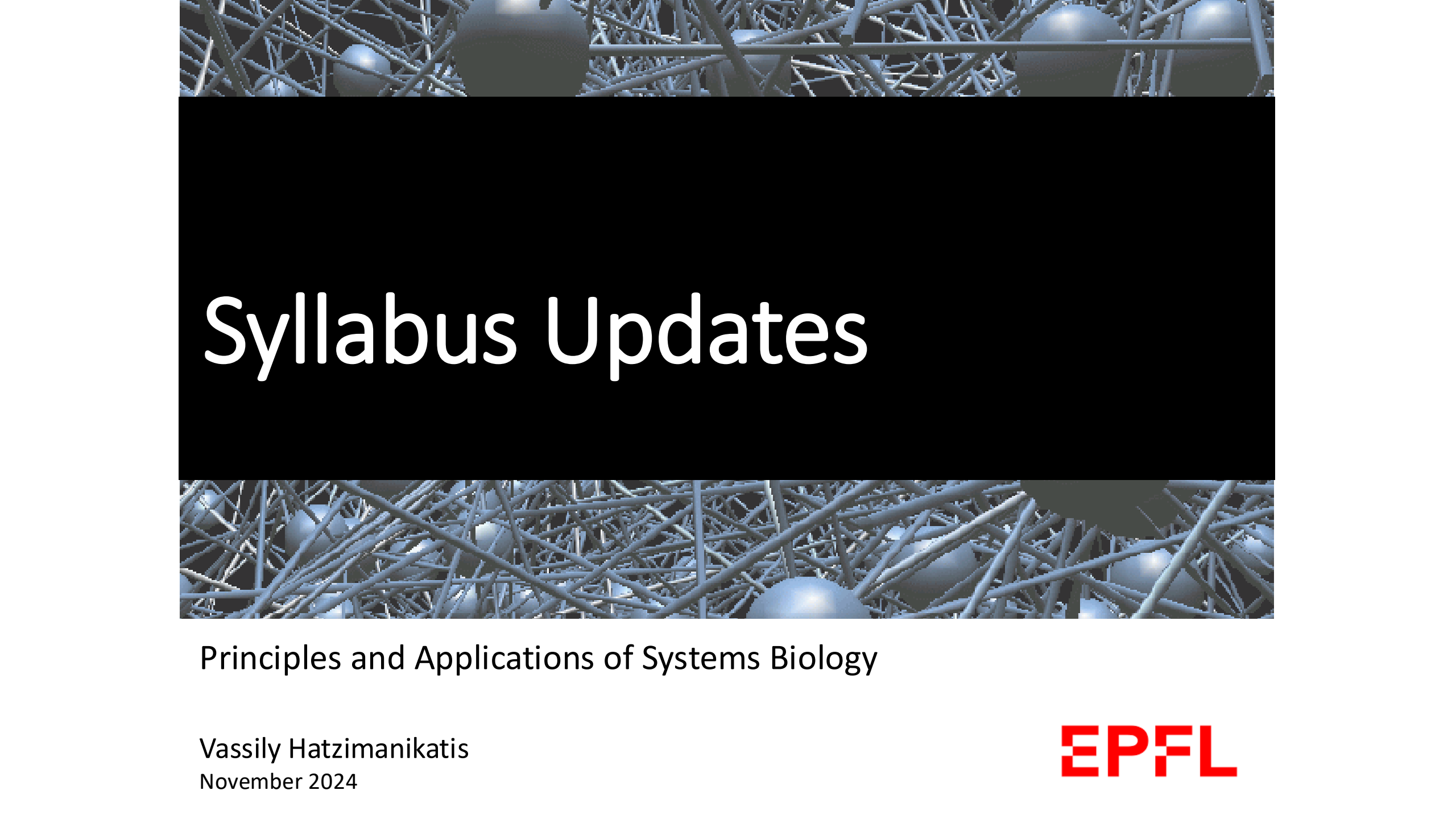




Syllabus Updates

Principles and Applications of Systems Biology

Vassily Hatzimanikatis
November 2024

EPFL

Syllabus

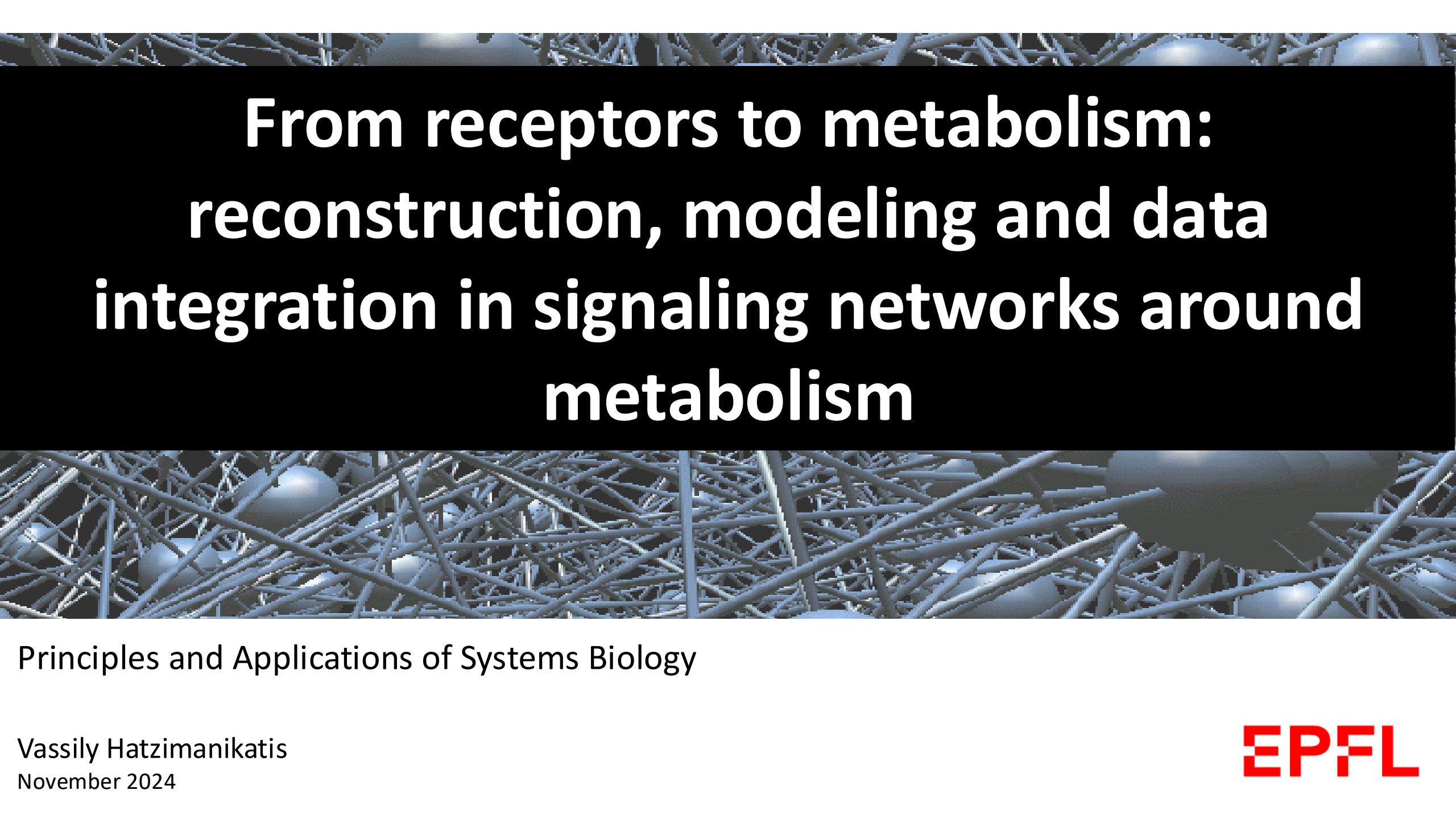
Date	Lecture	Exercise
14.09	Introduction to Systems Biology	-
20.09	FBA, Objective functions, Growth trade off (FAB1) Phenotypic phase plane, alternative flux distributions (FBA2),	-
27.09	Practical (Software; Simulation exercises)	Q/A MATLAB,Python
04.10	(From 20.09) + Thermodynamics flux balance analysis, and GCM (TFA2)	Q/A
11.10	Thermodynamics flux balance analysis, and GCM (TFA2)	Q/A
18.10	Thermodynamics flux balance analysis, and GCM (TFA2)	
01.11	Data integration of Stoichiometric models	Q/A
08.11	Enzyme kinetics/Large-scale modelling + Kinetic modeling of cancer metabolism	Q/A
15.11	Kinetic modeling of cancer metabolism + Signal transduction systems	Q/A
22.11	ME-models + Communities	Q/A
29.11	(Finish from 22.11) + Work on projects	Q/A
06.12	<i>Final Presentations</i>	
13.12	<i>Final Presentations</i>	
20.12	-	

Presentation Guide

Presentations should be maximum 20mins.

Your paper presentation should contain:

- Brief introduction to the topic
- Methods used in the paper
- Reproduction and discussion of the results
- Broader impact on the field



From receptors to metabolism: reconstruction, modeling and data integration in signaling networks around metabolism

Principles and Applications of Systems Biology

Vassily Hatzimanikatis
November 2024

EPFL

Introduction to Signaling

Introduction

Proc. Natl. Acad. Sci. USA
Vol. 78, No. 11, pp. 6840–6844, November 1981
Biochemistry

An amplified sensitivity arising from covalent modification in biological systems

(protein modification/metabolic regulation/switch mechanism/enzyme cascades)

ALBERT GOLDBETER[†] AND DANIEL E. KOSHLAND, JR.

Molecular Cell, Vol. 9, 957–970, May, 2002, Copyright ©2002 by Cell Press

Mathematical Models of Protein Kinase Signal Transduction

Reinhard Heinrich,¹ Benjamin G. Neel,²
and Tom A. Rapoport^{3,4}

¹University of Cologne

²University of Illinois

³University of Chicago

⁴University of Texas

distinct ways to elicit either a
response, which can have dram

atic effects on the cell

IOP PUBLISHING

Phys. Biol. 9 (2012) 045010 (13pp)

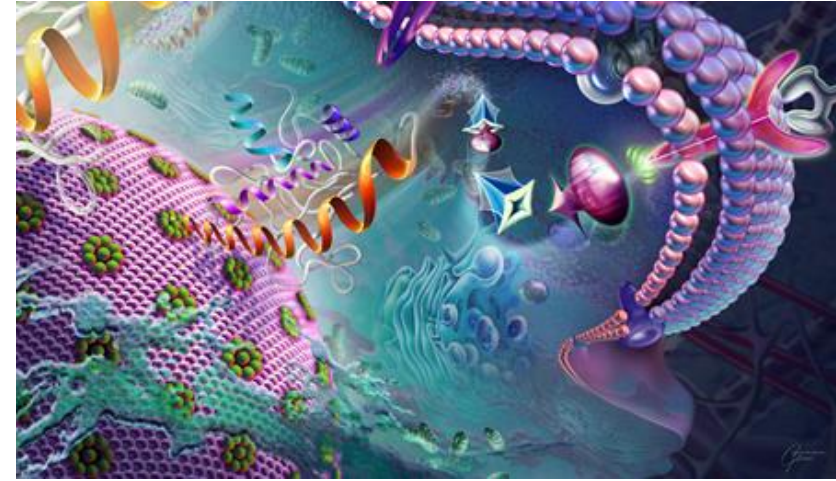
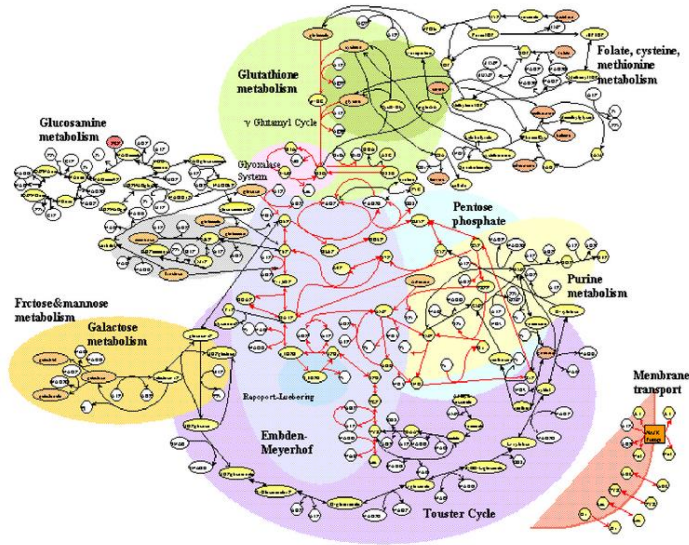
PHYSICAL BIOLOGY

[doi:10.1088/1478-3975/9/4/045010](https://doi.org/10.1088/1478-3975/9/4/045010)

Exploration of trade-offs between steady-state and dynamic properties in signaling cycles

A Radivojevic^{1,2}, B Chachuat³, D Bonvin² and V Hatzimanikatis¹

Introduction



Metabolic pathways

Transfer of **mass**

Determined by presence of enzymes catalyzing the reactions

Metabolites **μM - mM**

Enzyme vs. substrate: lower amount of enzyme

Signal transduction pathways

Transfer of **information**

Compounds of different types: highly organized complexes

10 - 10^4 molecules per cell

Enzyme vs. substrate: same order of magnitude

Introduction

Signaling Paradigm

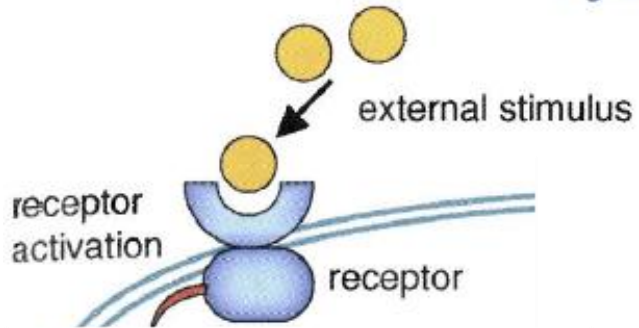


External stimuli:

- hormones
- pheromones
- heat
- cold
- light
- osmotic pressure
- appearance
- Change in concentration of substances
- (glucose, K⁺, Ca⁺, cAMP)

Introduction

Signaling Paradigm

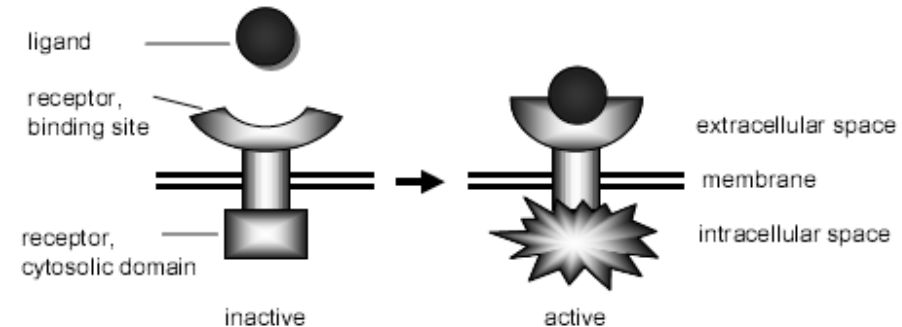


Receptor-ligand interactions:

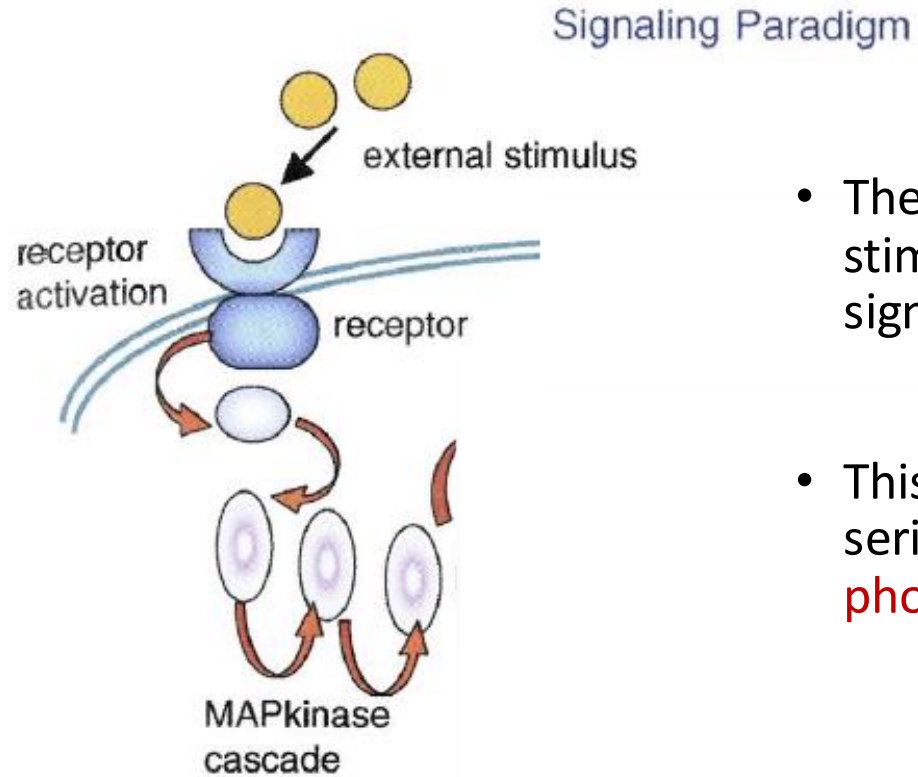
- Many receptors are **transmembrane proteins**; they receive the signal and transmit it.
- In the active form they are able to initiate a **downstream process** within the cell

Importing a signal:

- the stimulus may penetrate the cell membrane and bind to a respective receptor in the cell interior.
- Activation of the cytosolic domain

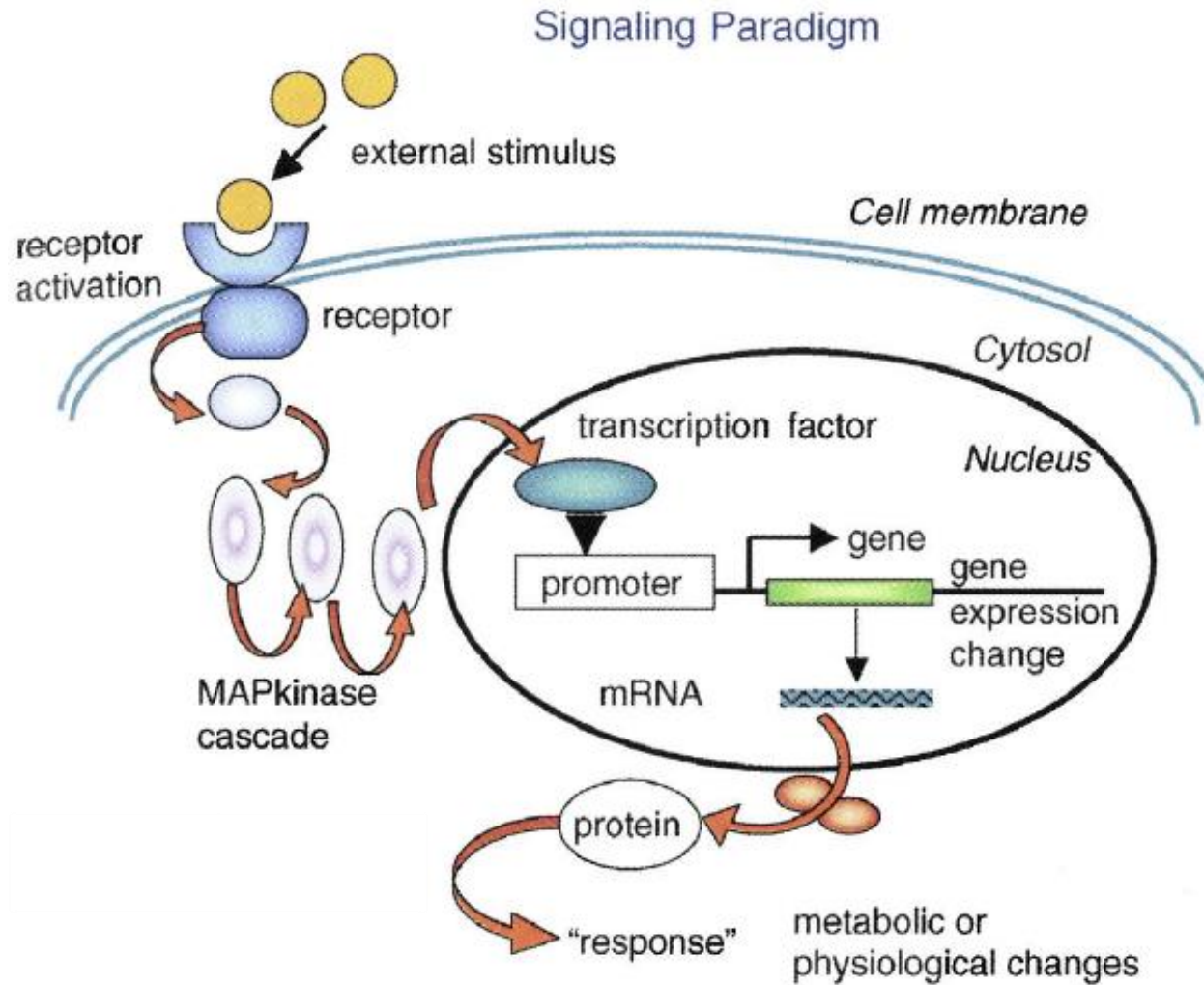


Introduction



- The active receptor stimulates an internal signaling cascade
- This cascade often includes a series of **changes in protein phosphorylation** states

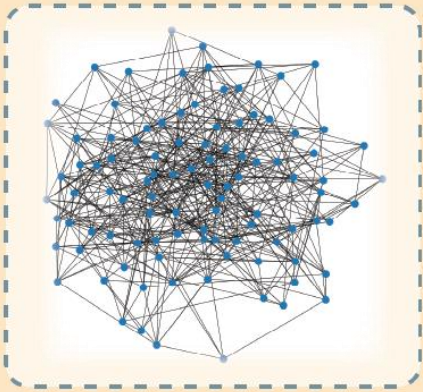
Introduction



Introduction to SIGMA method

Introduction

SIGMA Method



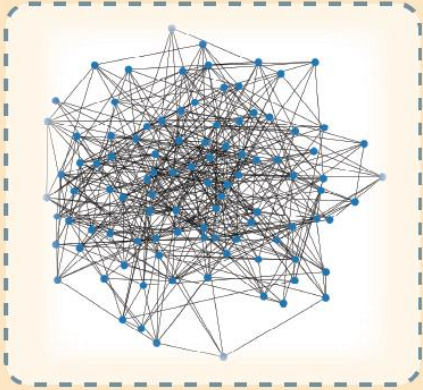
 reactome

Signaling network

- **Reconstruct signaling networks for specific pathways of interest.**

Introduction

SIGMA Method



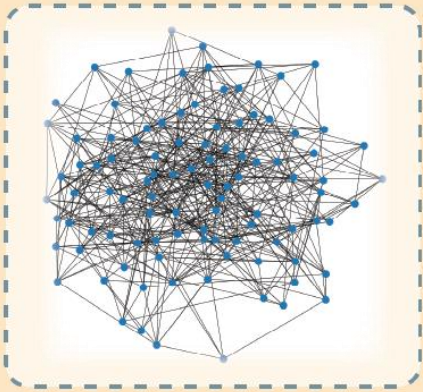
 reactome

Signaling network

- Reconstruct signaling networks for specific pathways of interest.
- Navigate the signaling network to study the *information flow*.

Introduction

SIGMA Method



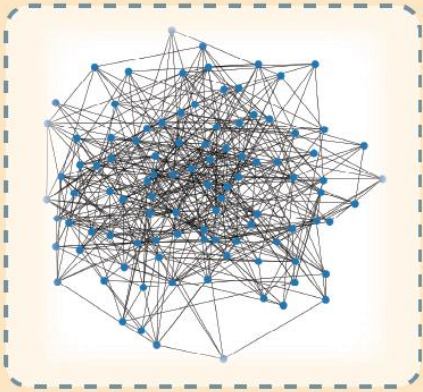
 reactome

Signaling network

- Reconstruct signaling networks for specific pathways of interest.
- Navigate the signaling network to study the *information flow*.
- Identify the flow of information from receptors to transcription factors or proteins.

Introduction

SIGMA Method



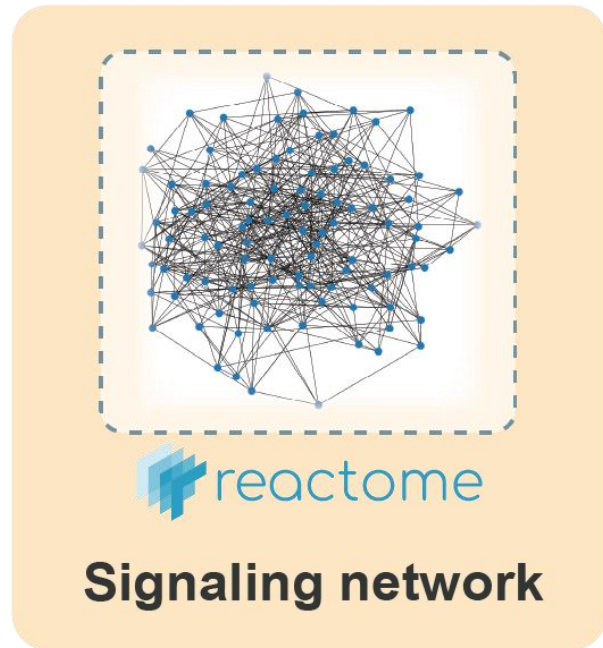
 reactome

Signaling network

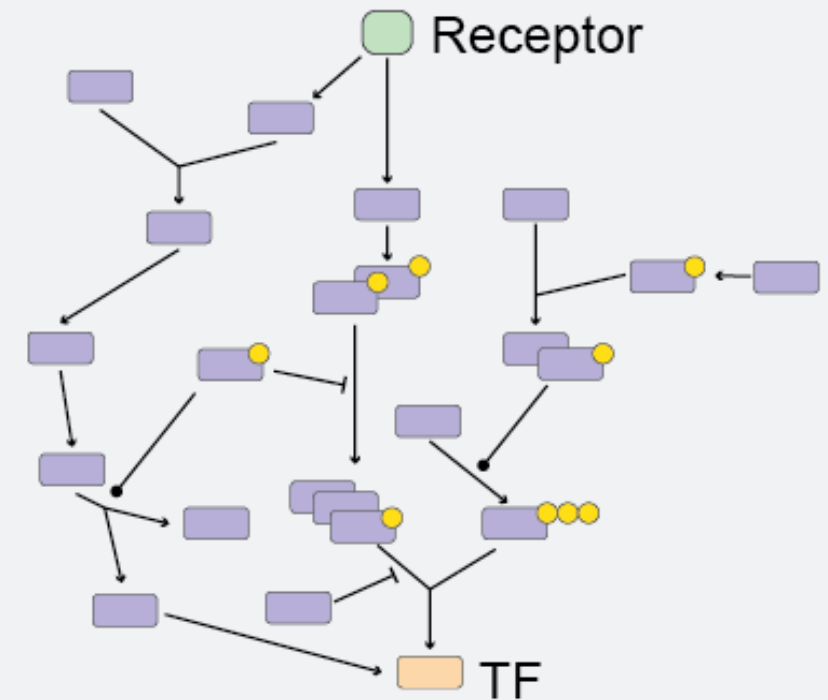
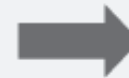
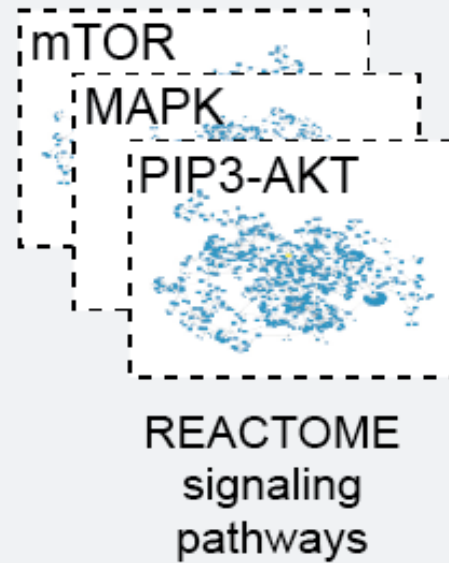
- Reconstruct signaling networks for specific pathways of interest.
- Navigate the signaling network to study the *information flow*.
- Identify the flow of information from receptors to transcription factors or proteins.
- Study deregulation in signaling pathways in a consistent manner.

Reconstruct signaling networks

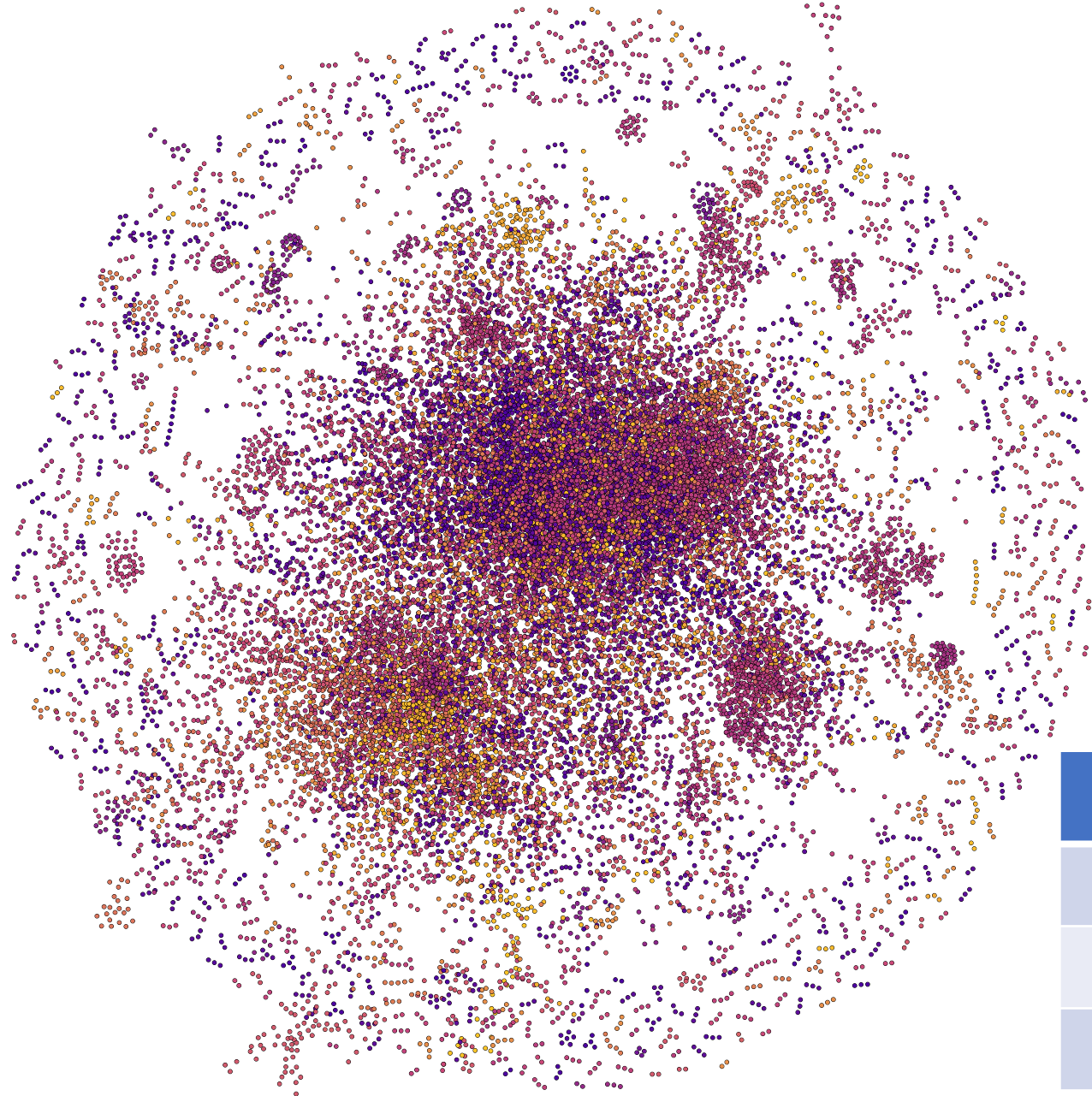
SIGMA Method



Signaling model



Reactome signaling network



Reactome	
Pathways	2610
Species	23830
Connections	46834

From Reactome to selected pathways network

The need for reconstructing signaling networks around selected pathways.

- 1) Allowing manual curation of the selected Reactome pathways.**
- 2) Reducing the network size and complexity.**

From Reactome to selected pathways network

The need for reconstructing signaling networks around selected pathways.

- 1) Allowing manual curation of the selected Reactome pathways.**
- 2) Reducing the network size and complexity.**

Signaling Network Reconstruction Workflow:

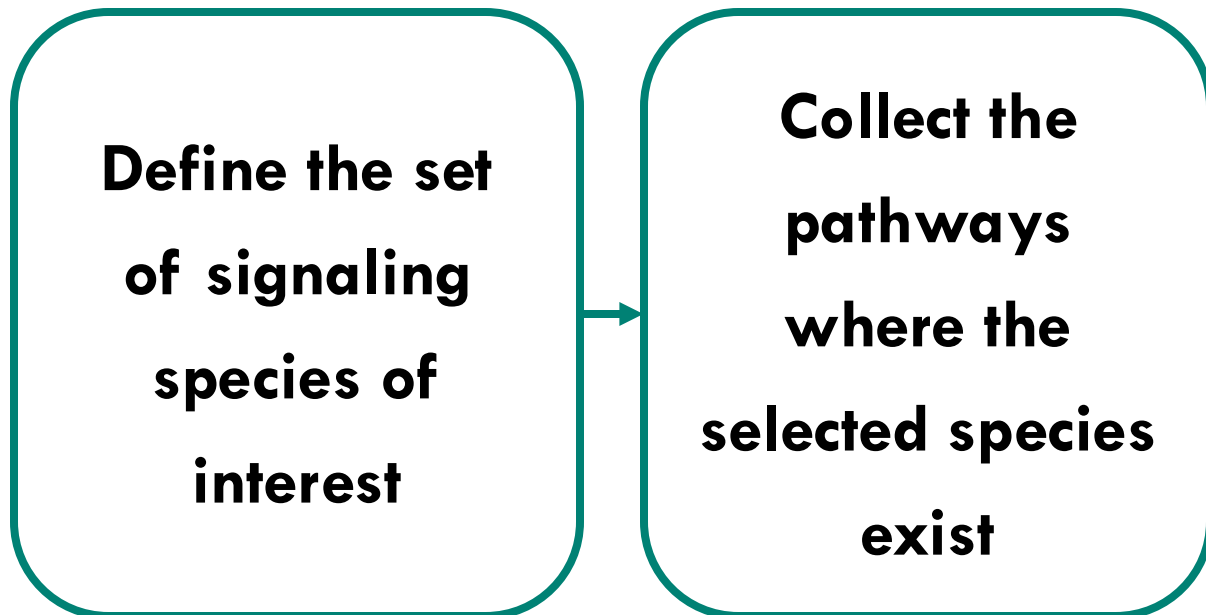
**Define the set
of signaling
species of
interest**

From Reactome to selected pathways network

The need for reconstructing signaling networks around selected pathways.

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Signaling Network Reconstruction Workflow:

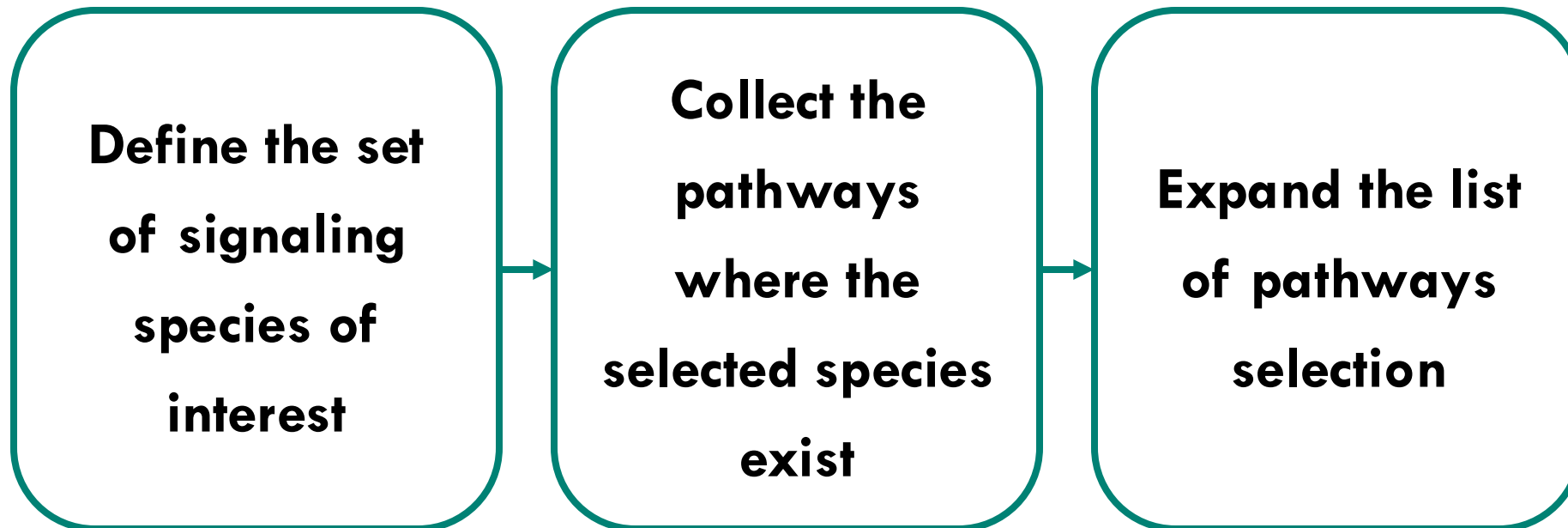


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Signaling Network Reconstruction Workflow:

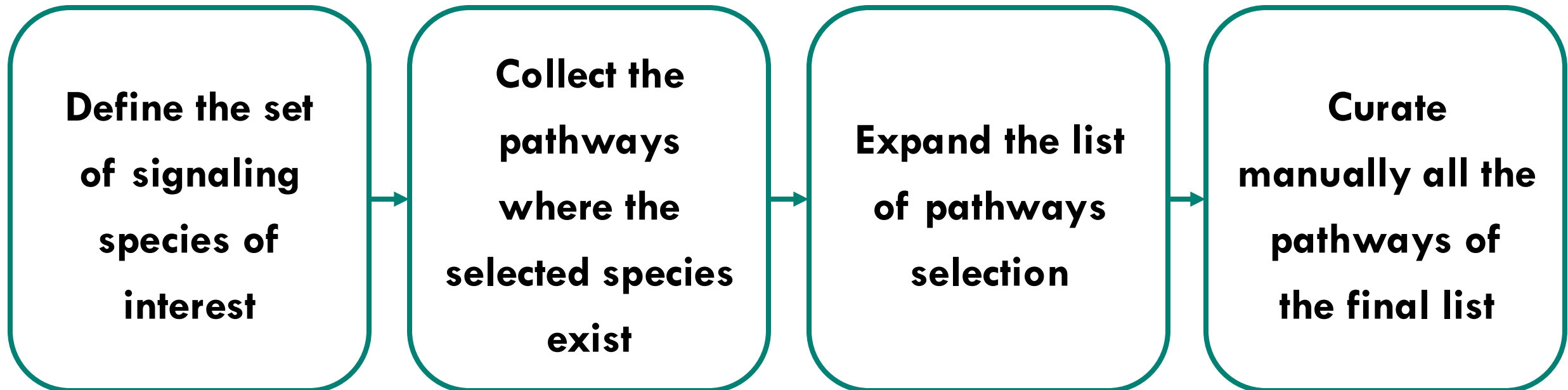


From Reactome to selected pathways network

The need for reconstructing signaling networks around selected pathways.

- 1) Allowing manual curation of the selected Reactome pathways.
- 2) Reducing the network size and complexity.

Signaling Network Reconstruction Workflow:



BDNF signaling in cancer

Received: 18 January 2019 | Revised: 13 April 2019 | Accepted: 18 April 2019
DOI: 10.1002/jcb.29050

RESEARCH ARTICLE

Journal of Cellular Biochemistry

WILEY

Brain-derived neurotrophic factor downregulation in gastric cancer

Farbod Esfandi¹ | Hamid Bouraghi² | Mark C Glassy³ | Mohammad Taheri⁴ | Mir Salar Kahaei¹ | Vahid Kholghi Oskoei¹ | Soudeh Ghafouri-Fard¹

ONCOLOGY REPORTS 40: 621-634, 2018

Proliferative role of BDNF/TrkB signaling is associated with anoikis resistance in cervical cancer

YUAN YUAN, HAI-QIONG YE and QIAN-CHUAN REN

Department of Obstetrics and Gynecology, The Affiliated Hospital of Southwest Medical University, Luzhou, Sichuan 644300, P.R. China

Received October 23, 2017; Accepted May 31, 2018

DOI: 10.3892/or.2018.6515

Received: 10 October 2018 | Revised: 7 April 2020 | Accepted: 20 April 2020
DOI: 10.1096/fj.201802159RRR

RESEARCH ARTICLE

THE FASEB JOURNAL

Activation of BDNF/TrkB pathway promotes prostate cancer progression via induction of epithelial-mesenchymal transition and anoikis resistance

Tao Li | Ying Yu | Yarong Song | Xuechao Li | Dongyang Lan | Peng Zhang | Yajun Xiao | Yifei Xing

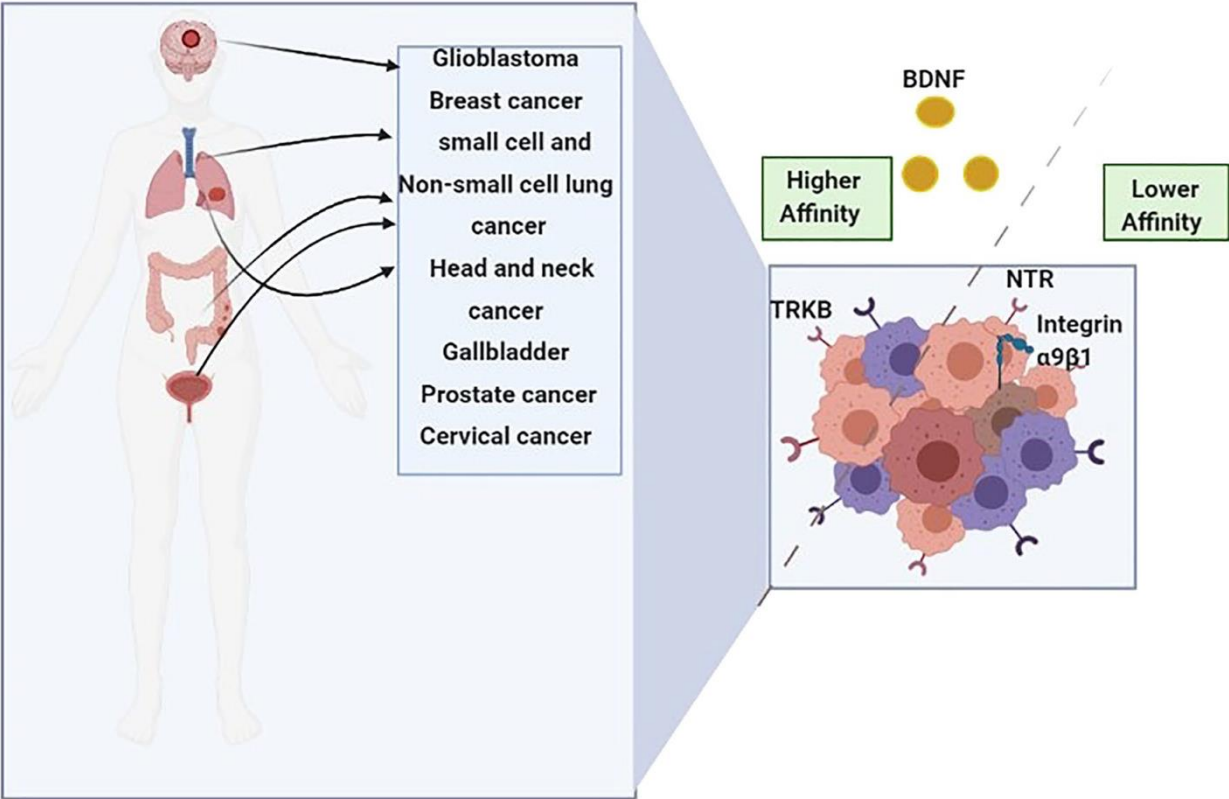
Journal of Cancer Research and Clinical Oncology (2023) 149:2621–2636
<https://doi.org/10.1007/s00432-022-04365-8>

REVIEW

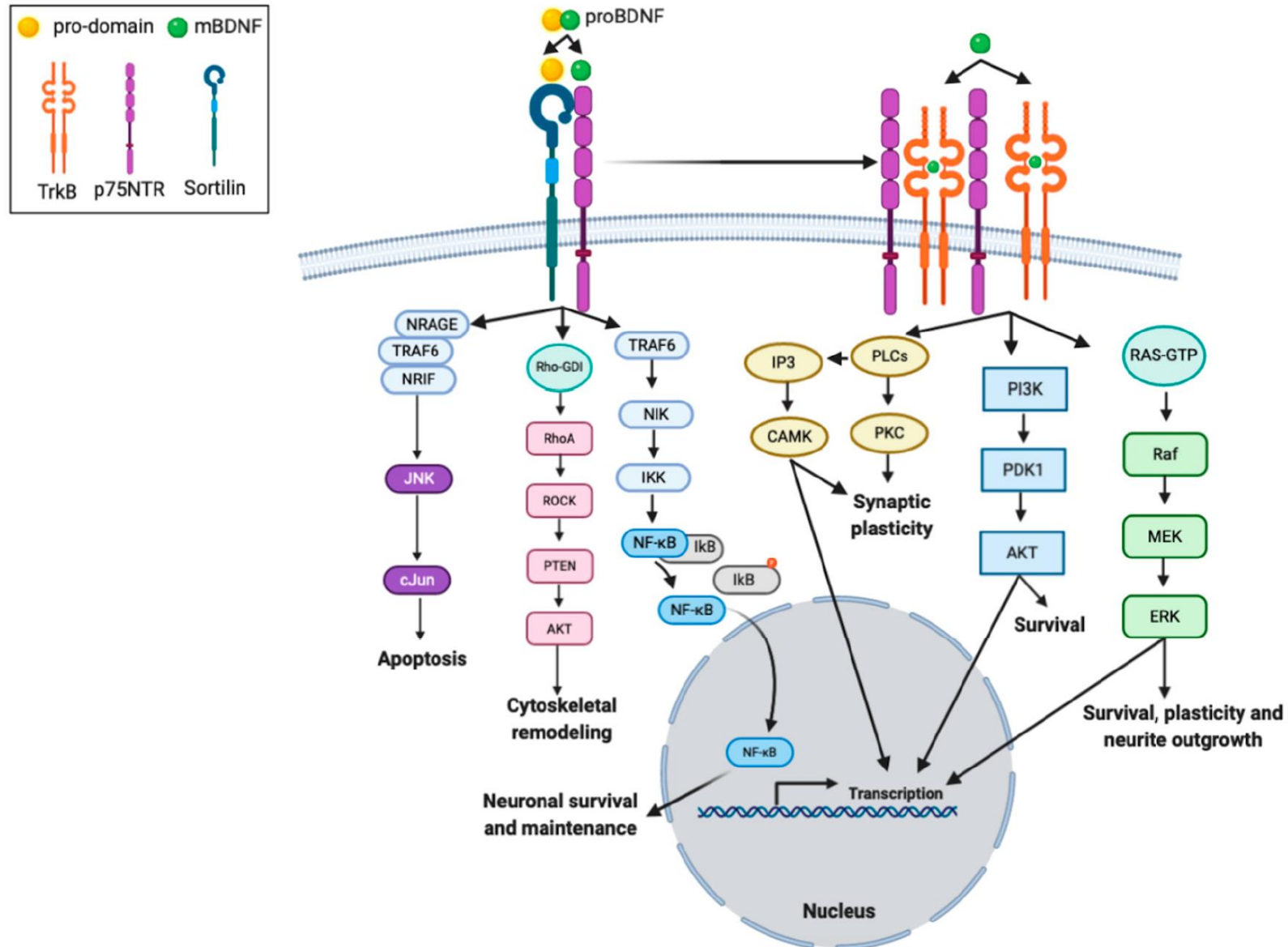
Check for updates

BDNF and its signaling in cancer

Mohammad Malekan^{1,2} · Sasan Salehi Nezamabadi^{3,4} · Elham Samami^{3,5} · Mehdi Mohebalizadeh^{6,7} · Amene Saghazadeh^{3,8} · Nima Rezaei^{8,9,10}



Brain-derived neurotrophic factor (BDNF)



mTOR signaling in cancer

Review Article | [Published: 13 November 2018](#)

mTOR signalling and cellular metabolism are mutual determinants in cancer

[Dirk Mossmann](#), [Sujin Park](#) & [Michael N. Hall](#) 

[Nature Reviews Cancer](#) **18**, 744–757 (2018) | [Cite this article](#)

Hua et al. *Journal of Hematology & Oncology* (2019) 12:71
<https://doi.org/10.1186/s13045-019-0754-1>

Journal of
Hematology & Oncology

REVIEW

Open Access

Targeting mTOR for cancer therapy

Hui Hua¹, Qingbin Kong², Hongying Zhang², Jiao Wang³, Ting Luo⁴ and Yangfu Jiang^{2*}



Zou et al. *Cell Biosci* (2020) 10:31
<https://doi.org/10.1186/s13578-020-00396-1>

Cell & Bioscience

REVIEW

Open Access

mTOR signaling pathway and mTOR inhibitors in cancer: progress and challenges

Zhilin Zou^{1,2,3†}, Tao Tao^{4†}, Hongmei Li^{3*} and Xiao Zhu^{1,2*} 



International Journal of
Molecular Sciences



Review

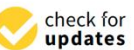
mTOR Signaling in Cancer and mTOR Inhibitors in Solid Tumor Targeting Therapy

Tian Tian, Xiaoyi Li and Jinhua Zhang *

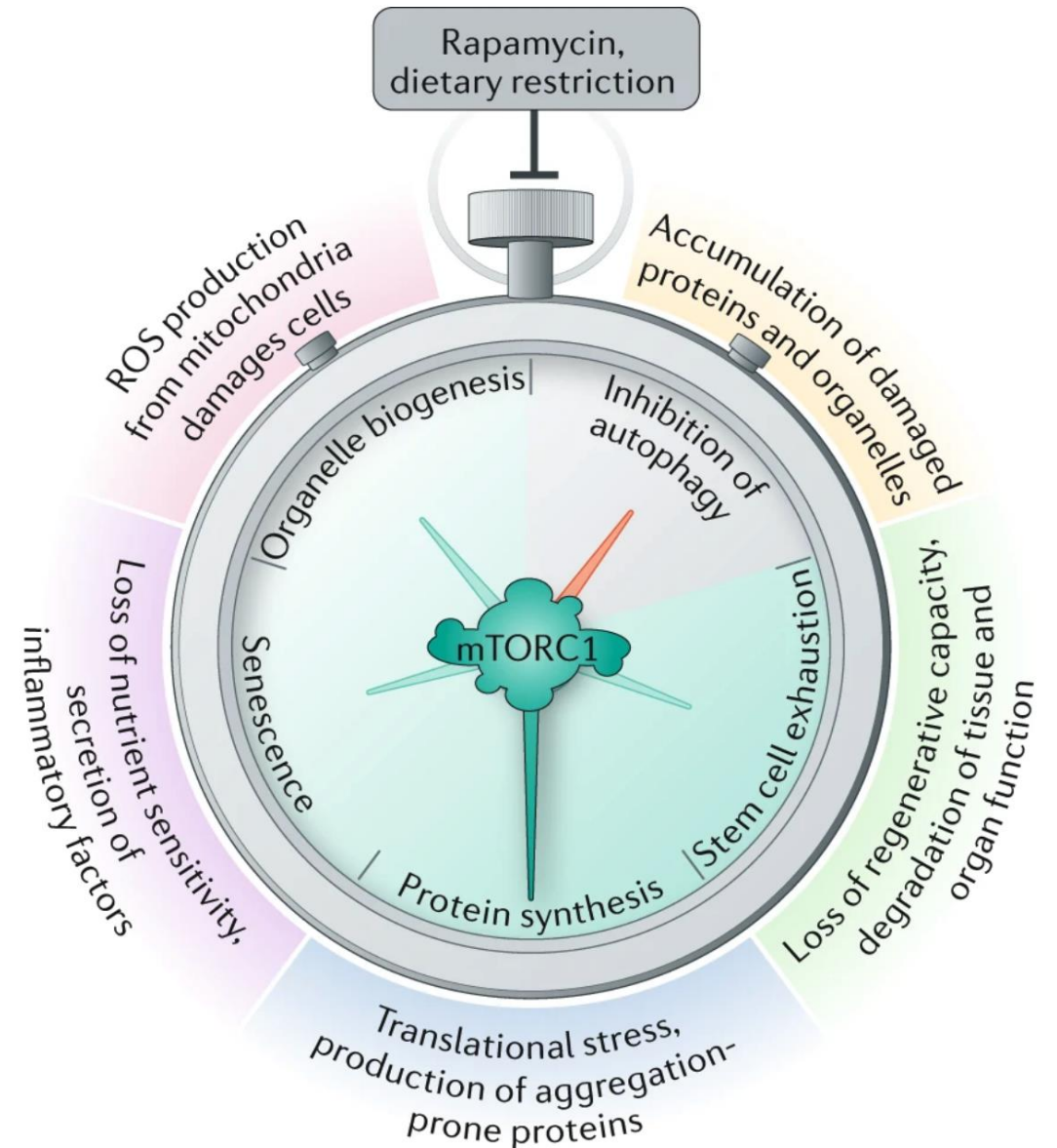
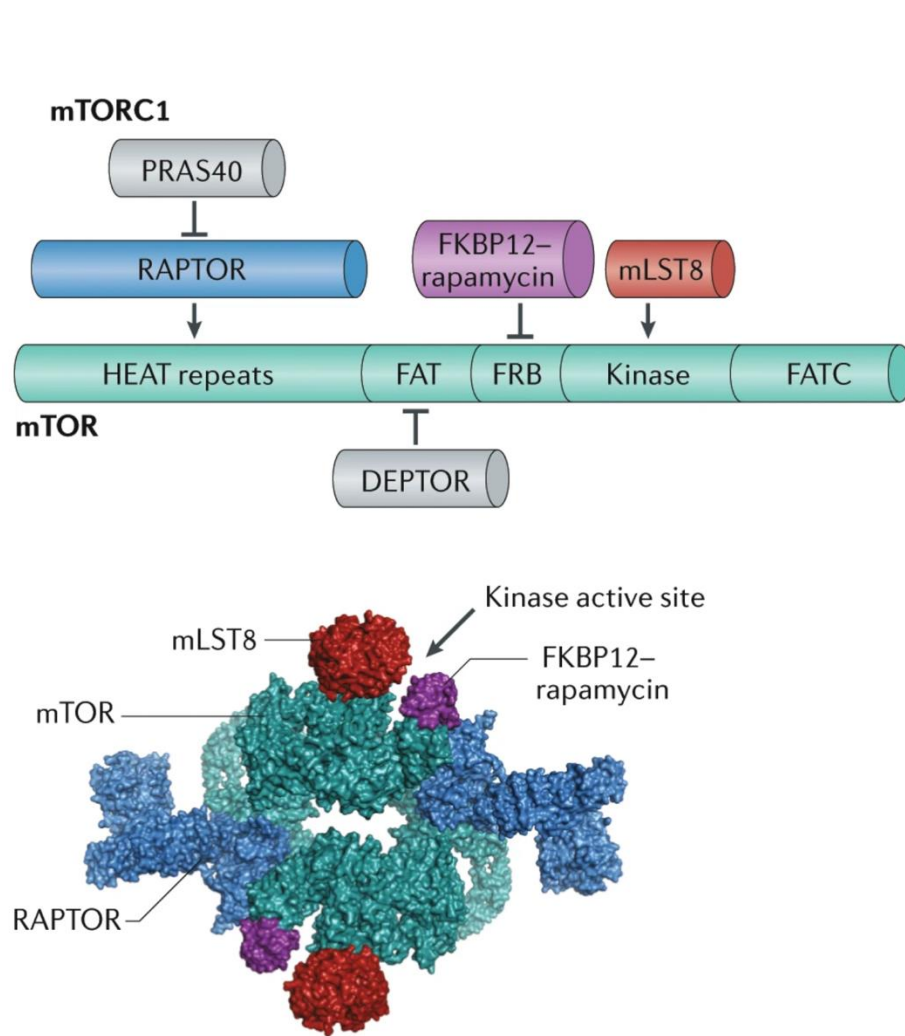
College of Life Science and Bioengineering, Beijing Jiaotong University, Beijing 100044, China;
ttian@bjtu.edu.cn (T.T.); 15271074@bjtu.edu.cn (X.L.)

* Correspondence: zhangjh@bjtu.edu.cn; Tel./Fax: +86-10-51684351

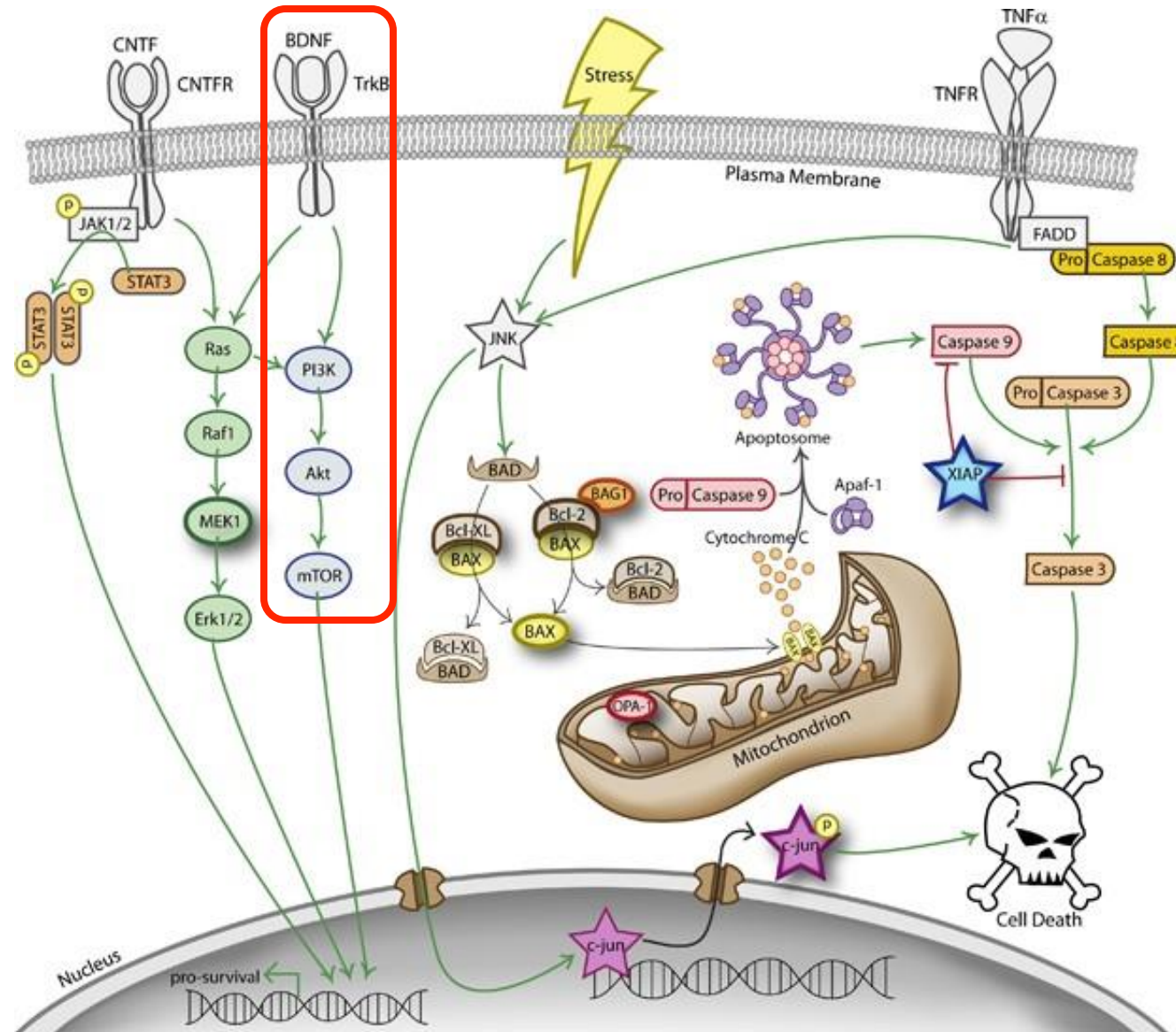
Received: 10 January 2019; Accepted: 1 February 2019; Published: 11 February 2019



Mammalian target of rapamycin (mTOR)



mTOR regulation through BDNF



AMPK in Cancer

Review Article | [Published: 13 March 2019](#)

AMP-activated protein kinase: the current landscape for drug development

[Gregory R. Steinberg](#)  & [David Carling](#)

Nature Reviews Drug Discovery **18**, 527–551 (2019) | [Cite this article](#)

OPEN
BIOLOGY

royalsocietypublishing.org/journal/rsob

The strange case of AMPK and cancer:
Dr Jekyll or Mr Hyde?[†]

Diana Vara-Ciruelos, Fiona M. Russell and D. Grahame Hardie

Division of Cell Signalling and Immunology, School of Life Sciences, University of Dundee, Dow Street,
Dundee DD1 5EH, UK

Yuan *et al. Journal of Hematology & Oncology* (2020) 13:113
<https://doi.org/10.1186/s13045-020-00949-4>

Journal of
Hematology & Oncology

REVIEW

Open Access

The MAPK and AMPK signalings: interplay and implication in targeted cancer therapy

Jimin Yuan^{1,2*}, Xiaoduo Dong³, Jiajun Yap⁴ and Jiancheng Hu^{4,5*} 



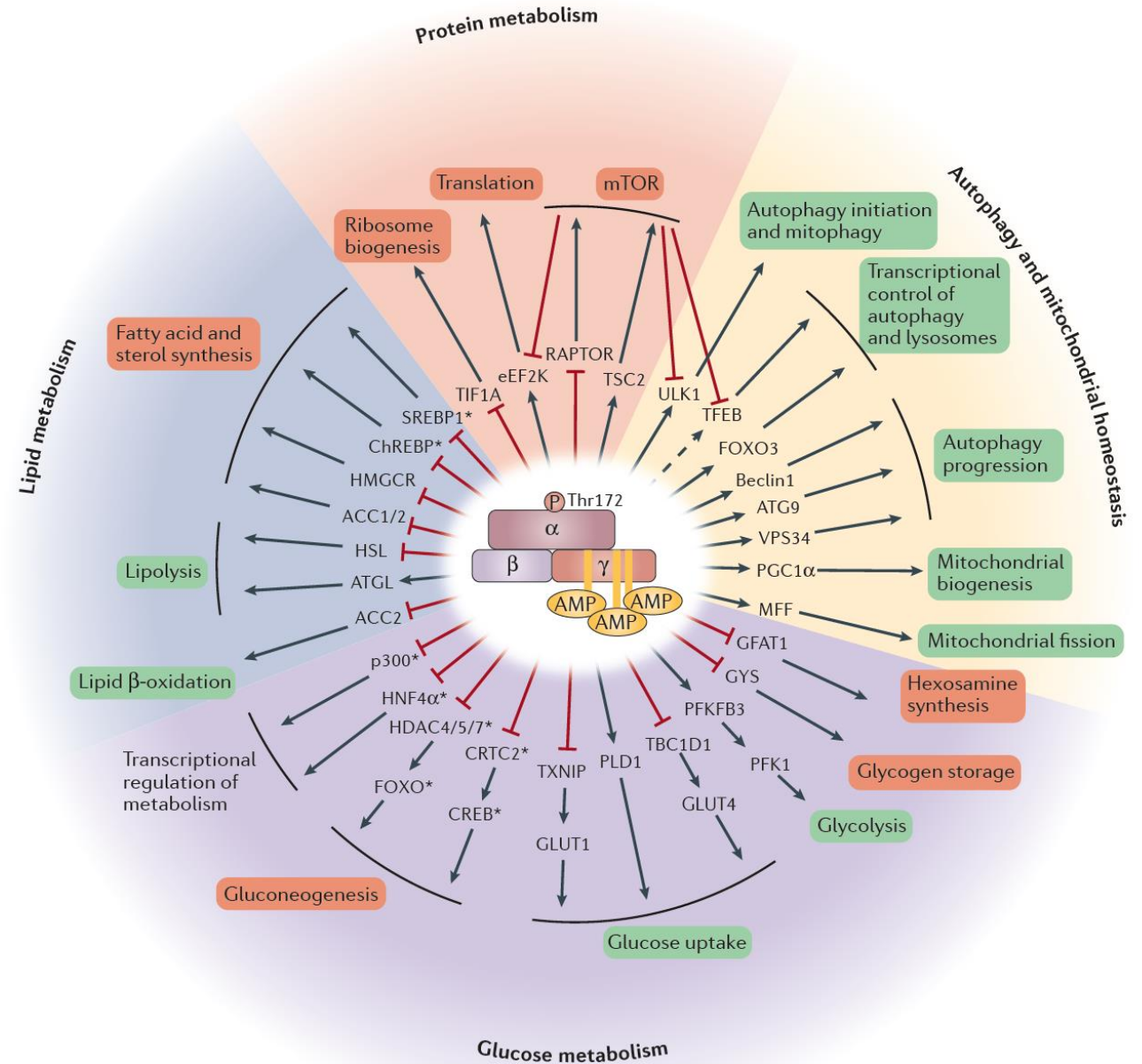
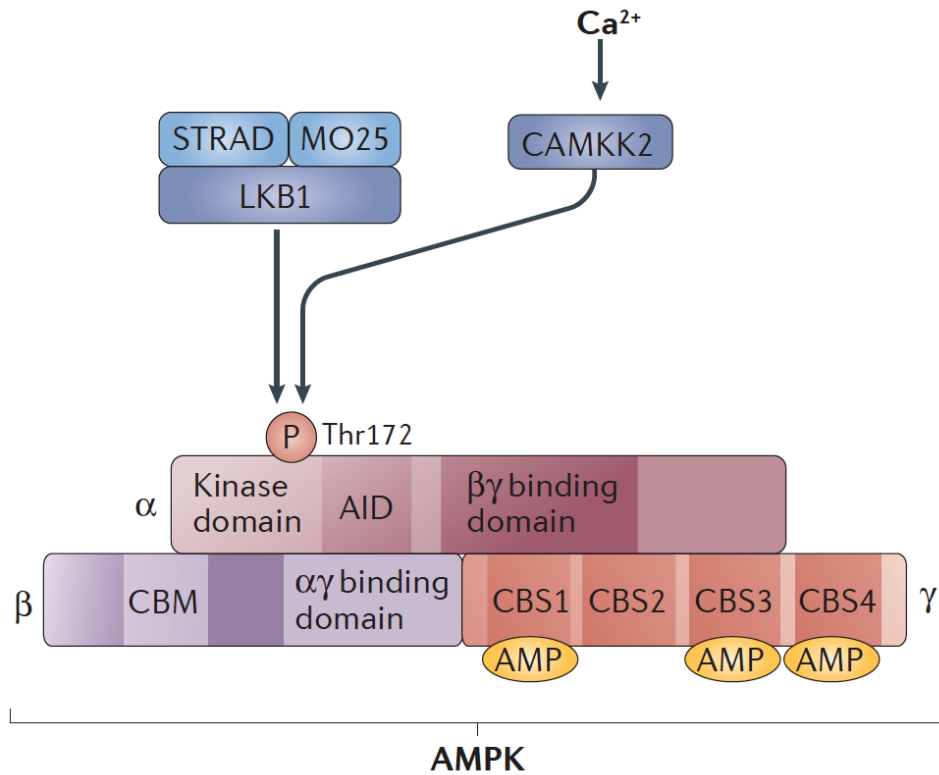
Hindawi
Oxidative Medicine and Cellular Longevity
Volume 2019, Article ID 8730816, 16 pages
<https://doi.org/10.1155/2019/8730816>

Review Article

LKB1/AMPK Pathway and Drug Response in Cancer: A Therapeutic Perspective

Francesco Ciccarese , Elisabetta Zulato , and Stefano Indraccolo 

AMP-activated protein kinase (AMPK)



