



Master in Electrical and Electronics Engineering

EE-517: Bio-Nano-Chip Design

Lecture #3

Probe/Target interactions

Lecture Outline

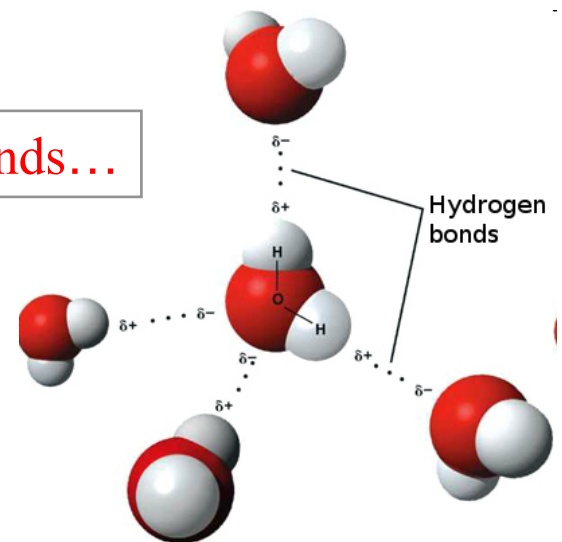
(Book Bio/CMOS: Chapter' paragraphs §4.4-16)

- DNA/RNA pairing & Bond energy
- Antibody/Antigen affinity & Bond energy
- Enzymes & Michaelis-Menten Kinetics

DNA Base pairing

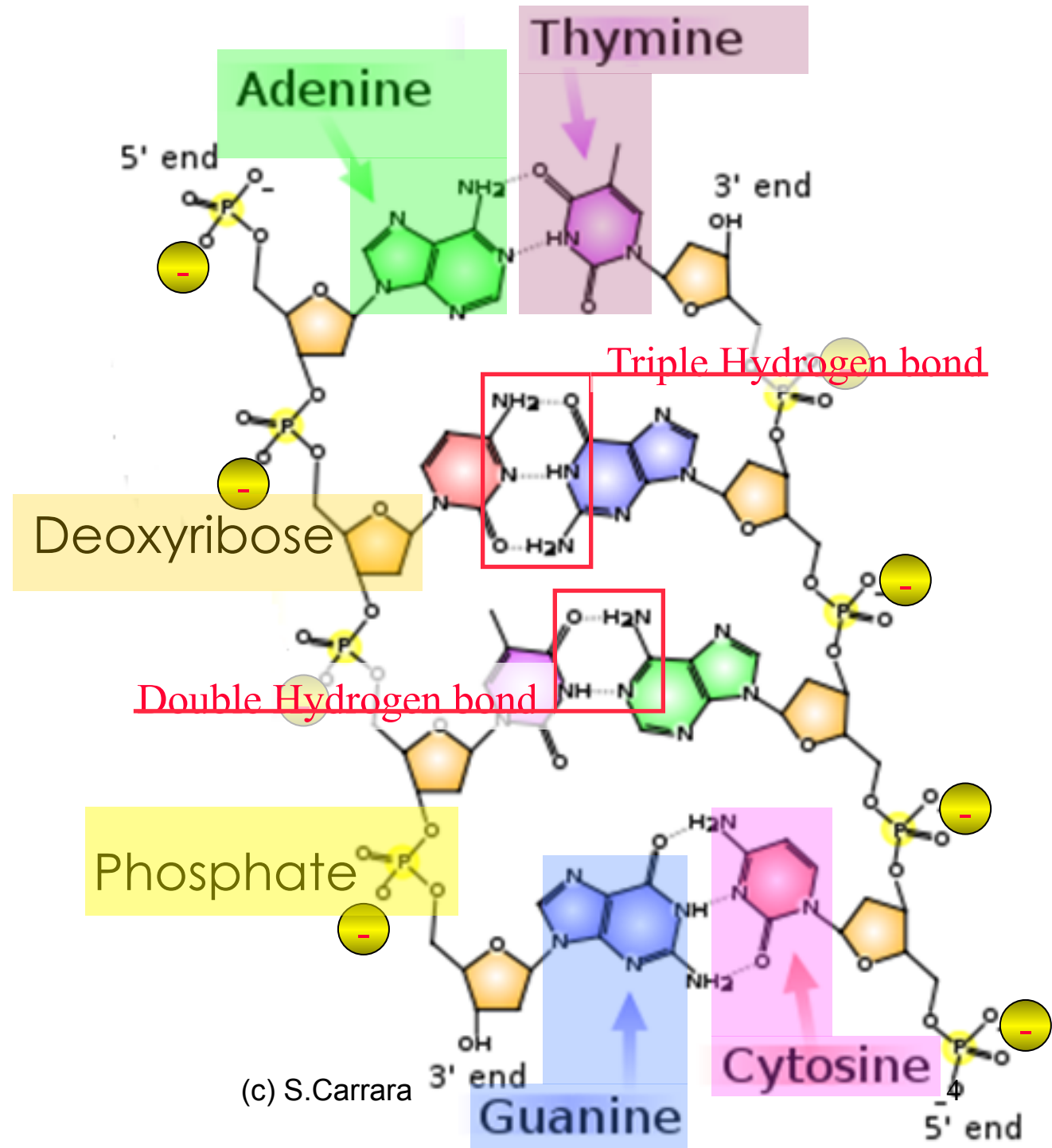
Each type of base on one strand forms a bond with just one type of base on the other strand. This is called **complementary base pairing**.

Thanks to Hydrogen Bonds...



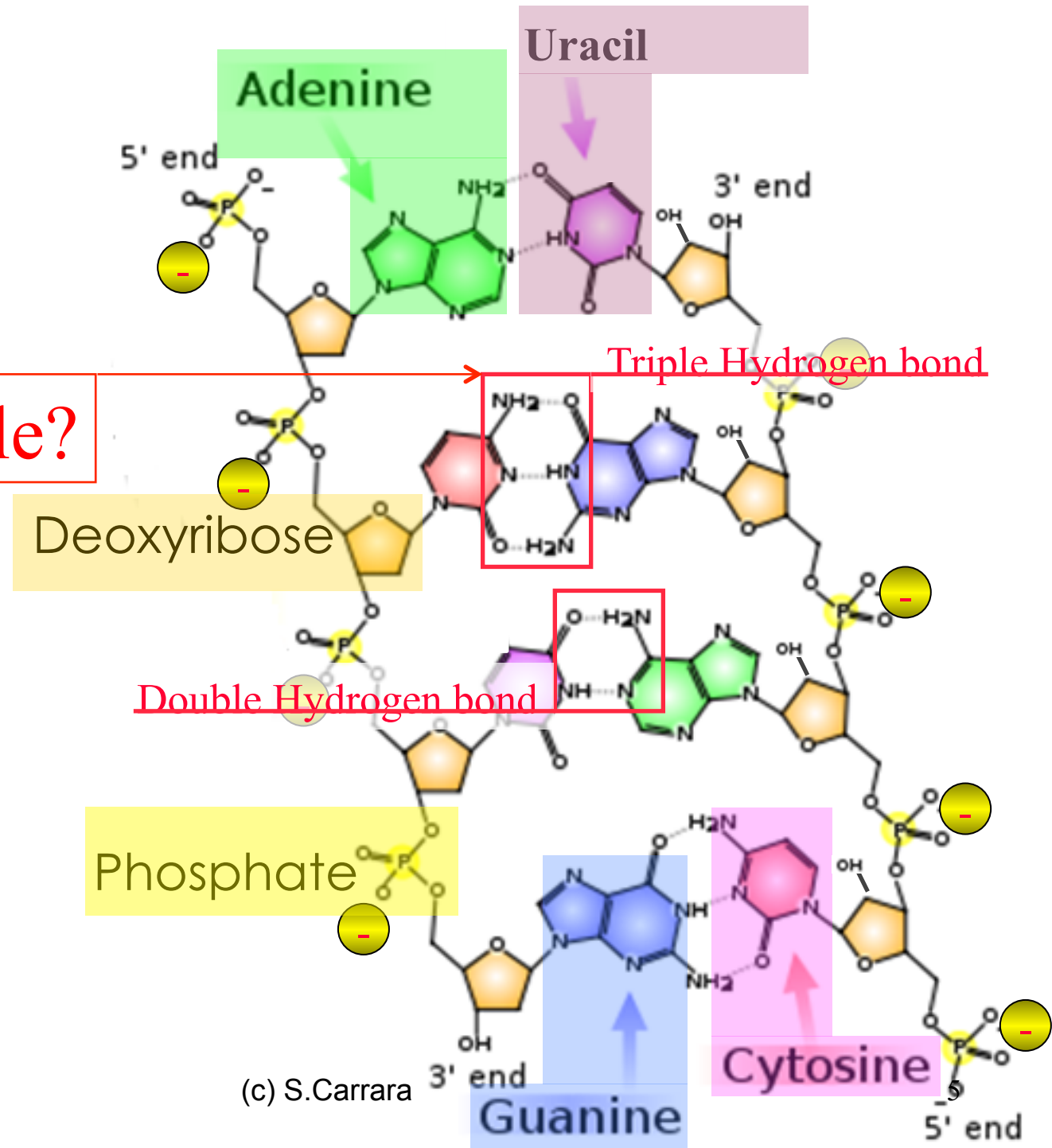
(c) S.Carrara

DNA-DNA Interaction



DNA-RNA Interaction

More stable?



