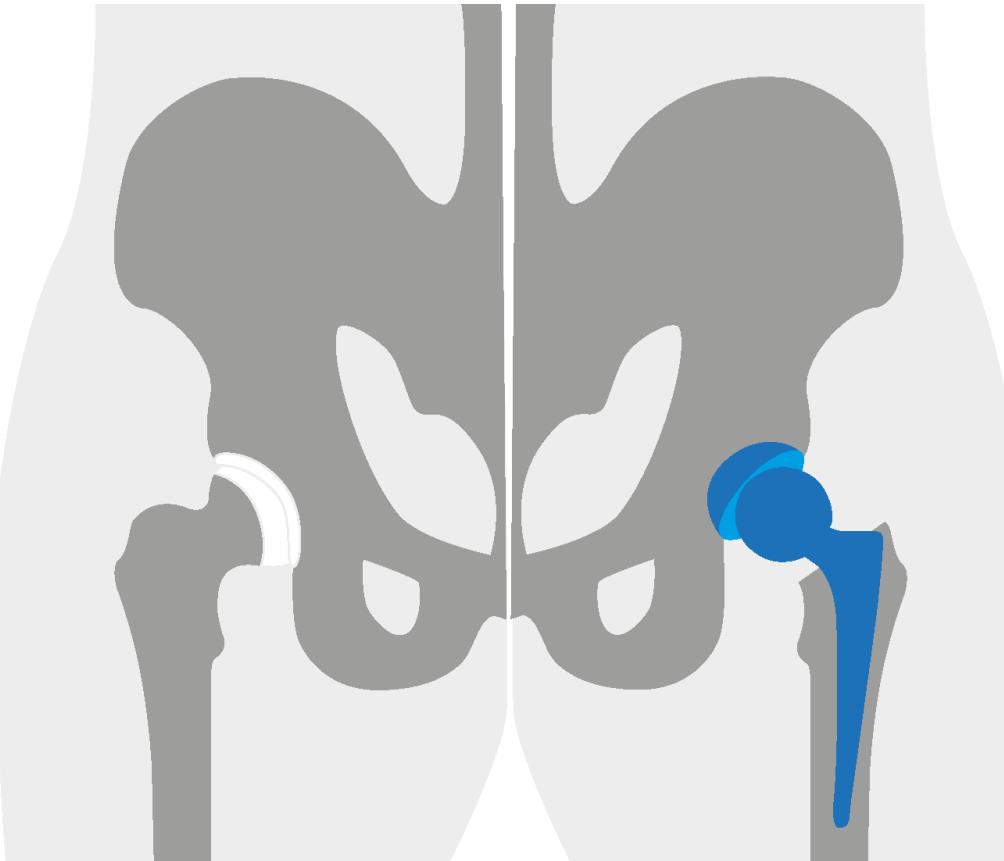


# Numerical Methods in Biomechanics

Alexandre Terrier, PhD

EPFL - Laboratory of Biomechanical Orthopedics

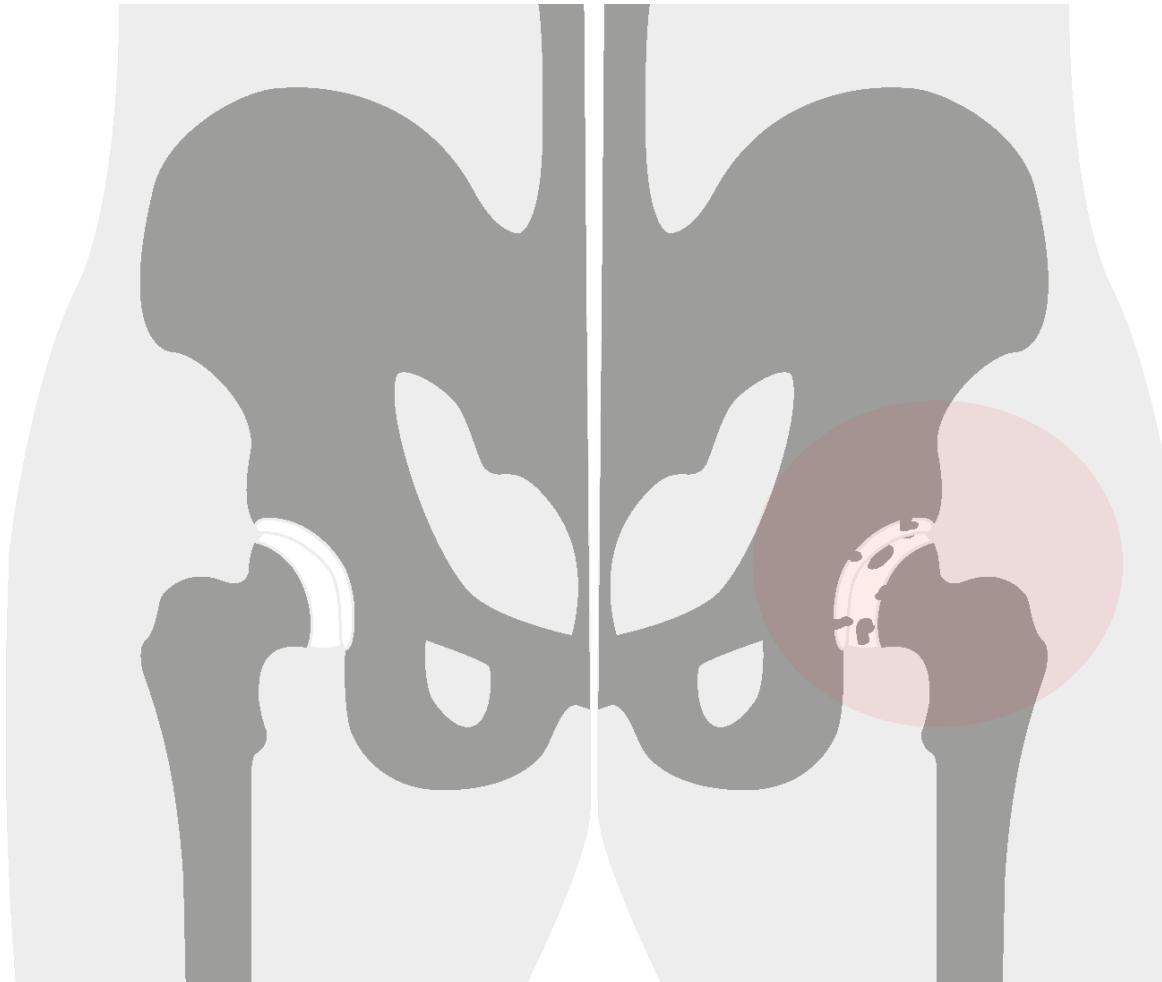


Numerical modeling  
to investigate  
aseptic loosening of  
hip implants

Valérie Malfroy Camine

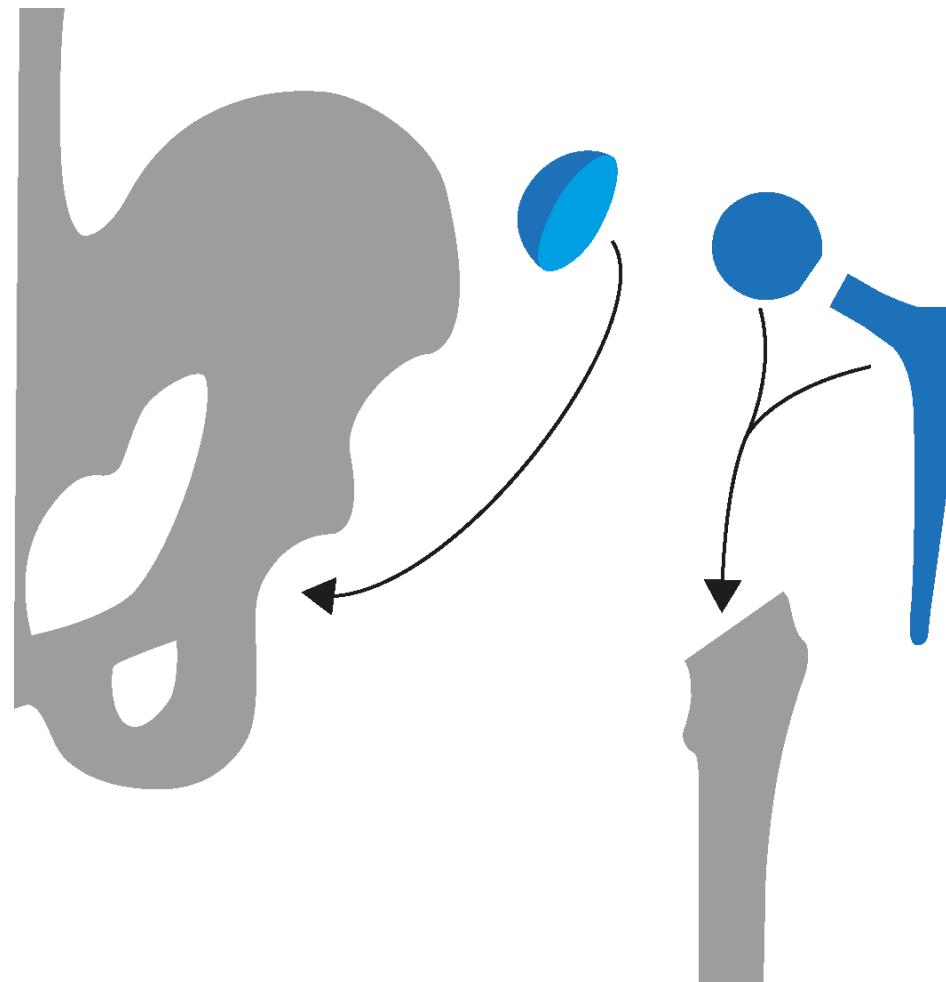