Signaling network reconstruction

1. Define the set of signaling species of interest

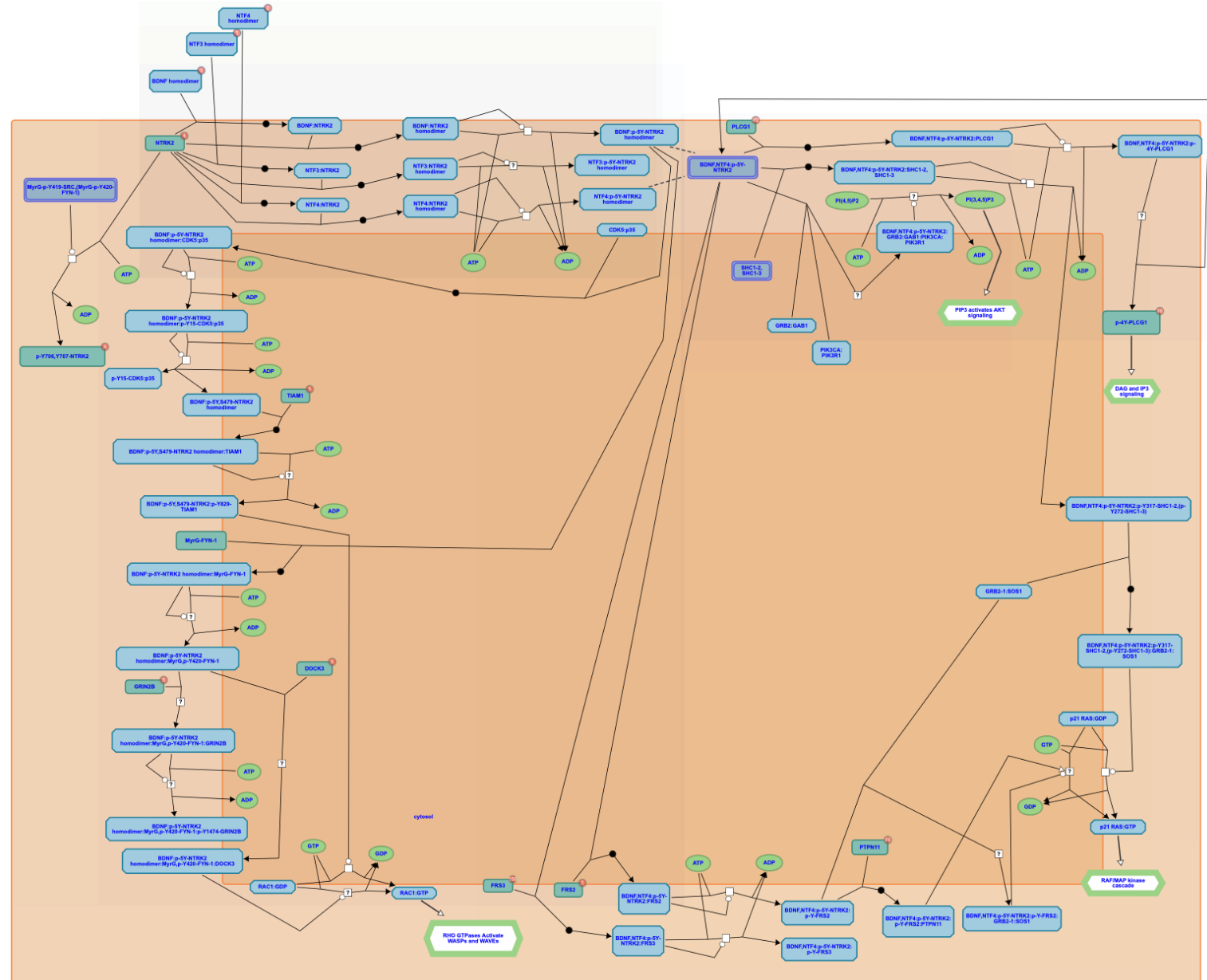
Signaling network reconstruction

1. Define the set of signaling species of interest

- **Brain-derived neurotrophic factor (BDNF)**

BDNF signaling in Reactome

Signaling by Neurotrophic Receptor Tyrosine Kinase 2 (NTRK2)



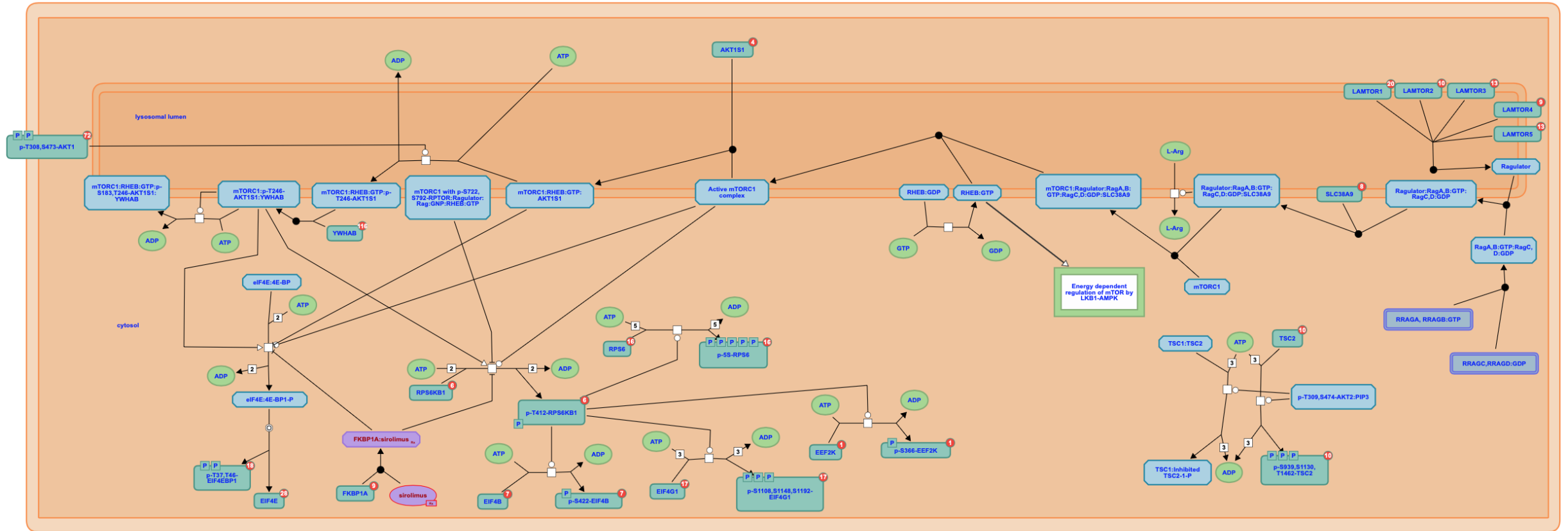
Signaling network reconstruction

1. Define the set of signaling species of interest

- **Brain-derived neurotrophic factor (BDNF)**
- **Mammalian target of rapamycin (mTOR)**

mTOR signaling in Reactome

mTOR signaling



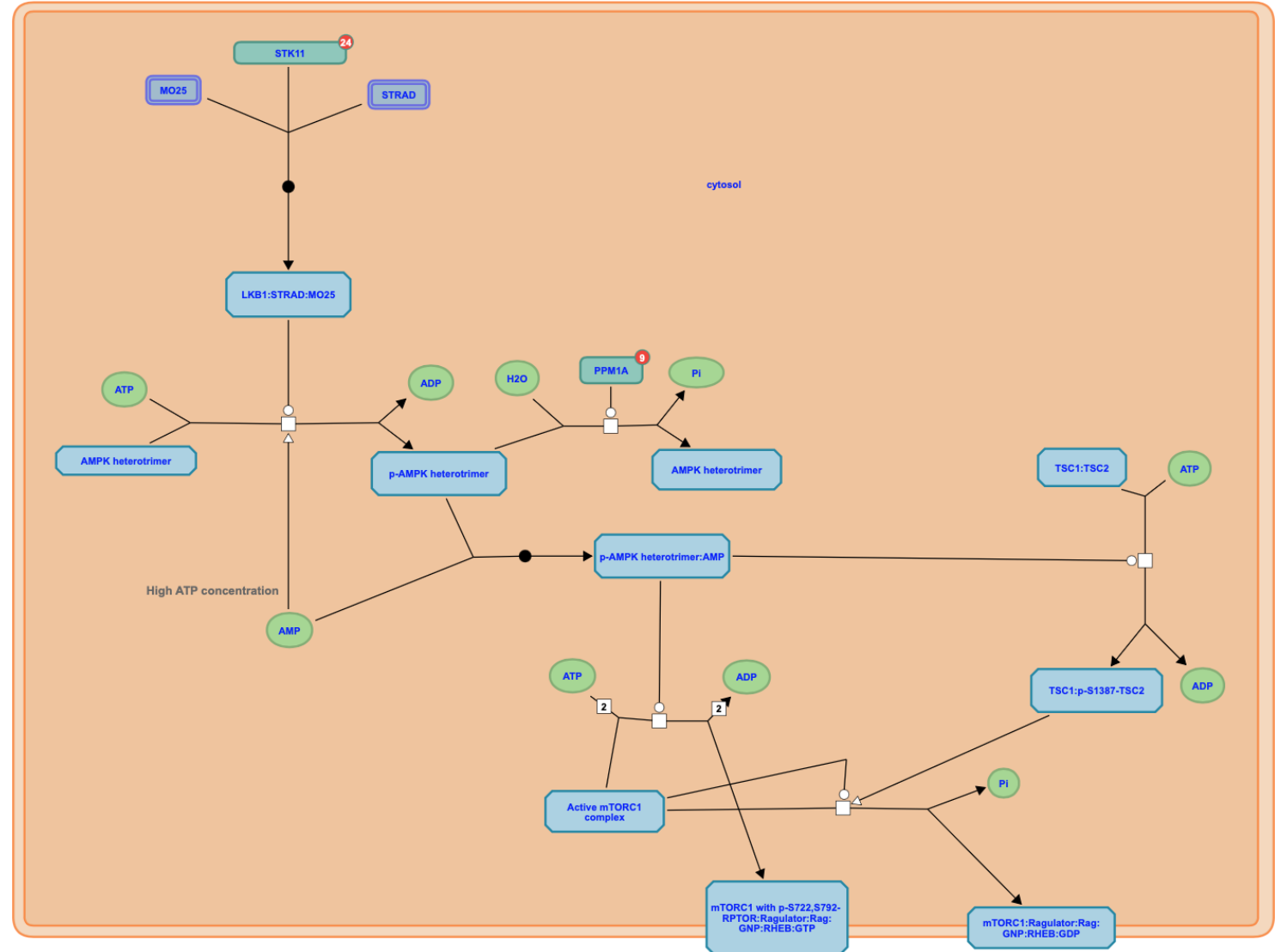
Signaling network reconstruction

1. Define the set of signaling species of interest

- **Brain-derived neurotrophic factor (BDNF)**
- **Mammalian target of rapamycin (mTOR)**
- **AMP-activated protein kinase (AMPK)**

AMPK signaling in Reactome

Energy dependent regulation of mTOR by LKB1-AMPK



Signaling network reconstruction

2. Collect the pathways where the selected species exist

Signaling network reconstruction

2. Collect the pathways where the selected species exist

BDNF

Signaling
by
NTRK2
(TRKB)

mTOR

PTEN
Regulation

Amino
acids
regulate
mTORC1

MTOR
signalling

Macroautophagy

Cellular
response
to heat
stress

TP53
Regulates
Metabolic
Genes

AMPK

Mitochondrial
biogenesis

Energy
dependent
regulation
of mTOR
by
LKB1-AMPK

TP53
Regulates
Metabolic
Genes

Integration
of energy
metabolism

Translocation
of
SLC2A4
(GLUT4)
to the
plasma
membrane

Macroautophagy

Lipophagy

Carnitine
metabolism

Activation
of NMDA
receptors
and
postsynaptic
events

Regulation
of TP53
Activity
through
Phosphorylation

Signaling network reconstruction

2. Collect the pathways where the selected species exist

BDNF

Signaling
by
NTRK2
(TRKB)

mTOR

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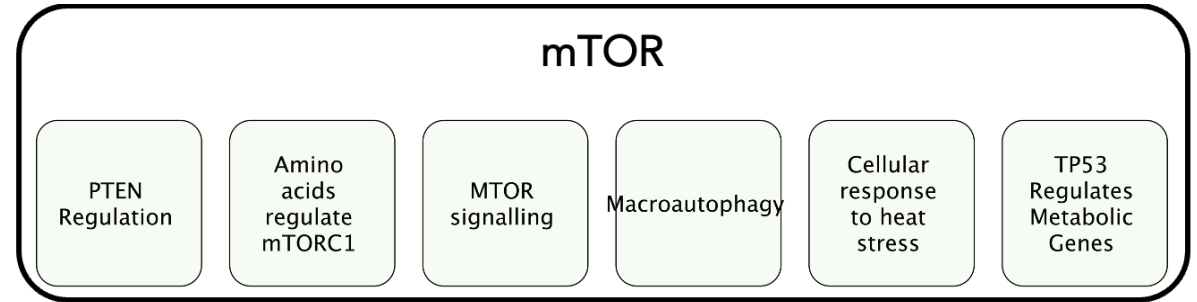
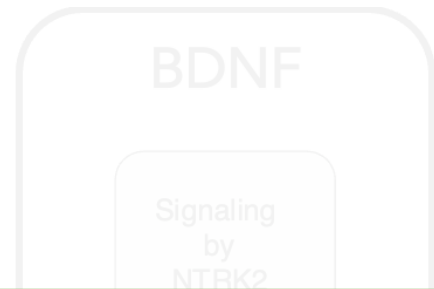
Regulation
of TP53
Activity
through
Phosphorylation



The pathway collection is not only based in the original state of the species but also in modified states of the species (e.g. phosphorylated state, ubiquitinated state, complex component)

Signaling network reconstruction

2. Collect the pathways where the selected species exist



Active mTORC1 complex [I]	mTORC1:Ragulator:RagA,B:GTP:RagC,D:GDP:SLC38A9 [I]
mTORC1 [c]	mTORC1:RHEB:GTP:AKT1S1 [I]
mTORC1 dimer [n]	mTORC1:RHEB:GTP:p-S183,T246-AKT1S1:YWHAB [I]
mTORC1 with p-S722,S792-RPTOR:Ragulator:Rag:GNP:RHEB:GTP [I]	MTORC1:RHEB:GTP:p-S758-ULK1:ATG13:RB1CC1:ATG101 [I]
MTORC1 with p-S722,S792-RPTOR:Ragulator:Rag:RHEB:GTP [I]	mTORC1:RHEB:GTP:p-T246-AKT1S1 [I]
MTORC1 with p-S722,S792-RPTOR:RHEB:GTP:p-S758-ULK1:ATG13:RB1CC1:ATG101 [I]	MTORC1:RHEB:GTP:ULK1:ATG13:RB1CC1:ATG101 [I]
mTORC1:p-T246-AKT1S1:YWHAB [I]	v-ATPase:Ragulator:RRAGA,B:GDP:RRAGC,D:GDP:mTORC1:RHEB:GTP [I]
mTORC1:Ragulator:Rag:GNP:RHEB:GDP [I]	v-ATPase:Ragulator:RRAGA,B:GTP:RRAGC,D:GDP:mTORC1 [I]
MTORC1:Ragulator:Rag:GNP:RHEB:GDP [I]	v-ATPase:Ragulator:RRAGA,B:GTP:RRAGC,D:GDP:mTORC1:RHEB:GTP [I]
MTORC1:Ragulator:Rag:GNP:RHEB:GTP [I]	

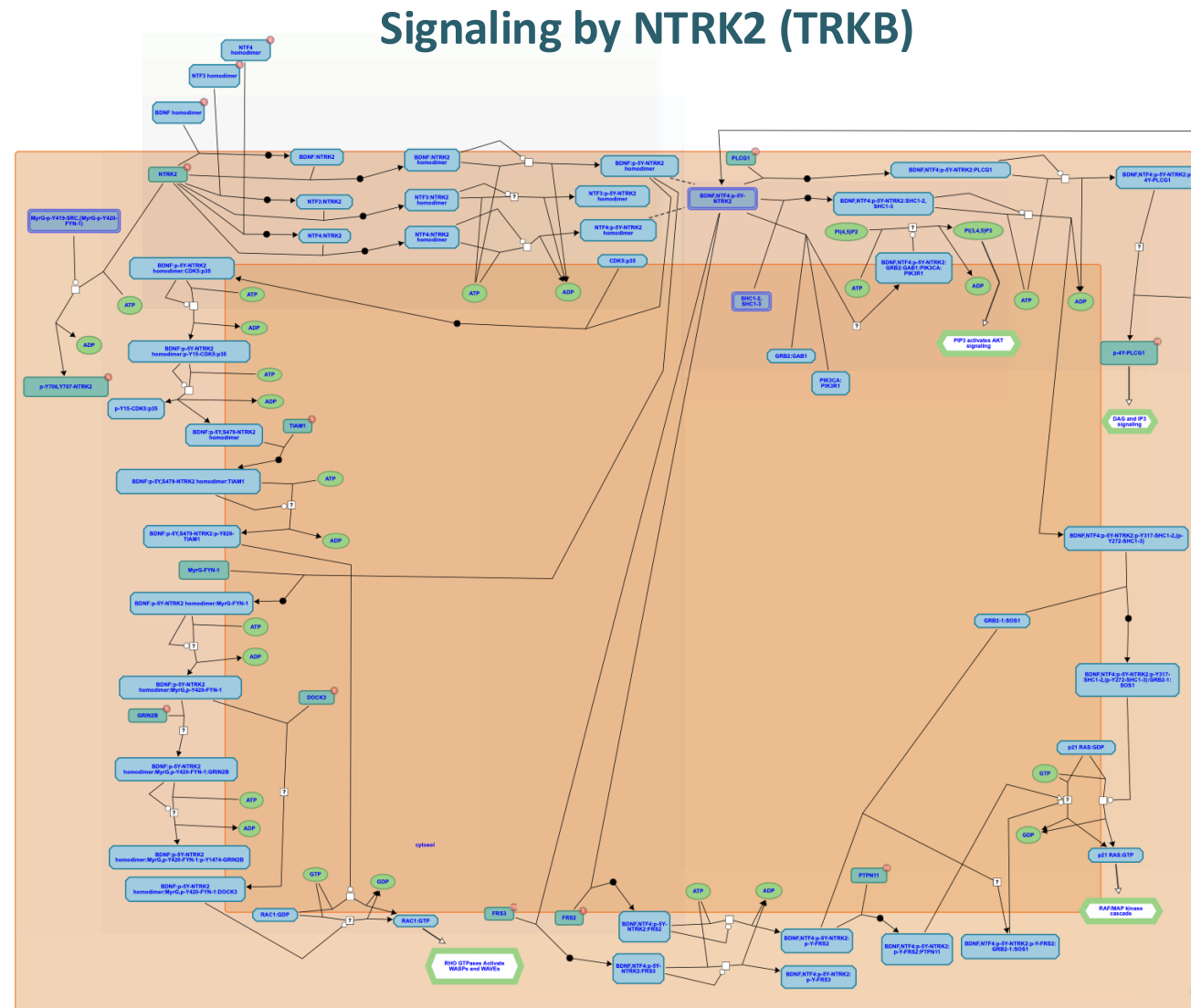


The pathway collection is not only based in the original state of the species but also in modified states of the species (e.g. phosphorylated state, ubiquitinated state, complex component)

Signaling network reconstruction

3. Expand the list of pathways selection

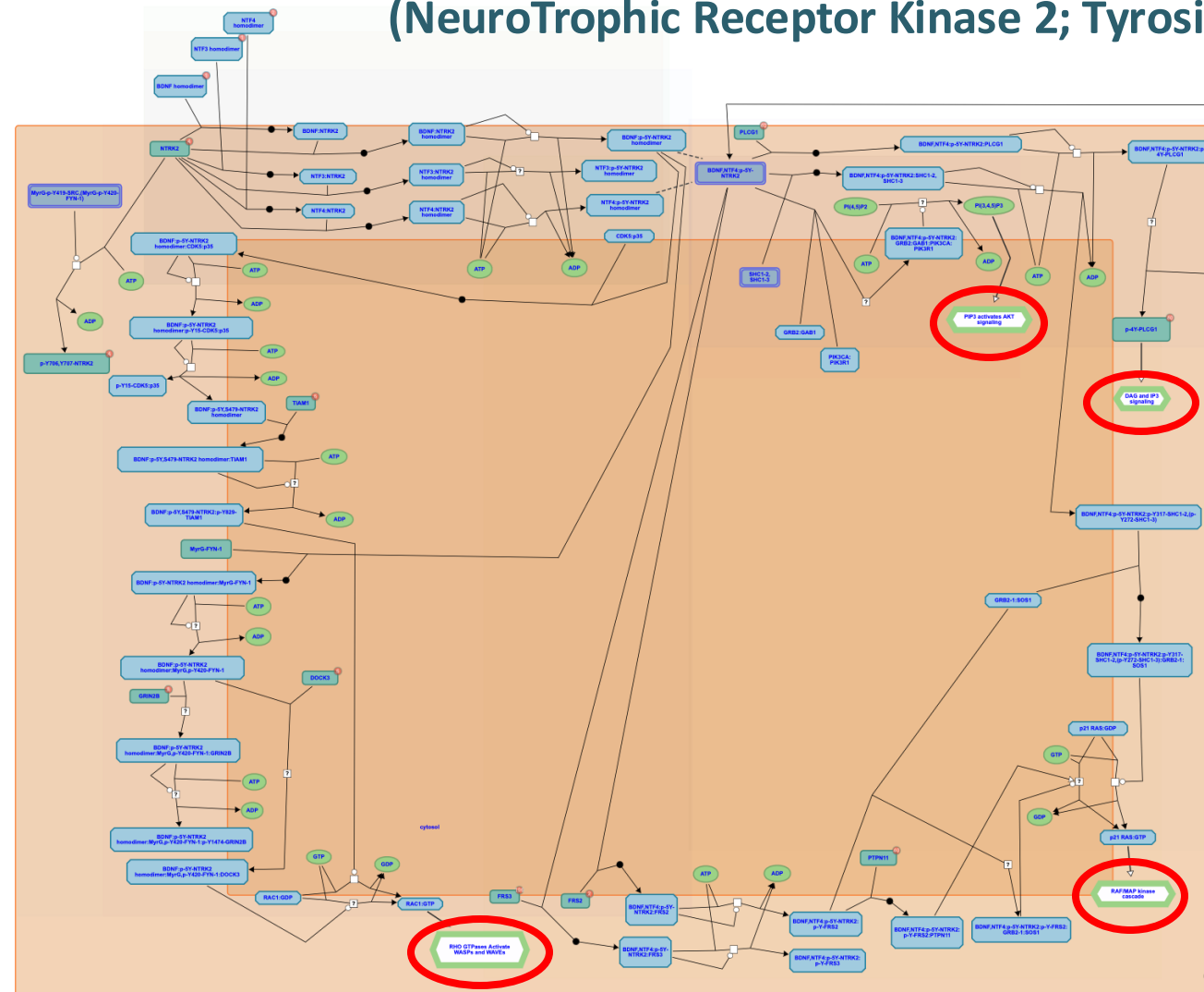
3. Expand the list of pathways selection



3. Expand the list of pathways selection

Signaling by NTRK2 (TRKB)

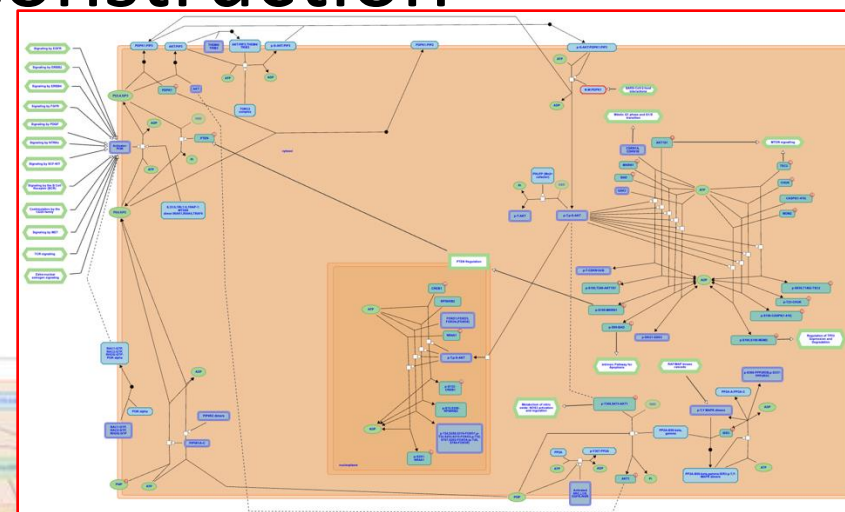
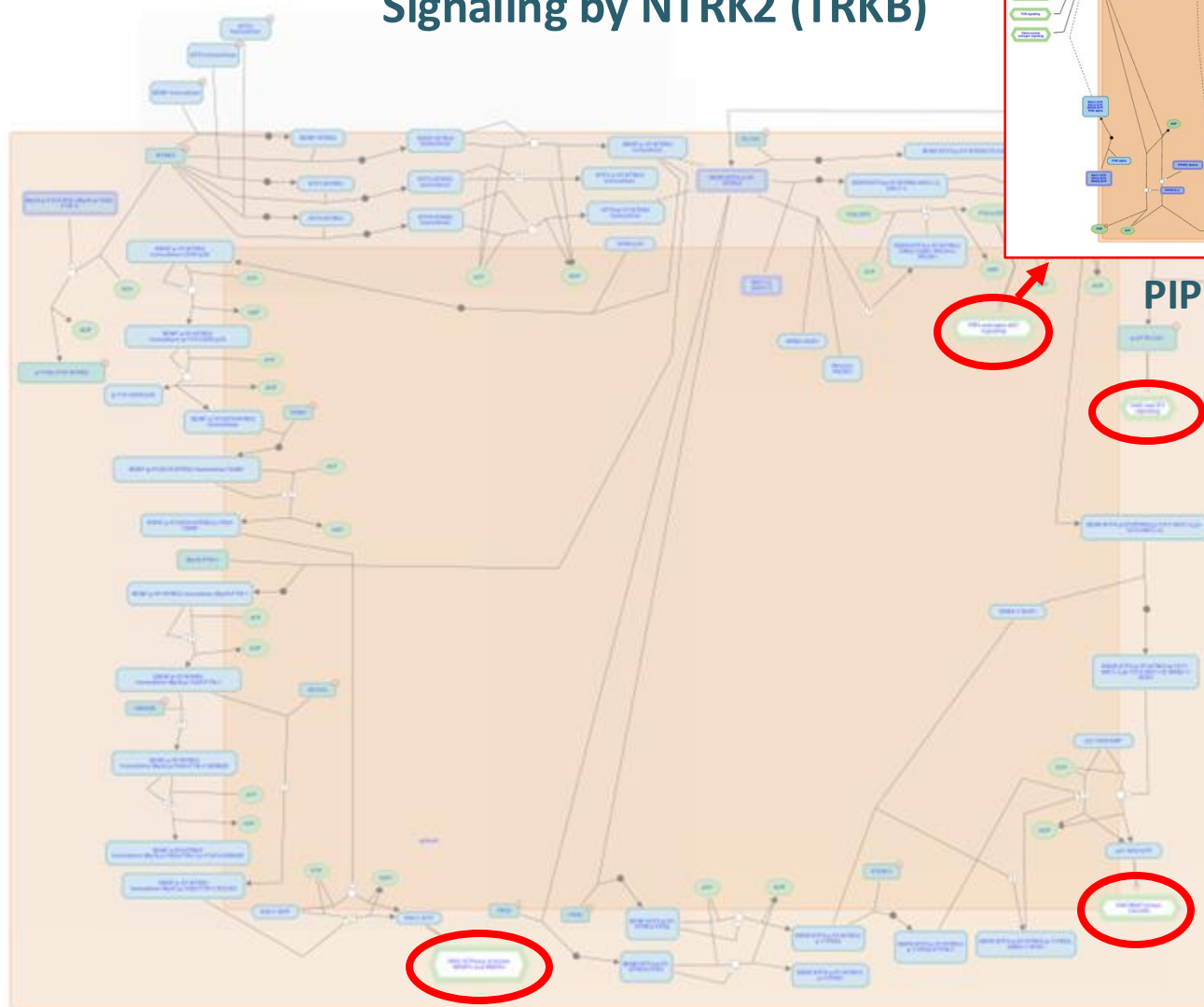
(NeuroTrophic Receptor Kinase 2; Tyrosine Protein Kinase))



Signaling network reconstruction

3. Expand the list of pathways selection

Signaling by NTRK2 (TRKB)



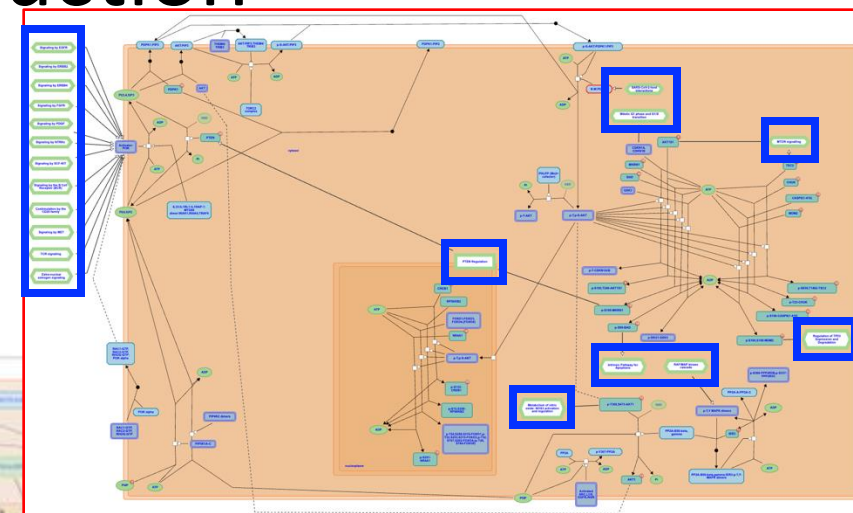
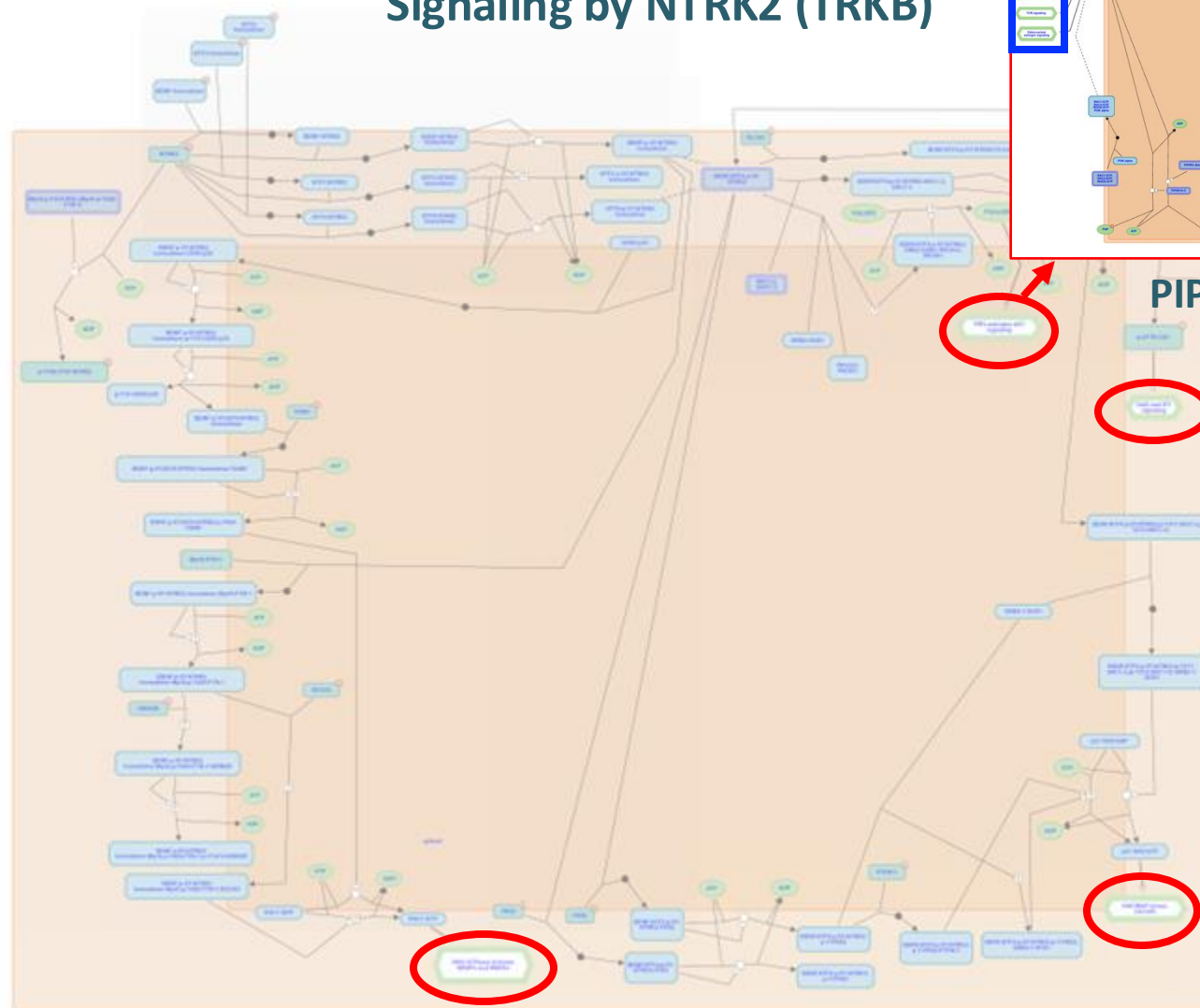
PIP3 activates AKT signaling

PI3K (phosphatidylinositol 3-kinase)
and Akt (protein kinase B)

Signaling network reconstruction

3. Expand the list of pathways selection

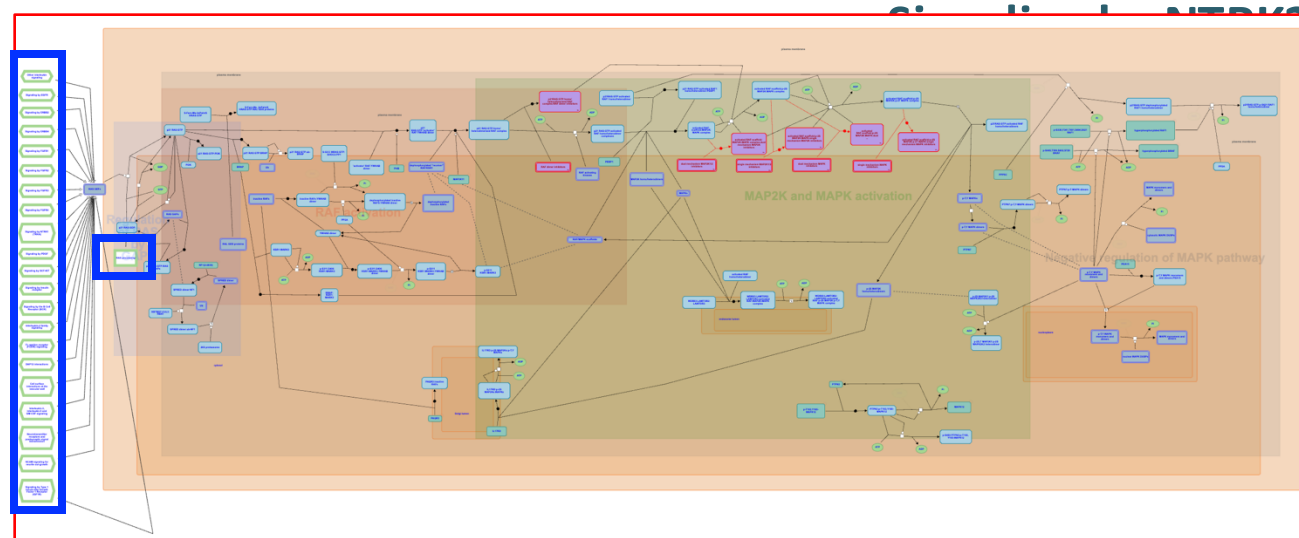
Signaling by NTRK2 (TRKB)



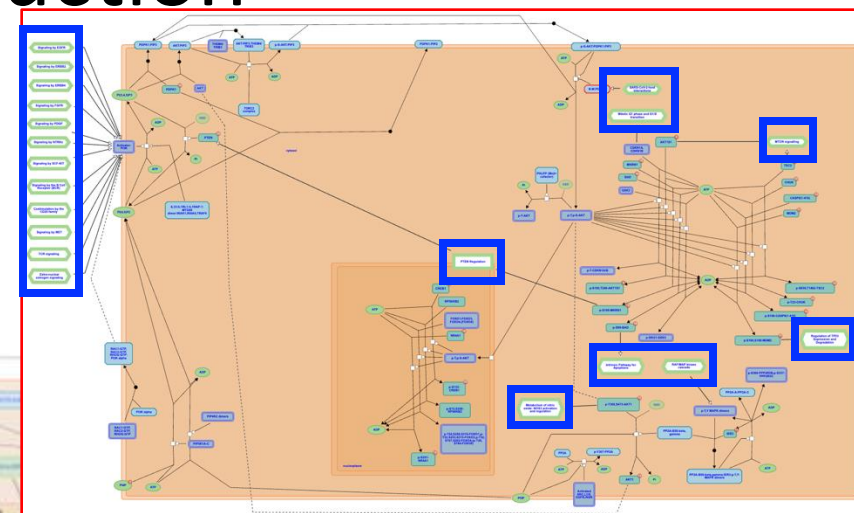
PIP3 activates AKT signaling

Signaling network reconstruction

3. Expand the list of pathways selection

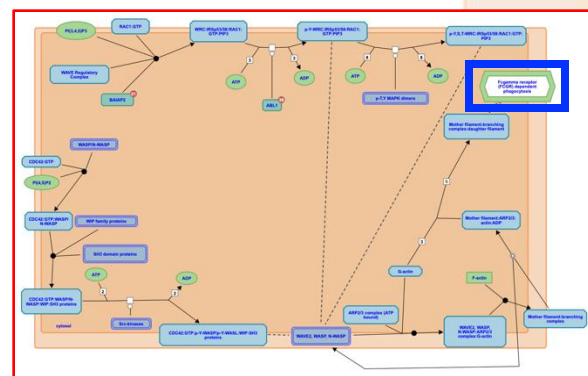


(TRKB)

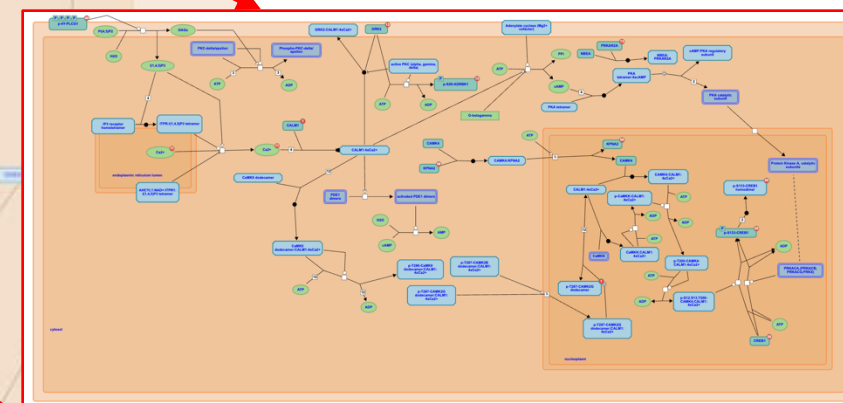


PIP3 activates AKT signaling

RAF/MAP kinase cascade



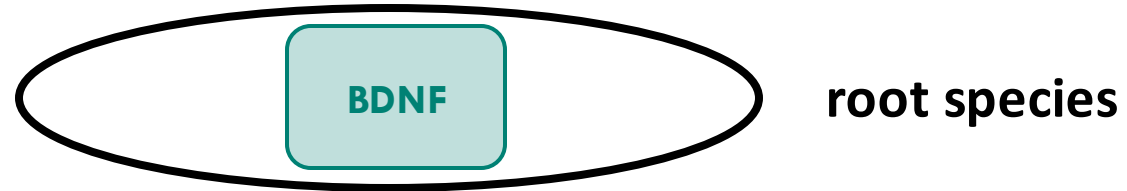
RHO GTPases Activate
WASPs and WAVES



DAG and IP3 signaling

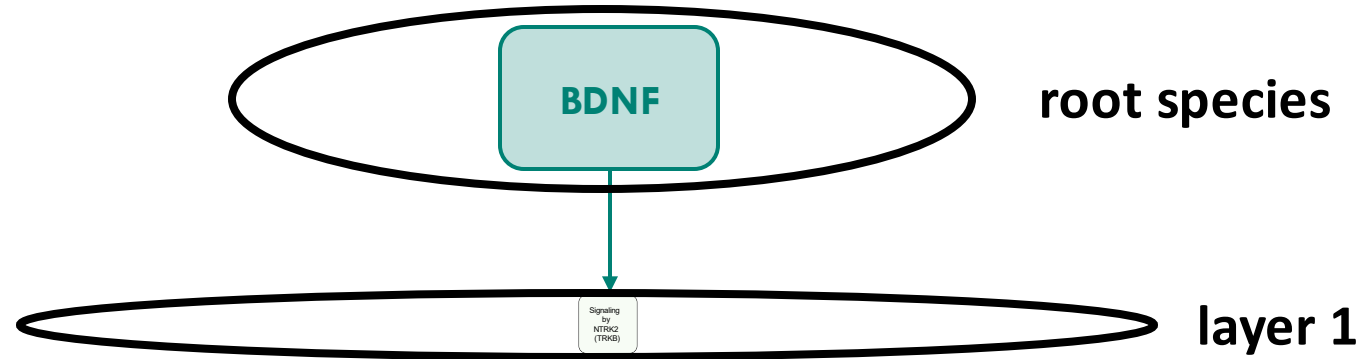
Signaling network reconstruction

3. Expand the list of pathways selection



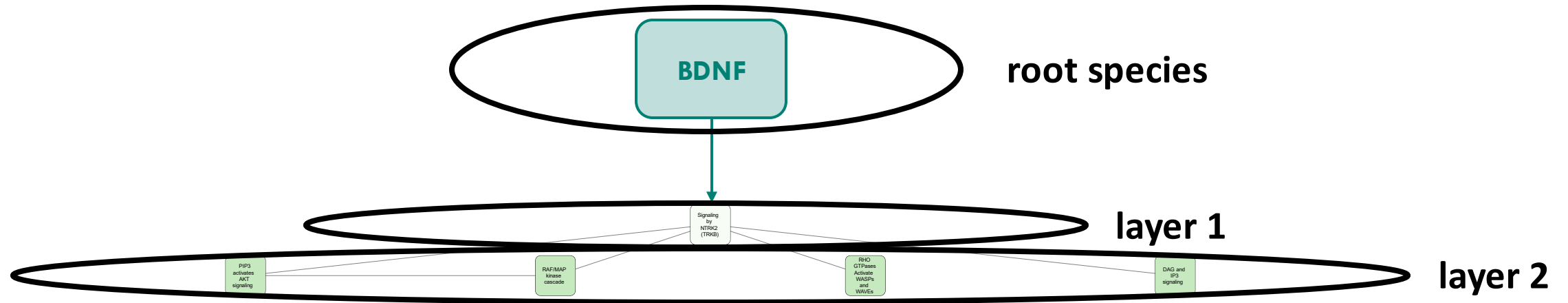
Signaling network reconstruction

3. Expand the list of pathways selection



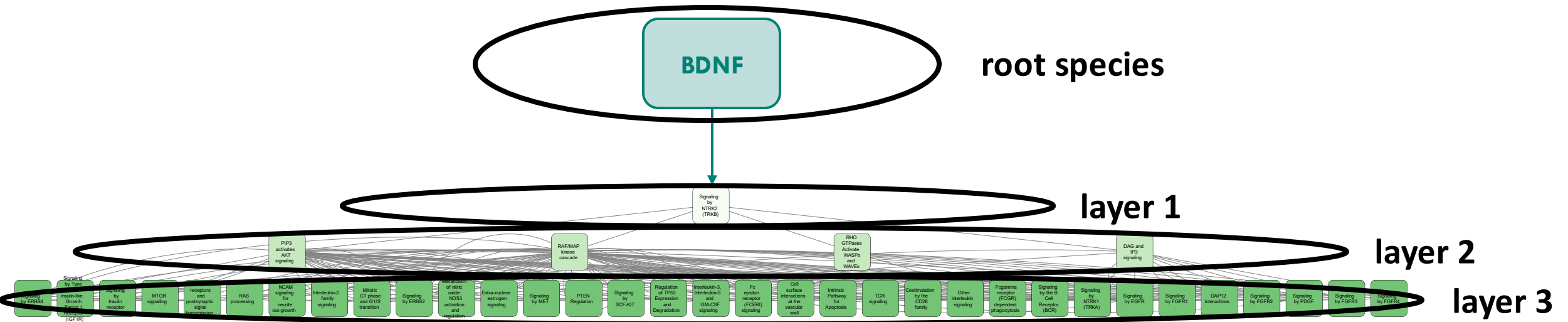
Signaling network reconstruction

3. Expand the list of pathways selection

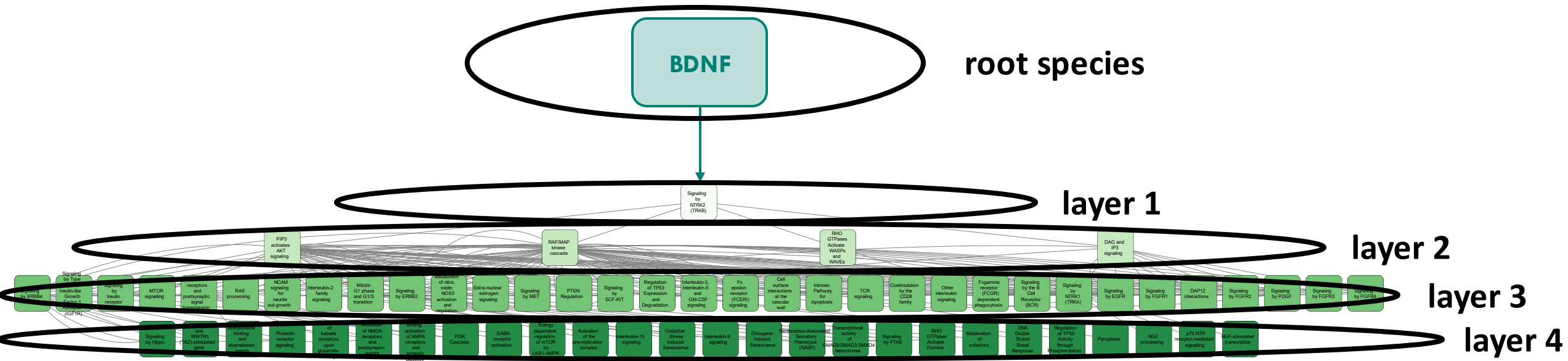


Signaling network reconstruction

3. Expand the list of pathways selection

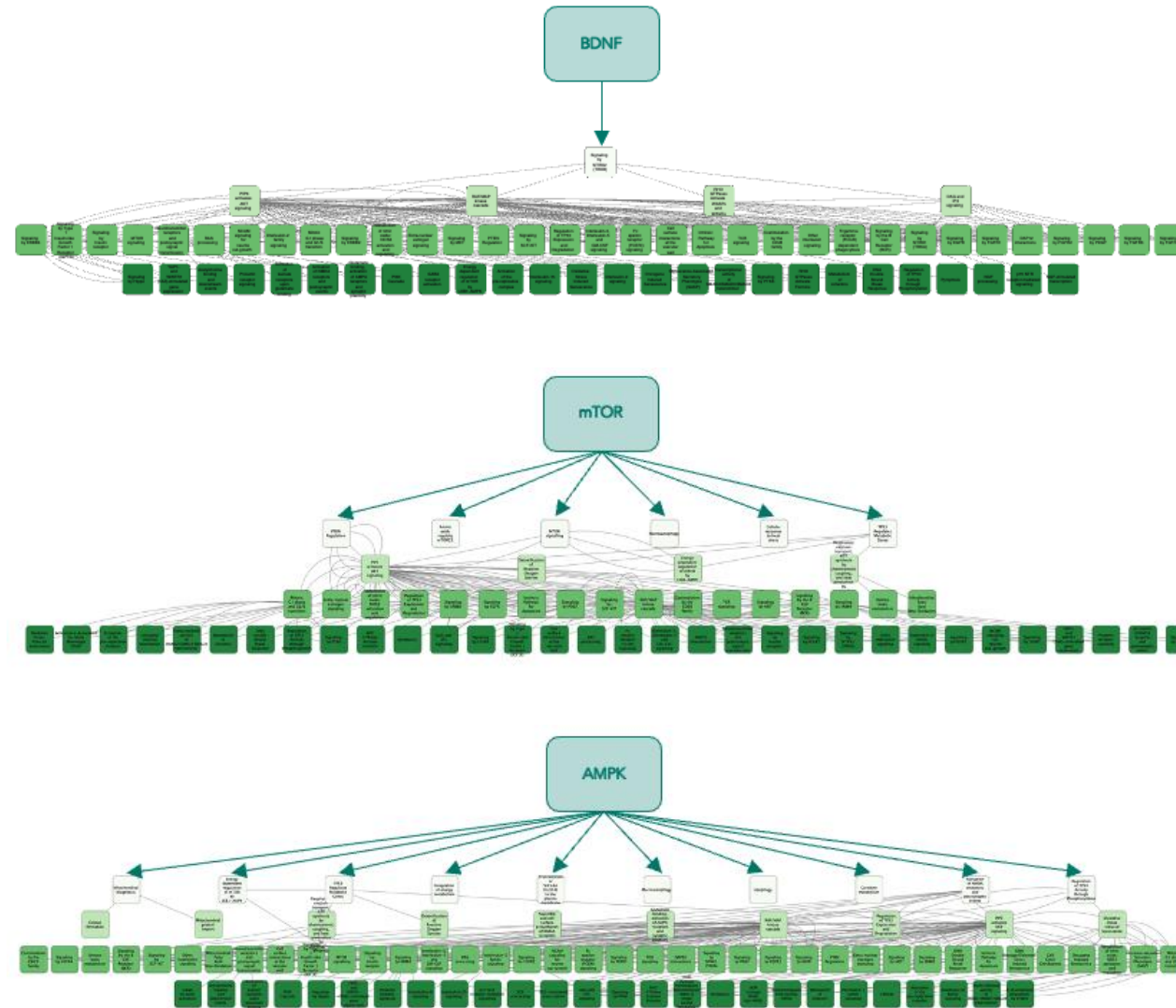


3. Expand the list of pathways selection



Signaling network reconstruction

3. Expand the list of pathways selection

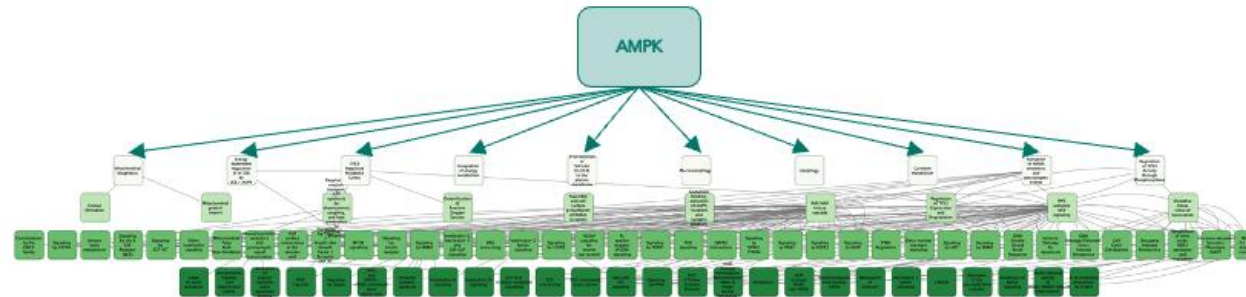
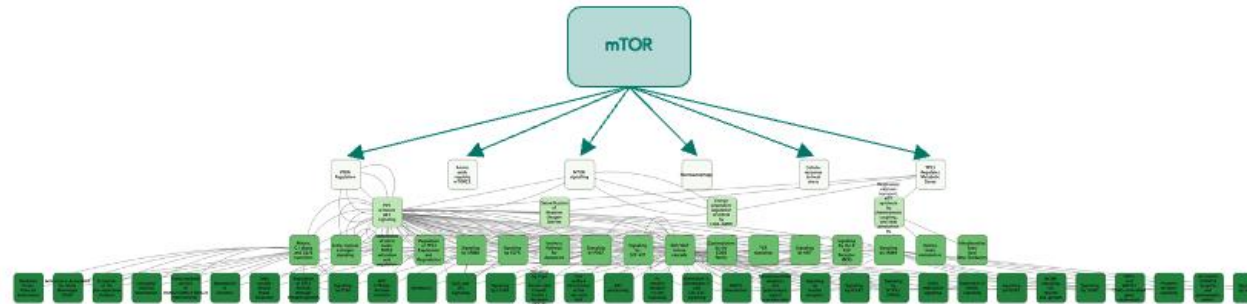
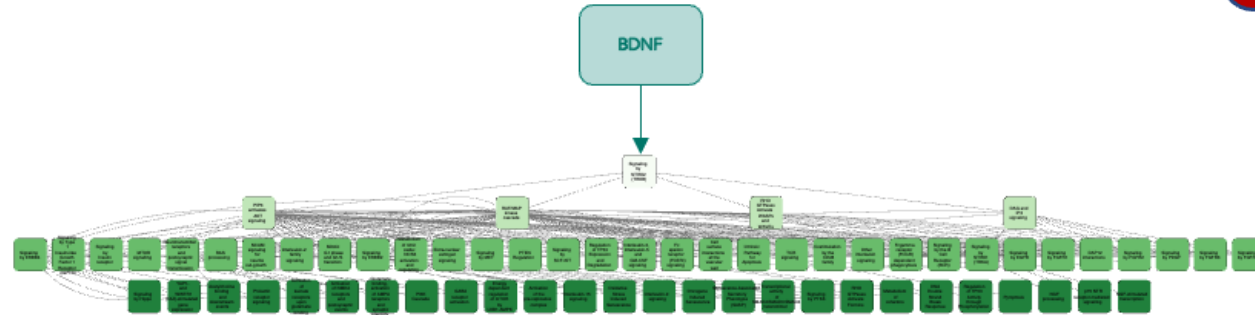


Signaling network reconstruction

3. Expand the list of pathways selection



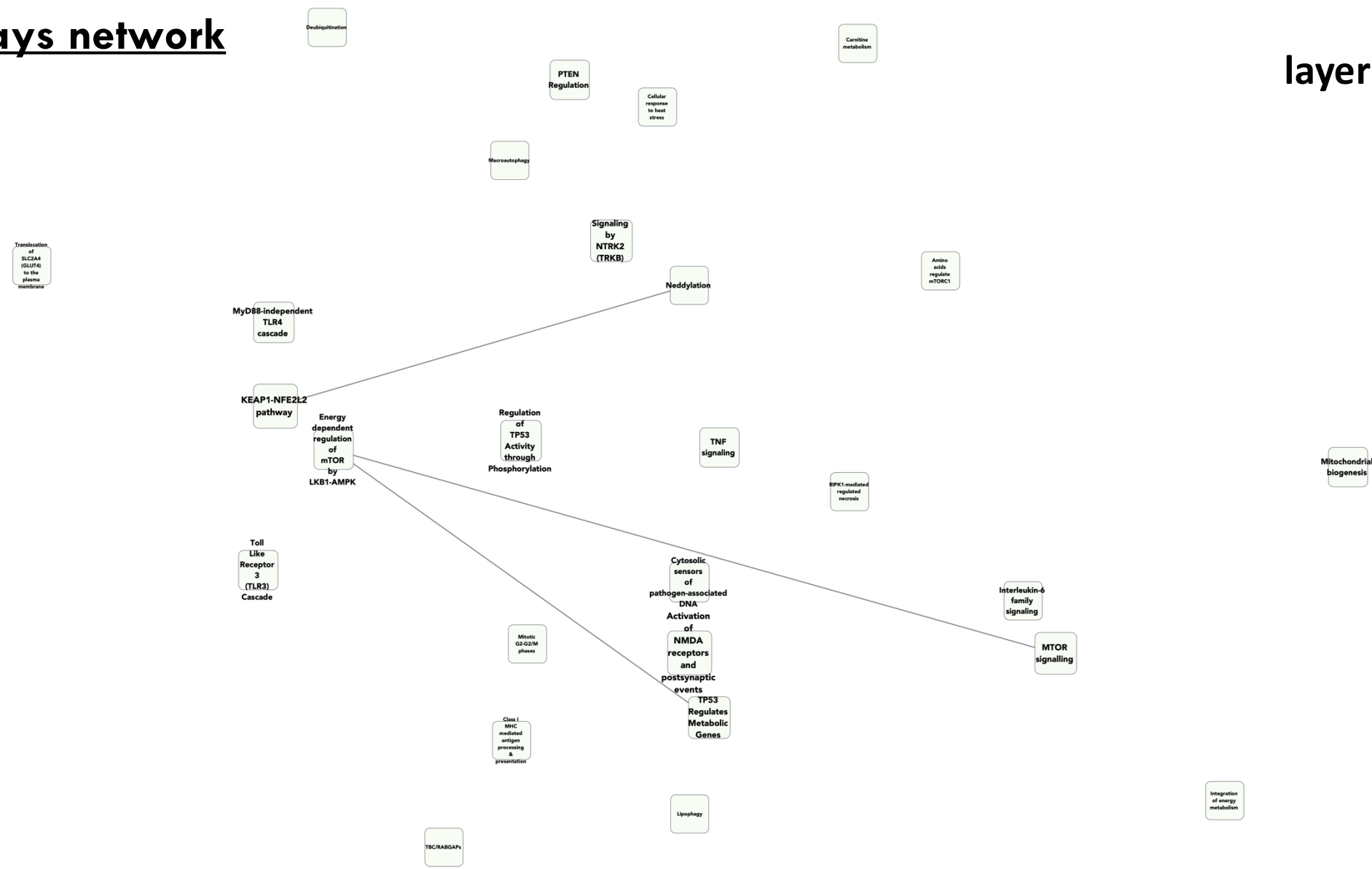
115 Pathways



Signaling network reconstruction

Final pathways network

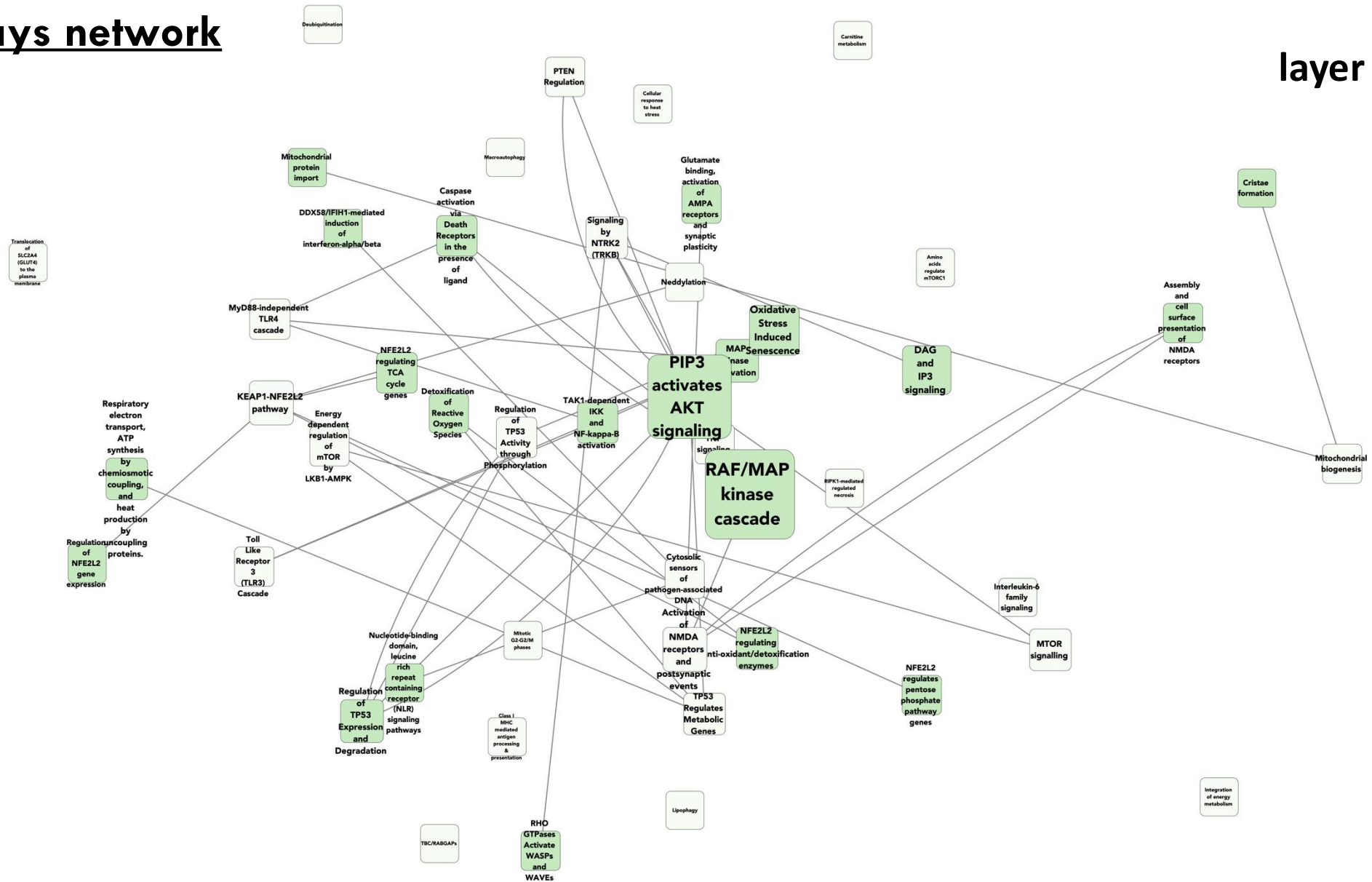
layer 1



Signaling network reconstruction

Final pathways network

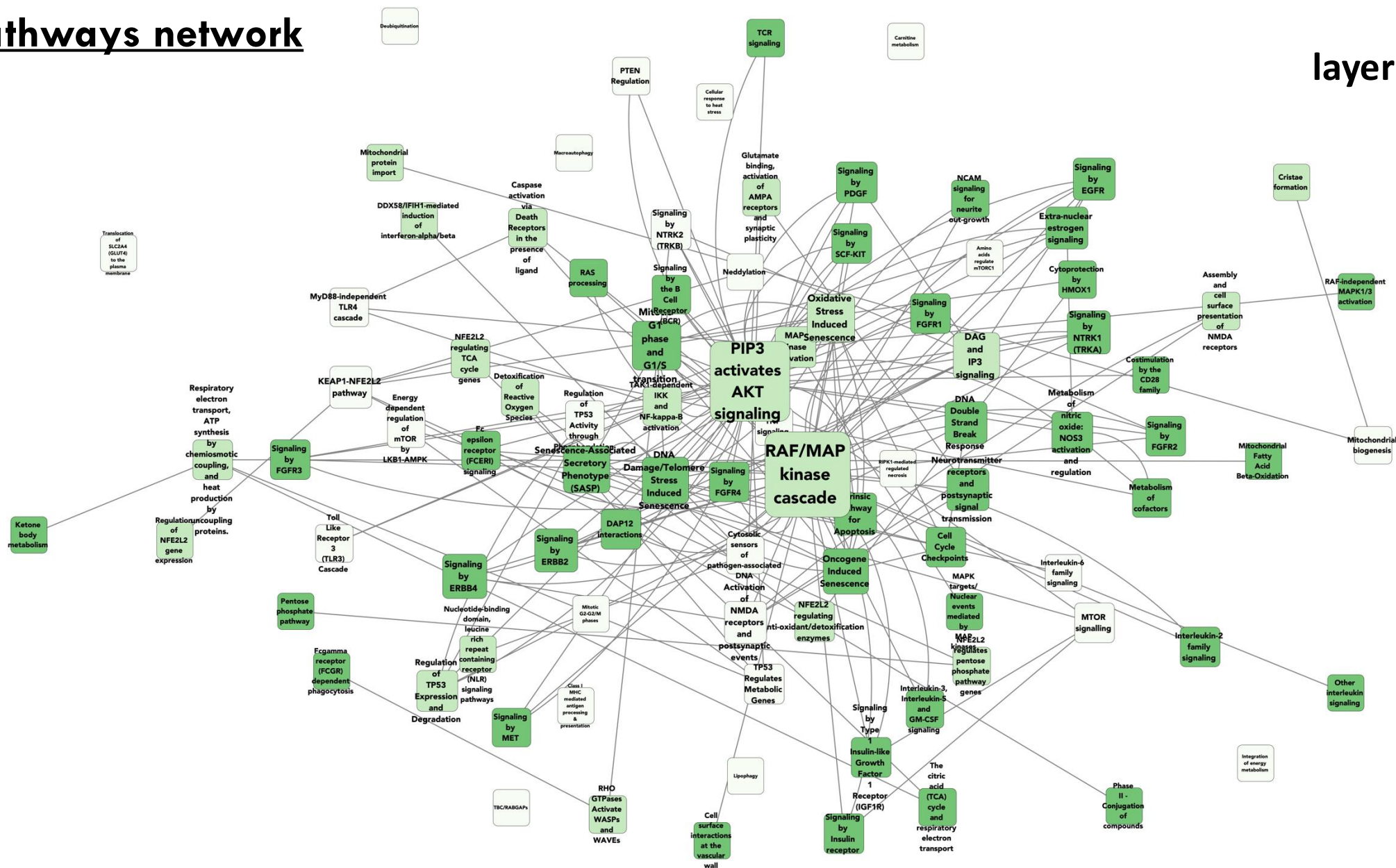
layer 2



Signaling network reconstruction

Final pathways network

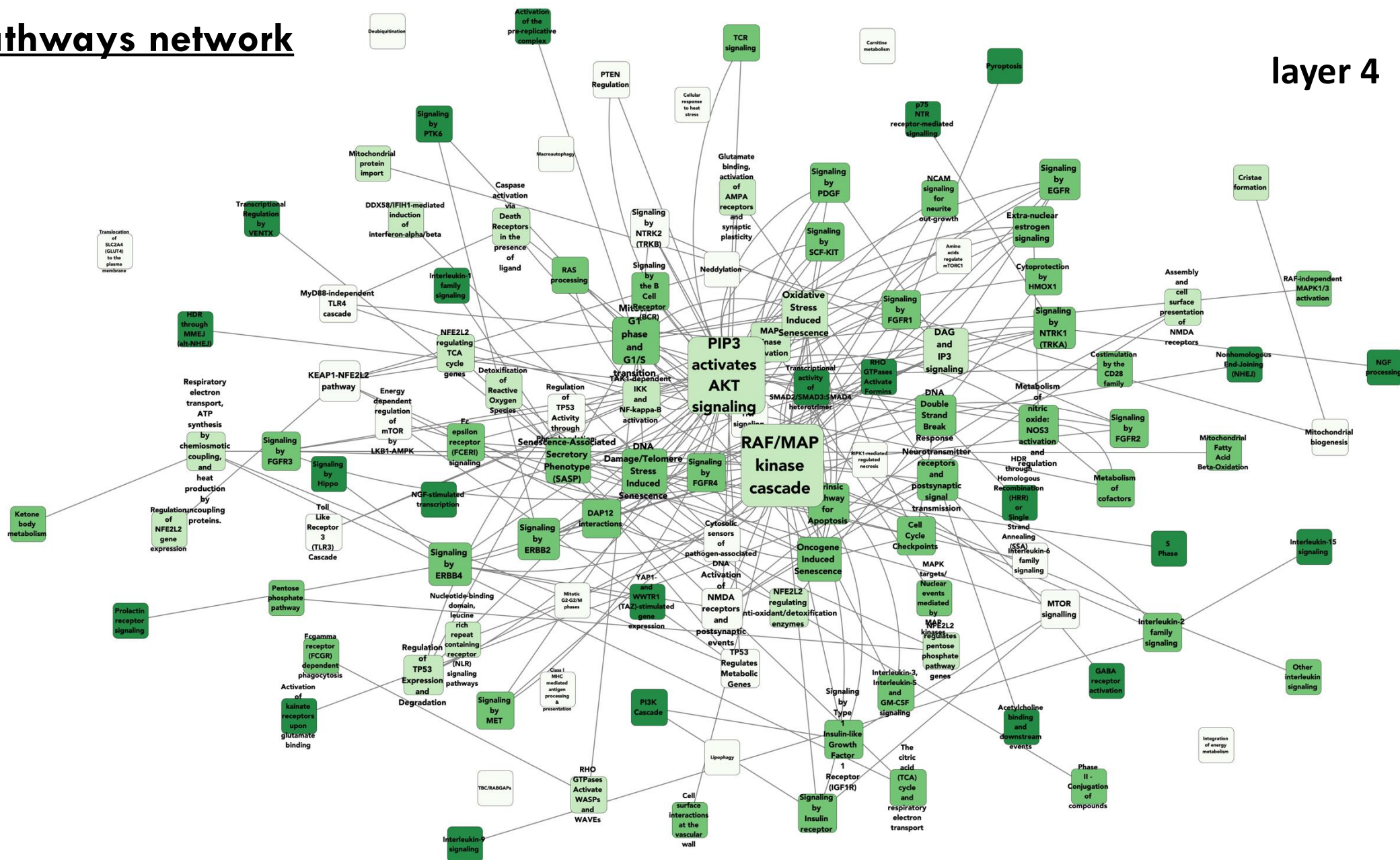
layer 3



Signaling network reconstruction

Final pathways network

layer 4



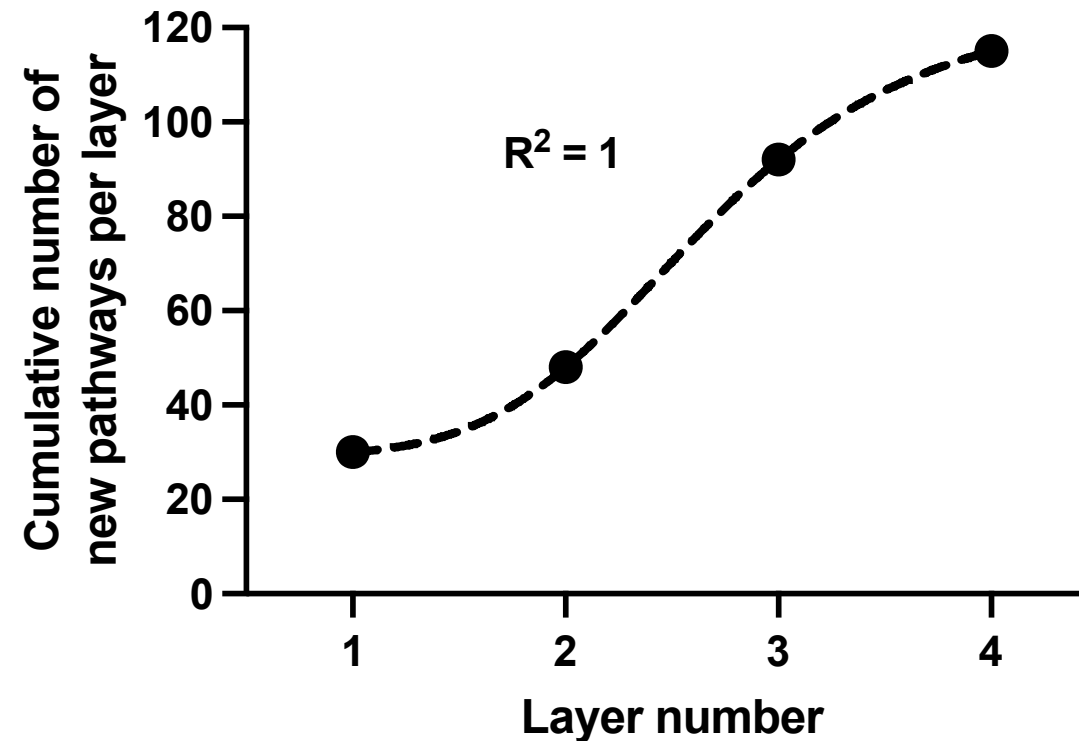
Signaling network reconstruction

Are 4 layers sufficient?

