

## Biomicroscopy II

Transmission  
Techniques

## Transmission

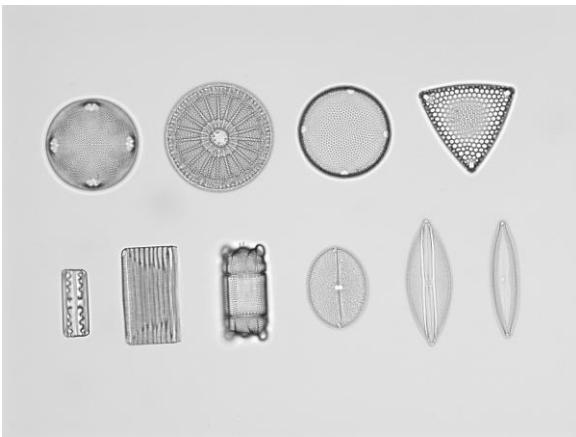
- Focus on the specimen
- Close field diaphragm
- Focus condenser until field diaphragm is seen sharp
- Center field diaphragm
- Open/close field diaphragm up to 80 –90 %
- Remove eyepiece, look down to the aperture diaphragm
- Center (if possible) aperture diaphragm
- Open aperture diaphragm up to 60 –70 %
- Start with low magnification objective. Repeat for every objective used

## Fluorescence

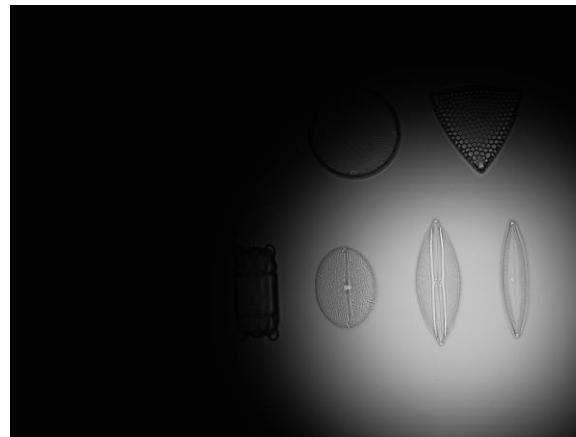
- Focus on the specimen
- Swing in focusing aid (if available)
- Focus image of arc sharply
- Swing out focusing aid
- Close field diaphragm
- Center field diaphragm

# Adjustment for Koehler illumination

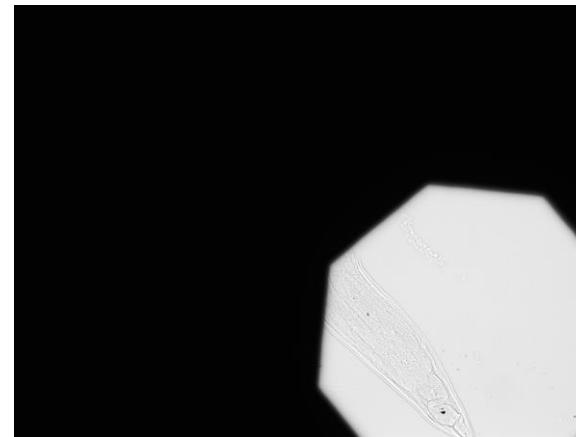
1. Focus on the sample



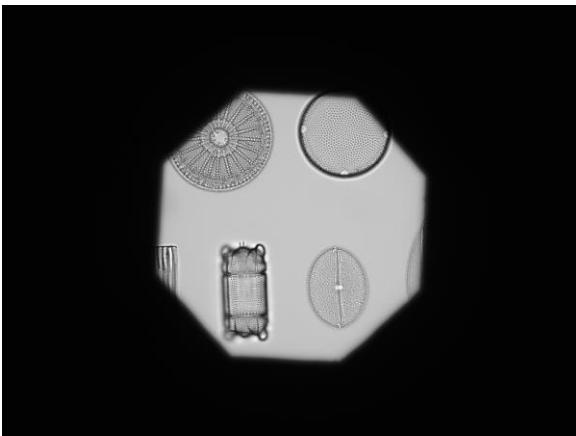
2. Close the field diaphragm



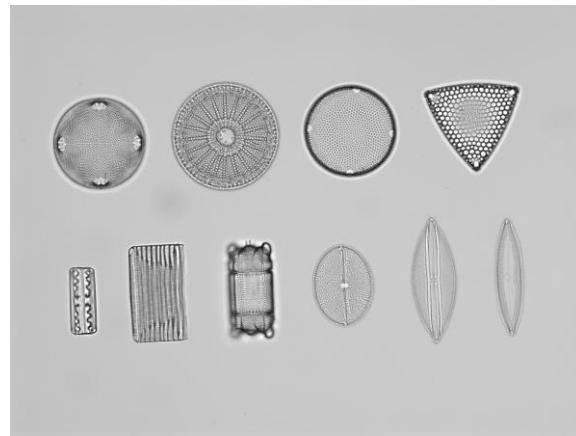
3. Set the condenser height in order to obtain a sharp spot