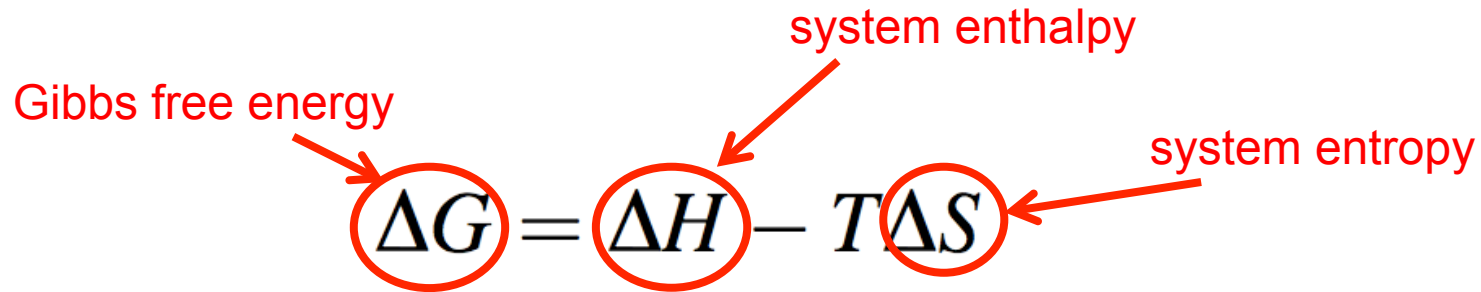
DNA/RNA pairing

Several interactions play a role in DNA-DNA and in DNA-RNA interactions. They include those between adjacent bases, those between electrons on the upper and lower parts of the base rings, and those between the phosphate groups in the nucleic acid backbone and the polar water molecules and other ions in the solution that interact directly with the paired molecule.

DNA/RNA pairing

Several interactions play a role in DNA-DNA and in DNA-RNA interactions. They include those between adjacent bases, those between electrons on the upper and lower parts of the base rings, and those between the phosphate groups in the nucleic acid backbone and the polar water molecules and other ions in the solution that interact directly with the paired molecule. **Entropic forces also play a role**

Gibbs free energy



The diagram shows the equation $\Delta G = \Delta H - T\Delta S$. Red circles are drawn around ΔG , ΔH , and $T\Delta S$. Red arrows point from text labels to these circled terms: 'Gibbs free energy' points to ΔG , 'system enthalpy' points to ΔH , and 'system entropy' points to $T\Delta S$.

$$\Delta G = \Delta H - T\Delta S$$

The Gibb free energy is the upper limit (some times called thermodynamic potential) of the total work obtainable by the system in processes at constant temperature and pressure

The enthalpy is related to variations in **stacking energy** related to bond strength, which includes but is not limited to **hydrogen bonds** strength

The entropy is kind of measure of the system disorder and $T\Delta S$ is the energy generated/adsorbed by the system in the conformational change

Gibbs free energy

The Gibb free energy is the upper limit (some times called thermodynamic potential) of the total work obtainable by the system in processes at constant temperature and pressure

$$\Delta G = \Delta H - T\Delta S$$

The first law of thermodynamics is the law of the energy conservation for the thermodynamic systems

$$\Delta U = Q - W.$$

Enthalpy & Entropy for different DNA dimers

| DNA dimer | ΔH [kJ/mol] | ΔS [kJ/(mol K)] |
|---|---------------------|-------------------------|
| $ \begin{array}{c} 5' \uparrow \text{A-T} \beta' \\ \quad \\ 3' \downarrow \text{T-A} \downarrow 5' \end{array} $ | -30.2 | -85.4 |
| $ \begin{array}{c} \uparrow \text{A-T} \\ \quad \\ \text{A-T} \downarrow \end{array} \text{ or } \begin{array}{c} \uparrow \text{T-A} \\ \quad \\ \text{T-A} \downarrow \end{array} $ | -33.1 | -92.9 |
| $ \begin{array}{c} \uparrow \text{T-A} \\ \quad \\ \text{A-T} \downarrow \end{array} $ | -30.2 | -89.2 |
| $ \begin{array}{c} \uparrow \text{G-C} \\ \quad \\ \text{C-G} \downarrow \end{array} $ | -41.0 | -102.1 |
| $ \begin{array}{c} \uparrow \text{G-C} \\ \quad \\ \text{G-C} \downarrow \end{array} \text{ or } \begin{array}{c} \uparrow \text{C-G} \\ \quad \\ \text{C-G} \downarrow \end{array} $ | -33.5 | -83.3 |
| $ \begin{array}{c} \uparrow \text{C-G} \\ \quad \\ \text{G-C} \downarrow \end{array} $ | -41.9 | -113.9 |

The Nearest-Neighbor Model

number of occurrences of each nearest neighbor

account for the initial pair

$$\Delta G = \sum_{\forall i} n_i \Delta G_i + \Delta G_i|_{init\ term\ A-T} + \Delta G_i|_{init\ term\ G-C} + \Delta G_{system}$$

Gibbs free energy change for the considered neighboring pairs

empirical parameter (+1.80 kJ/mol if the nucleic acid duplex is self-complementary, or equal to zero if not)

It introduces a computation for the total Gibbs free energy that takes into account the interactions from neighbours

Gibbs free energy

| duplex | <i>Experimental</i> ΔG [kJ/mol] |
|--------------------------|--|
| GGTTATTGG CCAATAACC | -26.8 |
| GGTTCCTTGG CCAAGAACC | -31.4 |
| GGTTTTTTGG CCAAAAAACC | -29.5 |
| GGTTATTGG CCAAAACC | -12.0 |
| GGTTCCTTGG CCAATAACC | -12.4 |
| GGTTTTTTGG CCAAGGAACC | -17.5 |

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Role of Solvent

$$\Delta G = \Delta G_{\text{double strand}}^{\text{intramol}} + \Delta G_{\text{double strand}}^{\text{solvent}} + \Delta G_{\text{single strand}}^{\text{solvent}}$$

solvation free energy of the double helix

intermolecular nucleic double strand acid interactions

solvation free energy of the single strands

The diagram shows the equation $\Delta G = \Delta G_{\text{double strand}}^{\text{intramol}} + \Delta G_{\text{double strand}}^{\text{solvent}} + \Delta G_{\text{single strand}}^{\text{solvent}}$. Three terms are circled in red. Red arrows point from descriptive text to each term: 'intermolecular nucleic double strand acid interactions' points to the first term, 'solvation free energy of the double helix' points to the second term, and 'solvation free energy of the single strands' points to the third term.

The solvent also plays a role in DNA/DNA and DNA/RNA pairing, thus, it contributes to the Gibbs free energy too

Role of Solvent

$$\Delta G = \Delta G_{double\ strand}^{intramol} + \Delta G_{double\ strand}^{solvent} + \Delta G_{single\ strand}^{solvent}$$

DNA/RNA hydrogen bonds interactions

$$\Delta G_{double\ strand}^{intramol} = \Delta G_{DNA-RNA}^{hydrogen\ bonds} + \Delta G_{DNA-RNA}^{Coulomb} + \Delta G_{DNA-RNA}^{van\ der\ Waals}$$

Coulomb interactions between the phosphate groups of the backbone and the other ions in the solution

van der Waals interactions

between the rings on the two sides of the duplex.

$$\Delta G_{single\ strand}^{solvent} = \Delta G_{DNA}^{solvent} + \Delta G_{RNA}^{solvent}$$

interactions between the different single strands and the solvent

DNA Denaturation

Melting of the DNA helix is the thermodynamic process that unwind the double helix and interrupts the non-covalent and sequence-dependent interactions between two single strands. The denaturation of DNA is another name for the same process.

The term **melting** is usually accomplished by heating the solutions containing the DNA. The word **denaturation** is more often used when the unwinding and opening of the double helix are performed by means of a pH change or by means of a change in ion content in the solution.

Melting Temperature

By definition, the maximum of double-helix unfolding and interaction interruption happens at the melting temperature.

Therefore, it is the moment of transformation where the enthalpy supplied by heating contributes to nucleic acid melting. Therefore, the Gibbs free energy is null at this temperature:

$$\Delta G = \Delta H - T\Delta S = 0.$$

The variation in the entropy starts from the system status where all the nucleic acids are in double-helix conformation:

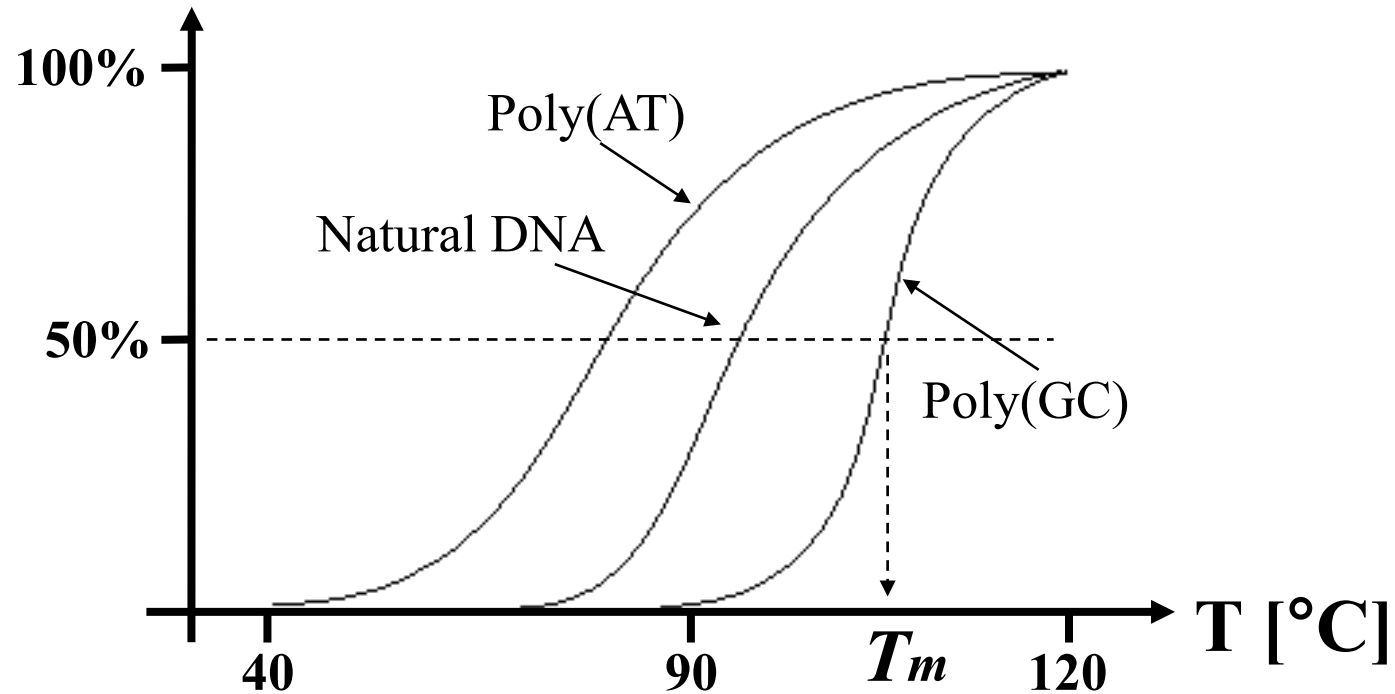
$$S_0 = R \ln(C_{DNA})$$

Thus, the melting temperature is:

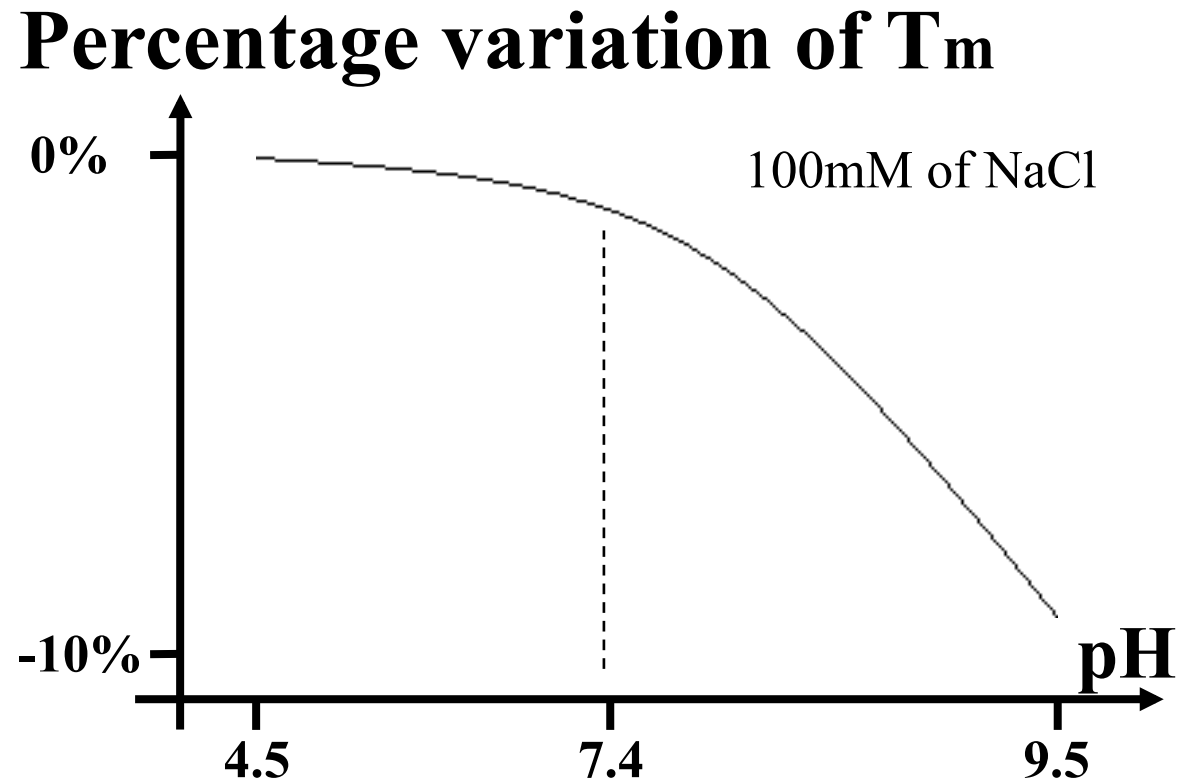
$$T_m = \frac{\Delta H}{\Delta S + R \ln C_{DNA}}$$

Melting processes for different DNA sequences

Percentage of double helix



Melting temperature of DNA versus pH



Antibody/Antigen interactions

Antibodies are a large class of y-shaped proteins called immunoglobulins. An antibody shows a Y-structure with three main protein regions: a constant region and two variable regions. The variable regions are suitable for blocking the antigens. The protein blocks the antigen by closing the b-sheets in the variable region like fingers of a human hand when grasping

An efficient trapping is assured by the perfect match between the steric shape of the antigen and the 3D conformation of the antibody. The right part of the antigen in direct contact with the antibody is called the **epitope**, while the right part of the antibody in contact with the antigen is called the **paratope**