Signaling network reconstruction

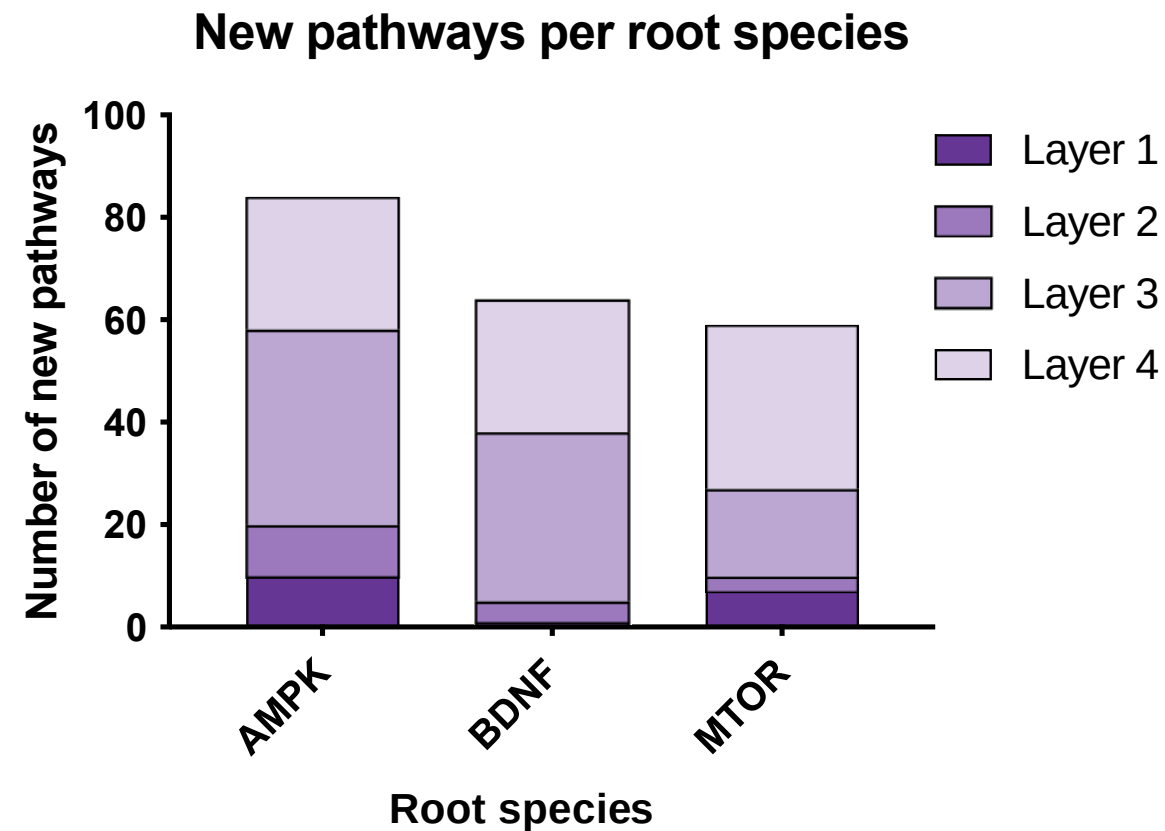
3. Expand the list of pathways selection

Number of pathways saturates as we increase the layers



Signaling network reconstruction

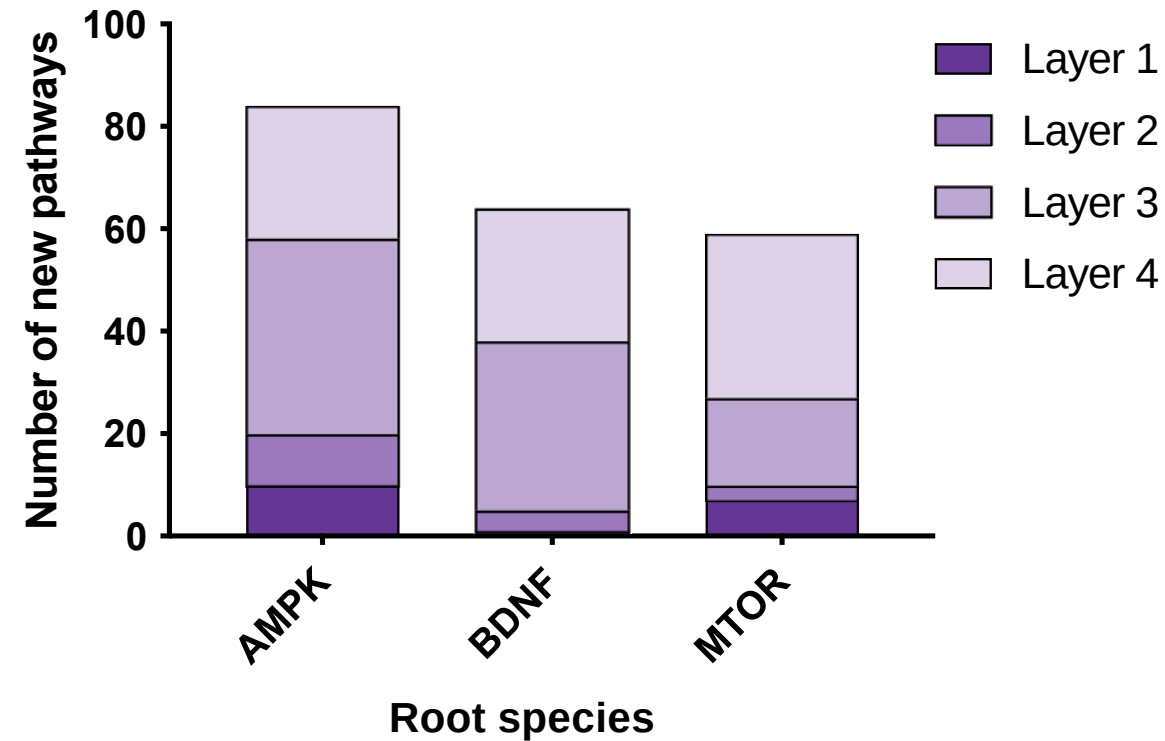
3. Expand the list of pathways selection



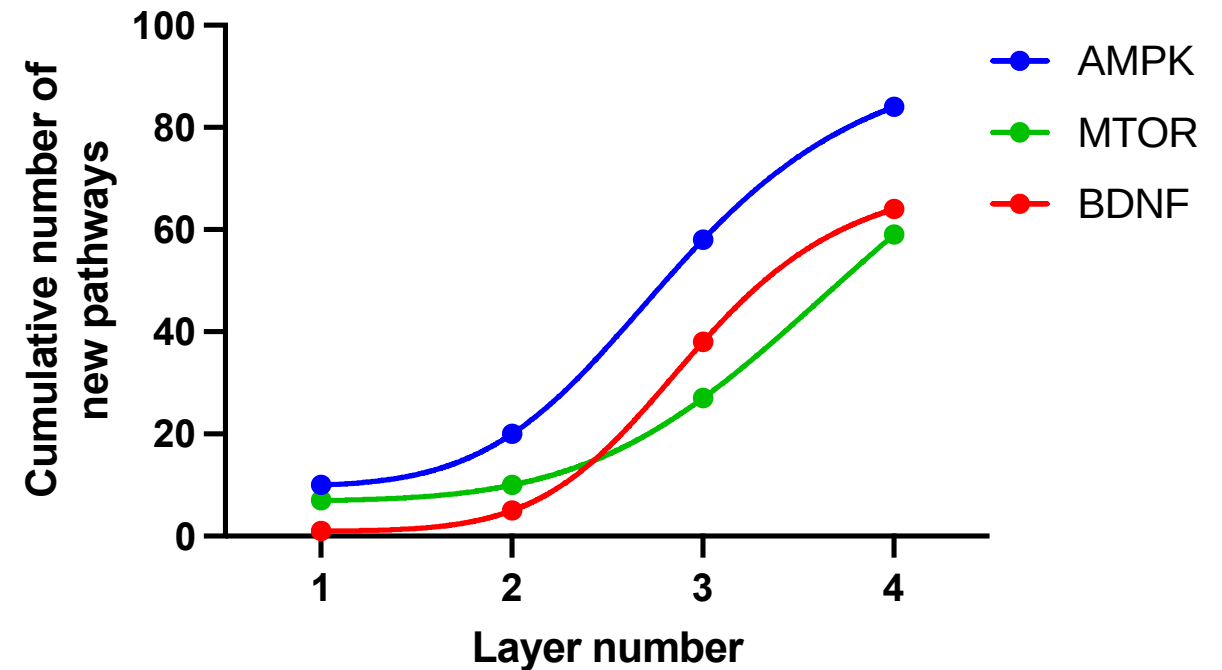
Signaling network reconstruction

3. Expand the list of pathways selection

New pathways per root species



New pathways per root species (cumulative)



Signaling network reconstruction

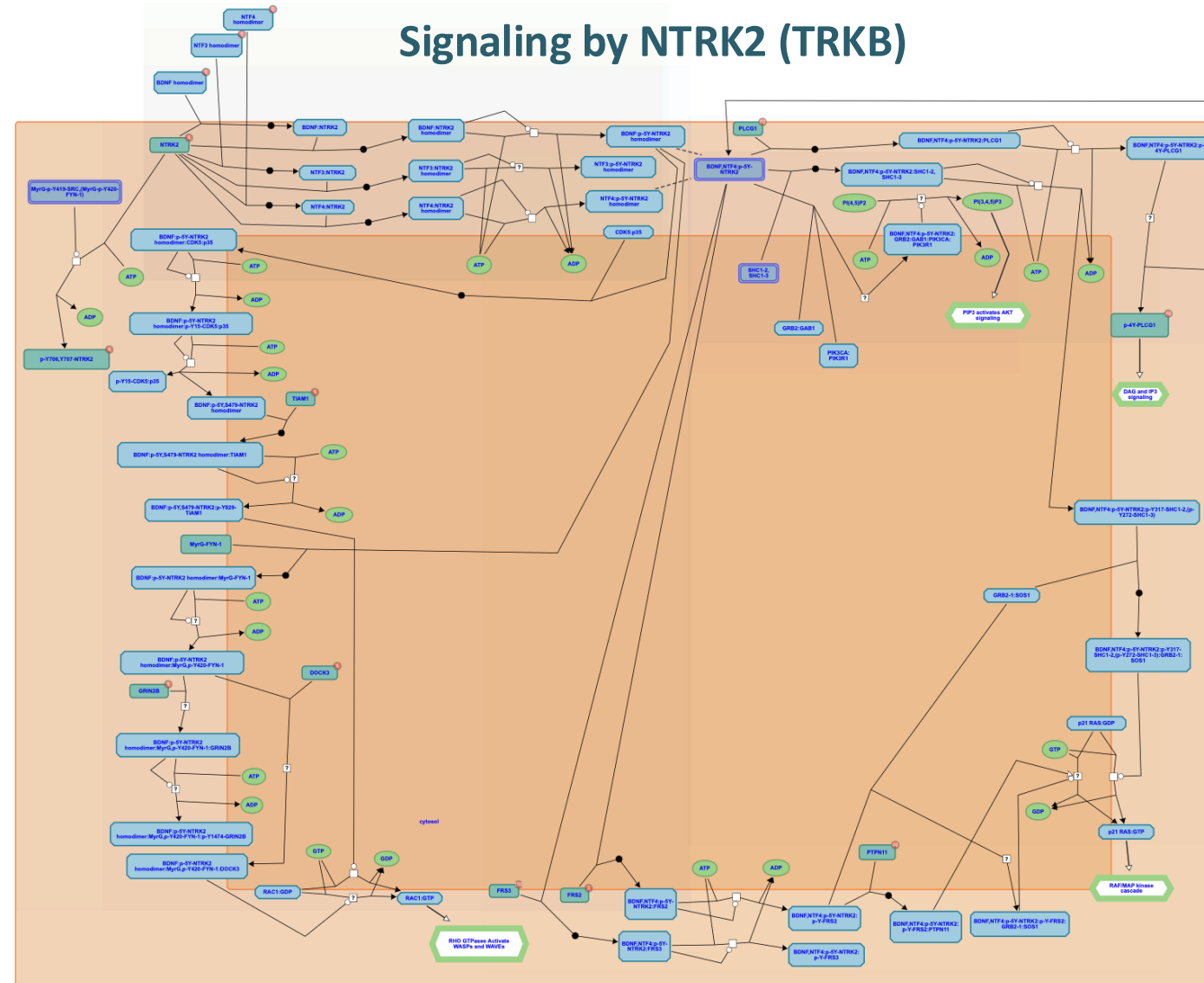
4. Curate manually all the pathways of the final list

Signaling network reconstruction

4. Curate manually all the pathways of the final list

Reactions and species information
can be downloaded automatically
but:

Signaling by NTRK2 (TRKB)



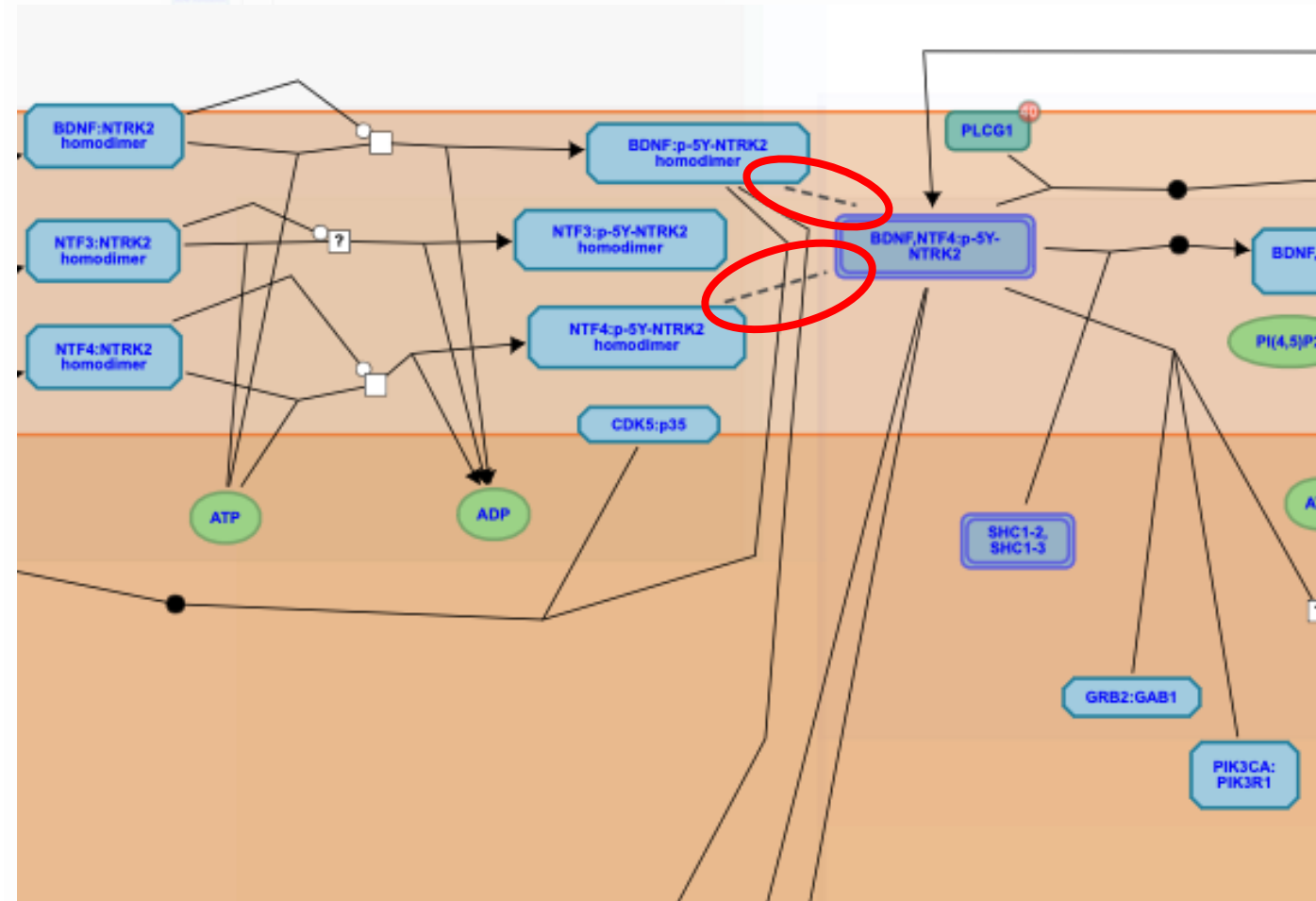
Signaling network reconstruction

4. Curate manually all the pathways of the final list

Reactions and species information can be downloaded automatically but:

1) Dashed lines in the graph represent flow of information but the data does not exist.

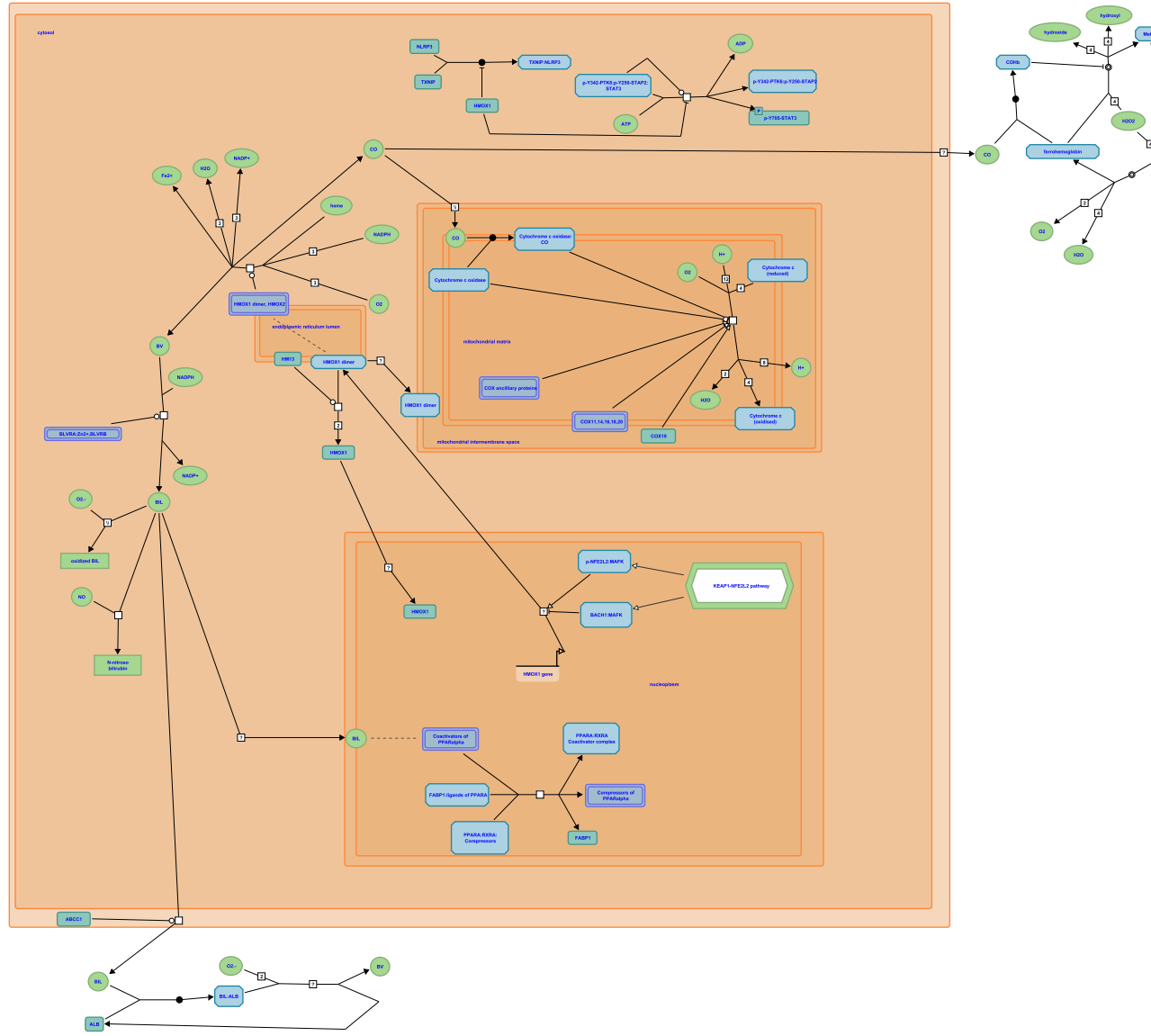
Signaling by NTRK2 (TRKB)



Signaling network reconstruction

4. Curate manually all the pathways of the final list

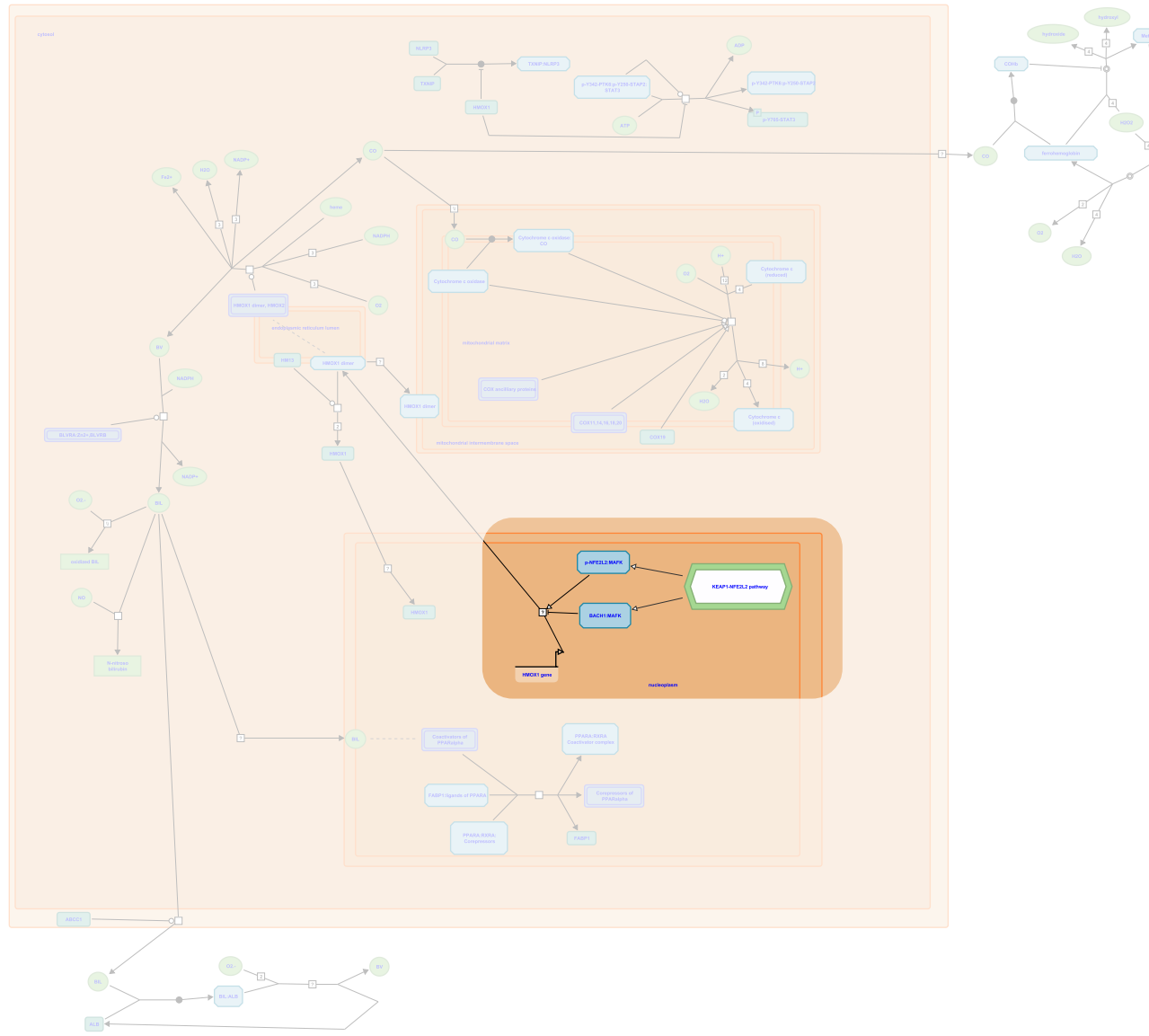
Cytoprotection by HMOX1



Signaling network reconstruction

4. Curate manually all the pathways of the final list

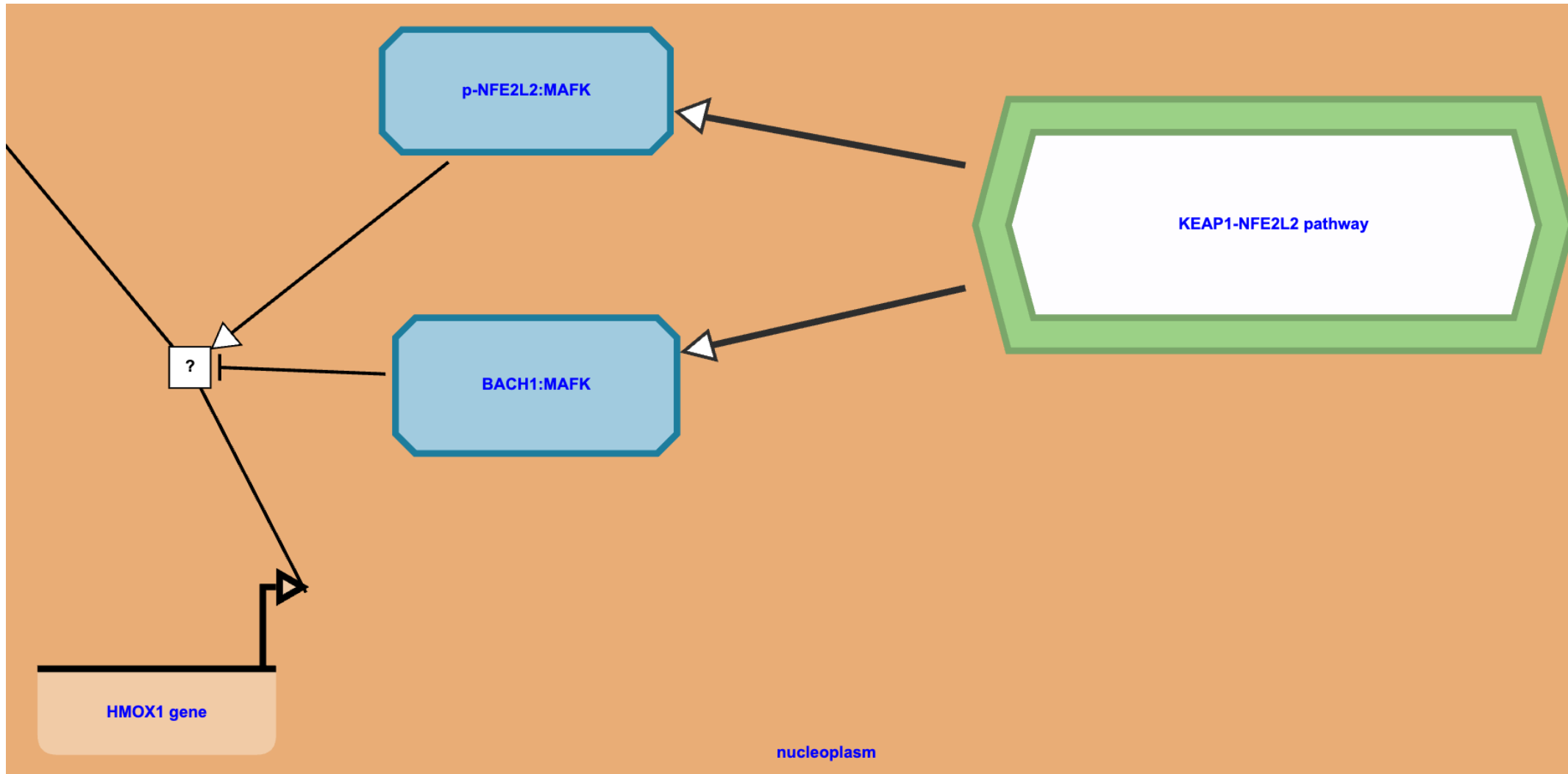
Cytoprotection by HMOX1



Signaling network reconstruction

4. Curate manually all the pathways of the final list

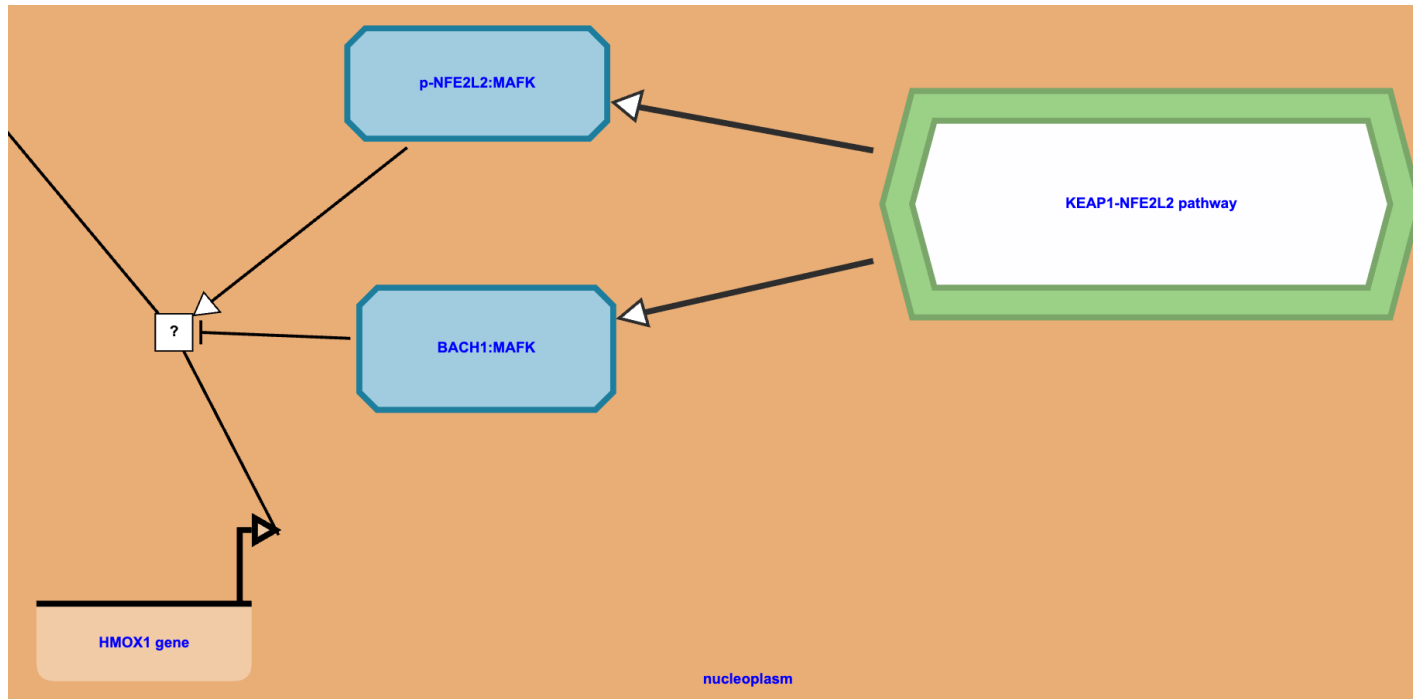
Cytoprotection by HMOX1



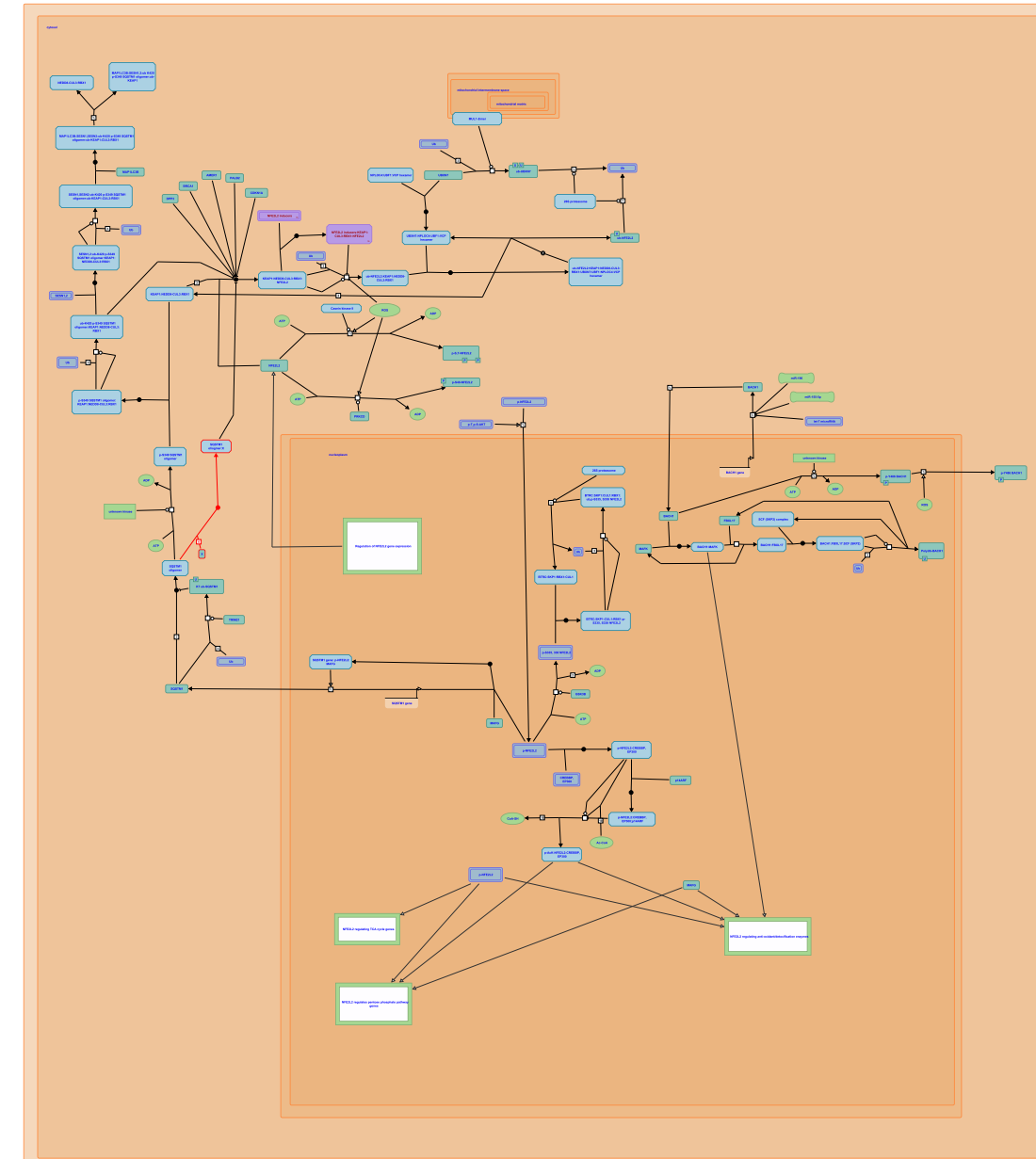
Signaling network reconstruction

4. Curate manually all the pathways of the final list

Cytoprotection by HMOX1



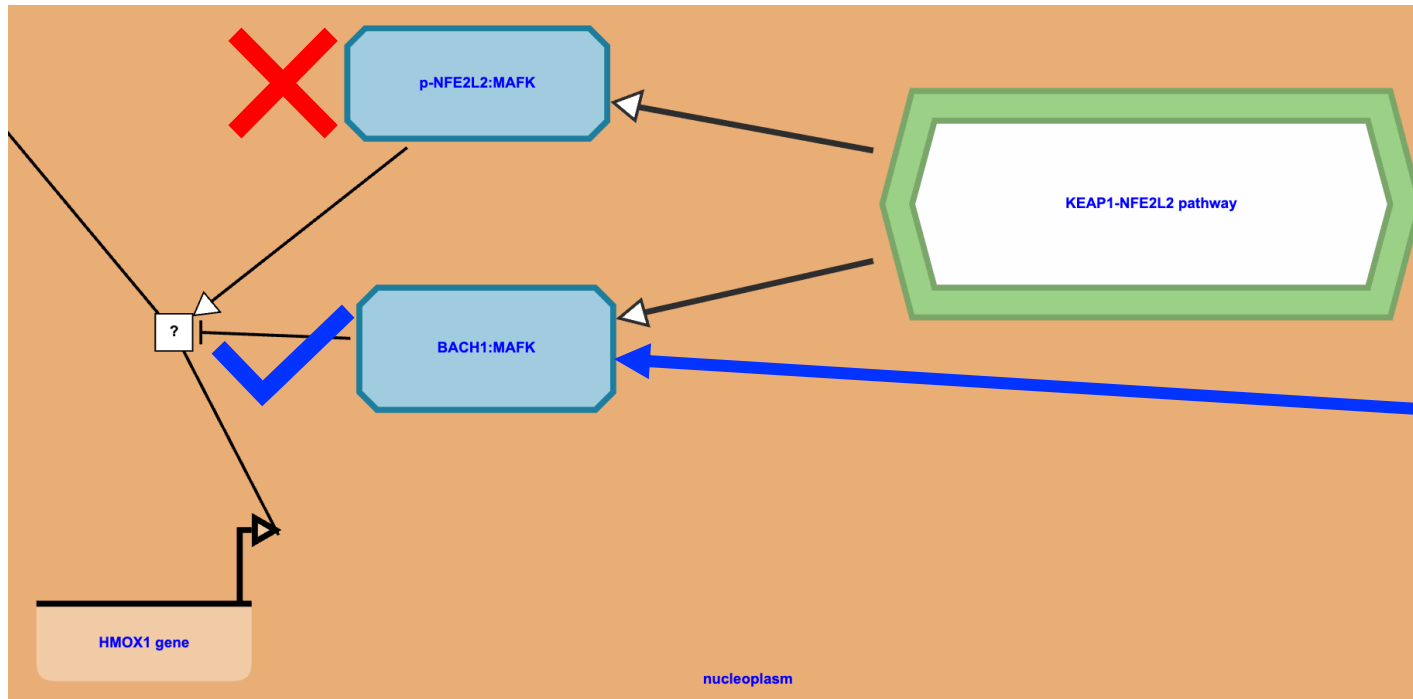
KEAP1-NFE2L2 pathway



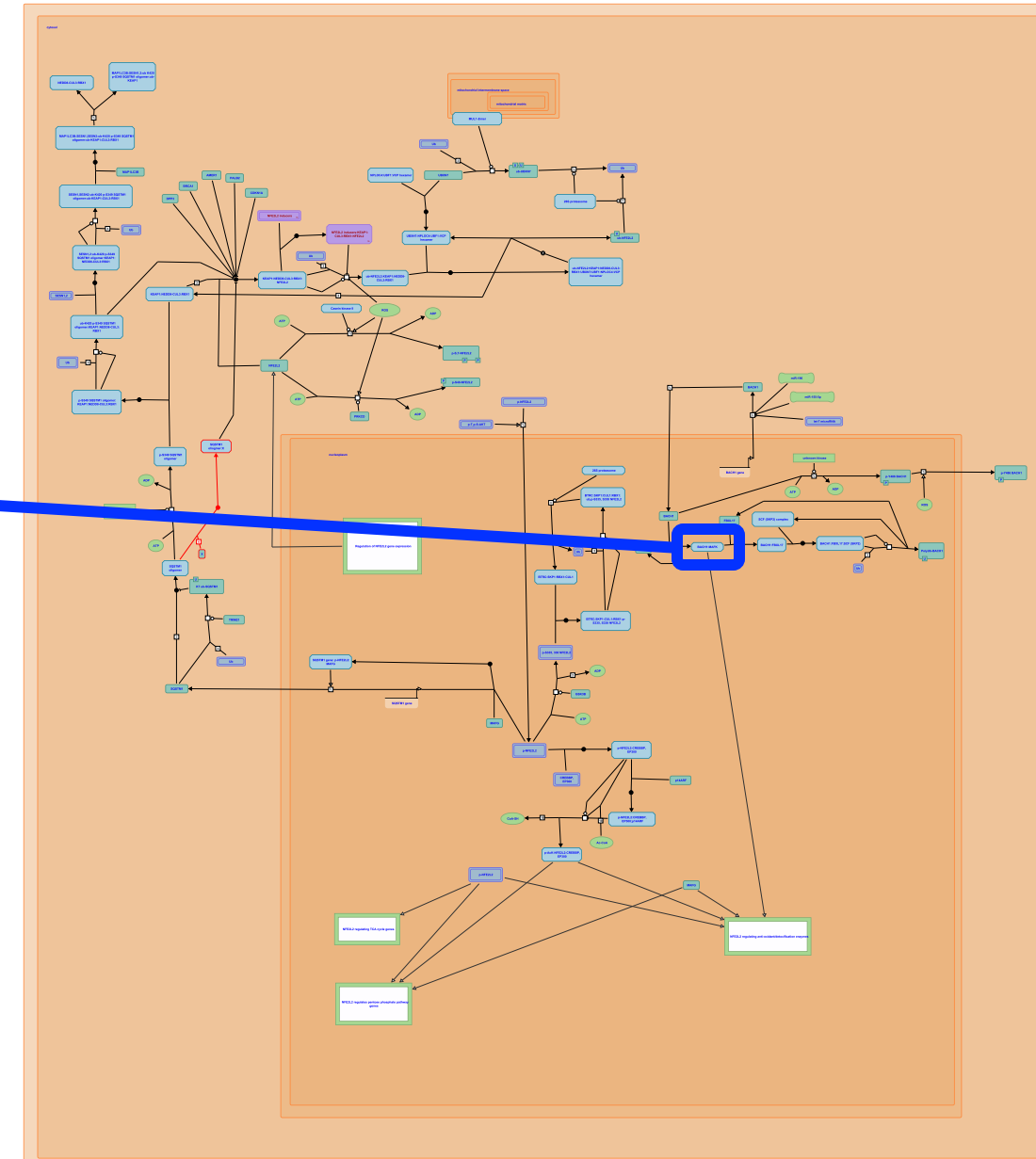
Signaling network reconstruction

4. Curate manually all the pathways of the final list

Cytoprotection by HMOX1



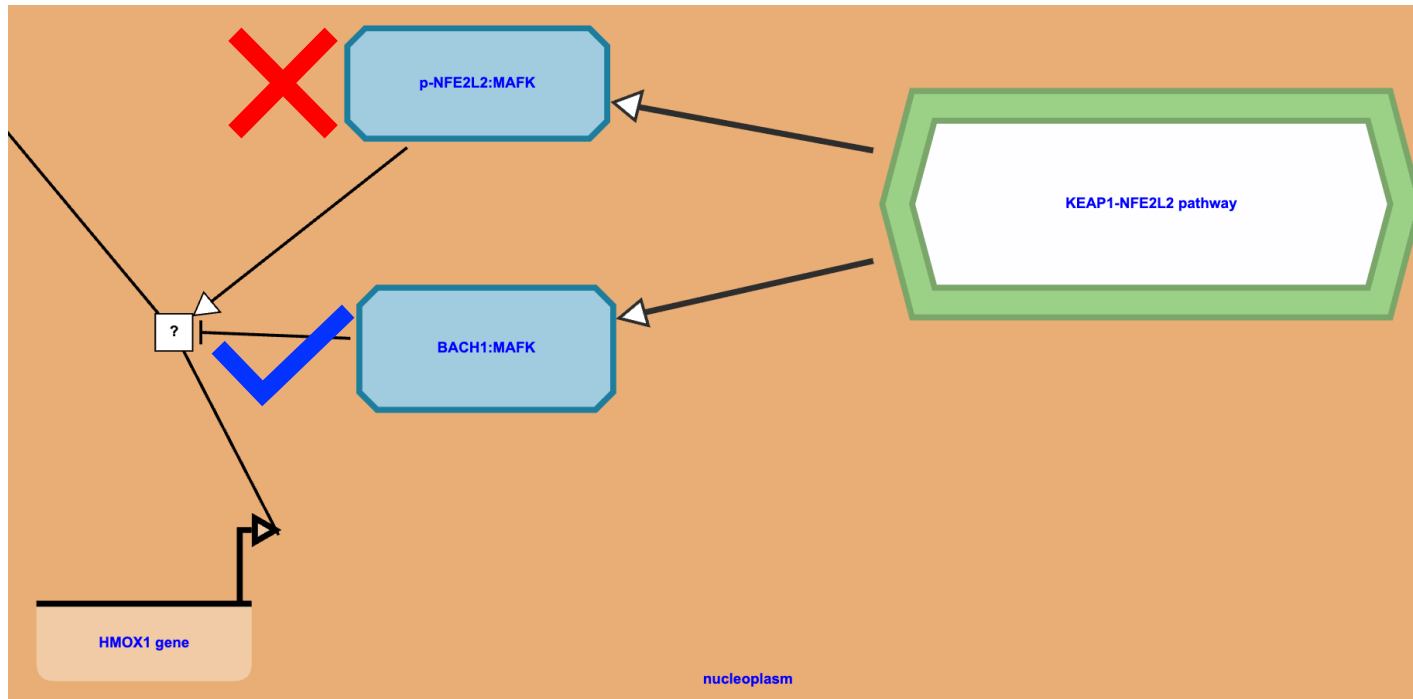
KEAP1-NFE2L2 pathway



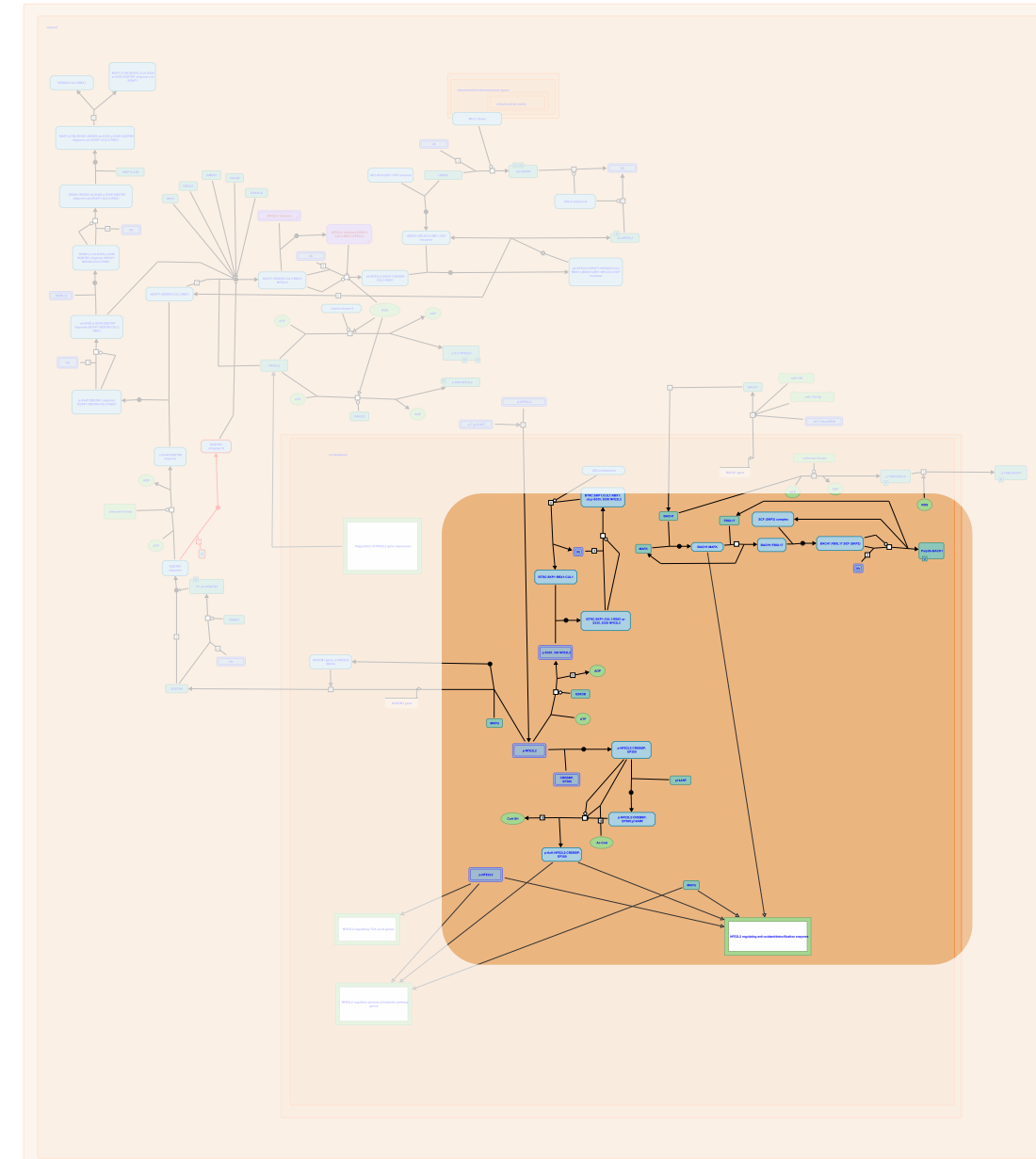
Signaling network reconstruction

4. Curate manually all the pathways of the final list

Cytoprotection by HMOX1



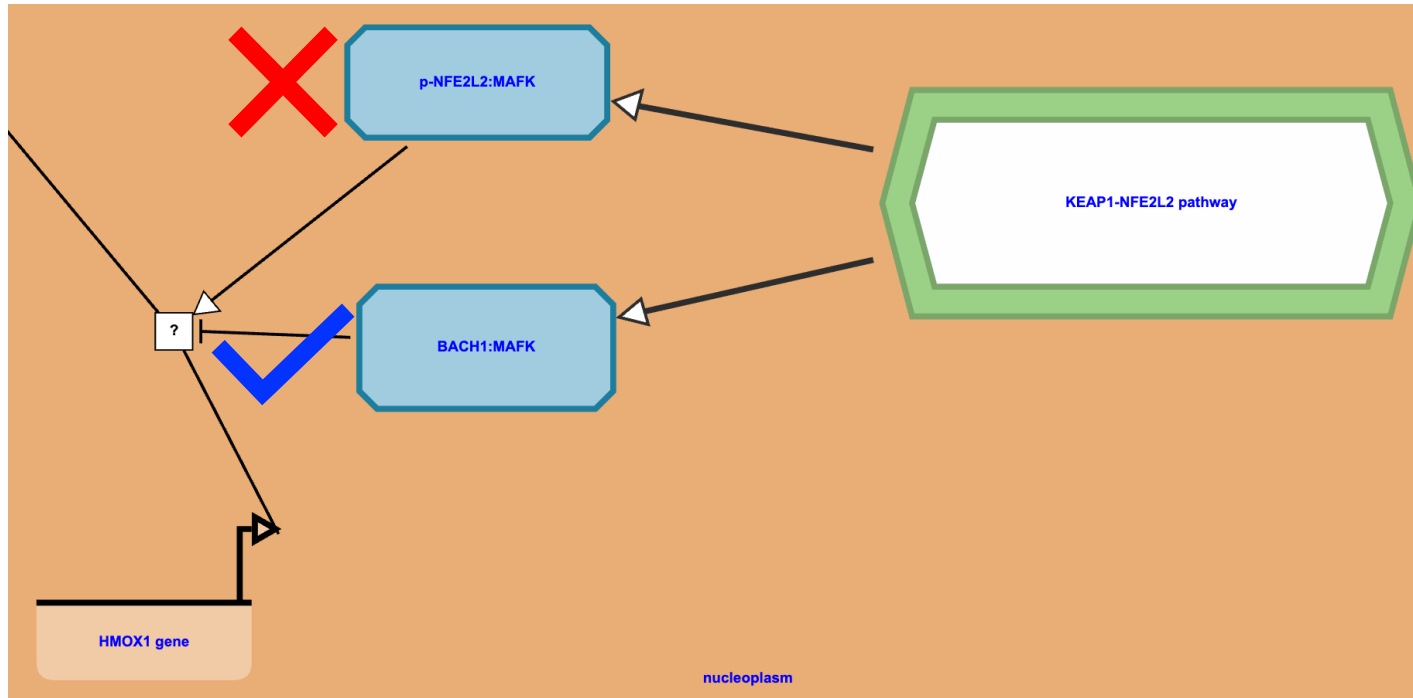
KEAP1-NFE2L2 pathway



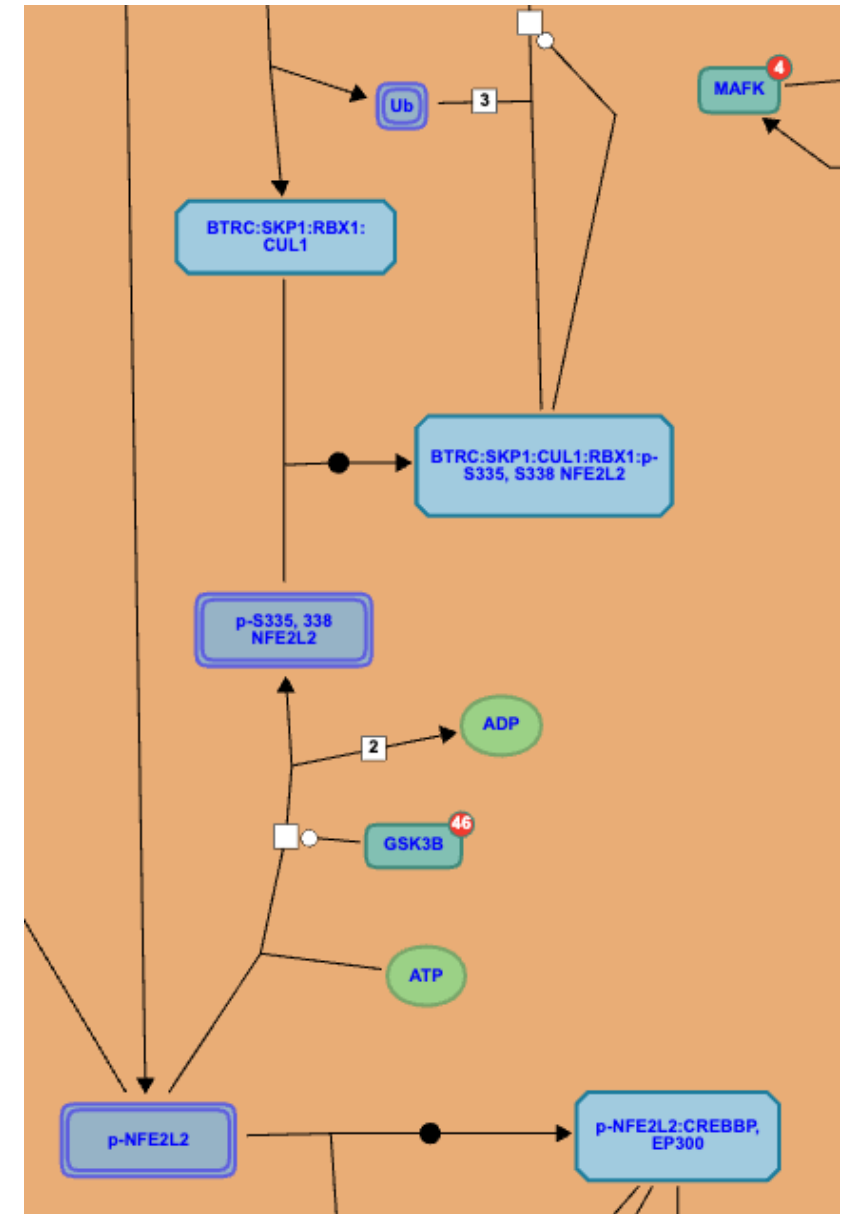
Signaling network reconstruction

4. Curate manually all the pathways of the final list

Cytoprotection by HMOX1



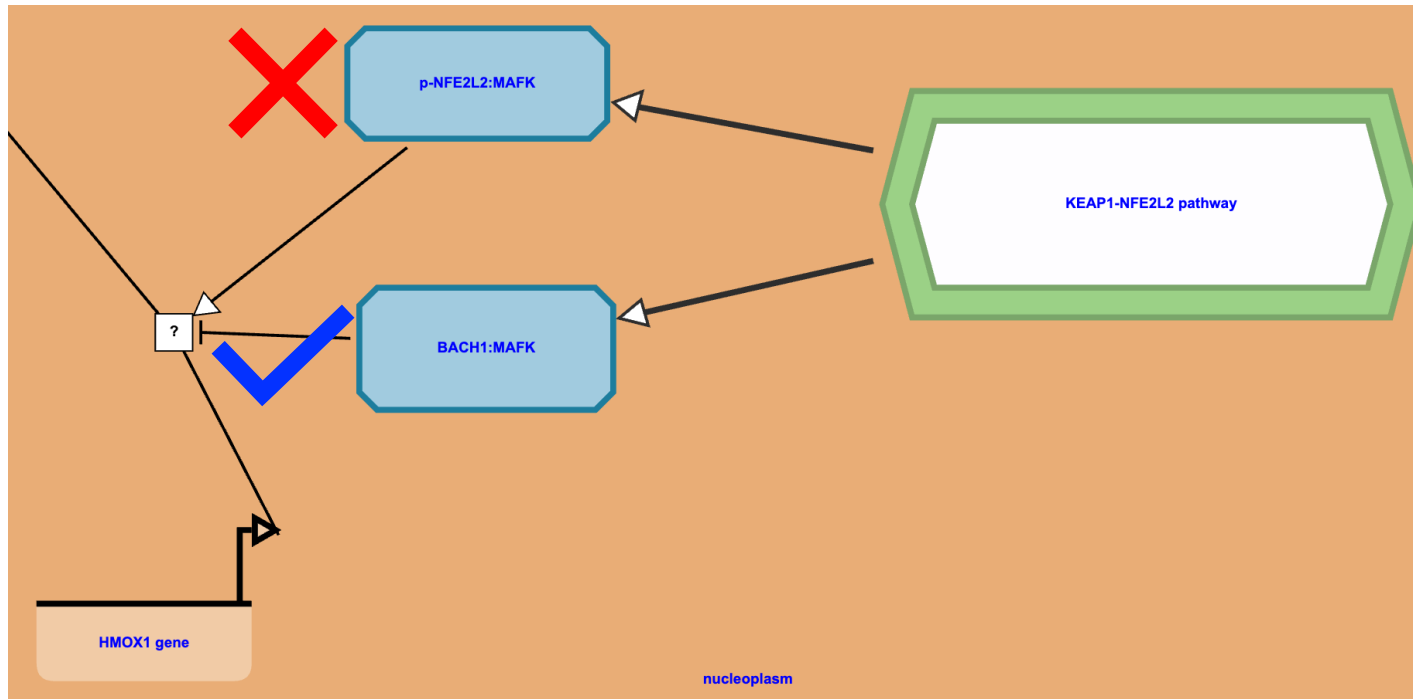
KEAP1-NFE2L2 pathway



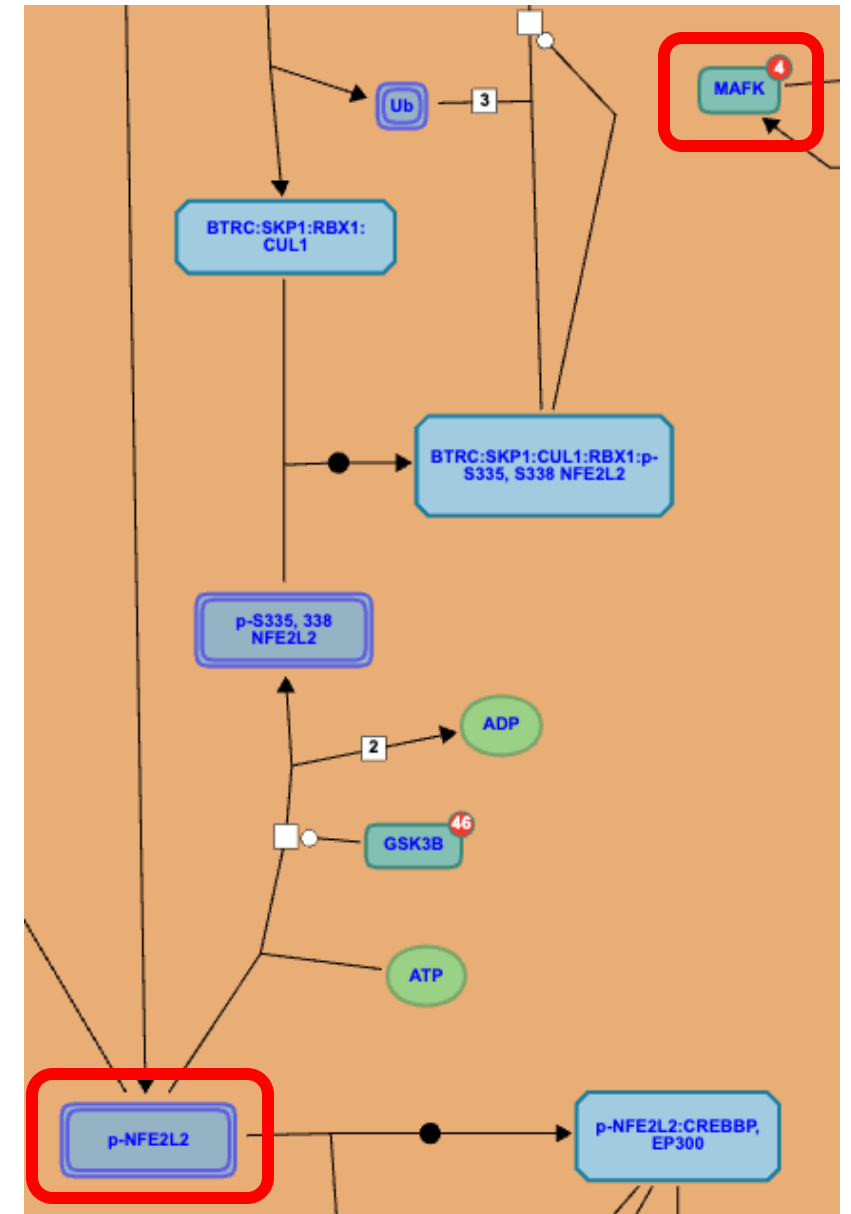
Signaling network reconstruction

4. Curate manually all the pathways of the final list

Cytoprotection by HMOX1



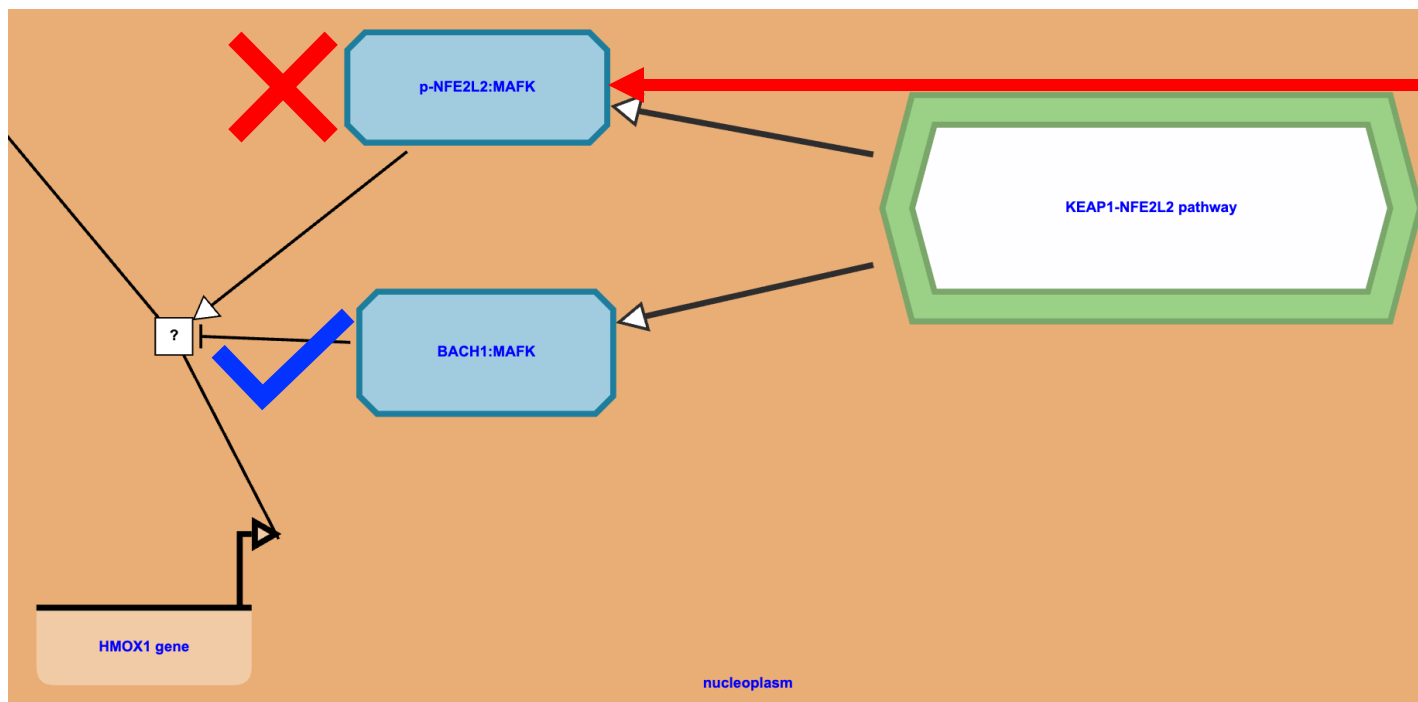
KEAP1-NFE2L2 pathway



Signaling network reconstruction

4. Curate manually all the pathways of the final list

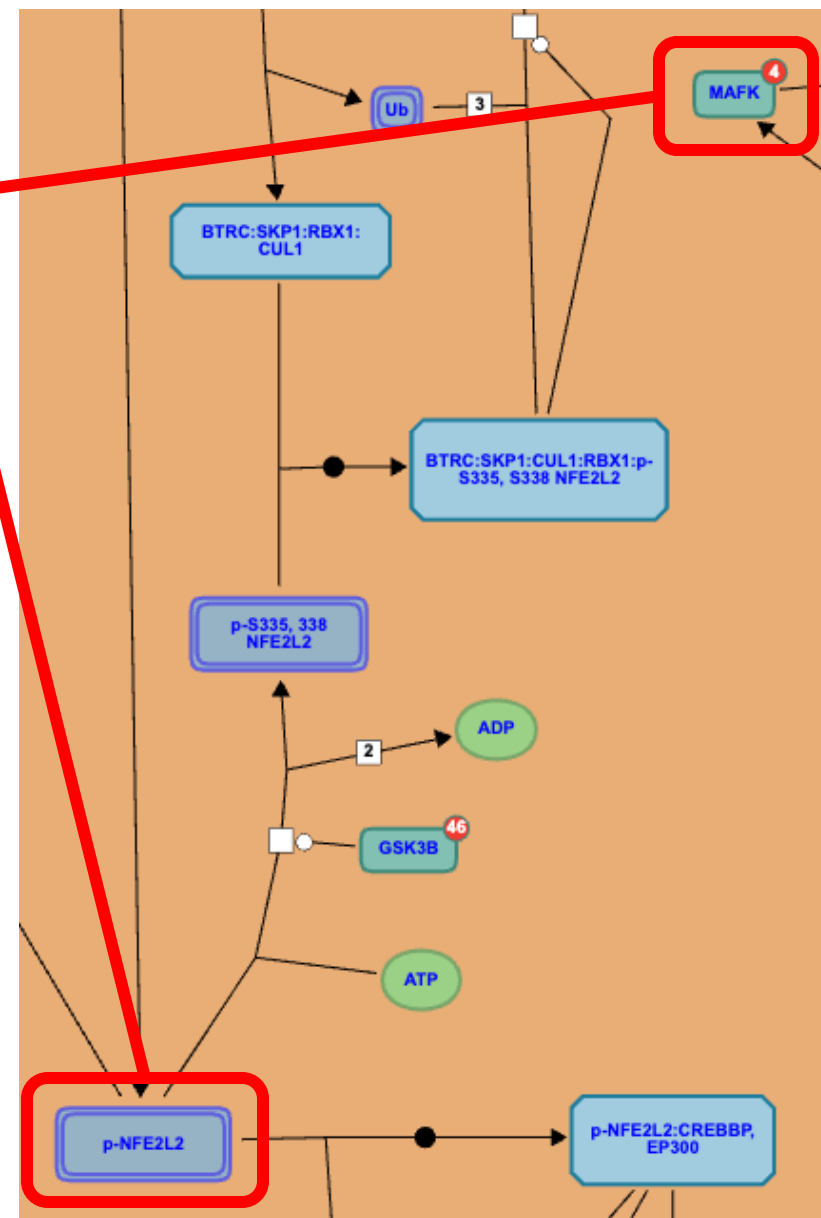
Cytoprotection by HMOX1



Create a reaction:



KEAP1-NFE2L2 pathway



Signaling network reconstruction

4. Curate manually all the pathways of the final list

Reactions and species information can be downloaded automatically but:

- 1) Dashed lines in the graph represent flow of information but the data does not exist.
- 2) The connection between different pathways should be checked.