# Osteoarthritis





# Total Hip Replacement



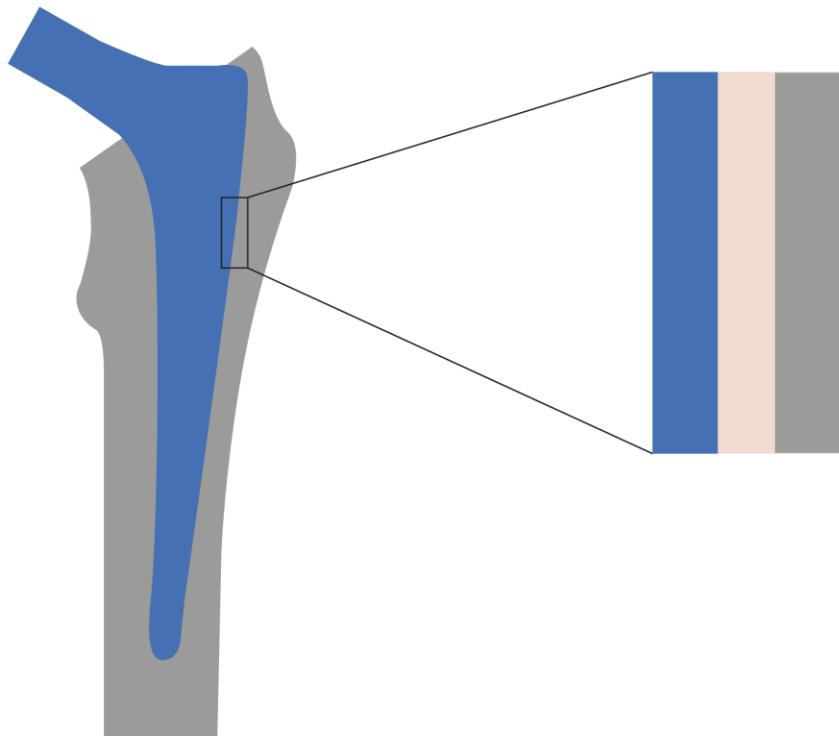


# Aseptic Loosening



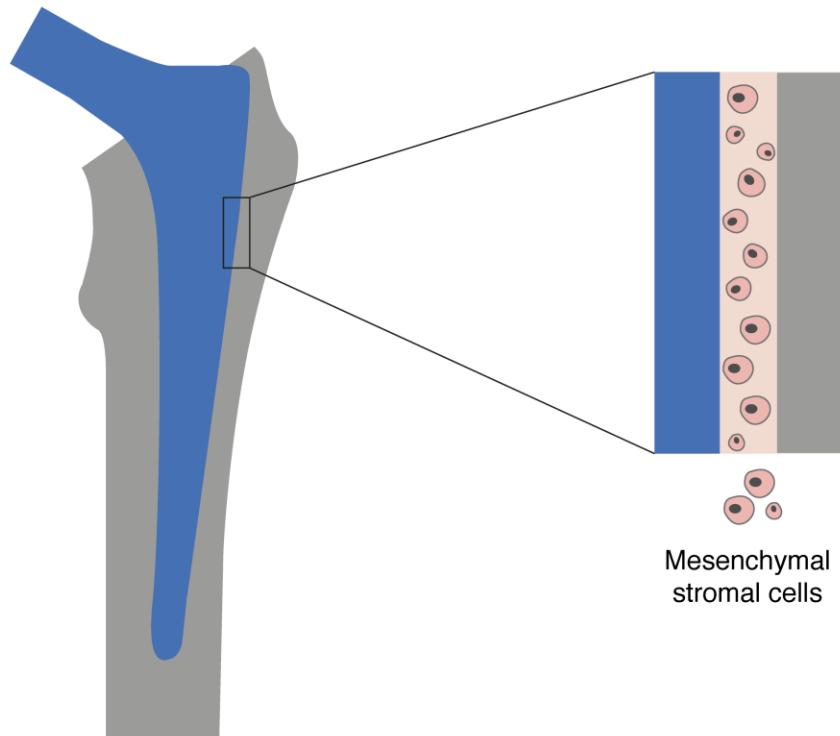


# Peri-implant Healing



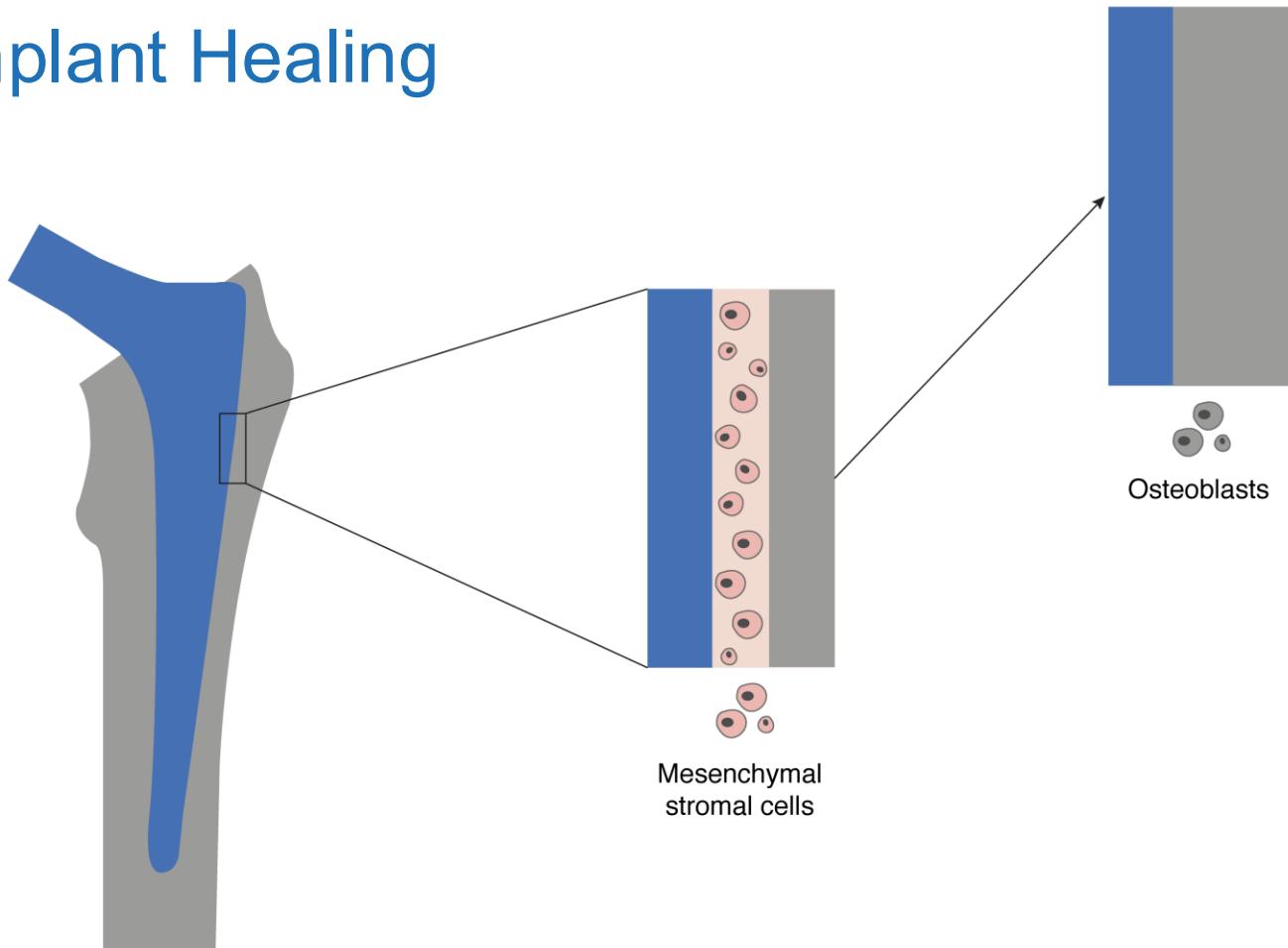


# Peri-implant Healing



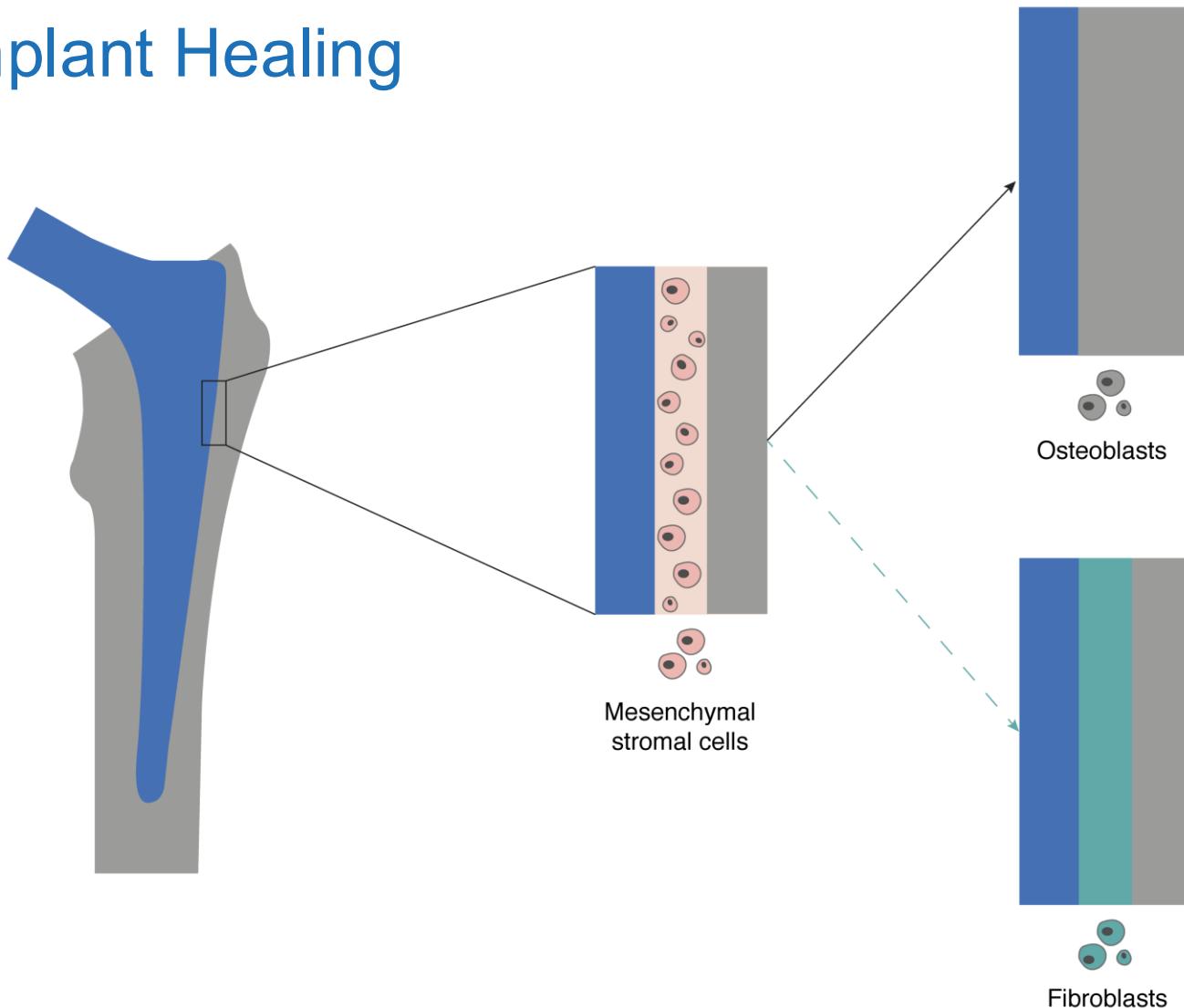


# Peri-implant Healing



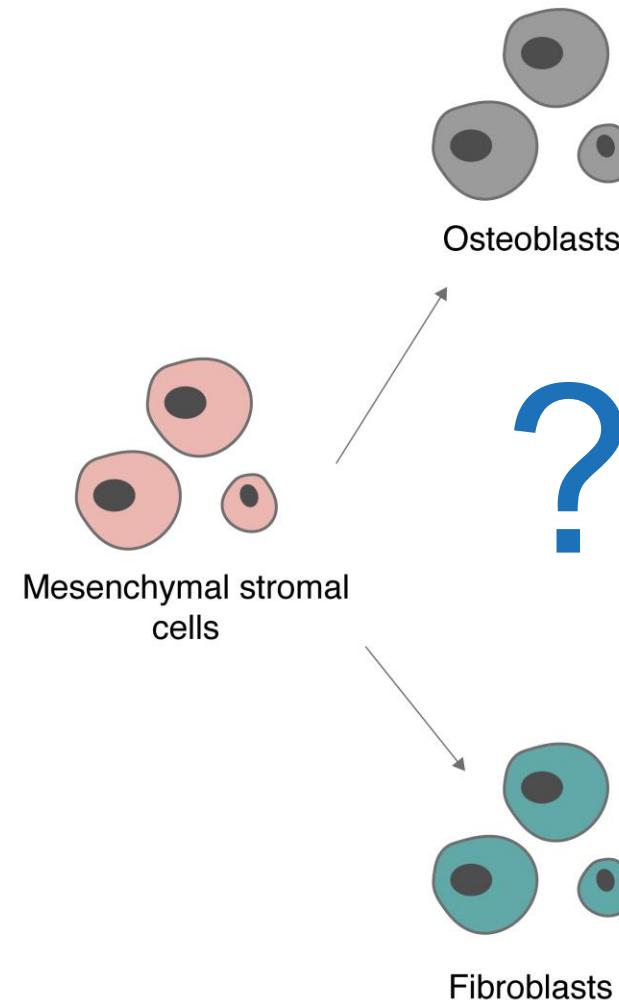