**CRYSTAL STRUCTURE OF
THE INTACT HUMAN IGG
B12 WITH BROAD AND
POTENT ACTIVITY
AGAINST PRIMARY HIV-1
ISOLATES**

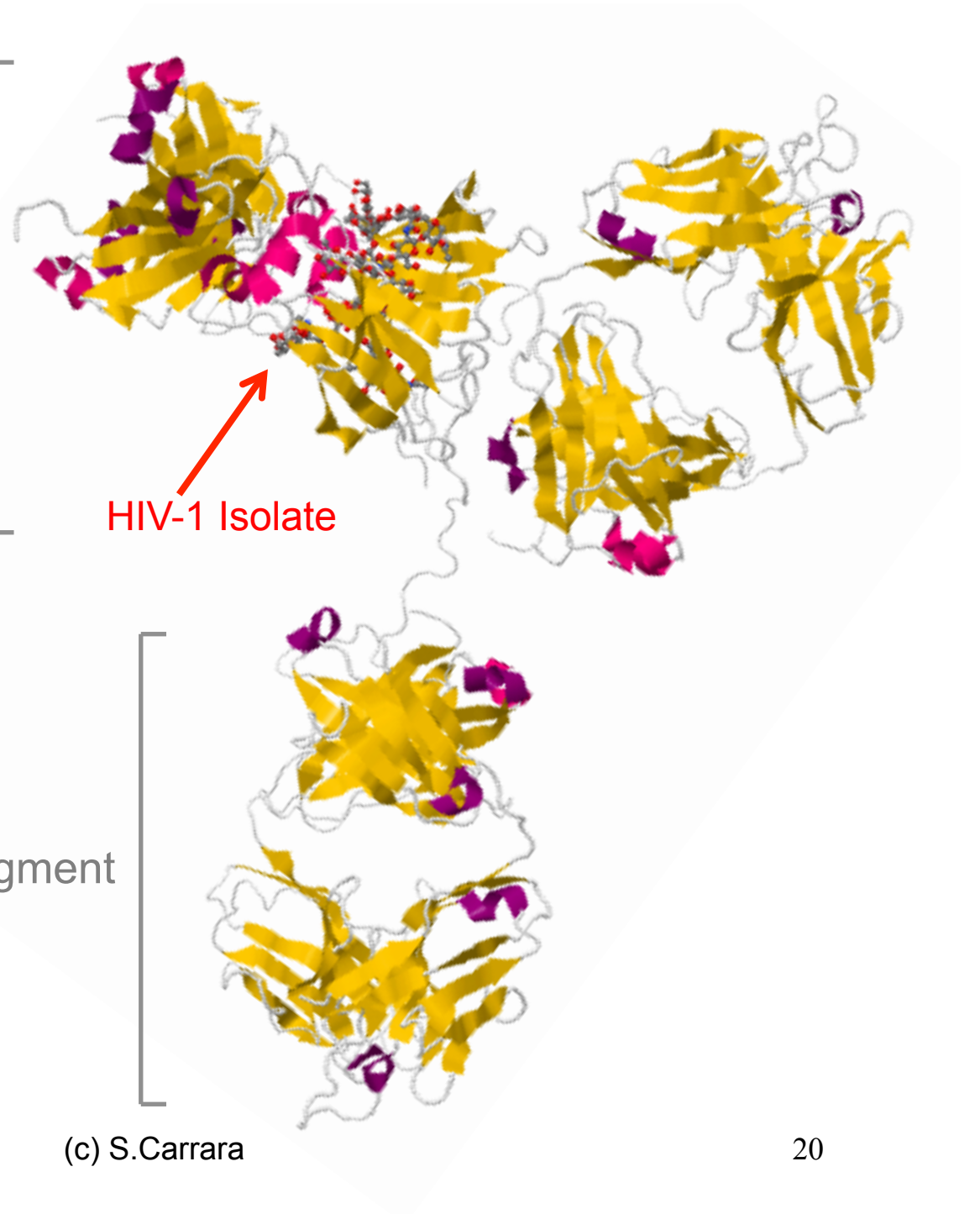
Variable Fragments

HIV-1 Isolate

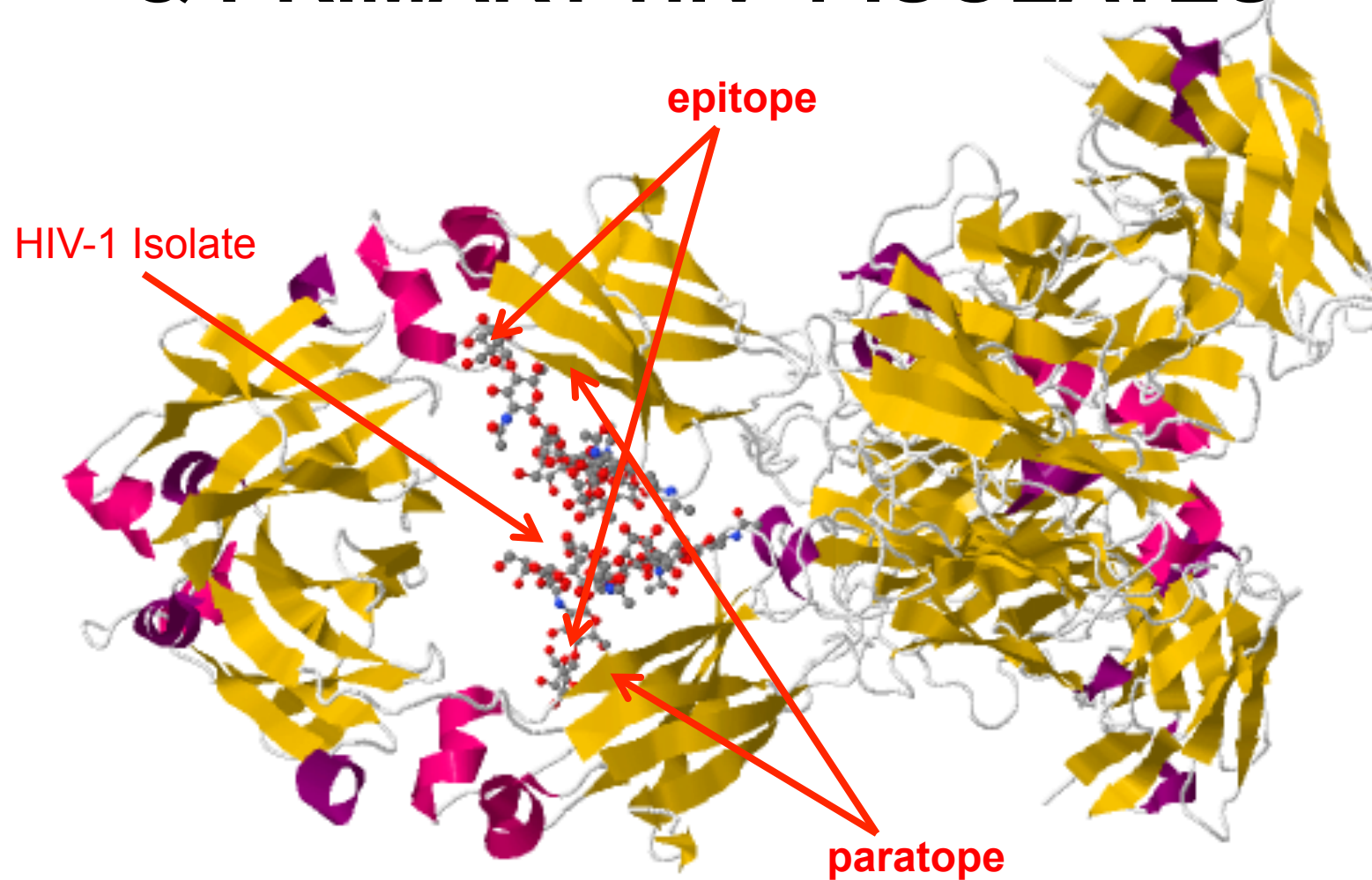
Constant Fragment

[Lateral View]

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CRYSTAL HUMAN IGG B12 & PRIMARY HIV-1 ISOLATES

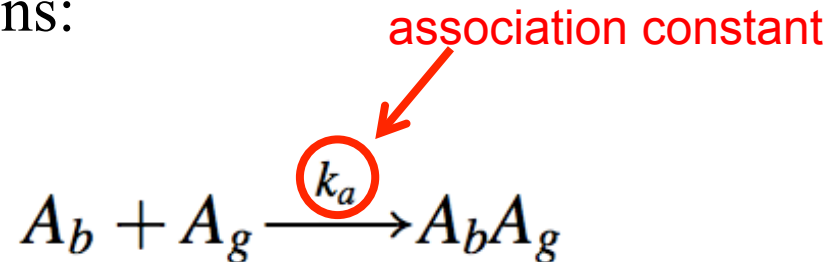


[Top View]

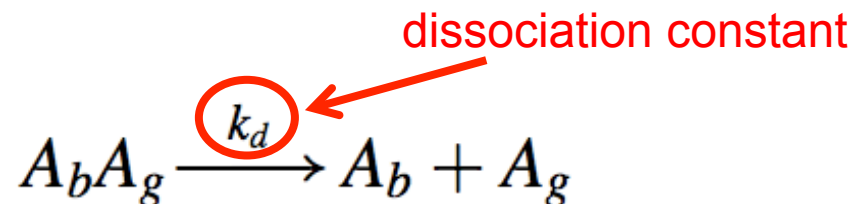
(c) S.Carrara

Uptake Process

The uptake of antigens is a process of association among antibodies and antigens:



That also has the reverse process of dissociation:



Both contributing to the net rate of formation of the complexes:

$$\frac{d[A_b A_g]}{dt} = k_a [A_b] [A_g] - k_d [A_b A_g]$$

Uptake Process

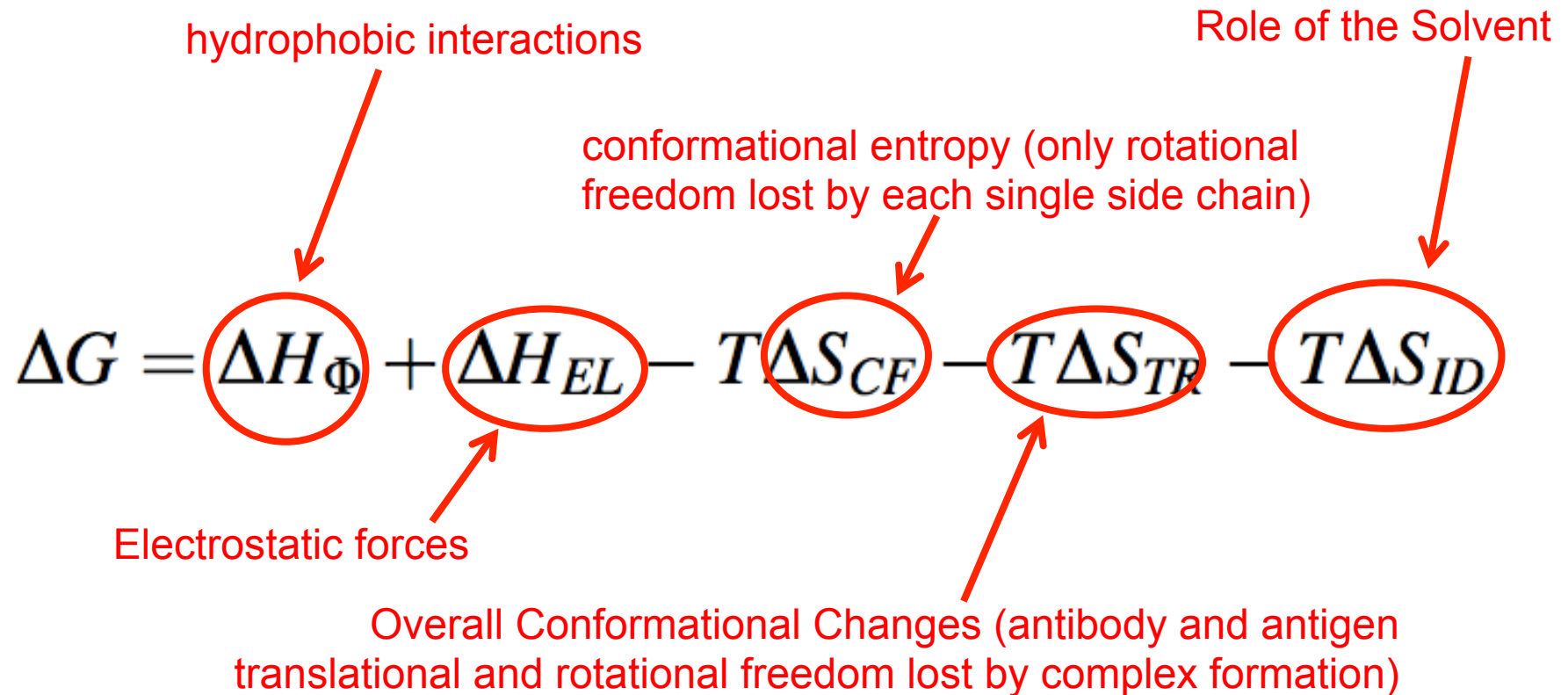
At equilibrium, we have: $\frac{d[A_bA_g]}{dt} = 0$

That defines the affinity constant: $K = \frac{k_a}{k_d} = \frac{[A_bA_g]}{[A_b][A_g]}$

Which contributes to the Gibbs free energy:

$$\Delta G = -RT \ln \left(\frac{[A_bA_g]}{[A_b][A_g]} \right)$$

Gibbs free energy contributions



The diagram shows the equation $\Delta G = \Delta H_{\Phi} + \Delta H_{EL} - T\Delta S_{CF} - T\Delta S_{TR} - T\Delta S_{ID}$ with five terms circled in red. Red arrows point from descriptive text to each circled term: 'hydrophobic interactions' to ΔH_{Φ} , 'Electrostatic forces' to ΔH_{EL} , 'conformational entropy (only rotational freedom lost by each single side chain)' to $T\Delta S_{CF}$, 'Overall Conformational Changes (antibody and antigen translational and rotational freedom lost by complex formation)' to $T\Delta S_{TR}$, and 'Role of the Solvent' to $T\Delta S_{ID}$.

$$\Delta G = \Delta H_{\Phi} + \Delta H_{EL} - T\Delta S_{CF} - T\Delta S_{TR} - T\Delta S_{ID}$$

hydrophobic interactions

conformational entropy (only rotational freedom lost by each single side chain)

