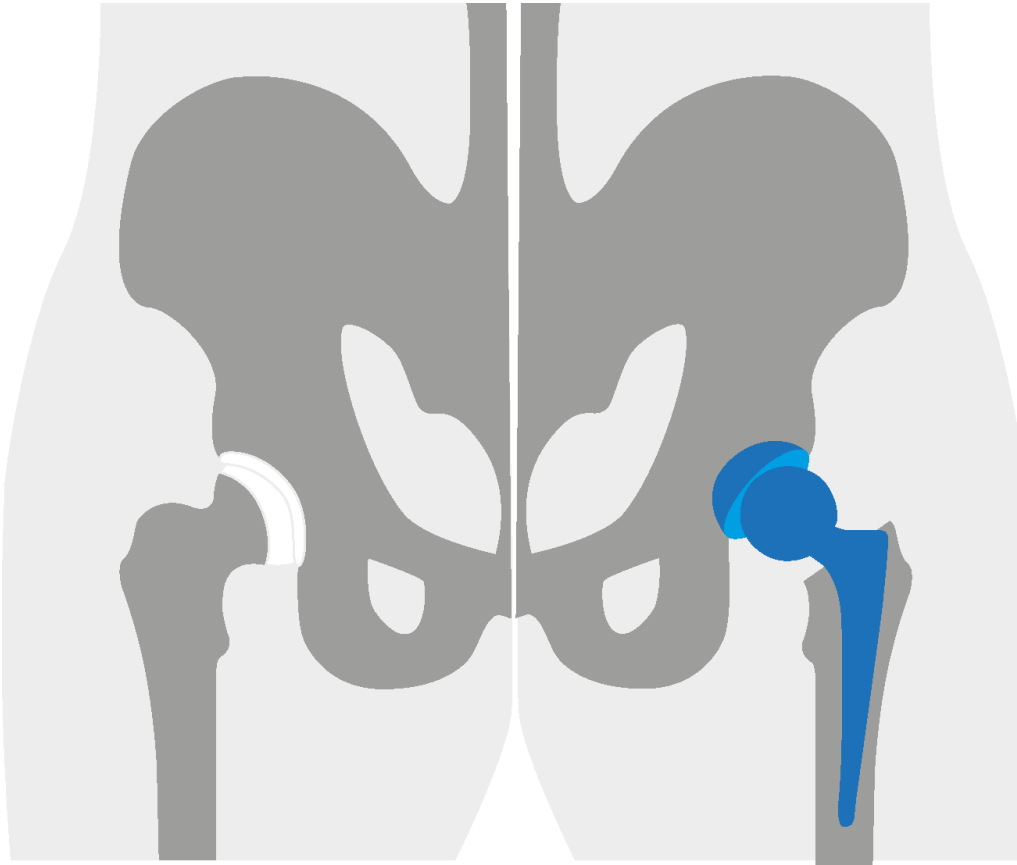


Numerical Methods in Biomechanics

Alexandre Terrier, PhD

EPFL - Laboratory of Biomechanical Orthopedics

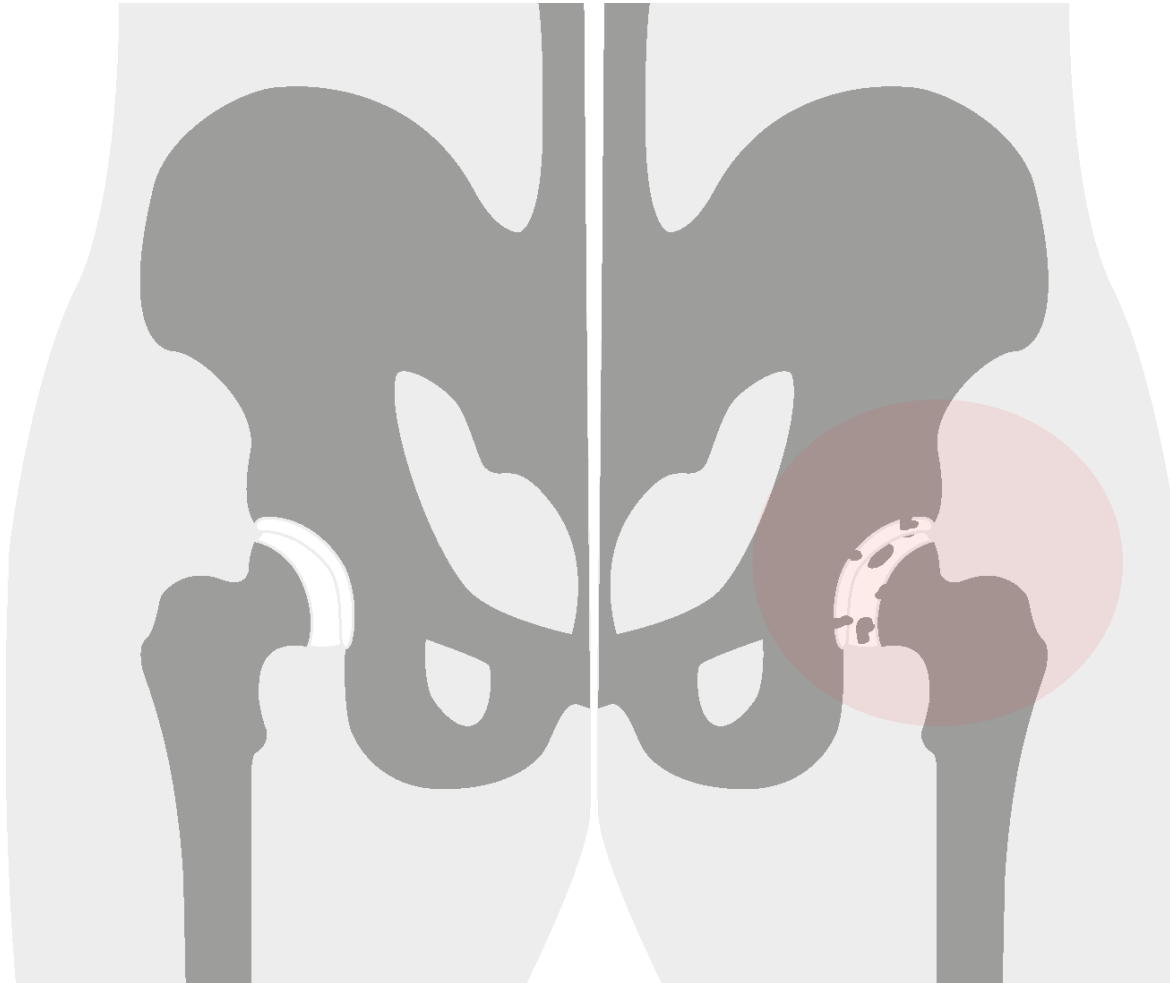


Numerical modeling
to investigate
aseptic loosening of
hip implants

Valérie Malfroy Camine

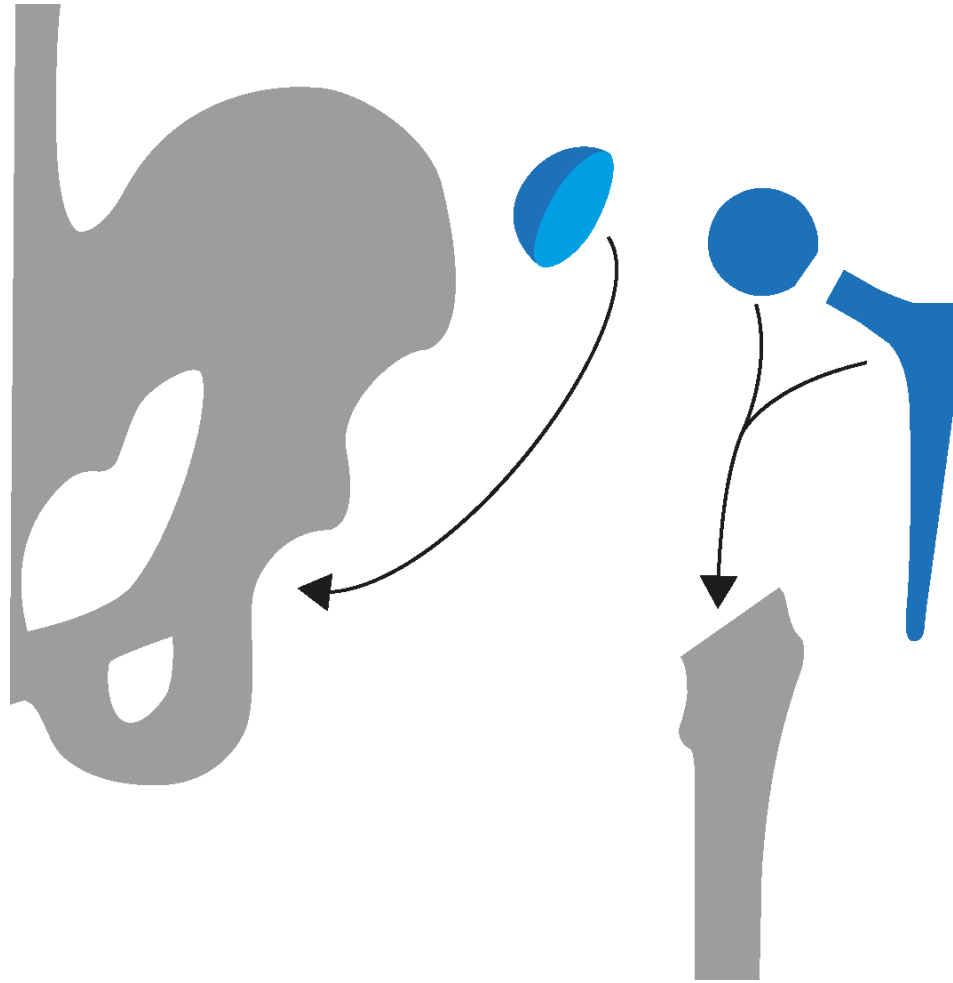


Osteoarthritis





Total Hip Replacement



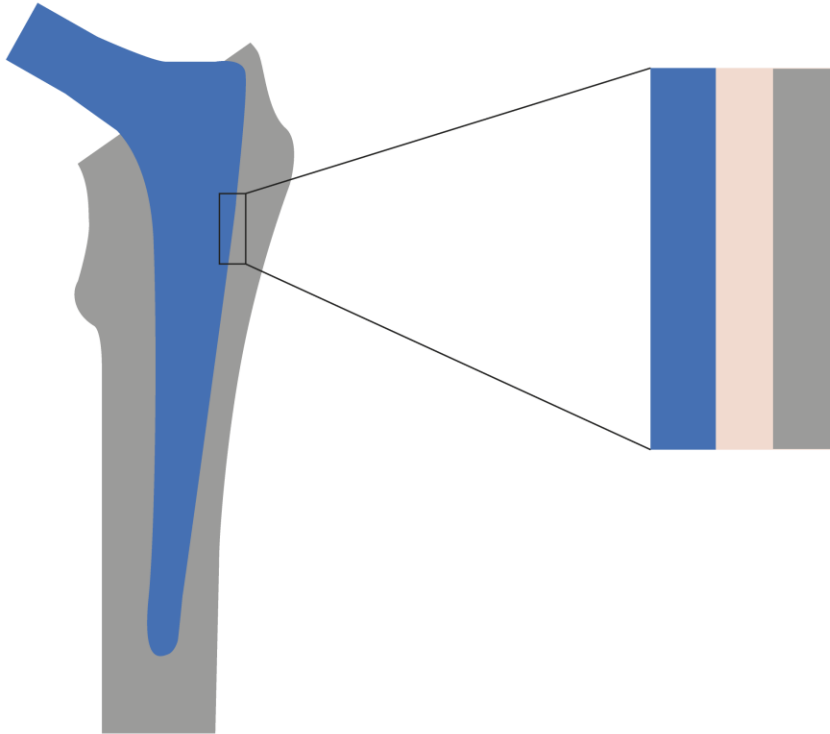