Signaling network reconstruction

4. Curate manually all the pathways of the final list

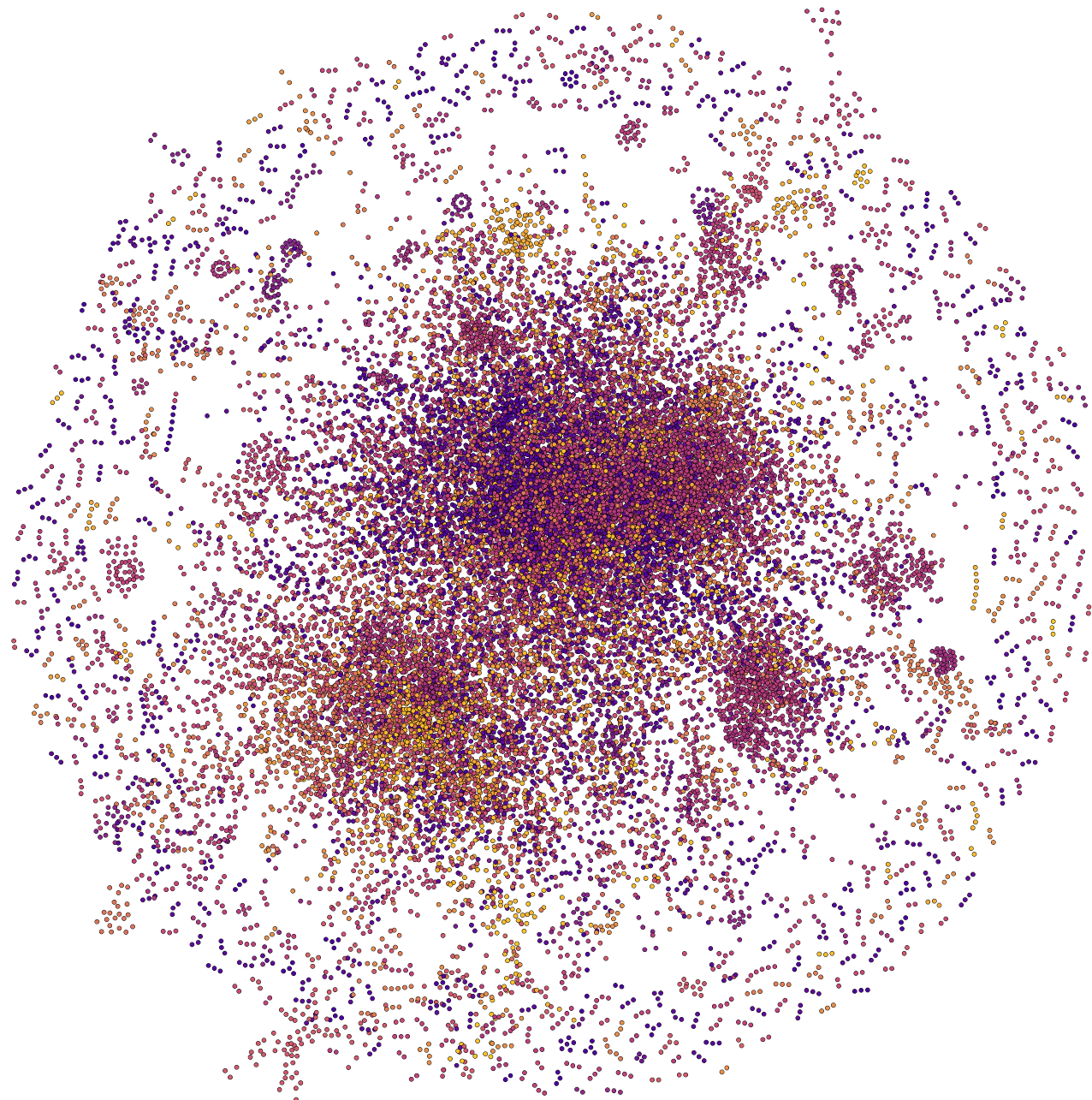
Reactions and species information can be downloaded automatically but:

- 1) Dashed lines in the graph represent flow of information but the data does not exist.
- 2) The connection between different pathways should be checked.

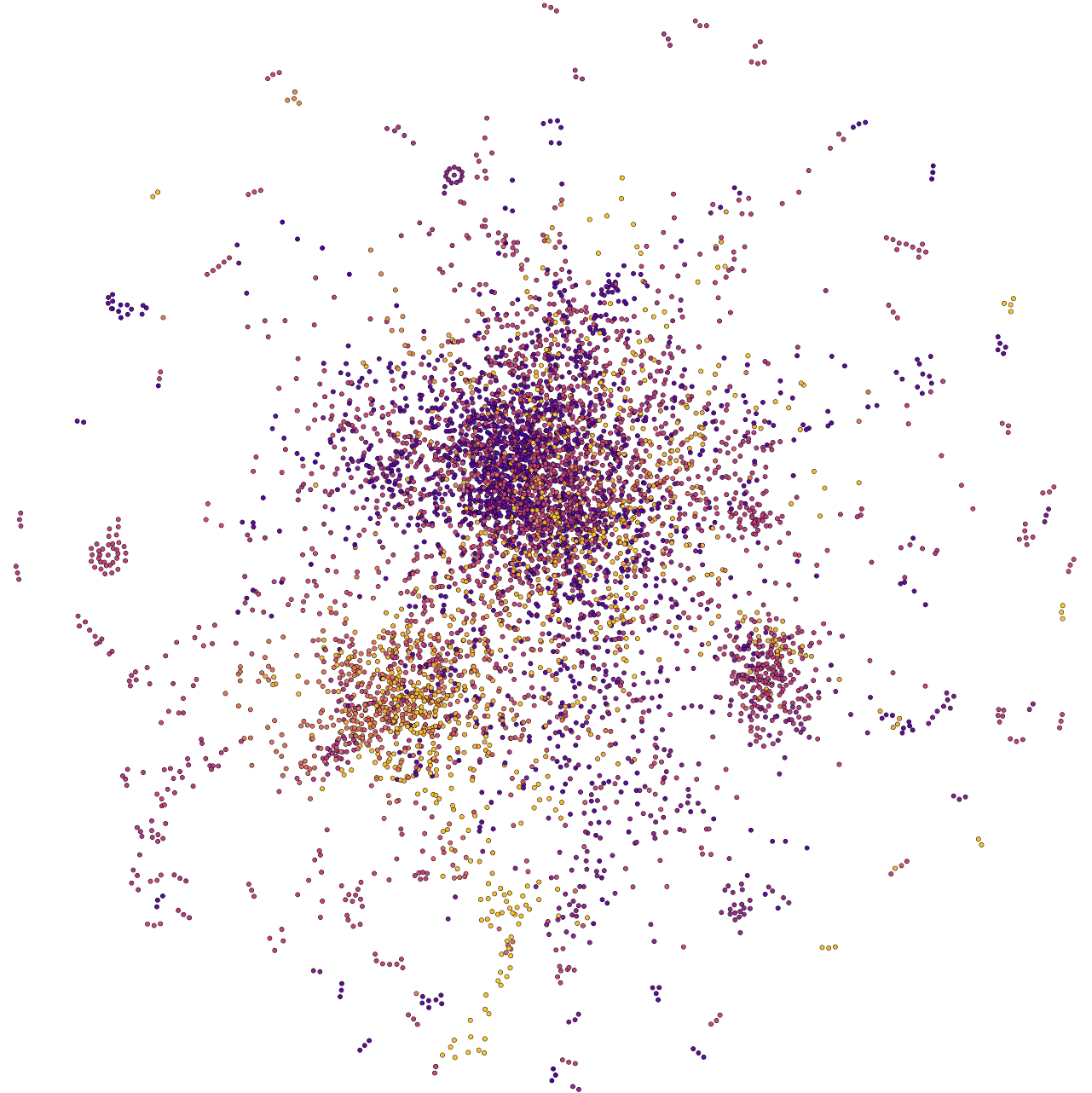
New reactions added to the network after manual curation:

- Reactions from dashed lines = 820
- Reactions between different pathways = 26

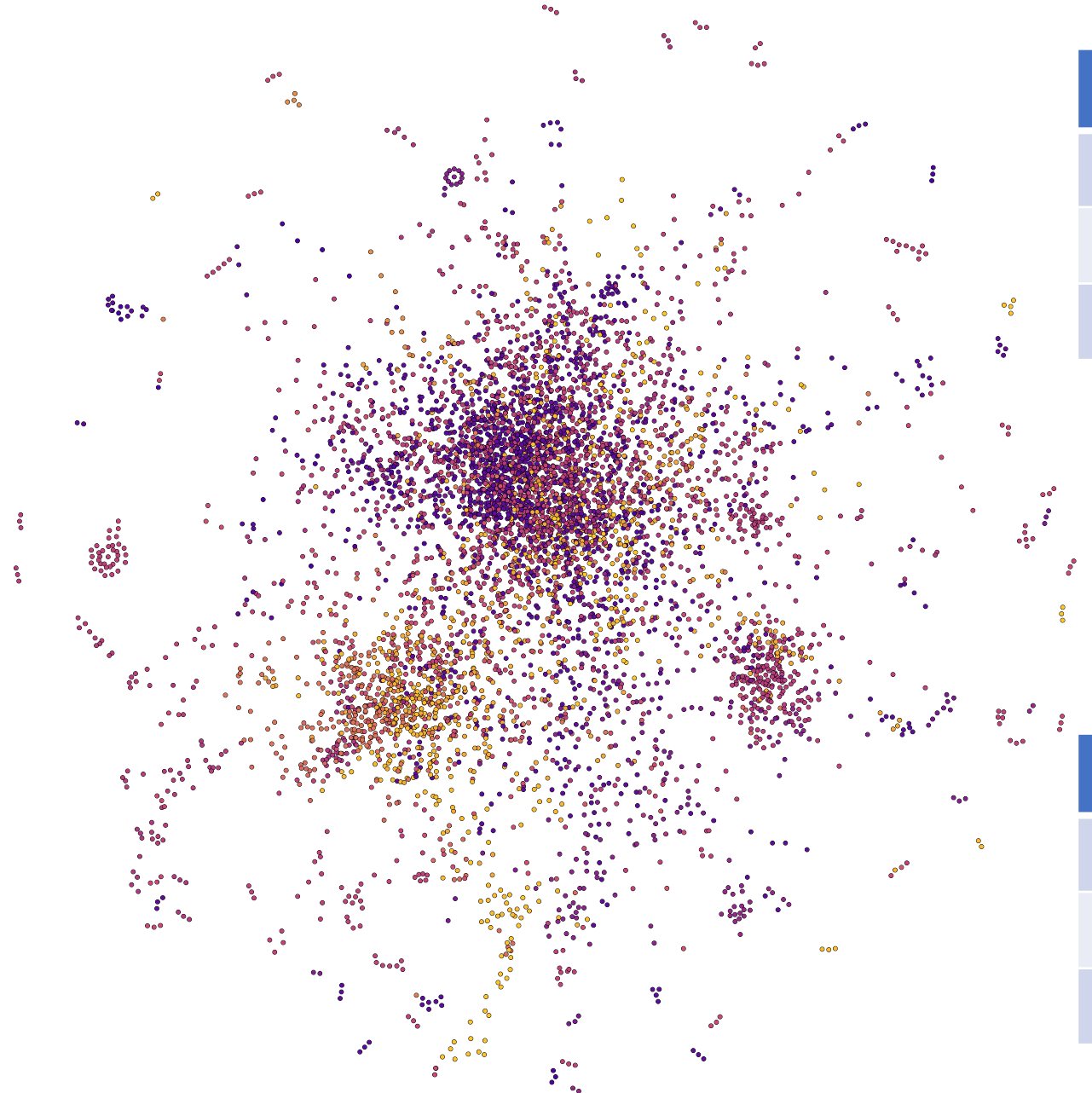
From Reactome ...



... to selected pathways network



From Reactome to selected pathways network



Reactome	
Pathways	2610
Species	23830
Connections	46834

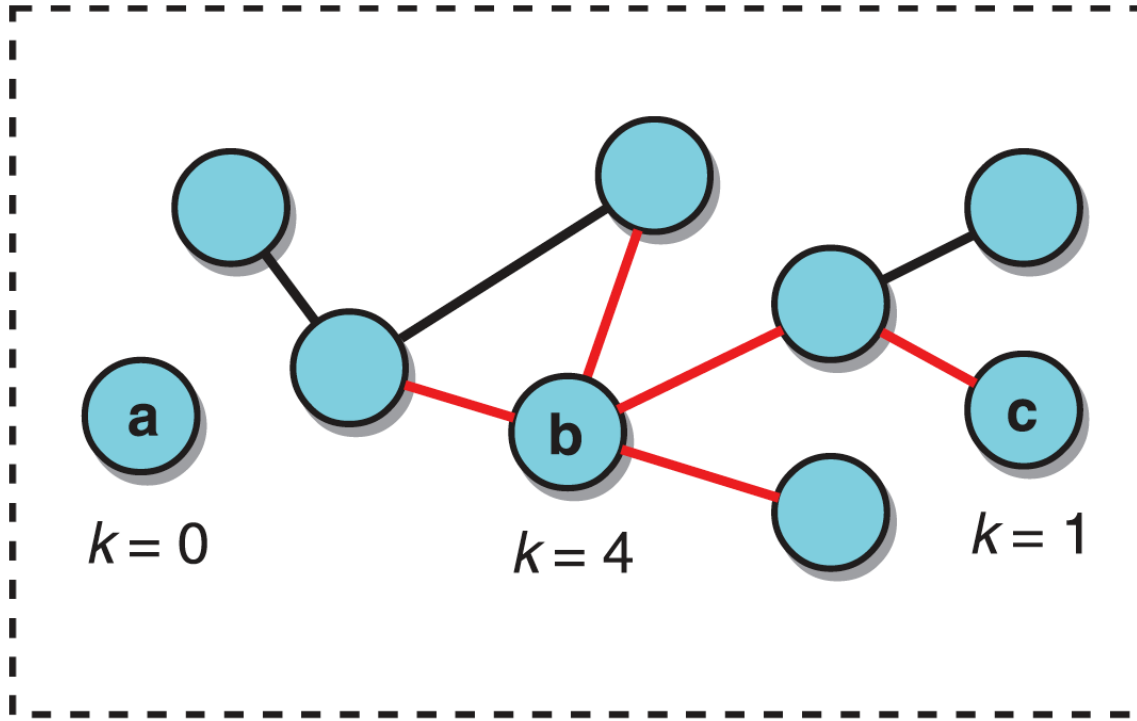


Selected network	
Pathways	115
Species	5947
Connections	12040

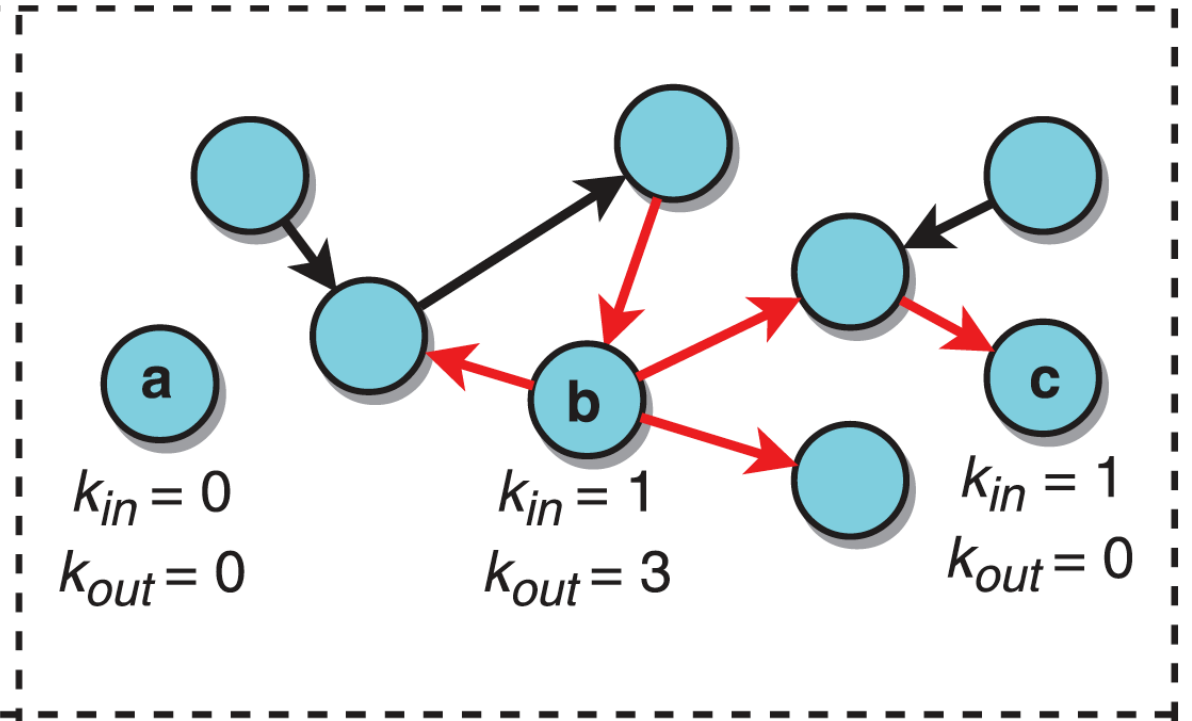
Analysis with concepts from network theory

Degree centrality

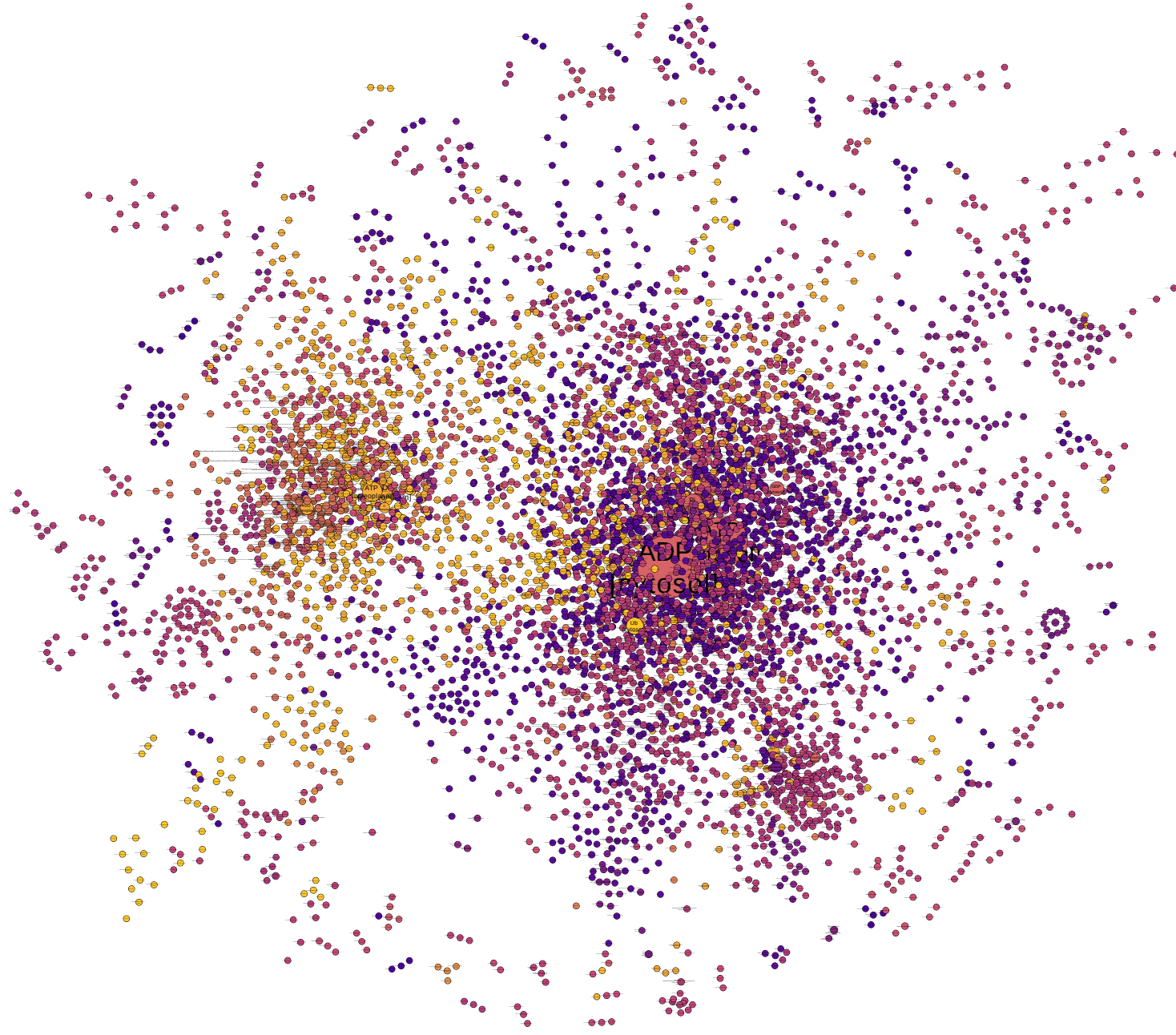
Undirected



Directed

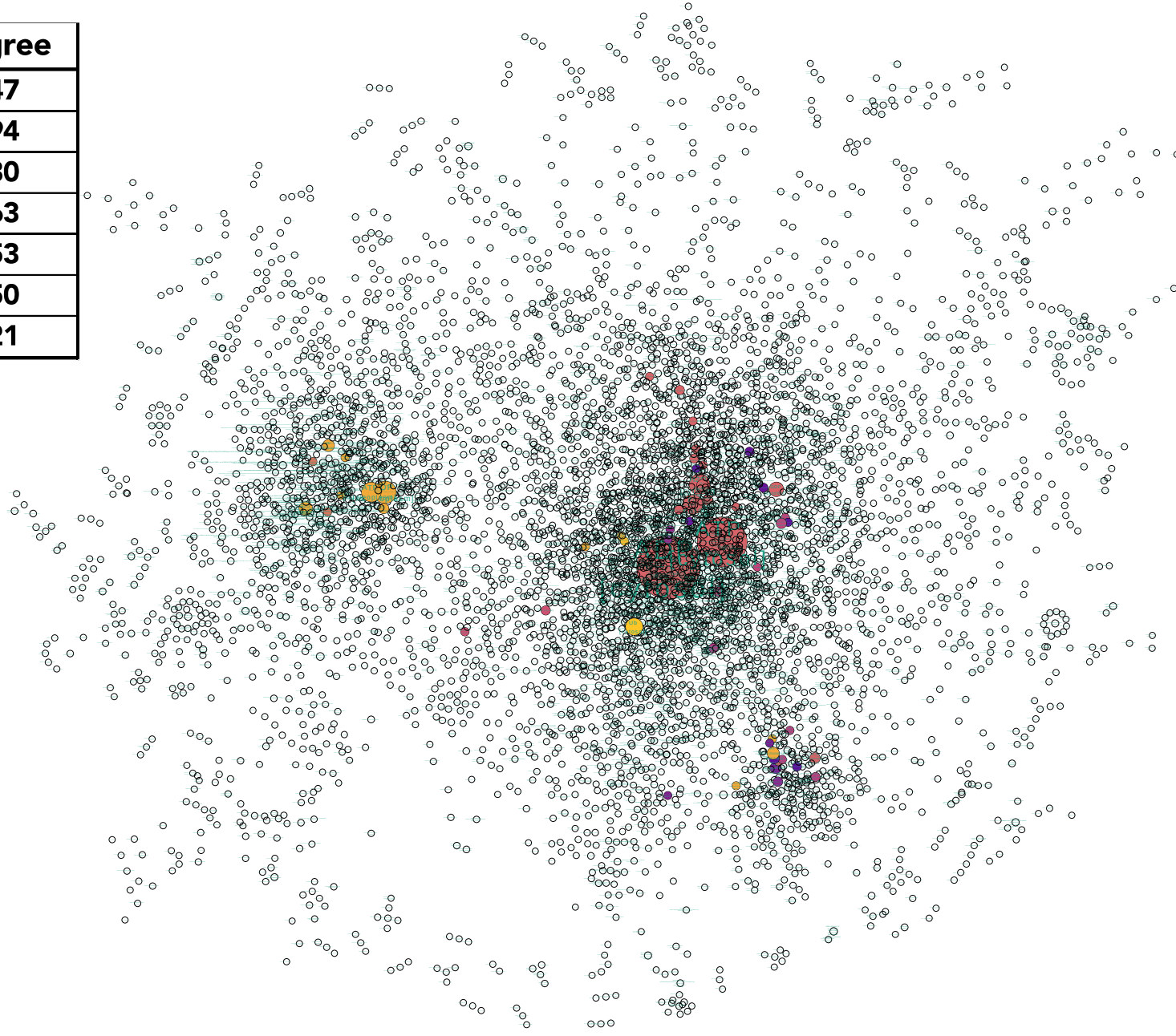


Degree centrality of the selected pathways network



Degree centrality of the selected pathways network

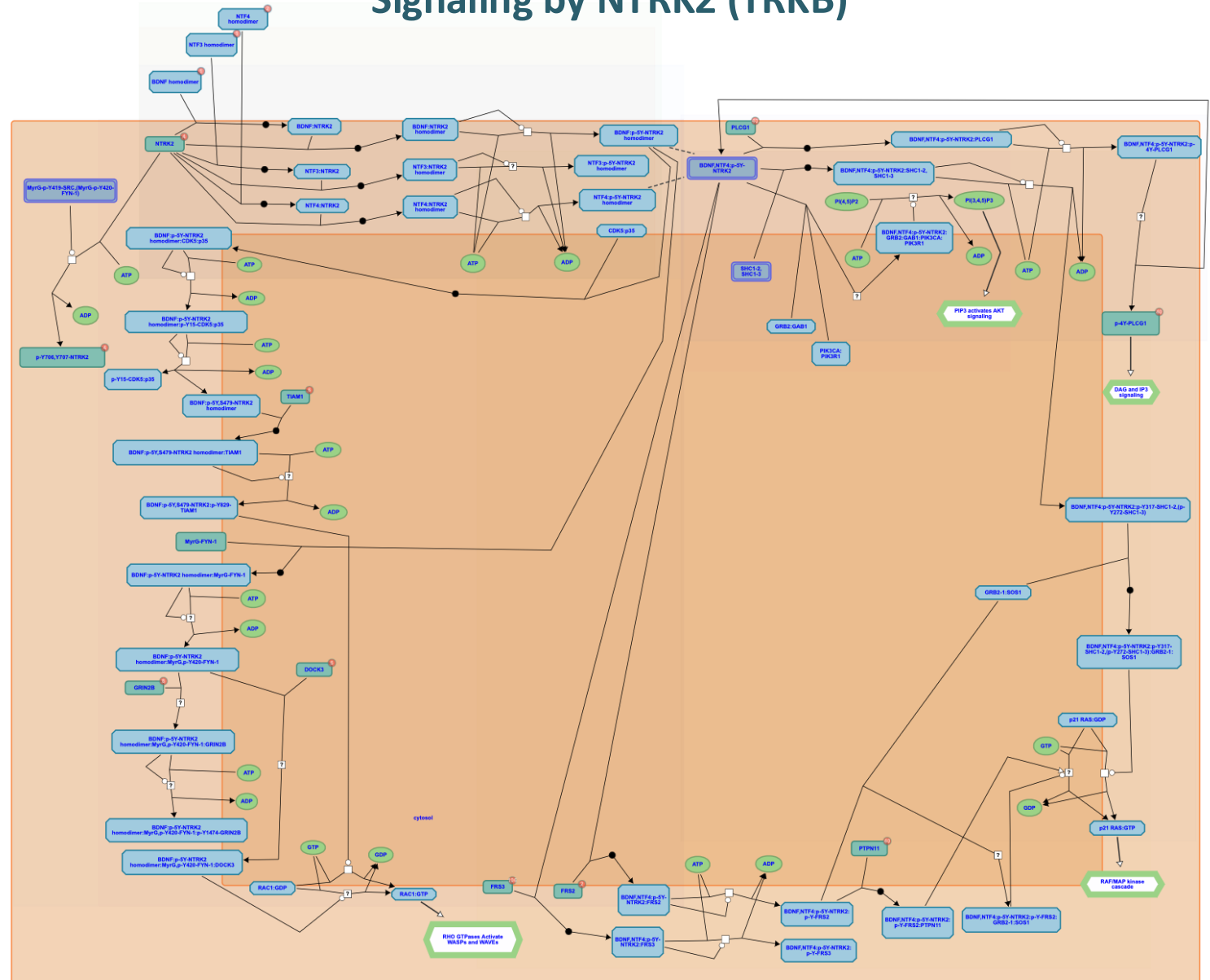
Species	Degree
ADP [cytosol]	647
ATP [cytosol]	494
ADP [nucleoplasm]	180
H2O [cytosol]	163
Pi [cytosol]	153
ATP [nucleoplasm]	150
Ub [cytosol]	121



Co-factors identification and removal

Why should we remove co-factors from the final network?

Signaling by NTRK2 (TRKB)

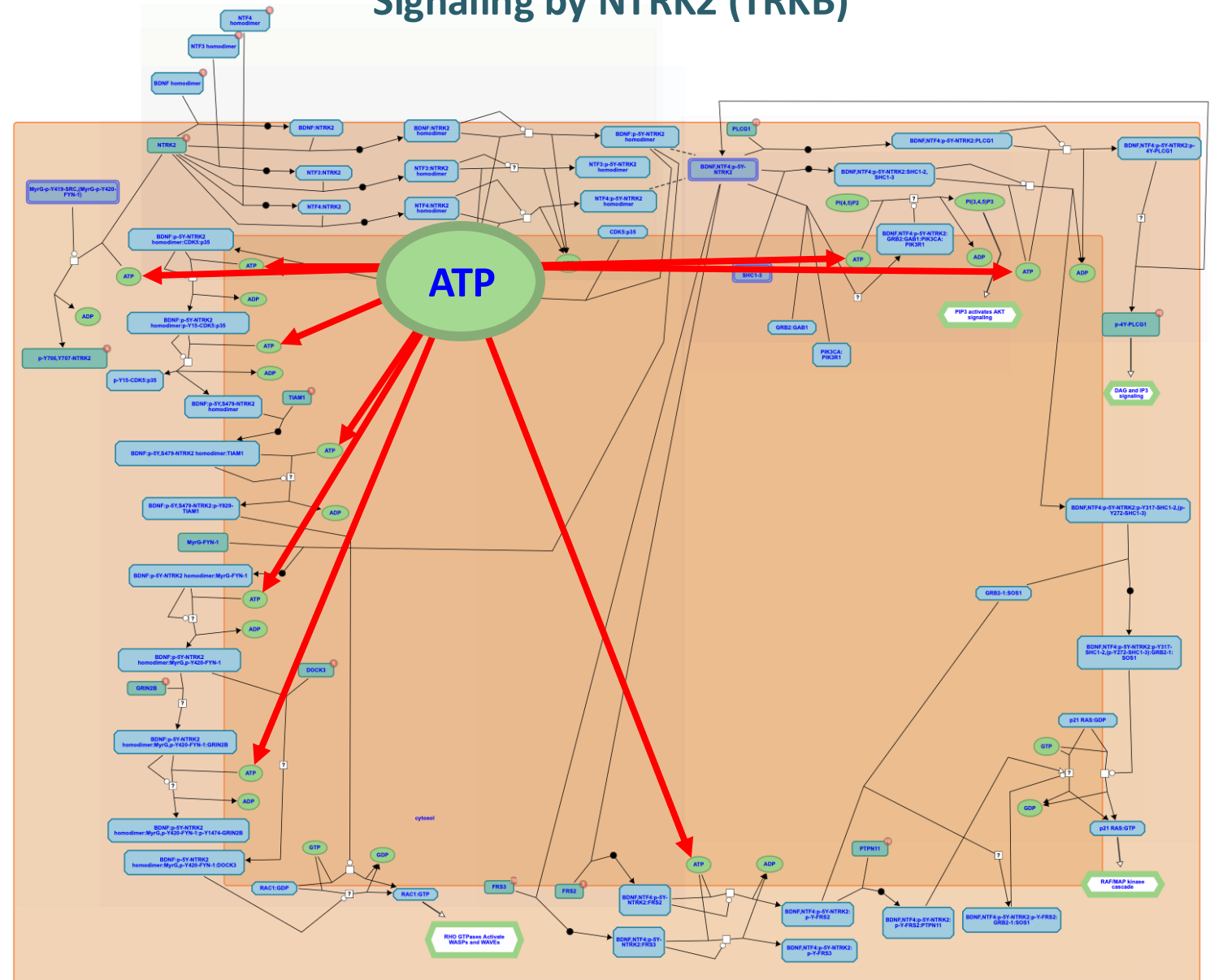


Co-factors identification and removal

Signaling by NTRK2 (TRKB)

Why should we remove co-factors from the final network?

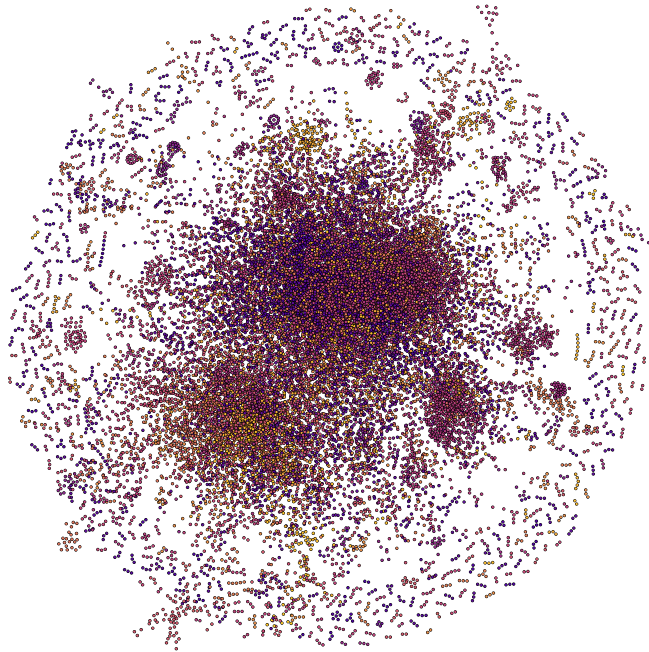
Avoid producing biological meaningless results.



Co-factors identification and removal

Selection of co-factor list for removal:

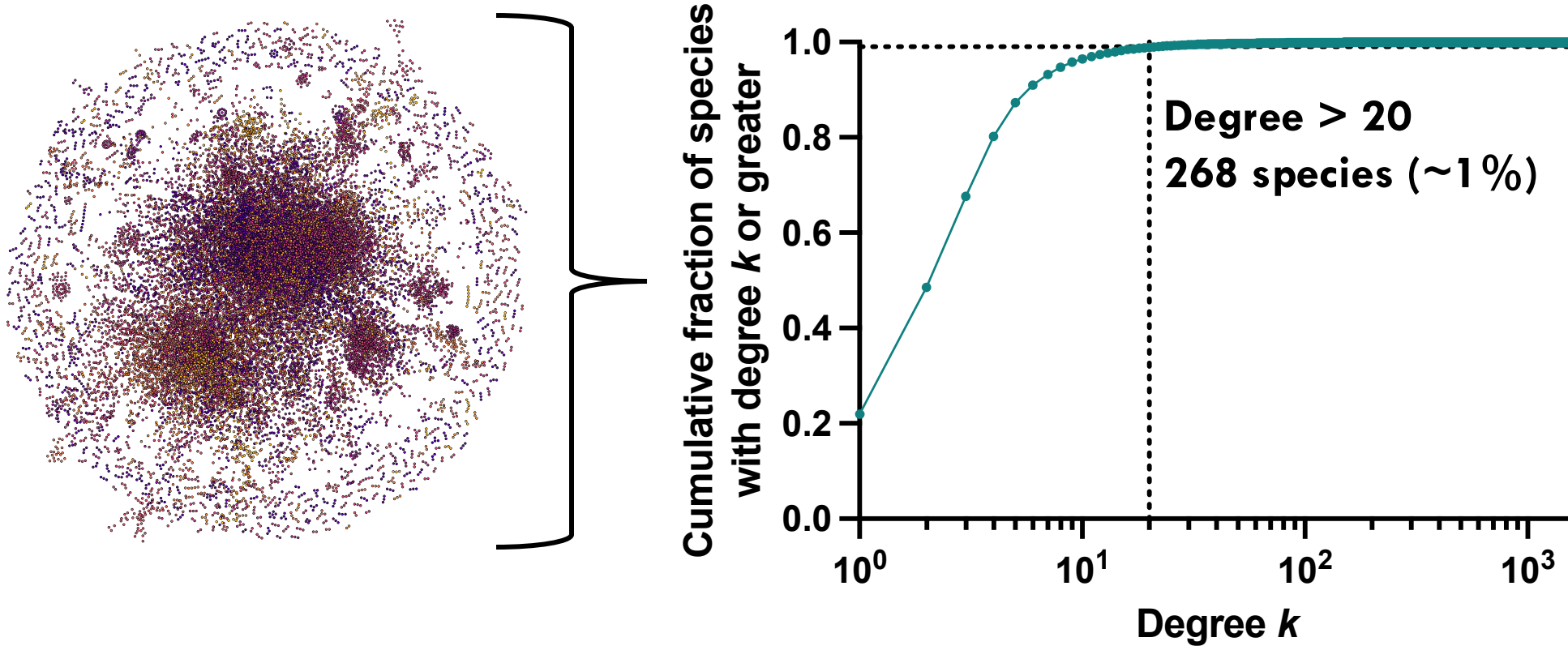
- Based on the degree ranking of the whole Reactome network.



Co-factors identification and removal

Selection of co-factor list for removal:

- Based on the degree ranking of the whole Reactome network.



Co-factors identification and removal

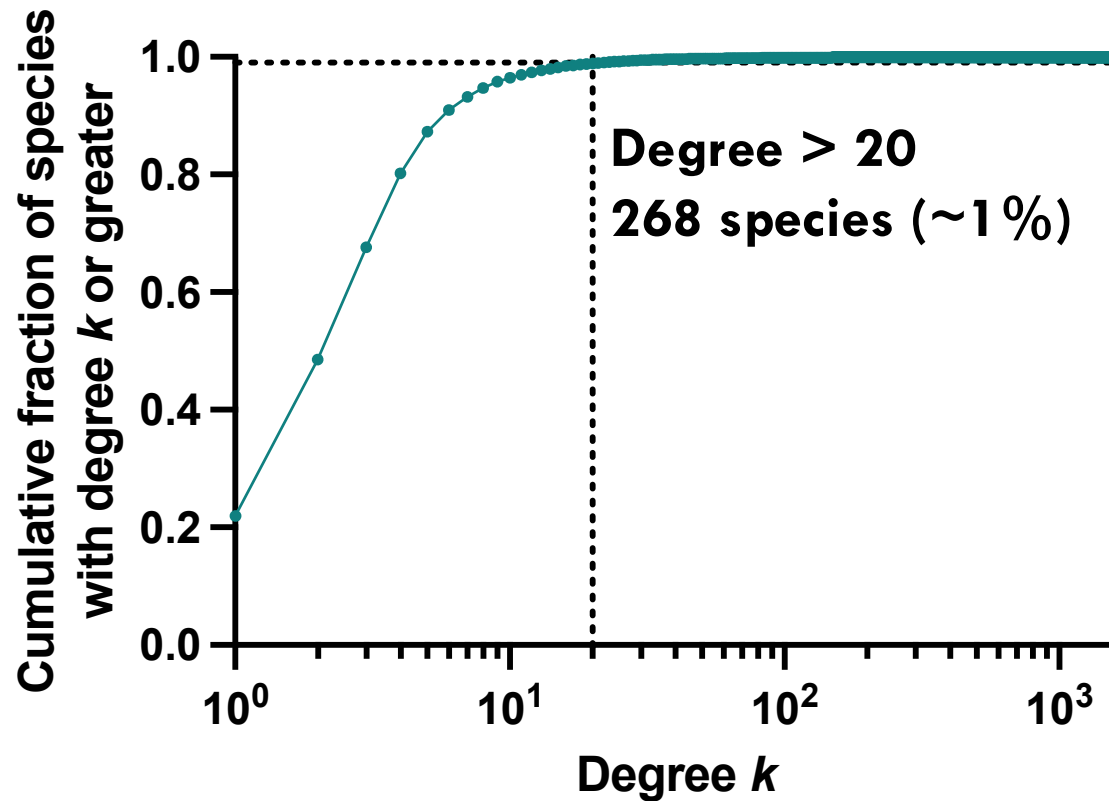
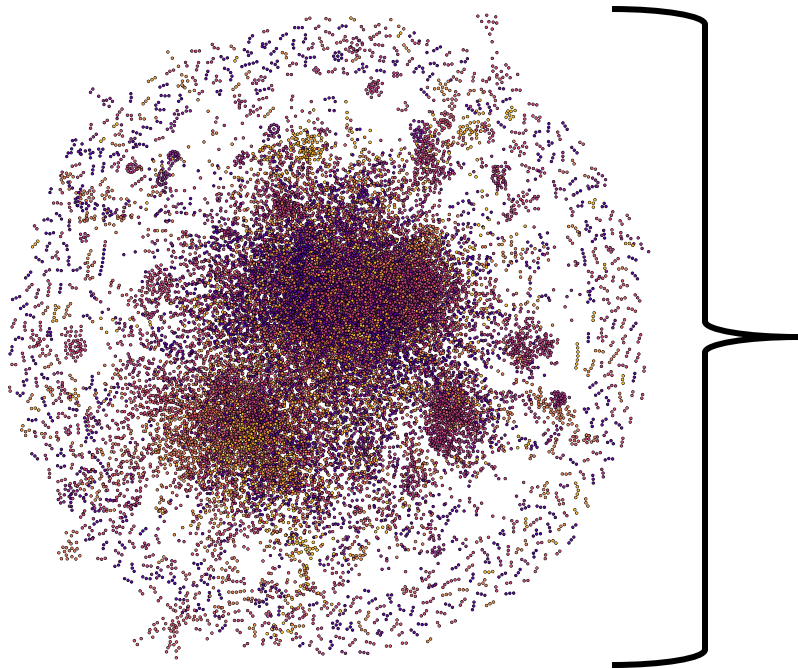
Selection of co-factor list for removal:

- Based on the degree ranking of the whole Reactome network.



Keep important small
second messengers:

- **cAMP**
- **cGMP**
- **IP₃**
- **DAG**
- **Ca²⁺**



Co-factors identification and removal

Selection of co-factor list for removal:

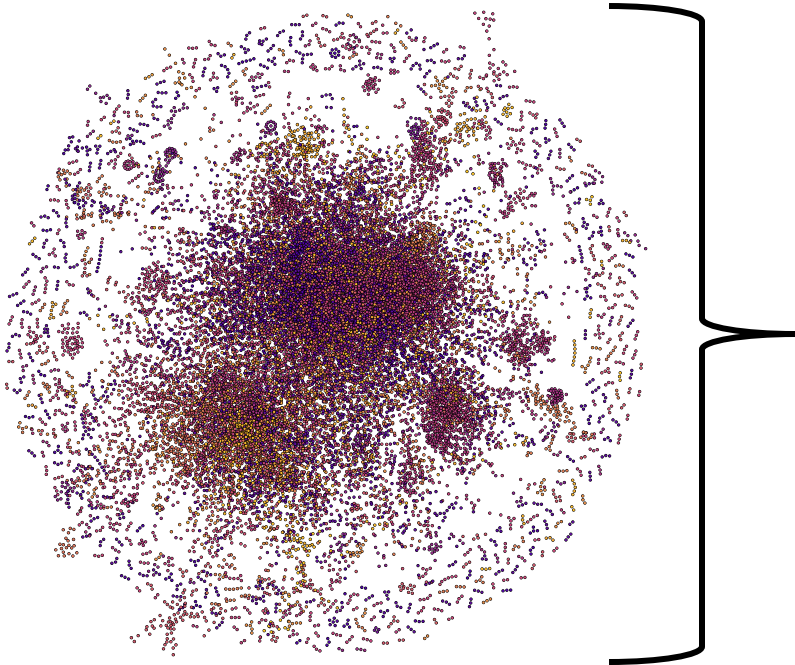
- Based on the degree ranking of the whole Reactome network.



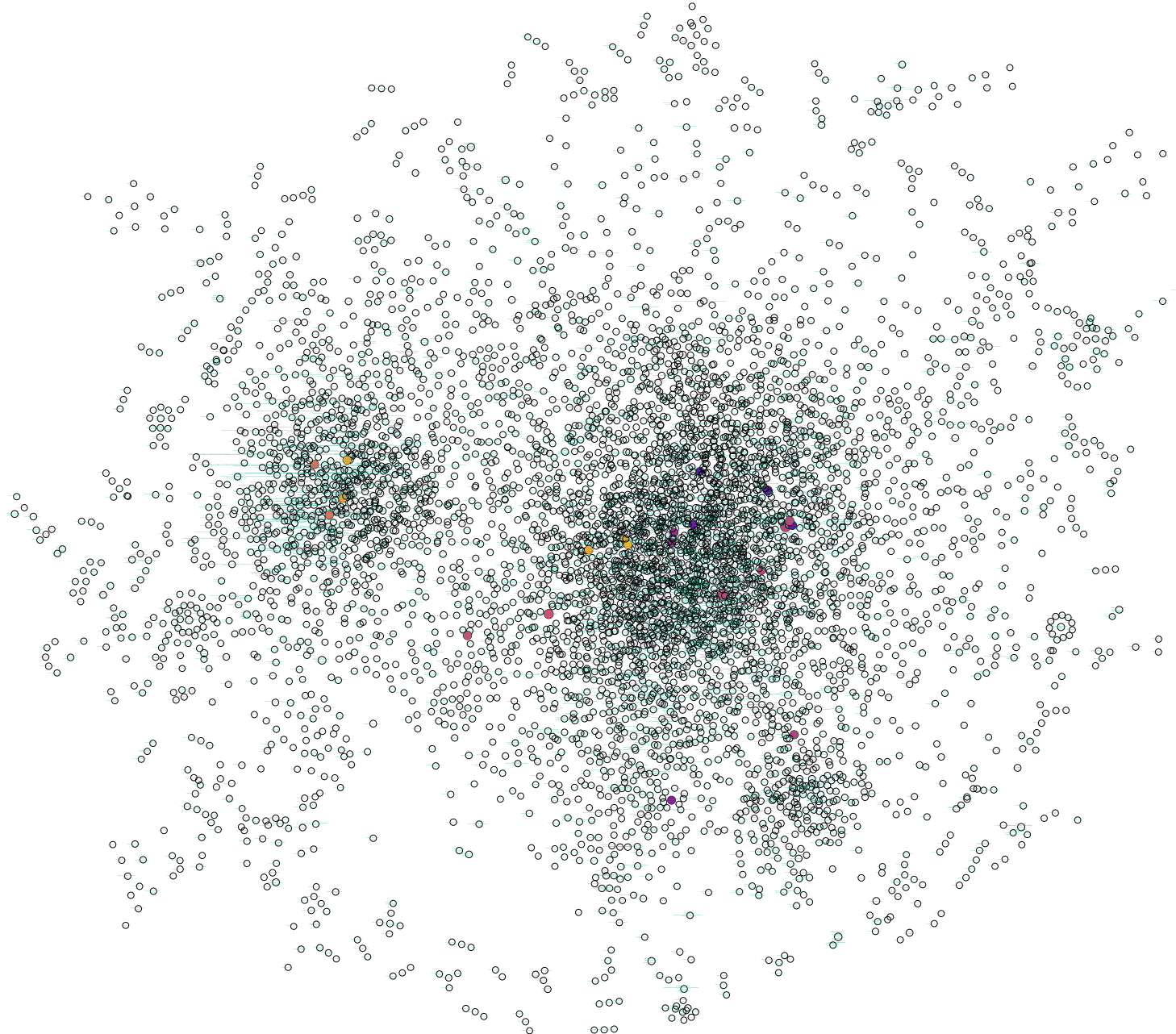
**Keep important small
second messengers:**

54 co-factors, small molecules

ATP	Pi	Ac-CoA	Fe ²⁺	IMP
ADP	PPi	GSH	Mg ²⁺	NTP(4-)
AMP	H ⁺	GSSG	Zn ²⁺	NO
CTP	H ₂ O	PAPS	NTP	HCOOH
CDP	Ub	PAP	K ⁺	QH ₂
CMP	NAD ⁺	AdoMet	Cl ⁻	THF
GTP	NADH	AdoHcy	NH ₄ ⁺	
GDP	NADP ⁺	K63polyUb	CH ₂ O	
GMP	NADPH	H ₂ O ₂	NH ₃	
UTP	O ₂	Na ⁺	dNTP	
UDP	CO ₂	FADH ₂	BH ₄	
UMP	CoA-SH	FAD	HCO ₃ ⁻	



Co-factors identification and removal



Selected network

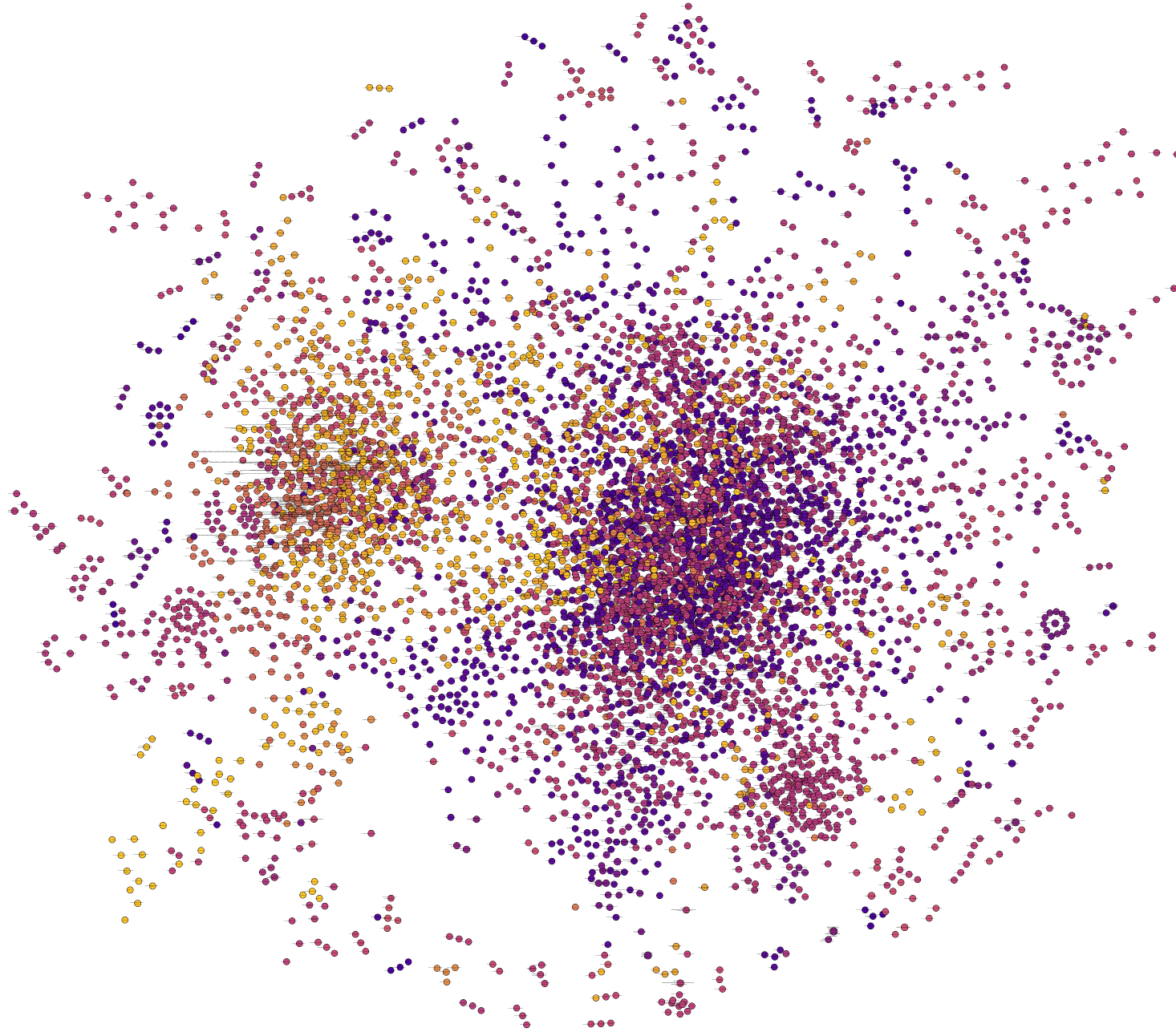
Pathways	115
Species	5947
Connections	12040

-136 species
-3344 connections

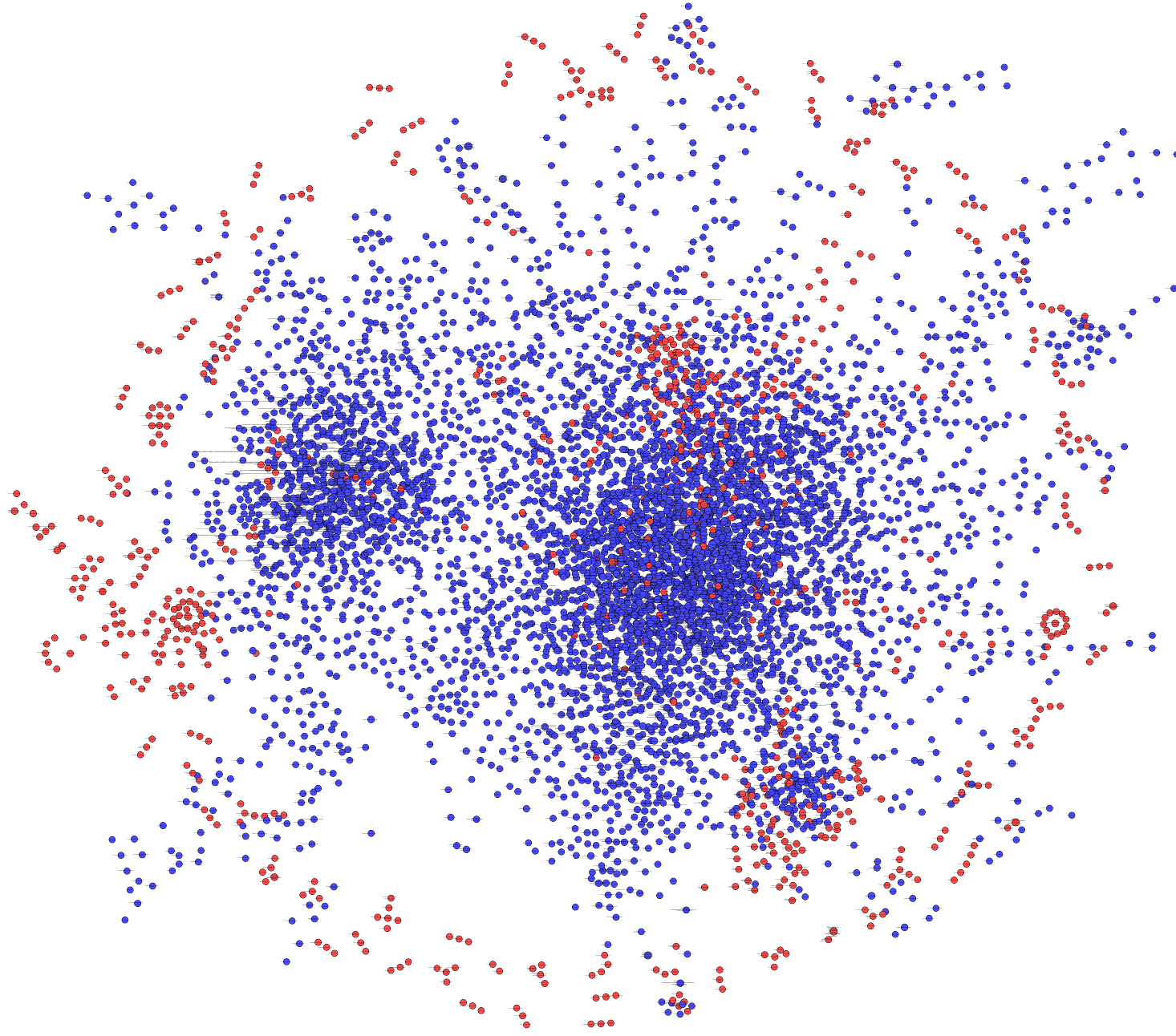
Final network

Pathways	115
Species	5811
Connections	8696

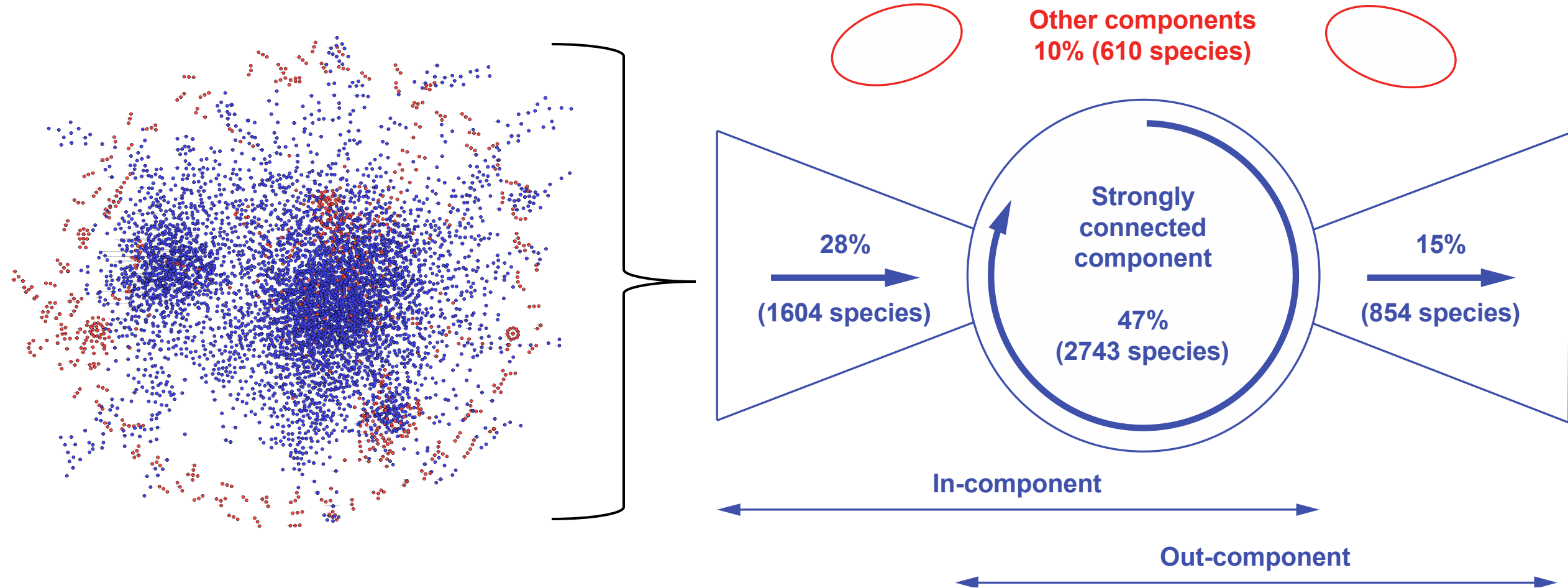
Final network of selected pathways



Analysis of the final network of selected pathways



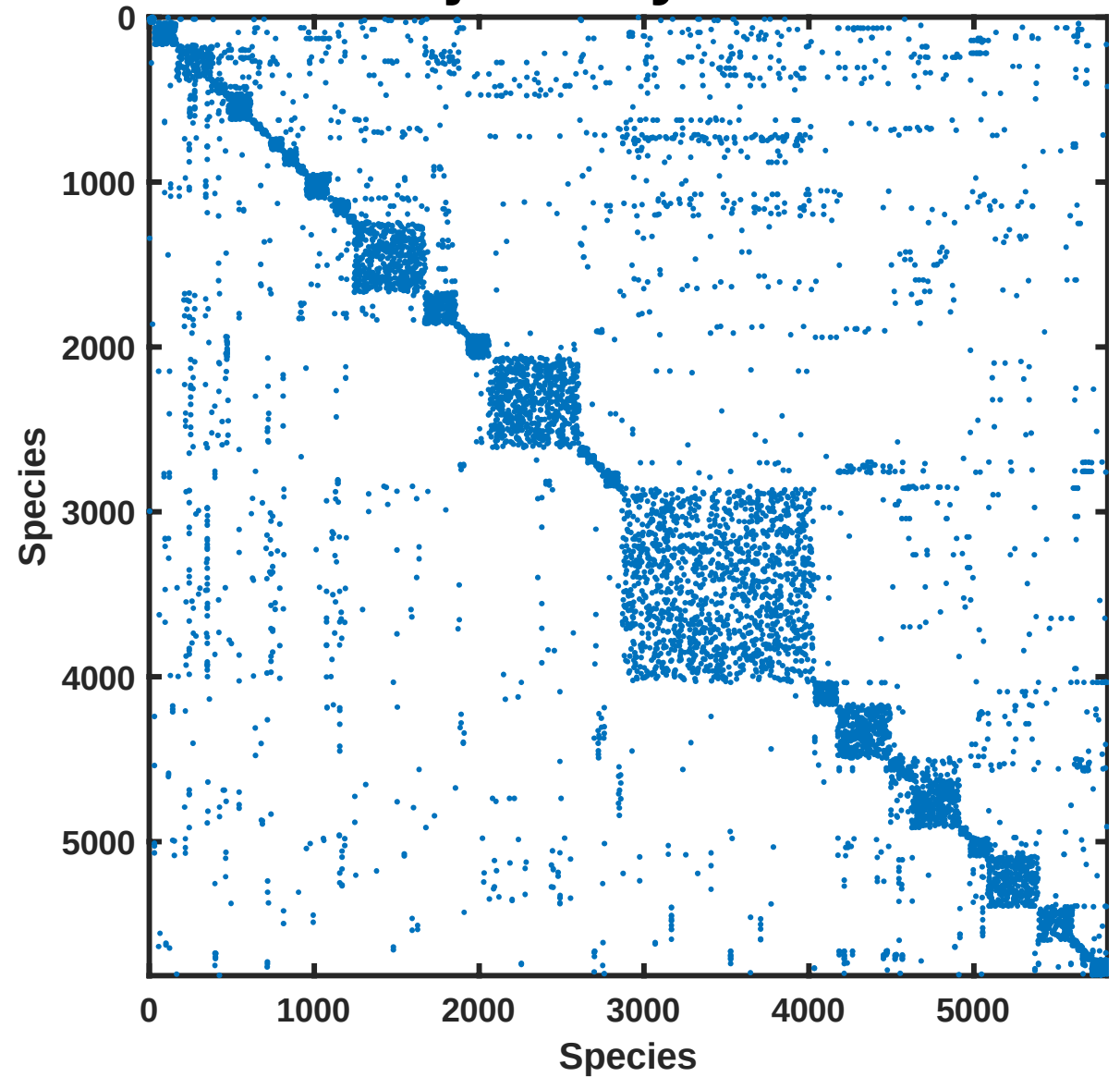
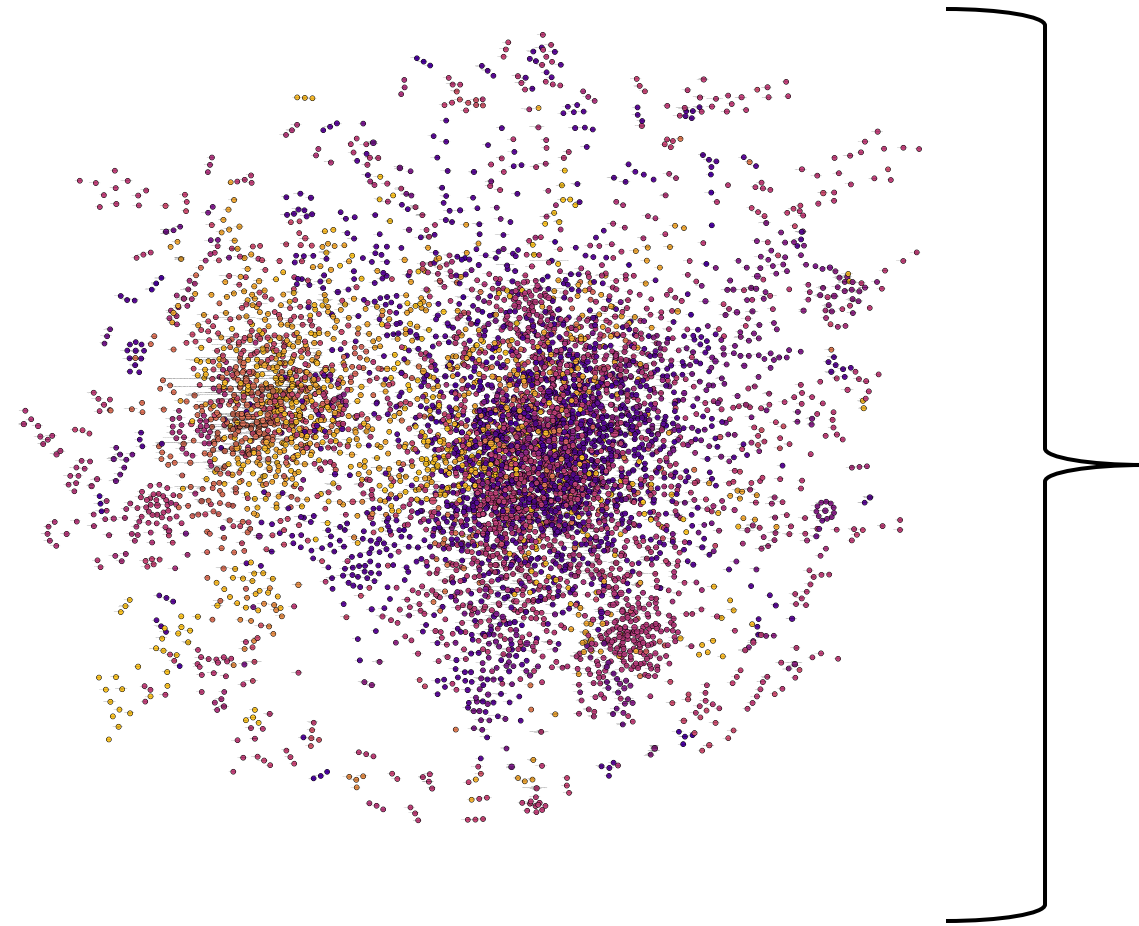
Analysis of the final network of selected pathways



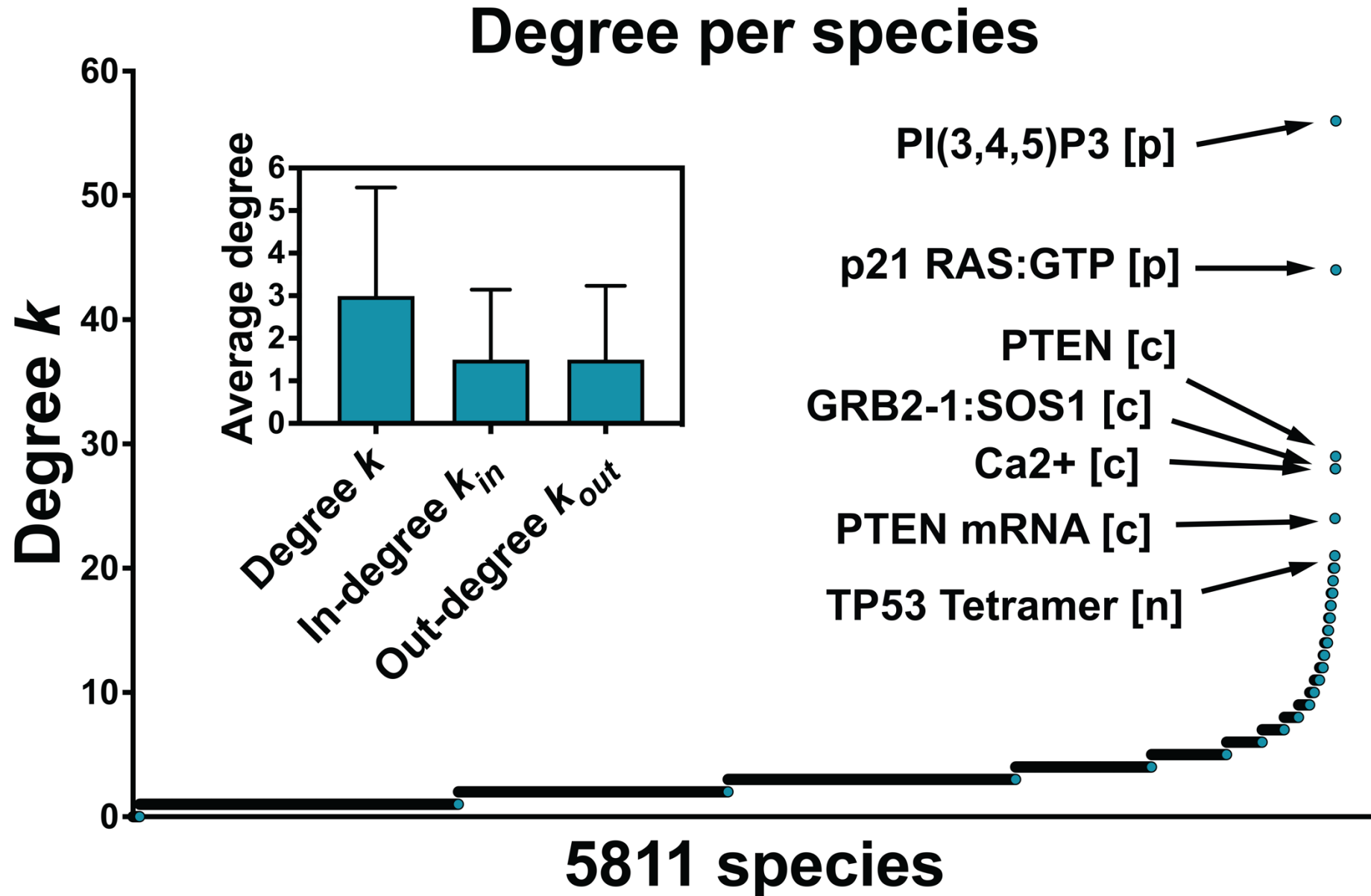
The “bow-tie” diagram

Analysis of the final network of selected pathways

adjacency matrix

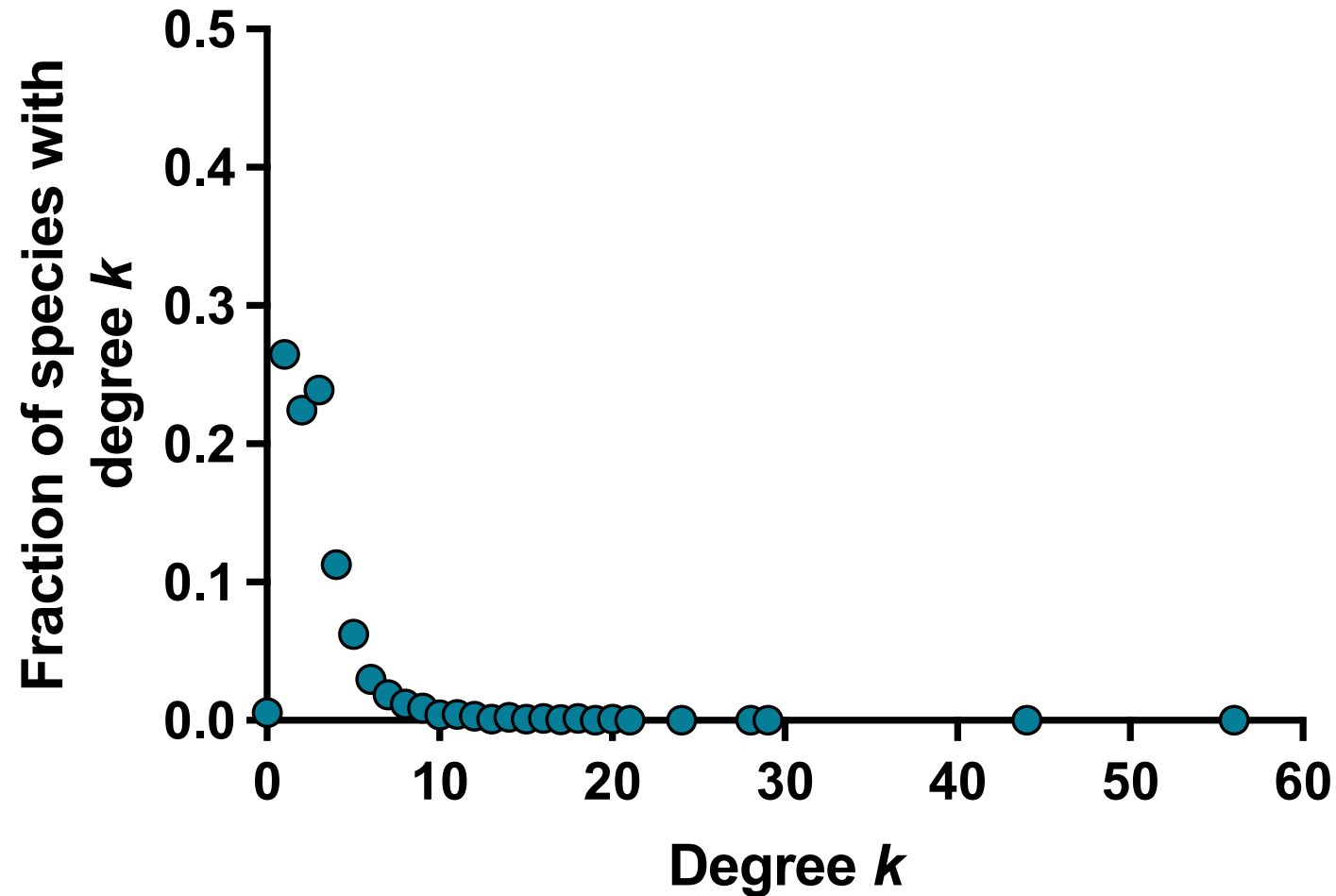


Analysis of the final network of selected pathways



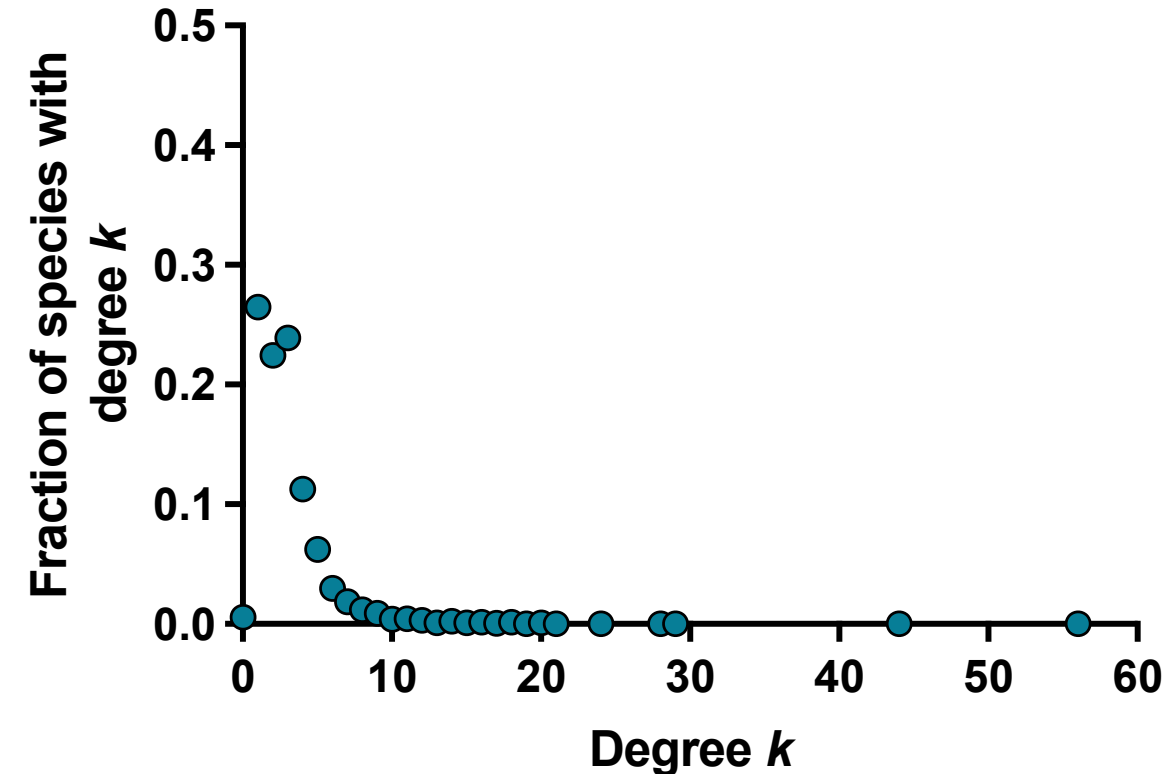
Analysis of the final network of selected pathways

