

# Course content

## Topics (lectures):

1. Introduction (1)
2. **Structure** (2-5)
3. **Single molecule mechanics** (6-9)
4. **Collective/emergent properties** (9-11)
5. **Student presentations** (11-13)

## Course structure:

1. Introduction to topic
2. Awardees (1-2 per week)
  - History, first-person, second-person accounts (C)
  - Article, analysis of scientific work (E)
3. Discussion of topic, outlook

# Lecture 6: Introduction to single molecule mechanics

Today's goal: Force spectroscopy, measurements & theory, history

- Force-extension curves
- Random walk models

PBOC Chapter 8.3

- History of nanocharacterization
- STM: Nobel 1986  
<https://www.nobelprize.org/prizes/physics/1986/press-release/>
- STM: the world's smallest movie  
<https://www.youtube.com/watch?v=oSCX78-8-q0>
- AFM
- Arthur Ashkin, optical tweezers (16:00-25:39)  
<https://www.nobelprize.org/prizes/physics/2018/ashkin/lecture/>
- Taekjip Ha, helicases via FRET + optical trapping  
<https://www.ibiology.org/biophysics/single-molecule-technologies/#part-3>

*“The operative industry of Nature is so prolific that machines will be eventually found not only unknown to us but also unimaginable by our mind.”*

Marcello Malpighi, *De Viscerum Structura*, 1666

# Models of the cell

Fig. 1.

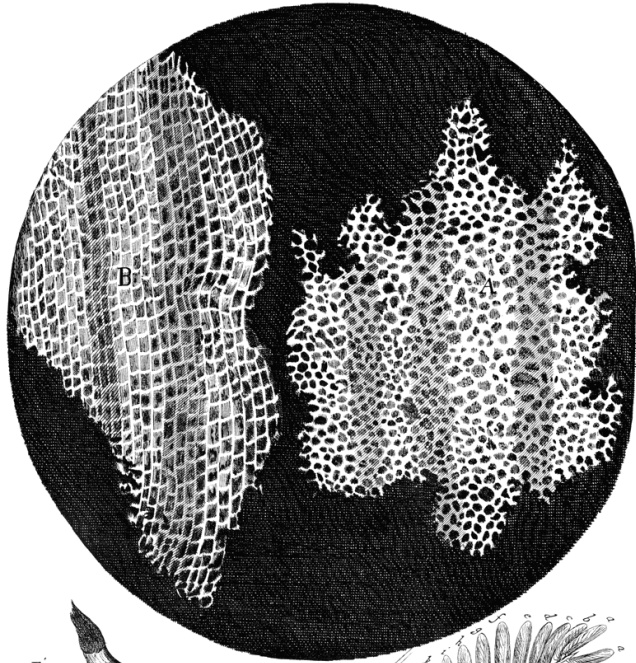
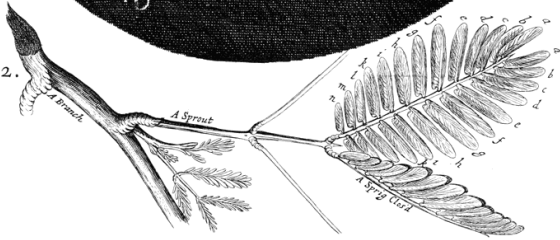


Fig. 2.



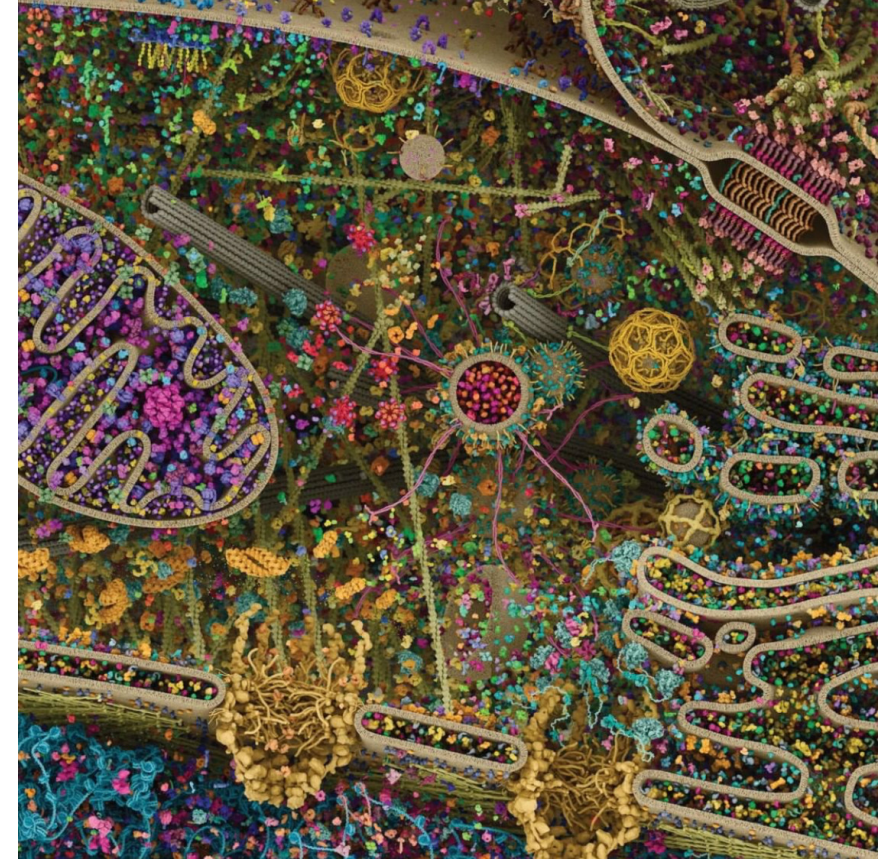
“[...] great many little Boxes.”