Aseptic Loosening



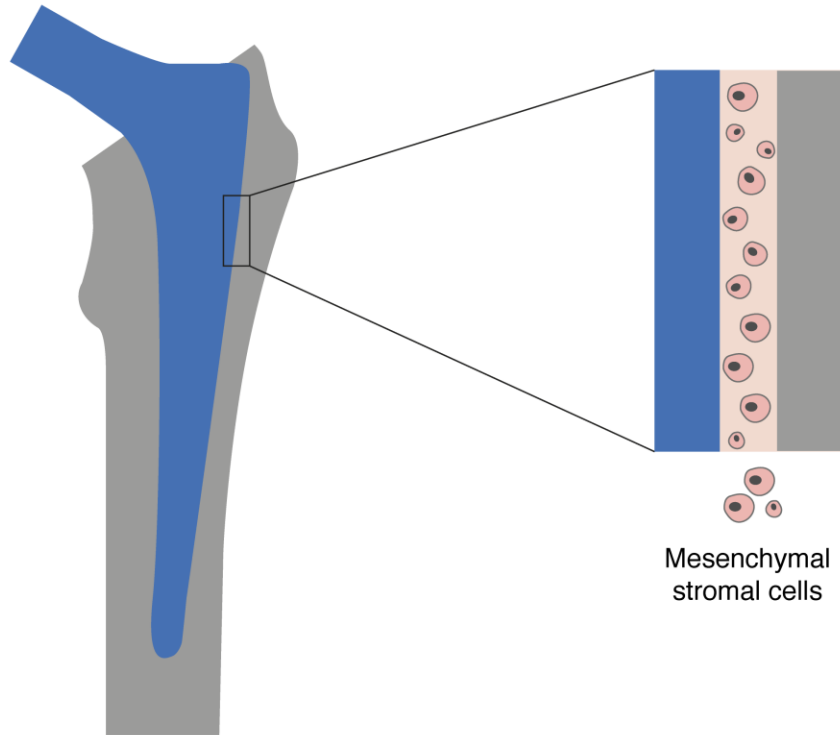


Peri-implant Healing



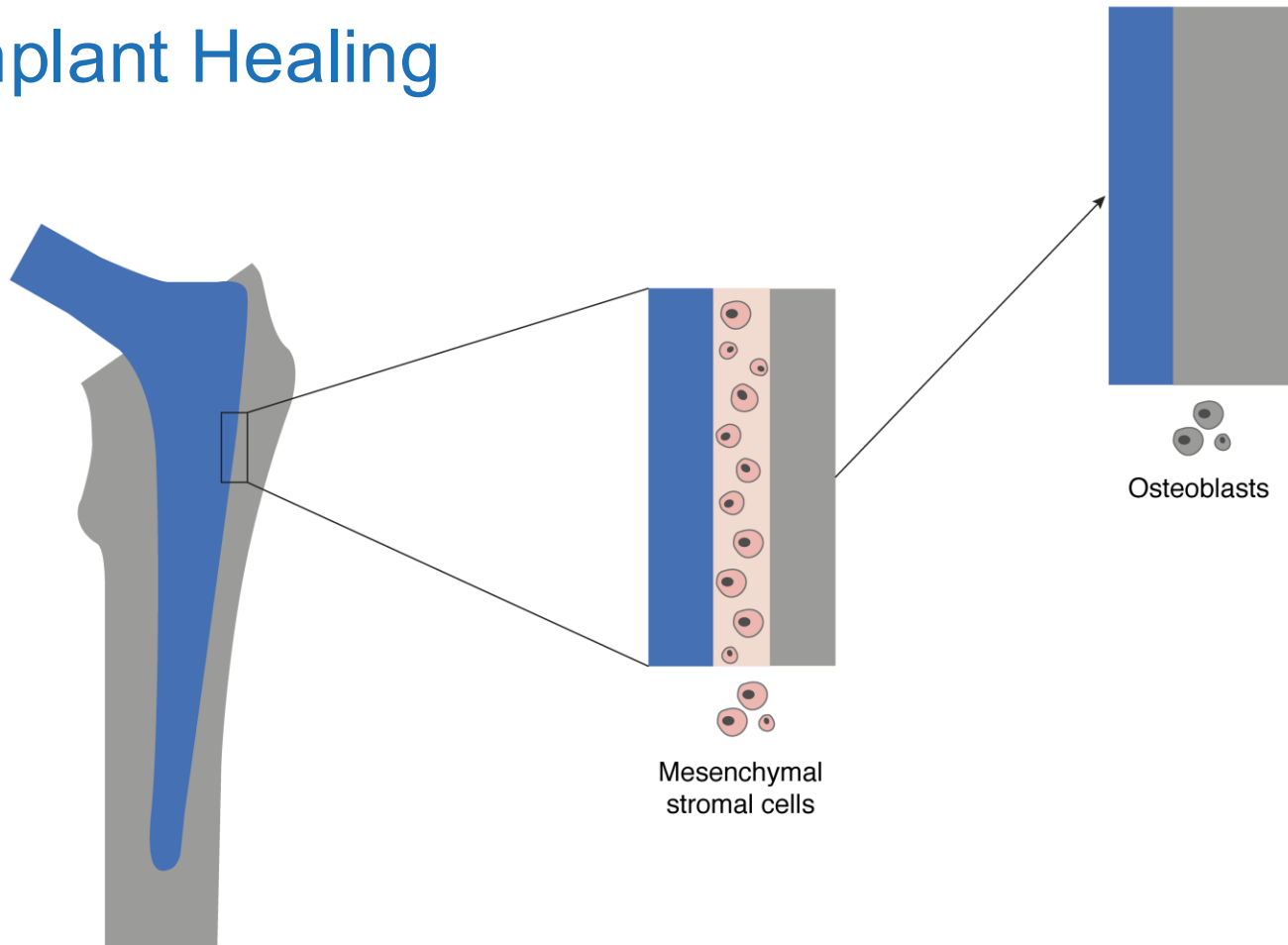


Peri-implant Healing



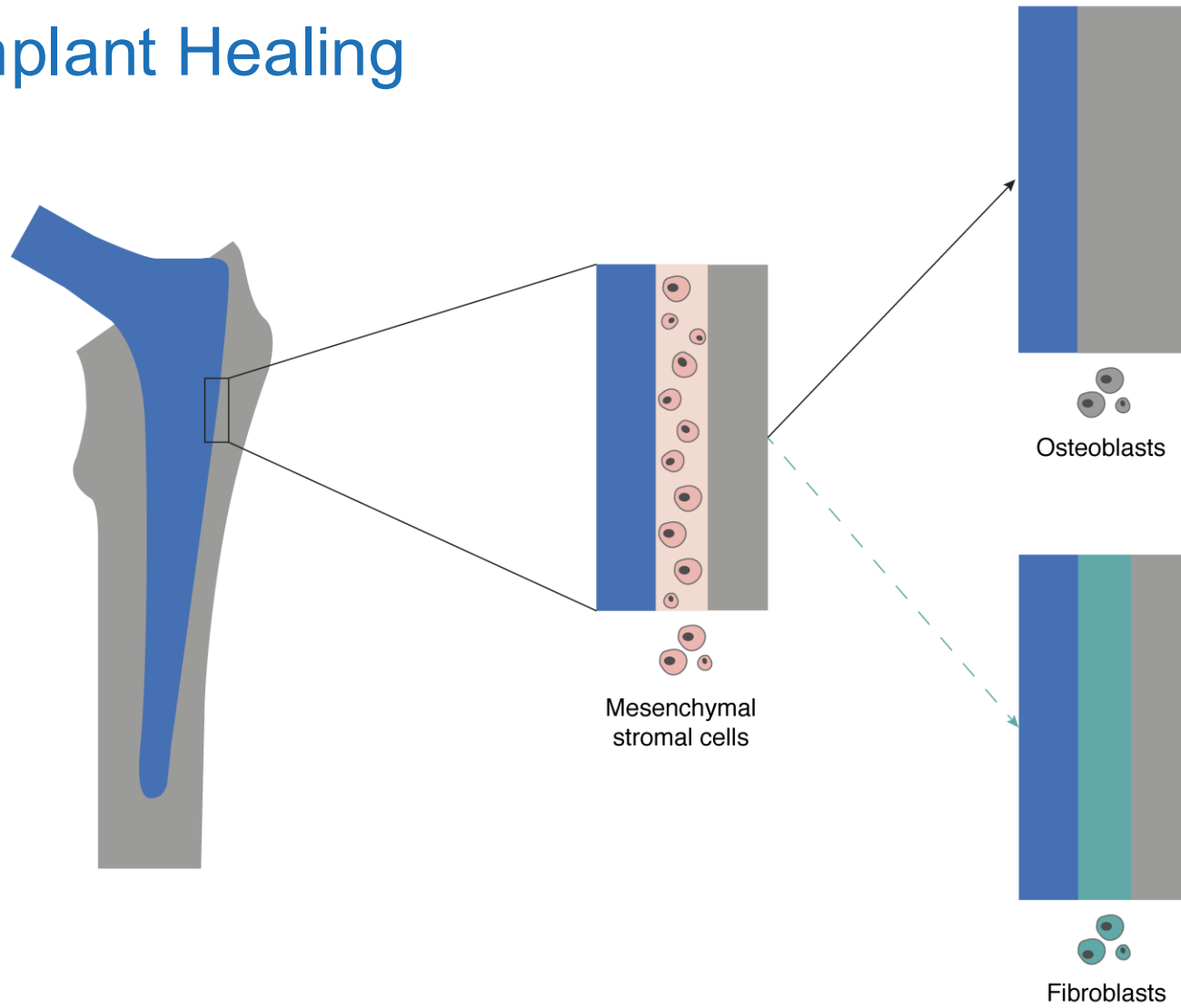


Peri-implant Healing



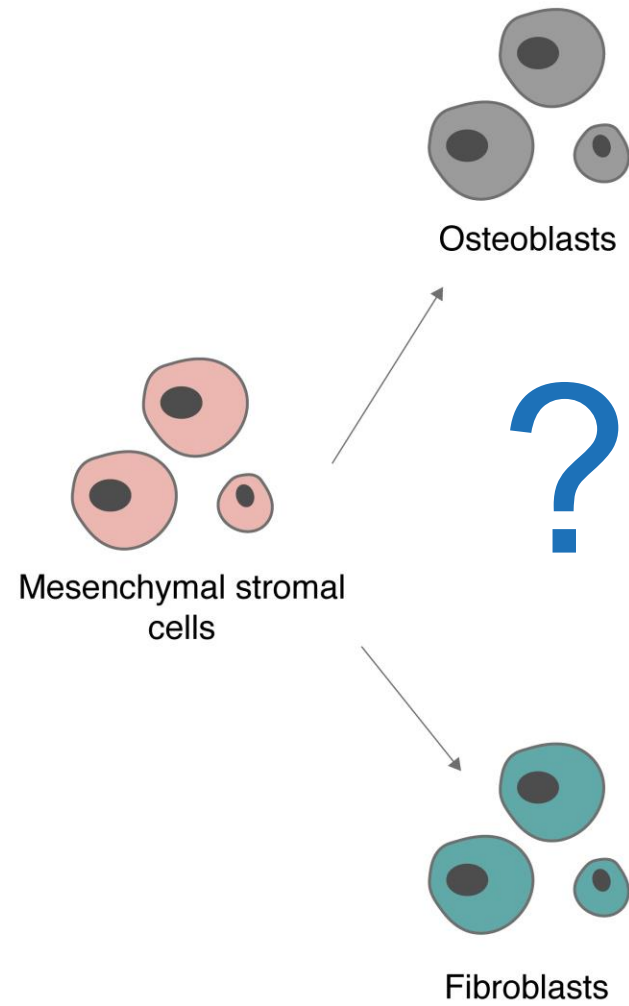


Peri-implant Healing



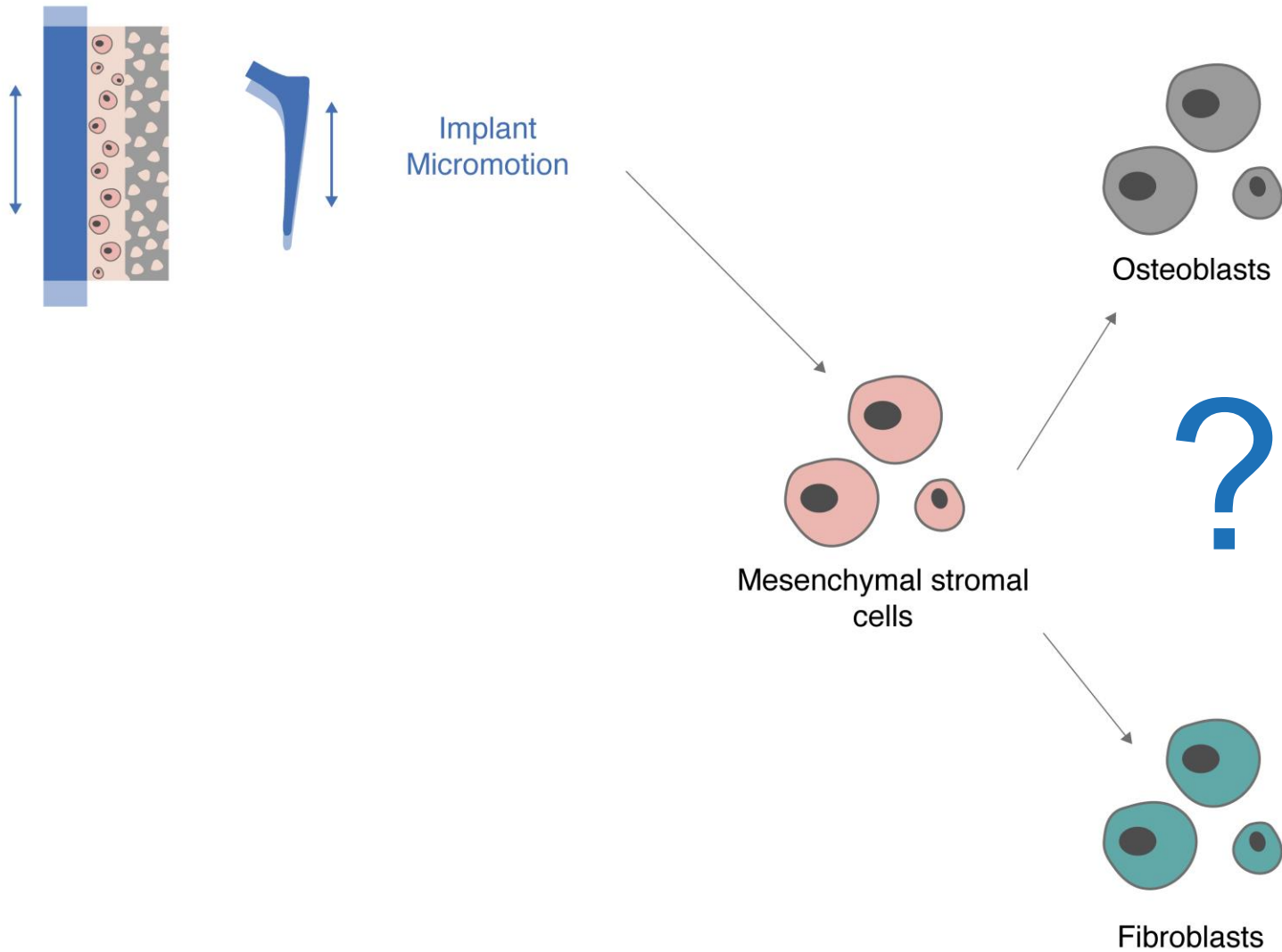


MSCs Differentiation



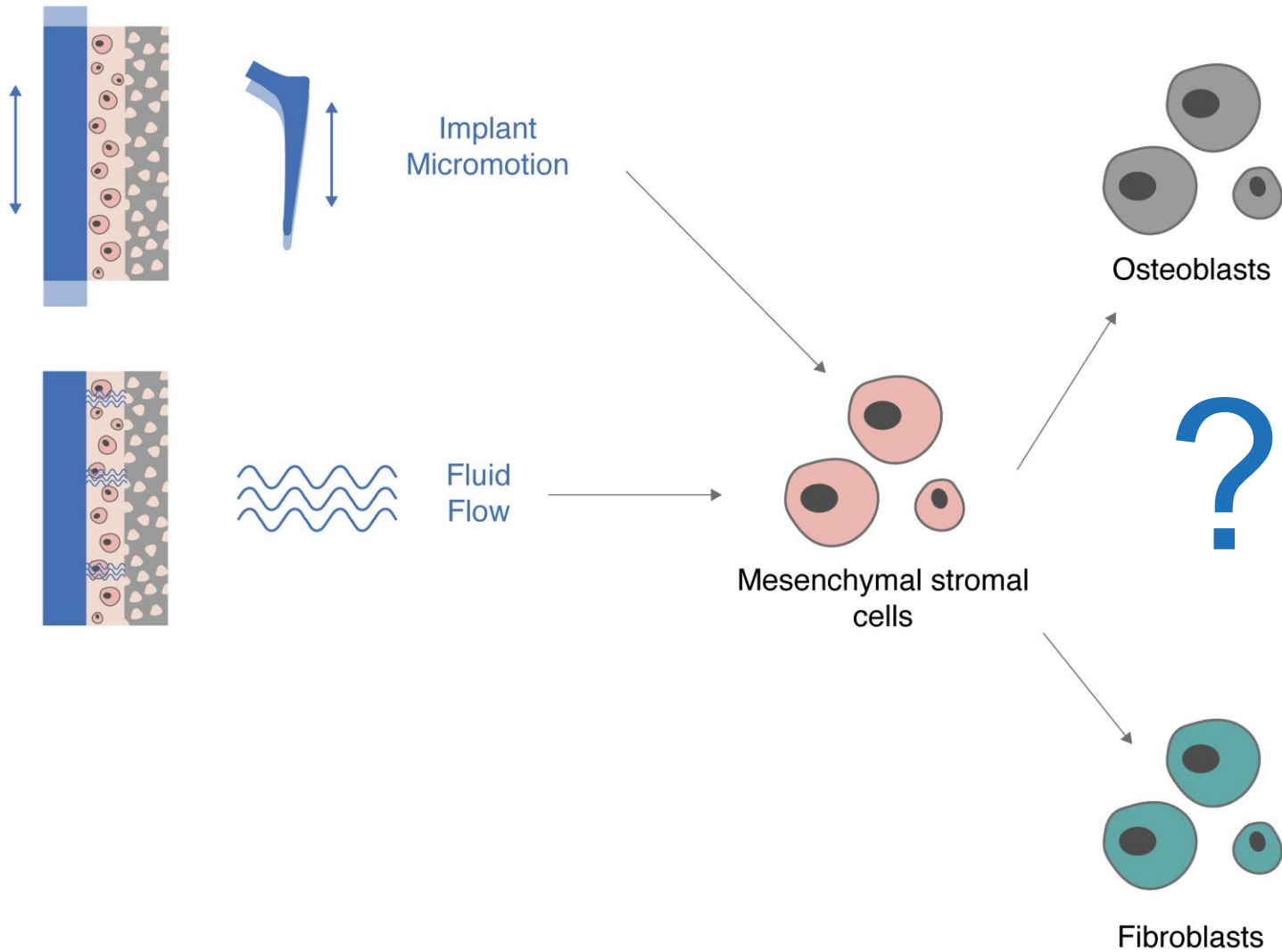


