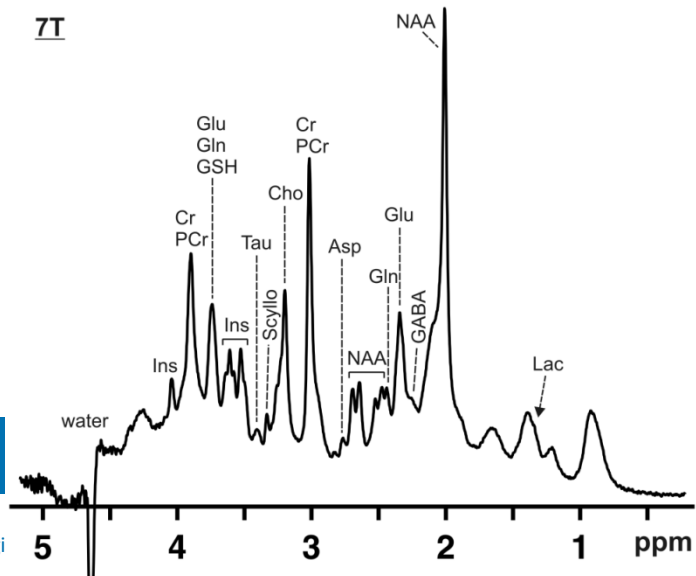
- Complexity of the spectra:
 - many resonances
 - peak overlap
 - contribution of macromolecules and residual water
- Unpredictable lineshape :
 - residual eddy currents
 - field inhomogeneity
- Unpredictable baseline:
 - macromolecules
 - lipid signals
 - partially suppressed water

QUANTIFICATION

In vivo spectra - high B_0
- short TE



Increased - sensitivity
- spectral resolution



Ralf Mekle & Giulio Gambarota

☹️ Vendors software - visualization of spectra

☹️ Peak fitting - !!!!! Overlapping peaks ☹️

😊 X nuclei

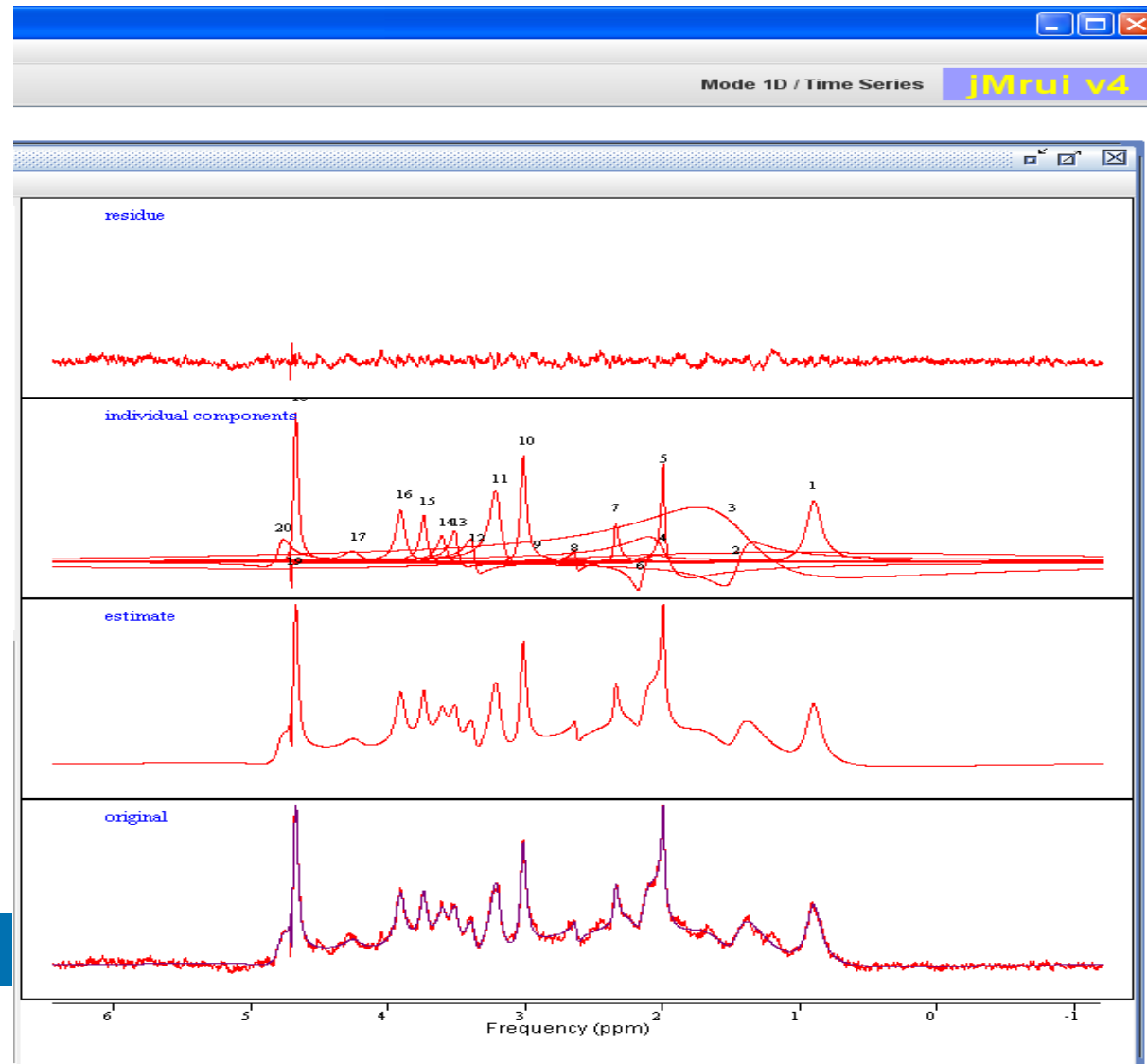
😊 “Basis-set” fitting – In vivo MRS spectrum is modelled as a linear combination of individual metabolite basis spectra