4. Center the spot with condenser centering screws



5. Open the field diaphragm



6. Remove eyepiece and set aperture diaphragme



**Dark field:** Fine structural features at, and even below, the resolution limit of a light microscope. Highly suitable for metallographic and crystallographic examinations with reflected light.

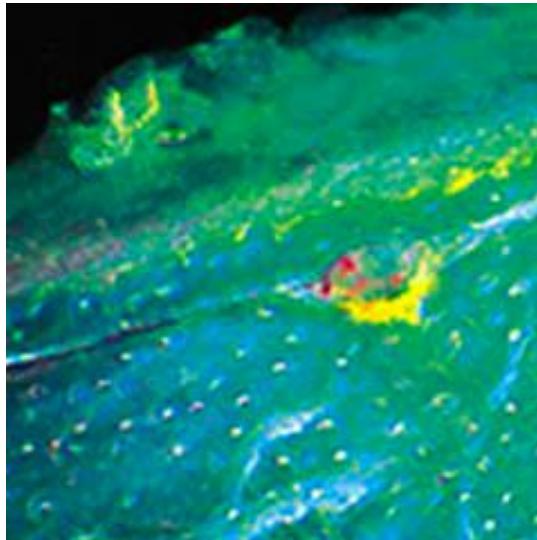
**Phase contrast:** Used for visualizing very fine structural features in tissues and single cells contained in very thin ( $< 5 \mu\text{m}$ ), non-stained specimens.

**DIC:** Method shows optical path differences in the specimen in a relief-like fashion. The method is excellently suited for thick, non-stained specimens ( $> 5 \mu\text{m}$ ). Can be used for optical sectioning.

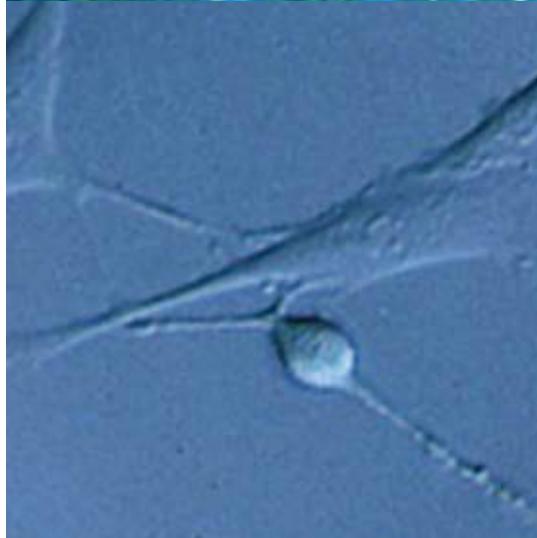
**PlasDIC:** The same specimen as conventional DIC but in plastic dishes.