Degree distribution
(linear scale)

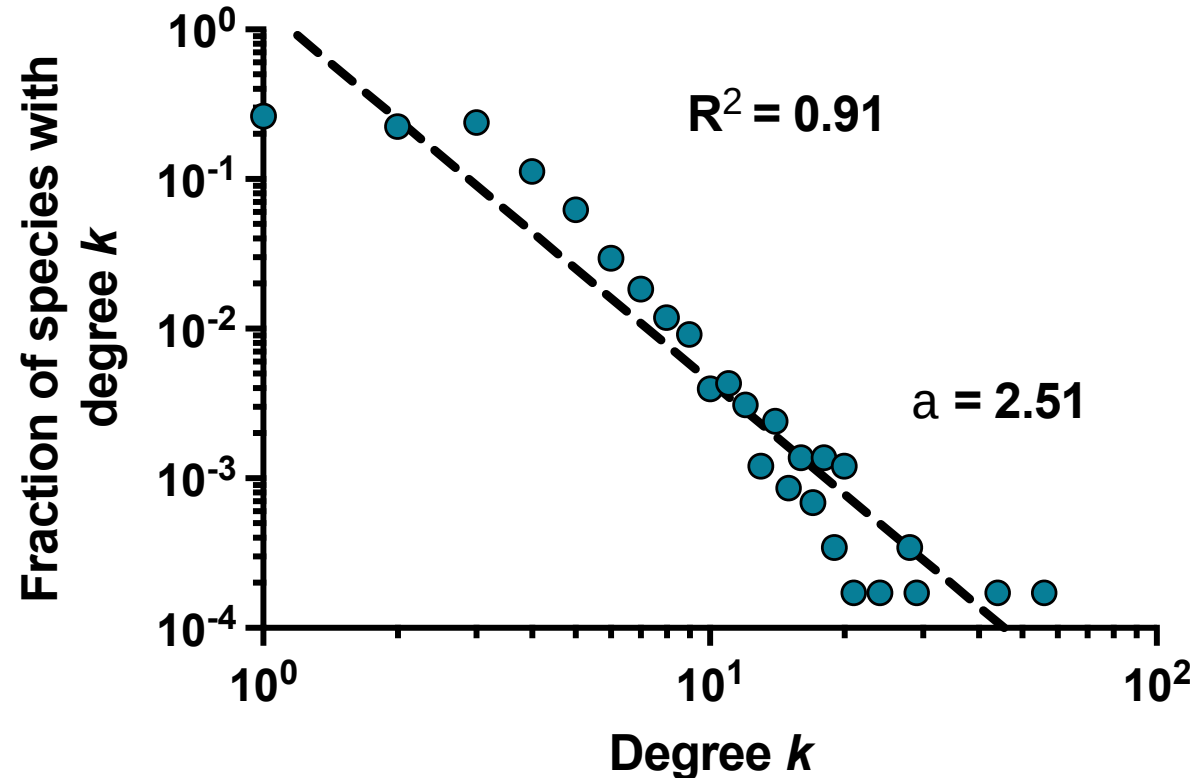


Analysis of the final network of selected pathways

Degree distribution
(linear scale)



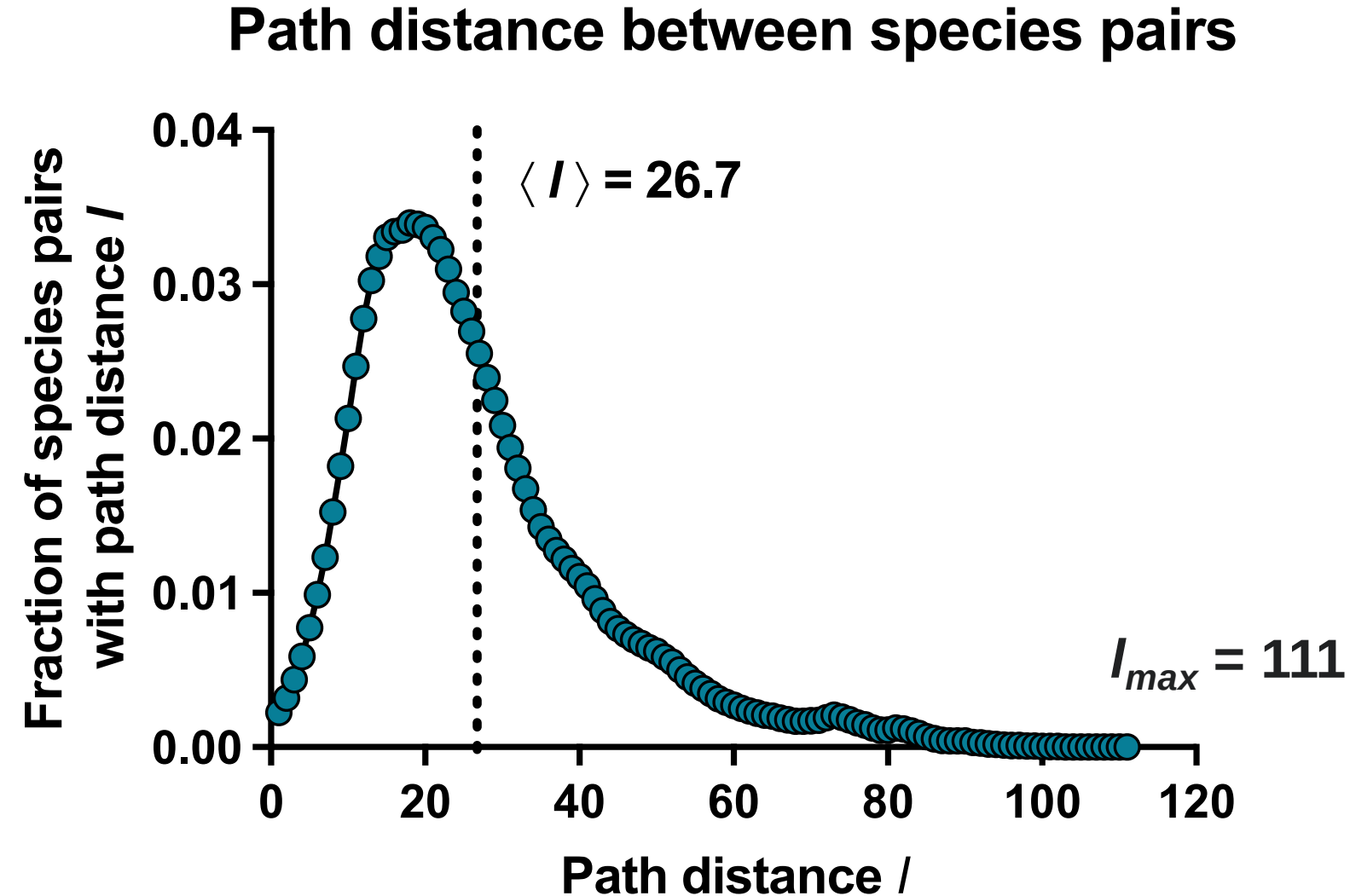
Degree distribution
(log scale - linear binning)



SCALE FREE

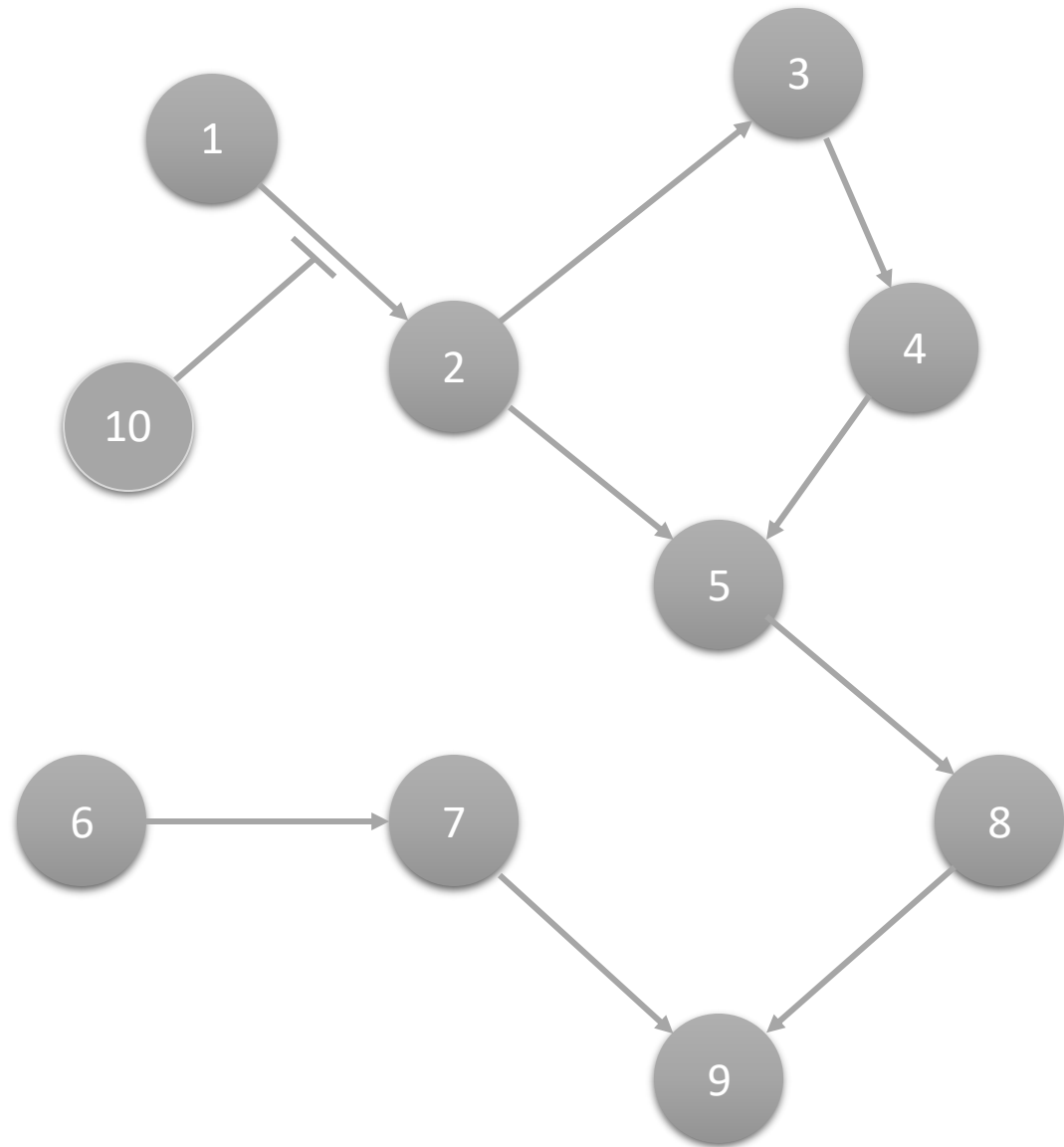
$$p_k \sim k^{-\alpha}$$
$$\log p_k \sim -\alpha \log k$$

Analysis of the final network of selected pathways



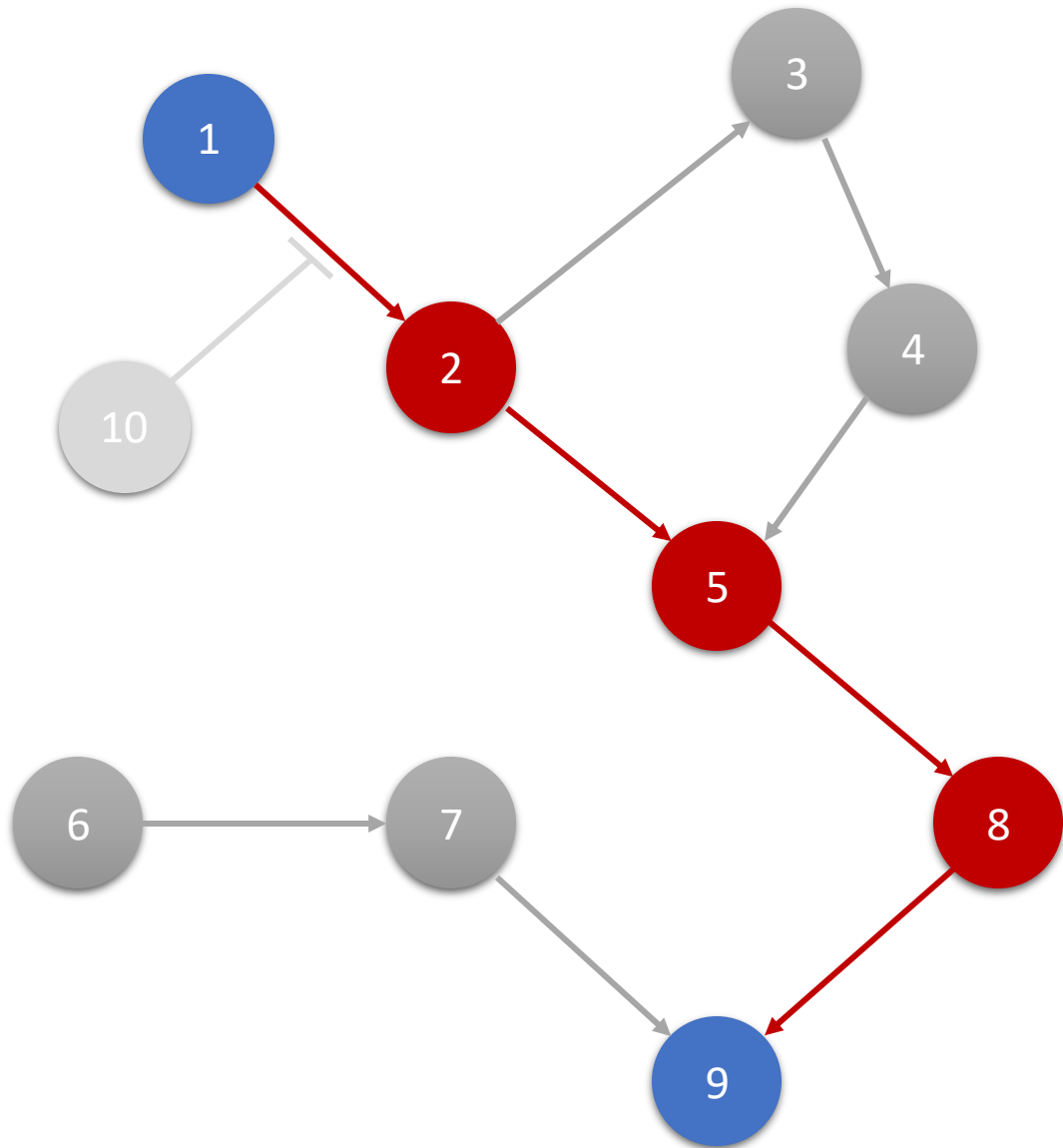
from Signaling Models to Information Flows

Method for connecting signaling network species



From species 1 to species 9

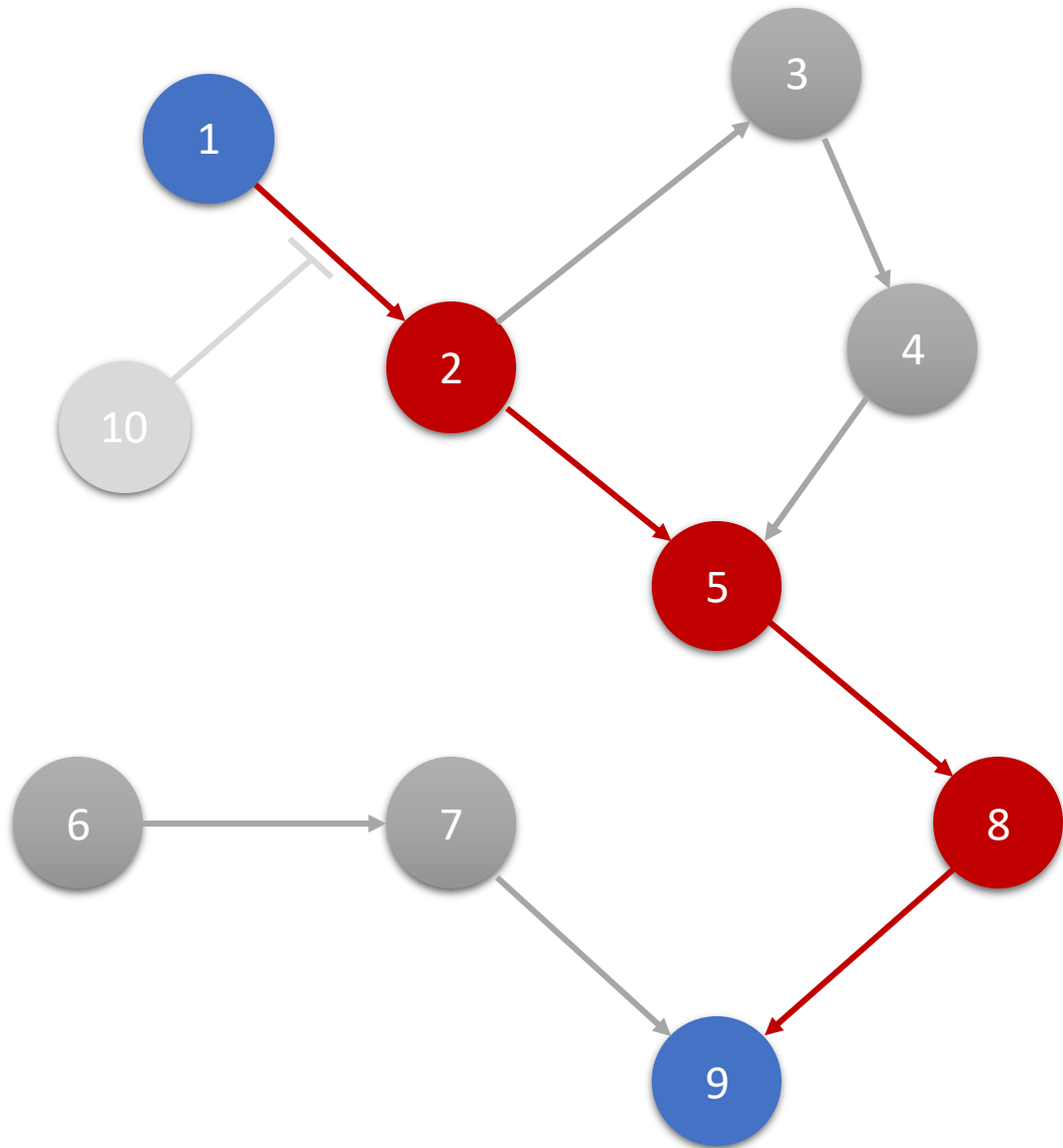
Method for connecting signaling network species



From species 1 to species 9

We enforce the start species (1) and the target species (9) to be active and minimize the number of active connections.

Method for connecting signaling network species

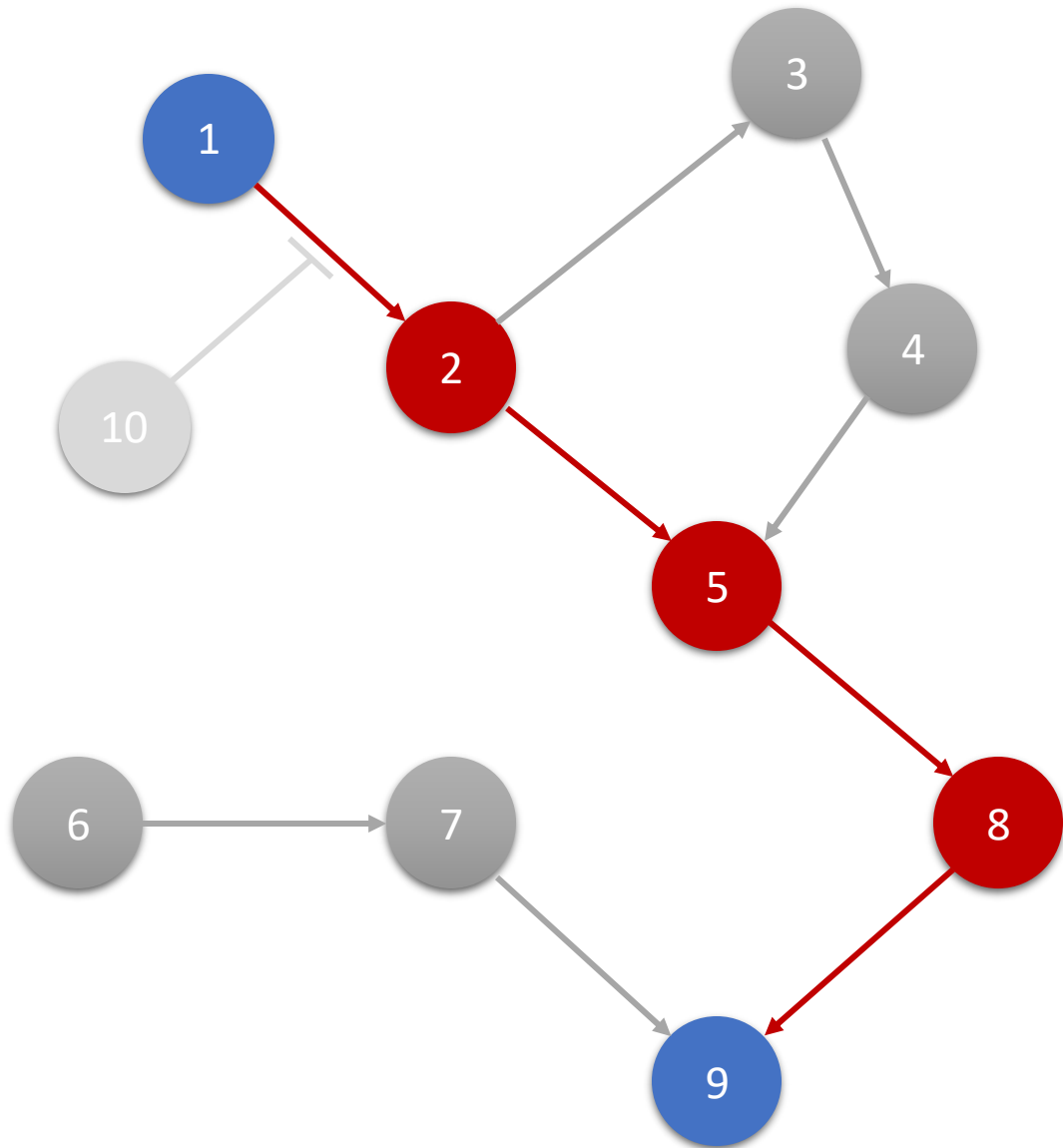


Why minimize?

1) Focus on the most essential species and connections of the information flow.

2) Identify the parsimonious way of cellular signal transduction.

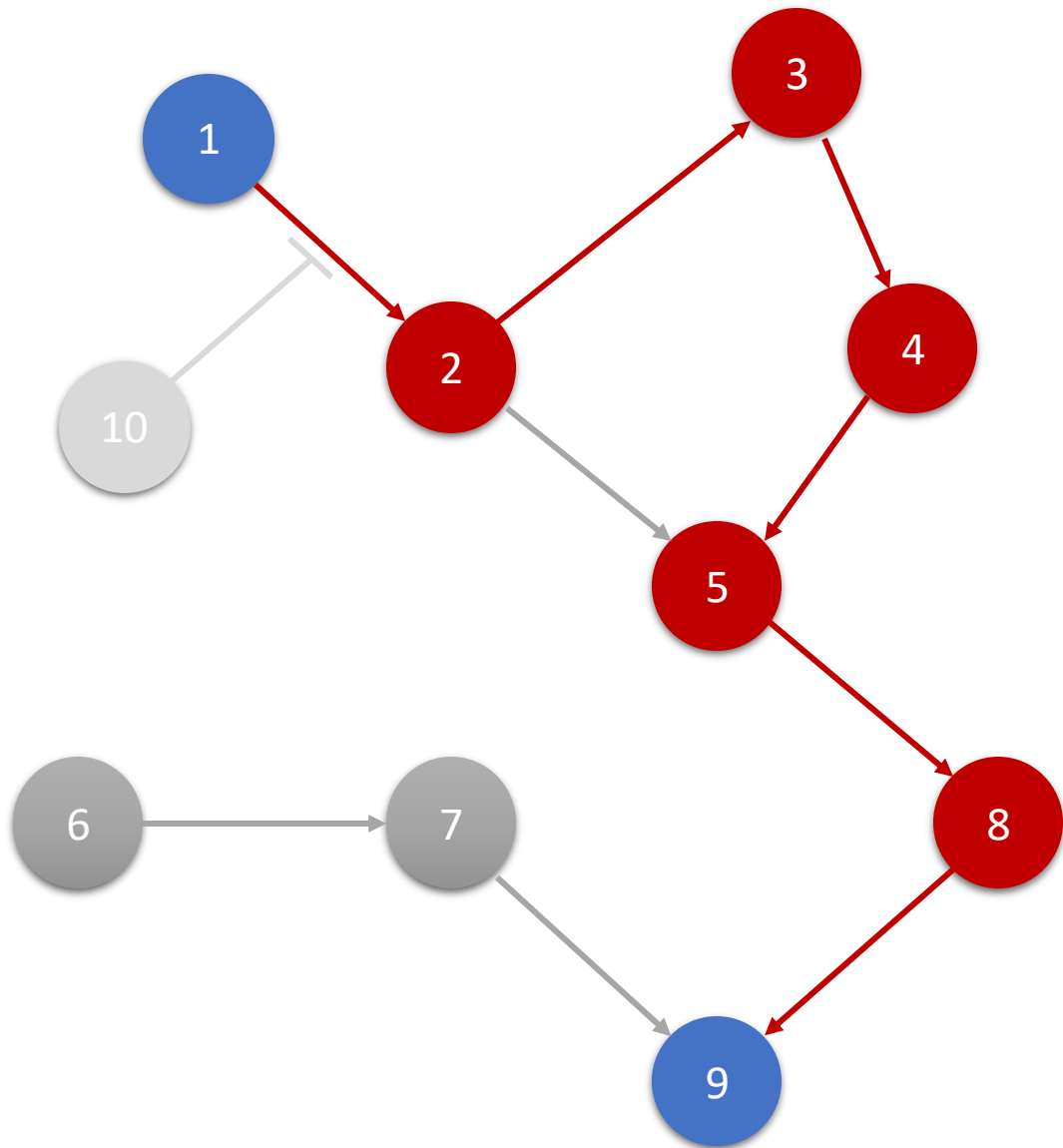
Method for connecting signaling network species



**We exhaustively enumerate
alternative paths!**

**Minimal path:
4 connections**

Method for connecting signaling network species



We exhaustively enumerate alternative paths!

**Minimal path:
4 connections**

**Alternative path:
Minimal path + 2**

Studying BDNF connection to mTOR

The start species is the BDNF ligand.

What about the target species?

Need for a systematic way to identify target species.

Studying BDNF connection to mTOR

Start	Target
BDNF homodimer [e]	Active mTORC1 complex [l]
BDNF homodimer [e]	mTORC1 [c]
BDNF homodimer [e]	mTORC1 dimer [n]
BDNF homodimer [e]	mTORC1 with p-S722,S792-RPTOR:Ragulator:Rag:GNP:RHEB:GTP [l]
BDNF homodimer [e]	MTORC1 with p-S722,S792-RPTOR:Ragulator:Rag:RHEB:GTP [l]
BDNF homodimer [e]	MTORC1 with p-S722,S792-RPTOR:RHEB:GTP:p-S758-ULK1:ATG13:RB1CC1:ATG101 [l]
BDNF homodimer [e]	mTORC1:p-T246-AKT1S1:YWHAB [l]
BDNF homodimer [e]	mTORC1:Ragulator:Rag:GNP:RHEB:GDP [l]
BDNF homodimer [e]	MTORC1:Ragulator:Rag:GNP:RHEB:GDP [l]
BDNF homodimer [e]	MTORC1:Ragulator:Rag:GNP:RHEB:GTP [l]
BDNF homodimer [e]	mTORC1:Ragulator:RagA,B:GTP:RagC,D:GDP:SLC38A9 [l]
BDNF homodimer [e]	mTORC1:RHEB:GTP:AKT1S1 [l]
BDNF homodimer [e]	mTORC1:RHEB:GTP:p-S183,T246-AKT1S1:YWHAB [l]
BDNF homodimer [e]	MTORC1:RHEB:GTP:p-S758-ULK1:ATG13:RB1CC1:ATG101 [l]
BDNF homodimer [e]	mTORC1:RHEB:GTP:p-T246-AKT1S1 [l]
BDNF homodimer [e]	MTORC1:RHEB:GTP:ULK1:ATG13:RB1CC1:ATG101 [l]
BDNF homodimer [e]	v-ATPase:Ragulator:RRAGA,B:GDP:RRAGC,D:GDP:mTORC1:RHEB:GTP [l]
BDNF homodimer [e]	v-ATPase:Ragulator:RRAGA,B:GTP:RRAGC,D:GDP:mTORC1 [l]
BDNF homodimer [e]	v-ATPase:Ragulator:RRAGA,B:GTP:RRAGC,D:GDP:mTORC1:RHEB:GTP [l]

Studying BDNF connection to mTOR

Start	Target	Result
BDNF homodimer [e]	Active mTORC1 complex [l]	NO
BDNF homodimer [e]	mTORC1 [c]	NO
BDNF homodimer [e]	mTORC1 dimer [n]	NO
BDNF homodimer [e]	mTORC1 with p-S722,S792-RPTOR:Ragulator:Rag:GNP:RHEB:GTP [l]	YES
BDNF homodimer [e]	MTORC1 with p-S722,S792-RPTOR:Ragulator:Rag:RHEB:GTP [l]	YES
BDNF homodimer [e]	MTORC1 with p-S722,S792-RPTOR:RHEB:GTP:p-S758-ULK1:ATG13:RB1CC1:ATG101 [l]	YES
BDNF homodimer [e]	mTORC1:p-T246-AKT1S1:YWHAB [l]	YES
BDNF homodimer [e]	mTORC1:Ragulator:Rag:GNP:RHEB:GDP [l]	YES
BDNF homodimer [e]	MTORC1:Ragulator:Rag:GNP:RHEB:GDP [l]	YES
BDNF homodimer [e]	MTORC1:Ragulator:Rag:GNP:RHEB:GTP [l]	NO
BDNF homodimer [e]	mTORC1:Ragulator:RagA,B:GTP:RagC,D:GDP:SLC38A9 [l]	NO
BDNF homodimer [e]	mTORC1:RHEB:GTP:AKT1S1 [l]	NO
BDNF homodimer [e]	mTORC1:RHEB:GTP:p-S183,T246-AKT1S1:YWHAB [l]	YES
BDNF homodimer [e]	MTORC1:RHEB:GTP:p-S758-ULK1:ATG13:RB1CC1:ATG101 [l]	YES
BDNF homodimer [e]	mTORC1:RHEB:GTP:p-T246-AKT1S1 [l]	YES
BDNF homodimer [e]	MTORC1:RHEB:GTP:ULK1:ATG13:RB1CC1:ATG101 [l]	YES
BDNF homodimer [e]	v-ATPase:Ragulator:RRAGA,B:GDP:RRAGC,D:GDP:mTORC1:RHEB:GTP [l]	NO
BDNF homodimer [e]	v-ATPase:Ragulator:RRAGA,B:GTP:RRAGC,D:GDP:mTORC1 [l]	NO
BDNF homodimer [e]	v-ATPase:Ragulator:RRAGA,B:GTP:RRAGC,D:GDP:mTORC1:RHEB:GTP [l]	NO

Studying BDNF connection to mTOR

Start	Target	Result	Min path
BDNF homodimer [e]	Active mTORC1 complex [l]	NO	-
BDNF homodimer [e]	mTORC1 [c]	NO	-
BDNF homodimer [e]	mTORC1 dimer [n]	NO	-
BDNF homodimer [e]	mTORC1 with p-S722,S792-RPTOR:Ragulator:Rag:GNP:RHEB:GTP [l]	YES	22
BDNF homodimer [e]	MTORC1 with p-S722,S792-RPTOR:Ragulator:Rag:RHEB:GTP [l]	YES	23
BDNF homodimer [e]	MTORC1 with p-S722,S792-RPTOR:RHEB:GTP:p-S758-ULK1:ATG13:RB1CC1:ATG101 [l]	YES	22
BDNF homodimer [e]	mTORC1:p-T246-AKT1S1:YWHAB [l]	YES	13
BDNF homodimer [e]	mTORC1:Ragulator:Rag:GNP:RHEB:GDP [l]	YES	23
BDNF homodimer [e]	MTORC1:Ragulator:Rag:GNP:RHEB:GDP [l]	YES	24
BDNF homodimer [e]	MTORC1:Ragulator:Rag:GNP:RHEB:GTP [l]	NO	-
BDNF homodimer [e]	mTORC1:Ragulator:RagA,B:GTP:RagC,D:GDP:SLC38A9 [l]	NO	-
BDNF homodimer [e]	mTORC1:RHEB:GTP:AKT1S1 [l]	NO	-
BDNF homodimer [e]	mTORC1:RHEB:GTP:p-S183,T246-AKT1S1:YWHAB [l]	YES	14
BDNF homodimer [e]	MTORC1:RHEB:GTP:p-S758-ULK1:ATG13:RB1CC1:ATG101 [l]	YES	26
BDNF homodimer [e]	mTORC1:RHEB:GTP:p-T246-AKT1S1 [l]	YES	12
BDNF homodimer [e]	MTORC1:RHEB:GTP:ULK1:ATG13:RB1CC1:ATG101 [l]	YES	25
BDNF homodimer [e]	v-ATPase:Ragulator:RRAGA,B:GDP:RRAGC,D:GDP:mTORC1:RHEB:GTP [l]	NO	-
BDNF homodimer [e]	v-ATPase:Ragulator:RRAGA,B:GTP:RRAGC,D:GDP:mTORC1 [l]	NO	-
BDNF homodimer [e]	v-ATPase:Ragulator:RRAGA,B:GTP:RRAGC,D:GDP:mTORC1:RHEB:GTP [l]	NO	-

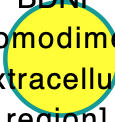
Studying BDNF connection to mTOR

Start	Target	Min path	No Alts Pmin	No Alts Pmin+1	No Alts Pmin+2	No Alts Pmin+3	Target Pathway
BDNF homodimer [e]	mTORC1:RHEB:GTP:p-T246-AKT1S1 [l]	12	1	1	-	-	MTOR
BDNF homodimer [e]	mTORC1:p-T246-AKT1S1:YWHAB [l]	13	1	1	-	-	MTOR
BDNF homodimer [e]	mTORC1:RHEB:GTP:p-S183,T246-AKT1S1:YWHAB [l]	14	1	1	-	-	MTOR
BDNF homodimer [e]	mTORC1 with p-S722,S792-RPTOR:Ragulator:Rag:GNP:RHEB:GTP [l]	22	2	6	10	11	MTOR AMPK
BDNF homodimer [e]	mTORC1:Ragulator:Rag:GNP:RHEB:GDP [l]	23	2	6	10	11	MTOR AMPK TP53
BDNF homodimer [e]	MTORC1 with p-S722,S792-RPTOR:RHEB:GTP:p-S758- ULK1:ATG13:RB1CC1:ATG101 [l]	22	2	6	10	11	Macroautophagy
BDNF homodimer [e]	MTORC1 with p-S722,S792-RPTOR:Ragulator:Rag:RHEB:GTP [l]	23	2	6	10	11	Macroautophagy
BDNF homodimer [e]	MTORC1:Ragulator:Rag:GNP:RHEB:GDP [l]	24	2	6	10	11	Macroautophagy
BDNF homodimer [e]	MTORC1:RHEB:GTP:ULK1:ATG13:RB1CC1:ATG101 [l]	25	2	6	10	11	Macroautophagy
BDNF homodimer [e]	MTORC1:RHEB:GTP:p-S758-ULK1:ATG13:RB1CC1:ATG101 [l]	26	2	6	10	11	Macroautophagy

Studying BDNF connection to mTOR

Start	Target	Min path	No Alts Pmin	No Alts Pmin+1	No Alts Pmin+2	No Alts Pmin+3	Target Pathway
BDNF homodimer [e]	mTORC1:RHEB:GTP:p-T246-AKT1S1 [l]	12	1	1	-	-	MTOR
BDNF homodimer [e]	mTORC1:p-T246-AKT1S1:YWHAB [l]	13	1	1	-	-	MTOR
BDNF homodimer [e]	mTORC1:RHEB:GTP:p-S183,T246-AKT1S1:YWHAB [l]	14	1	1	-	-	MTOR
BDNF homodimer [e]	mTORC1 with p-S722,S792-RPTOR:Ragulator:Rag:GNP:RHEB:GTP [l]	22	2	6	10	11	MTOR AMPK
BDNF homodimer [e]	mTORC1:Ragulator:Rag:GNP:RHEB:GDP [l]	23	2	6	10	11	MTOR AMPK TP53
BDNF homodimer [e]	MTORC1 with p-S722,S792-RPTOR:RHEB:GTP:p-S758- ULK1:ATG13:RB1CC1:ATG101 [l]	22	2	6	10	11	Macroautophagy
BDNF homodimer [e]	MTORC1 with p-S722,S792-RPTOR:Ragulator:Rag:RHEB:GTP [l]	23	2	6	10	11	Macroautophagy
BDNF homodimer [e]	MTORC1:Ragulator:Rag:GNP:RHEB:GDP [l]	24	2	6	10	11	Macroautophagy
BDNF homodimer [e]	MTORC1:RHEB:GTP:ULK1:ATG13:RB1CC1:ATG101 [l]	25	2	6	10	11	Macroautophagy
BDNF homodimer [e]	MTORC1:RHEB:GTP:p-S758-ULK1:ATG13:RB1CC1:ATG101 [l]	26	2	6	10	11	Macroautophagy

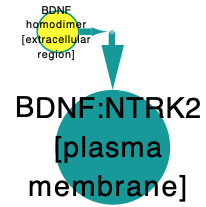
Studying BDNF connection to mTOR



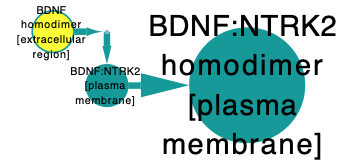
BDNF
homodimer
[extracellular
region]

A diagram showing a BDNF homodimer in its extracellular region. It is represented by a yellow circle with a blue outline. The text 'BDNF homodimer [extracellular region]' is positioned to the left of the circle.

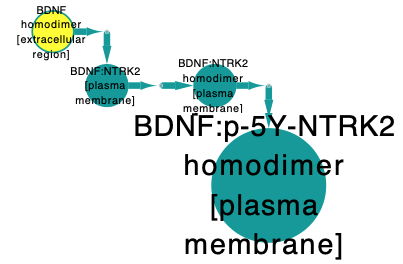
Studying BDNF connection to mTOR



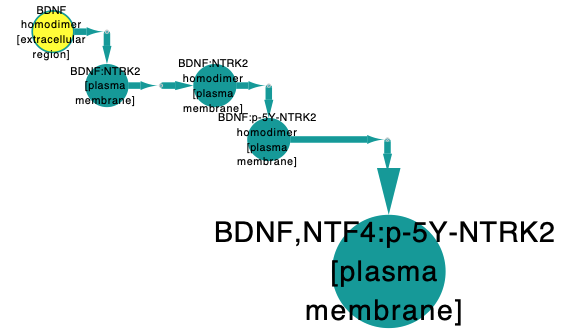
Studying BDNF connection to mTOR



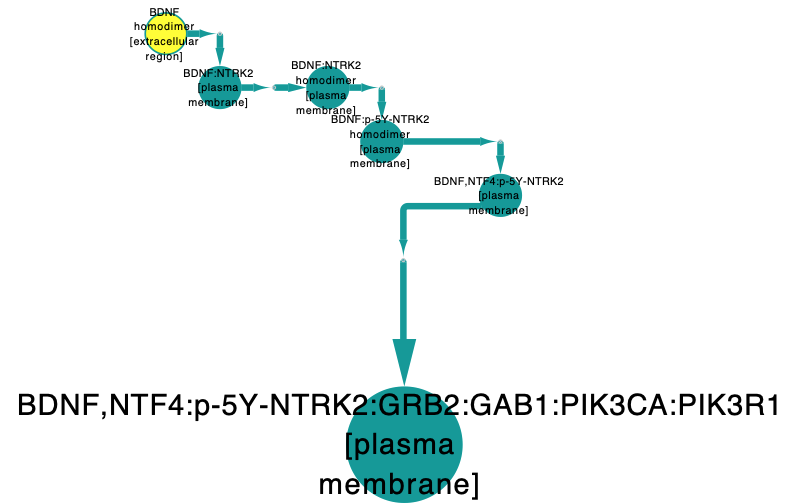
Studying BDNF connection to mTOR



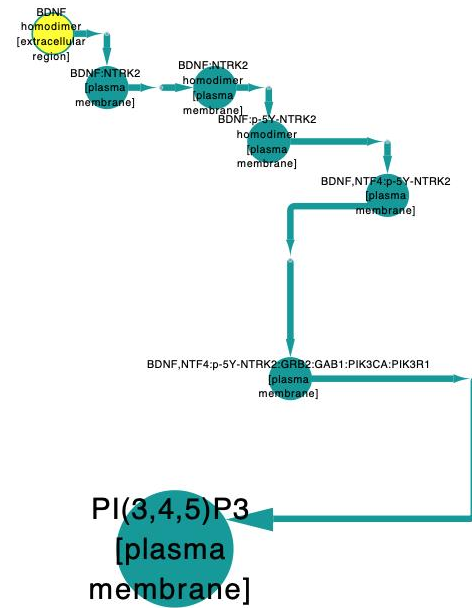
Studying BDNF connection to mTOR



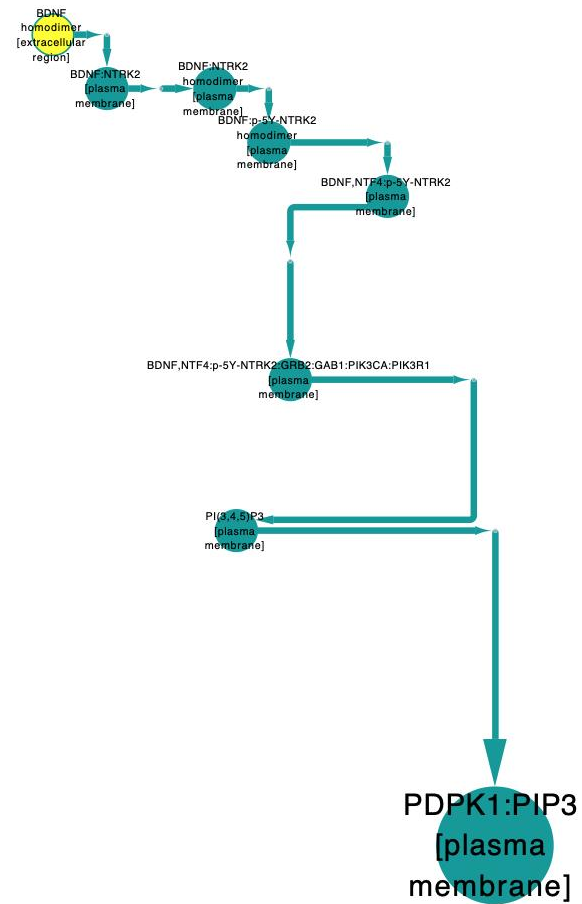
Studying BDNF connection to mTOR



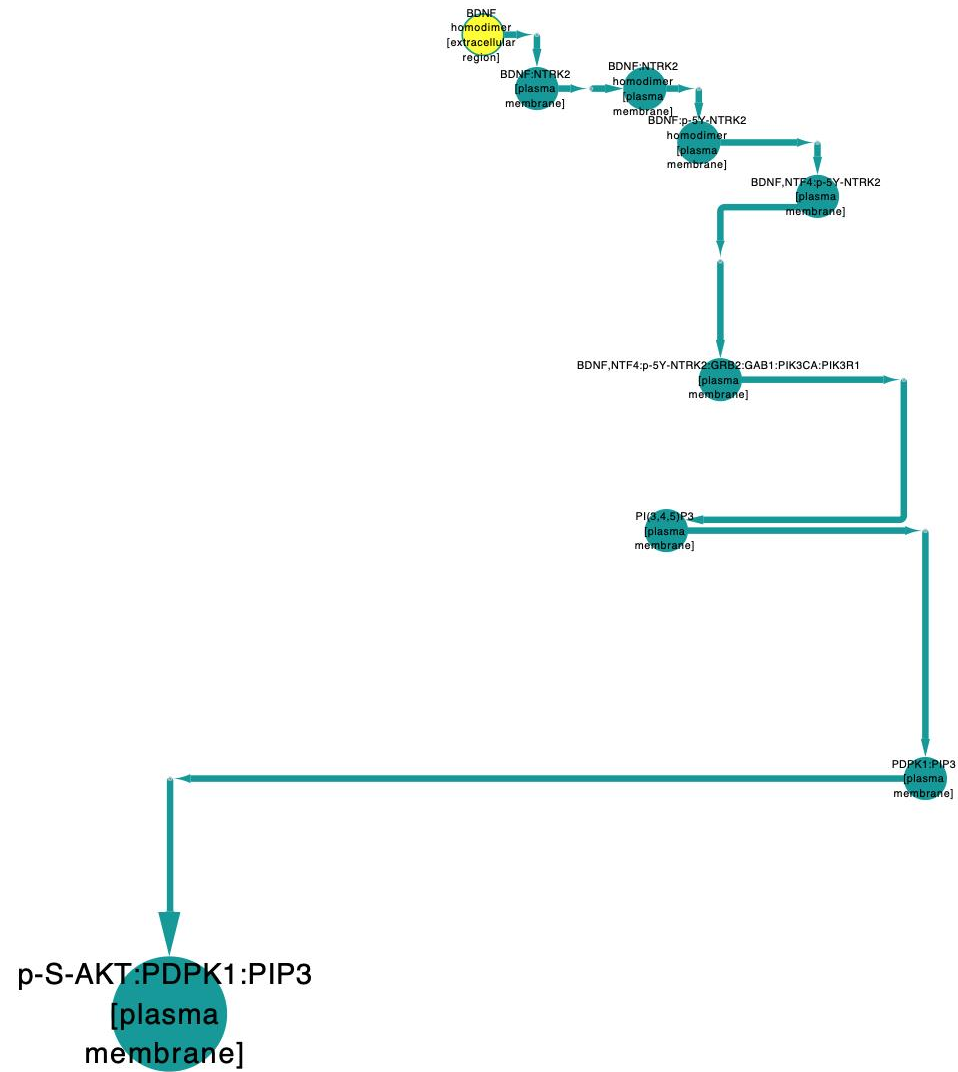
Studying BDNF connection to mTOR



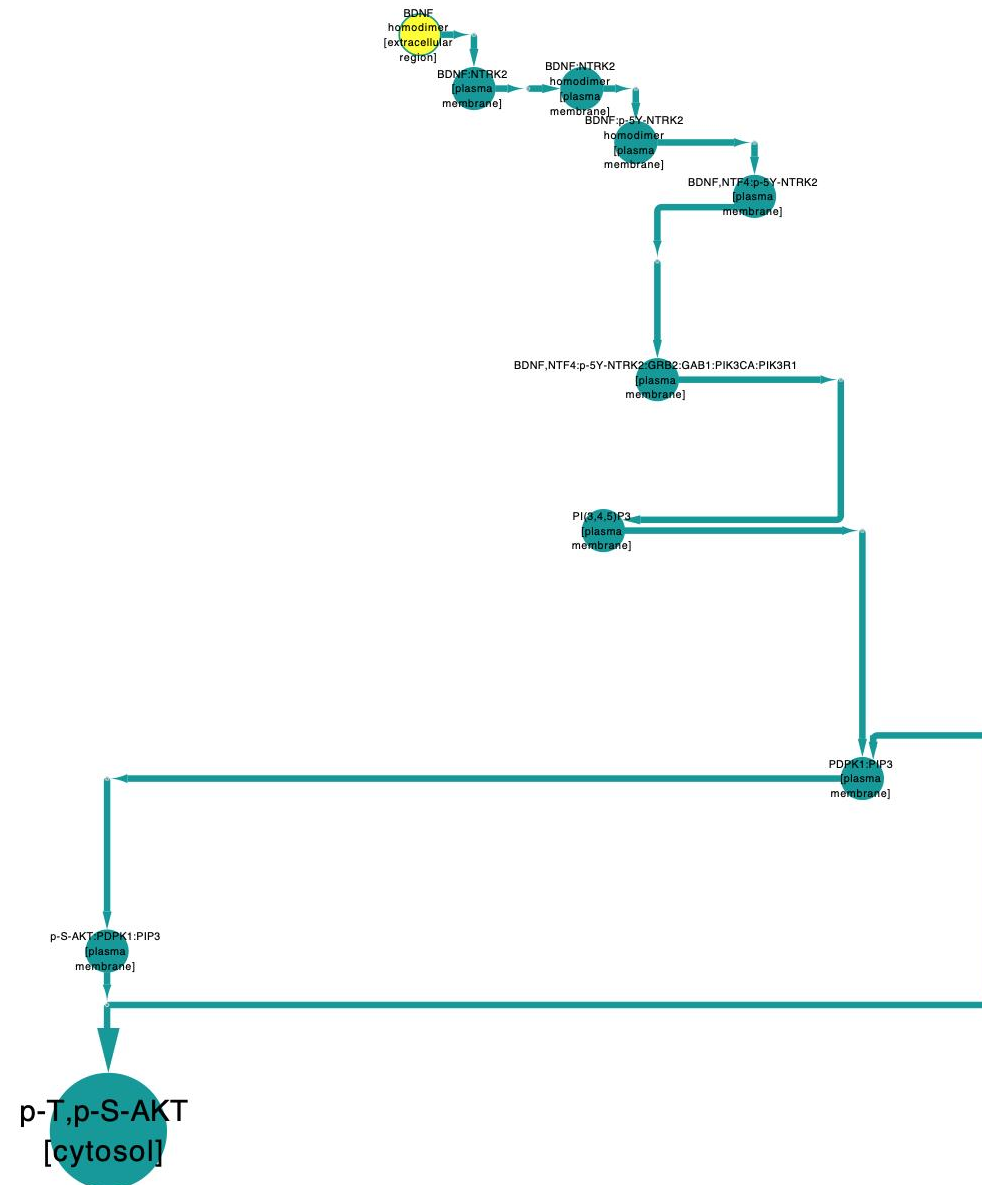
Studying BDNF connection to mTOR



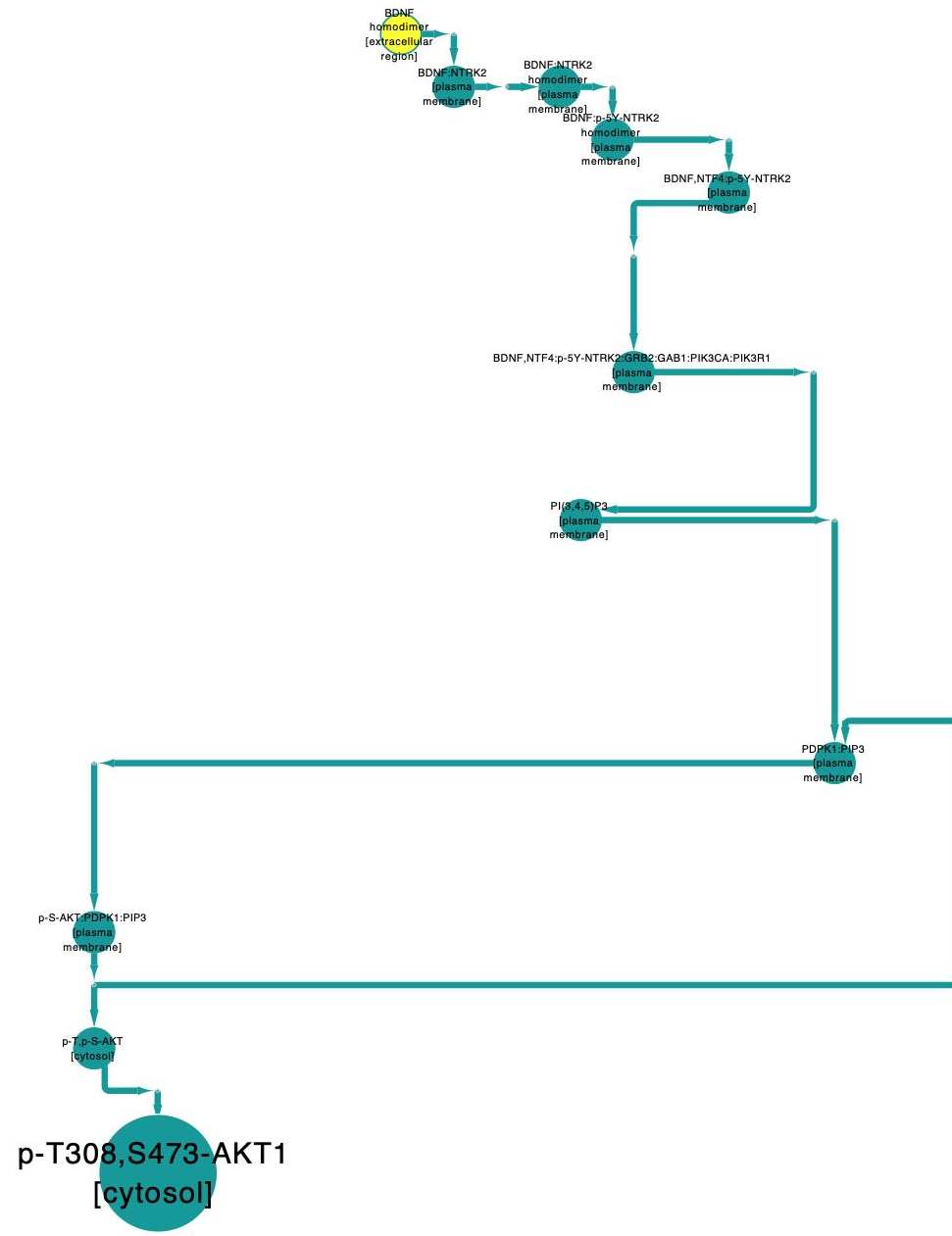
Studying BDNF connection to mTOR



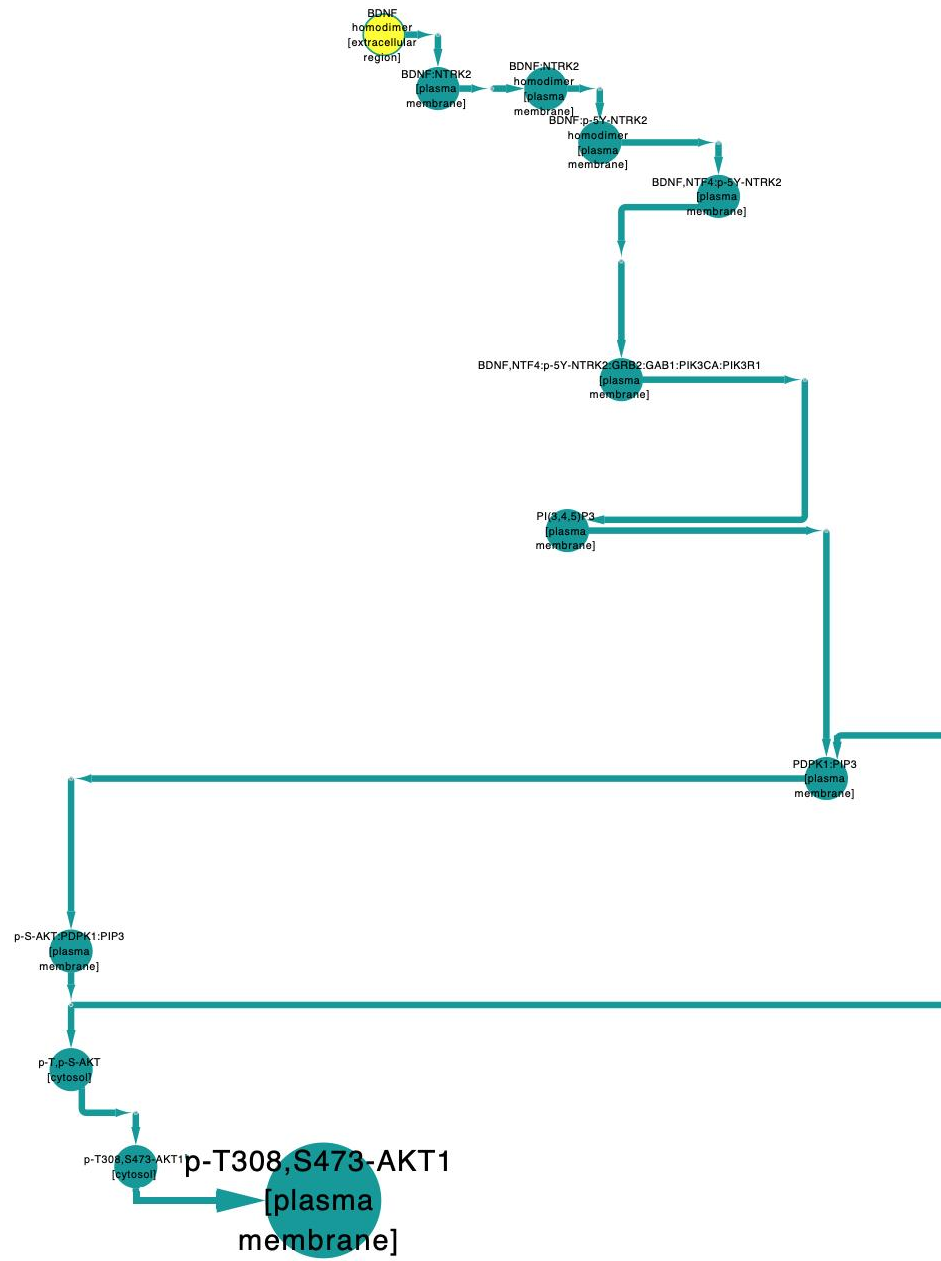
Studying BDNF connection to mTOR



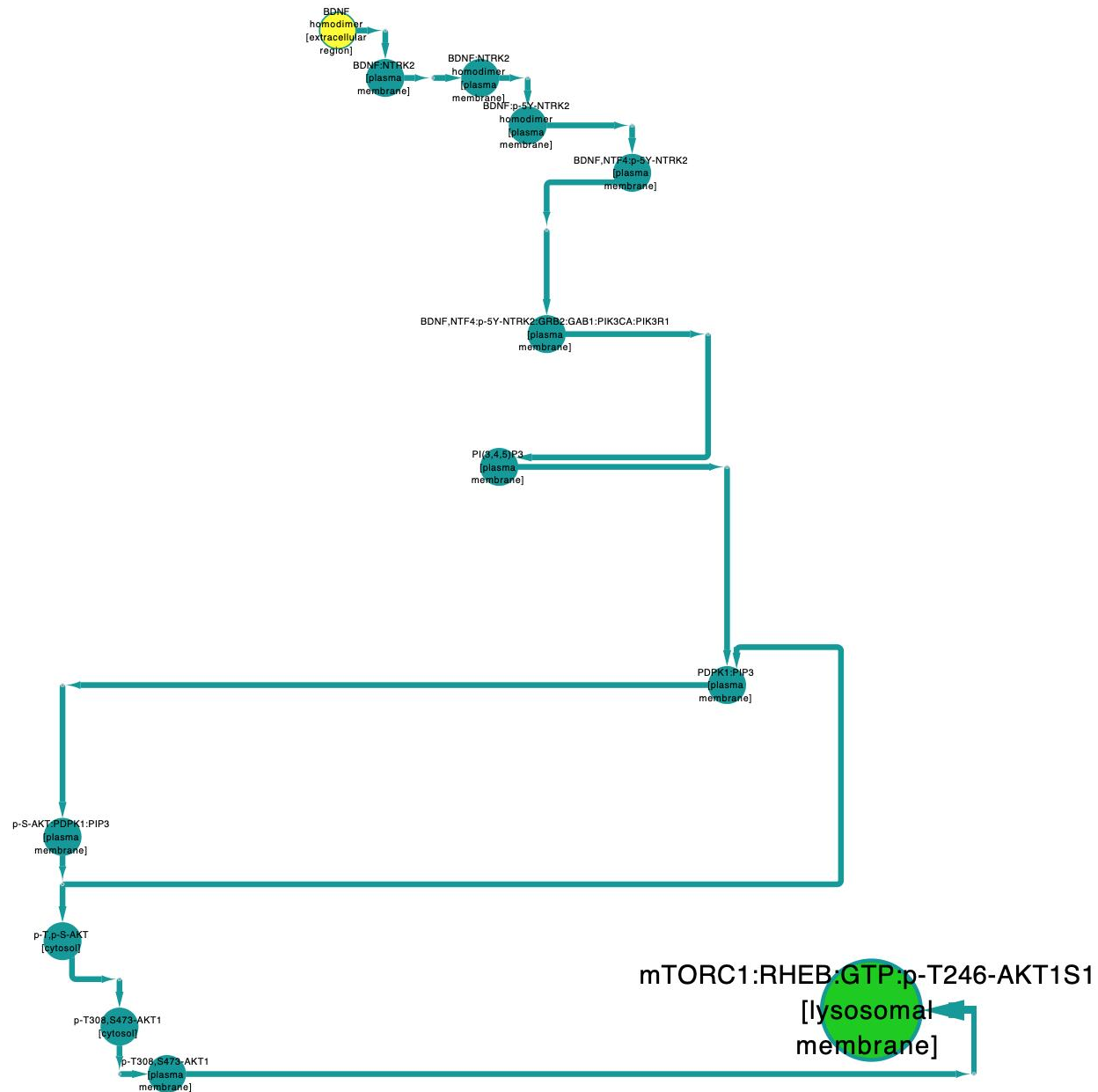
Studying BDNF connection to mTOR



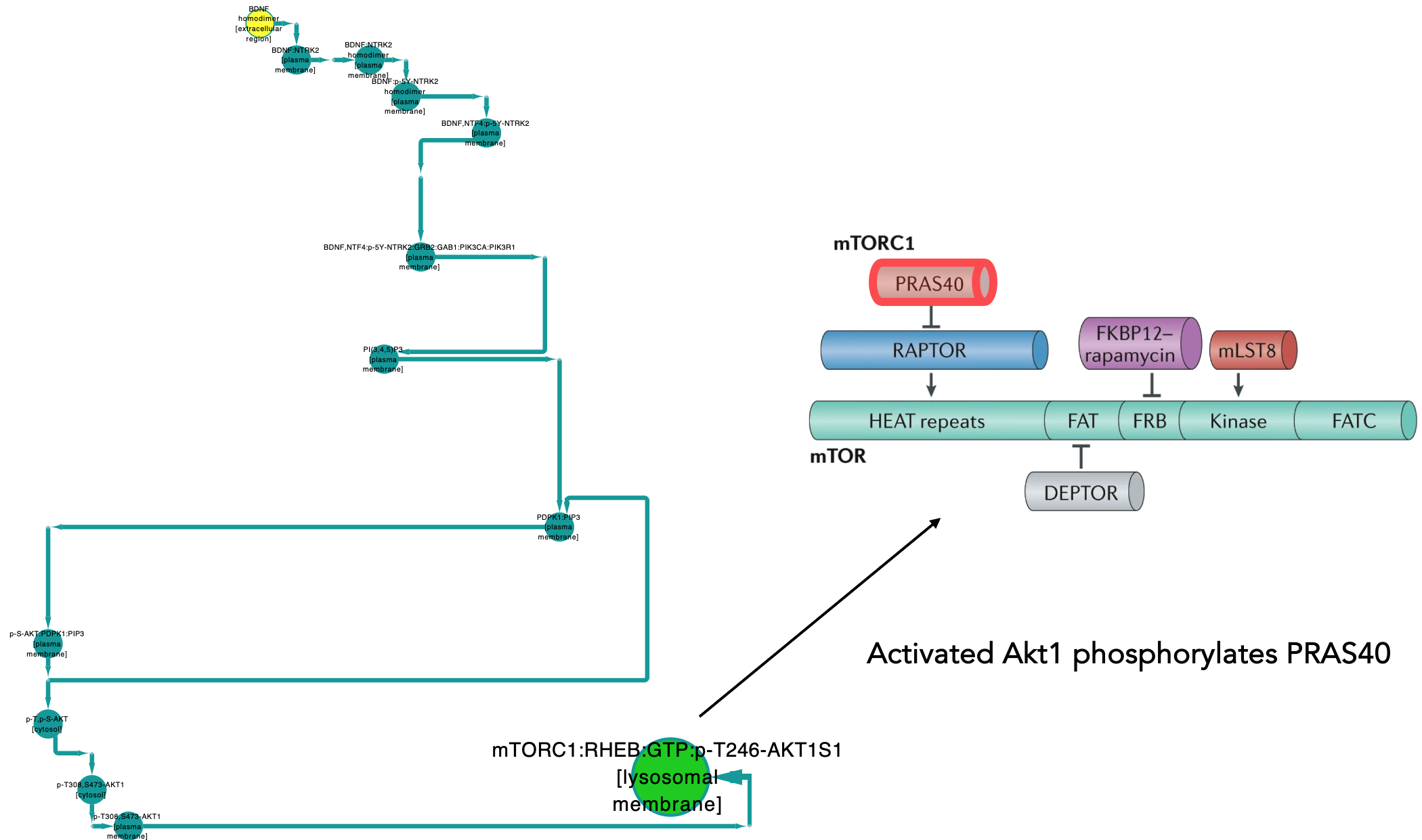
Studying BDNF connection to mTOR



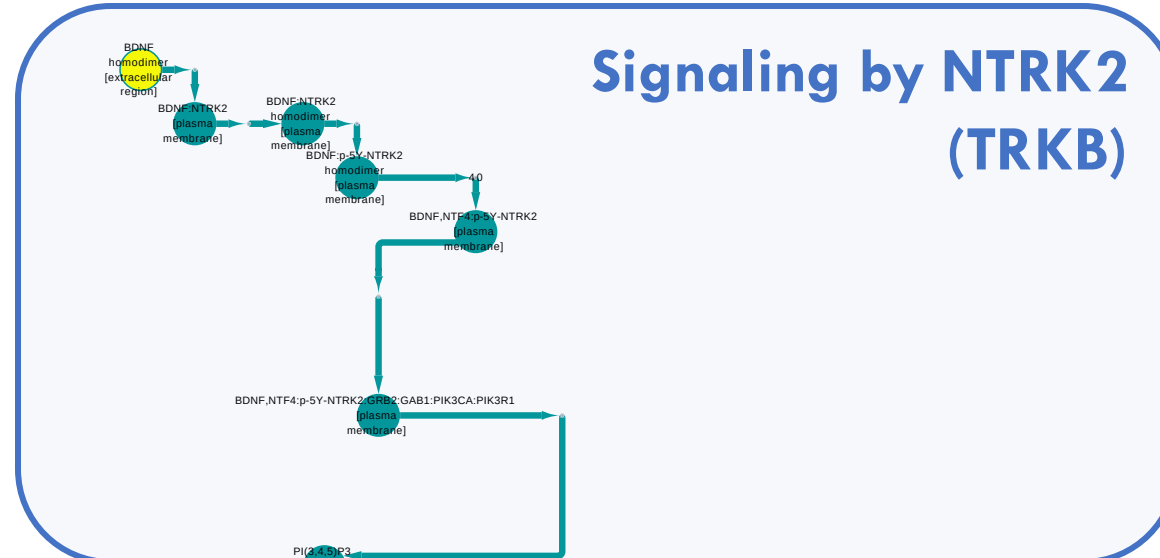
Studying BDNF connection to mTOR



Studying BDNF connection to mTOR



Studying BDNF connection to mTOR

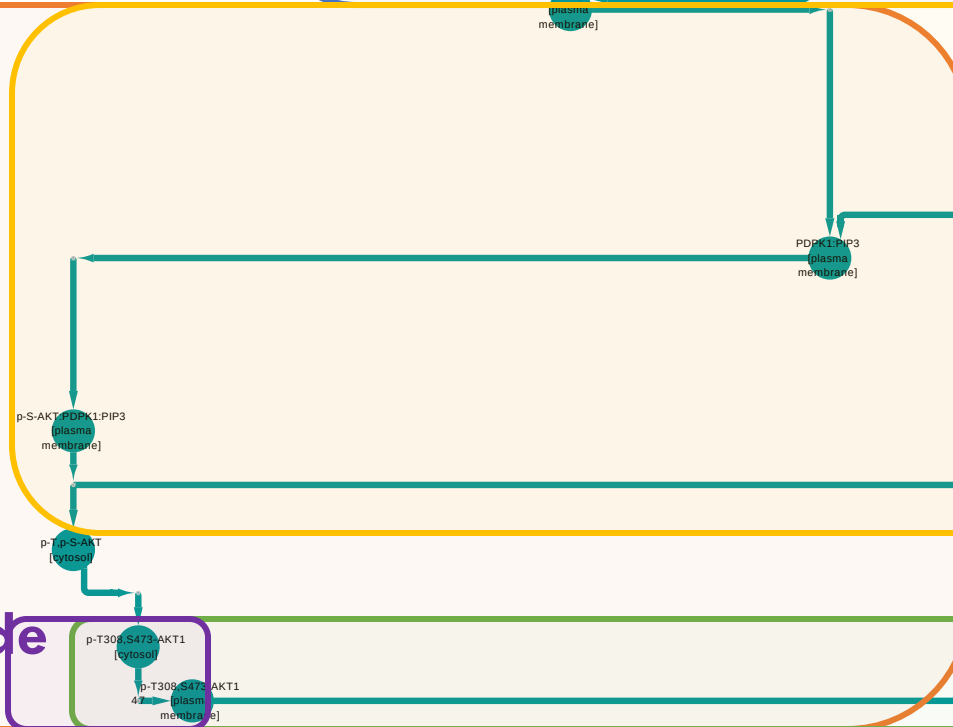


12 connections
13 species
5 pathways

**PIP3
activates
AKT
signaling**

**Costimulation
by the CD28
family**

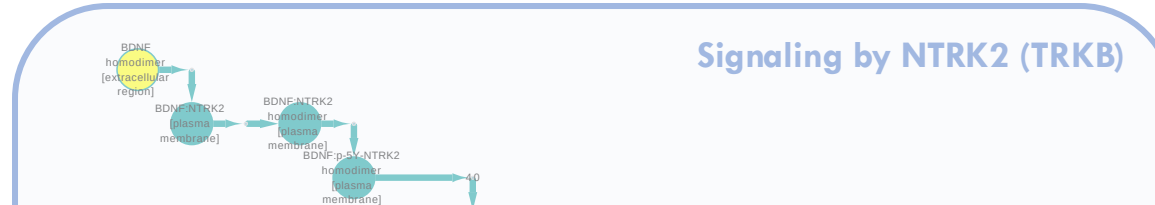
**Metabolism
of nitric oxide**



mTORC1:RHEB, GTP, p-T246-AKT1S1
[mitochondrion membrane]

MTOR signaling

Studying BDNF connection to mTOR

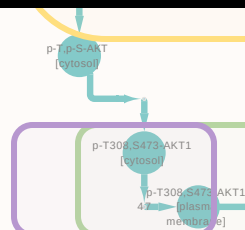


What are the surrounding species that have to be active in order for this path to be active?