Robert Hooke  
Micrographia, 1665



Small bags containing a concentrated solution of macromolecules undergoing second-order reactions.

magnetic resonance and cryo-electron microscopy data



[...] a factory that contains an elaborate network of interlocking assembly lines, each of which is composed of a set of large protein machines.

Bruce Alberts, The Cell as a Collection of Protein Machines: Preparing the Next Generation of Molecular Biologists, *Cell* **92** (1998)

# Why study single molecules?

*“There are two breads.  
You eat both, I eat none.  
Average consumption:  
One bread per person.”*

Nicanor Parra

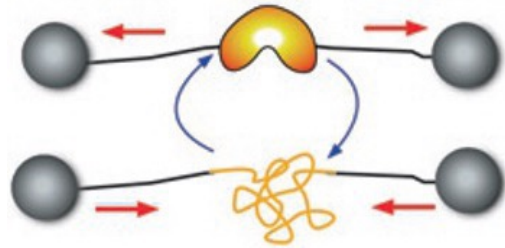
Averaging events in ensemble experiments hides the true dynamics of the actors involved.

The “molecular trajectories” recorded by single molecule experiments are more readily interpretable in the mechanistic framework.

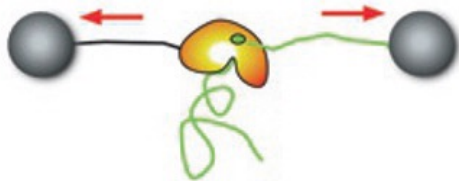
By exerting forces and measuring displacements of single molecule, it is possible to monitor simultaneously the dynamics and the energetic landscape of a system.

# Why study single molecules?

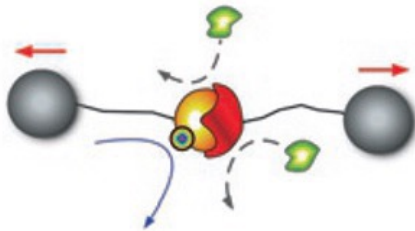
Measure forces exerted by:



protein folding  
unfolding



DNA binding  
proteins



receptor-ligand  
binding



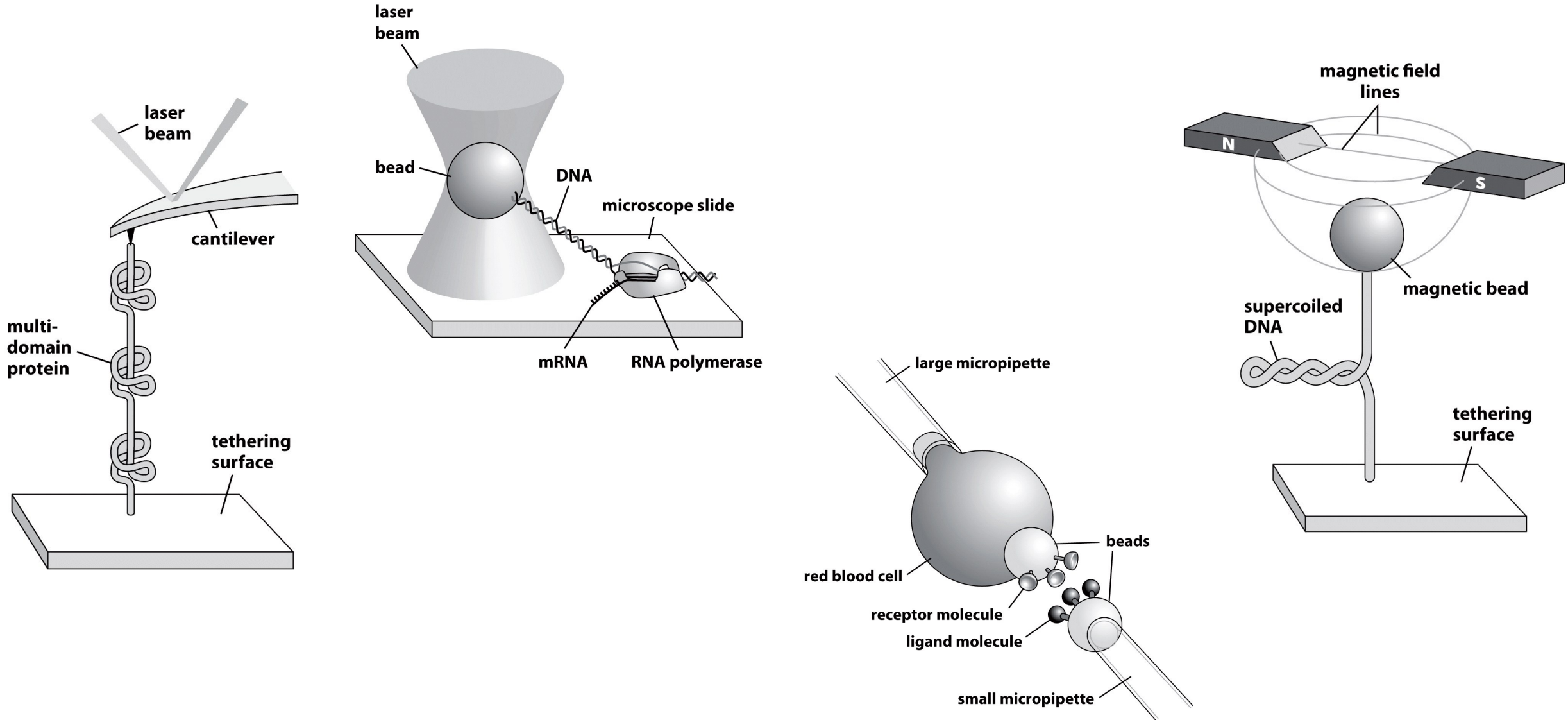
motor proteins

## Examples:

Packing DNA into a virus  
Moving cargo along cytoskeleton  
DNA replication, transcription



# How to study single molecules?



# How to study single molecules?

**NanoScope®** Scanning Probe Microscopes  
We Have Science Covered

**And Price Too!**  
\$35,000\* For A Complete NanoScope II AFM System, Including Computer, Control Electronics, Contact AFM and 12µm Scanner!