# Peri-implant Healing



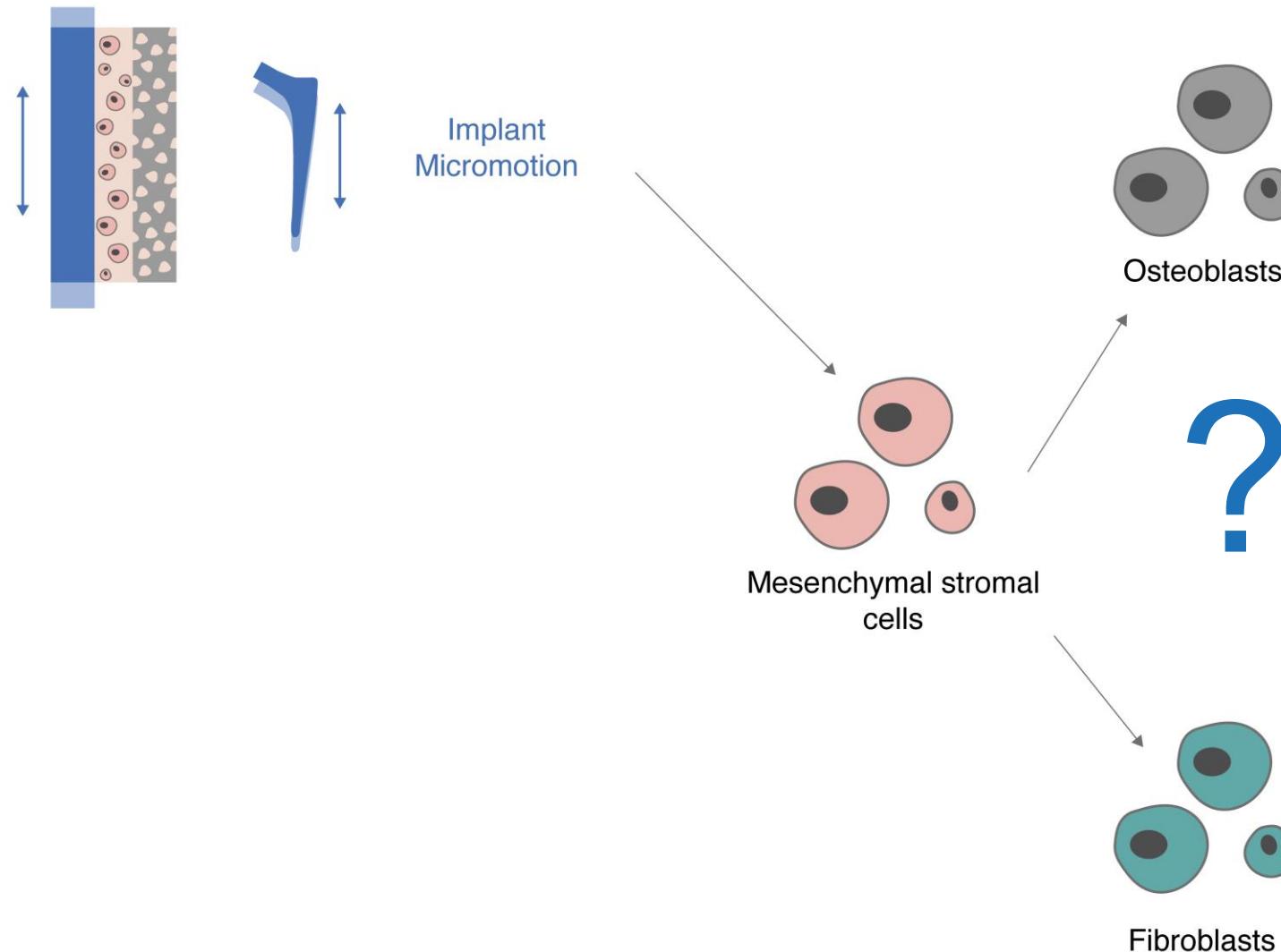


# MSCs Differentiation



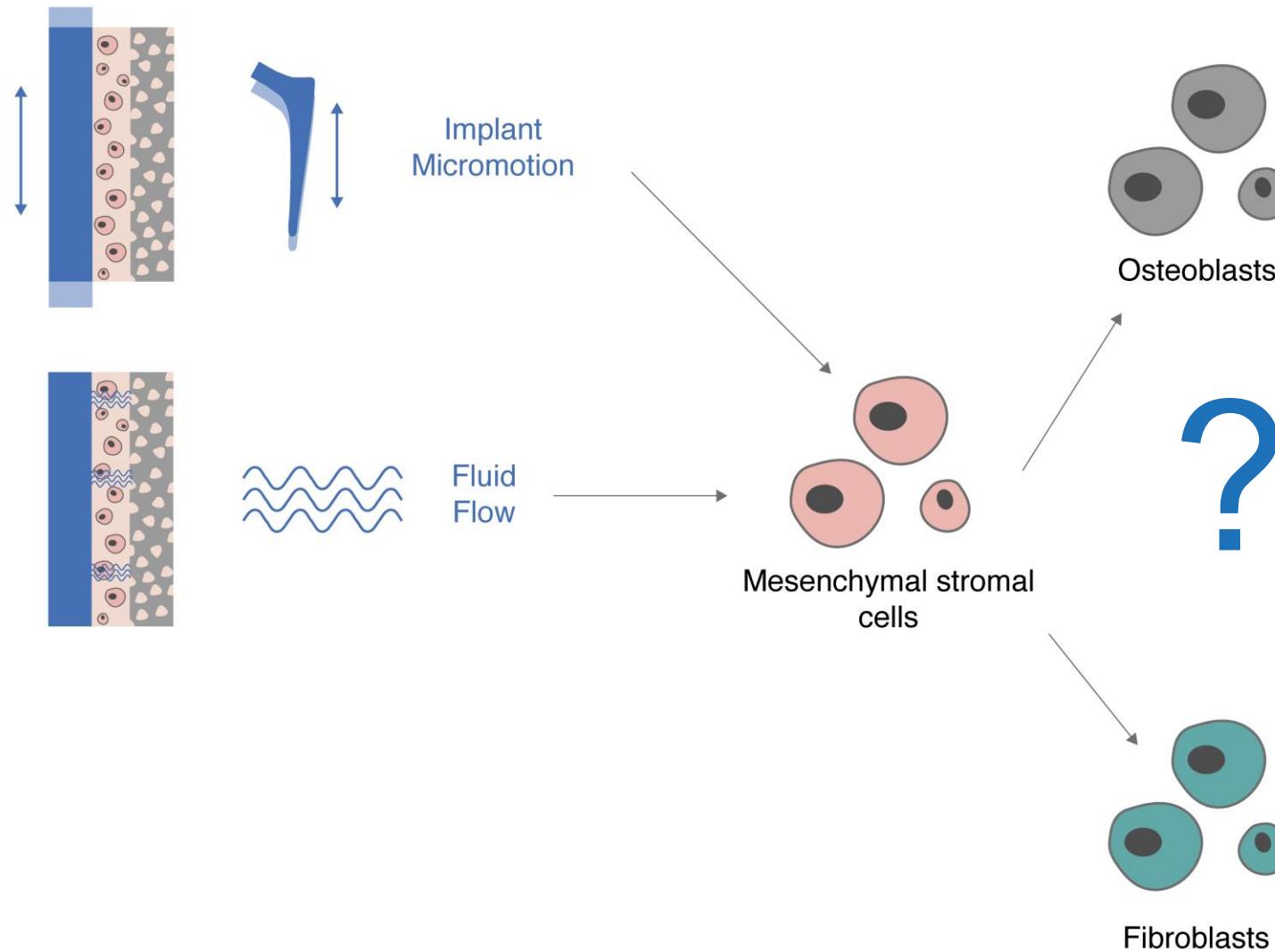


# MSCs Differentiation



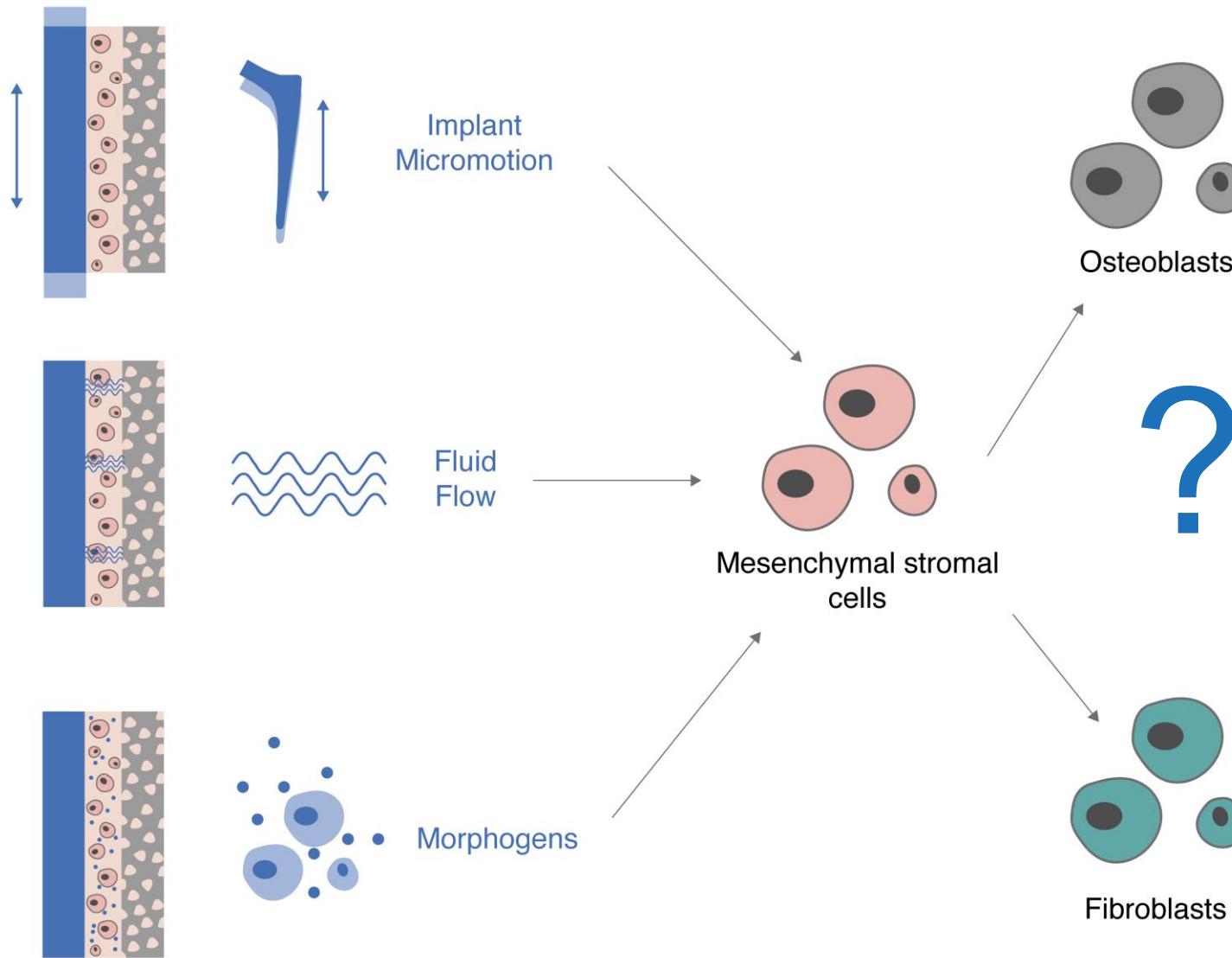


# MSCs Differentiation



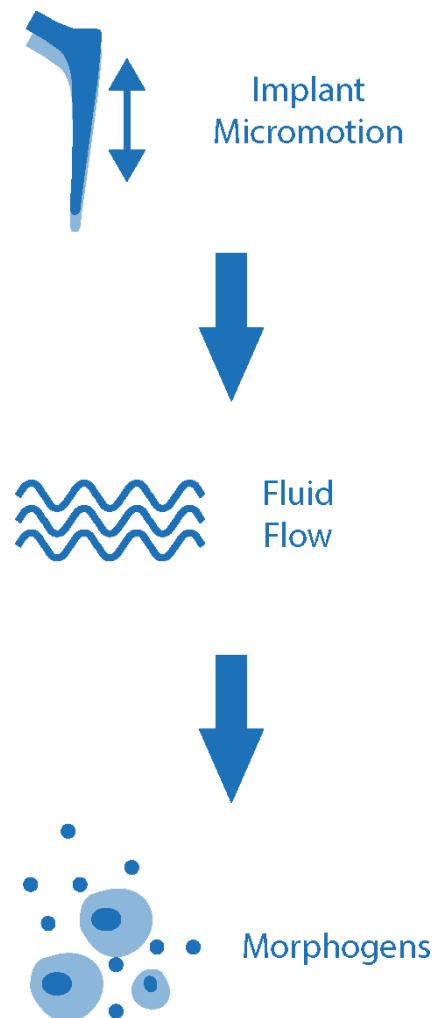


# MSCs Differentiation



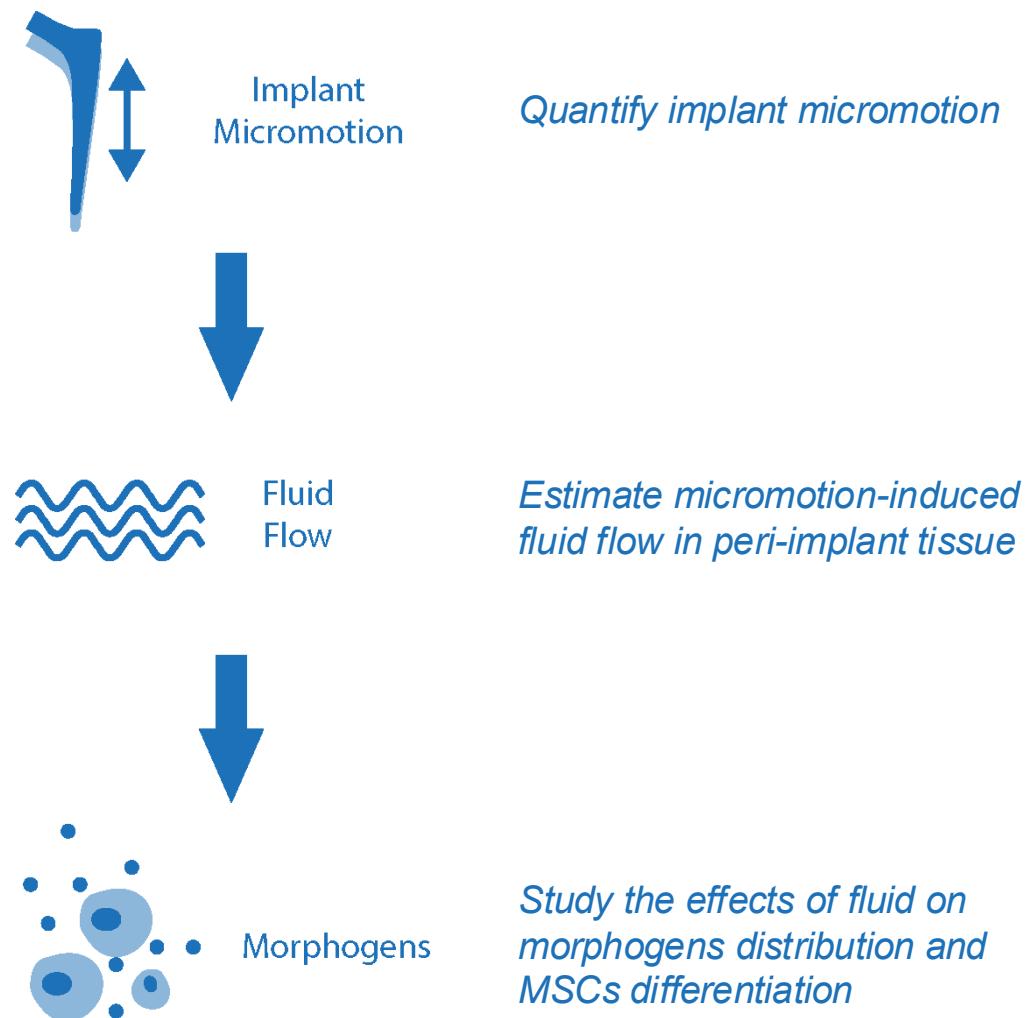


# Hypothesis





# Modeling Strategy





Quantify  