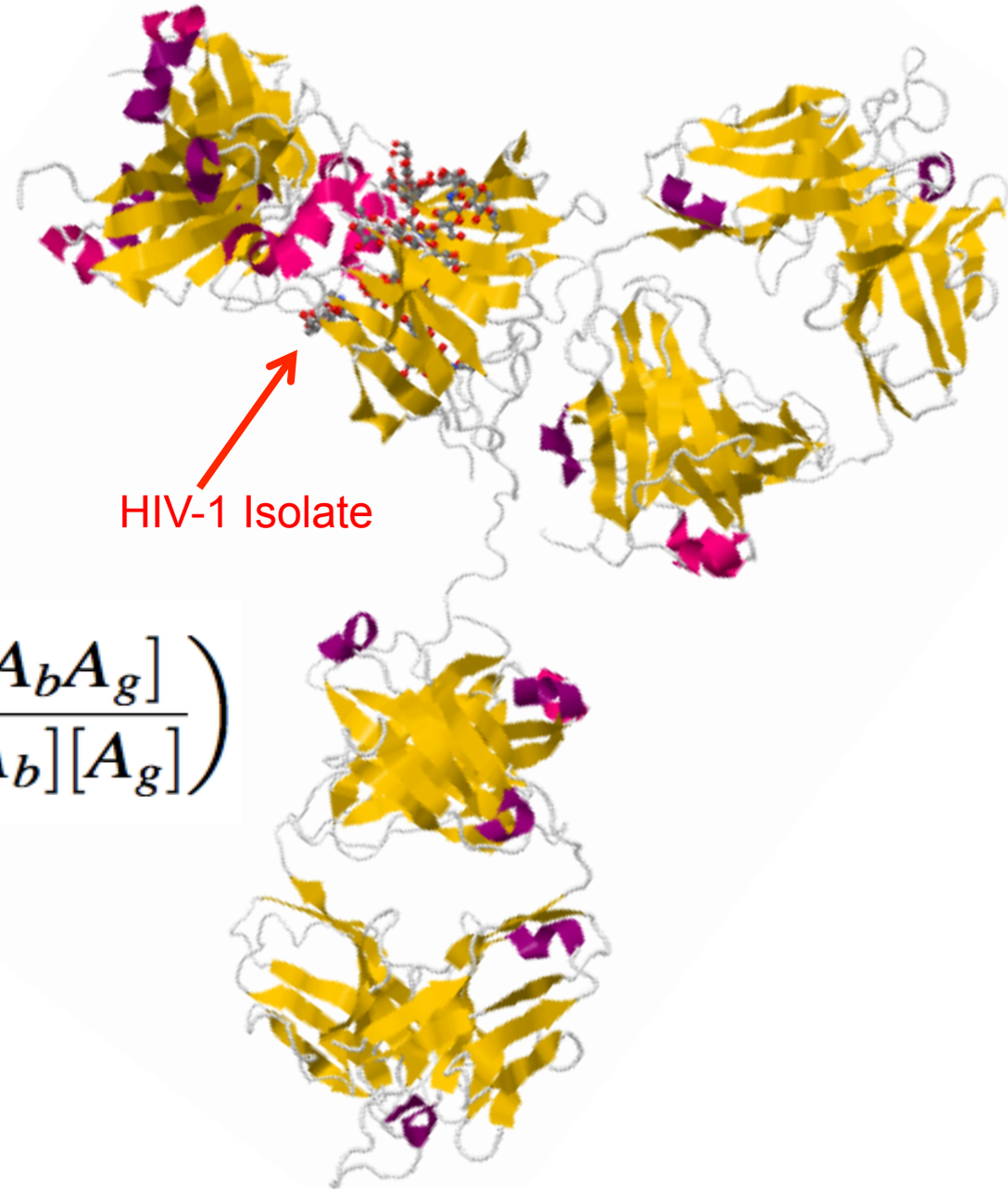
Role of the Solvent

Electrostatic forces

Overall Conformational Changes (antibody and antigen translational and rotational freedom lost by complex formation)

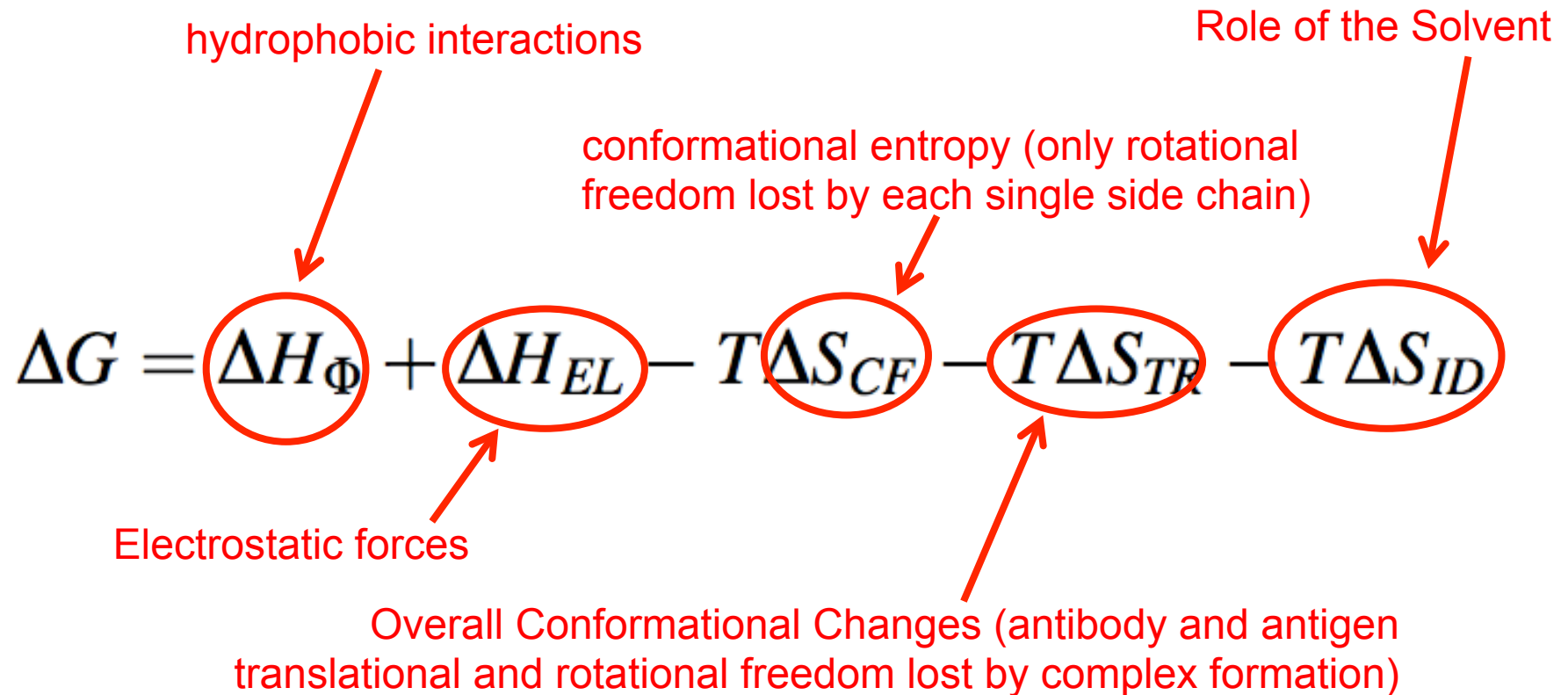
The total Gibbs free energy is related to various sources of molecular interactions

**CRYSTAL STRUCTURE OF THE
INTACT HUMAN IGG B12 WITH
BROAD AND POTENT ACTIVITY
AGAINST PRIMARY HIV-1
ISOLATES**



$$\Delta G = -RT \ln \left(\frac{[A_b A_g]}{[A_b][A_g]} \right)$$

Gibbs free energy contributions



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hydrophobic interactions

conformational entropy (only rotational freedom lost by each single side chain)

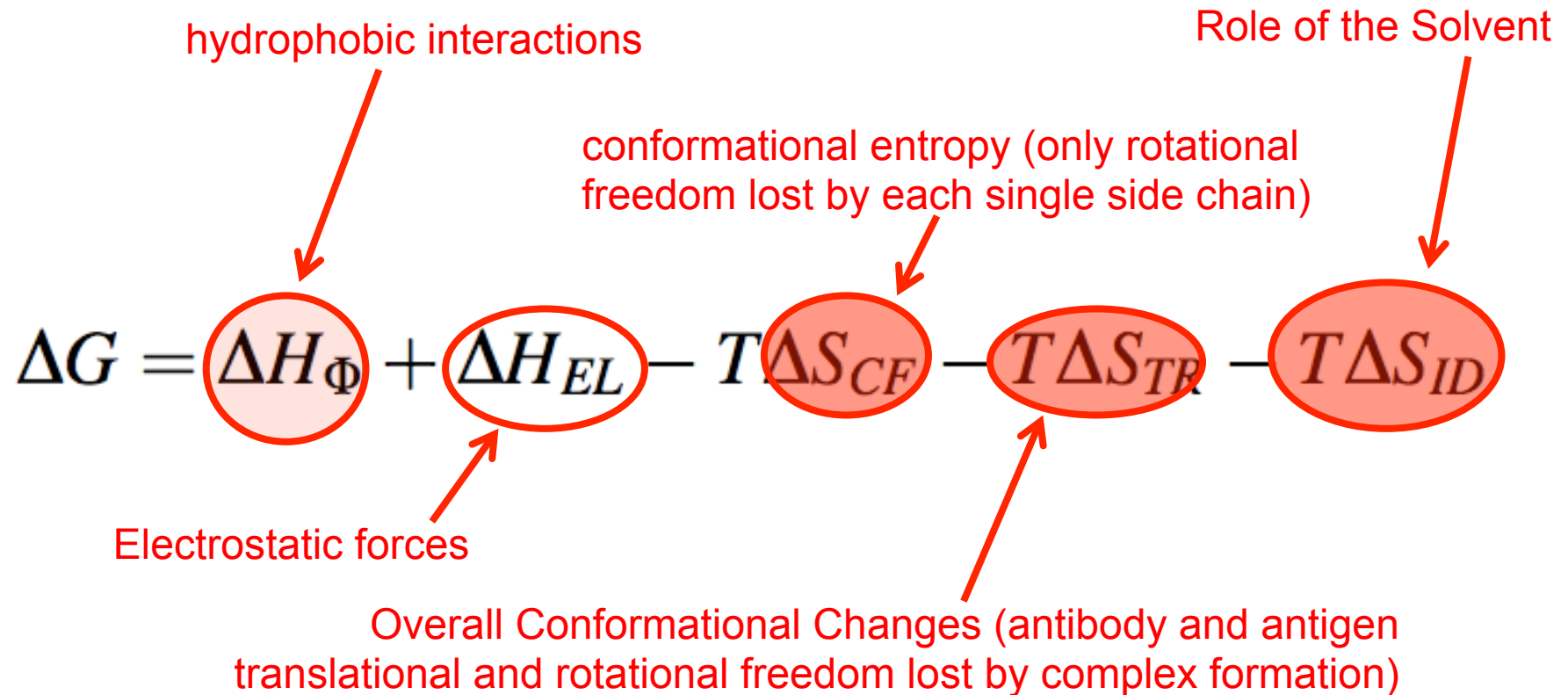
Role of the Solvent

Electrostatic forces

Overall Conformational Changes (antibody and antigen translational and rotational freedom lost by complex formation)