Can we identify crosstalks?

PIP3 active
AKT
signaling

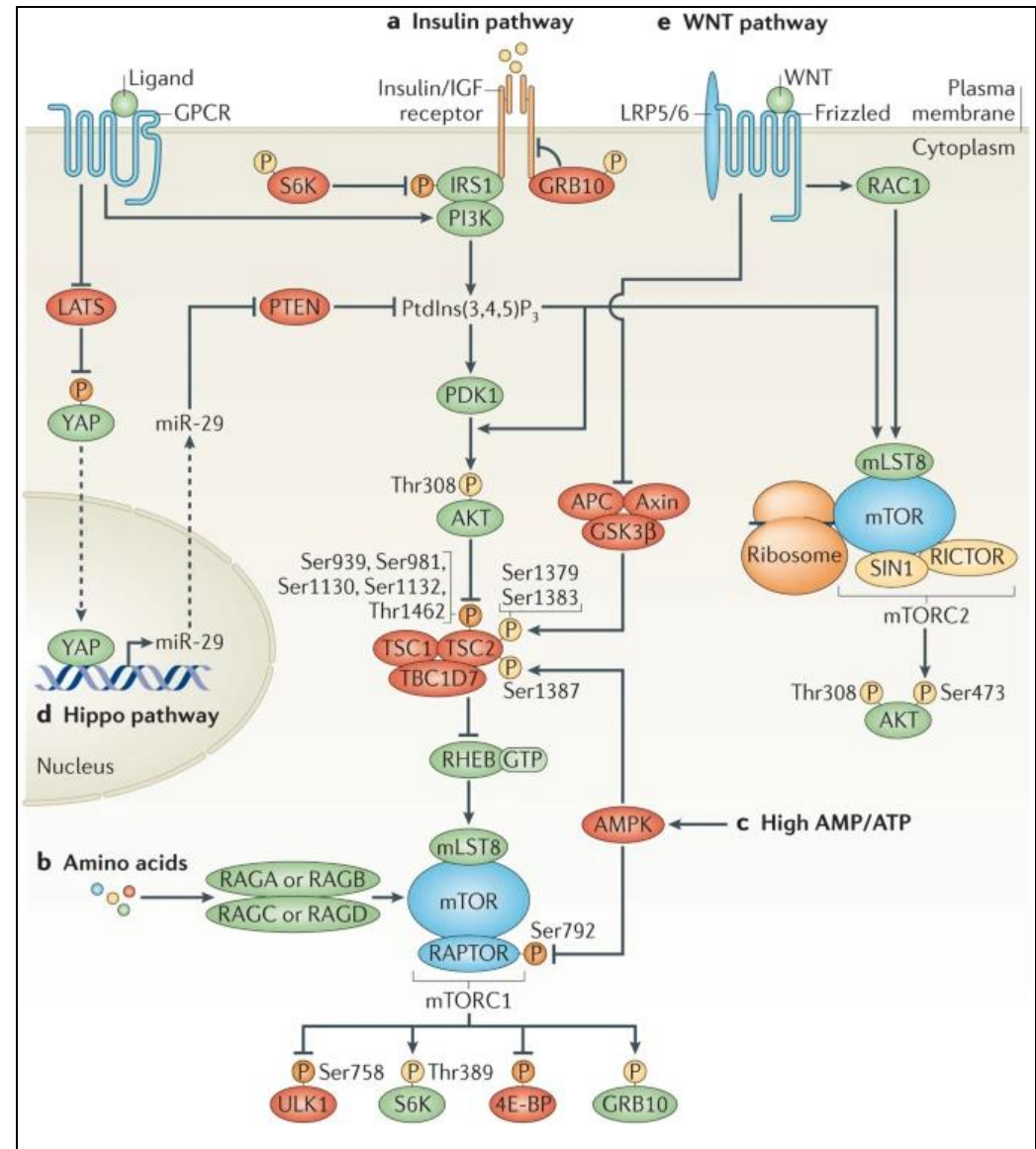
Metabolism of
nitric oxide:
NOS3
activation and
regulation



mTORC1-RHEB-GTP-p-T246-AKT1S1
[cytosol]
[plasma membrane]

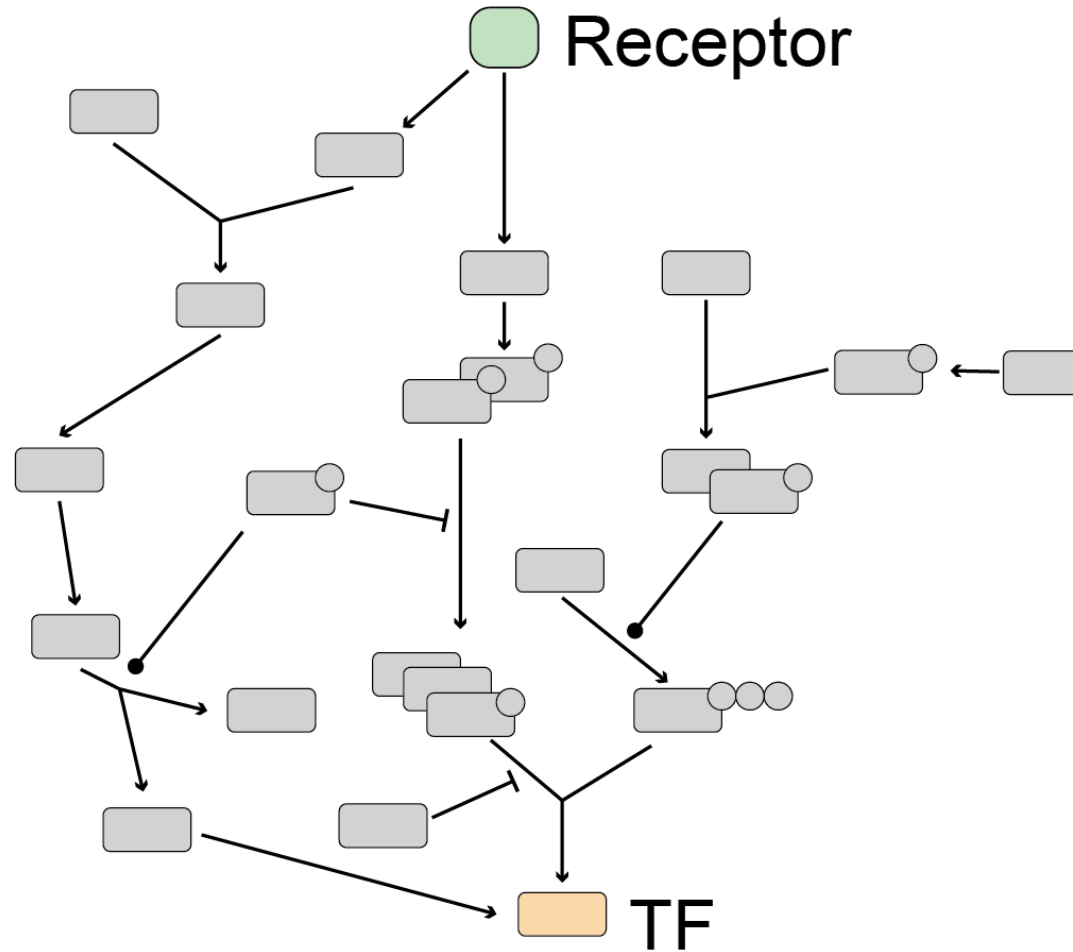
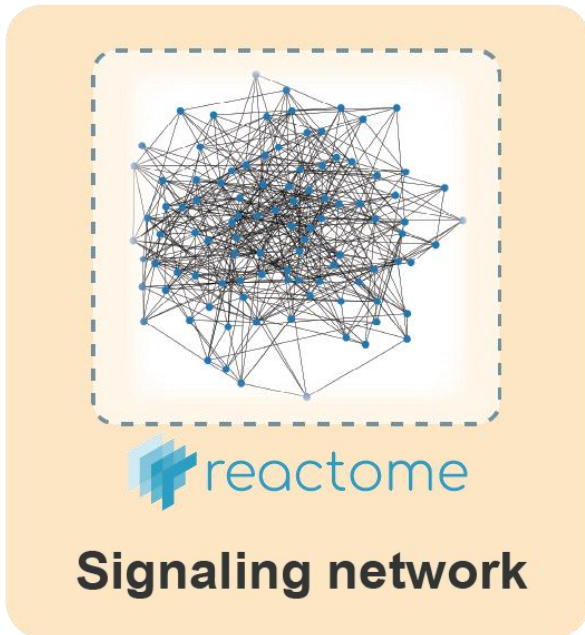
MTOR signaling

Crosstalks in signaling pathways



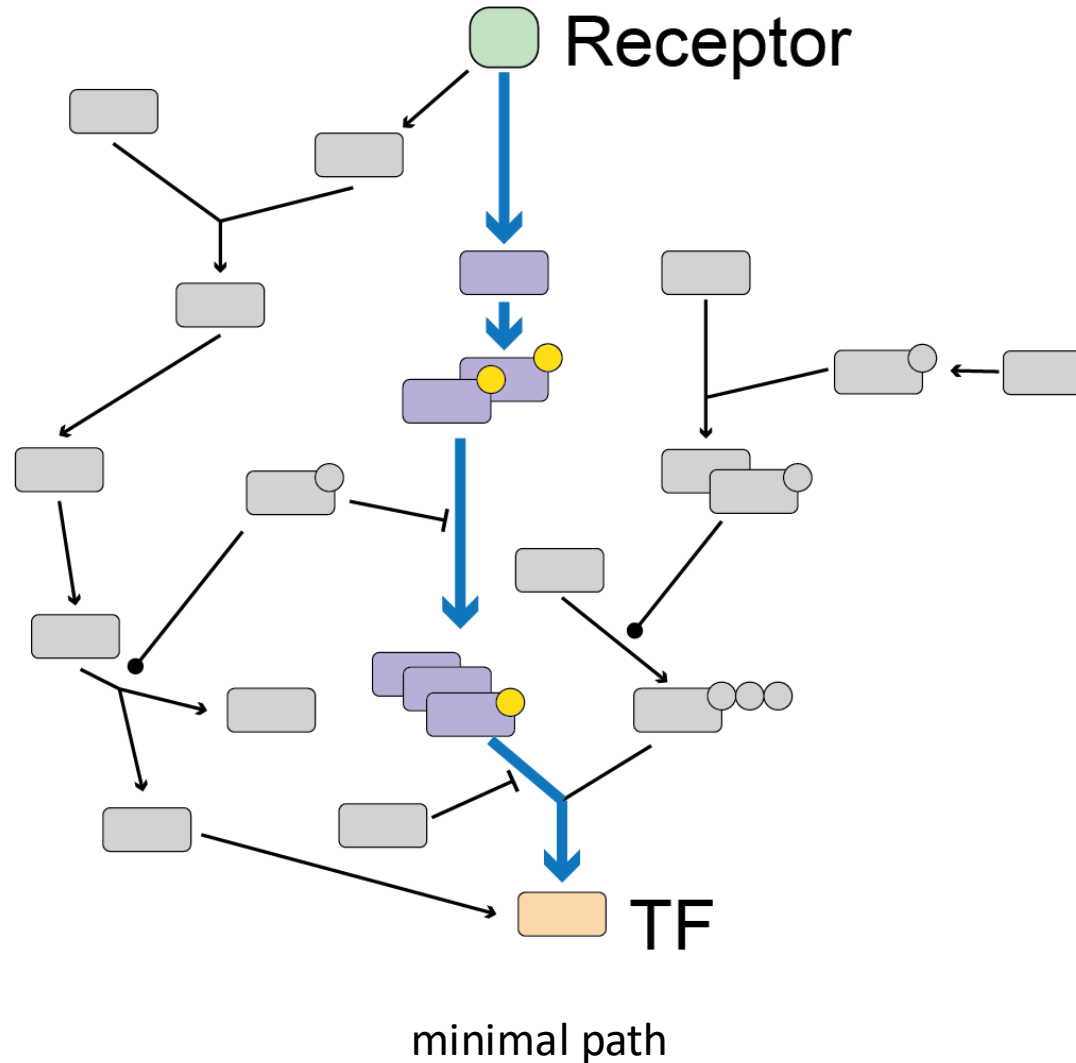
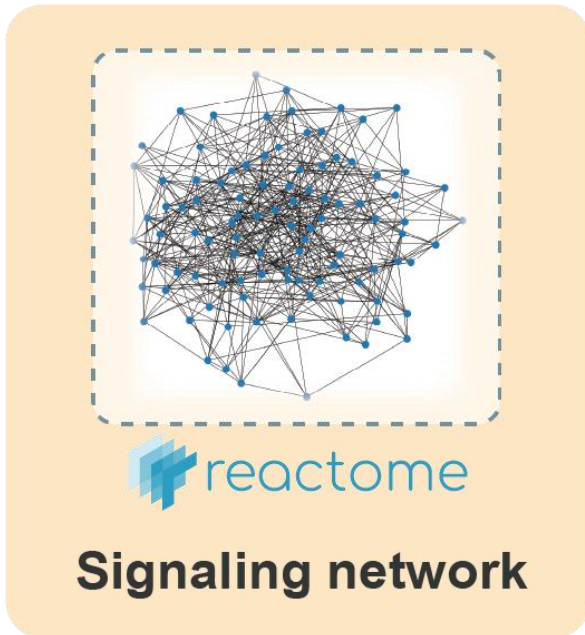
Create balanced signaling networks

SIGMA Method



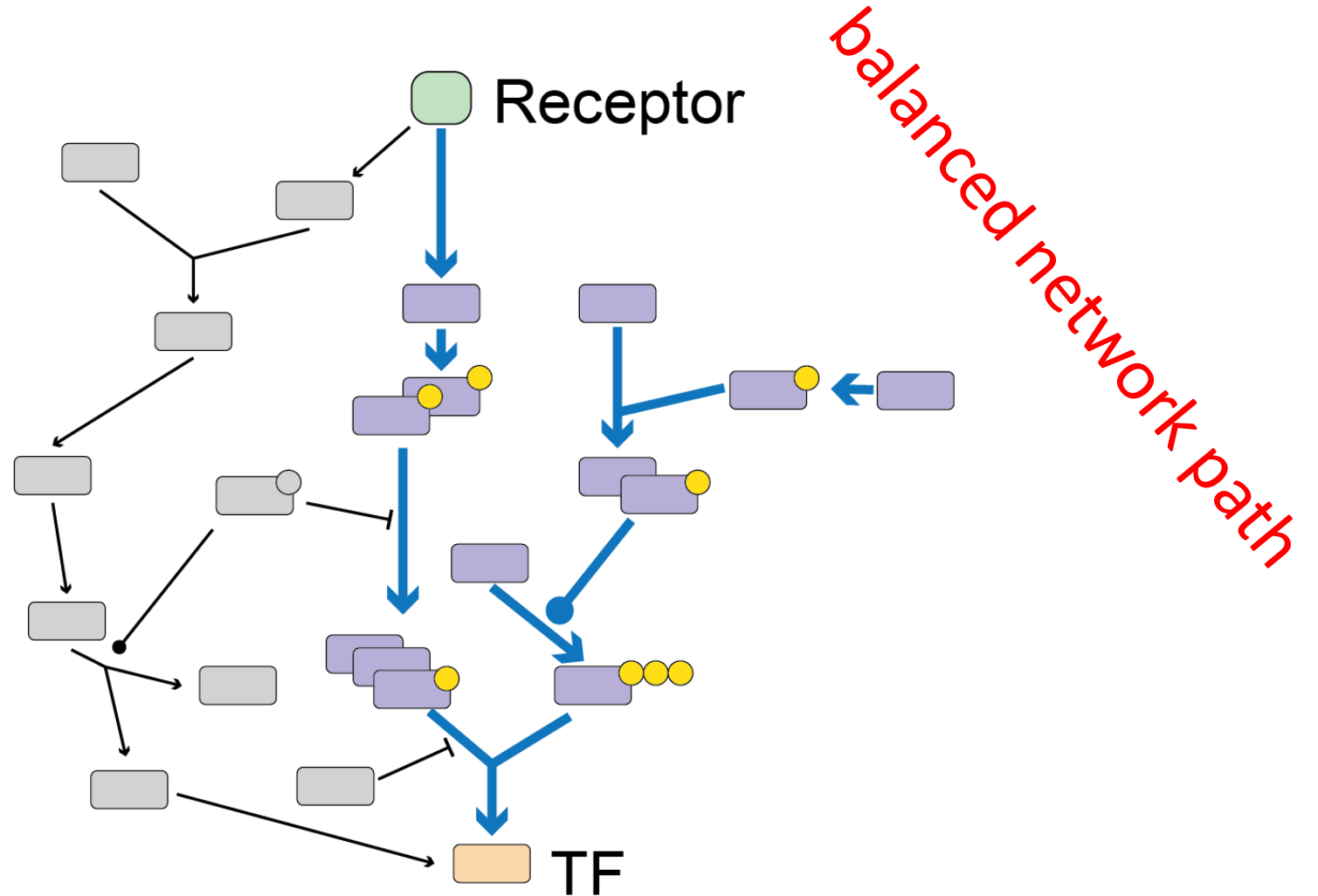
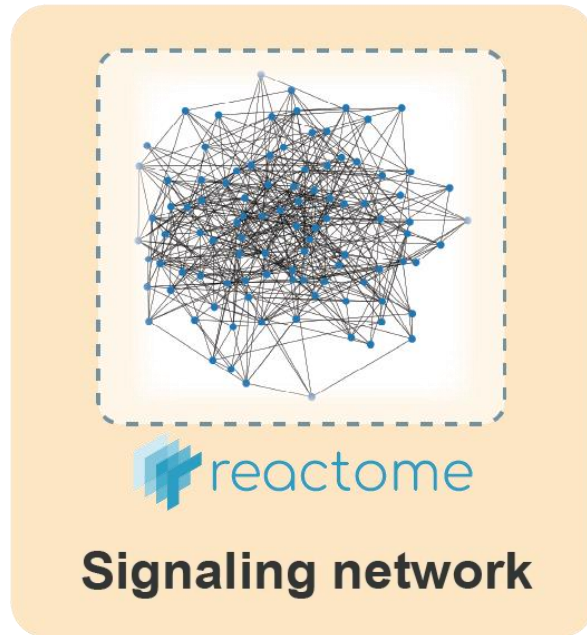
Create balanced signaling networks

SIGMA Method



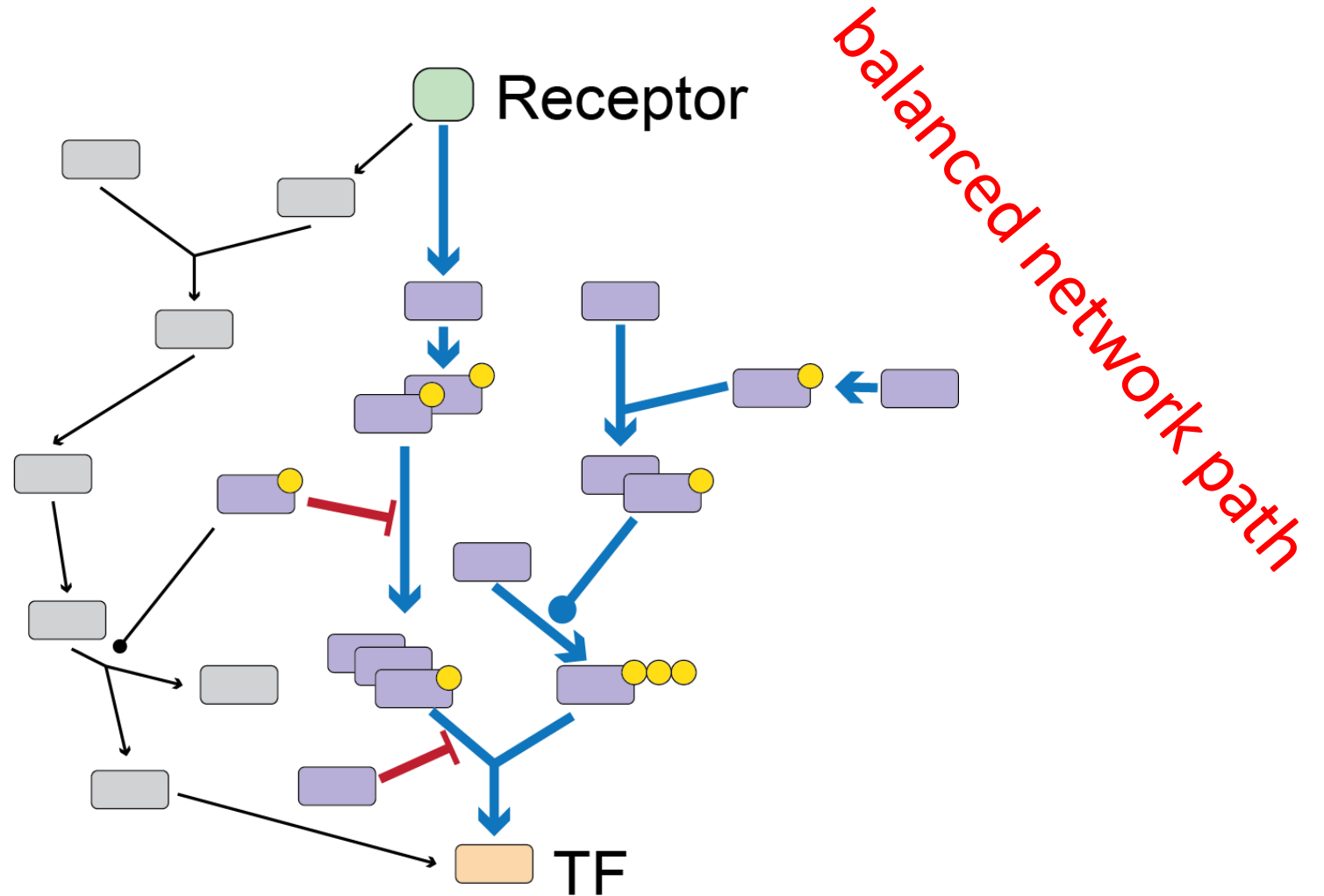
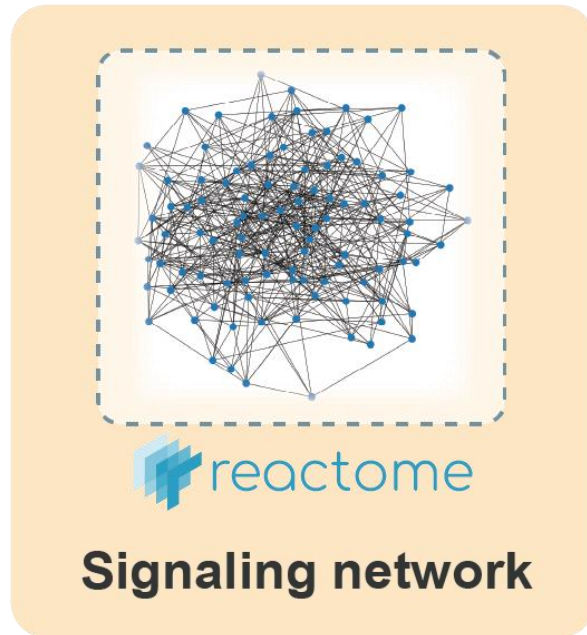
Create balanced signaling networks

SIGMA Method



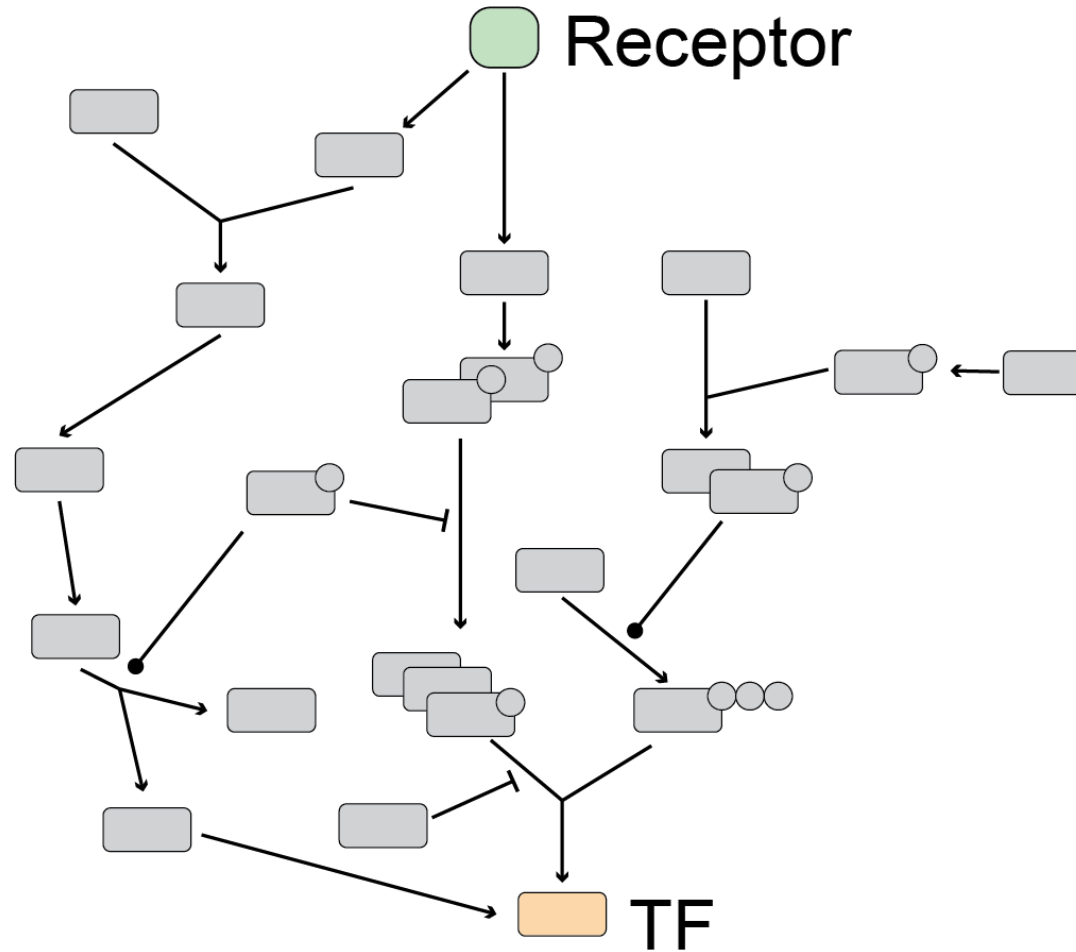
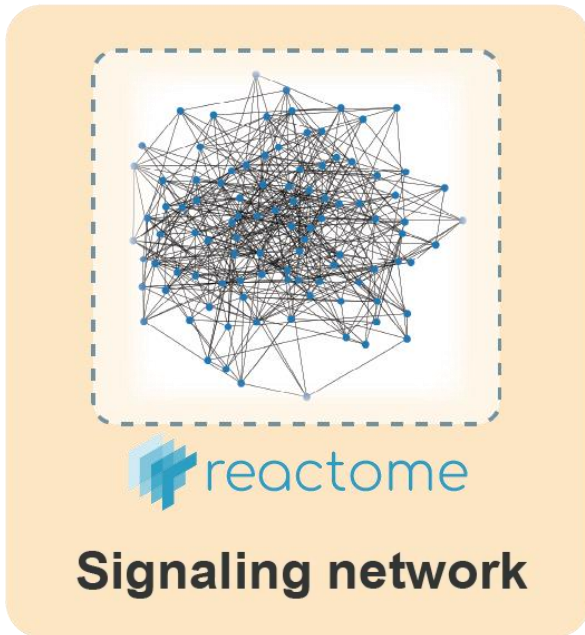
Create balanced signaling networks

SIGMA Method



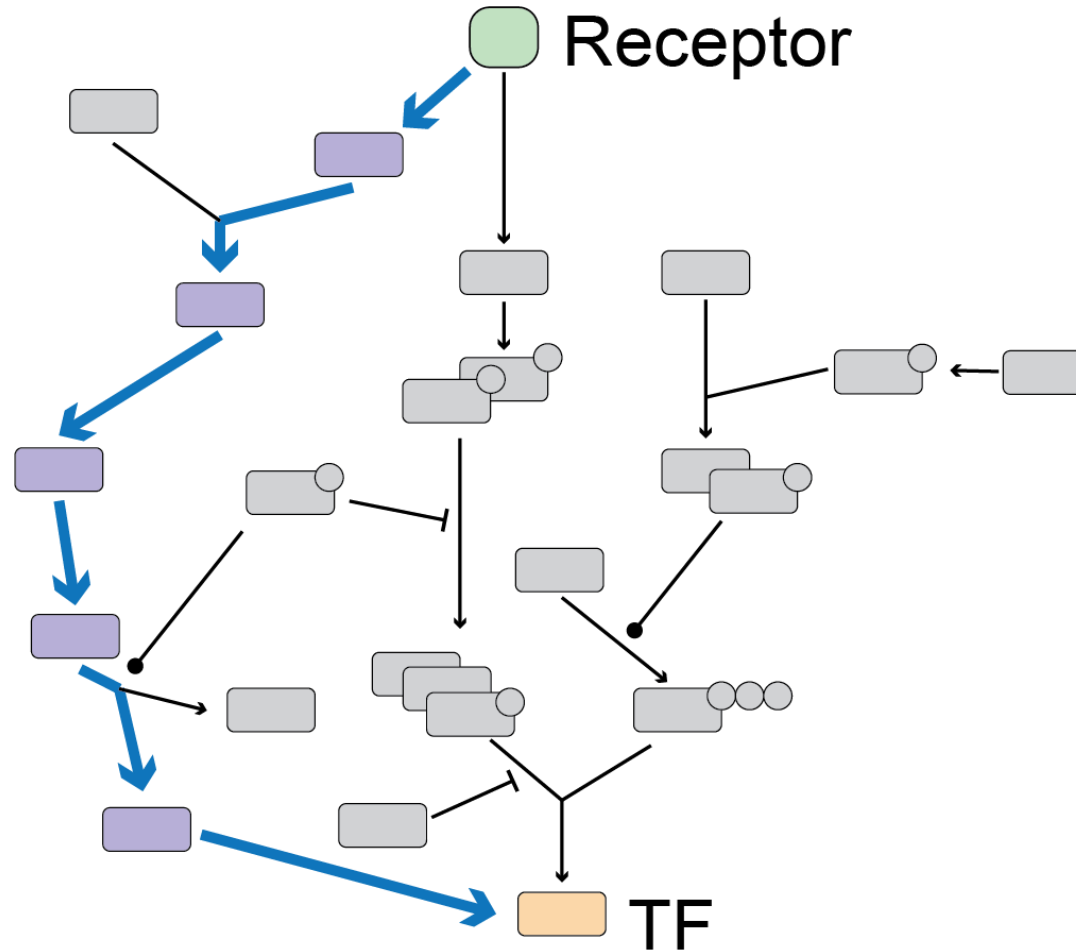
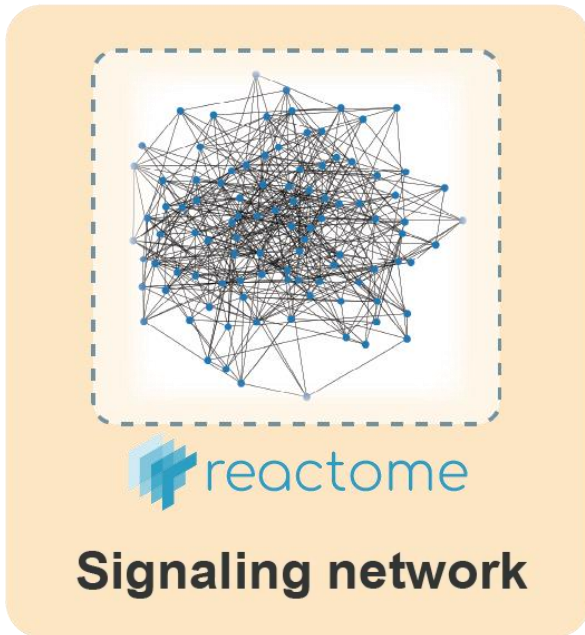
Create balanced signaling networks

SIGMA Method



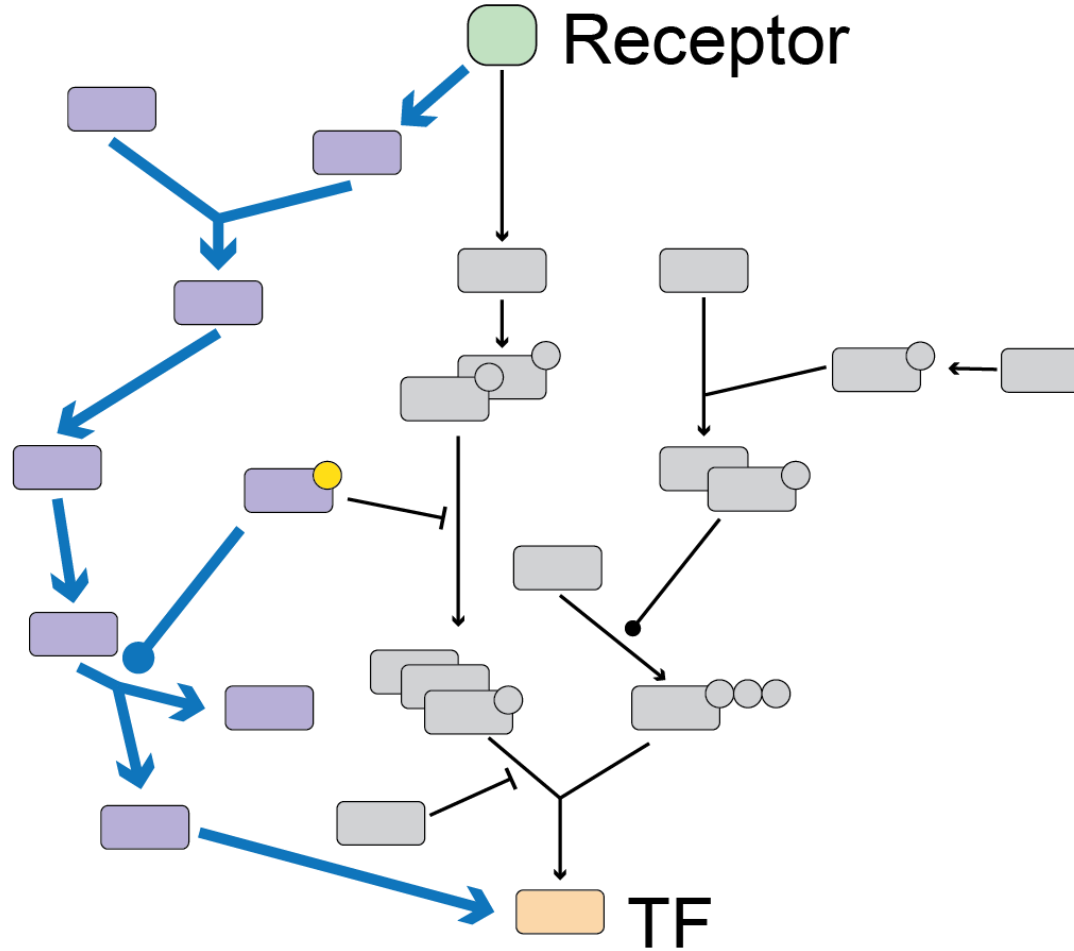
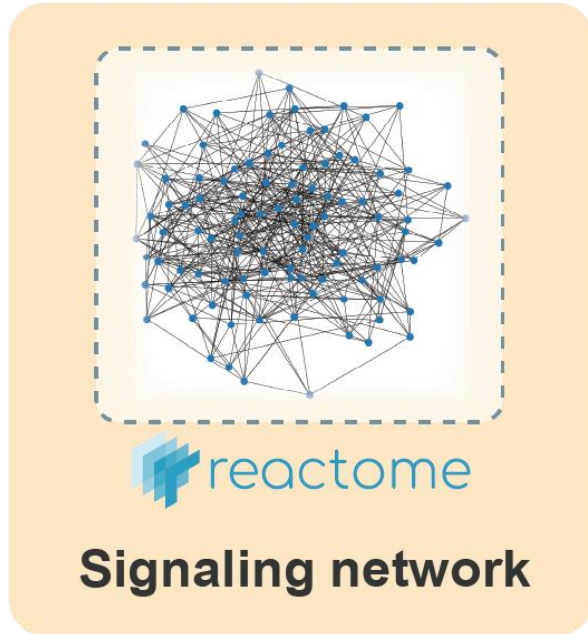
Create balanced signaling networks

SIGMA Method



Create balanced signaling networks

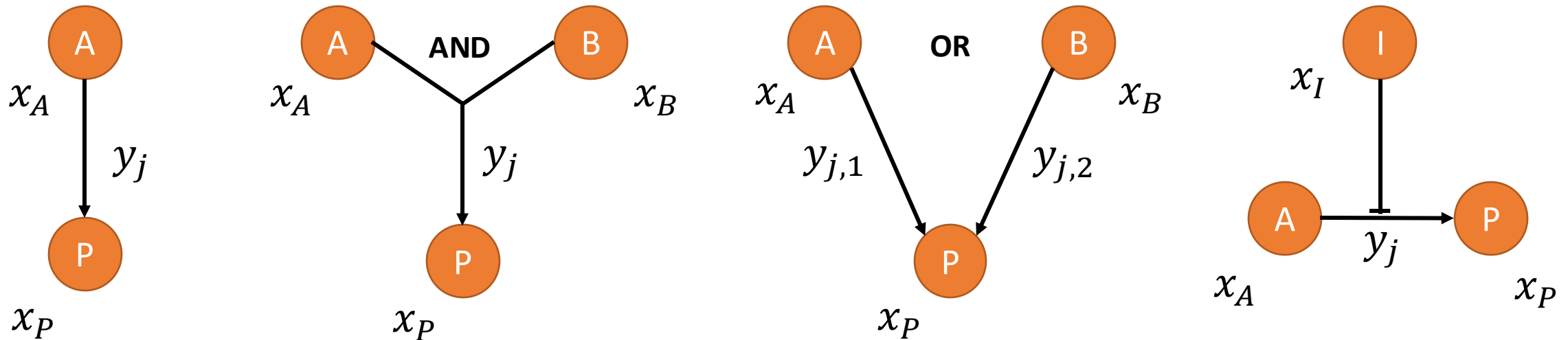
SIGMA Method



Method for generating balanced signaling networks

From species connections to reactions

- It is based on optimizing pathway topology
- The reactant-reaction-product relations are the motifs



Method for generating balanced signaling networks

A hybrid sequential method

1) PathTracer algorithm to identify the minimal path between two species.

**Biotechnology
Journal**
www.biotechnology-journal.com

Biotechnol. J. 2016, 11, 648–661

Research Article

MapMaker and PathTracer for tracking carbon in genome-scale metabolic models

Christopher J. Tervo and Jennifer L. Reed

Method for generating balanced signaling networks

A hybrid sequential method

1) PathTracer algorithm to identify the minimal path between two species.

2) Oftadeh et al. formulation of the Mitsos et al. method to identify the minimal network that balances the minimal path.

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OPEN ACCESS Freely available online

PLOS COMPUTATIONAL BIOLOGY

Identifying Drug Effects via Pathway Alterations using an Integer Linear Programming Optimization Formulation on Phosphoproteomic Data

Alexander Mitsos¹, Ioannis N. Melas², Paraskeuas Siminelakis², Aikaterini D. Chairakaki², Julio Saez-Rodriguez^{3,4}, Leonidas G. Alexopoulos^{2*}

Journal of Receptors and Signal Transduction

Volume 38, 2018 - Issue 5-6

Research Article

Accounting for robustness in modeling signal transduction responses

Mohammad Omid Oftadeh & Sayed-Amir Marashi 

Pages 442-447 | Received 20 Aug 2018, Accepted 25 Dec 2018, Published online: 22 Feb 2019

Method for generating balanced signaling networks

Network for minimal path

Pathways

115

Species

5811

Reactions

8696

**From connections
to reactions**



Network for balanced path

Pathways

115

Species

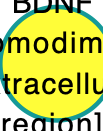
5947

Reactions

4409

Studying BDNF connection to mTOR

BDNF
homodimer
[extracellular
region]



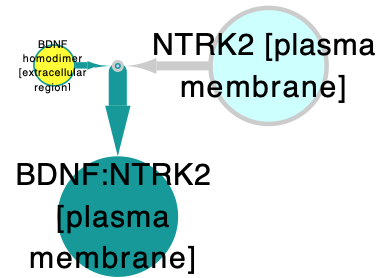
Minimal path

12 connections
13 species
5 pathways

Pathways in
minimal path

1. Signaling by NTRK2 (TRKB)
2. PIP3 activates AKT signaling
3. Costimulation by the CD28 family
4. Metabolism of nitric oxide: NOS3 activation and regulation
5. MTOR signaling

Studying BDNF connection to mTOR



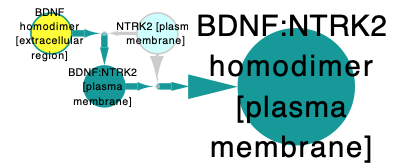
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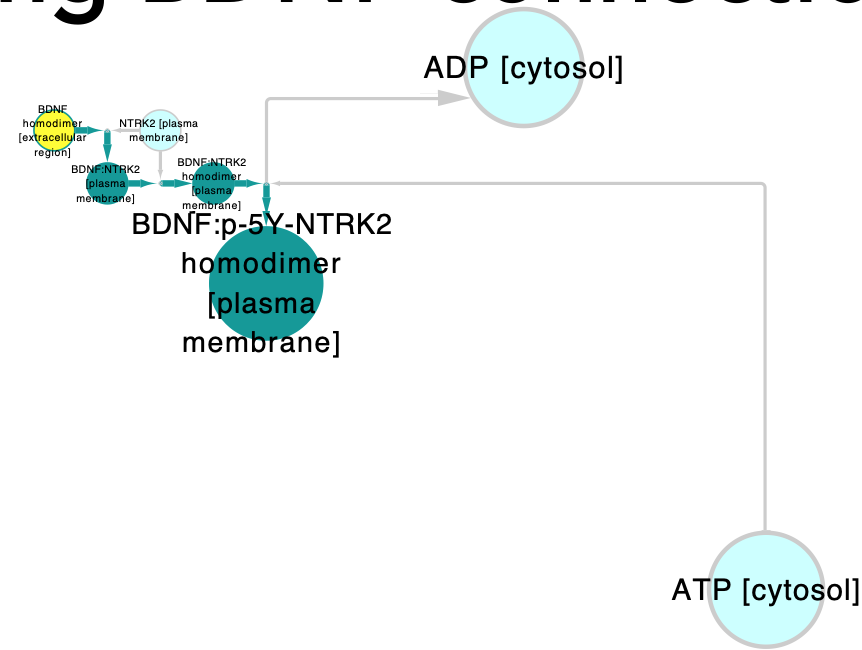
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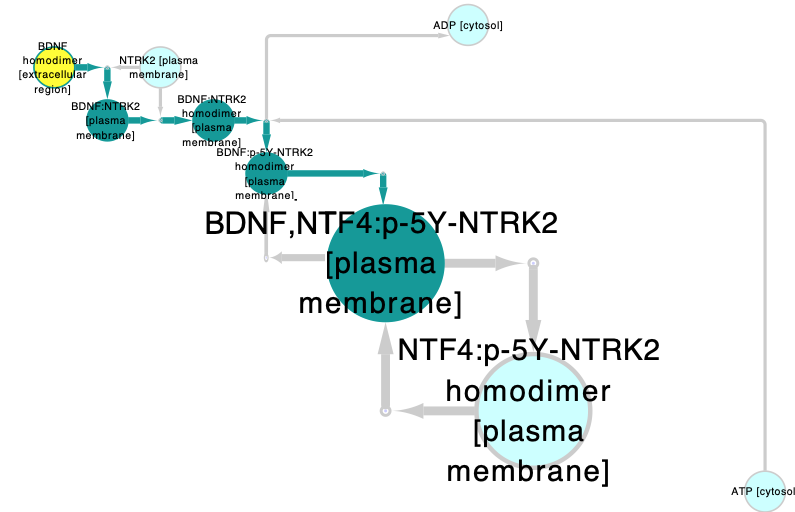
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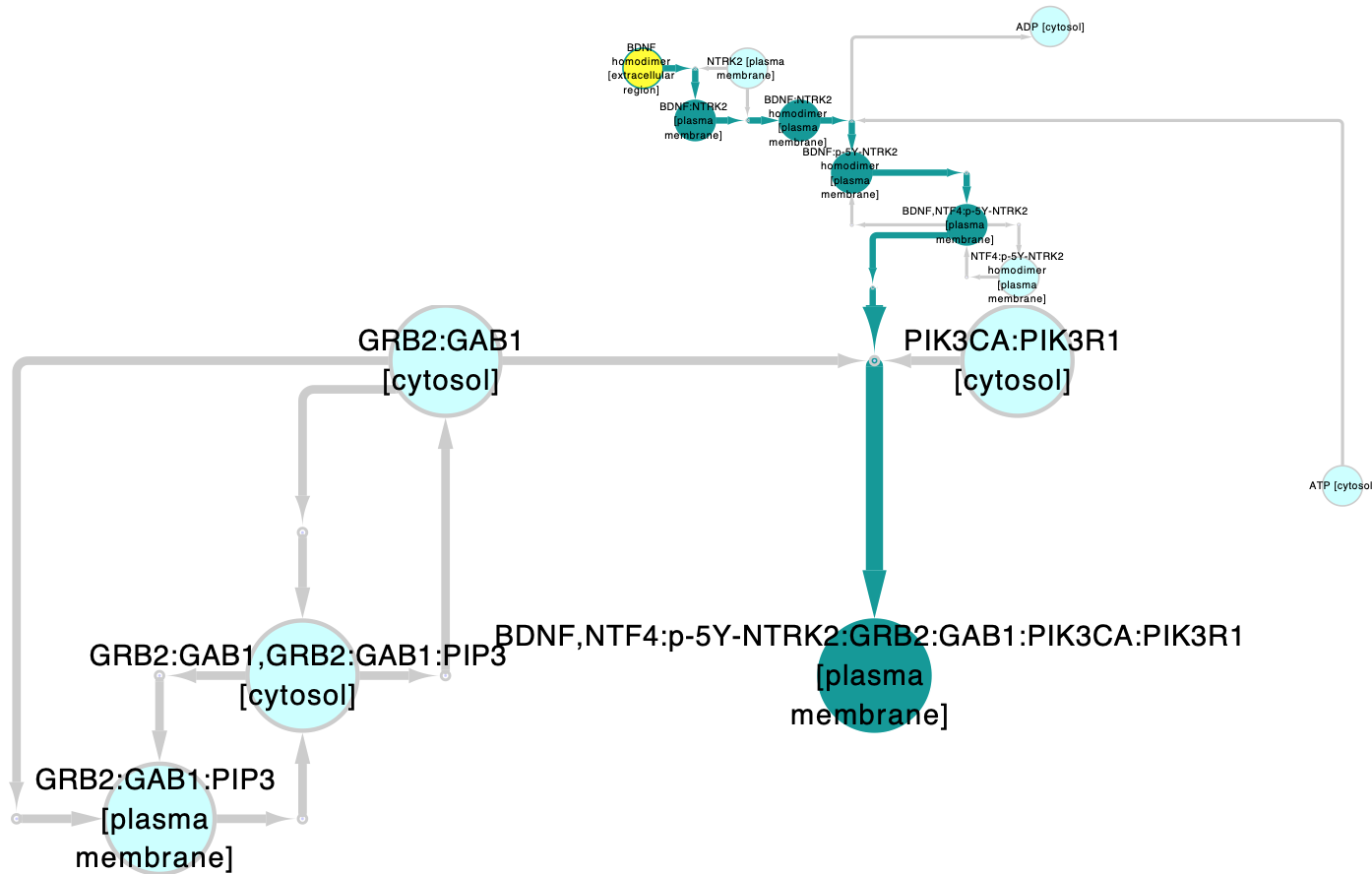
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Studying BDNF connection to mTOR



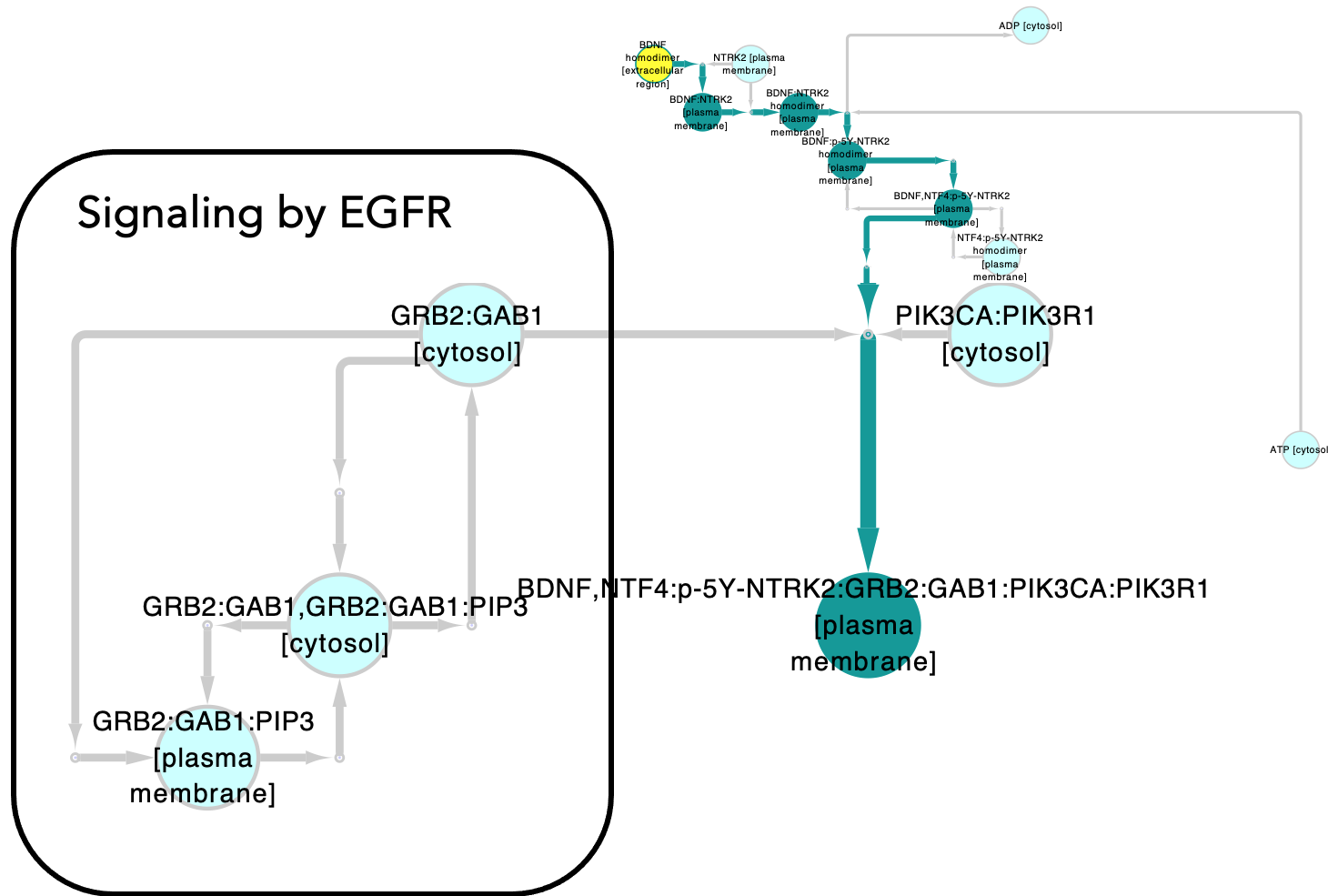
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Studying BDNF connection to mTOR



Minimal path

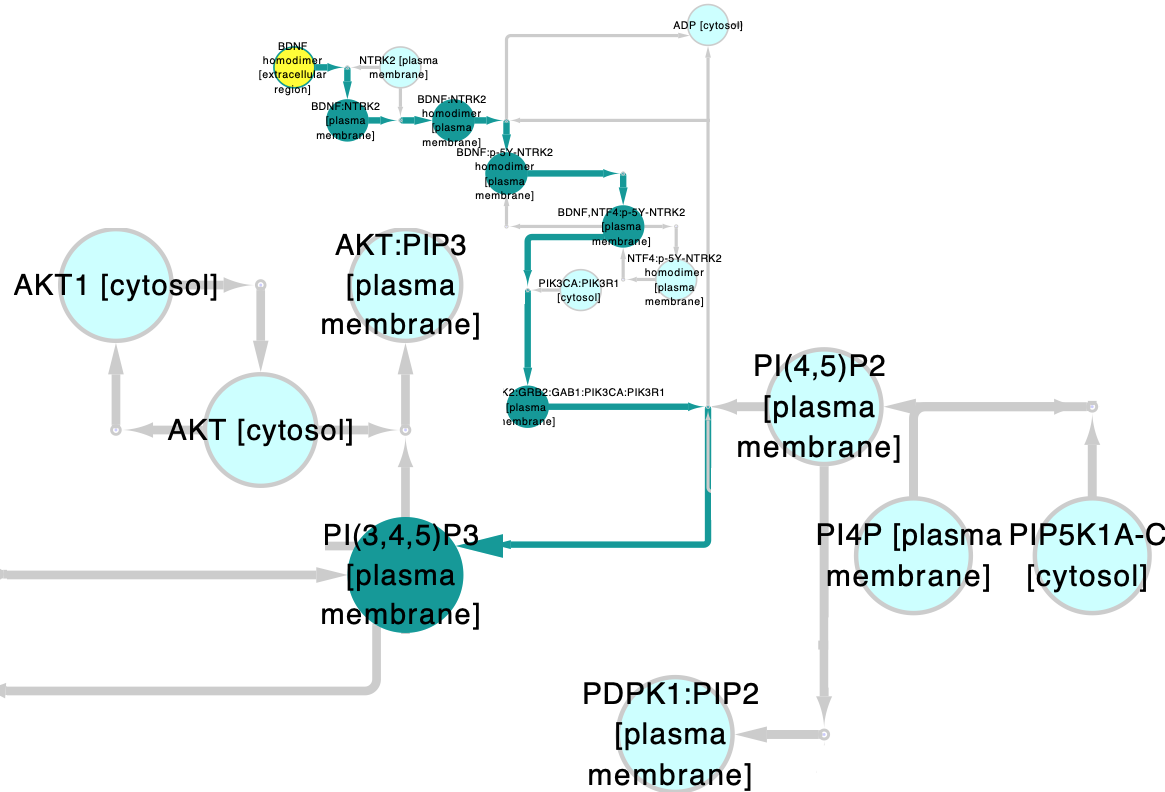
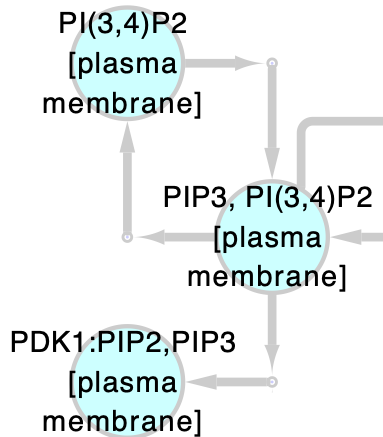
12 connections
13 species
5 pathways

Pathways in
minimal path

1. Signaling by NTRK2 (TRKB)
2. PIP3 activates AKT signaling
3. Costimulation by the CD28 family
4. Metabolism of nitric oxide: NOS3 activation and regulation
5. MTOR signaling

Studying BDNF connection to mTOR

- TCR signaling
- Fc epsilon receptor (FCERI) signaling



Minimal path

12 connections
13 species
5 pathways

Pathways in
minimal path

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Studying BDNF connection to mTOR

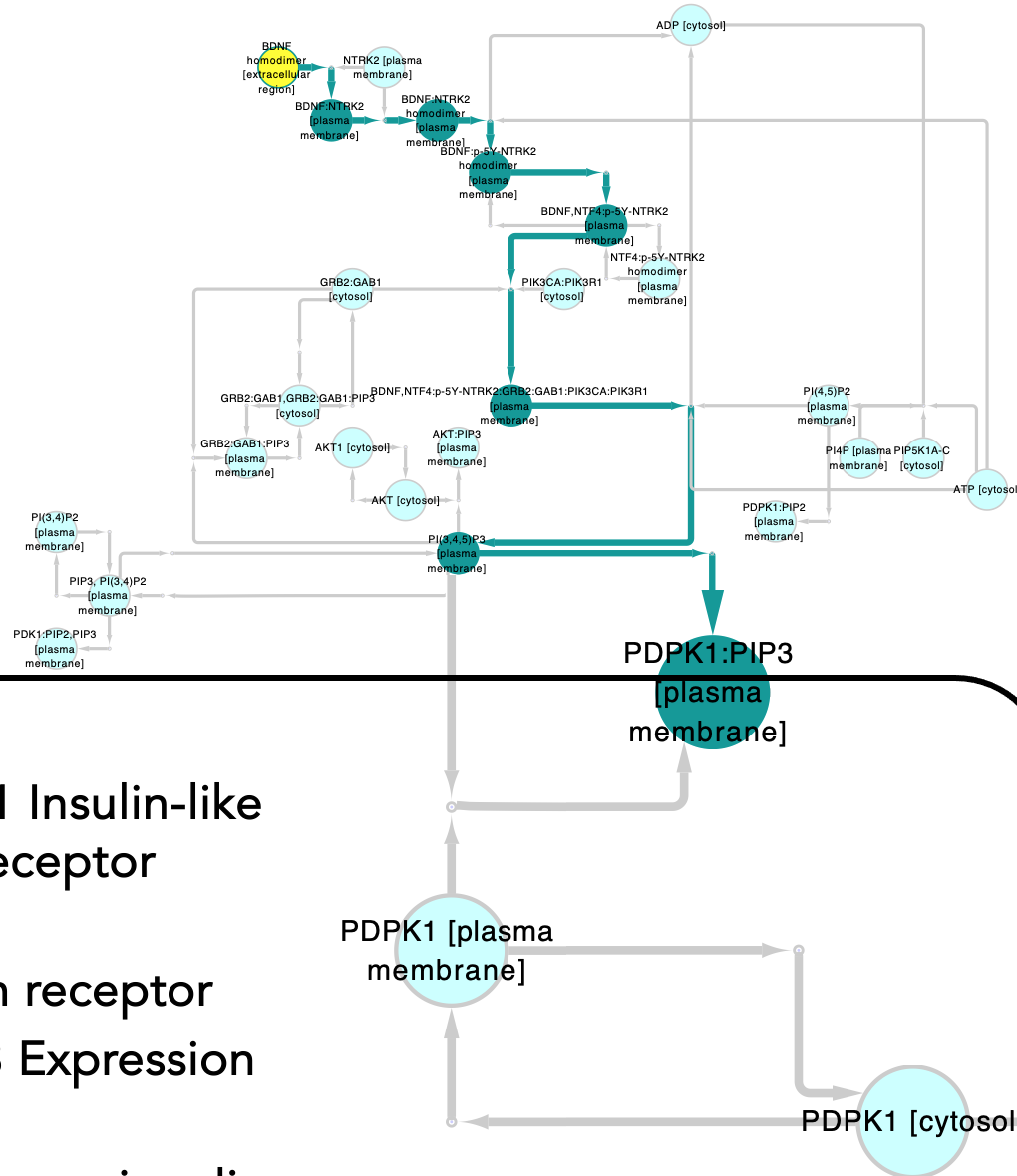
Minimal path

12 connections
13 species
5 pathways

Pathways in
minimal path

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3. Costimulation by the CD28 family
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5. MTOR signaling

- PI3K Cascade
- Signaling by Type 1 Insulin-like Growth Factor 1 Receptor (IGF1R)
- Signaling by Insulin receptor
- Regulation of TP53 Expression and Degradation
- Extra-nuclear estrogen signaling



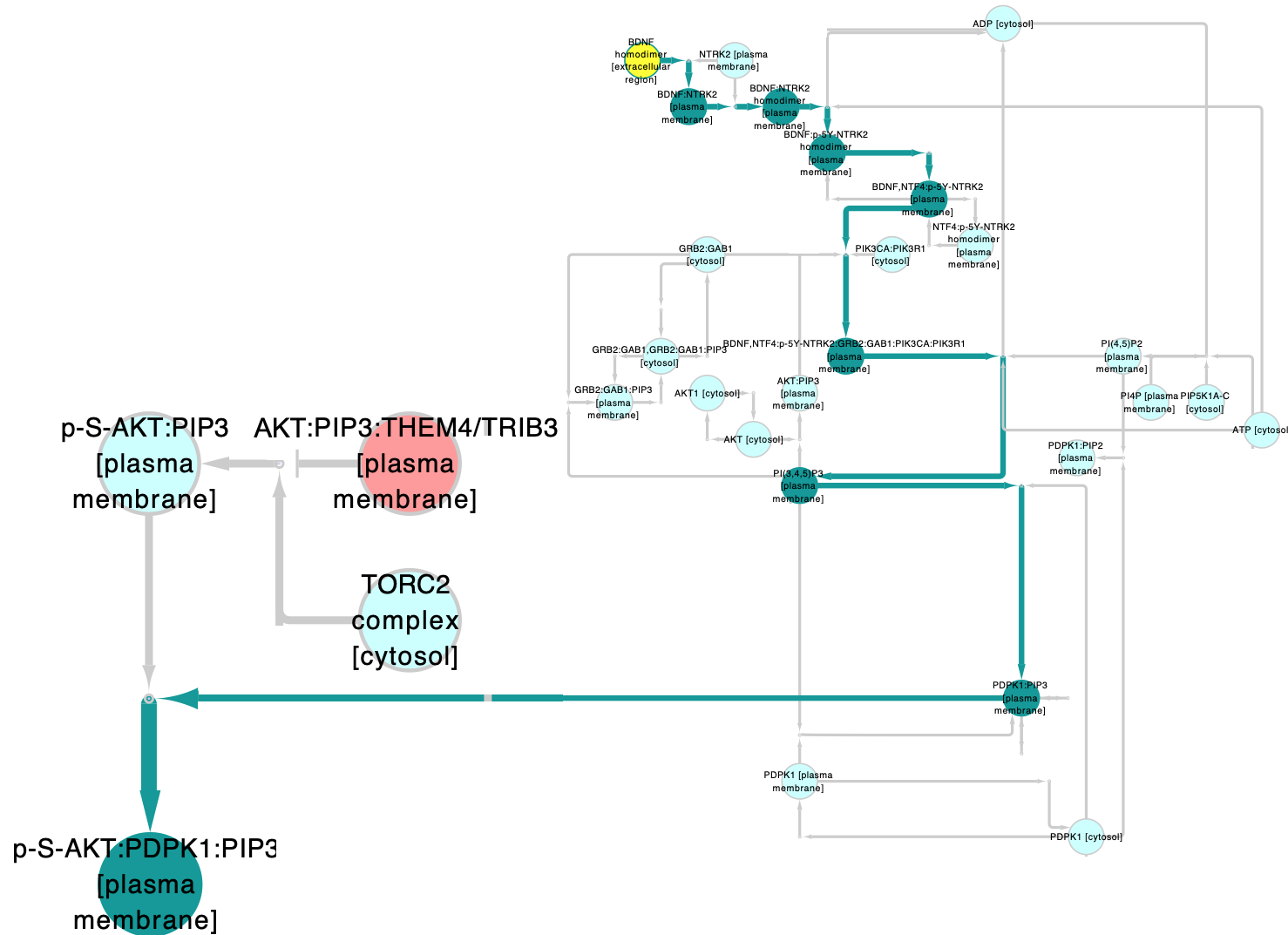
Studying BDNF connection to mTOR

Minimal path

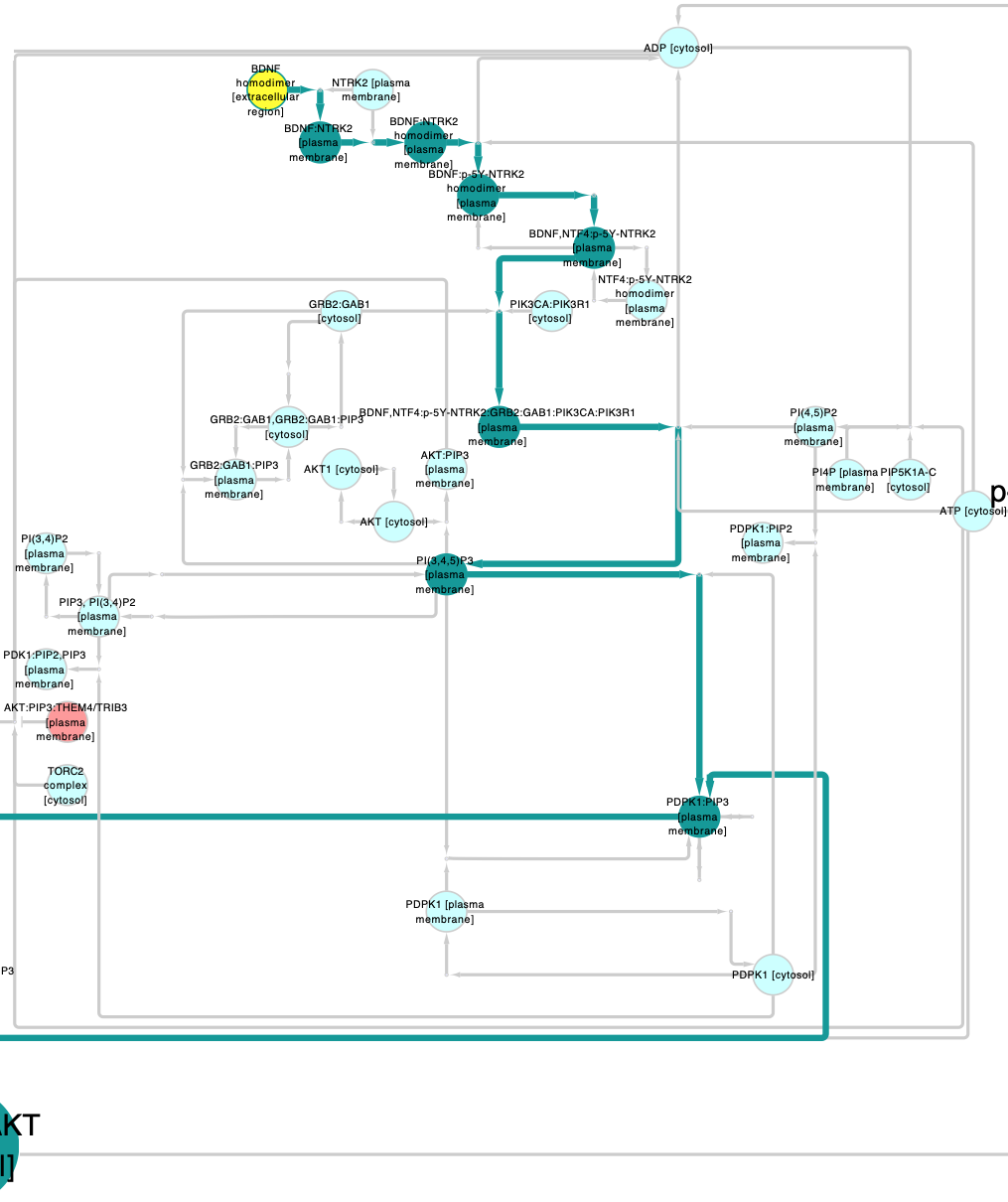
12 connections
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Pathways in minimal path