MSCs Differentiation



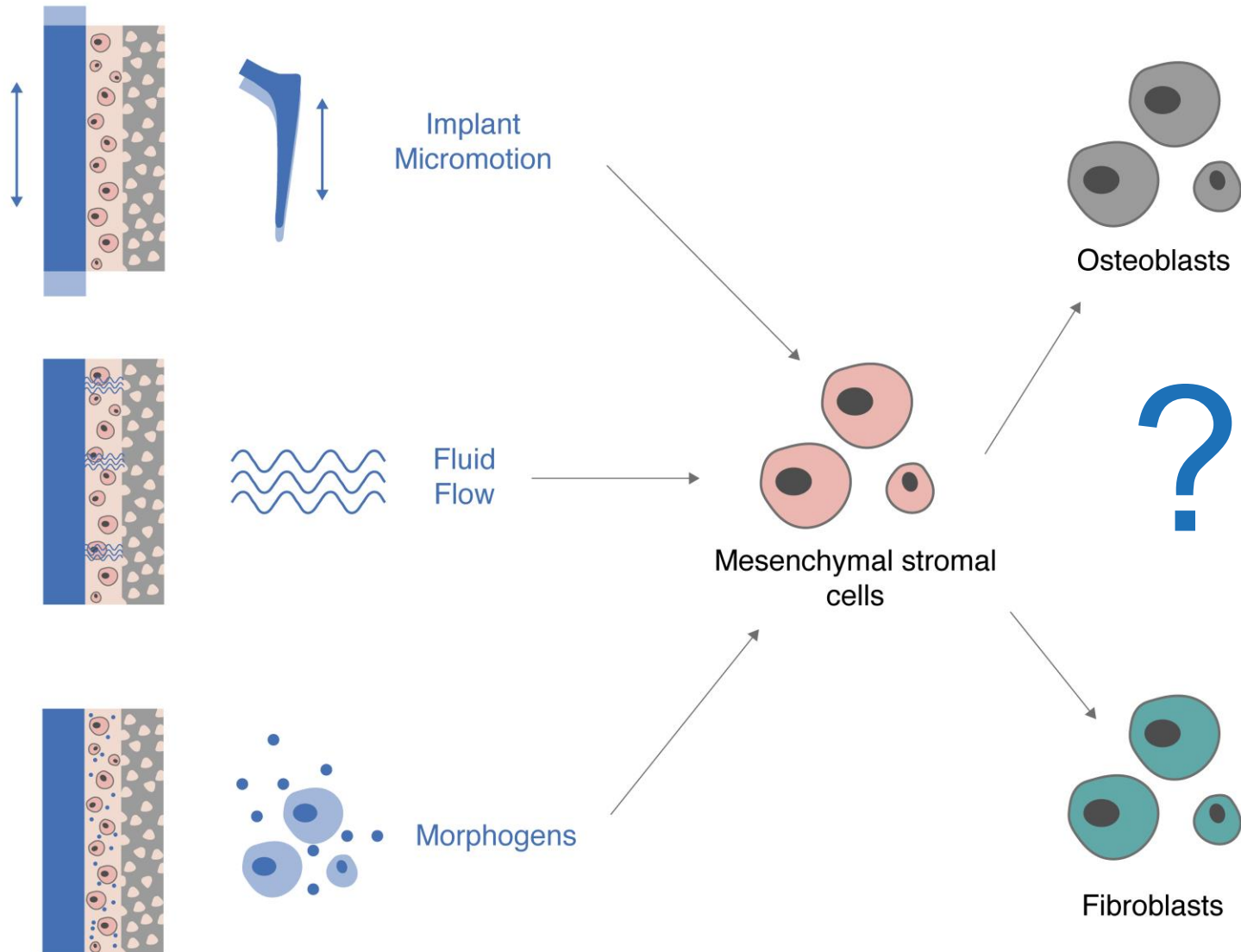


MSCs Differentiation



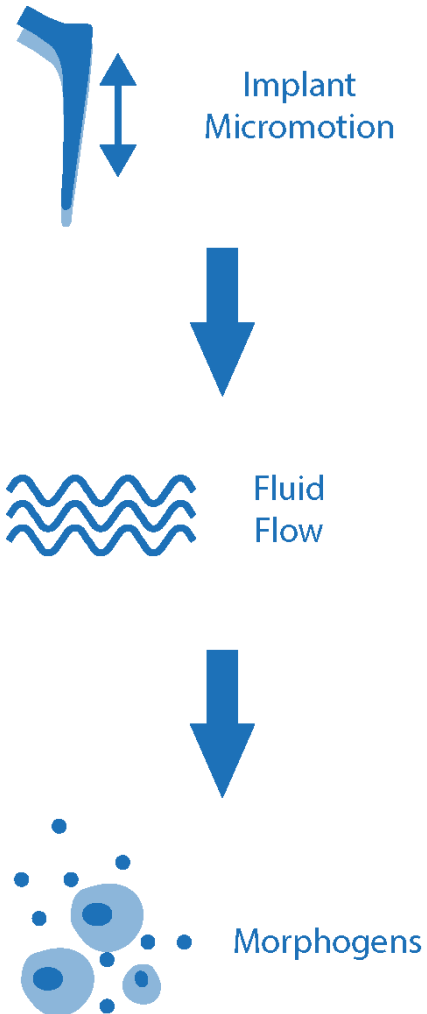


MSCs Differentiation





Hypothesis





Modeling Strategy



Implant
Micromotion

Quantify implant micromotion



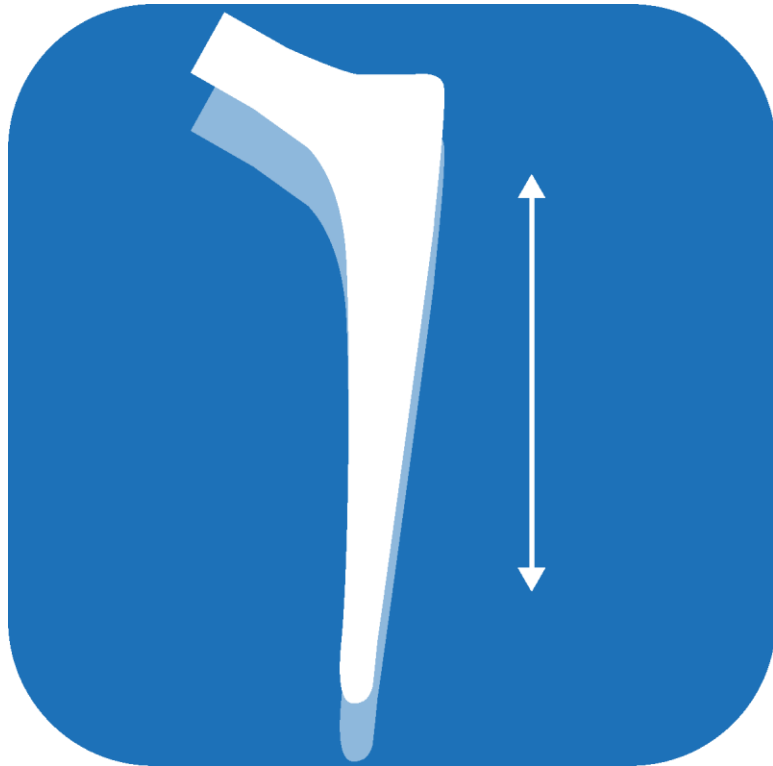
Fluid
Flow

*Estimate micromotion-induced
fluid flow in peri-implant tissue*



Morphogens

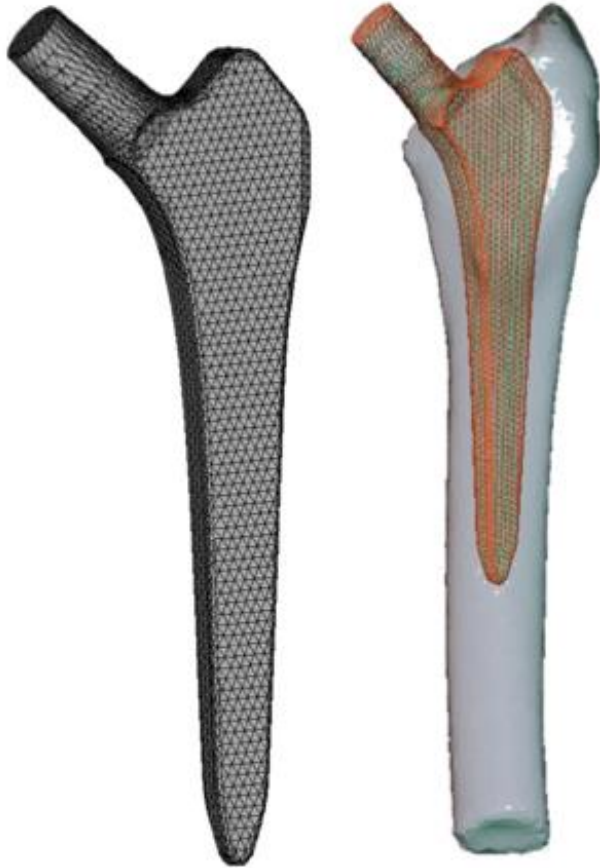