- Experimentally
- Simulated
 - User-friendly software packages: NMRScopeB, Vespa, GAMMA, GAVA,
- Macromolecules contribution
- Lipids contribution (i.e. malignant brain tumors)

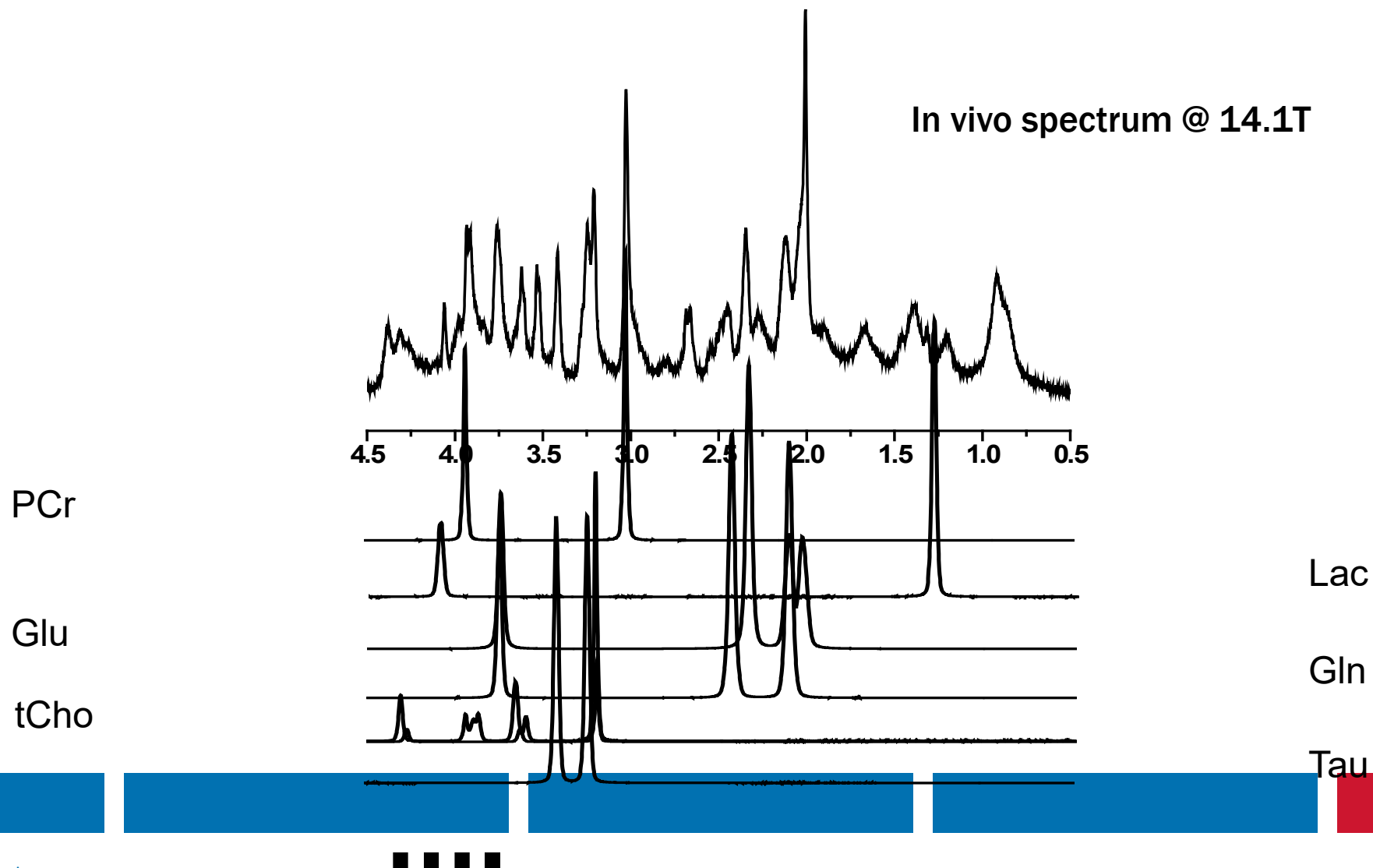
SHORT DESCRIPTION OF THE ALGORITHMS

HLSVD quantification

Amplitude	sd. Amp.
666.88	0.2707
676.74	0.1729
333.53	2.40
140.71	0.6707
1.971E4	0.9626
1.111E3	3.45
1.455E3	2.14
479.78	138.56
475.13	34.35
420.84	46.61
551.35	18.94
837.73	11.91
351.08	226.13
1.112E3	0.2797
33.12	0.0265
614.40	0.0191