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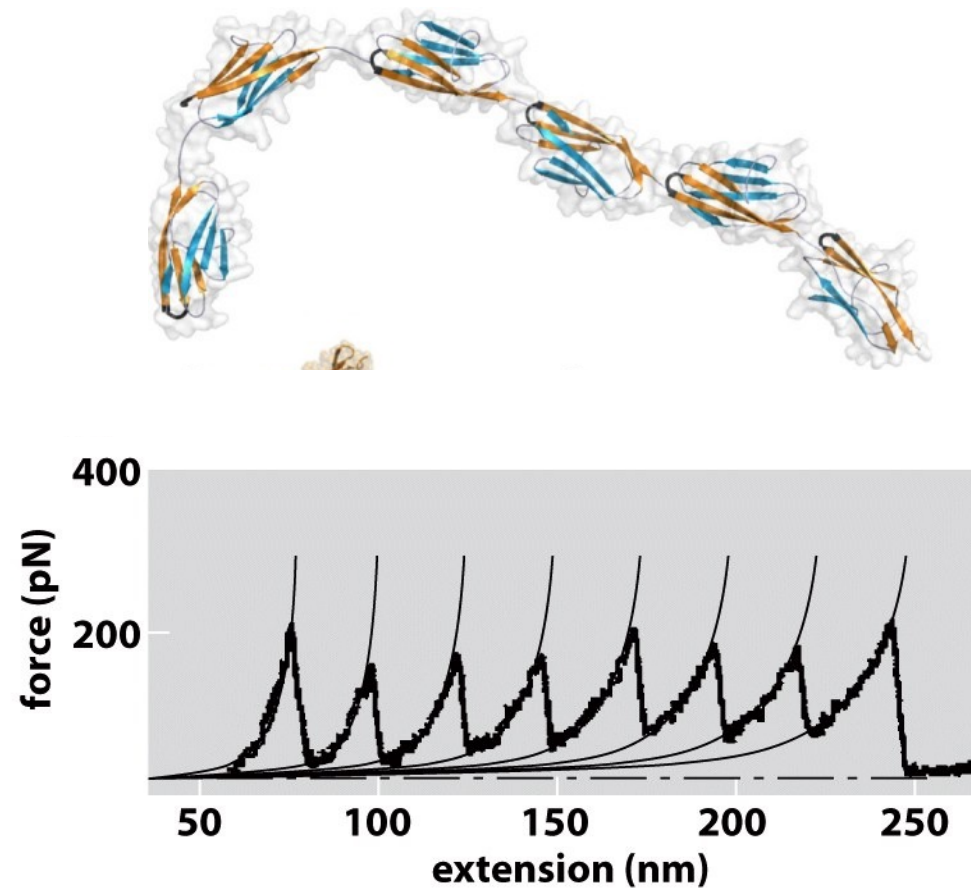
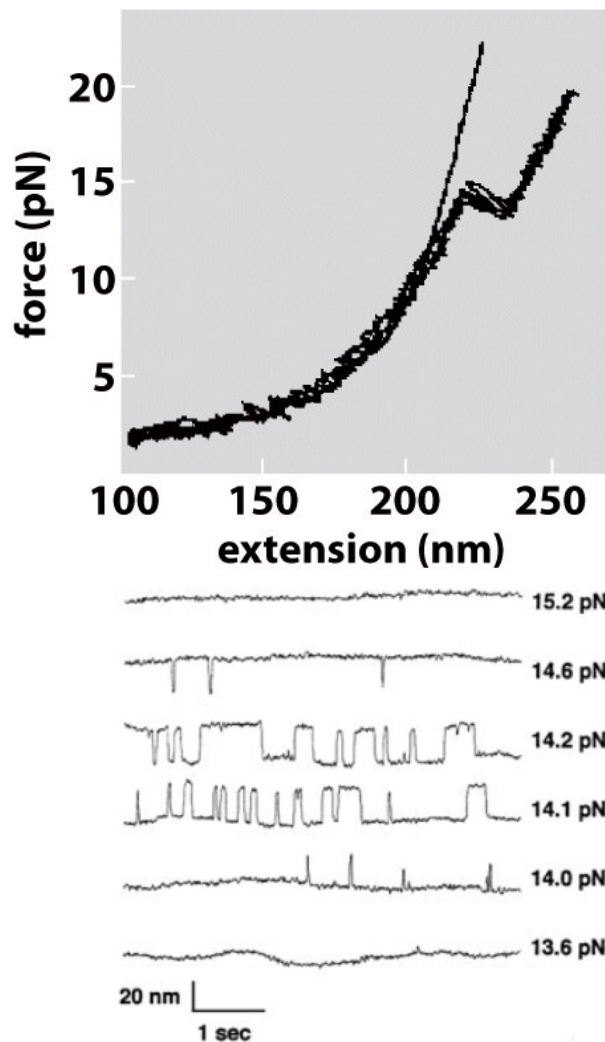
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\*U.S. Domestic Price Only.  
Not to be construed as an endorsement by the journal SCIENCE.

	Optical tweezers	Magnetic (electromagnetic) tweezers	AFM
Spatial resolution (nm)	0.1–2	5–10 (2–10)	0.5–1
Temporal resolution (s)	10 <sup>-4</sup>	10 <sup>-1</sup> –10 <sup>-2</sup> (10 <sup>-4</sup> )	10 <sup>-3</sup>
Stiffness (pN nm <sup>-1</sup> )	0.005–1	10 <sup>-3</sup> –10 <sup>-6</sup> (10 <sup>-4</sup> )	10–10 <sup>5</sup>
Force range (pN)	0.1–100	10 <sup>-3</sup> –10 <sup>2</sup> (0.01–10 <sup>4</sup> )	10–10 <sup>4</sup>
Displacement range (nm)	0.1–10 <sup>5</sup>	5–10 <sup>4</sup> (5–10 <sup>5</sup> )	0.5–10 <sup>4</sup>
Probe size (µm)	0.25–5	0.5–5	100–250
Typical applications	3D manipulation	Tethered assay	High-force pulling and interaction assays
	Tethered assay	DNA topology	
	Interaction assay	(3D manipulation)	
Features	Low-noise and low-drift dumbbell geometry	Force clamp	High-resolution imaging
		Bead rotation	
		Specific interactions	
Limitations	Photodamage	No manipulation	Large high-stiffness probe
	Sample heating	(Force hysteresis)	Large minimal force Nonspecific
	Nonspecific		