*Study the effects of fluid on
morphogens distribution and
MSCs differentiation*



Quantify
implant micromotion



Geometry

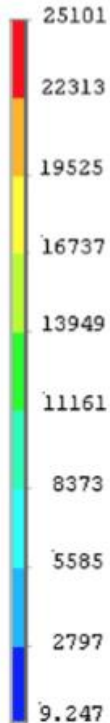
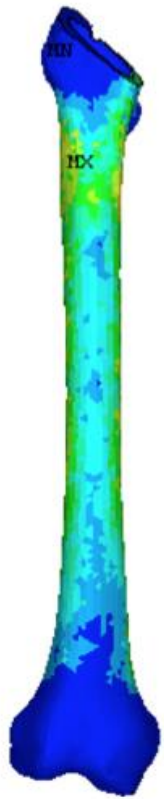


Bone: From patients CT-scans

Implant: From CAD file



Material



Elastic
Modulus



Mineral
Density

Bone: Linear elastic with an elastic modulus dependent on mineral density obtained from the CT scan

Implant: Linear elastic (titanium alloy)



Governing Equations

Solid Mechanics

Navier's equation for solid:

$$-\nabla \sigma = F$$

Hooke's law constitutive
equation for linear elastic
material:

$$\sigma = C : \varepsilon$$



Boundary Conditions

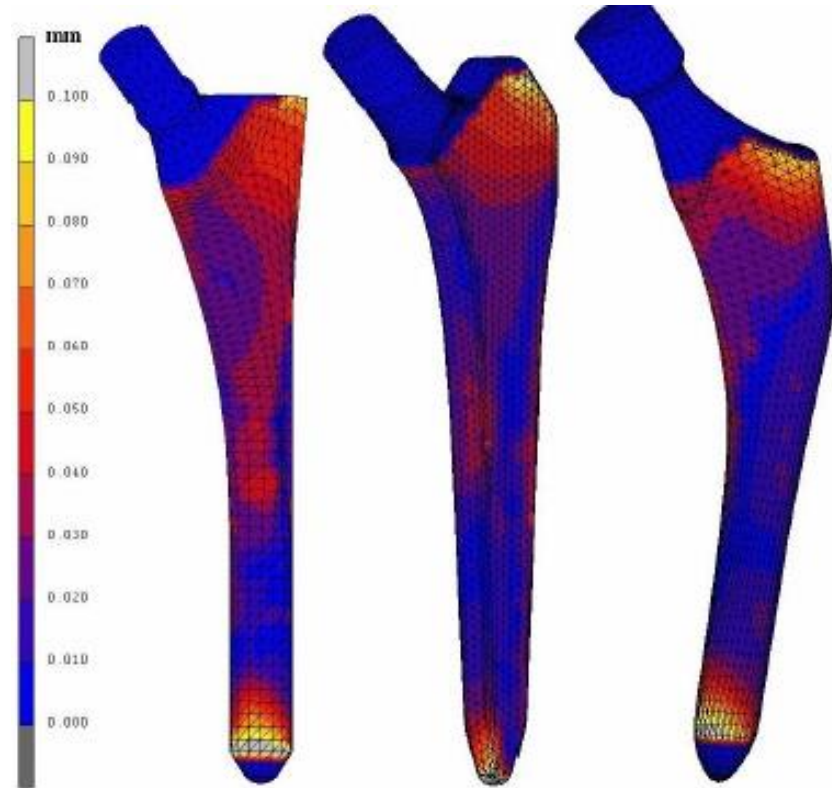


Loads and constraints:

- Experimental measurements in instrumented prostheses
- ISO standards for implant testing