QUANTIFICATION WITH A BASIS SET



MRSI parameters

☒ NRATIO

LCModel path Basis set

Configs Original ☒

☐ NSIMUL ☐ VITRO

PPM start PPM end DKNTMN WCONC

NRATIO

Relative metabolite Relative concentration

Licence
Owner
Key

Config name

Combination
NCOMB

Index	Combination
1	NAA+NAAG
2	Glu+Gln
3	GPC+PCho
4	Cr+PCr

Omission
NOMIT

Index	Omitted
1	-CrCH2
2	Gua
3	Ser
4	Lip13a
5	Lip13b
6	Lip09
7	MM09

Use
NUSE

Index	Used
1	NAA
2	Gln
3	PCr
4	Cr
5	Ins
6	Tau
7	PCho

Registration

See all video recordings available on gdocs

QUANTIFICATION SOFTWARE

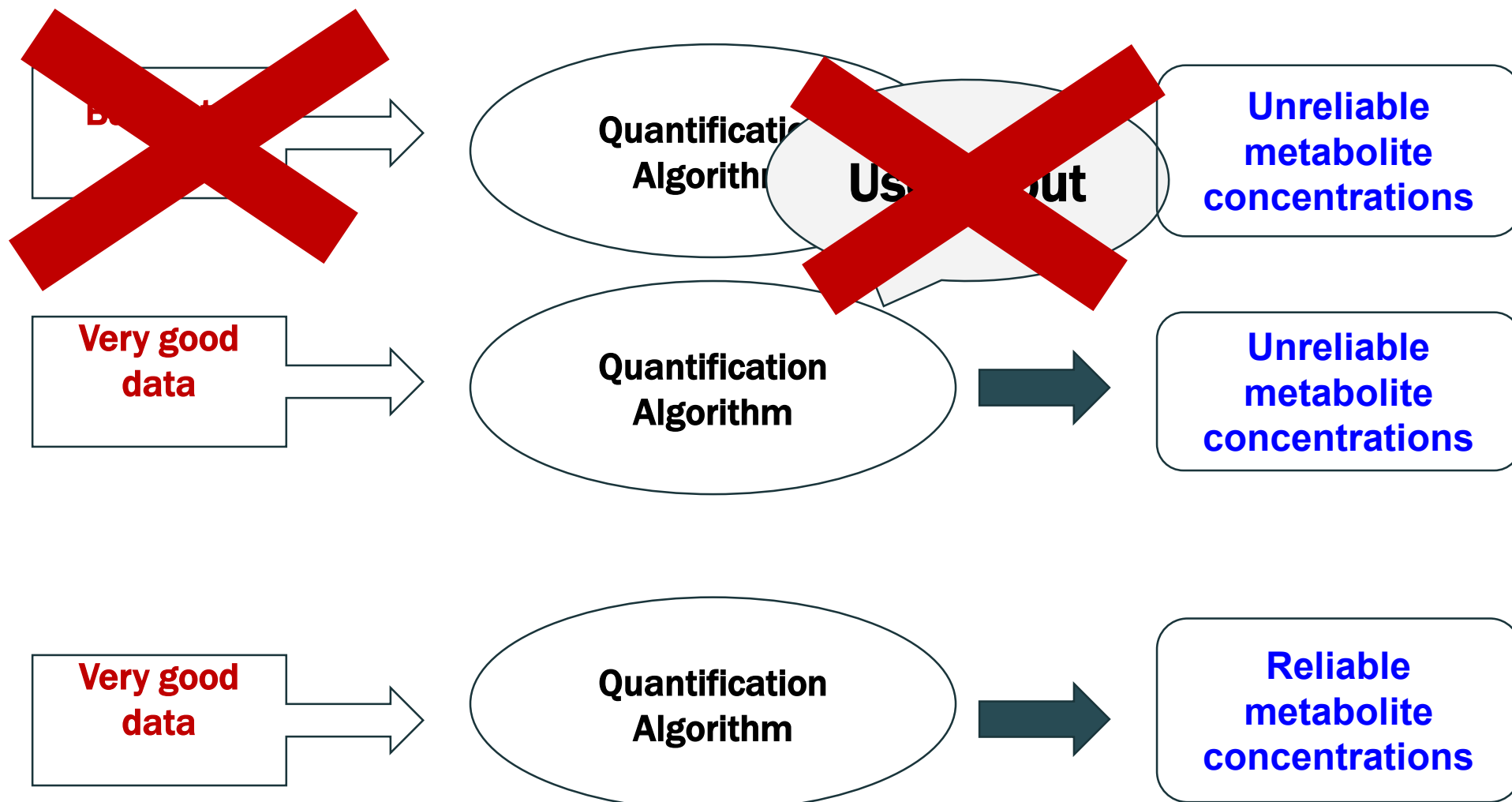