The total Gibbs free energy in antibody/antigen uptake is related to various sources of molecular interactions

Gibbs free energy contributions



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$$\Delta G = \Delta H_{\Phi} + \Delta H_{EL} - T\Delta S_{CF} - T\Delta S_{TR} - T\Delta S_{ID}$$

hydrophobic interactions

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Role of the Solvent

Electrostatic forces

Overall Conformational Changes (antibody and antigen translational and rotational freedom lost by complex formation)

The total Gibbs free energy in antibody/antigen uptake is related to various sources of molecular interactions

Hydrophobic Interactions

First antibody/antigen interaction is based on hydrophobic interactions the epitope and the paratope:

$$\Delta H_{\Phi} = \alpha A_{contact}$$

The coefficient of proportionality in the previous equation is empirically determined in:

$$\alpha = -104.5 \frac{kJ}{mol \text{ \AA}^2}$$

The typical contact area of an antibody–antigen interaction ranges from 150 to 690 Å². Thus, the hydrophobic energy ranges from 15.7 kJ/mol to 72.1 kJ/mol

Electrostatic Interactions

van der Waals forces and hydrogen bonds contribute to the electrostatic interactions:

$$\Delta H_{EL} = - \sum_{\forall i,j} \frac{q_i q_j}{16\pi r_{i,j}}$$

This electrostatic enthalpy is usually in the range of 88 to 163 kJ/mol for antibody/antigen interactions.

Conformational Changes of protein residues

Conformational changes due to rearrangements in the 3D organization of protein residues both in the epitope and in the paratope contribute to the entropy of the system. The torsional degrees of freedom are three in 3D space and, thus, we have for N side chains that lost their rotational freedom:

$$T\Delta S_{CF} = -RT \ln(3^N) = -NRT \ln(3)$$

This conformational entropy returns a value of 2.7 kJ/mol for each side chain.

Other contributions

The overall translational and rotational freedom that was partially blocked by the complex formation is usually empirically estimated in:

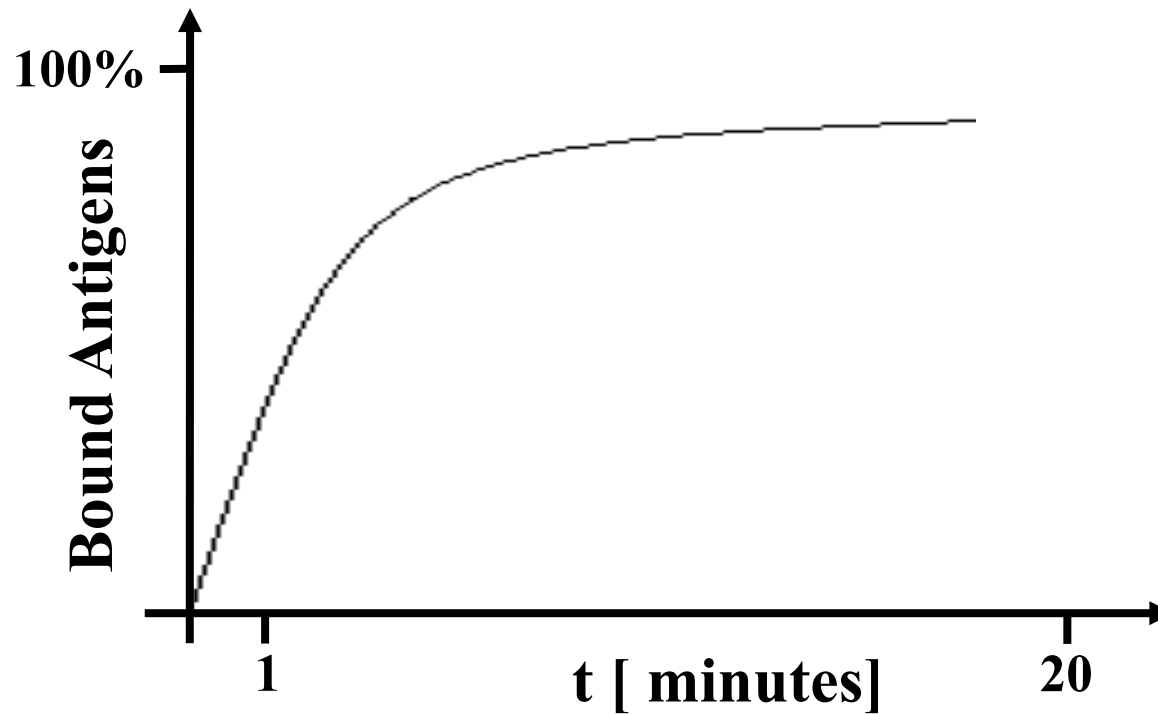
$$-T\Delta S_{TR}^{Empirical} = 29 \div 46 \text{ kJ/mol}$$

While the Role of the Solvent may be roughly estimated by considering an approximation to infinite dilutions:

$$T\Delta S_{ID} \approx -8.4 \text{ kJ/mol}$$

So, the typical **total Gibbs free energy** of a complex antibody–antigen interaction is in a range of **27.6 to 60.3 kJ/mol**

Typical trend of antigen binding versus time



Different Kinds of Antibody

Dealing with real cases results in a bit more complex situation than just adding antigens to antibodies with a unique perfect match. **Monoclonal antibodies** are, then, all antibodies that have exactly the same specificity because they are from the same cloned single cell.

However, antibodies are in general secreted in blood plasma by cells that are from different cell lines. Therefore, it is easy to obtain antibodies that are all against the same antigen but that do not have exactly the same specificity: these are **polyclonal antibodies**. Different kinds of antibodies means different kinetic on the same antigen.

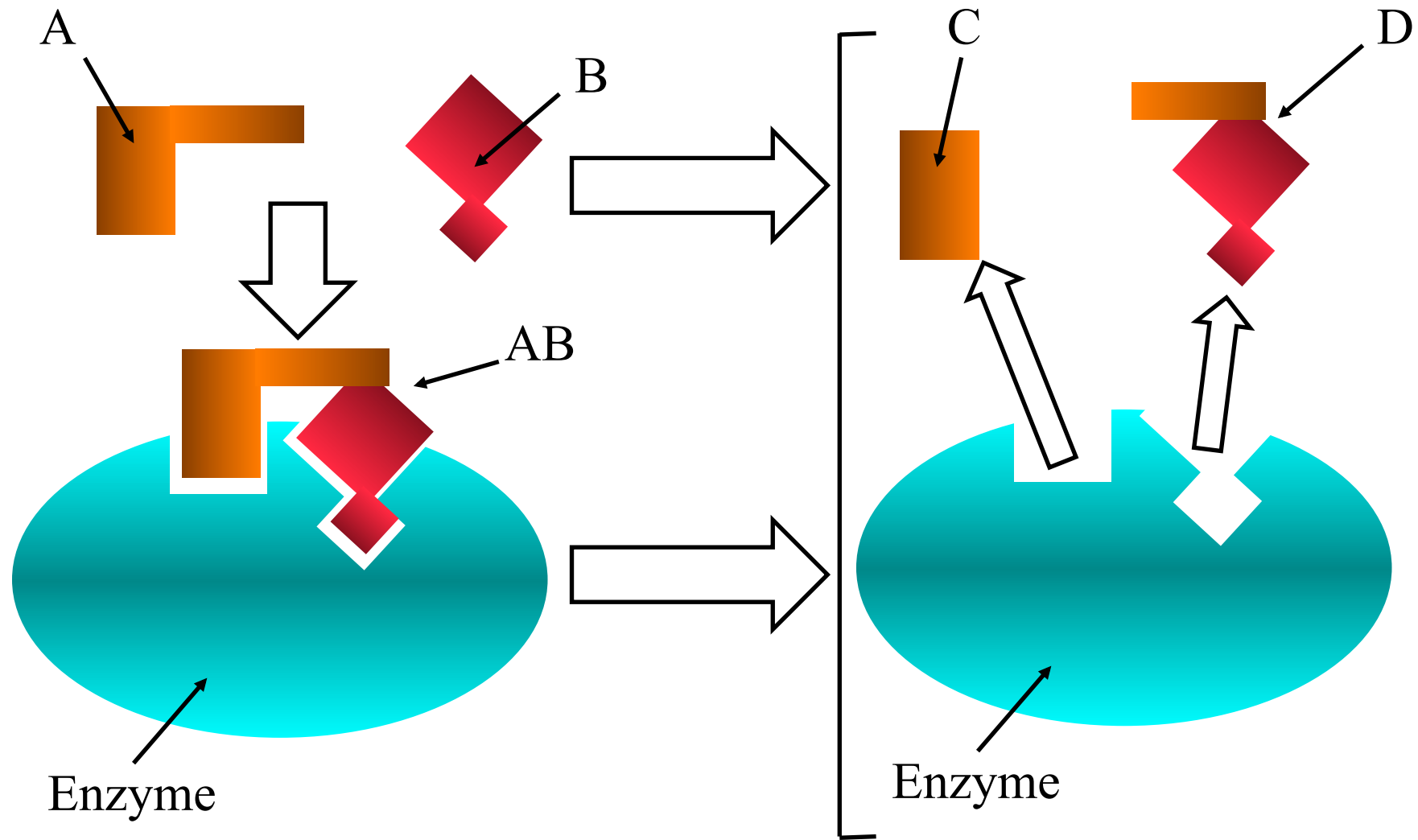
We then have different kinetics by involving **polyclonal antibodies** since they present **different paratopes** to address different epitopes of the same antigen

Enzyme/Substrate interactions

Enzymes are highly selective catalysts, accelerating both the rate and specificity of metabolic reactions. Almost all chemical reactions in a biological cell need enzymes in order to occur at a sufficient rates compatible with life. Most enzymes are proteins, although some catalytic RNA molecules have been identified.