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Studying BDNF connection to mTOR



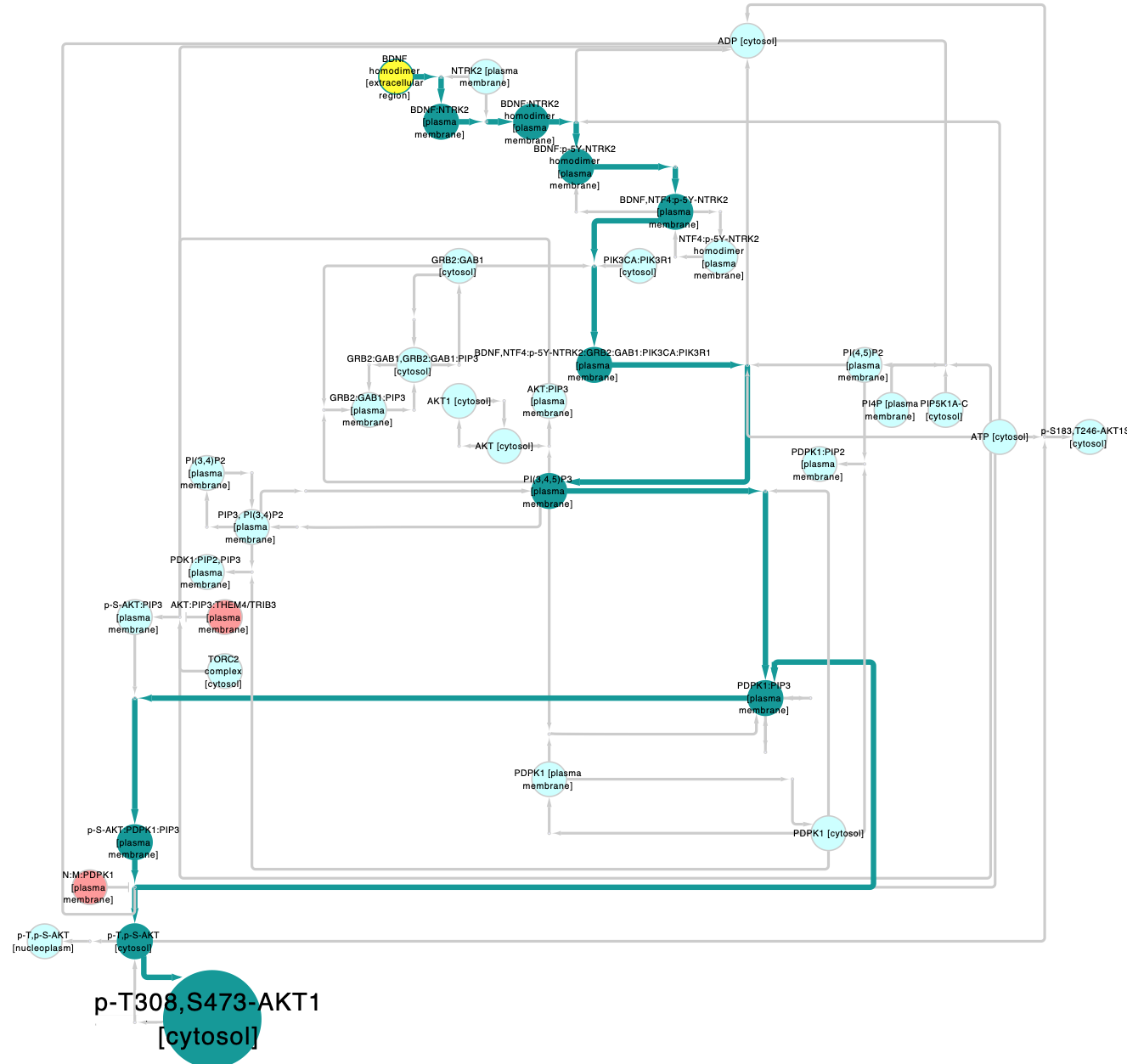
Minimal path

12 connections
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Studying BDNF connection to mTOR



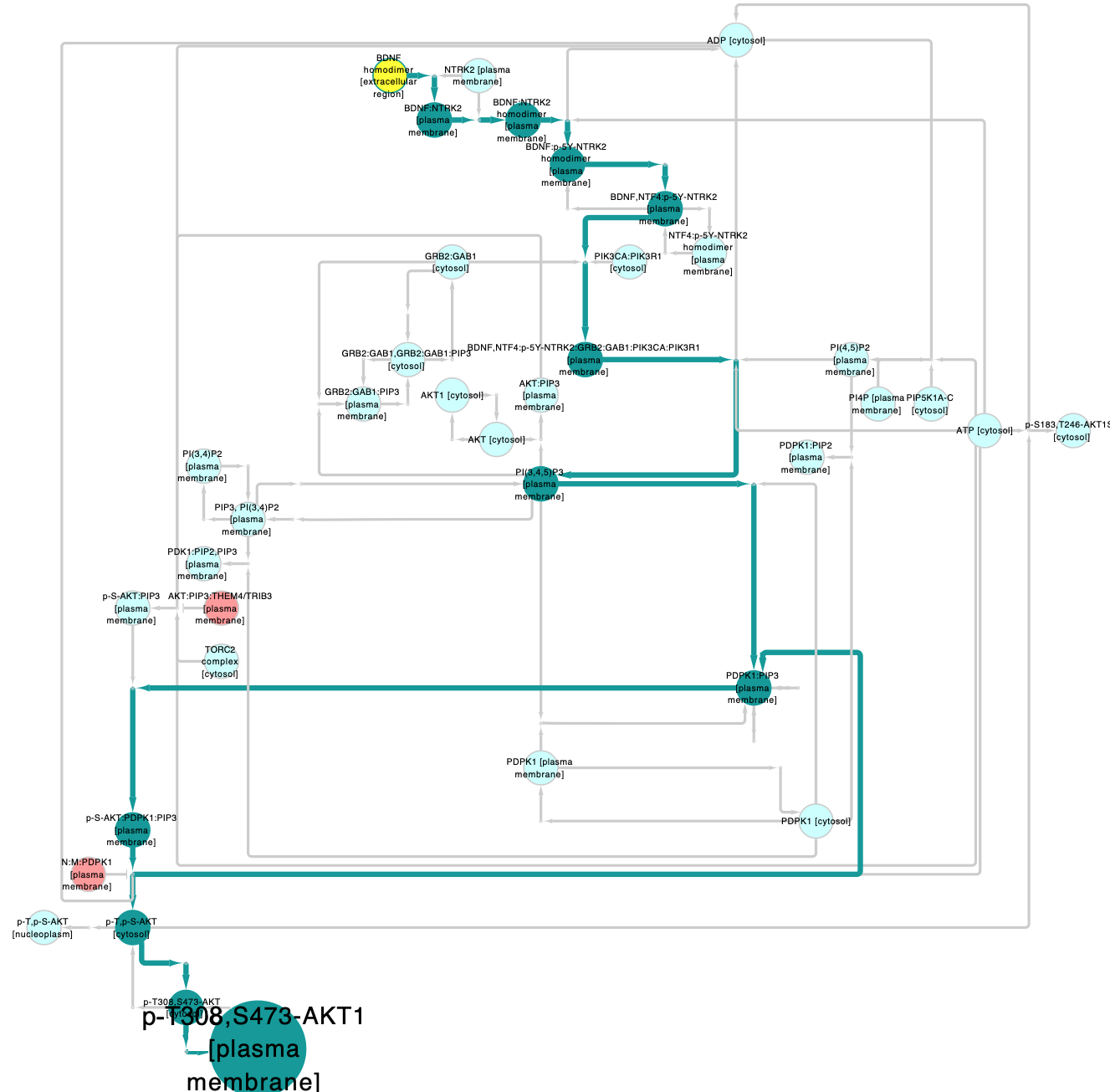
Minimal path

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Pathways in
minimal path

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Studying BDNF connection to mTOR



Minimal path

12 connections
13 species
5 pathways

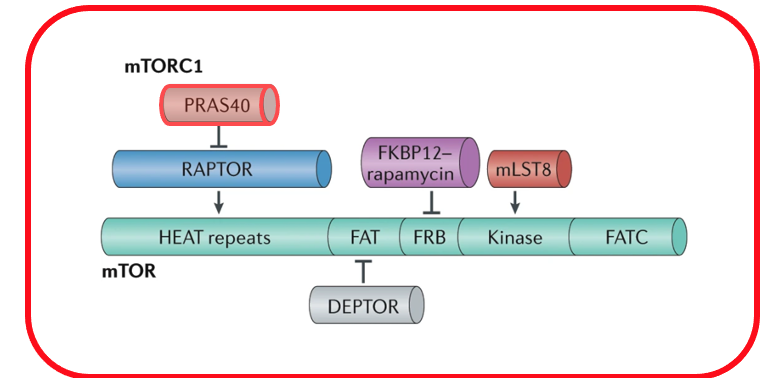
Pathways in
minimal path

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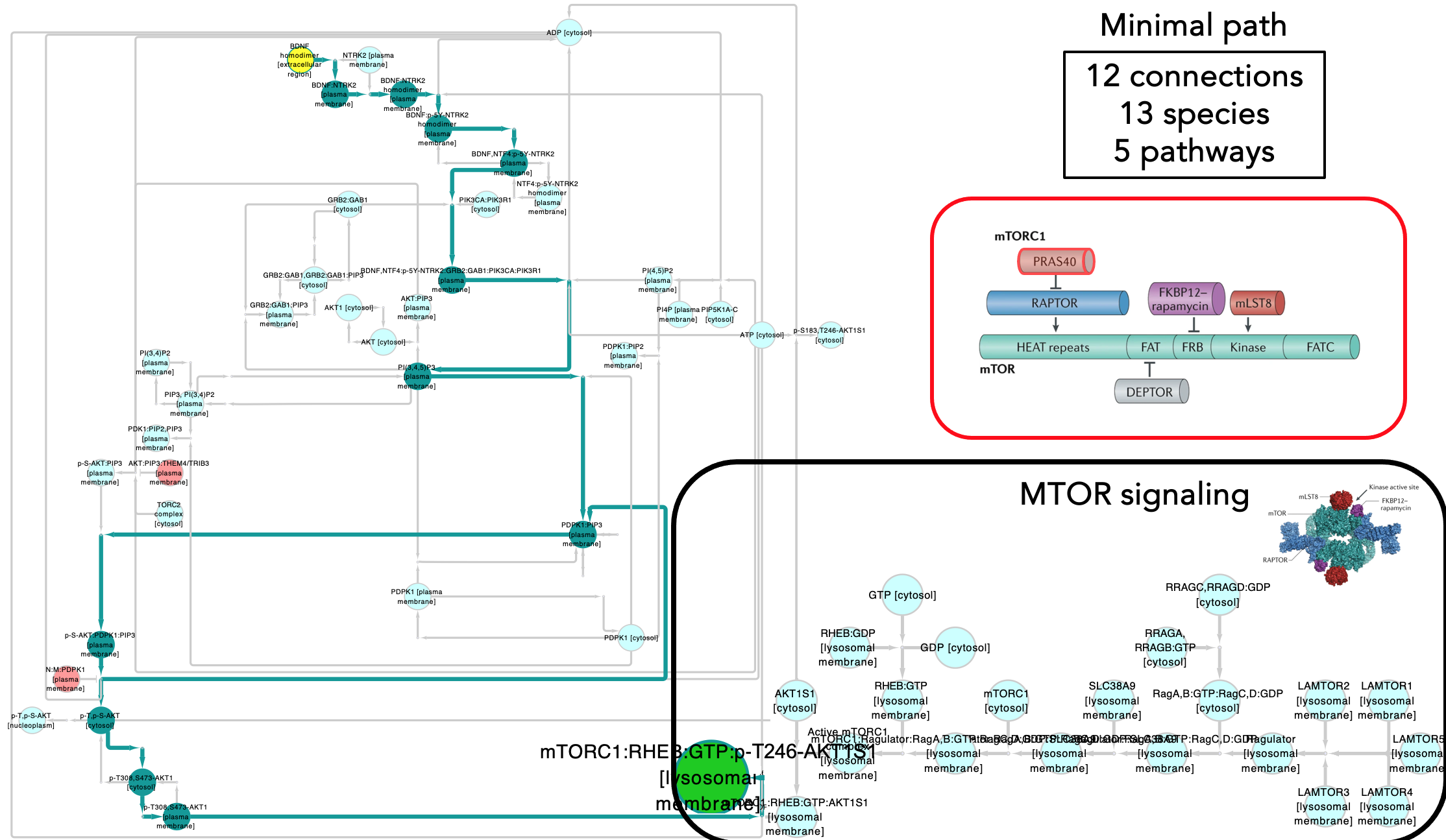
Studying BDNF connection to mTOR

Minimal path

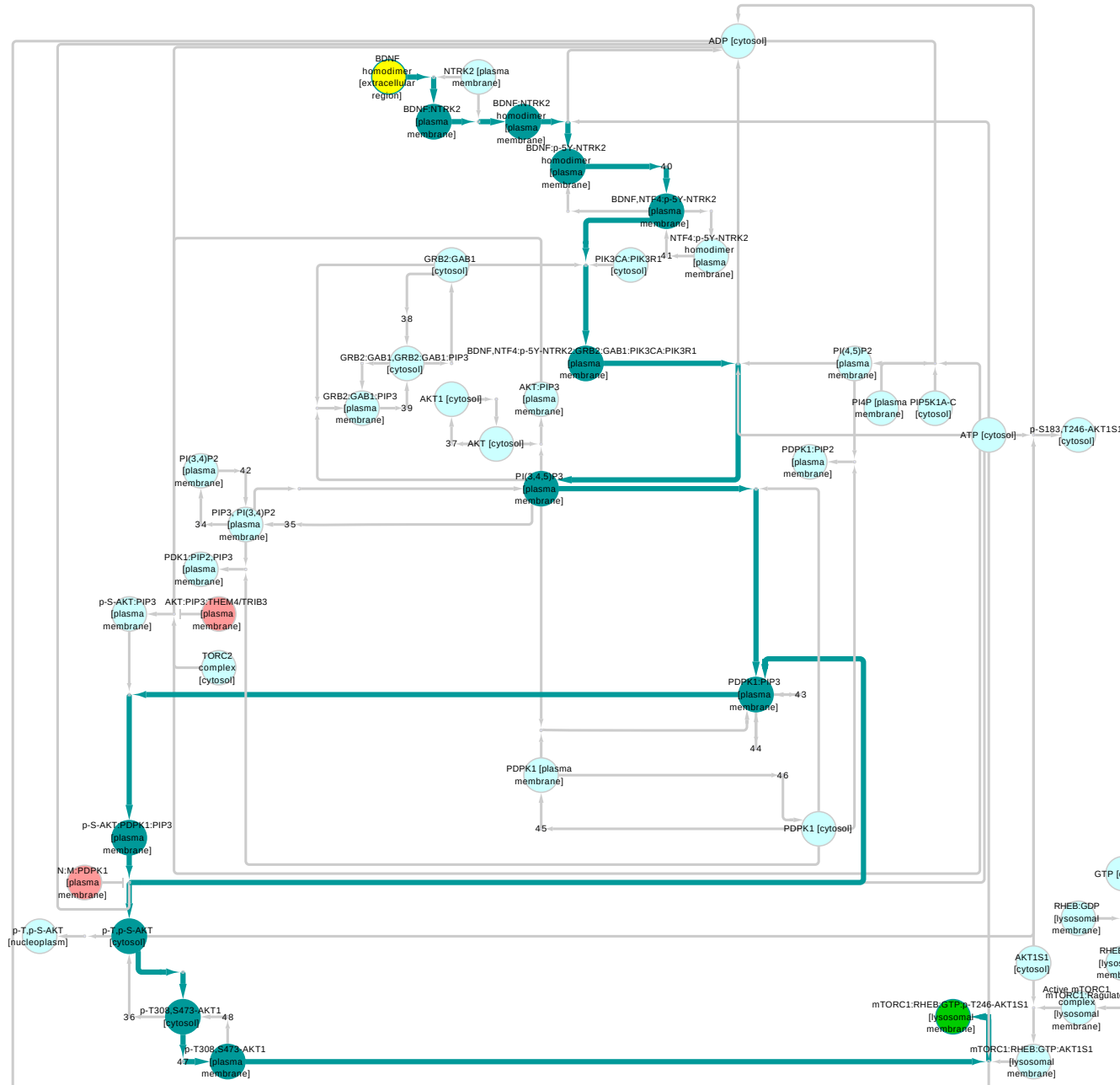
12 connections
13 species
5 pathways



MTOR signaling



Studying BDNF connection to mTOR



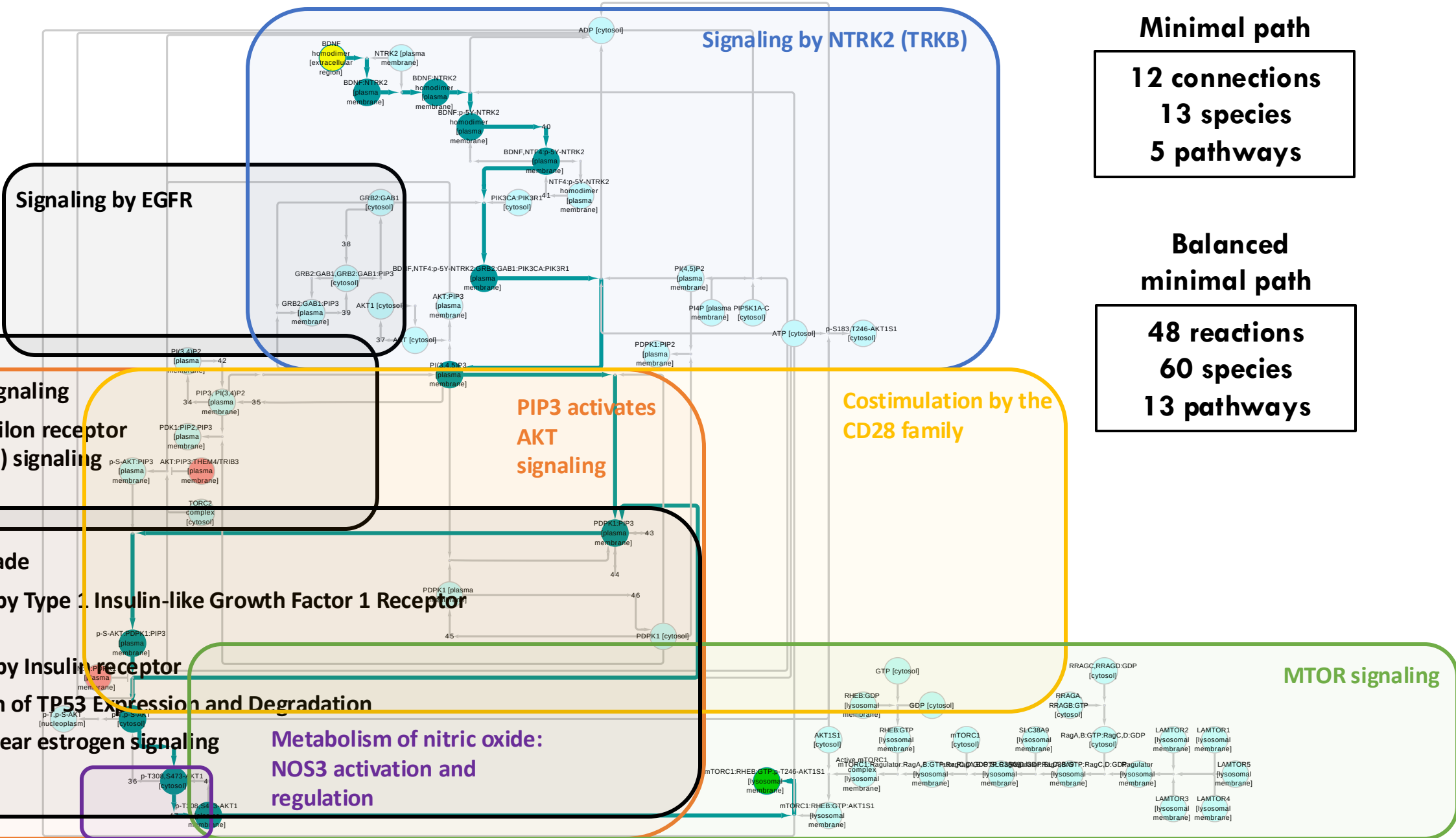
Minimal path

12 connections
13 species
5 pathways

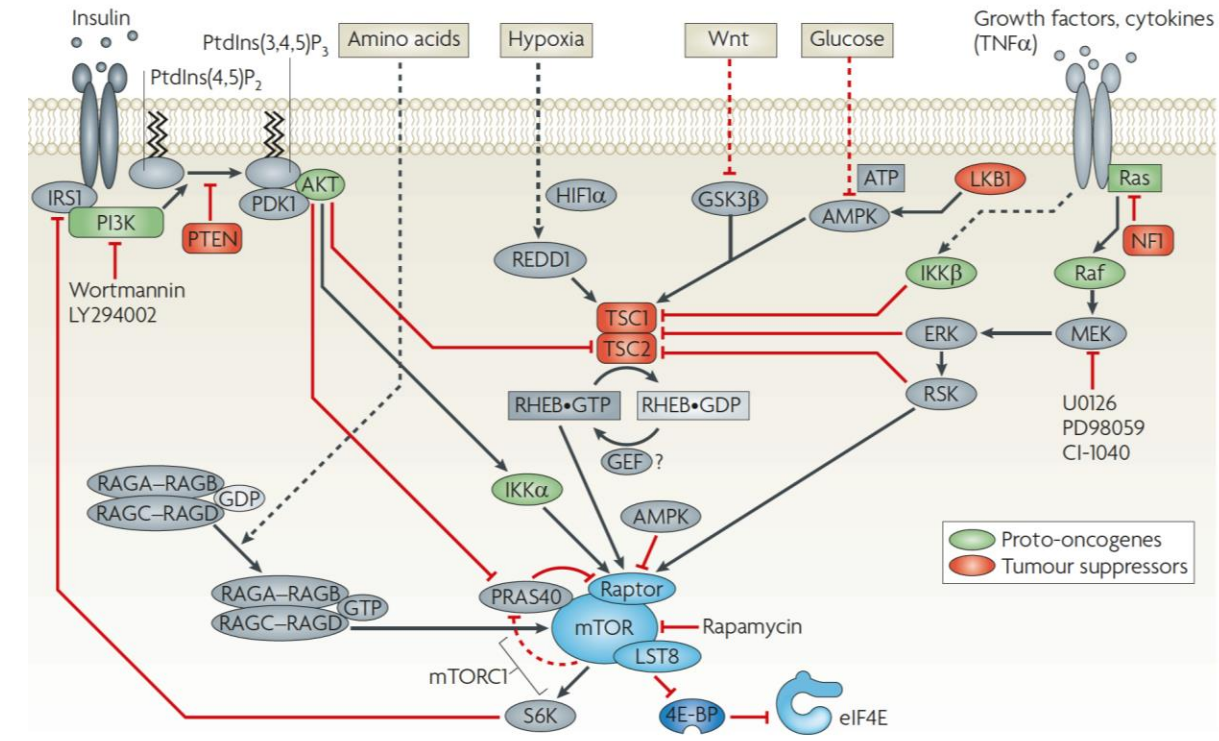
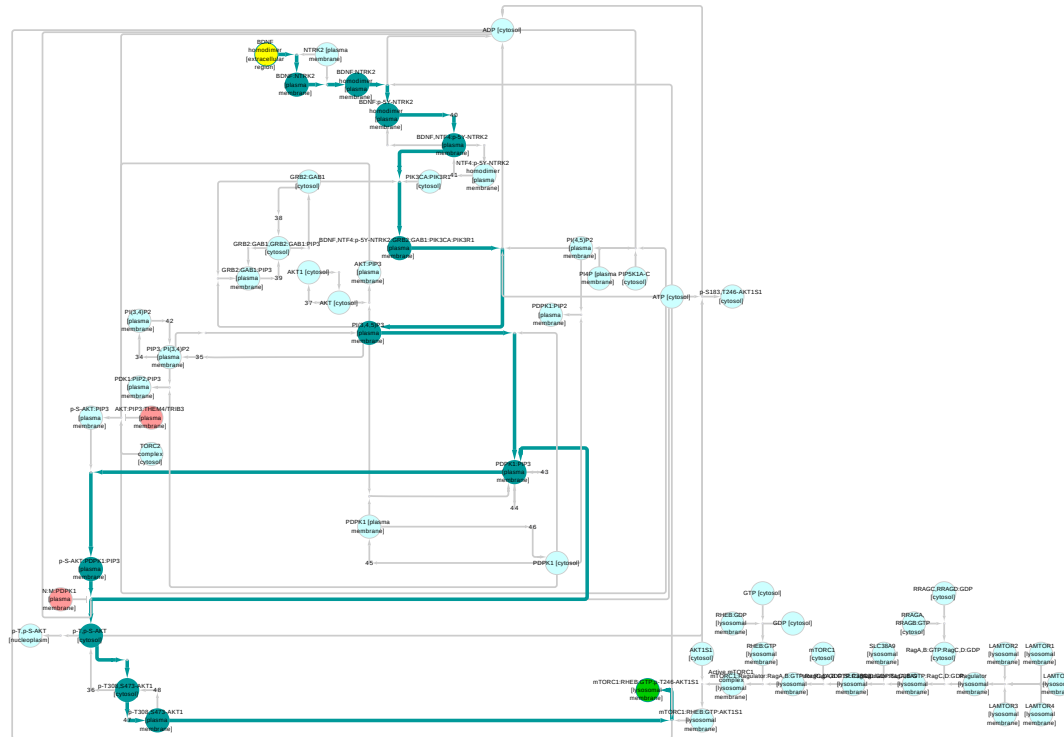
Balanced minimal path

48 reactions
60 species
13 pathways

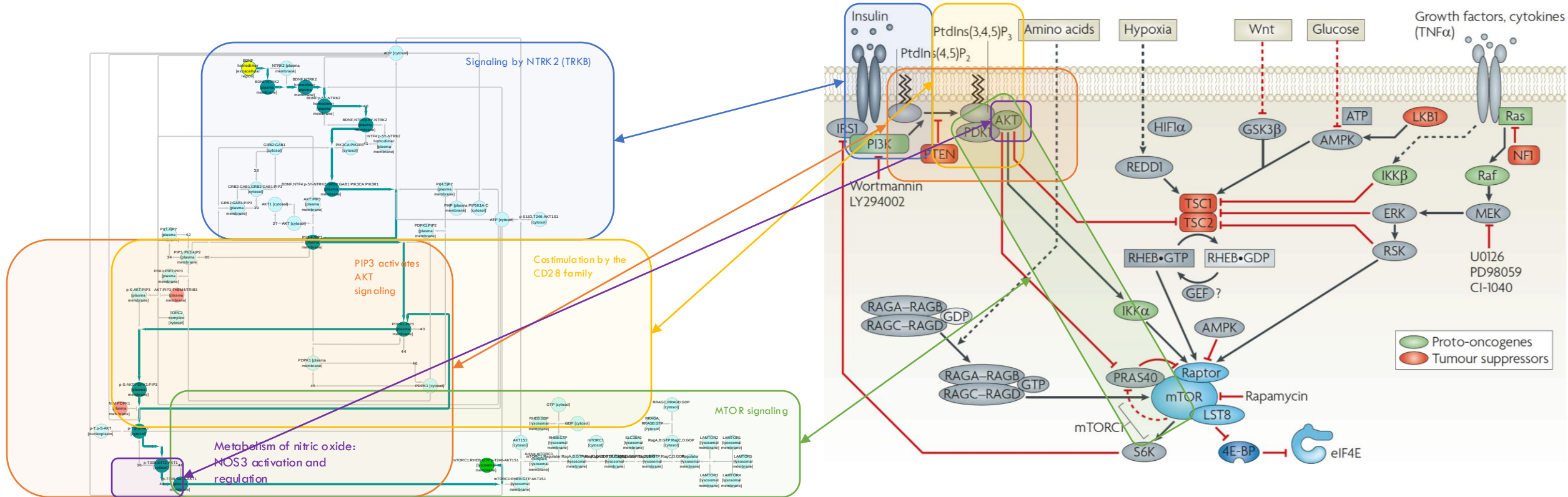
Studying BDNF connection to mTOR



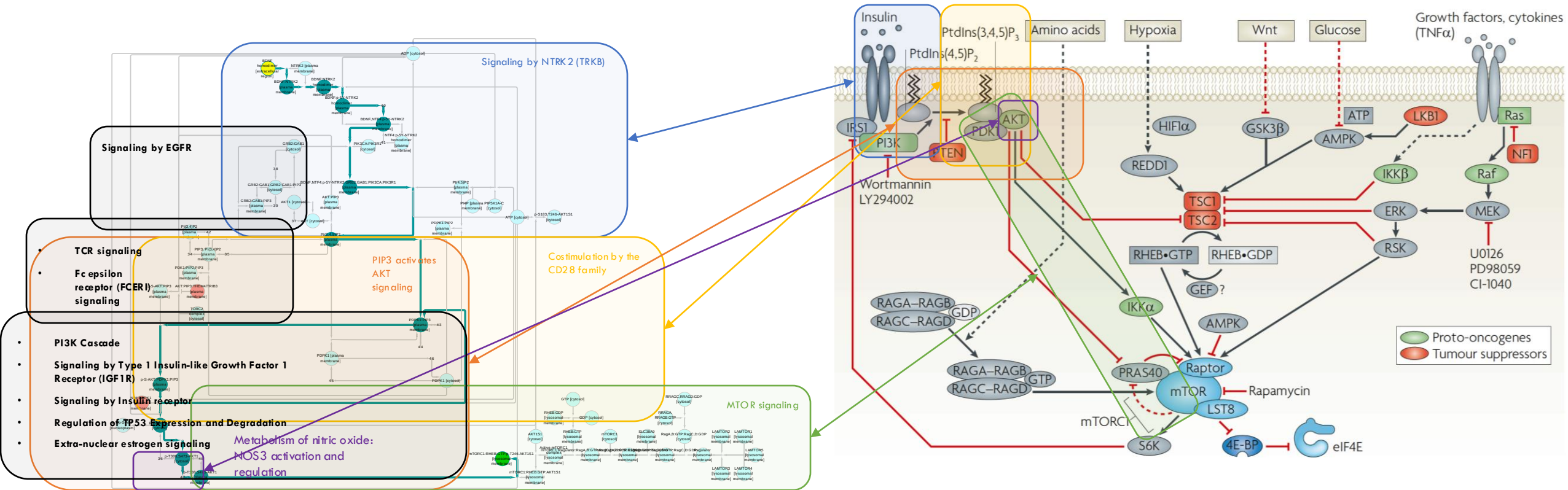
Studying BDNF connection to mTOR



Studying BDNF connection to mTOR



Studying BDNF connection to mTOR



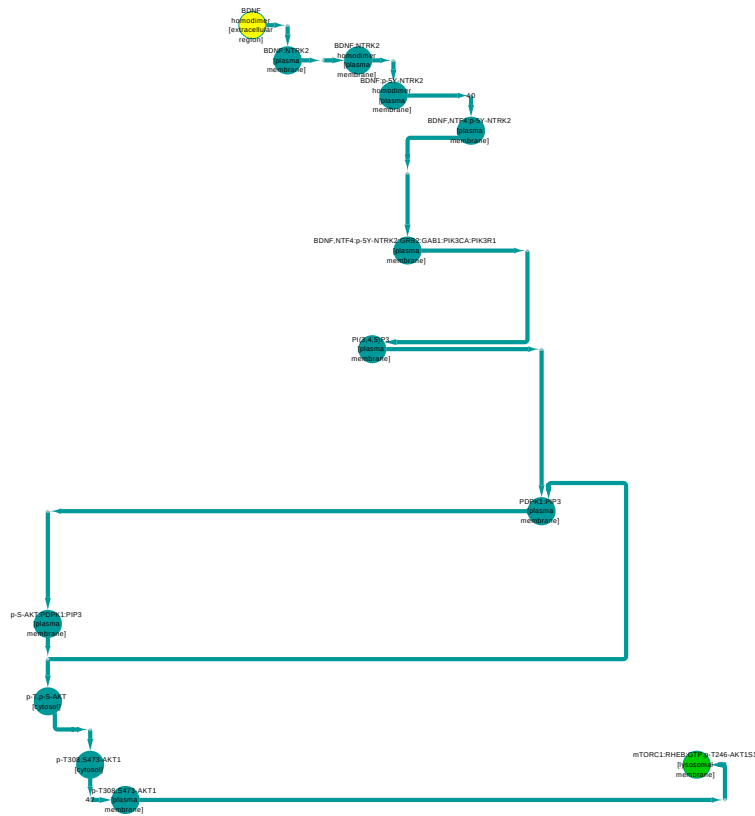
Alternative BDNF connections to mTOR

Minimal path

12 connections

13 species

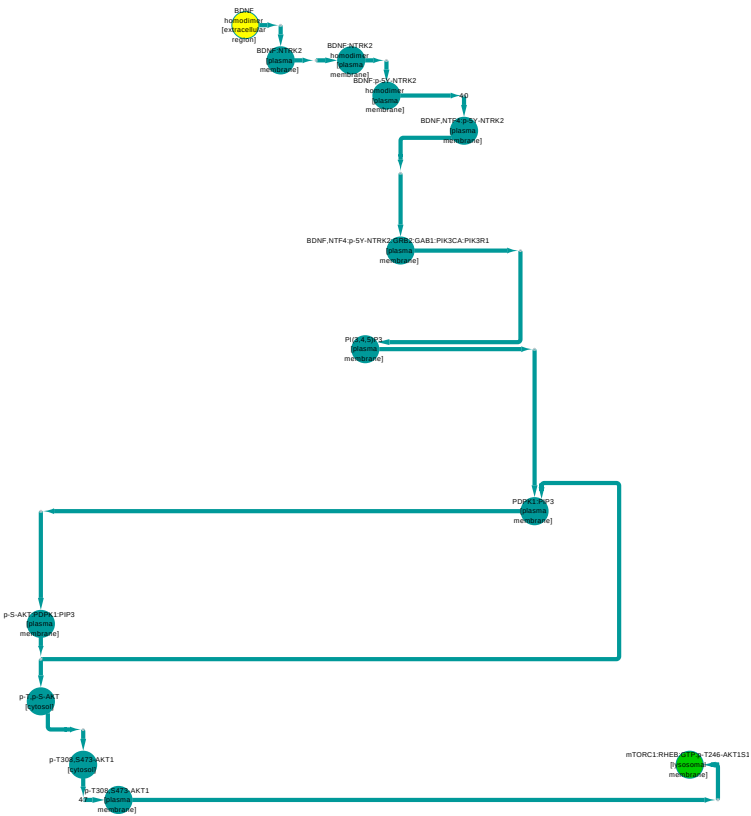
5 pathways



Alternative BDNF connections to mTOR

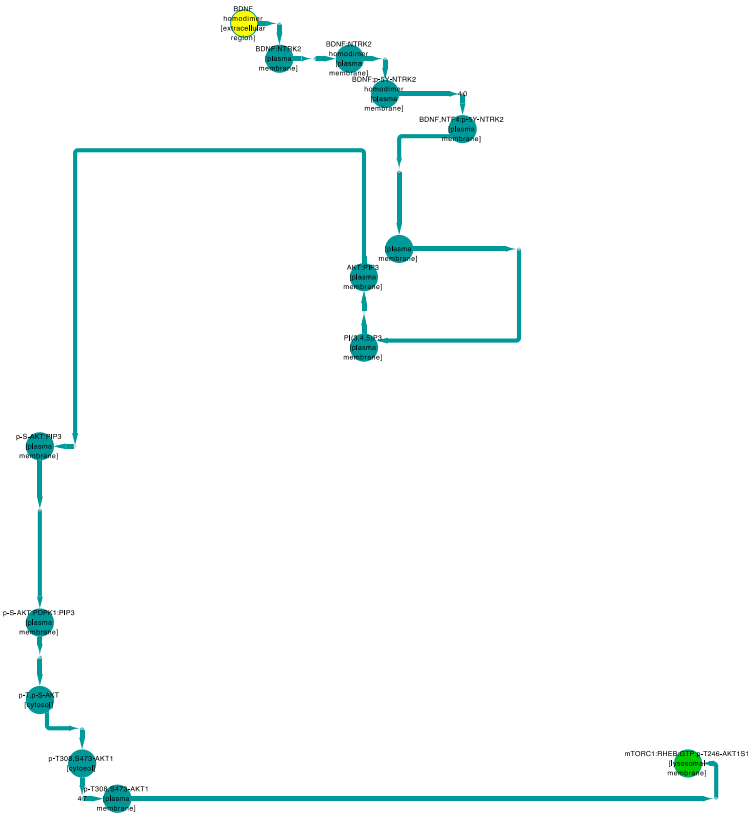
Minimal path

12 connections
13 species
5 pathways



Minimal path+1

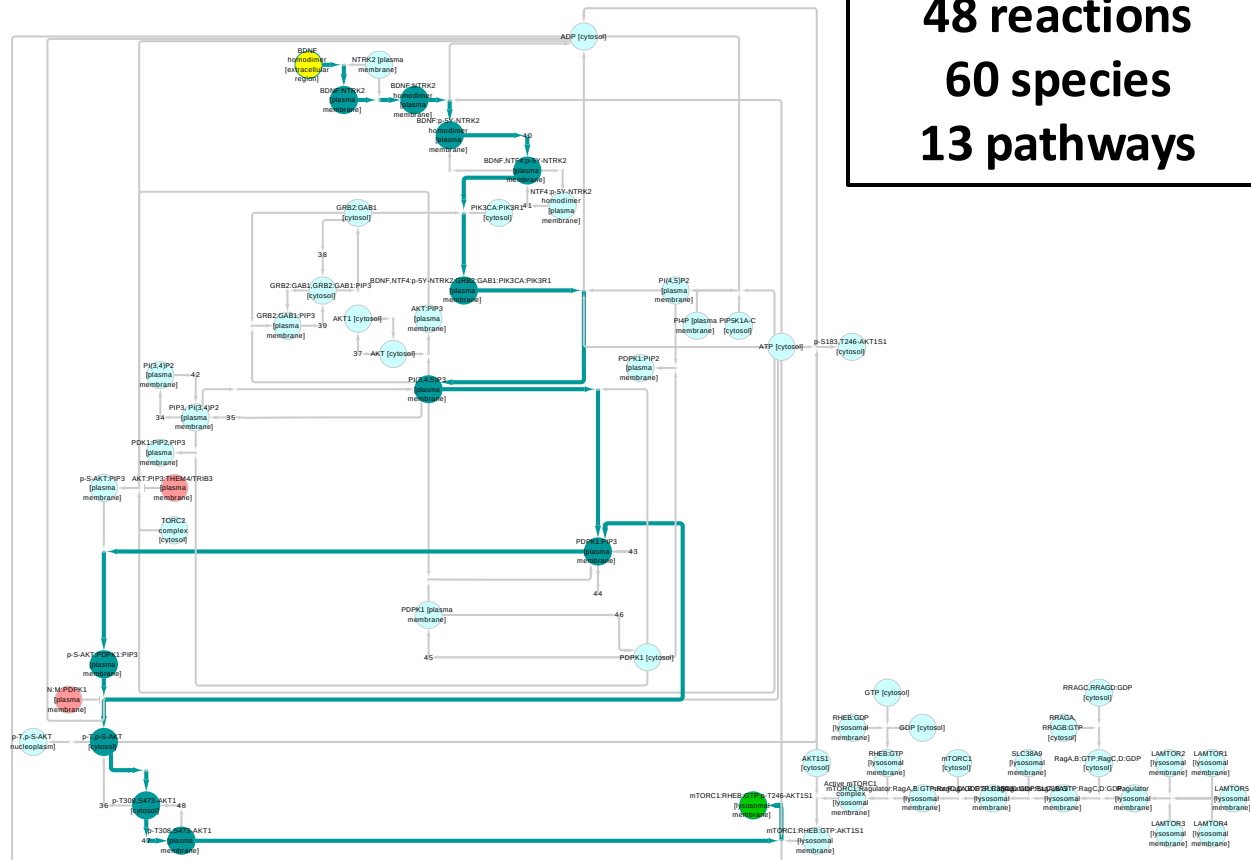
13 connections
14 species
5 pathways



Alternative BDNF connections to mTOR

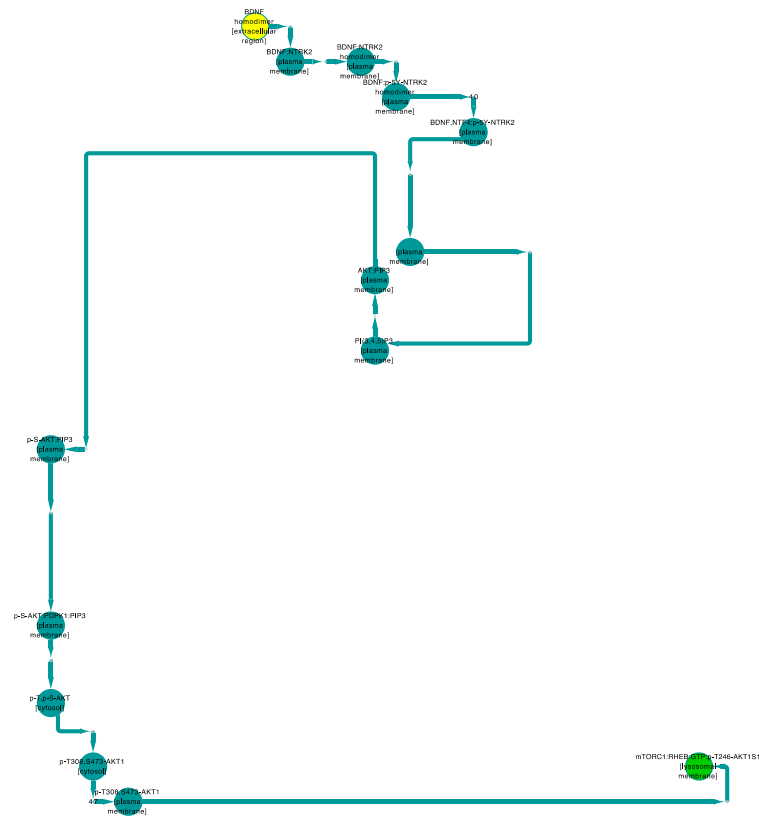
Minimal path

48 reactions
60 species
13 pathways



Minimal path+1

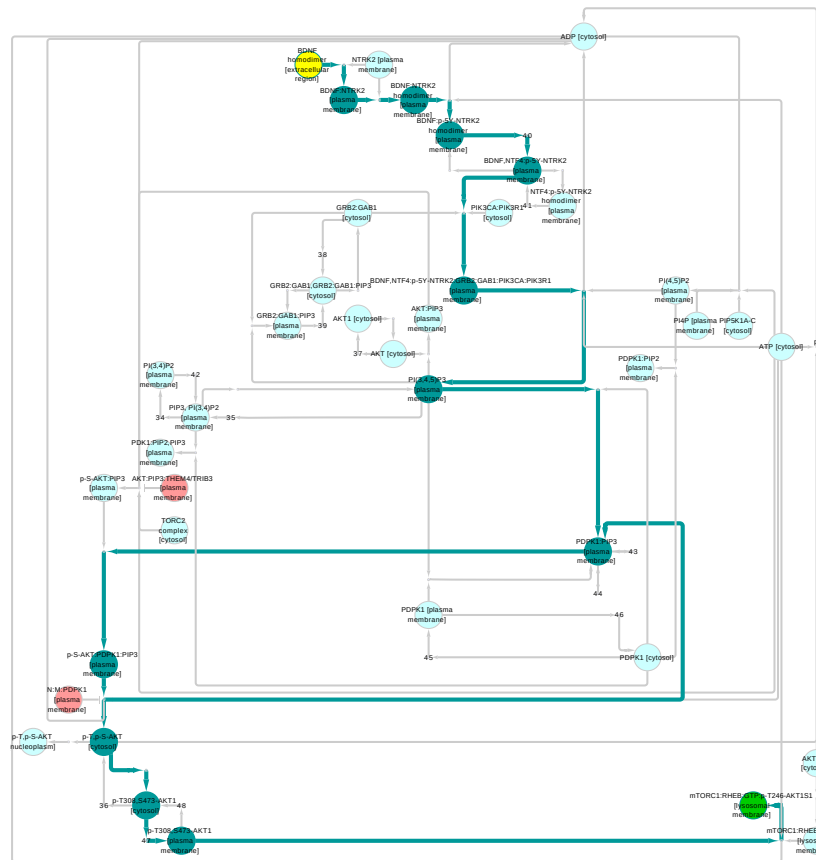
13 connections
14 species
5 pathways



Alternative BDNF connections to mTOR

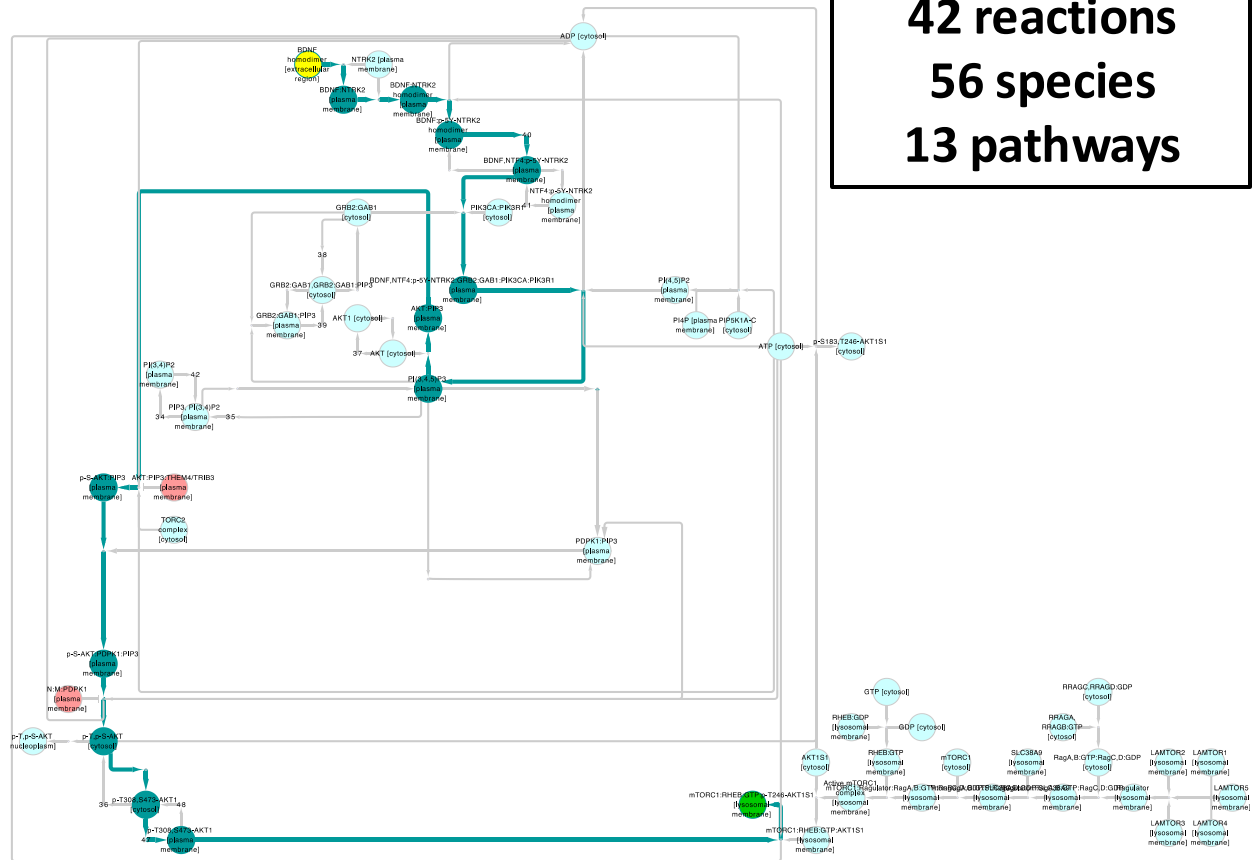
Minimal path

48 reactions
60 species
13 pathways



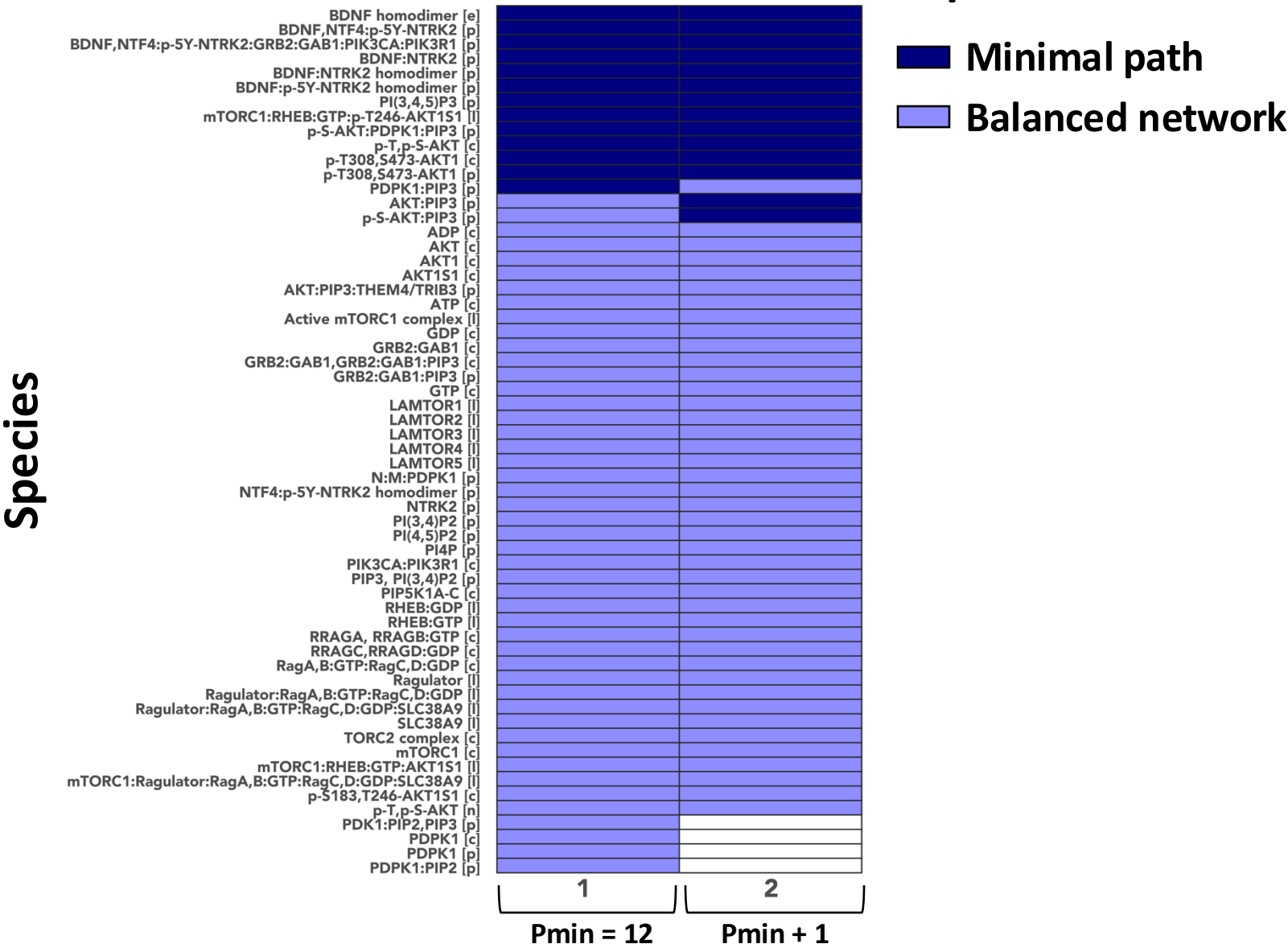
Minimal path+1

42 reactions
56 species
13 pathways



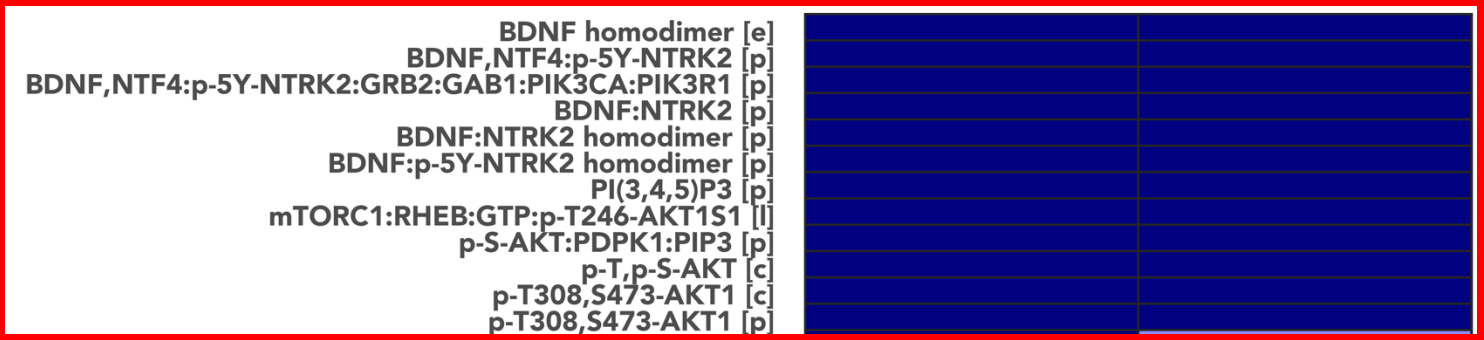
Alternative BDNF connections to mTOR

BDNF to mTORC1:RHEB:GTP:p-T246-AKT1S1

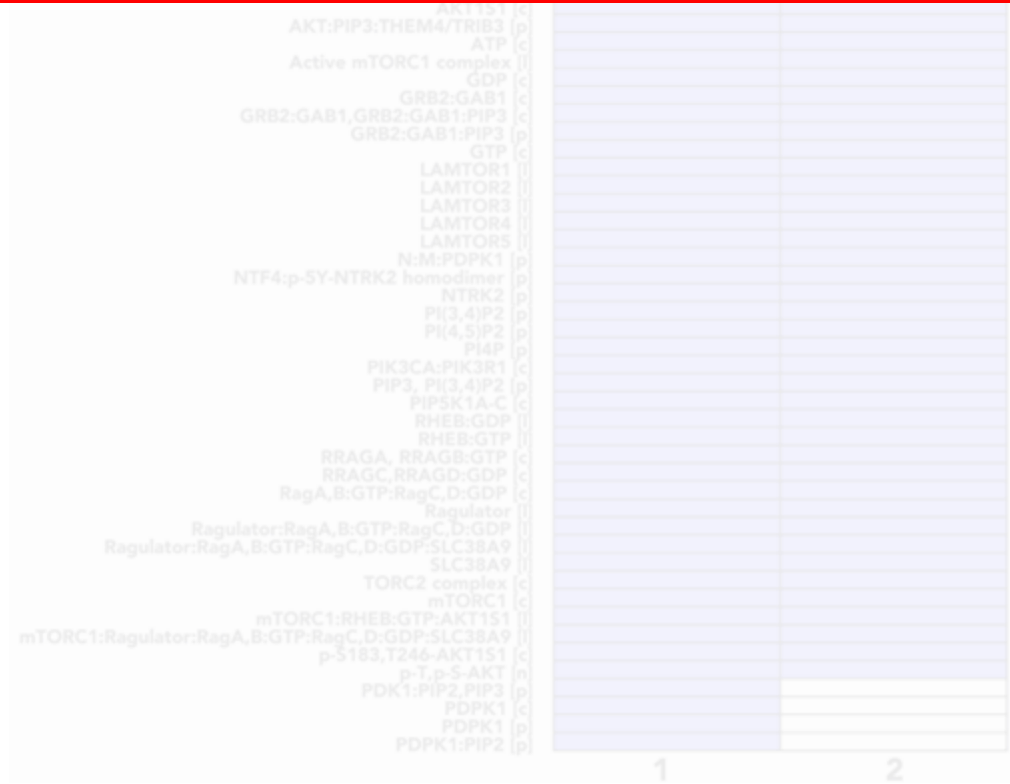


Alternative BDNF connections to mTOR

BDNF to mTORC1:RHEB:GTP:p-T246-AKT1S1



Minimal path
Balanced network



Alternative BDNF connections to mTOR

BDNF to mTORC1:RHEB:GTP:p-T246-AKT1S1



Alternative BDNF connections to mTOR

BDNF to mTORC1:RHEB:GTP:p-T246-AKT1S1

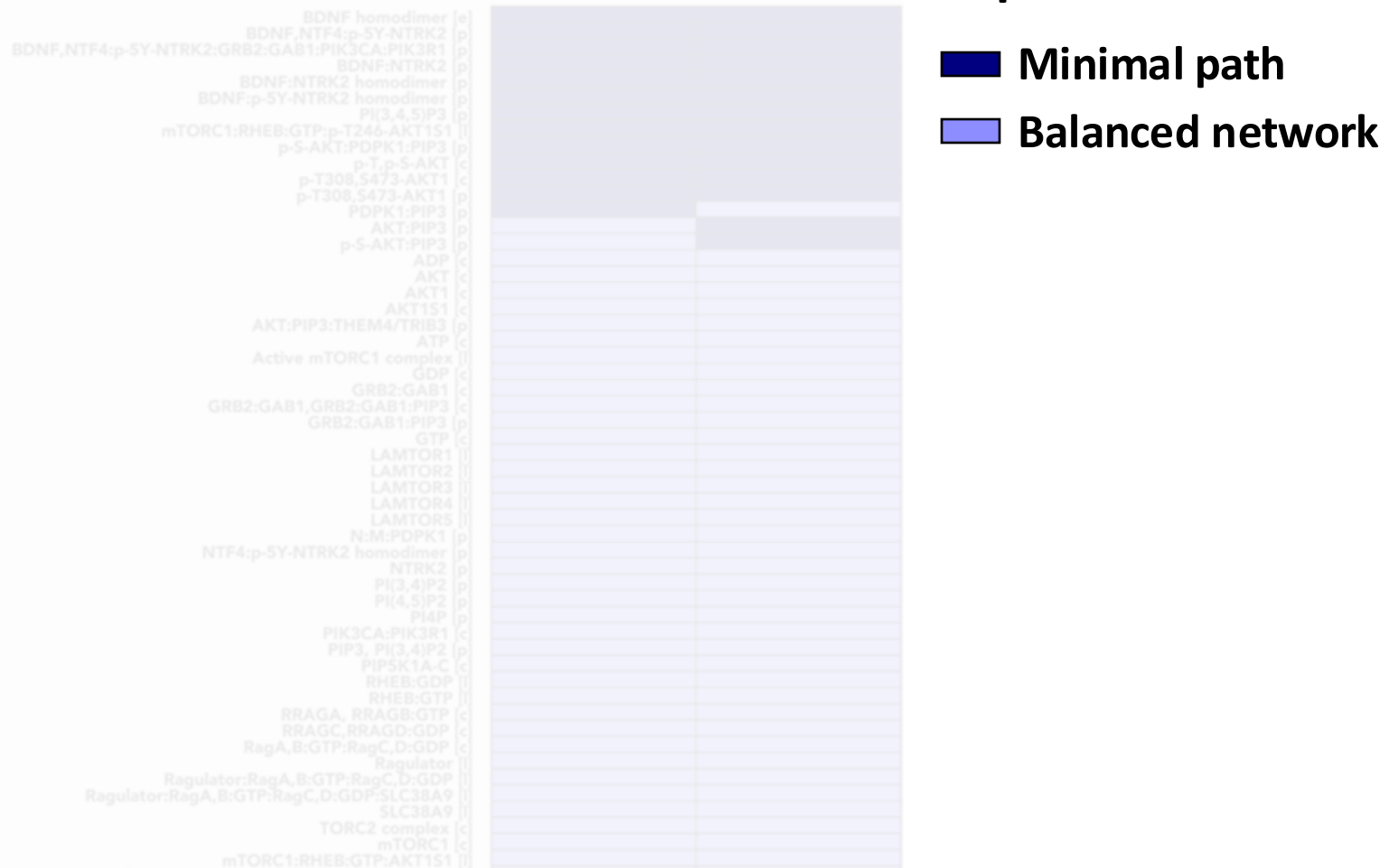
ADP	[c]		
AKT	[c]		
AKT1	[c]		
AKT1S1	[c]		
AKT:PI(3,4,5)P3:RHOA	[p]		
ATP	[c]		
Active mTORC1 complex	[l]		
GDP	[c]		
GRB2:GAB1	[c]		
GRB2:GAB1,GRB2:GAB1:PI(3,4,5)P3	[c]		
GRB2:GAB1:PI(3,4,5)P3	[p]		
GTP	[c]		
LAMTOR1	[l]		
LAMTOR2	[l]		
LAMTOR3	[l]		
LAMTOR4	[l]		
LAMTOR5	[l]		
N:M:PDK1	[p]		
NTF4:p-5Y-NTRK2 homodimer	[p]		
NTRK2	[p]		
PI(3,4,5)P2	[p]		
PI(4,5)P2	[p]		
PI4P	[p]		
PIK3CA:PIK3R1	[c]		
PI(3,4,5)P3, PI(3,4,5)P2	[p]		
PIP5K1A-C	[c]		
RHEB:GDP	[l]		
RHEB:GTP	[l]		
RRAGA, RRAGB:GTP	[c]		
RRAGC,RRAGD:GDP	[c]		
RagA,B:GTP:RagC,D:GDP	[c]		
Ragulator	[l]		
Ragulator:RagA,B:GTP:RagC,D:GDP	[l]		
Ragulator:RagA,B:GTP:RagC,D:GDP:SLC38A9	[l]		
SLC38A9	[l]		
TORC2 complex	[c]		
mTORC1	[c]		
mTORC1:RHEB:GTP:AKT1S1	[l]		
mTORC1:Ragulator:RagA,B:GTP:RagC,D:GDP:SLC38A9	[l]		
p-S183,T246-AKT1S1	[c]		
p-T,p-S-AKT	[n]		

■ Minimal path

■ Balanced network

Alternative BDNF connections to mTOR

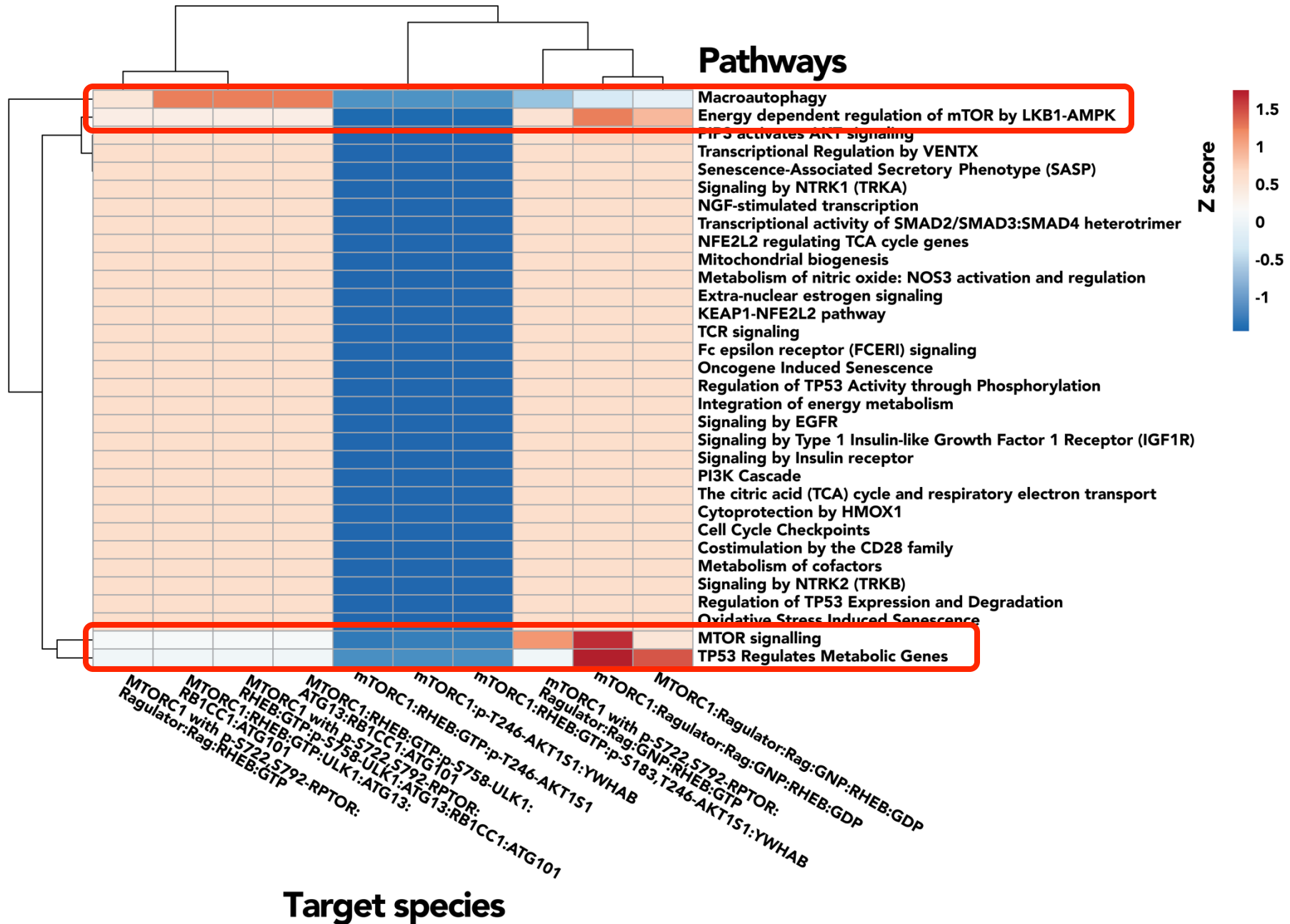
BDNF to mTORC1:RHEB:GTP:p-T246-AKT1S1



PDK1:PIP2,PIP3	[p]		
PDPK1	[c]		
PDPK1	[p]		
PDPK1:PIP2	[p]		

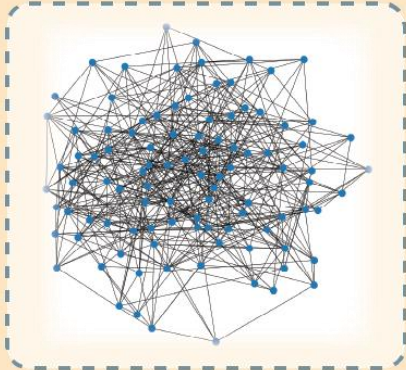
[illegible]

Target species

[illegible]