implant micromotion



# Geometry

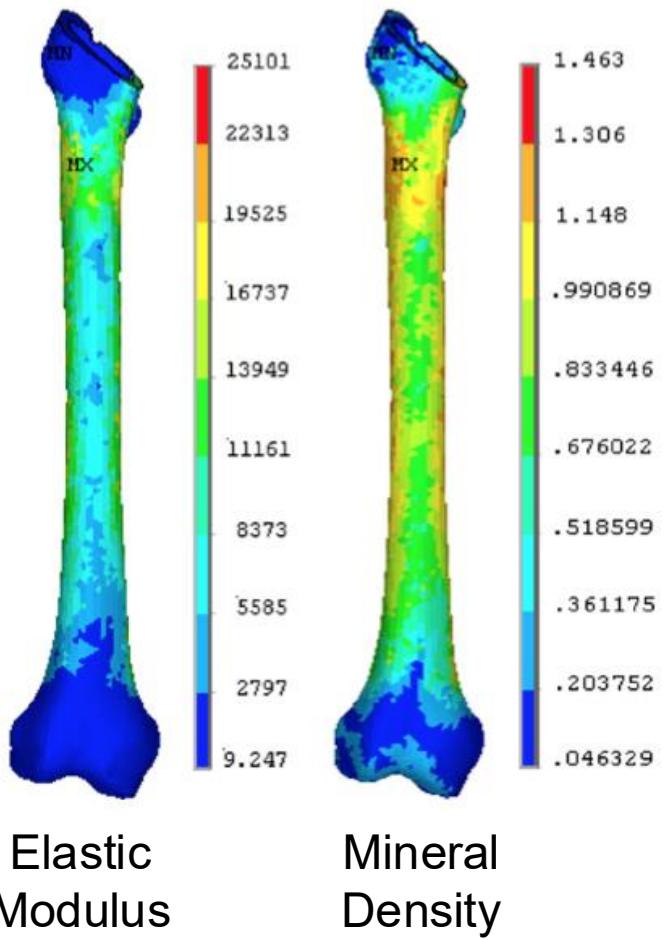


**Bone:** From patients CT-scans

**Implant:** From CAD file



# Material



**Bone:** Linear elastic with an elastic modulus dependent on mineral density obtained from the CT scan

**Implant:** Linear elastic (titanium alloy)



# Governing Equations

## Solid Mechanics

Navier's equation for solid:

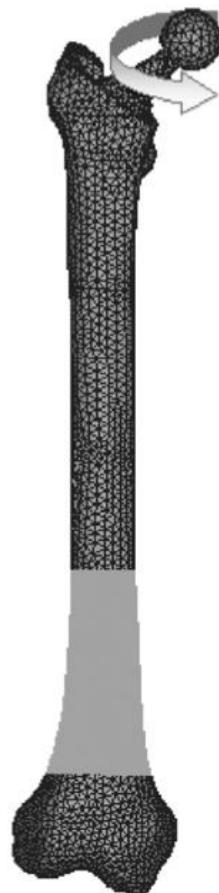
$$-\nabla \sigma = F$$

Hooke's law constitutive equation for linear elastic material:

$$\sigma = C : \varepsilon$$



# Boundary Conditions

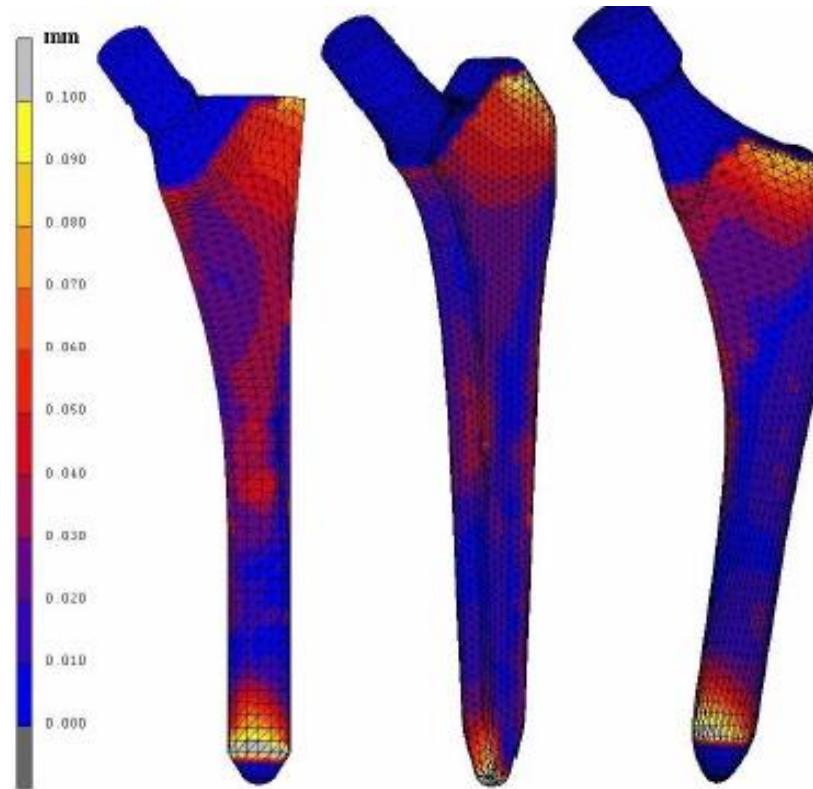


Loads and constraints:

- Experimental measurements in instrumented prostheses
- ISO standards for implant testing