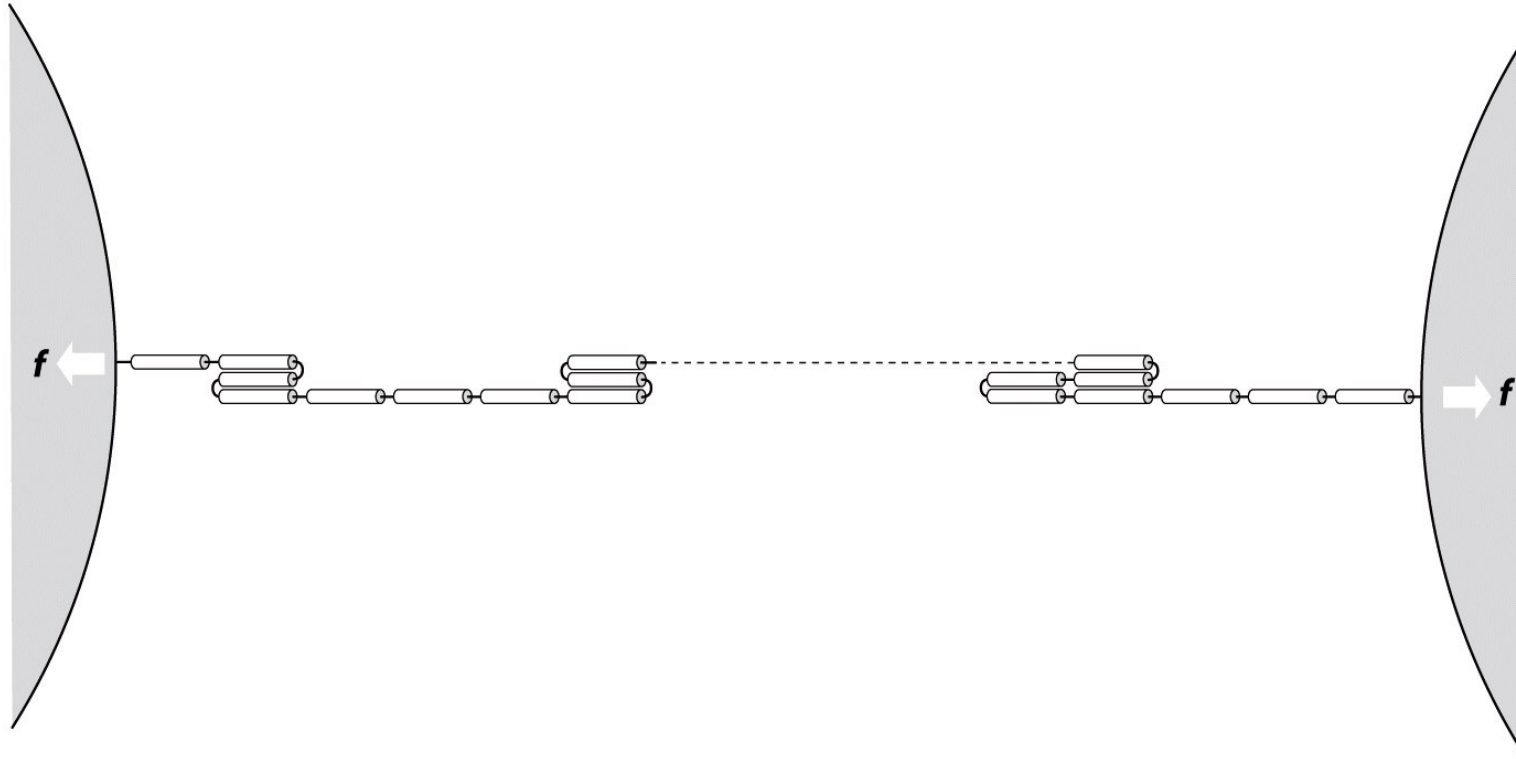
## Experiments:

titin (human cardiac protein)