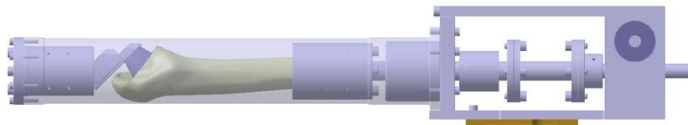
Results



Micromotion extends locally from a few μm to 100 μm



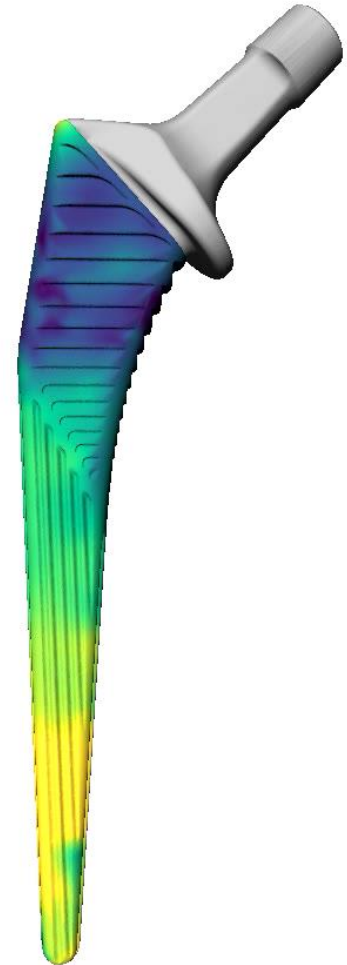
Experimental Validation

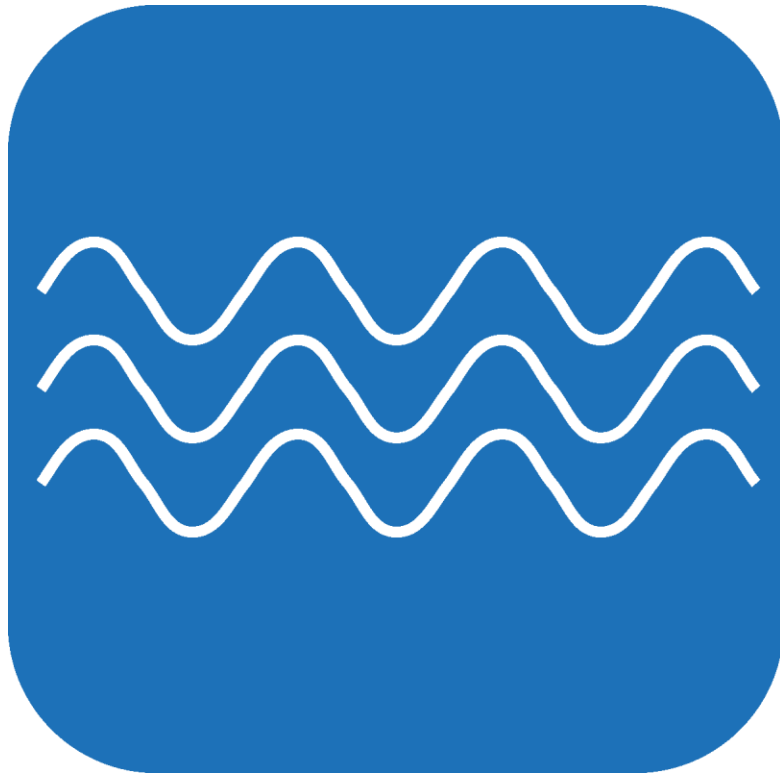


Experimental loading of implanted cadaveric femur



Analyze the displacements of radiopaque markers in a μ -CT scan

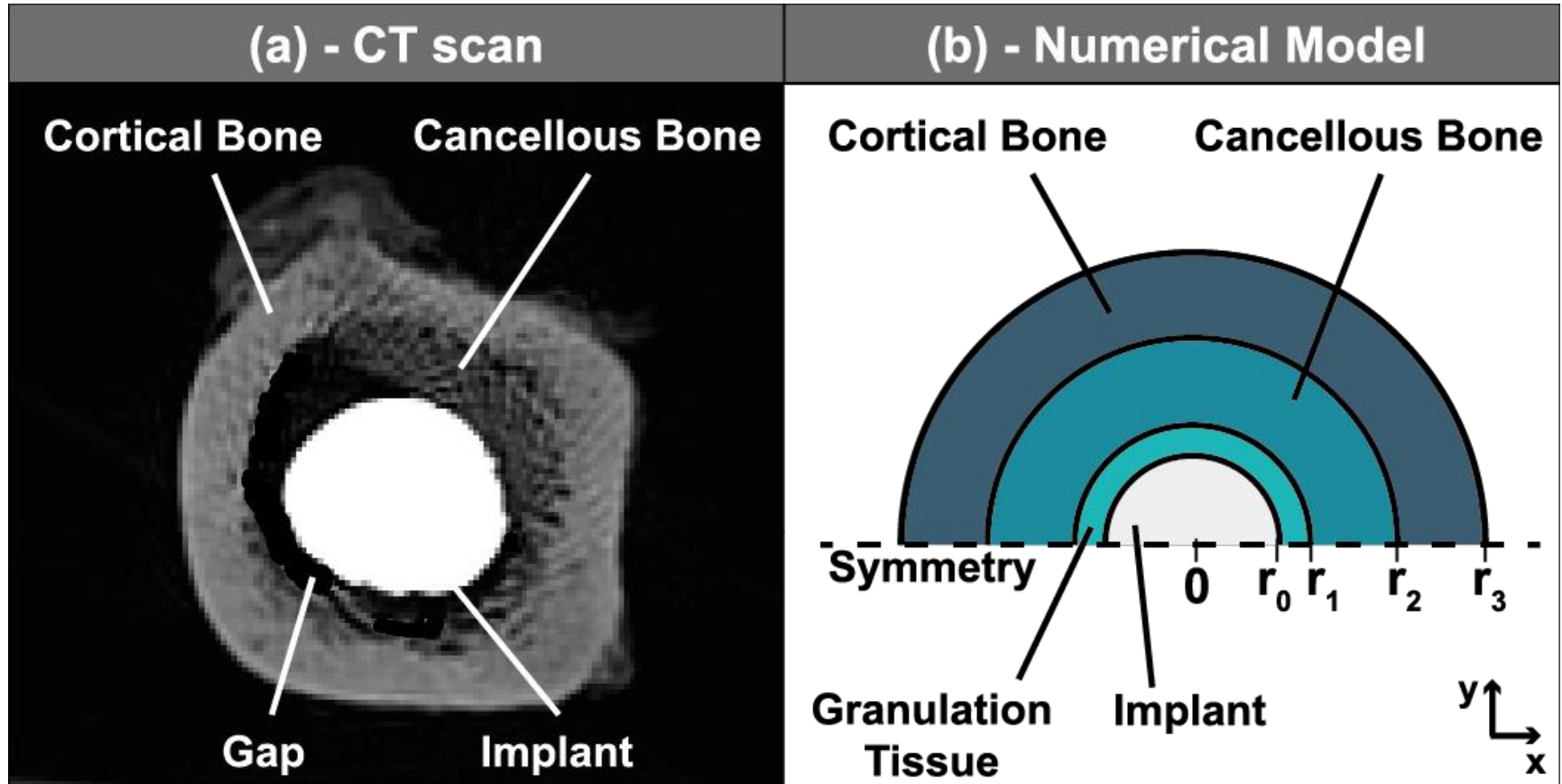




Quantify
micromotion-induced
fluid flow with a
poroelastic FE model



Geometry



<http://dx.doi.org/10.1080/10255842.2017.1296954>



Governing Equations

Biot's poroelasticity

Navier's equation for solid:

$$-\nabla \sigma = 0$$

Darcy's law combined with continuity equation:

Darcy's (fluid) velocity

↓

$$S \frac{\partial p_f}{\partial t} + \nabla \cdot \left[-\frac{\kappa}{\mu} \nabla p_f \right] = -\alpha_B \frac{\partial \varepsilon_{vol}}{\partial t}$$

Coupled through Biot's constitutive relations:

$$\sigma = \mathbf{C}(E, \nu) \varepsilon(\mathbf{u}) - \alpha_B p_f$$

$$p_f = \frac{1}{S} (\zeta - \alpha_B \varepsilon_{vol})$$

σ : stress tensor

ε : strain tensor

ε_{vol} : volumetric strain

\mathbf{C} : elastic tensor

E : Young modulus

ν : Poisson ratio

p_f : fluid (pore) pressure

κ : permeability

μ : viscosity

α_B : Biot-Willis coefficient

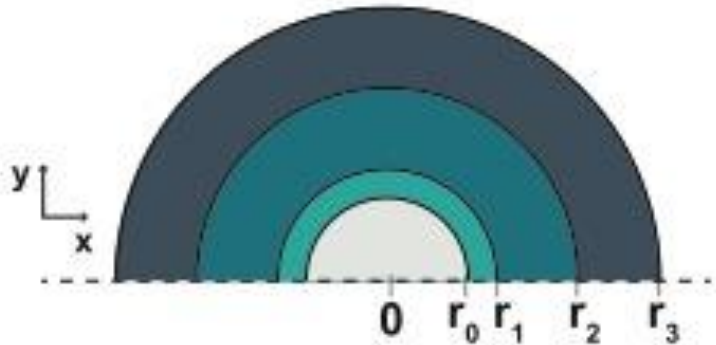
S : storage coefficient

$S = S(\alpha, \text{porosity, fluid bulk modulus, solid bulk modulus})$

ζ : fluid volume per unit volume



Initial and Boundary Conditions



Initial conditions:

$$p_f = 0, \mathbf{u} = 0$$

Boundary conditions:

Solid

$$\begin{cases} \mathbf{u} = 0 & , \forall \mathbf{r} = \mathbf{r}_3 \\ \mathbf{u} = \mathbf{u}_0 \cdot \frac{1}{2} \sin(2\pi f t - \frac{\pi}{2}) & , \forall \mathbf{r} = \mathbf{r}_0 \end{cases}$$

Fluid

$$\begin{cases} \mathbf{n} \cdot \nabla p_f = 0 & , \forall \mathbf{r} = \mathbf{r}_0, \mathbf{r}_3 \\ p_f = 0 & , \forall \mathbf{r} = \mathbf{r}_1, \mathbf{r}_2 \end{cases}$$