	Cost	Type of data	Preprocessing	Simulations	Lineshape model	MM
Vespa	free	all	yes	yes	yes	yes
TARQUIN	free	all	yes	Yes – basic	Voigt	Baseline –TD Add
jMRUI	free	all	yes	yes	Lorentzian	Add TD
LCModel	Yes	all	yes	No Basis sets	estimated	Splines

Anke Henning. eMagRes, 2016, Vol 5: 981–994. DOI 10.1002/9780470034590.emrstm1472

Dirk van Ormondt, et al, eMagRes, 2015, Vol 4: 651–662. DOI 10.1002/9780470034590.emrstm1427

Jamie Near, Magnetic Resonance Spectroscopy Elsevier 2014

Jamie Near, et al, NMR Biomed, 2020 <https://doi.org/10.1002/nbm.4257>



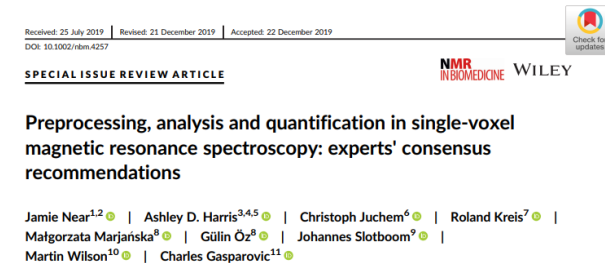
QUANTIFICATION (ABSOLUTE)