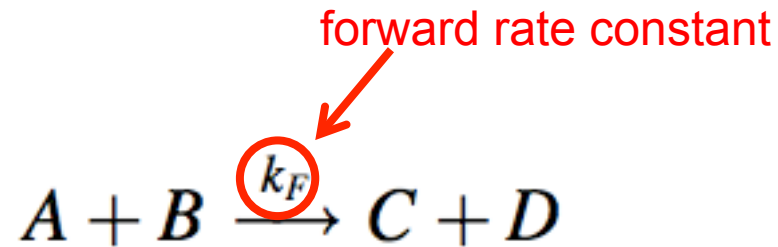
At the beginning of an enzymatic reactions, is called **substrates** the molecules which transformation is catalysed. This molecules are converted into different ones, called **products**.

Mechanism of Enzymatic reactions

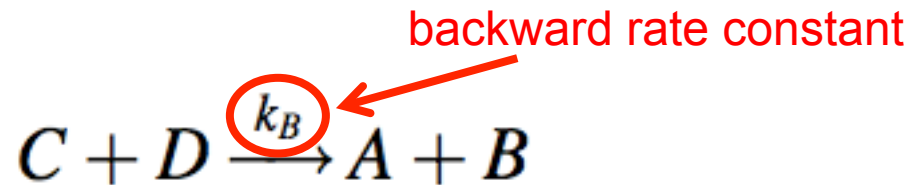


Equilibrium Constant

In any chemical reaction from two compounds to products, we have:



And we also have the reverse reaction as well:



Both defining the equilibrium constant of the overall reaction:

$$K = \frac{k_F}{k_B} = \frac{[C][D]}{[A][B]}$$

Enzymatic Process

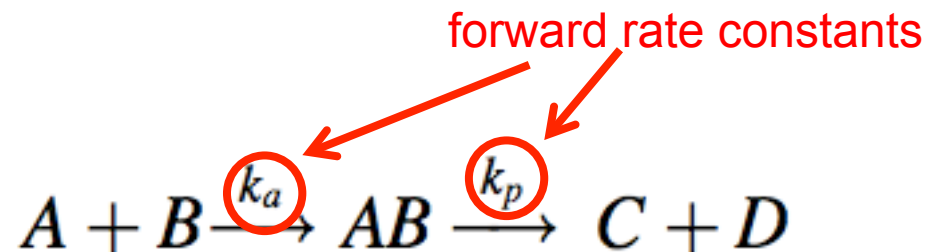
$$K = \frac{k_F}{k_B} = \frac{[C][D]}{[A][B]}$$

It is worth noting that the presence of an enzyme changes neither the equilibrium constant nor the two forward and backward rates

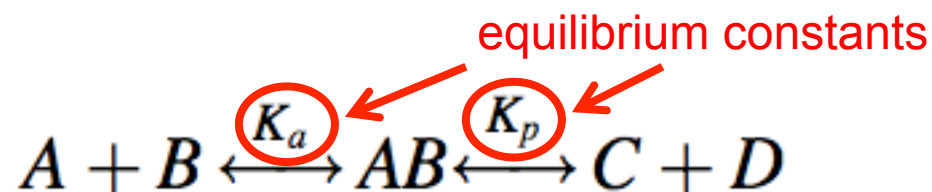
What the enzyme changes is the time within which the chemical reaction occurs. If the equilibrium is reached in, let say, 1 h in the absence of an enzyme, then it might be reached in few seconds in the presence of the suitable enzyme!

Enzymatic Process

In the presence of the enzyme, we have in the same chemical reaction :



And we also have the reverse reaction as well:



Thus, the rate of products' formation now depends by the formation of the complex:

$$\frac{d([C][D])}{dt} = k_p[AB]$$


Enzymatic Process

AB is obtained by A and B with the equilibrium constant K_a ., and thus the Gibbs free energy of the reaction is now related to the equilibrium constant of the formation of the intermediate state:

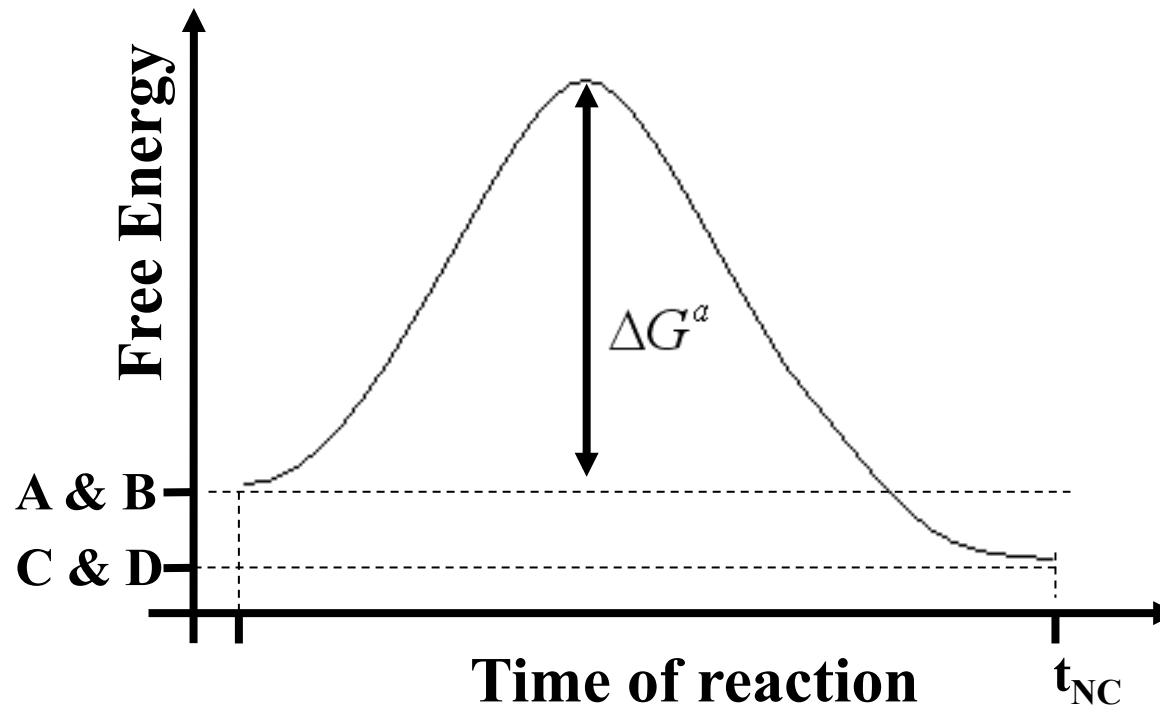
$$\Delta G^a = -RT \ln K_a$$

By neglecting the possibility for C and D to give back the intermediate state AB, we can now write the net rate of products formation as:

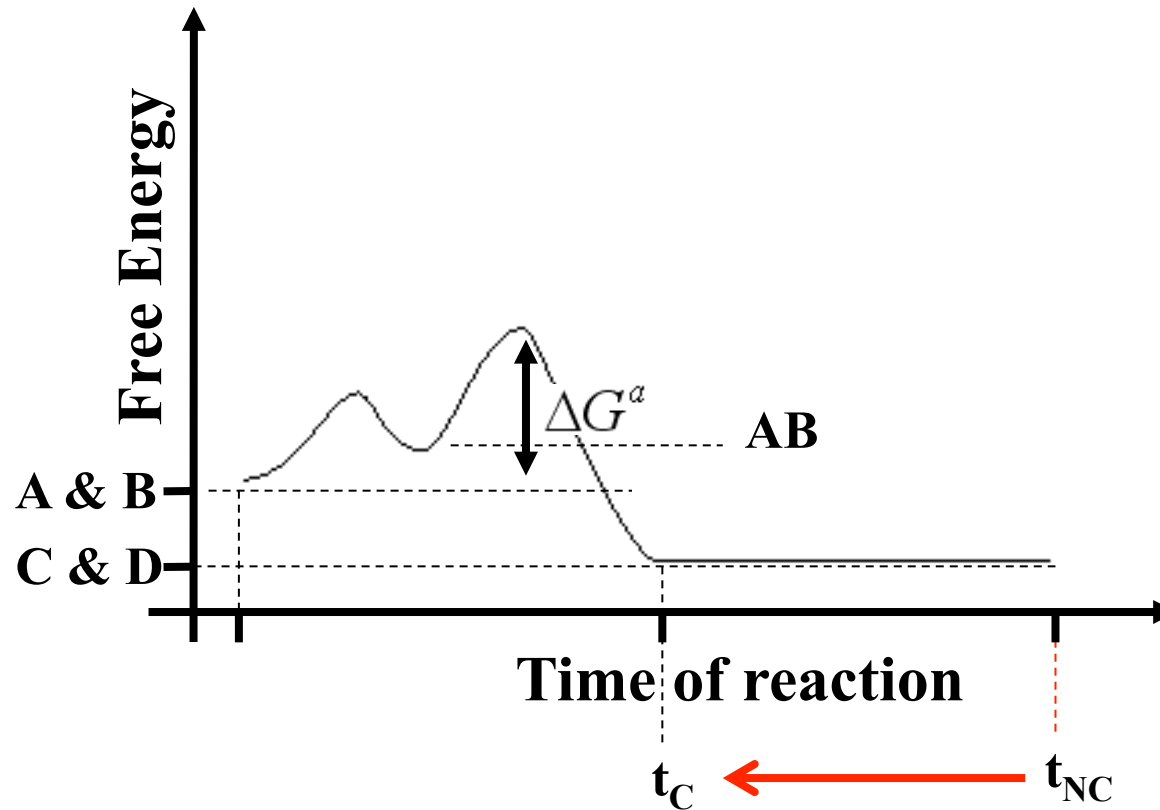
Gibbs free energy


$$\frac{d([C][D])}{dt} = k_p[AB] = k_p[A][B] e^{-\frac{\Delta G^a}{RT}}$$

Free energy of a reaction without enzymes

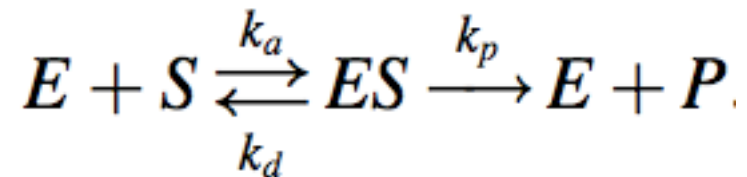


Free energy of a reaction with enzymes



Michaelis-Menten Kinetics

More often, the compound B is a common molecule, such as oxygen, or water, or proton, that is not specific to the enzyme action. Therefore, the enzymatic process is re-written in a form that explicitly introduce the presence of the enzyme:



By neglecting the possibility for E and P to give back the intermediate state AB, we can now write the net rate of product formation as:

$$\frac{d[P]}{dt} = k_p[ES]$$

Michaelis-Menten Kinetics

While the rate of complex formation:

$$\frac{d[ES]}{dt} = k_a[E][S]$$

And the rate of complex breakdown:

$$\frac{d([E][S])}{dt} + \frac{d[P]}{dt} = (k_d + k_p)[ES]$$

Need to be equal at the steady state of the enzymatic reaction:

$$k_a[E][S] = (k_d + k_p)[ES]$$

Michaelis-Menten Kinetics

Thus, the concentration of the complex is:

$$[ES] = \frac{k_a}{(k_d + k_p)} [E][S]$$

Which defines the so-called **Michaelis' constant**:

$$K_M = \left(\frac{k_a}{(k_d + k_p)} \right)^{-1}$$

The Michaelis constant is so important because it shows how efficient is the enzyme in forming the intermediate state that is required to catalyze the reaction.

Michaelis-Menten Kinetics

Usually, there is a certain amount of available enzymes that are still free to accept further substrates:

$$[E] = [E]_0 - [ES]$$

With $[E]_0$ as the initial concentration of total enzymes, which enables to re-write the concentration of the complex:

$$[ES] = \frac{([E]_0 - [ES])[S]}{K_M}$$

Or:

$$[ES] = [E]_0 \frac{[S]}{[S] + K_M}$$

Michaelis-Menten Kinetics

Now, by defining the reaction velocity V and the maximum reaction velocity V_m , respectively, as:

$$V = \frac{d[P]}{dt} \quad V_{\max} = k_p[E]_0$$

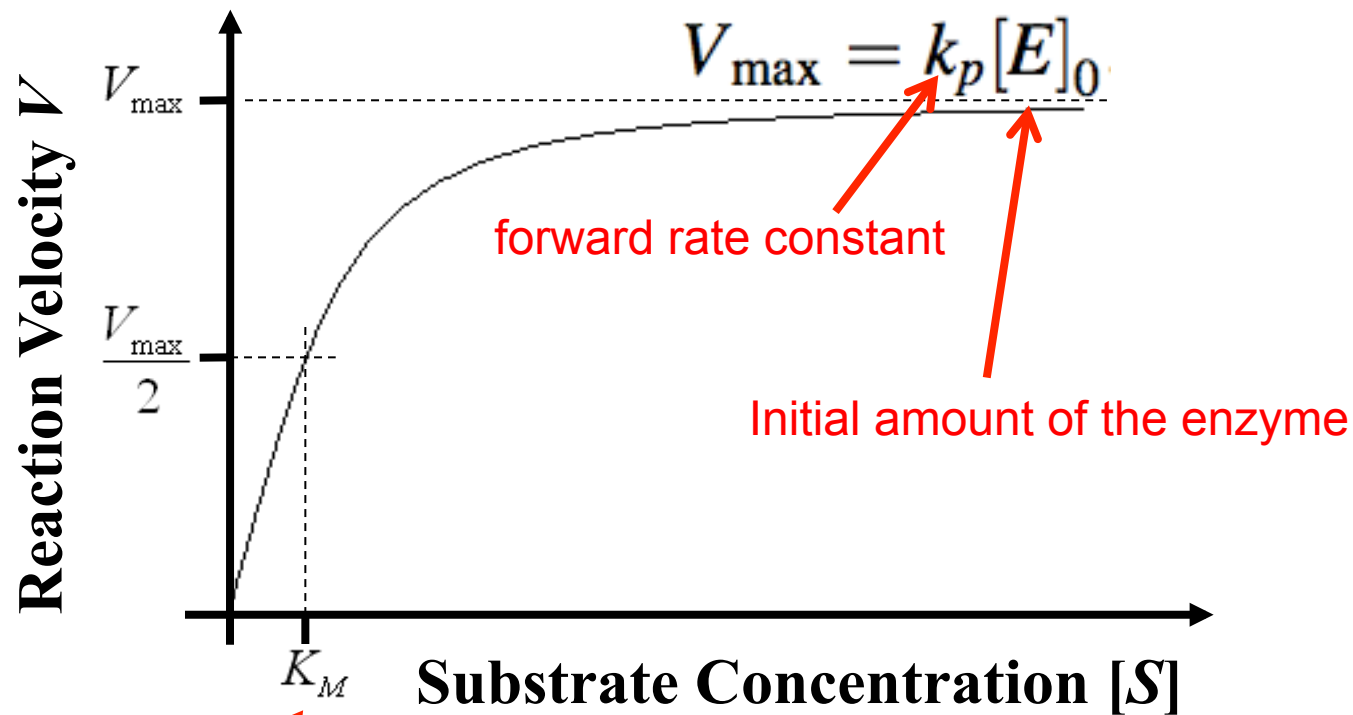
If we, then, go back to couple of previous equations:

$$\frac{d[P]}{dt} = k_p[ES] \quad [ES] = [E]_0 \frac{[S]}{[S] + K_M}$$

We can finally write the the so-called **Michaelis–Menten equation**:

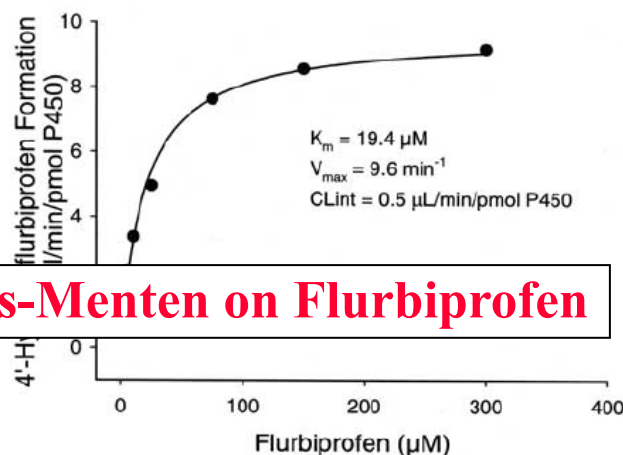
$$V = V_{\max} \frac{[S]}{[S] + K_M}$$

Michaelis-Menten Kinetics

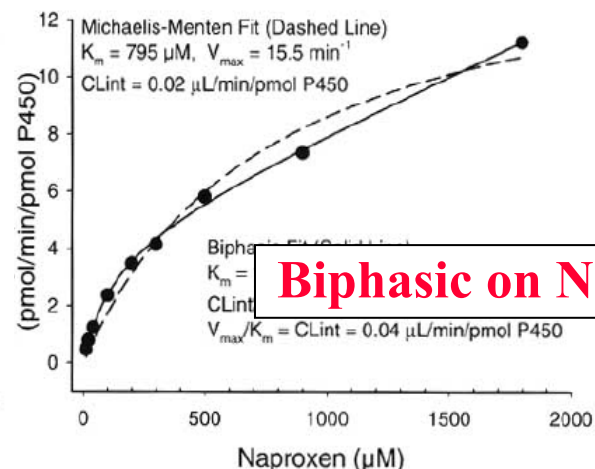


Depends only by the association and dissociation constants and by the forward rate

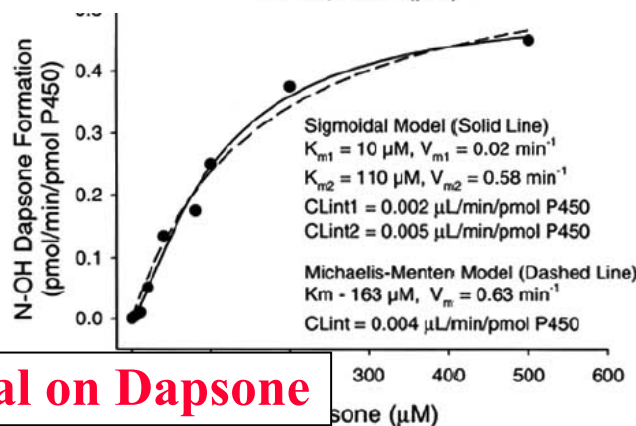
Atypical Enzymatic Reactions



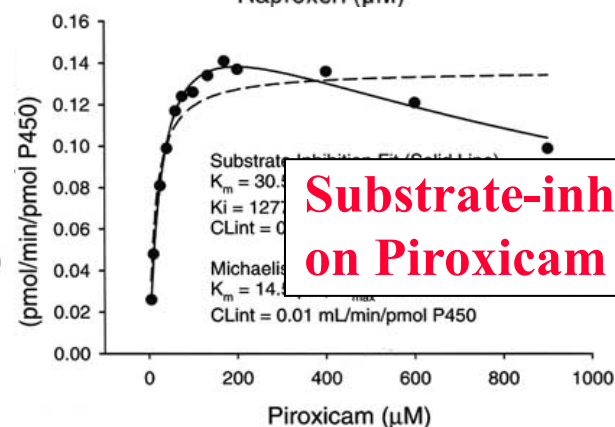
Michaelis-Menten on Flurbiprofen



Biphasic on Naproxen



Sigmoidal on Dapsone



Substrate-inhibition on Piroxicam

Timothy S. Tracy, Current Drug Metabolism, 2003, Vol. 4, No. 5

Some enzymes (e.g., the cytochrome P450 2C9) also present kinetics different than Michaelis-Menten on different substrates!