$$\mathbf{u}_0 = \text{Icon of a person with a vertical double-headed arrow}$$

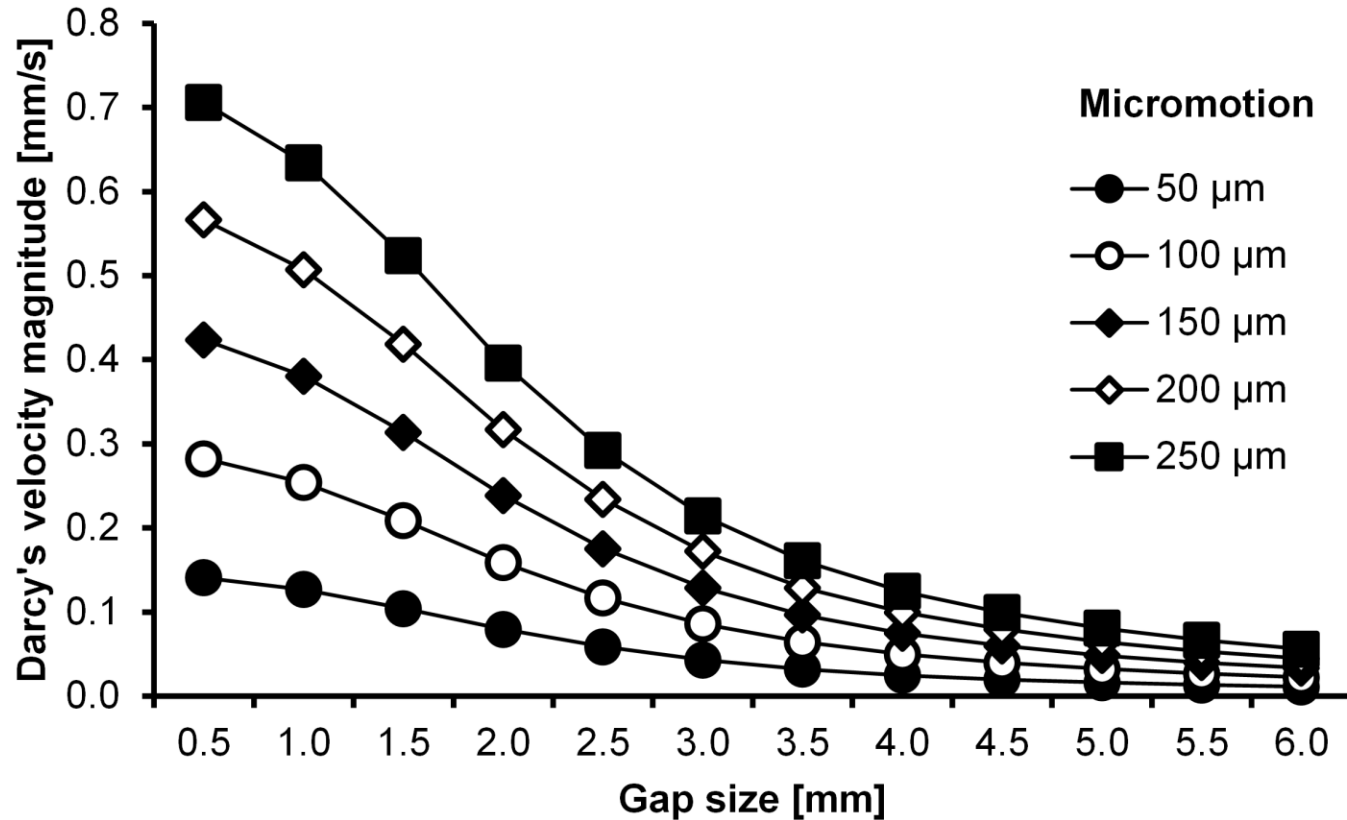


Material

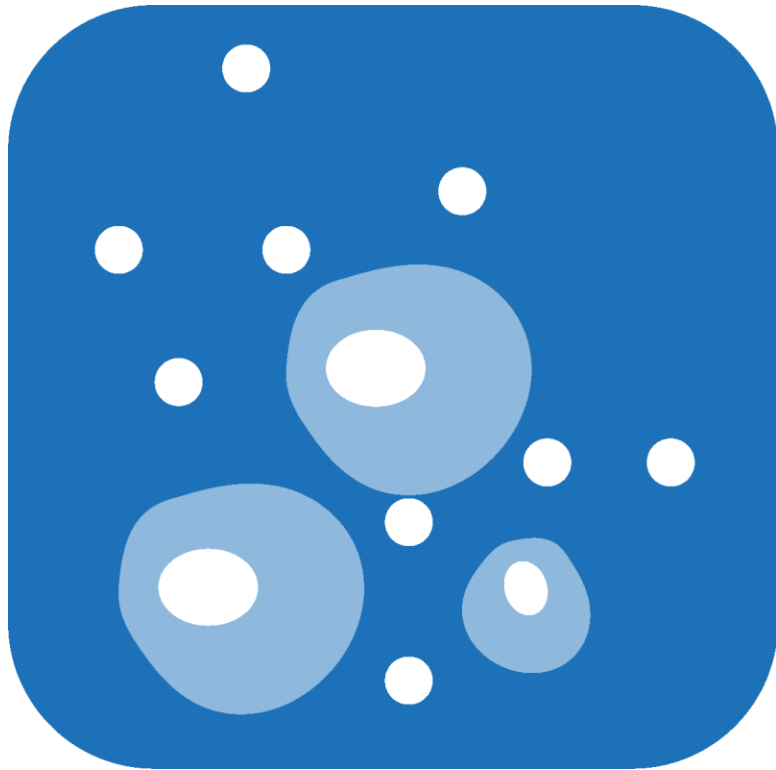
Property	Material		
	Granulation Tissue	Cancellous Bone	Cortical Bone
Young's Modulus (E)	1 MPa	6 GPa	15.75 GPa
Poisson's Ratio (ν)	0.167	0.325	0.325
Porosity (ϵ_p)	0.8	0.8	0.04
Permea- bility (κ)	$1e^{-17} \text{ m}^2$	$3.7e^{-16} \text{ m}^2$	$1e^{-20} \text{ m}^2$



Results



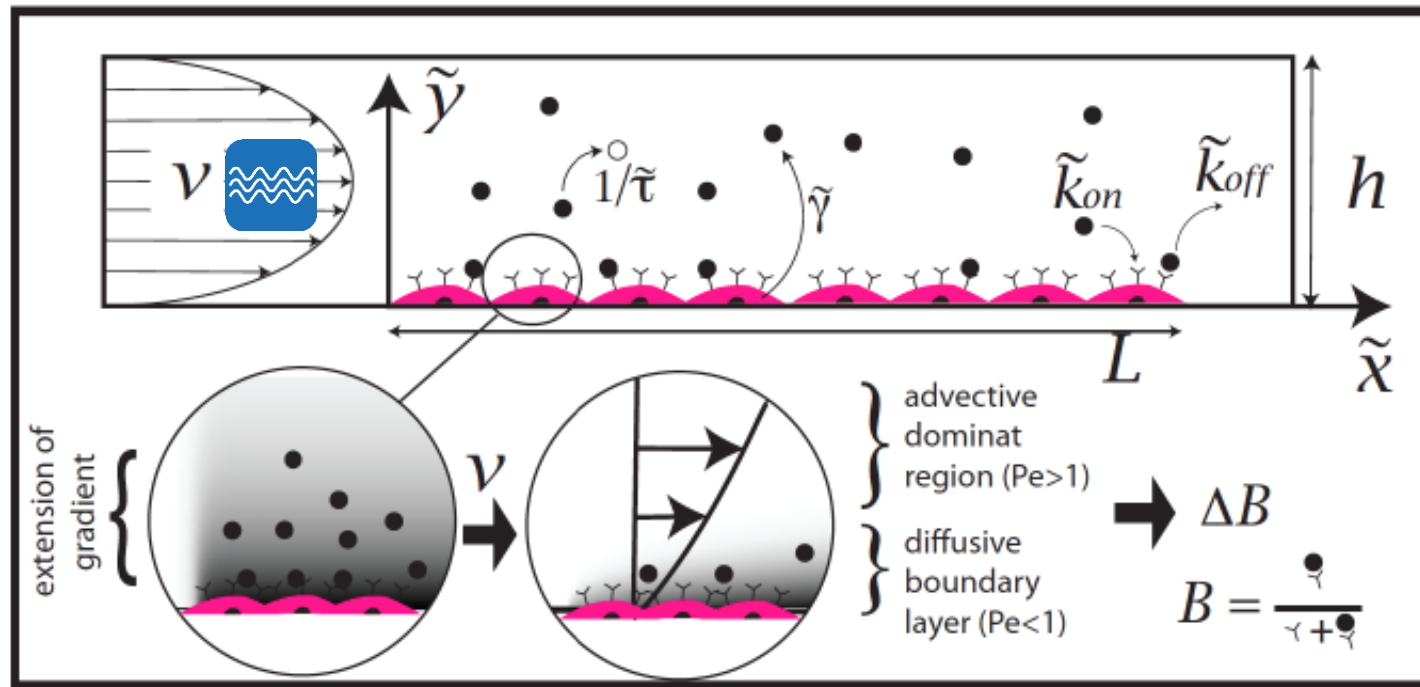
Micromotion-induced fluid flow in granulation tissue extends 1 $\mu\text{m/s}$ to 700 $\mu\text{m/s}$



Study the effects of
fluid on morphogens
distribution and
MSCs differentiation



Model



$$B = \frac{\text{surface density of } bound \text{ receptors}}{\text{surface density of receptors}}$$



Governing Equations

Mass transport

Diffusion (**D**) – Advection (**A**) – Reaction (**R**) in bulk:

$$\frac{\partial C}{\partial t} = \underbrace{\nabla \cdot [D(\nabla C)]}_{\mathbf{D}} - \underbrace{\nabla \cdot (C u)}_{\mathbf{A}} - \underbrace{\frac{1}{\tau} C}_{\mathbf{R}}$$

C - concentration

R_t – total number of receptors

B – number of bound receptors

Binding (**B**) – Unbinding (**UB**) reaction on the wall :