Signal amplitudes
Peak area



Concentrations
mmol/kg_{ww}

- External concentration reference
- Internal concentration reference
 - Ratios to tCr or NAA or Cho - They might change – disease
 - Water internal reference
 - Corrections for T1, T2 and water content
 - Very short TE – T2 correction could be neglected
 - Fully relaxed signals – long TR – T1 neglected
 - MT



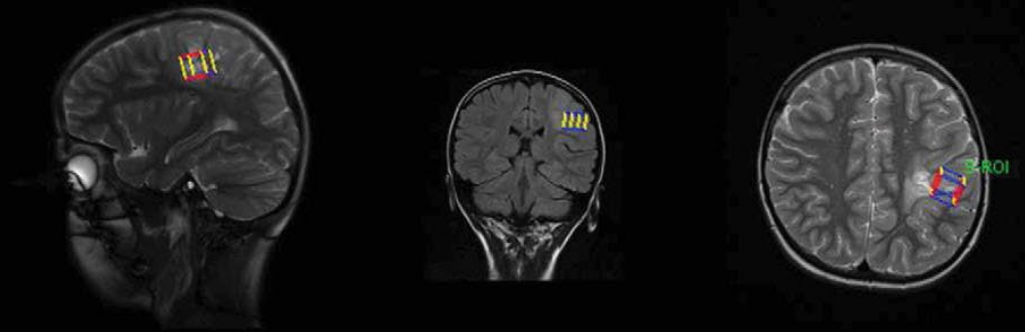
- CRLB – theoretical estimation
 - Assumes that the model is perfect
 - R Kreis, MRM 75, 2016 – CRLB % !!! – low conc metabolites
 - CRLB vs mean CRLB in normal cohort
- Fit residuals

QUANTIFICATION: RESULTS DISPLAY

Patient name : anonymous	Accession number : 2699671	Series description : svs_se_135_ws	Scanner type : Verio
Date of Birth :	Study description : Kopf_12_Kanal	Series number : 12	Echo time TE : 135000
Patient sex : M	Study date : 20110630	Series time : 155149.015000	Repetition time TR : 1500000
Patient ID :			Voxel dimension : [15, 15, 15]

Metabolite Name	Concentrations (2 comp) [mmol/kg ww]	Normal (2 comp) [mmol/kg ww]
Choline (Cho)	8.8 ± 0.1	1.5 ± 0.1
Creatine (Cr)	7.7 ± 0.2	5.2 ± 0.1
Glutamate (Glu)	6.6 ± 0.4	3.7 ± 0.2
Glutamine (Gln)	4.4 ± 0.3	2.2 ± 0.1
Lactate (Lac)	1.2 ± 0.2	0.3 ± 0.1
Myo-Inositol (m-Ins)	20.7 ± 1.2	7.8 ± 0.4
N-Acetyl Aspartate (NAA)	4.9 ± 0.2	7.5 ± 0.2

For scientific purposes only!
Report created with jMRUI.



b

FURTHER READING

SOFTWARE & CODE

How to access code

Contribute your code

SOFTWARE PACKAGES

COMPLETE LIST

Analysis & Quantification
Data Simulation & Basis Set
Generation
Deidentification
Input/Output
Processing
Reconstruction
Reproducible Workflows
Visualization

Software & Code

How to access code

You can browse the various software packages by clicking on a topic in the column to the left. Each topic takes you to a list of associated software packages.

Links in each package entry will take you to either an external web site (typical for applications that already have a website) or to various GitHub repositories that 'live' on the MRSHub.

You don't have to be familiar with GitHub to download code from this site. Three quick clicks and you can have a ZIP file downloaded to your computer:

SUMMARY & ACKNOWLEDGMENTS

MRS & MRSI is incredibly rich & versatile 😊

Thank you for listening! Questions?

For any question you might have later on, please write me an email:

cristina.cudalbu@epfl.ch



THANK YOU FOR YOUR ATTENTION



C I B M . C H