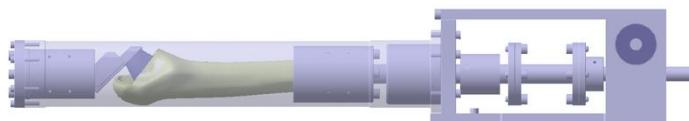
# Results



Micromotion extends locally from a few  $\mu\text{m}$  to 100  $\mu\text{m}$



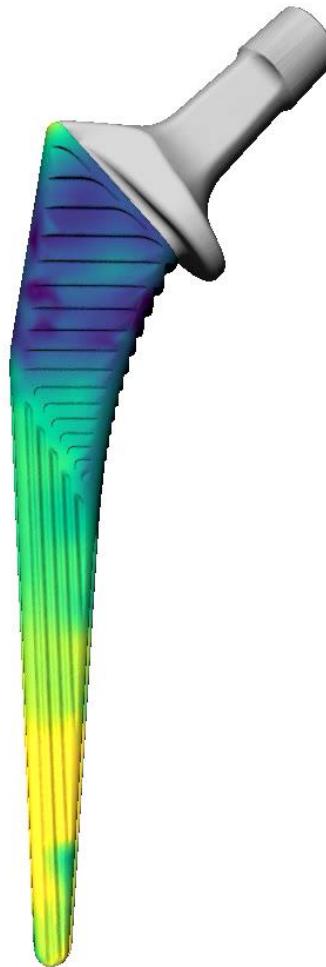
# Experimental Validation

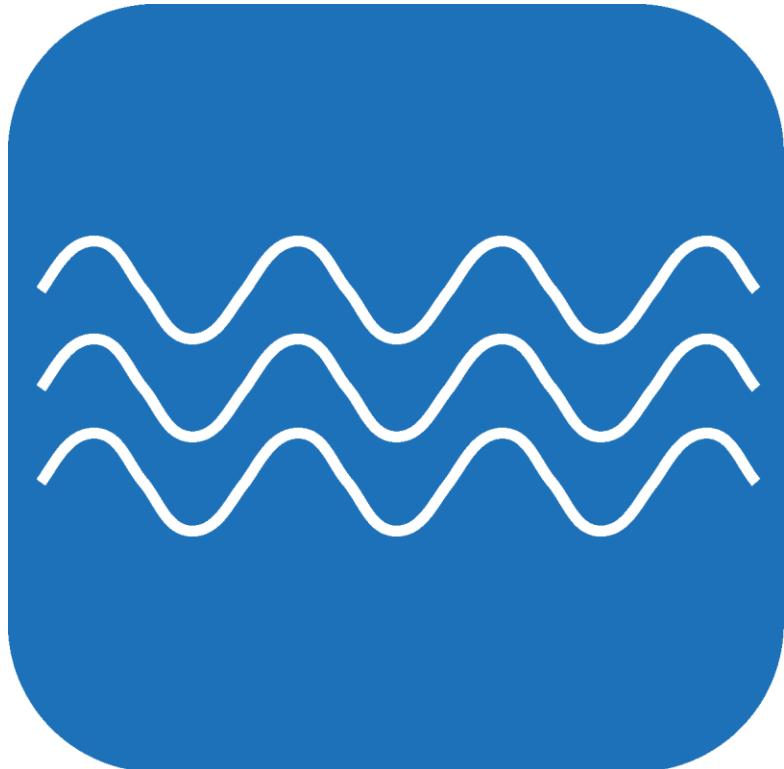


Experimental loading of implanted cadaveric femur



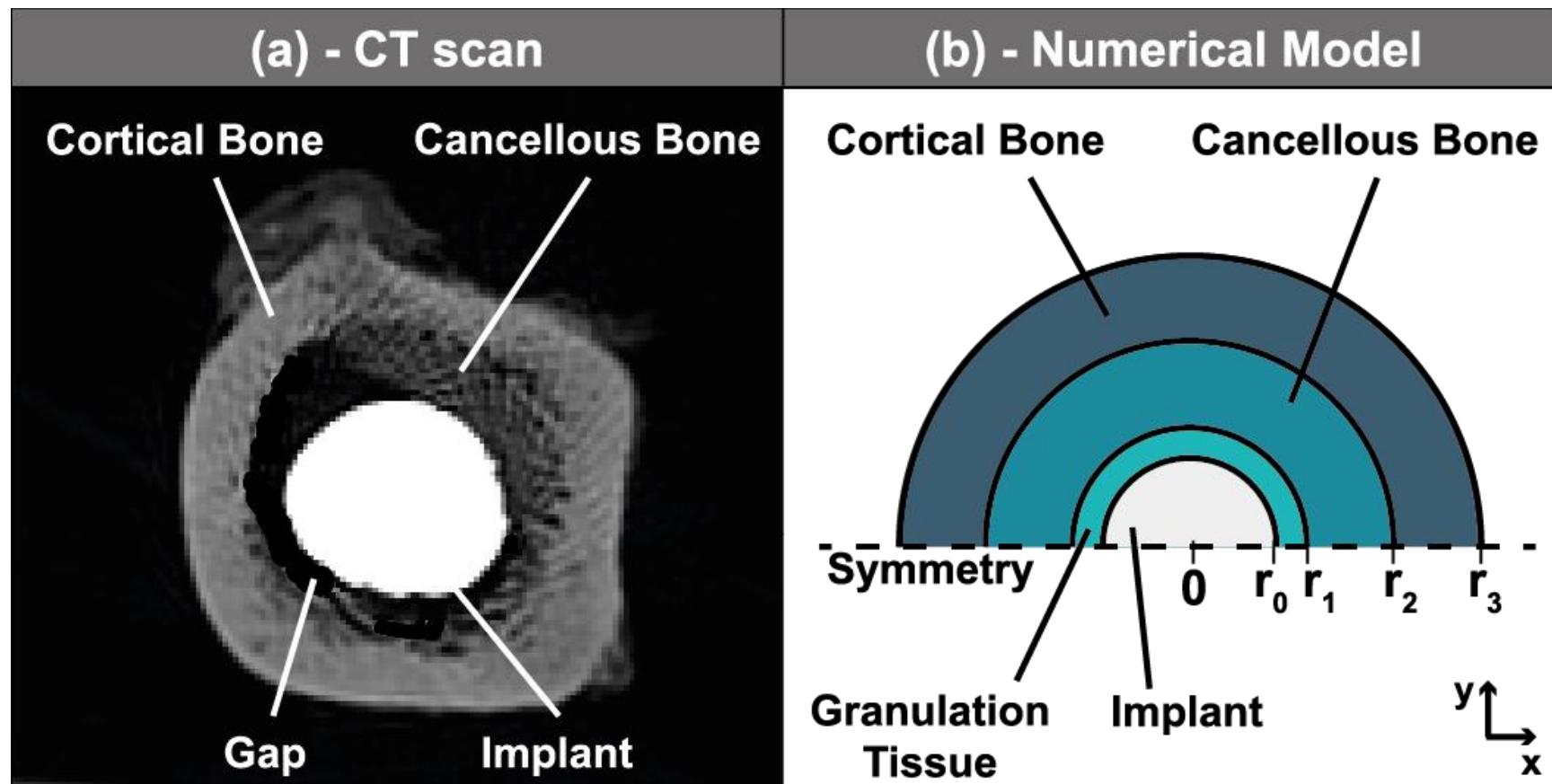
Analyze the displacements of radiopaque markers in a  $\mu$ -CT scan





Quantify  
micromotion-induced  
fluid flow with a  
poroelastic FE model

# Geometry





# Governing Equations

## Biot's poroelasticity

Navier's equation for solid:

Darcy's law combined with continuity equation:

Coupled through Biot's constitutive relations:

$\sigma$  : stress tensor

$\varepsilon$  : strain tensor

$\varepsilon_{vol}$  : volumetric strain

$C$  : elastic tensor

$E$  : Young modulus

$\nu$  : Poisson ratio

Darcy's (fluid) velocity

$$-\nabla \sigma = 0$$

$$S \frac{\partial p_f}{\partial t} + \nabla \cdot \left[ -\frac{\kappa}{\mu} \nabla p_f \right] = -\alpha_B \frac{\partial \varepsilon_{vol}}{\partial t}$$

$$\sigma = \mathbf{C}(E, \nu) \varepsilon(\mathbf{u}) - \alpha_B p_f$$

$$p_f = \frac{1}{S} (\zeta - \alpha_B \varepsilon_{vol})$$

$p_f$  : fluid (pore) pressure

$\kappa$  : permeability

$\mu$  : viscosity

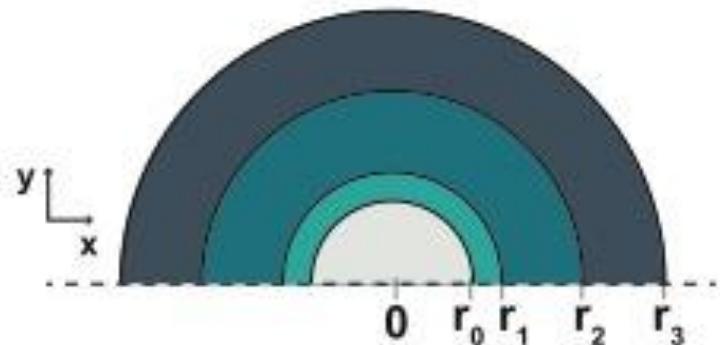
$\alpha_B$  : Biot-Willis coefficient

$S$  : storage coefficient

$S = S(\alpha, \text{porosity, fluid bulk modulus, solid bulk modulus})$



# Initial and Boundary Conditions



$$\mathbf{u}_0 =$$

Initial conditions:

$$p_f = 0, \mathbf{u} = 0$$

Boundary conditions:

*Solid*

$$\begin{cases} \mathbf{u} = 0 & , \forall \mathbf{r} = \mathbf{r}_3 \\ \mathbf{u} = \mathbf{u}_0 \cdot \frac{1}{2} \sin(2\pi ft - \frac{\pi}{2}) & , \forall \mathbf{r} = \mathbf{r}_0 \end{cases}$$

*Fluid*

$$\begin{cases} \mathbf{n} \cdot \nabla p_f = 0 & , \forall \mathbf{r} = \mathbf{r}_0, \mathbf{r}_3 \\ p_f = 0 & , \forall \mathbf{r} = \mathbf{r}_1, \mathbf{r}_2 \end{cases}$$