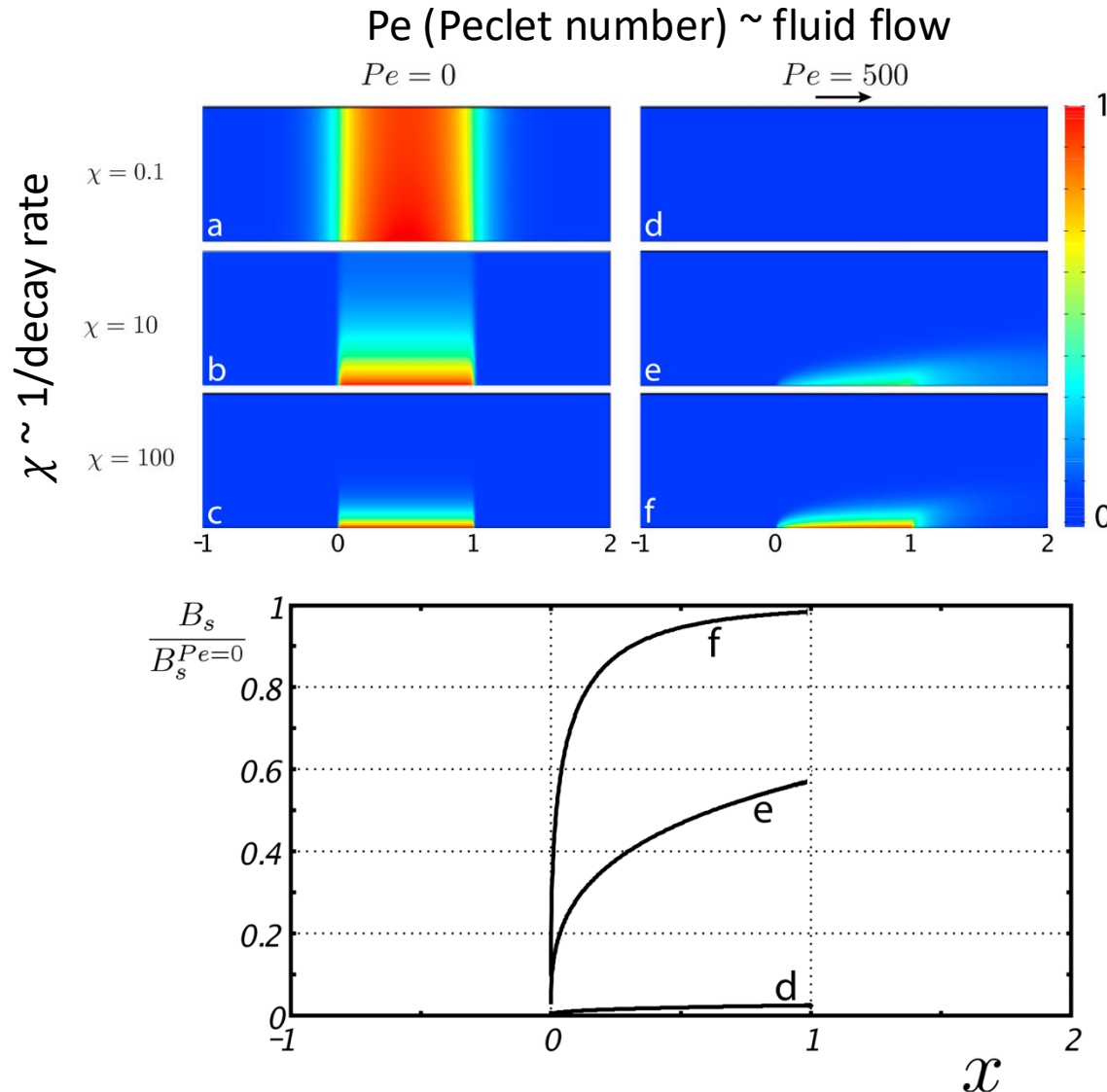
$$\frac{\partial B}{\partial t} = \underbrace{k_{on}(R_t - B)C|_{y=0}}_{\mathbf{B}} - \underbrace{k_{off}B}_{\mathbf{UB}}$$

Secretion(**S**) of morphogens:

$$\underbrace{\gamma}_{\mathbf{S}} - \frac{\partial B}{\partial t} = -D \frac{\partial}{\partial y} C \Big|_{y=0}$$



Results



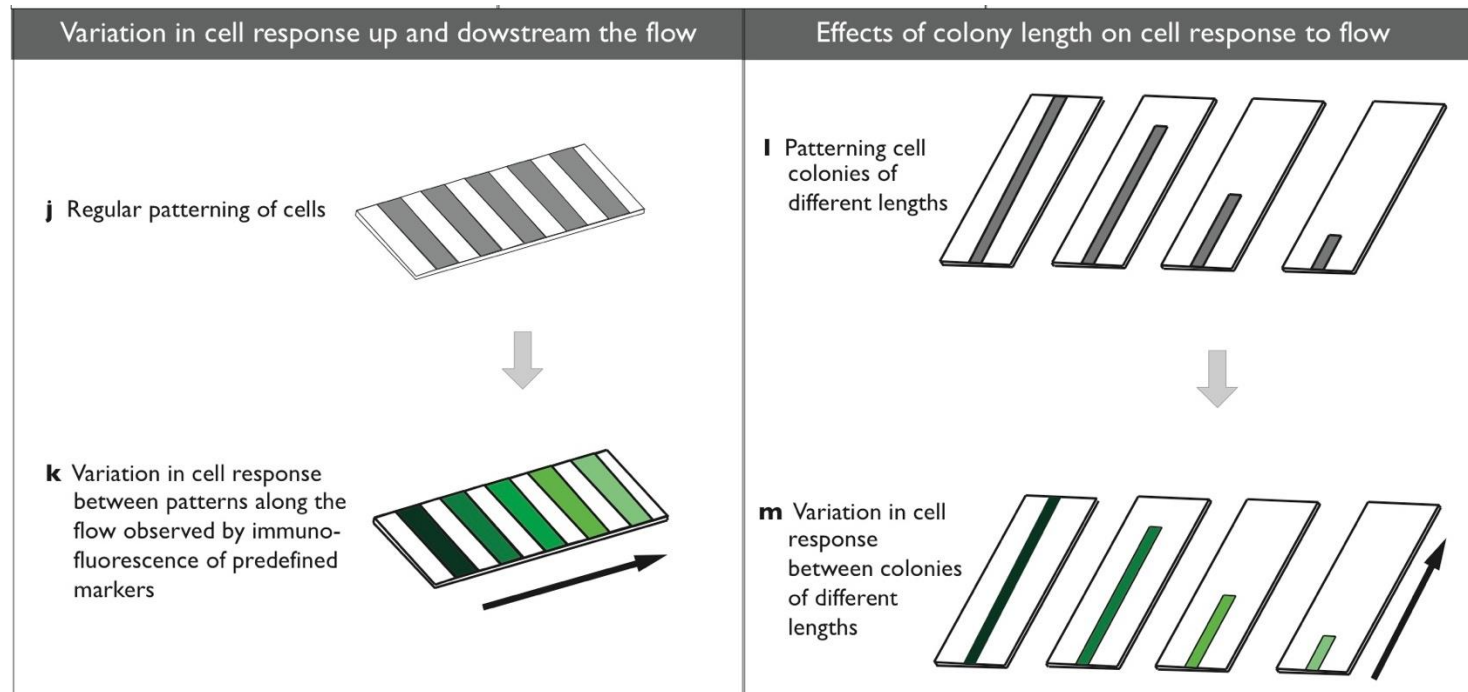
Fluid flow is strong enough to:

- Disturb the concentration profile of morphogens
- Change the number of bound receptors (i.e. have an effect on cell differentiation)



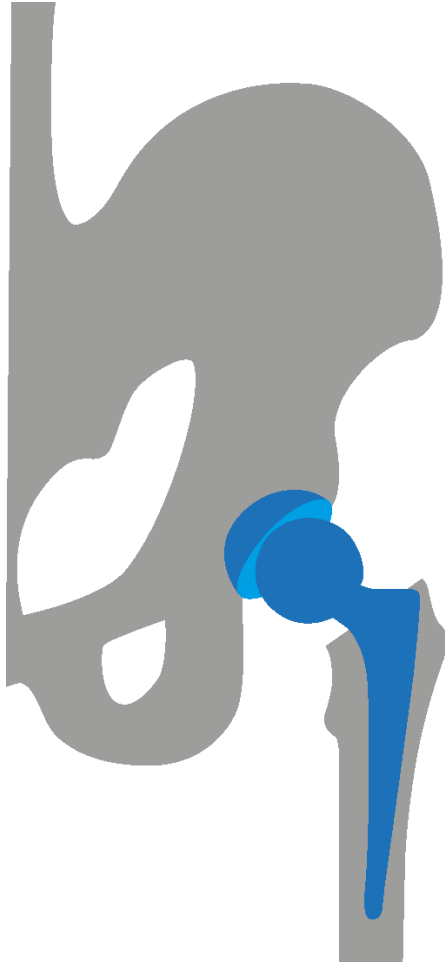
Experimental Validation

Microfluidics experiments





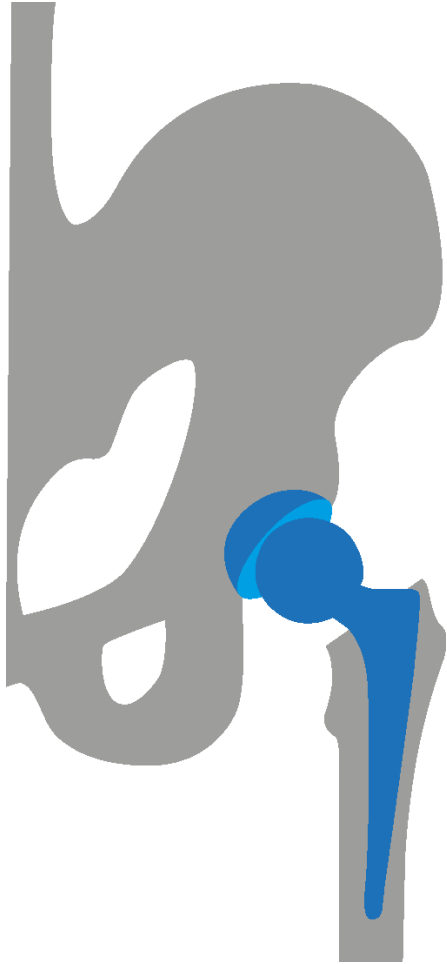
Conclusion



A better understanding of conditions promoting aseptic loosening of hip implants can lead to better implant designs or surgical techniques and benefit patients



Conclusion



Numerical modeling helps to investigate complex multi-scale hypotheses.

However, experimental validation is essential to assess the predictive capabilities of models.