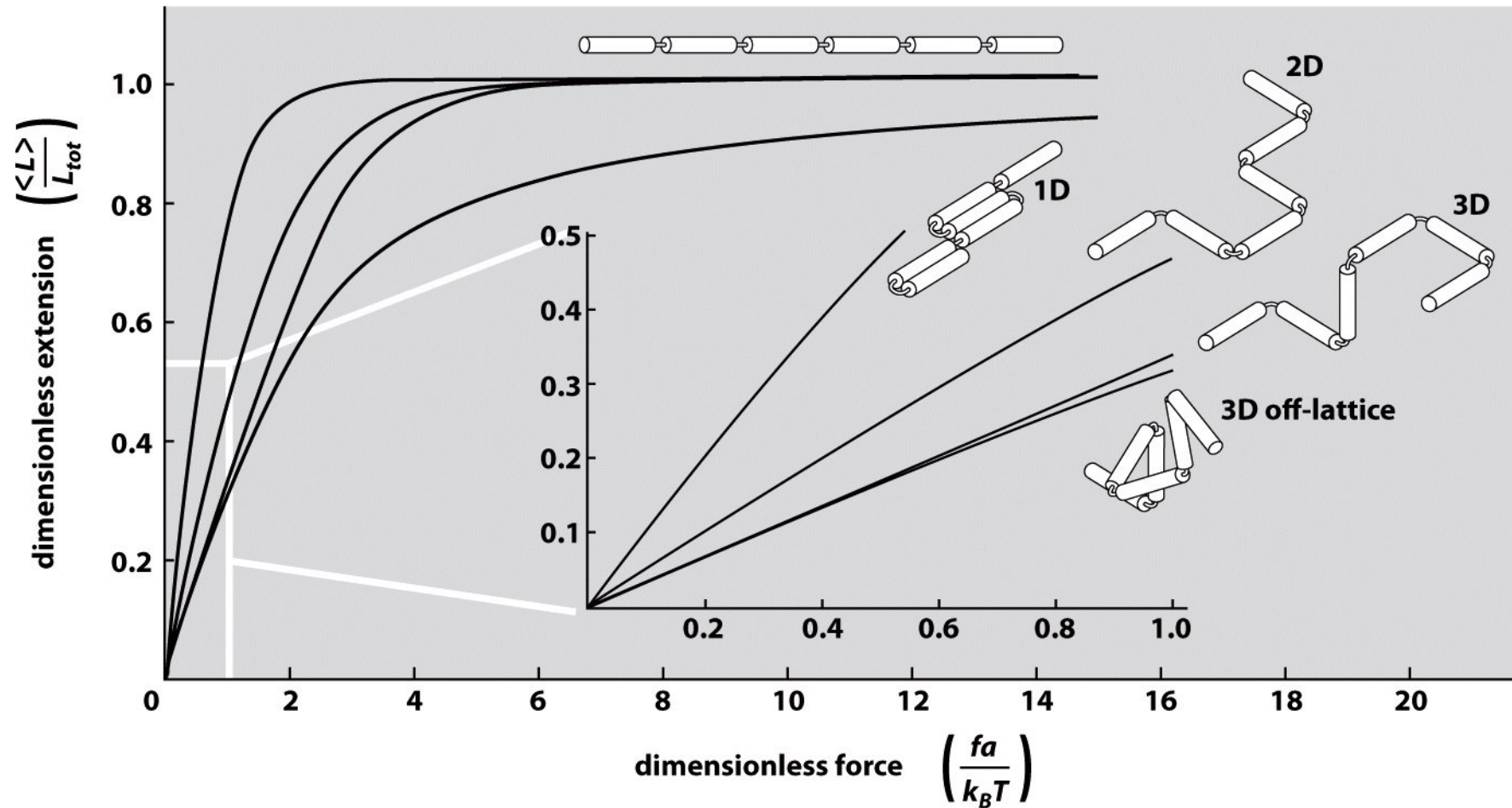
# Force-extension curves

Models:



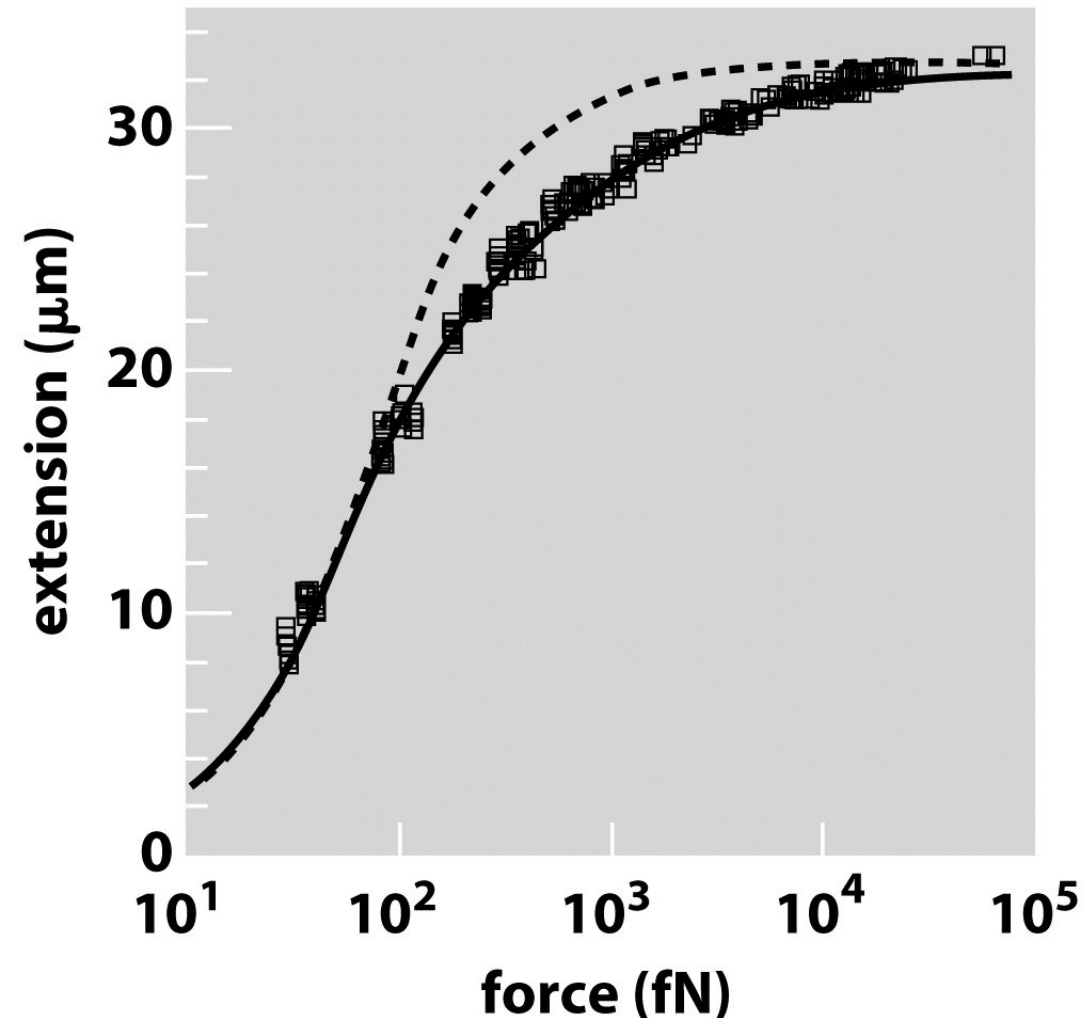
# Force-extension curves

Models:

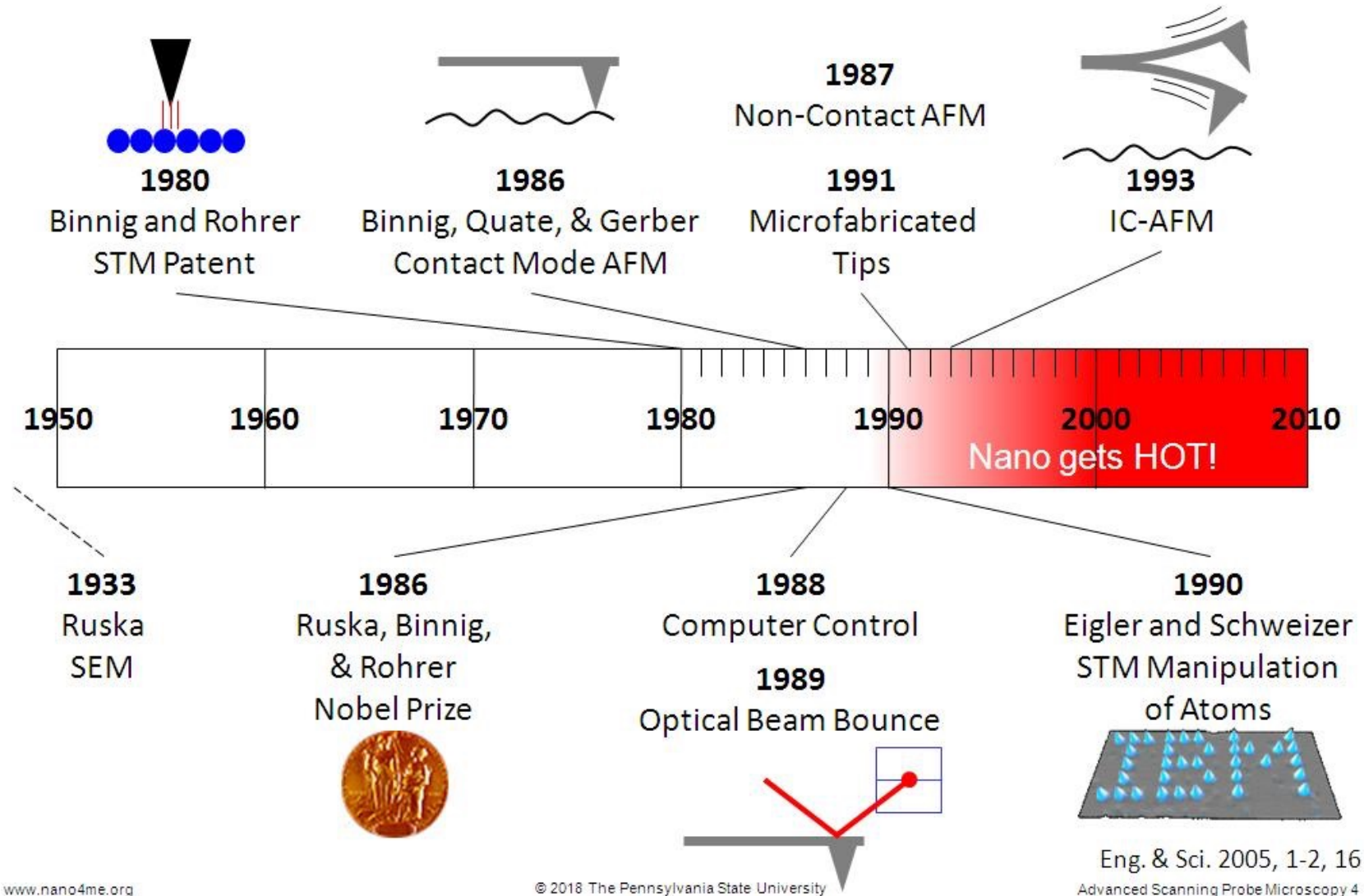


# Force-extension curves

Models and measurements:

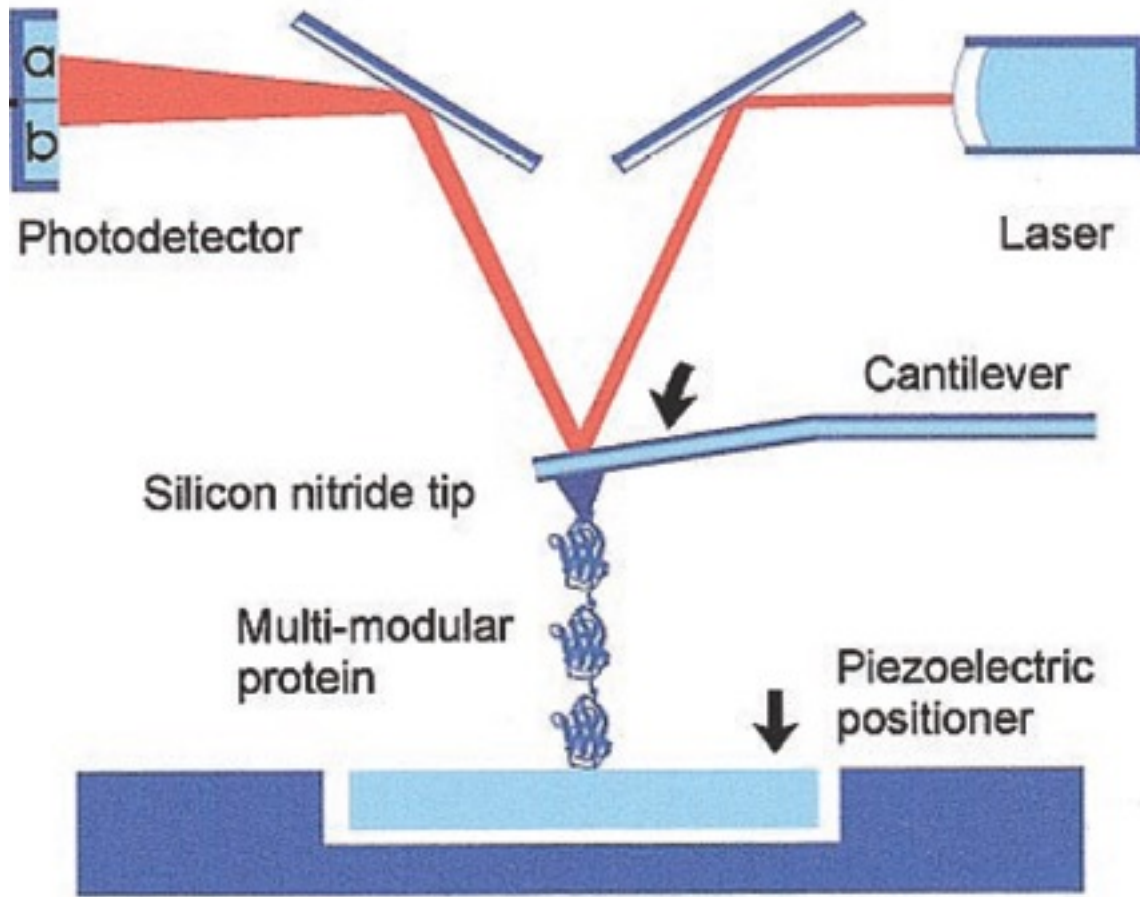


# History of nanocharacterization





# Atomic force microscope



- AFM cantilever probes by moving its tip along surface, or pulling on a protein
- Cantilever movement is detected with a focused laser beam that refracts into a photodetector
- The deflection of the cantilever deflects the laser correspondingly and can map the surface
- In single molecule force spectroscopy, the cantilever is pressed against a layer of proteins attached to a substrate, the tip adsorbs a single protein molecule, which is then extended.
- Extension of the molecule by retraction of the piezoelectric positioner results in deflection of the cantilever.

# Lectures 7, 8

- Paul Hansma: Development of AFMs to monitor individual protein molecules, in liquids (1990-2000)

*"For pioneering contributions to the development of biological scanning probe microscopy and for the molecular resolution imaging of biological molecules in aqueous solutions."* (2000)

- Carlos Bustamante: Study of DNA, RNA, and protein molecular mechanics (1990-2000)

*"For his pioneering work in single molecule biophysics and the elucidation of the fundamental physics principles underlying the mechanical properties and forces involved in DNA replication and transcription."* (2002)