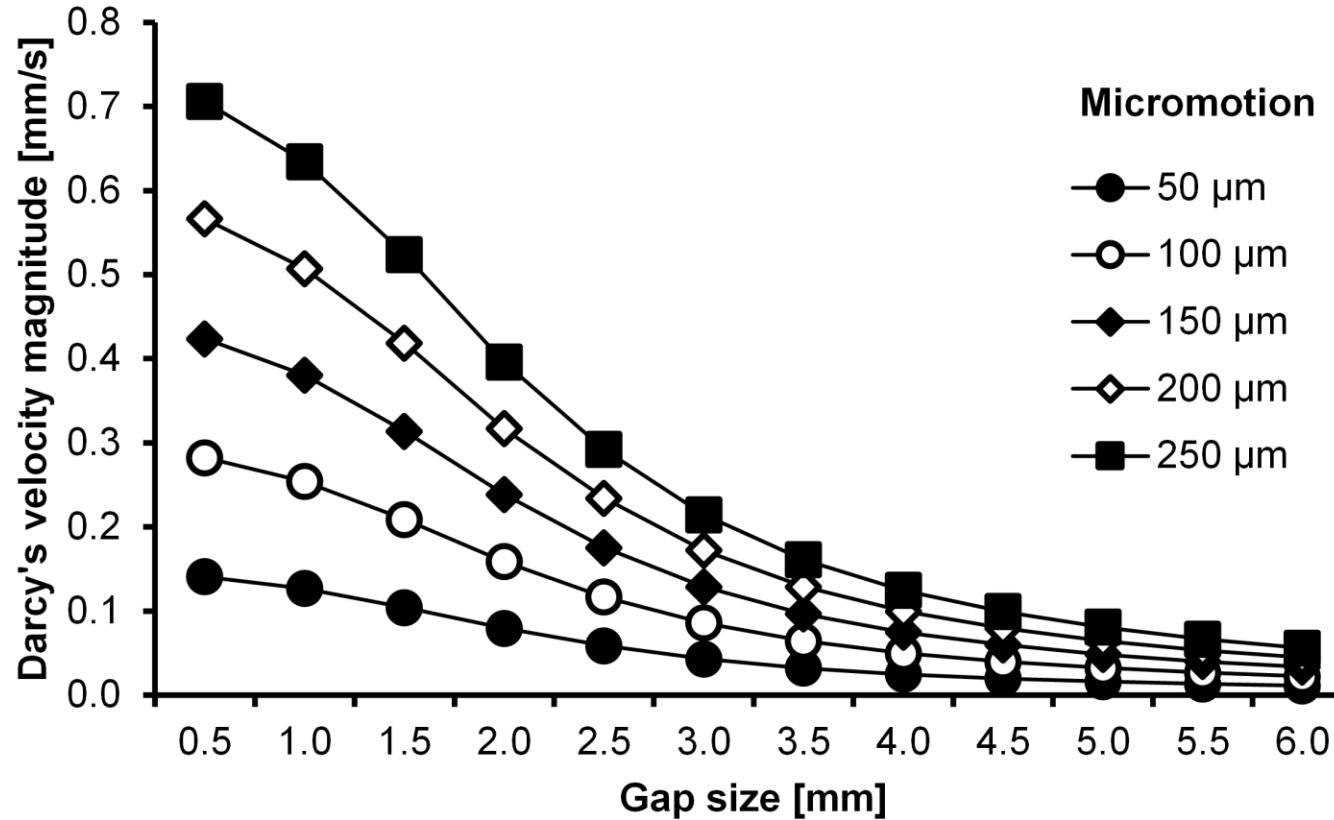


# Material

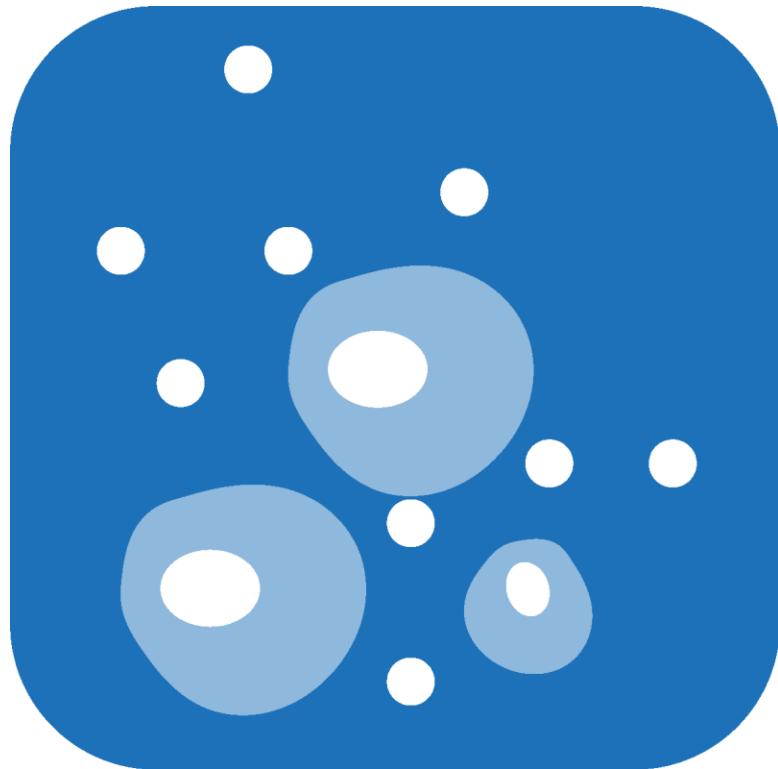
Property	Material		
	Granulation Tissue	Cancellous Bone	Cortical Bone
Young's Modulus (E)	1 MPa	6 GPa	15.75 GPa
Poisson's Ratio ( $\nu$ )	0.167	0.325	0.325
Porosity ( $\epsilon_p$ )	0.8	0.8	0.04
Permeability ( $\kappa$ )	$1e^{-17}$ m <sup>2</sup>	$3.7e^{-16}$ m <sup>2</sup>	$1e^{-20}$ m <sup>2</sup>



# Results



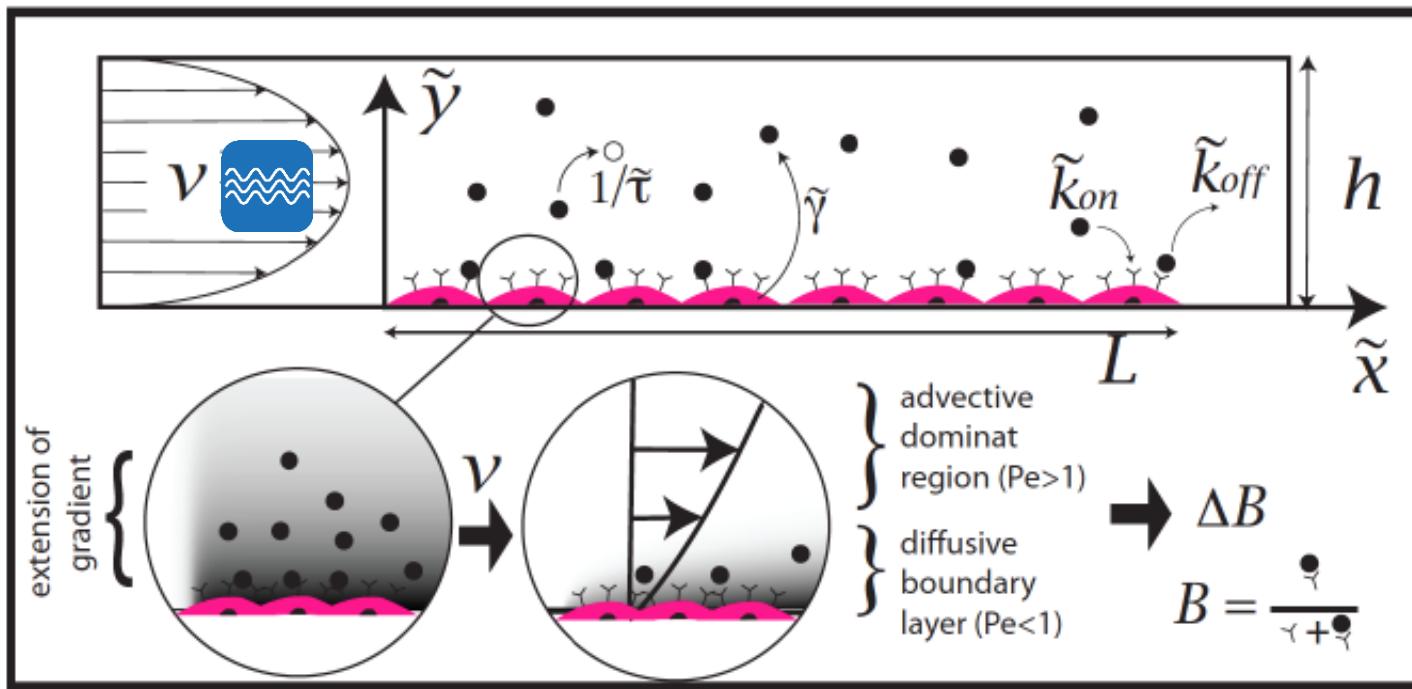
Micromotion-induced fluid flow in granulation tissue extends 1  $\mu\text{m/s}$  to 700  $\mu\text{m/s}$



Study the effects of  
fluid on morphogens  
distribution and  
MSCs differentiation



# Model



$$B = \frac{\text{surface density of } bound \text{ receptors}}{\text{surface density of receptors}}$$



# Governing Equations

## Mass transport

Diffusion (D) – Advection (A) – Reaction (R) in bulk:

$$\frac{\partial}{\partial t} C = \text{D} - \text{A} - \text{R}$$

D     
 A     
 R

Binding (B) – Unbinding (UB) reaction on the wall :

$$\frac{\partial B}{\partial t} = \text{B} - \text{UB}$$

B     
 UB

Secretion(S) of morphogens:

$$\gamma - \frac{\partial B}{\partial t} = -D \frac{\partial}{\partial y} C \Big|_{y=0}$$