# Examples of contrasting methods

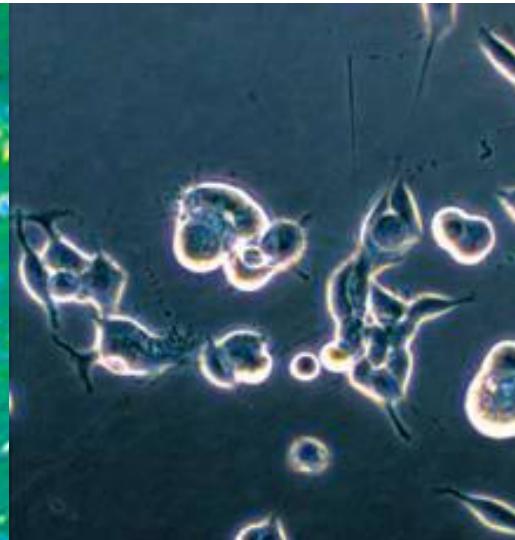
Dark field:  
Bone thin  
section



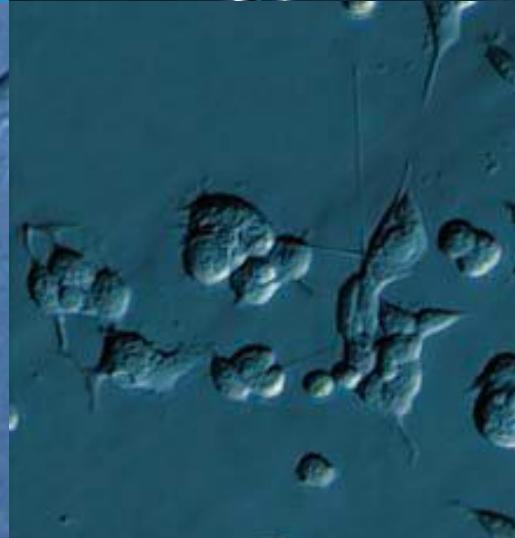
DIC:  
Neurons



Phase contrast:  
HEK cells



PlasDIC:  
HEK cells



# Adjustment for DIC for an inverted microscope

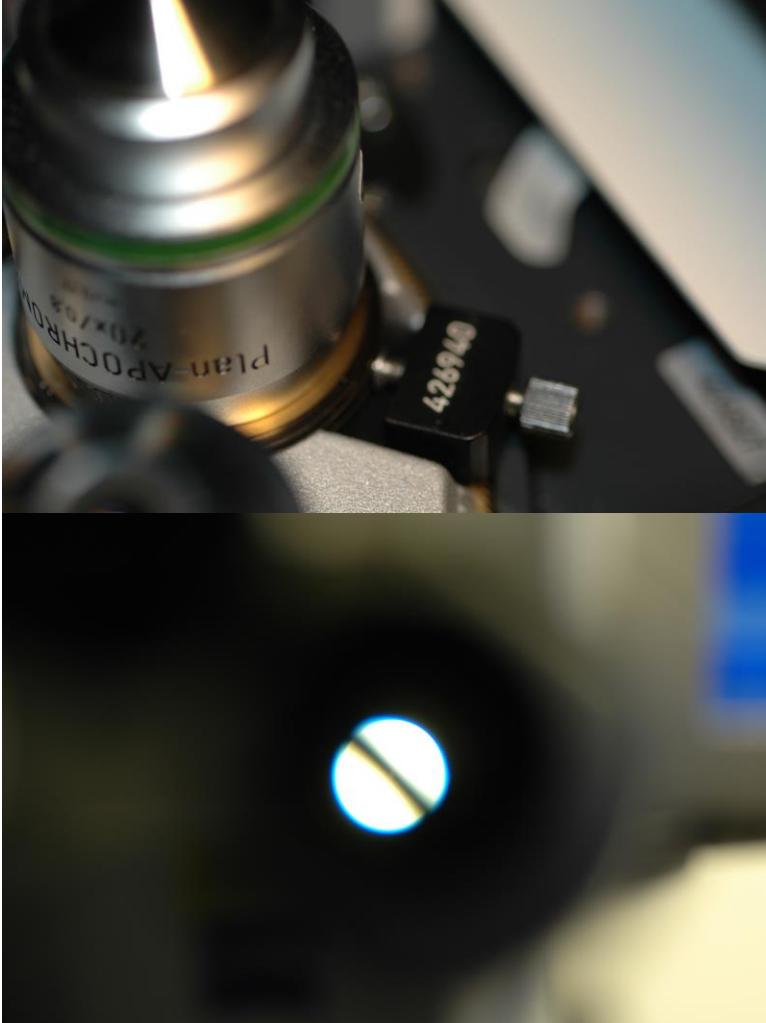
- Set up Koehler illumination in transmission
- Move specimen out of the light path
- Insert polarizer, analyzer and lower Wollaston prism
- Shift Wollaston prism to place dark line in the center
- Turn polarizer until the line is seen mostly dark
- Insert eyepiece back into eyetube
- Insert upper Wollaston prism (in condenser)
- Move the specimen back into the light path
- Move lower Wollaston prism to get required contrast
- Options (if available)
- Rotate the stage to highlight desired area in the sample
- Insert the lambda plate if color staining is required
- Repeat procedure for each objective being used

# Adjustment for DIC



- 1 Lower prism
- 2 Polariser
- 3 Upper prism
- 4 Analyser

# Adjustment for DIC



Insert polarizer, analyzer and lower Wollaston prism

Shift Wollaston prism to place dark line in the center

Turn polarizer until the line is seen mostly dark