S

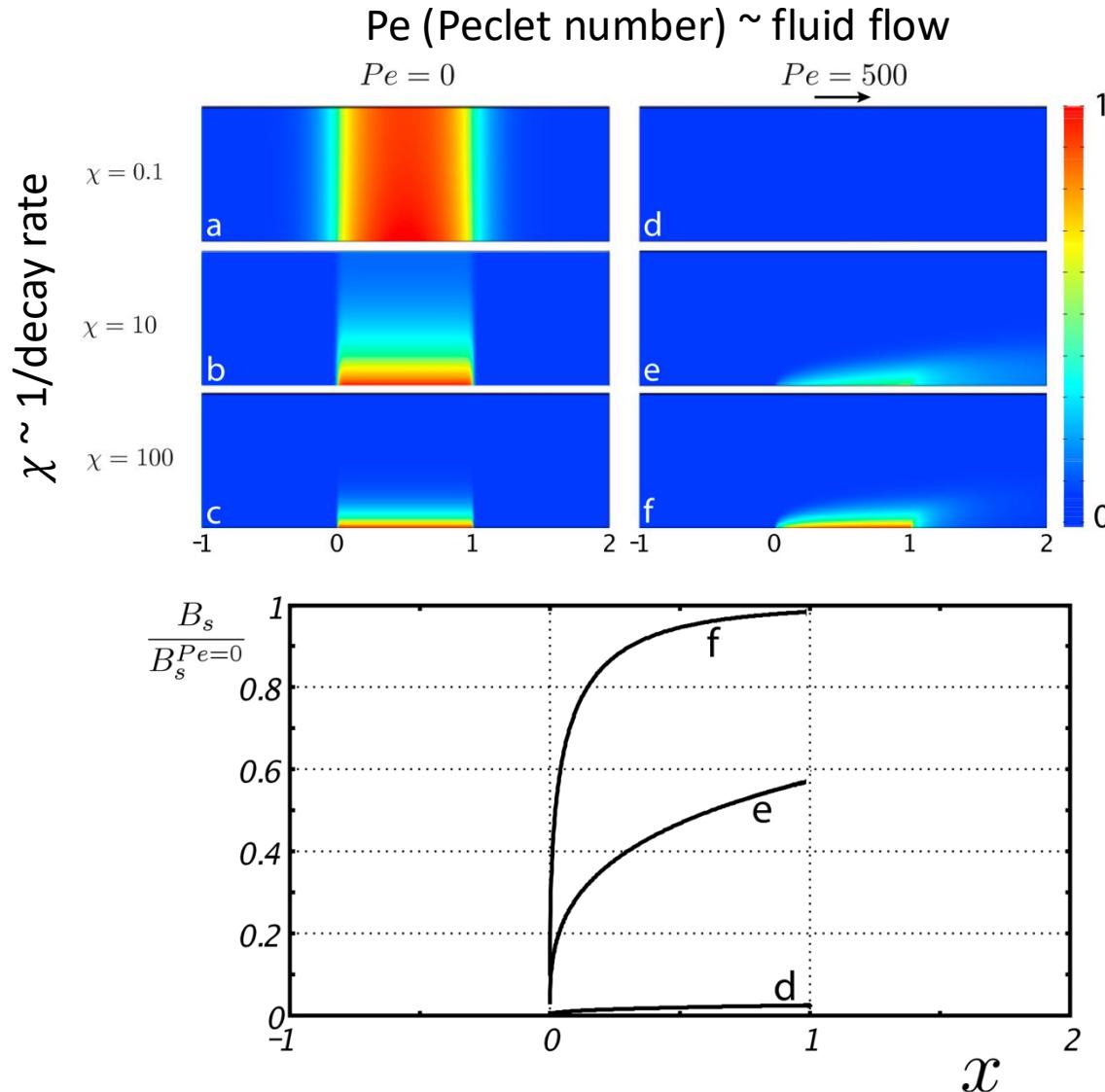
C - concentration

$R_t$  – total number of receptors

B – number of bound receptors



# Results



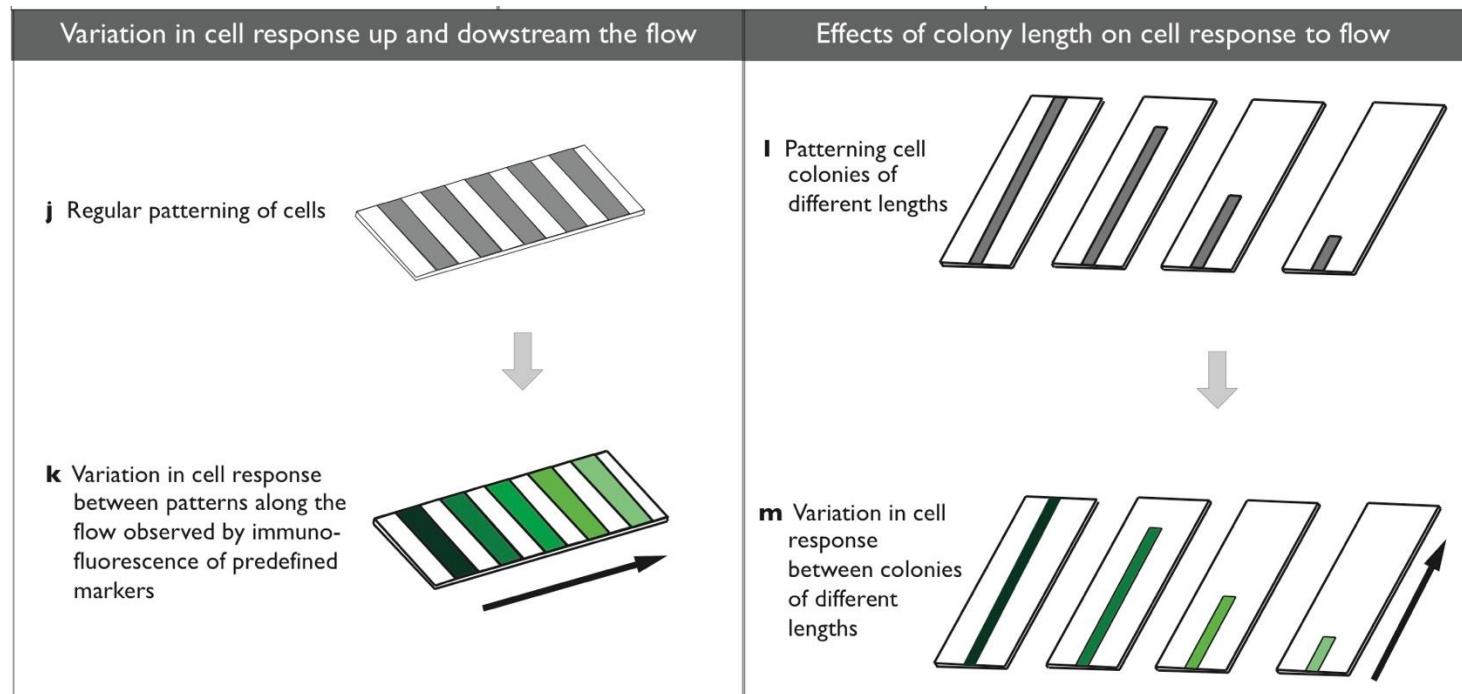
Fluid flow is strong enough to:

- Disturb the concentration profile of morphogens
- Change the number of bound receptors (i.e. have an effect on cell differentiation)



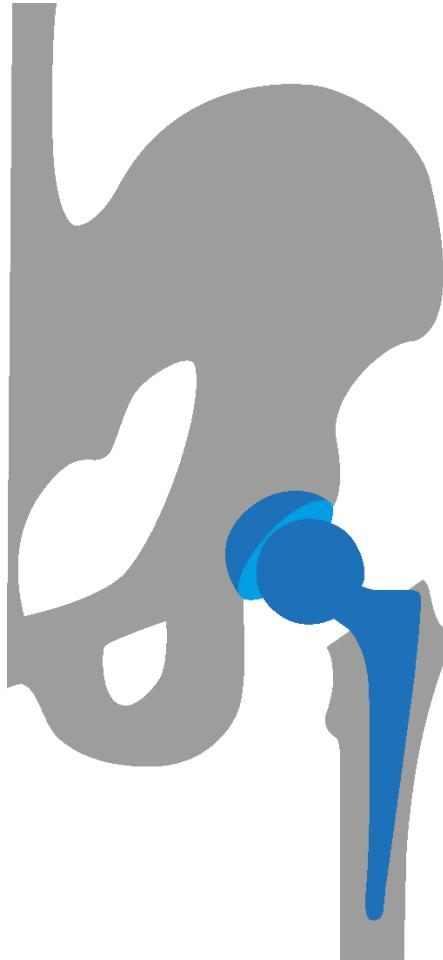
# Experimental Validation

## Microfluidics experiments





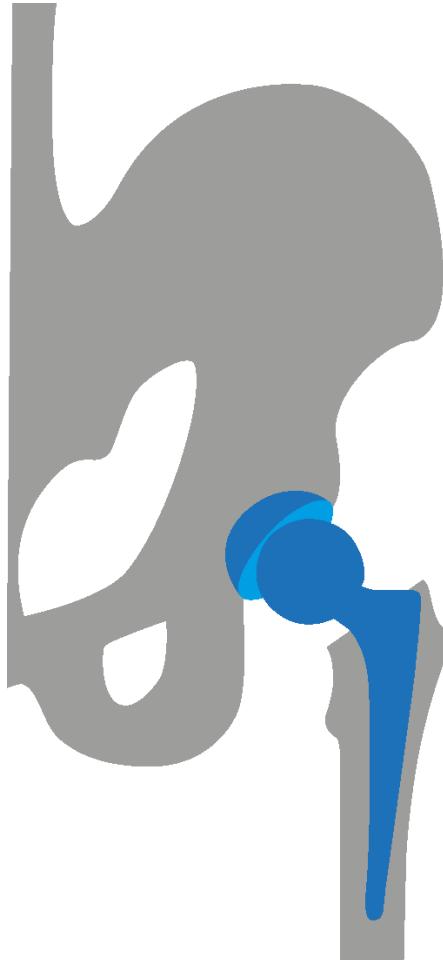
# Conclusion



A better understanding of conditions promoting aseptic loosening of hip implants can lead to better implant designs or surgical techniques and benefit patients



# Conclusion



Numerical modeling helps to investigate complex multi-scale hypotheses.

However, experimental validation is essential to assess the predictive capabilities of models.