Study deregulation in signaling pathways

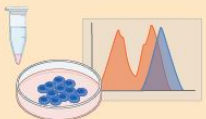
SIGMA Method



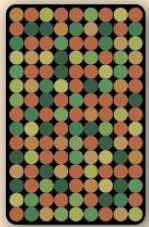
reactome

Signaling network

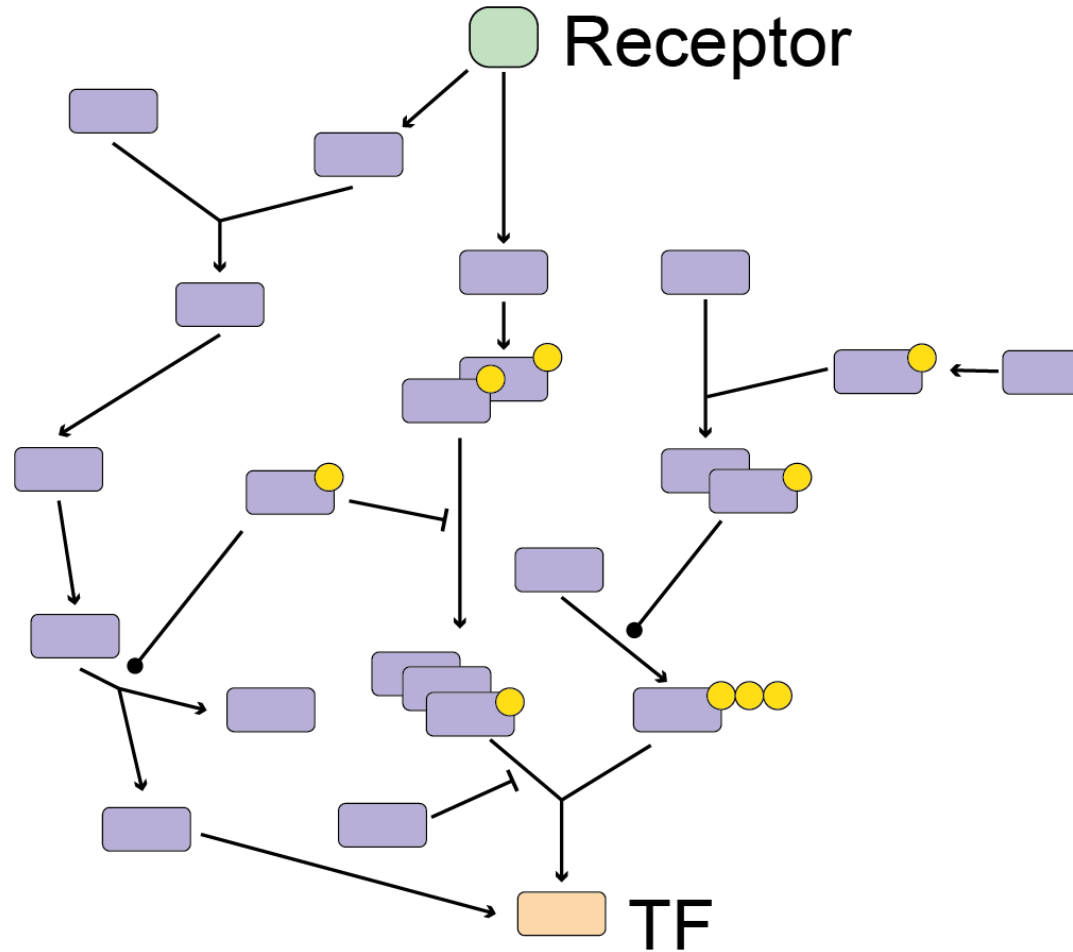
omics data



(sc)RNA-seq data



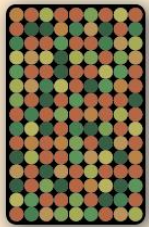
phosphoproteomics



SIGMA Method



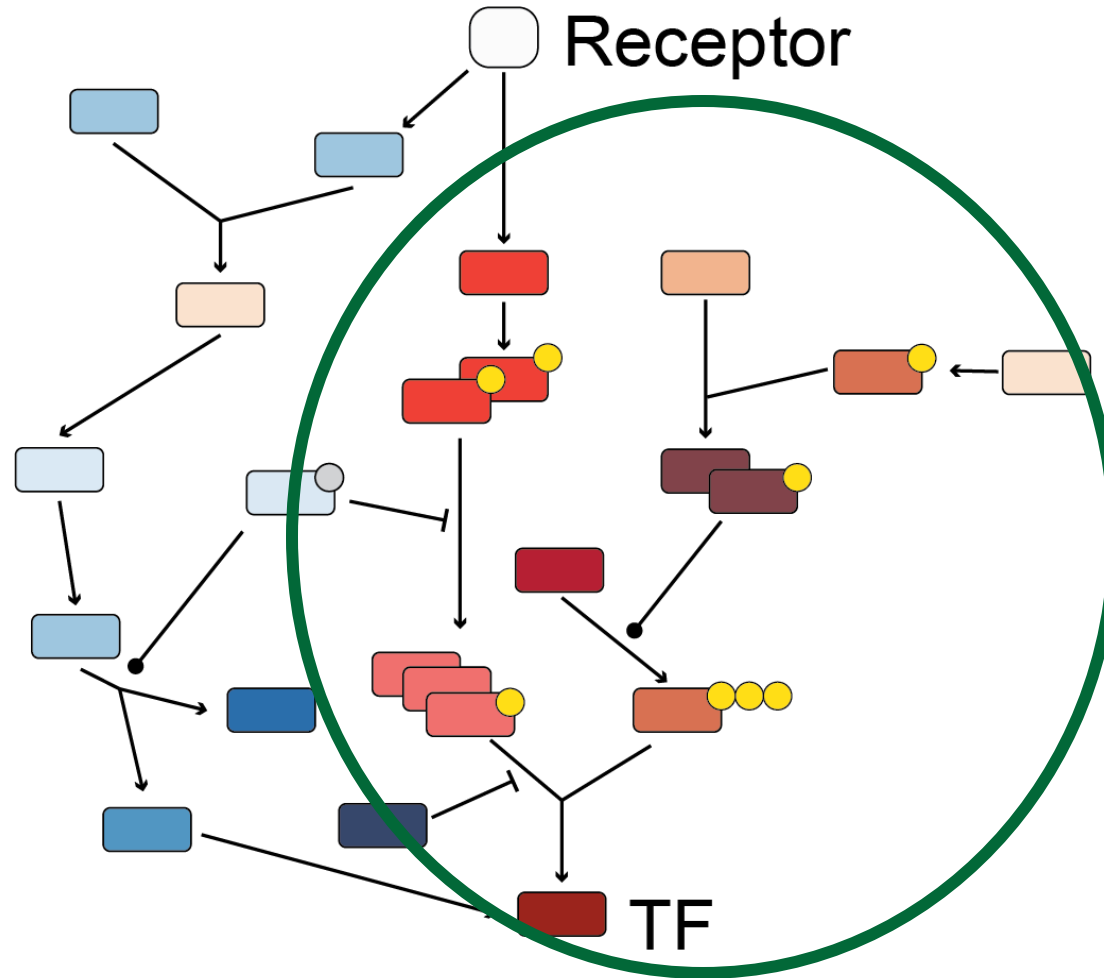
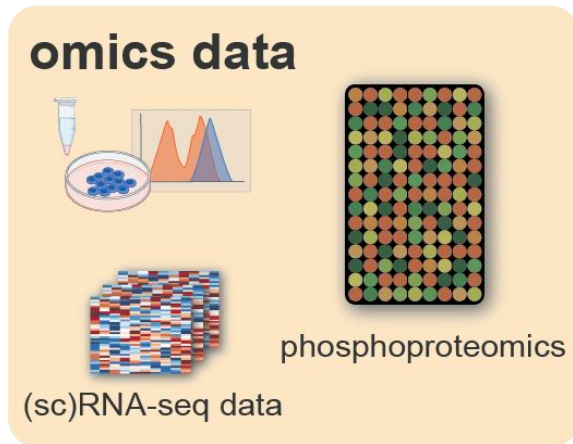
omics data



(sc)RNA-seq data

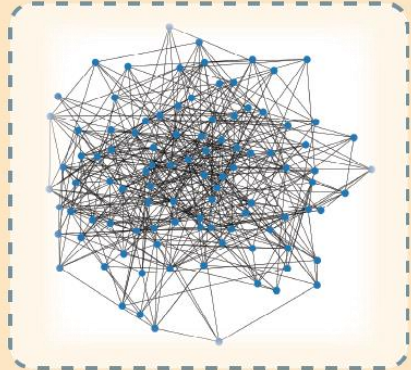
[illegible]

SIGMA Method



Study deregulation in signaling pathways

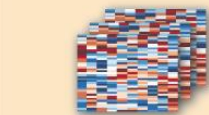
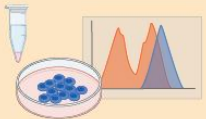
SIGMA Method



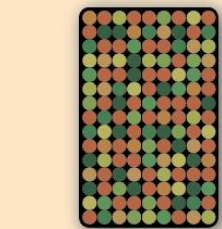
reactome

Signaling network

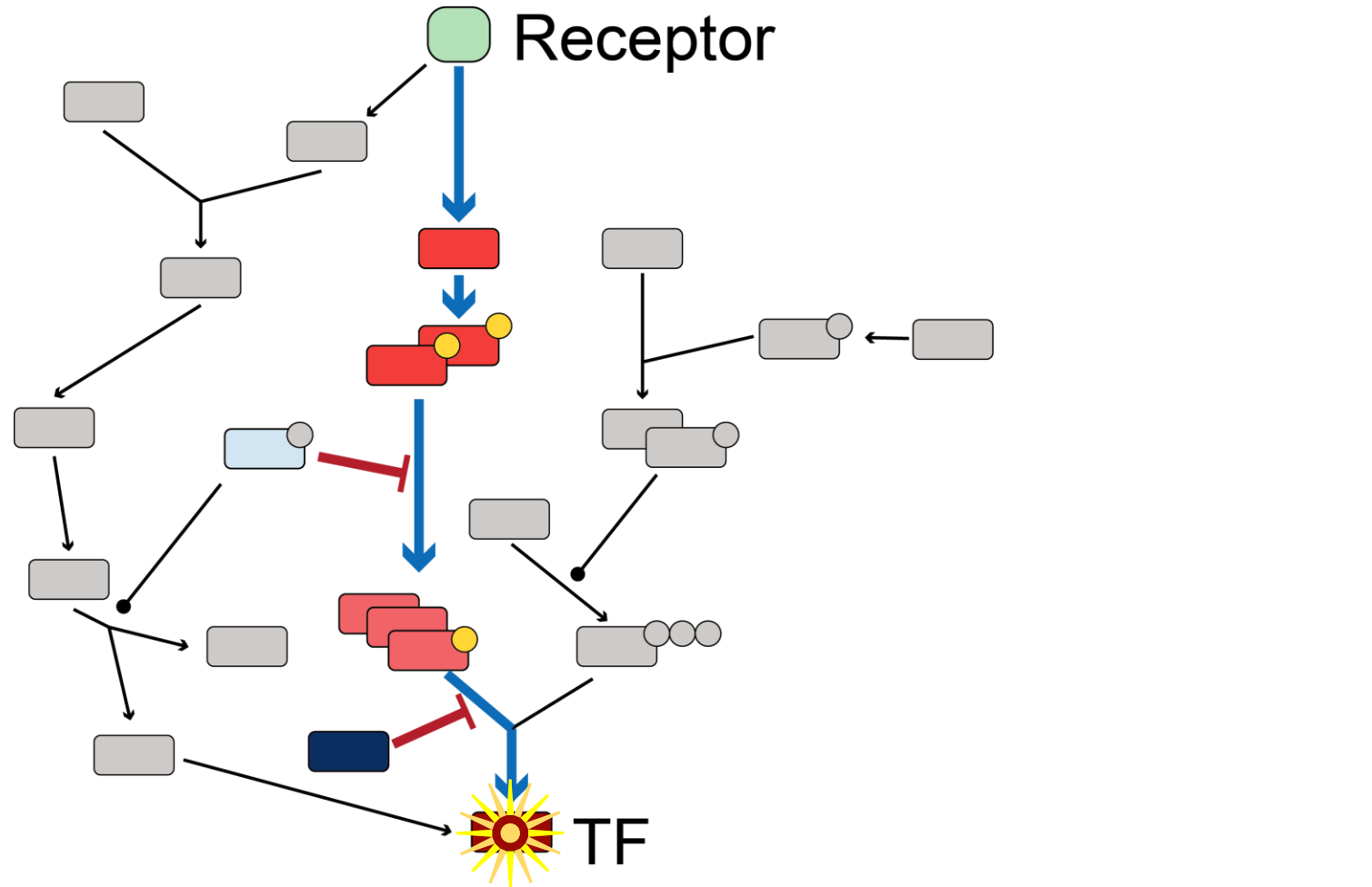
omics data



(sc)RNA-seq data



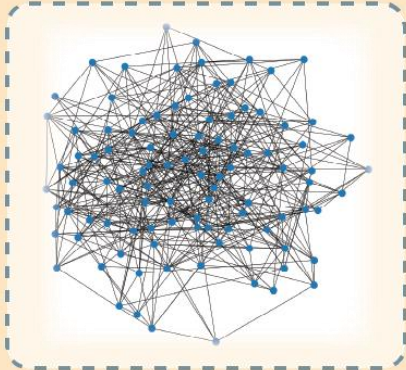
phosphoproteomics



Study deregulation in signaling pathways

SIGMA Method

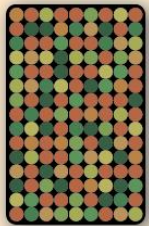
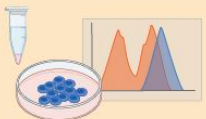
peripheral pathways



reactome

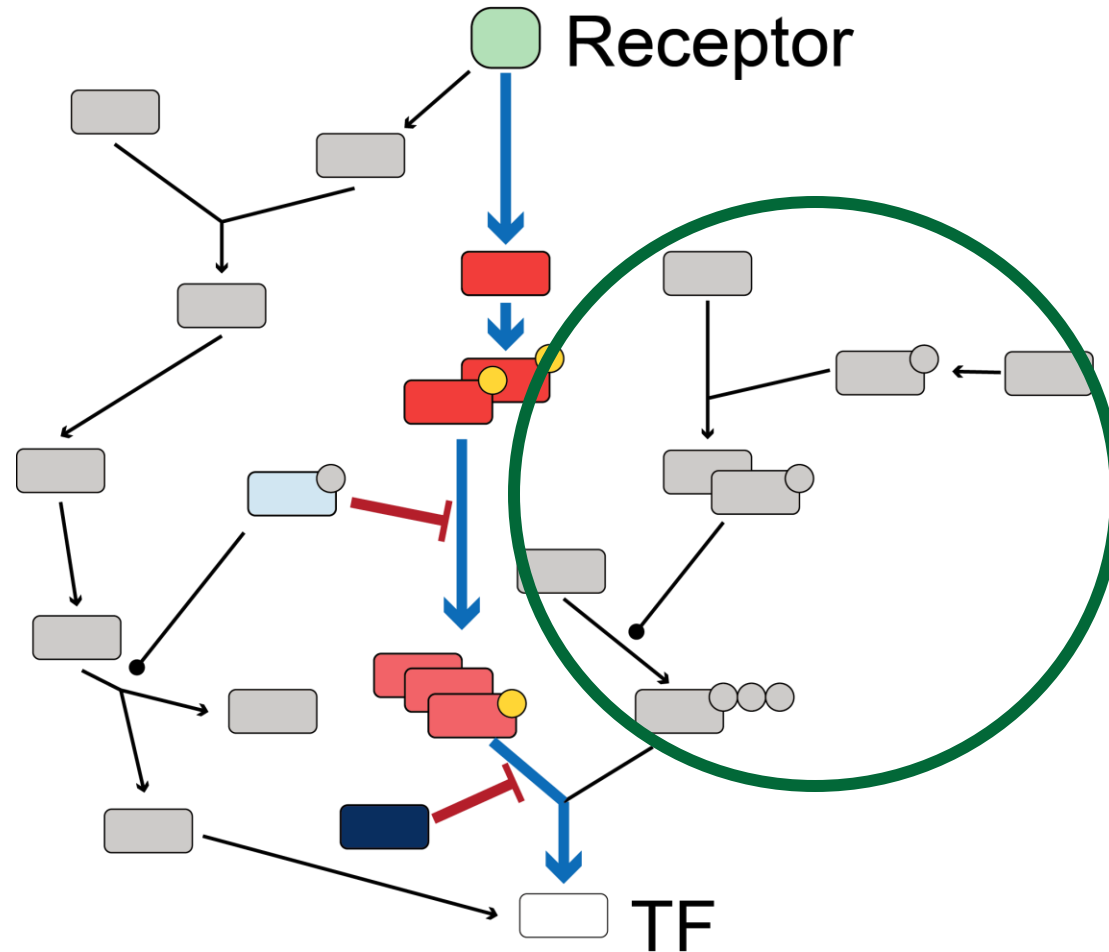
Signaling network

omics data



phosphoproteomics

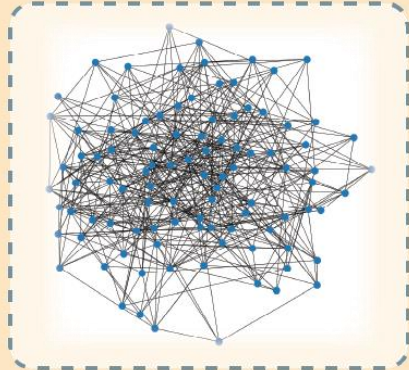
(sc)RNA-seq data



Study deregulation in signaling pathways

SIGMA Method

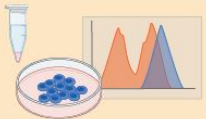
pathway cross-talk



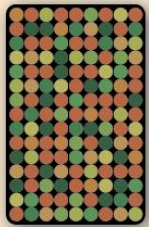
reactome

Signaling network

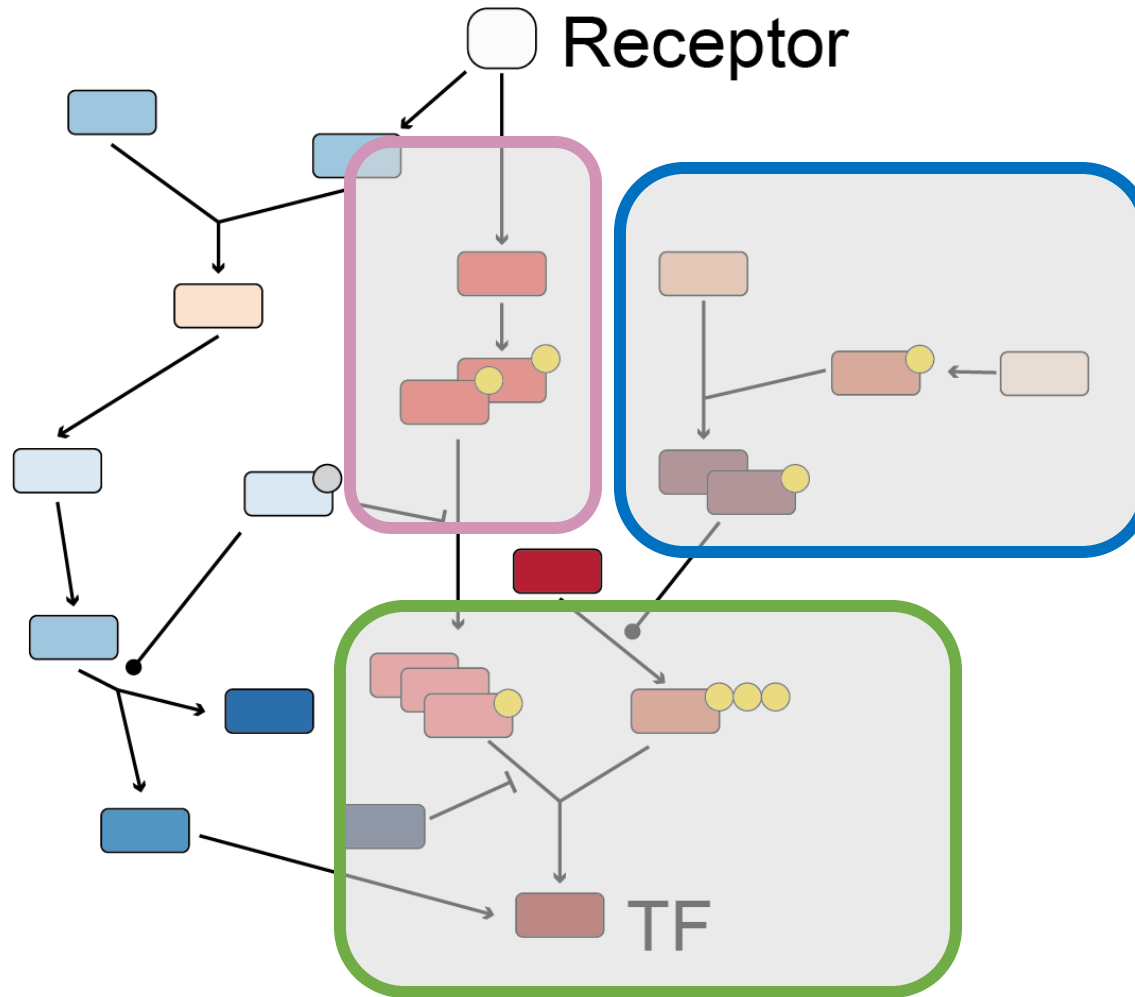
omics data



(sc)RNA-seq data



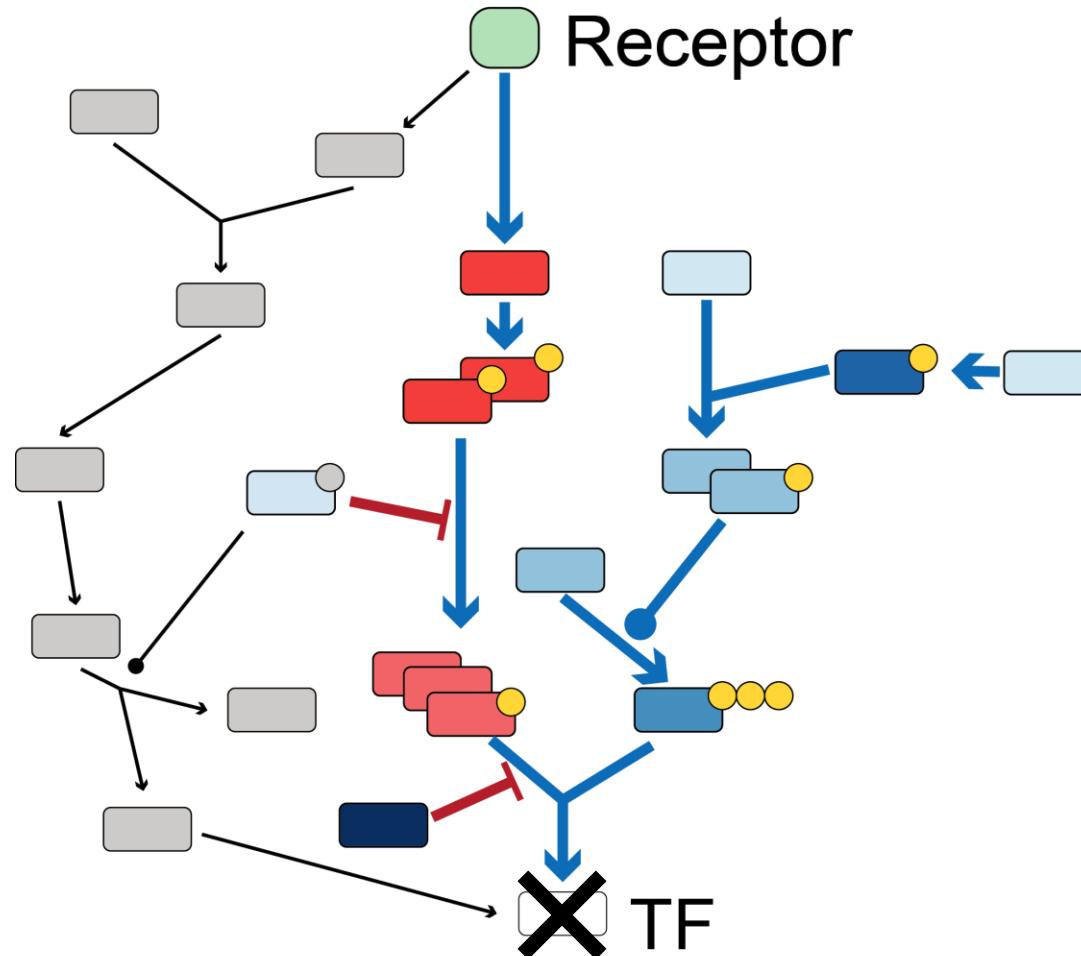
phosphoproteomics



SIGMA Method

Signaling network

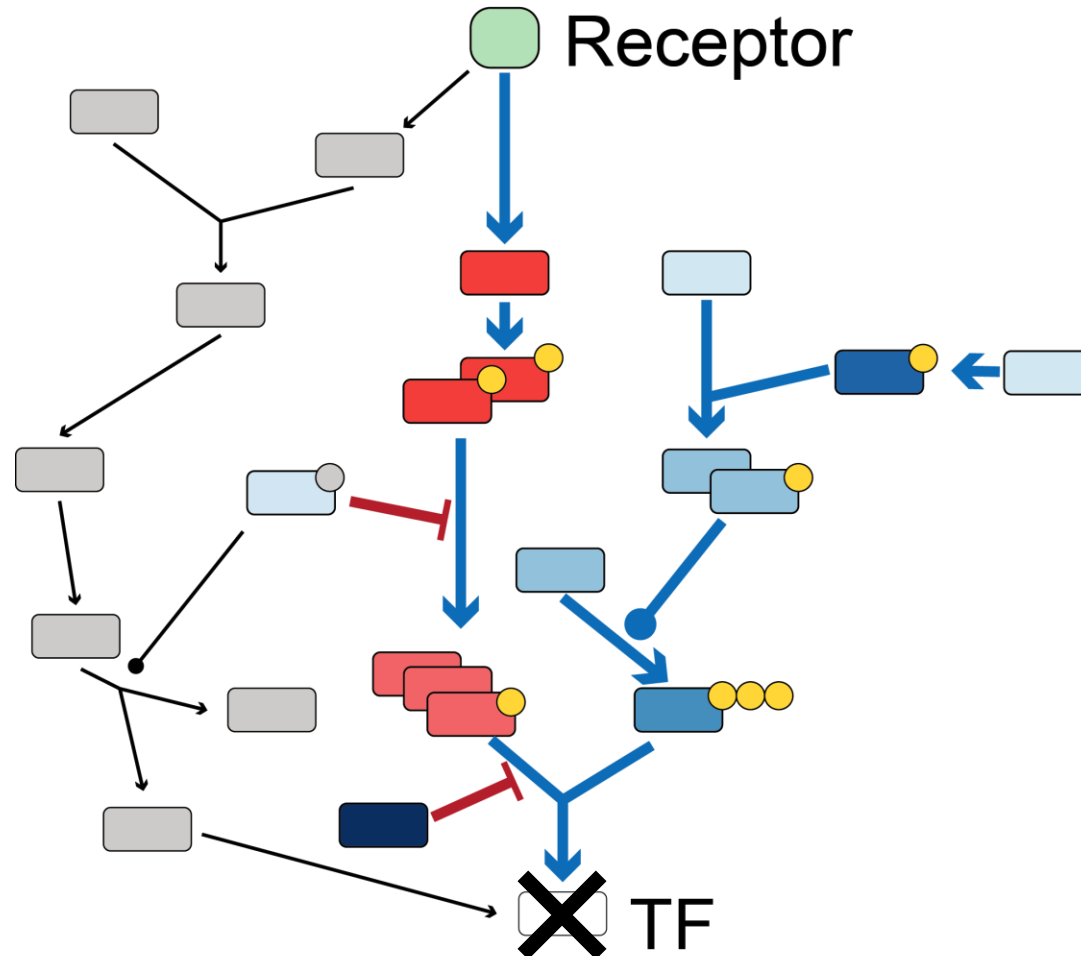
A 10x10 grid of colored circles. The circles are arranged in a pattern that is mostly uniform but has some variations in color. The colors are red, green, and yellow. The grid is filled with these colors in a way that creates a complex, non-repeating pattern.



SIGMA Method

Signaling network

A 10x10 grid of colored circles. The circles are arranged in a pattern that is mostly uniform but has some variations in color. The colors are red, green, and yellow. The grid is filled with these colors in a way that creates a complex, non-repeating pattern.



Neuron
NeuroResource

CellPress

Single-Cell Profiles of Retinal Ganglion Cells Differing in Resilience to Injury Reveal Neuroprotective Genes

Neuron NeuroResource

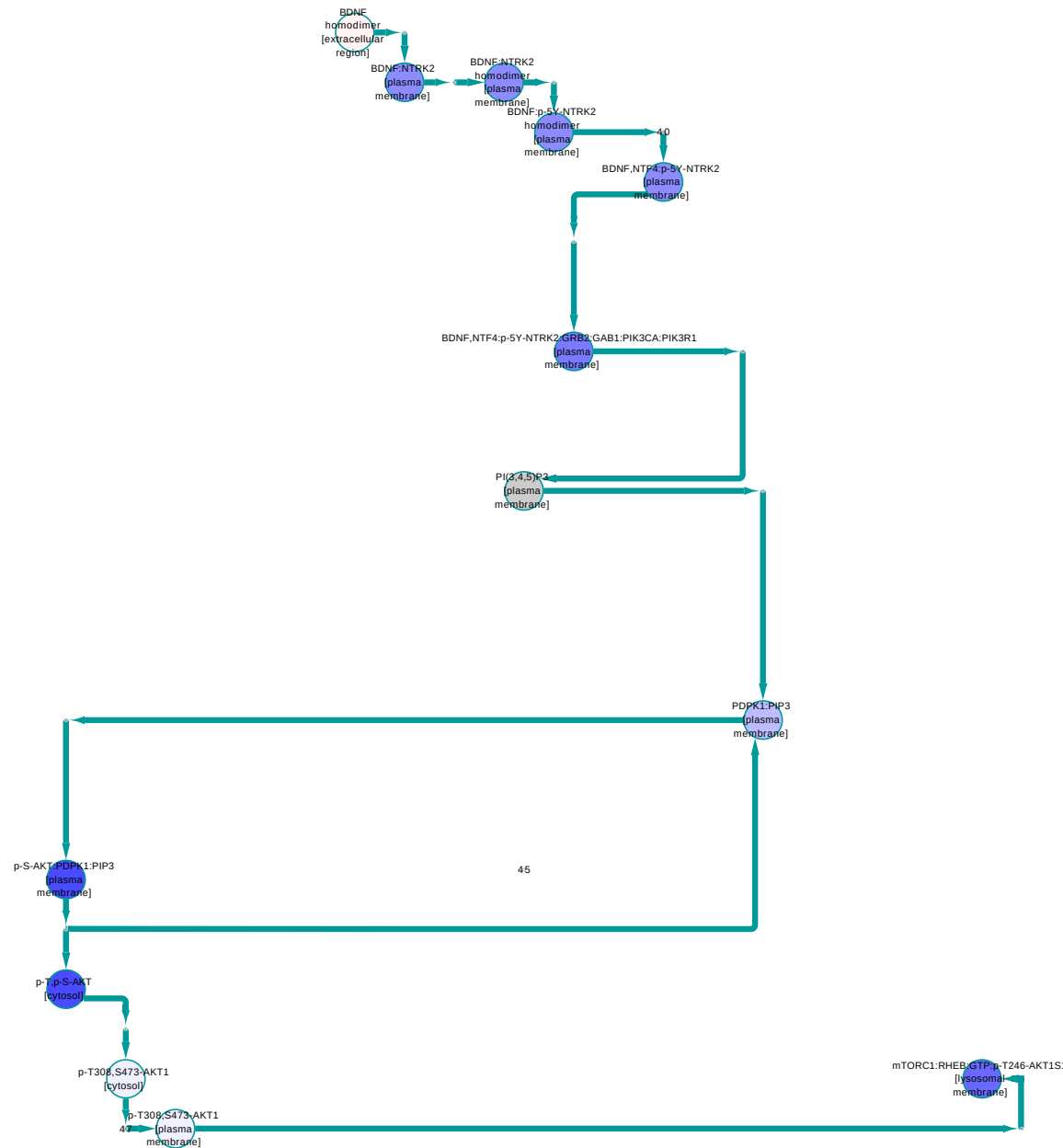
Single-Cell Profiles of Retinal Ganglion Cells Differing in Resilience to Injury Reveal Neuroprotective Genes

Nicholas M. Tran,^{1,7} Karthik Shekhar,^{2,7} Irene E. Whitney,^{1,7} Anne Jacobi,^{3,7} Inbal Ben-Zur,¹ Wenjun Yan,¹ Xian Adiconis,² McKinzie E. Arnold,³ Jung Min Lee,⁴ Joshua Z. Levin,¹ Charles M. Lieber,⁴ Aviv Regev,^{2,6} Zhigang He,³ and Joshua R. Sanes^{1,3,*}

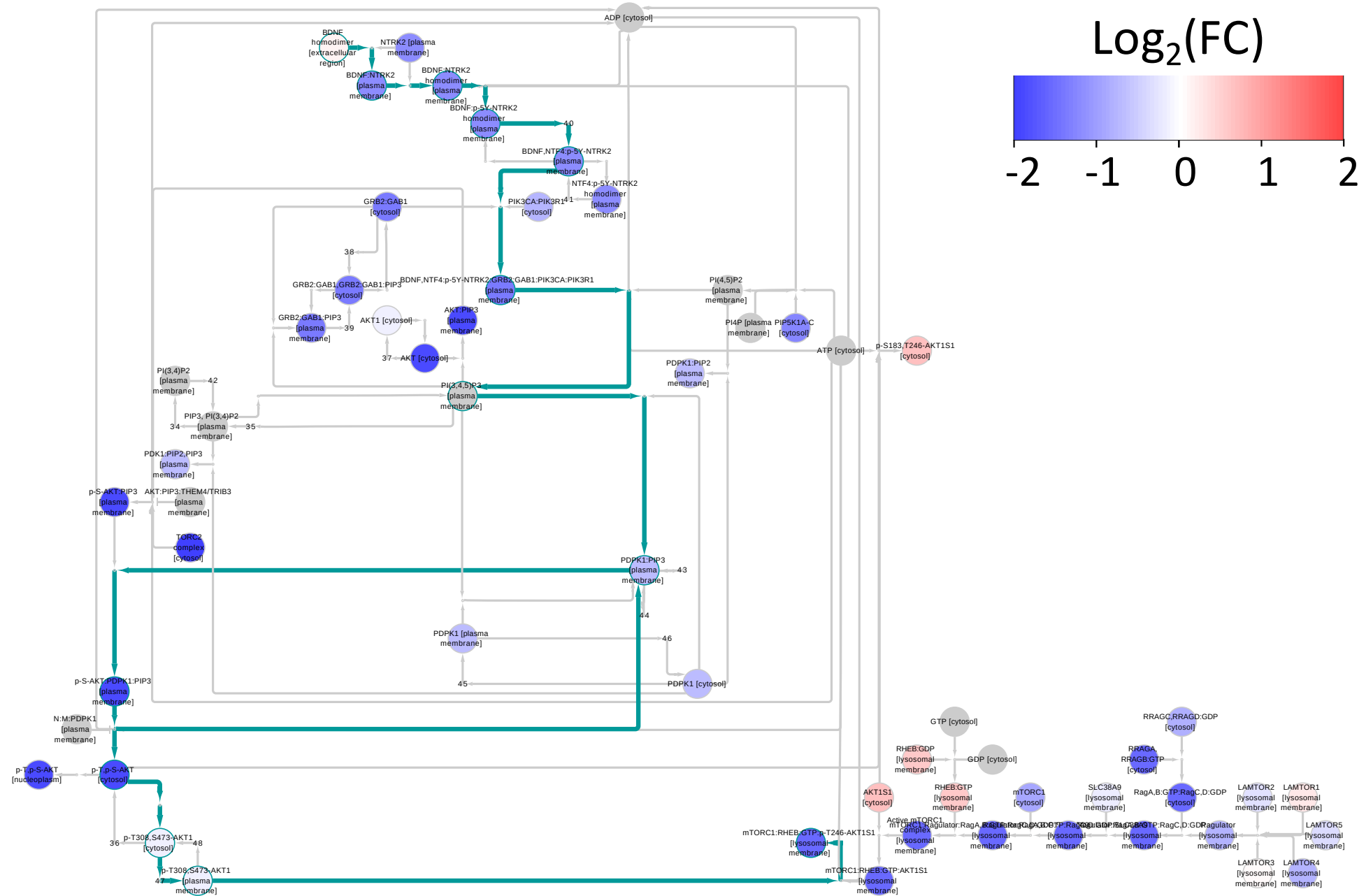
¹Center for Brain Science and Department of Molecular and Cellular Biology, Harvard University, Cambridge, MA 02138, USA
²Broad Institute of Harvard and MIT, Cambridge, MA 02142, USA
³F.M. Kirby Neurobiology Center, Department of Neurology, Boston Children's Hospital, Harvard Medical School, Boston, MA 02130, USA
⁴Department of Chemistry and Chemical Biology, Harvard University, Cambridge, MA 02138, USA
⁵Department of Material Science and Engineering and Wu Tsai Neurosciences Institute, Stanford University, Stanford, CA 94305, USA
⁶Howard Hughes Medical Institute, Chevy Chase, MD 20815, USA and Department of Biology, Johns Hopkins University, Baltimore, MD 21218, USA
⁷These authors contributed equally
⁸These authors contributed equally
⁹Lead Contact
^{*}Correspondence: sanesj@mcb.harvard.edu
<https://doi.org/10.1016/j.neuron.2019.11.006>

scRNA-seq data in

Studying BDNF connection to mTOR



Studying BDNF connection to mTOR



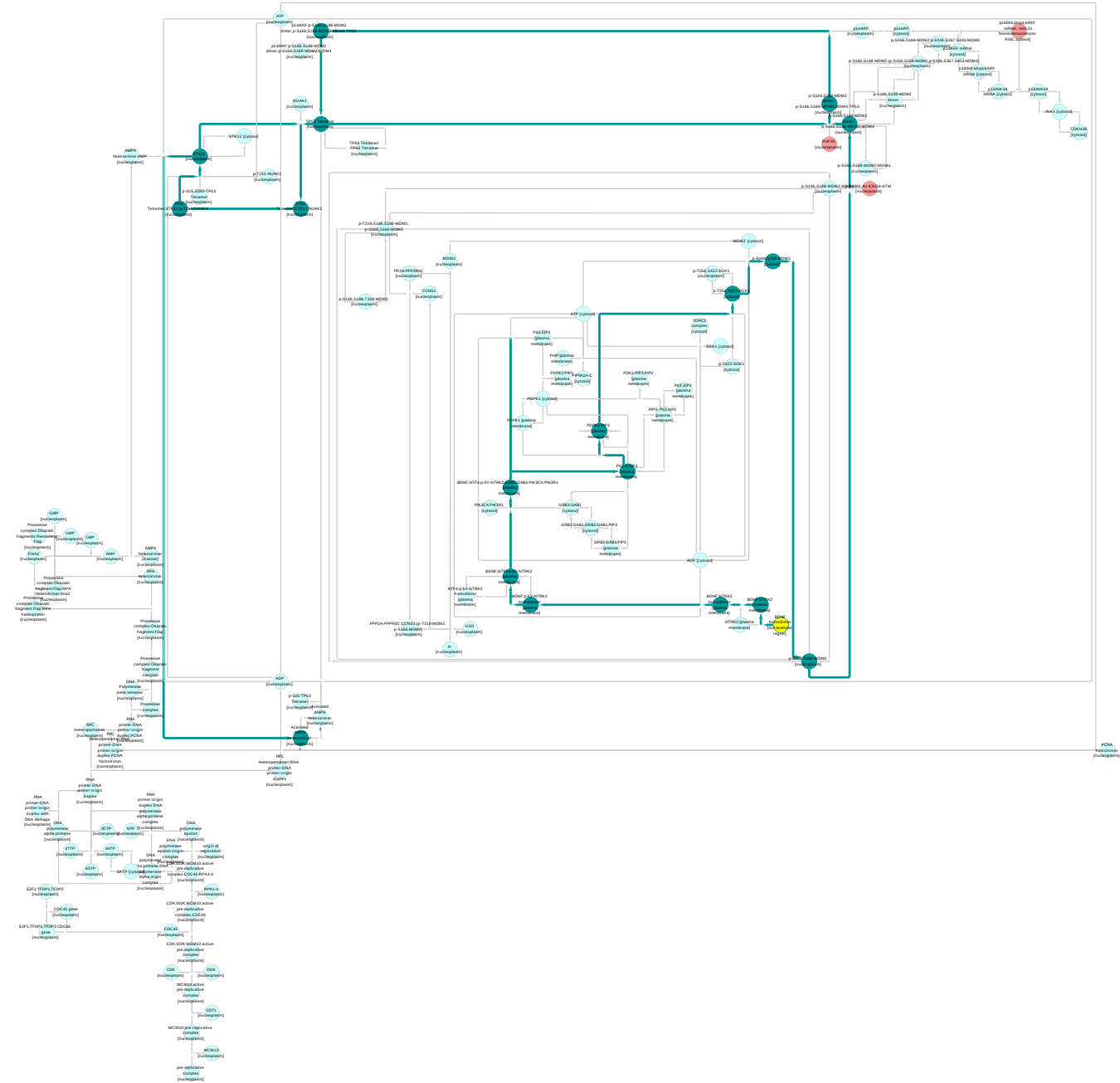
Studying BDNF connection to AMPK

Start	Target	Result	Min path
BDNF homodimer [e]	Activated AMPK heterotrimer [n]	YES	18
BDNF homodimer [e]	AMPK heterotrimer (active) [c]	NO	-
BDNF homodimer [e]	AMPK heterotrimer (inactive) [n]	NO	-
BDNF homodimer [e]	AMPK heterotrimer [c]	YES	21
BDNF homodimer [e]	AMPK heterotrimer:AMP [n]	NO	-
BDNF homodimer [e]	AMPK-alpha2:AMPK-beta:AMPK-gamma:AMP [c]	NO	-
BDNF homodimer [e]	p-AMPK heterotrimer [c]	YES	20
BDNF homodimer [e]	p-AMPK heterotrimer:AMP [c]	YES	21
BDNF homodimer [e]	p-AMPK heterotrimer:AMP [n]	YES	22
BDNF homodimer [e]	p-AMPK heterotrimer:AMP:p-S317,467,556,638,T575-ULK1: ATG13:RB1CC1:ATG101 [c]	YES	23
BDNF homodimer [e]	p-AMPK heterotrimer:AMP:p-T180,S317,467,556,638,T575-ULK1: ATG13:RB1CC1:ATG101 [c]	YES	24
BDNF homodimer [e]	p-AMPK heterotrimer:AMP:p-T180,S317,467,556,638,T575-ULK1: ATG13:RB1CC1:ATG101 [er]	YES	26
BDNF homodimer [e]	p-AMPK heterotrimer:AMP:p-T180,S317,467,556,638,T575-ULK1: p-ATG13:p-RB1CC1:ATG101 [c]	YES	25
BDNF homodimer [e]	p-AMPK heterotrimer:AMP:ULK1:ATG13:RB1CC1:ATG101 [c]	YES	22
BDNF homodimer [e]	SESN1,2,3:p-AMPK heterotrimer:AMP [c]	YES	17

Studying BDNF connection to AMPK

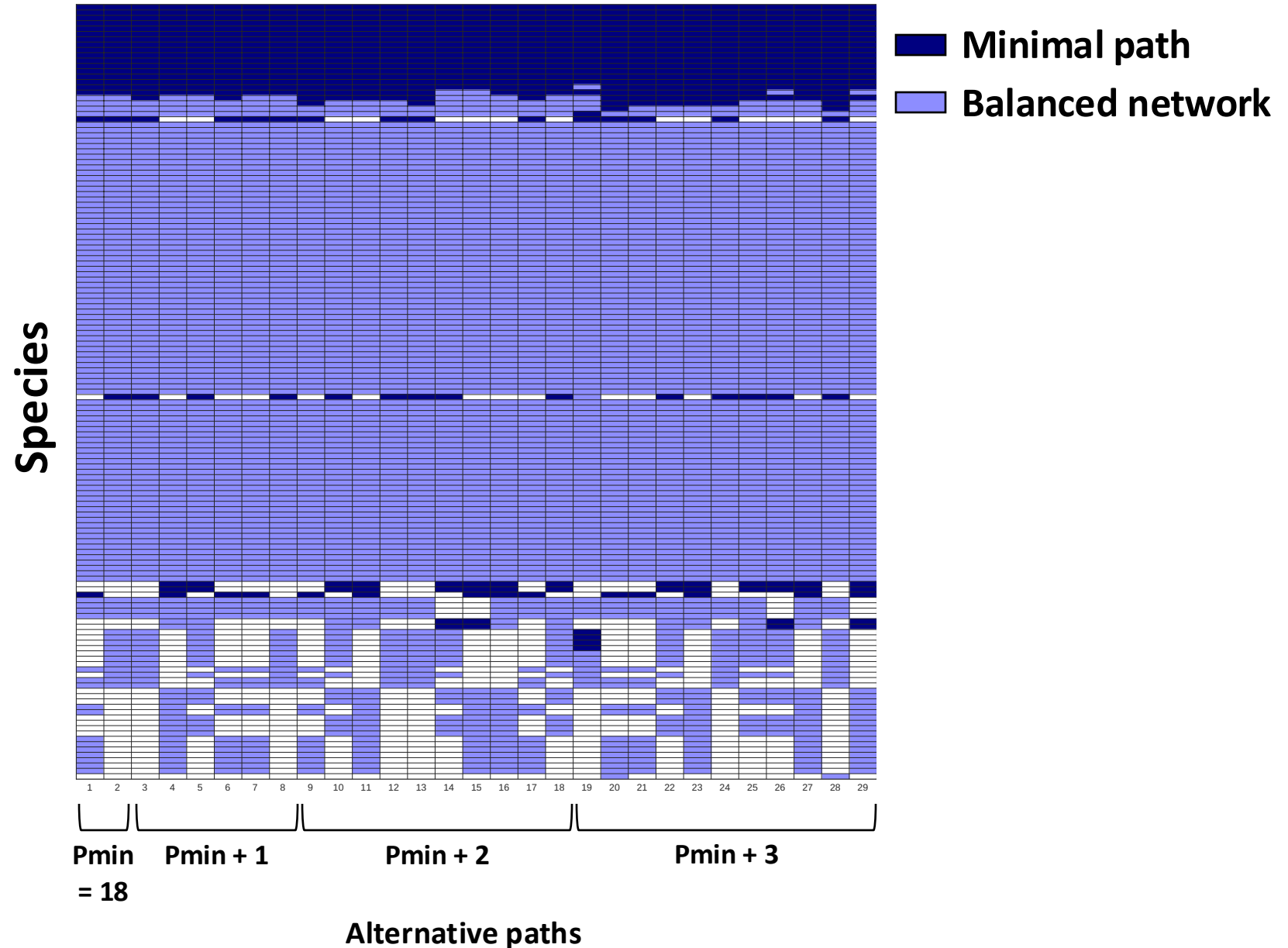
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BDNF homodimer [e]	AMPK heterotrimer (inactive) [n]	NO	-
BDNF homodimer [e]	AMPK heterotrimer [c]	YES	21
BDNF homodimer [e]	AMPK heterotrimer:AMP [n]	NO	-
BDNF homodimer [e]	AMPK-alpha2:AMPK-beta:AMPK-gamma:AMP [c]	NO	-
BDNF homodimer [e]	p-AMPK heterotrimer [c]	YES	20
BDNF homodimer [e]	p-AMPK heterotrimer:AMP [c]	YES	21
BDNF homodimer [e]	p-AMPK heterotrimer:AMP [n]	YES	22
BDNF homodimer [e]	p-AMPK heterotrimer:AMP:p-S317,467,556,638,T575-ULK1: ATG13:RB1CC1:ATG101 [c]	YES	23
BDNF homodimer [e]	p-AMPK heterotrimer:AMP:p-T180,S317,467,556,638,T575-ULK1: ATG13:RB1CC1:ATG101 [c]	YES	24
BDNF homodimer [e]	p-AMPK heterotrimer:AMP:p-T180,S317,467,556,638,T575-ULK1: ATG13:RB1CC1:ATG101 [er]	YES	26
BDNF homodimer [e]	p-AMPK heterotrimer:AMP:p-T180,S317,467,556,638,T575-ULK1: p-ATG13:p-RB1CC1:ATG101 [c]	YES	25
BDNF homodimer [e]	p-AMPK heterotrimer:AMP:ULK1:ATG13:RB1CC1:ATG101 [c]	YES	22
BDNF homodimer [e]	SESN1,2,3:p-AMPK heterotrimer:AMP [c]	YES	17

Studying BDNF connection to AMPK



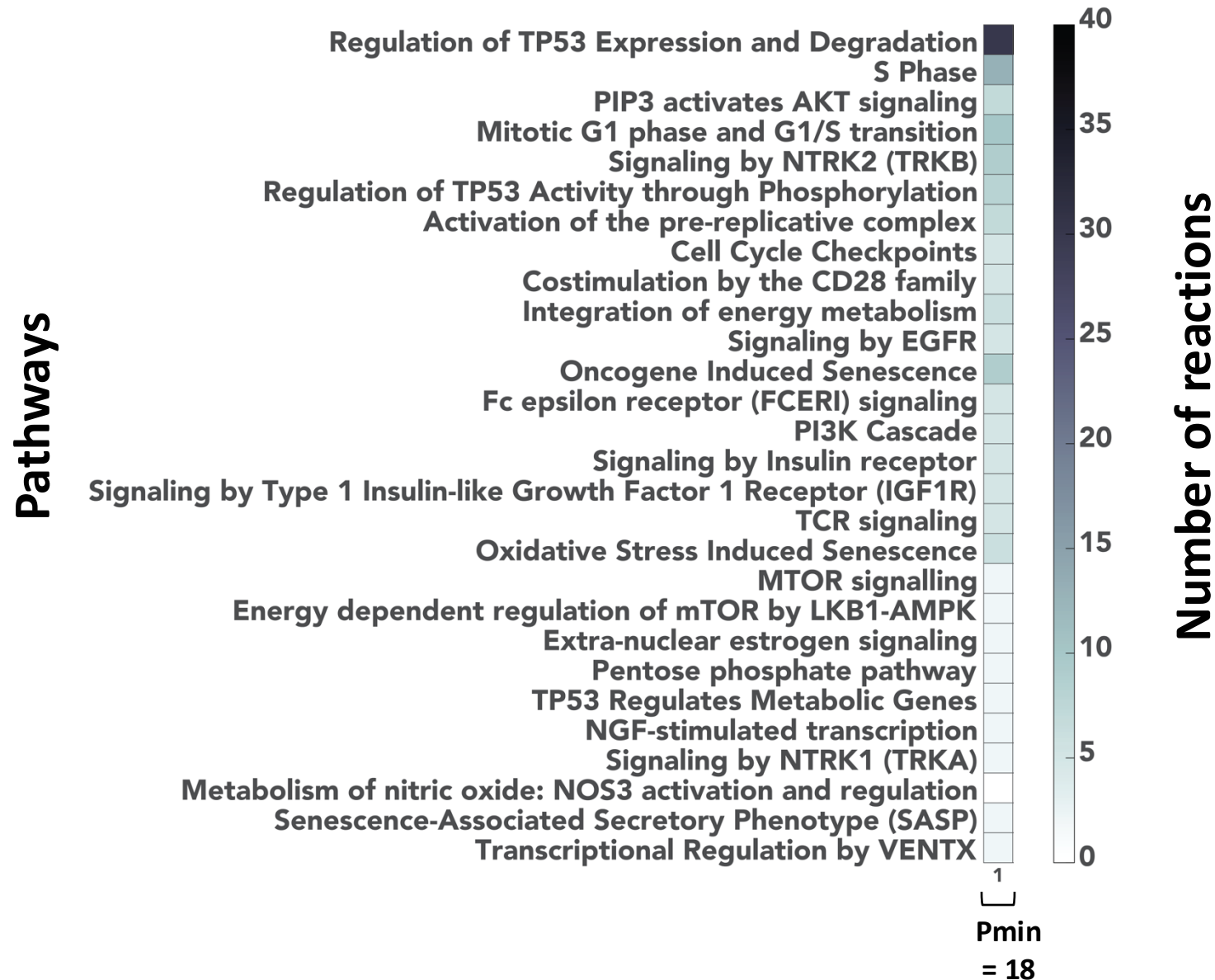
Studying BDNF connection to AMPK

BDNF to Activated AMPK heterotrimer



Studying BDNF connection to AMPK

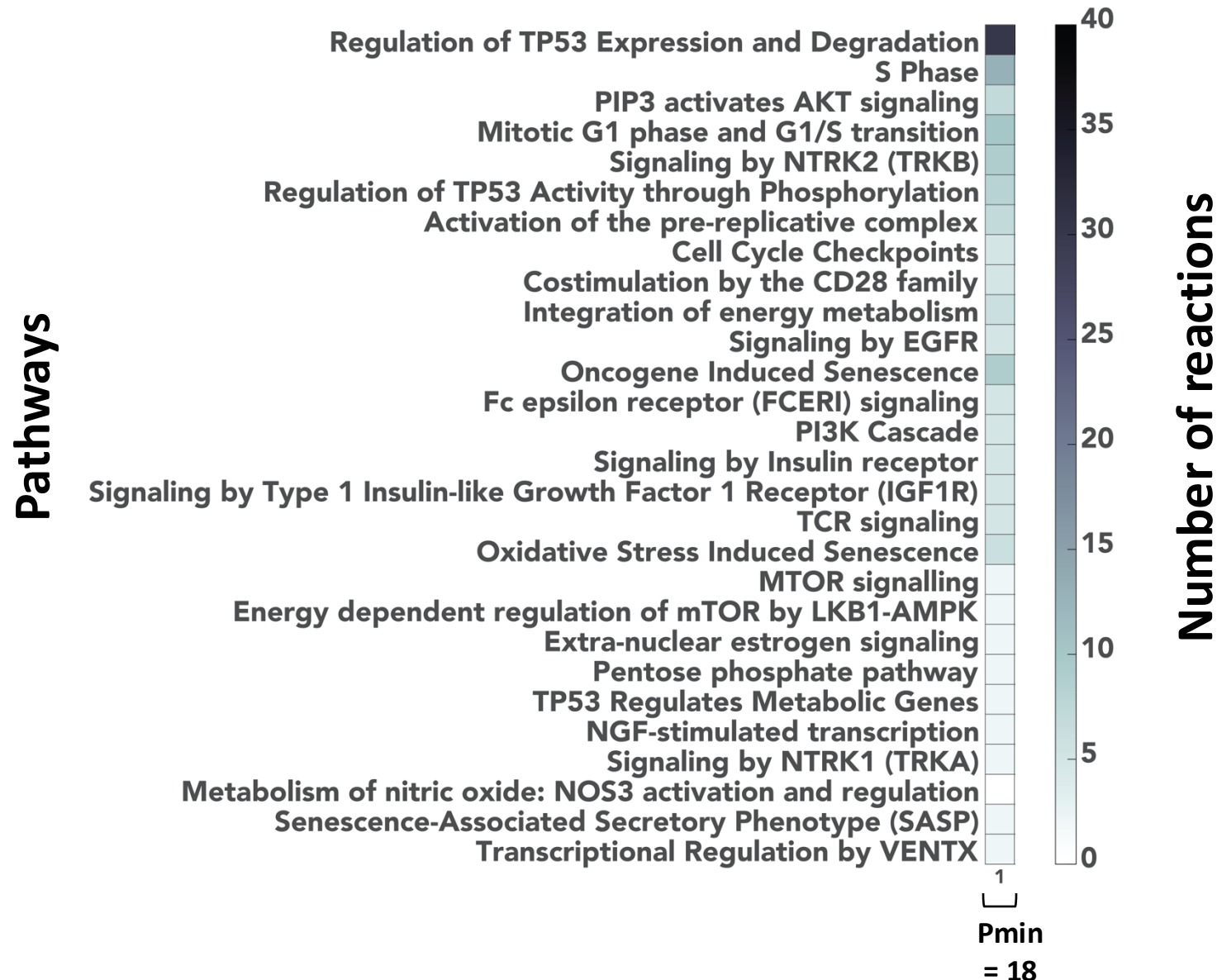
BDNF to Activated AMPK heterotrimer



- 18 steps from BDNF to AMPK

Studying BDNF connection to AMPK

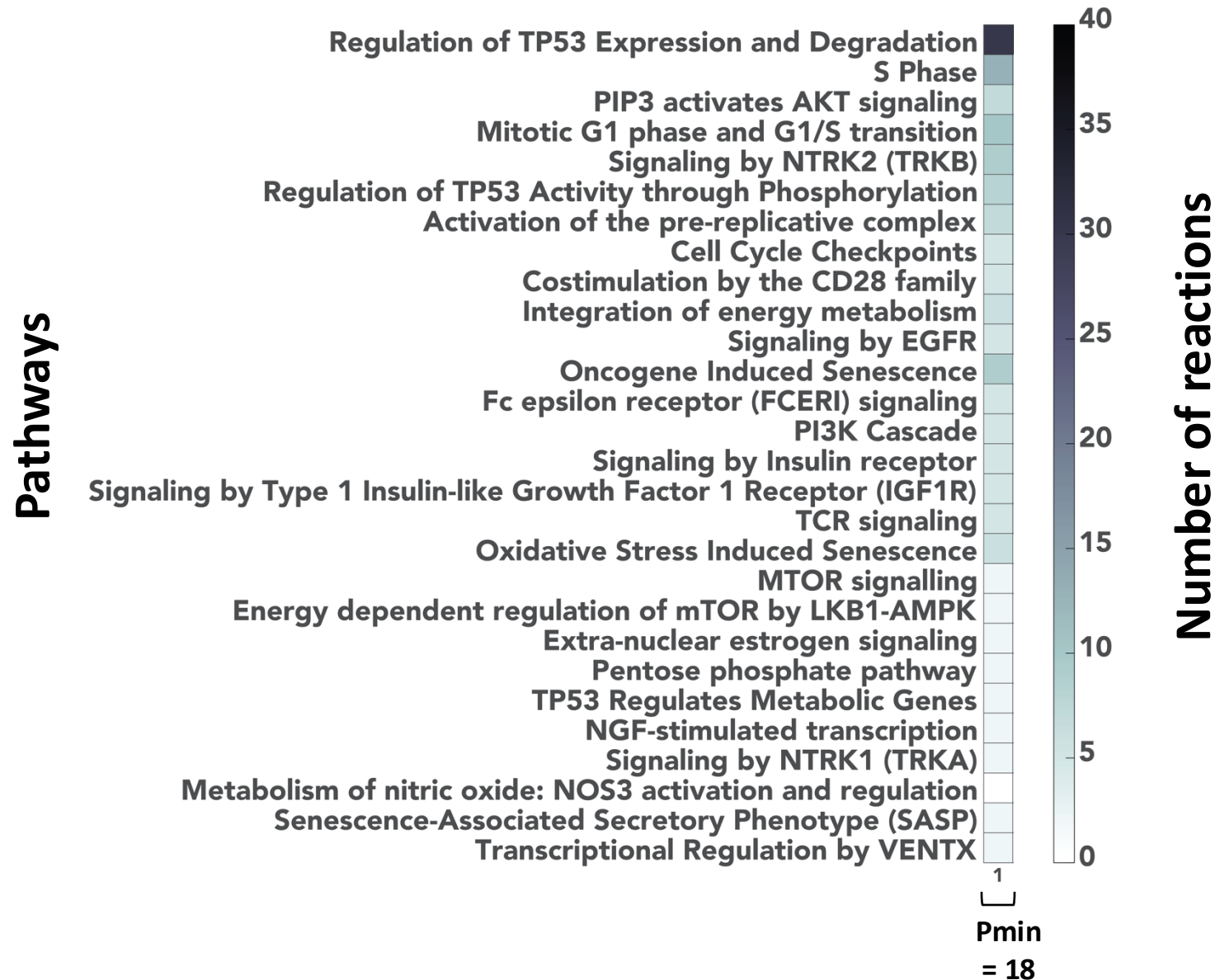
BDNF to Activated AMPK heterotrimer



- 18 steps from BDNF to AMPK
- Total of 104 reactions (including the peripheral)

Studying BDNF connection to AMPK

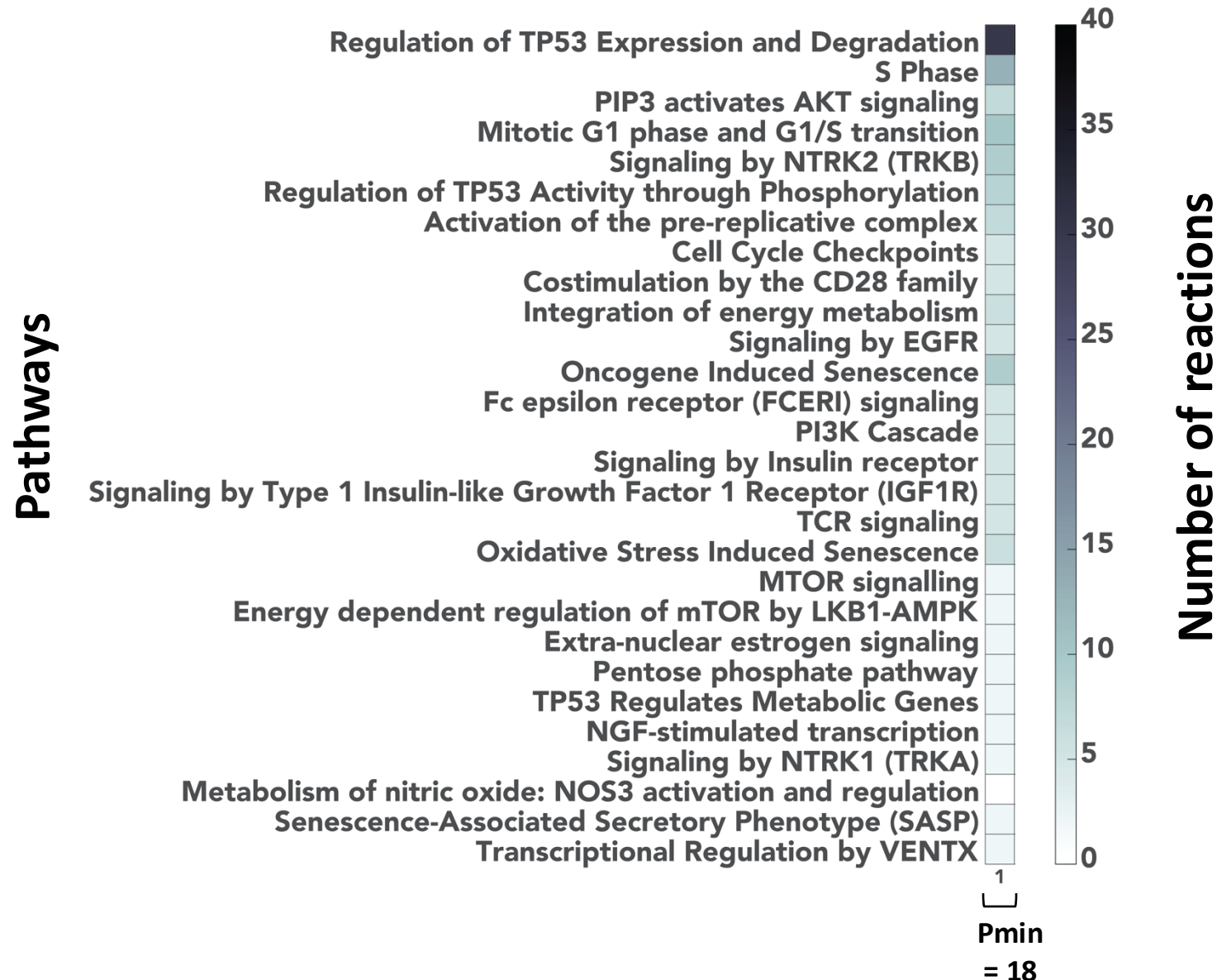
BDNF to Activated AMPK heterotrimer



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- Total of 104 reactions (including the peripheral)
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Studying BDNF connection to AMPK

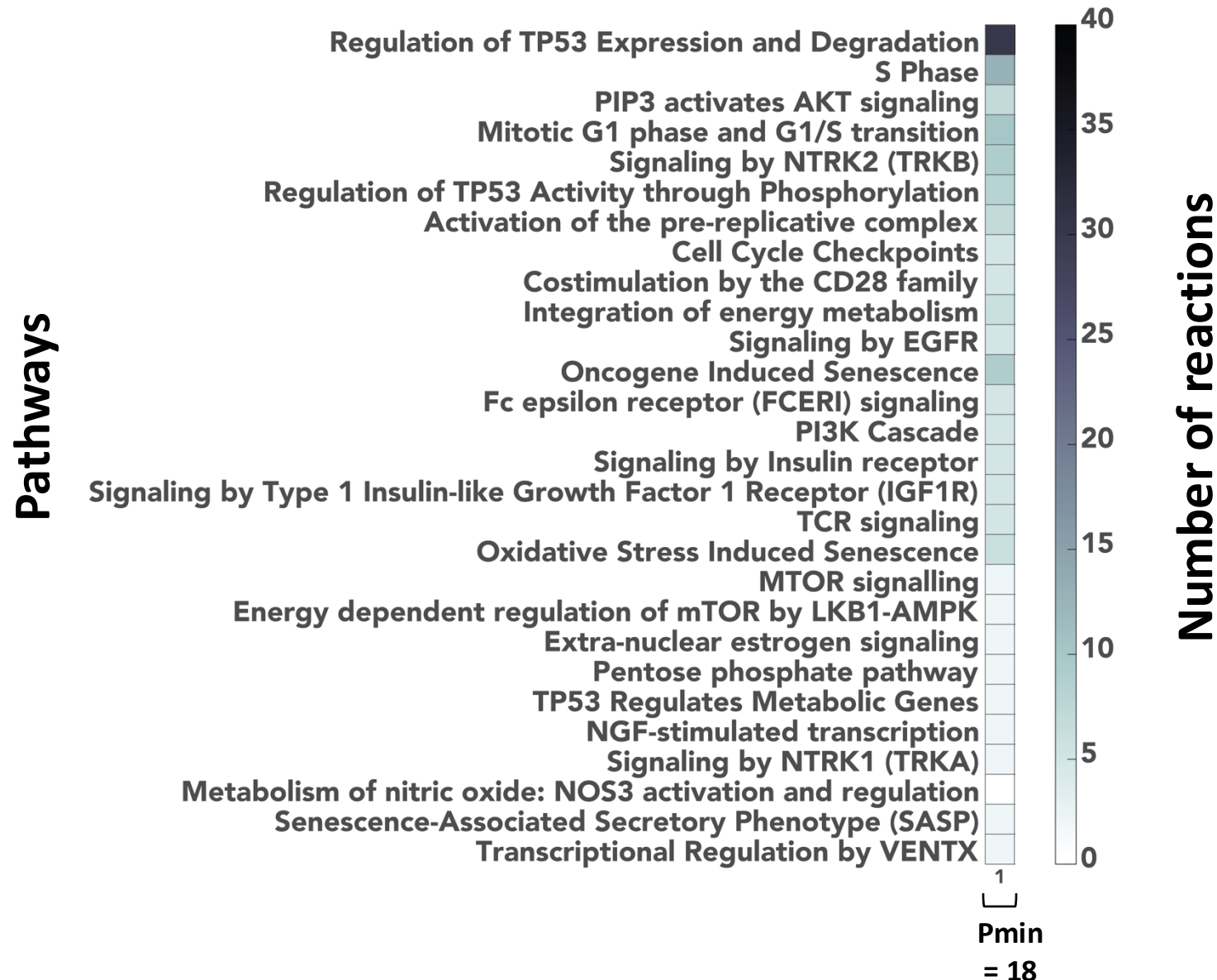
BDNF to Activated AMPK heterotrimer



- 18 steps from BDNF to AMPK
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- Max of 30 reactions in Regulation of TP53

Studying BDNF connection to AMPK

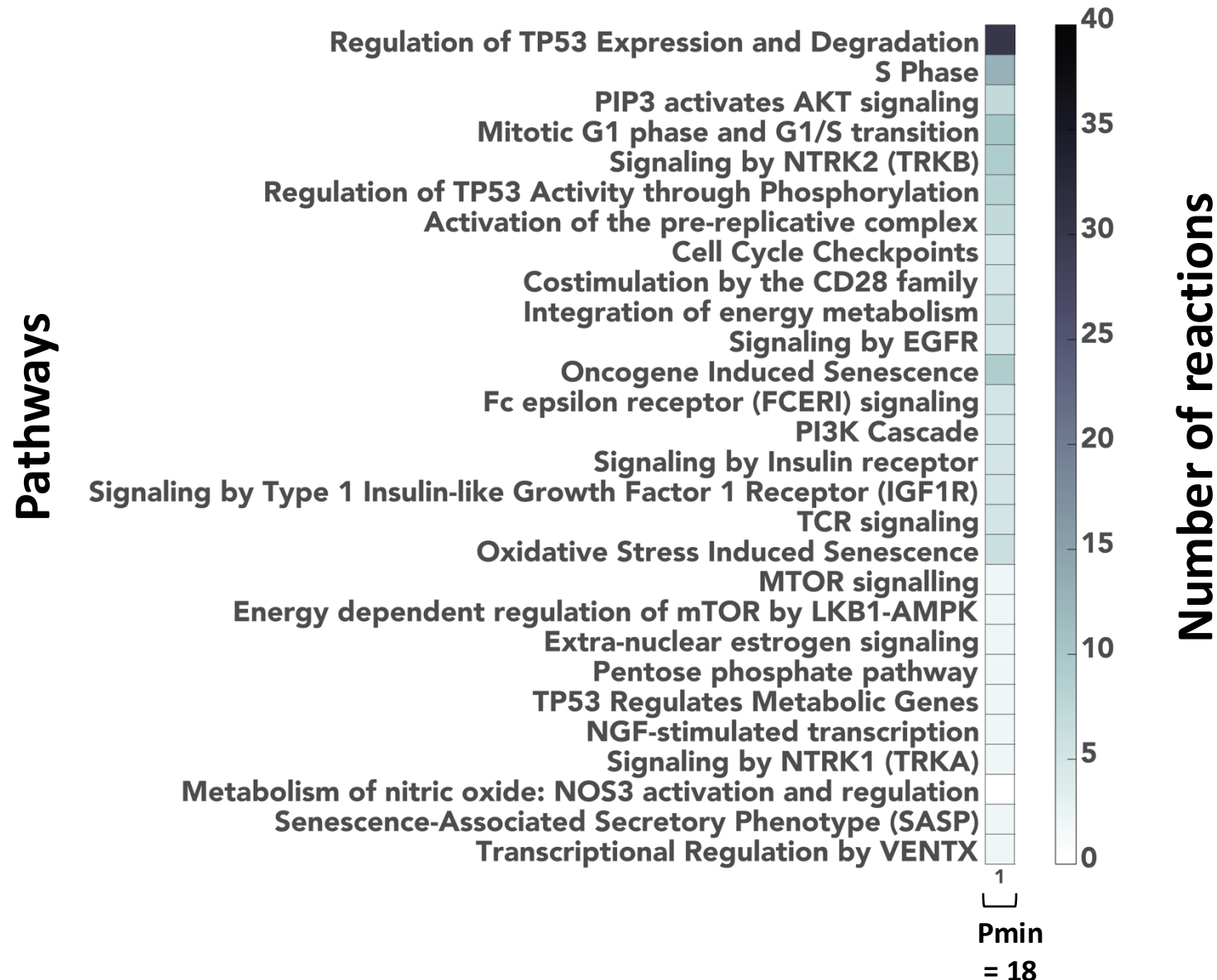
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- 18 steps from BDNF to AMPK
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Studying BDNF connection to AMPK

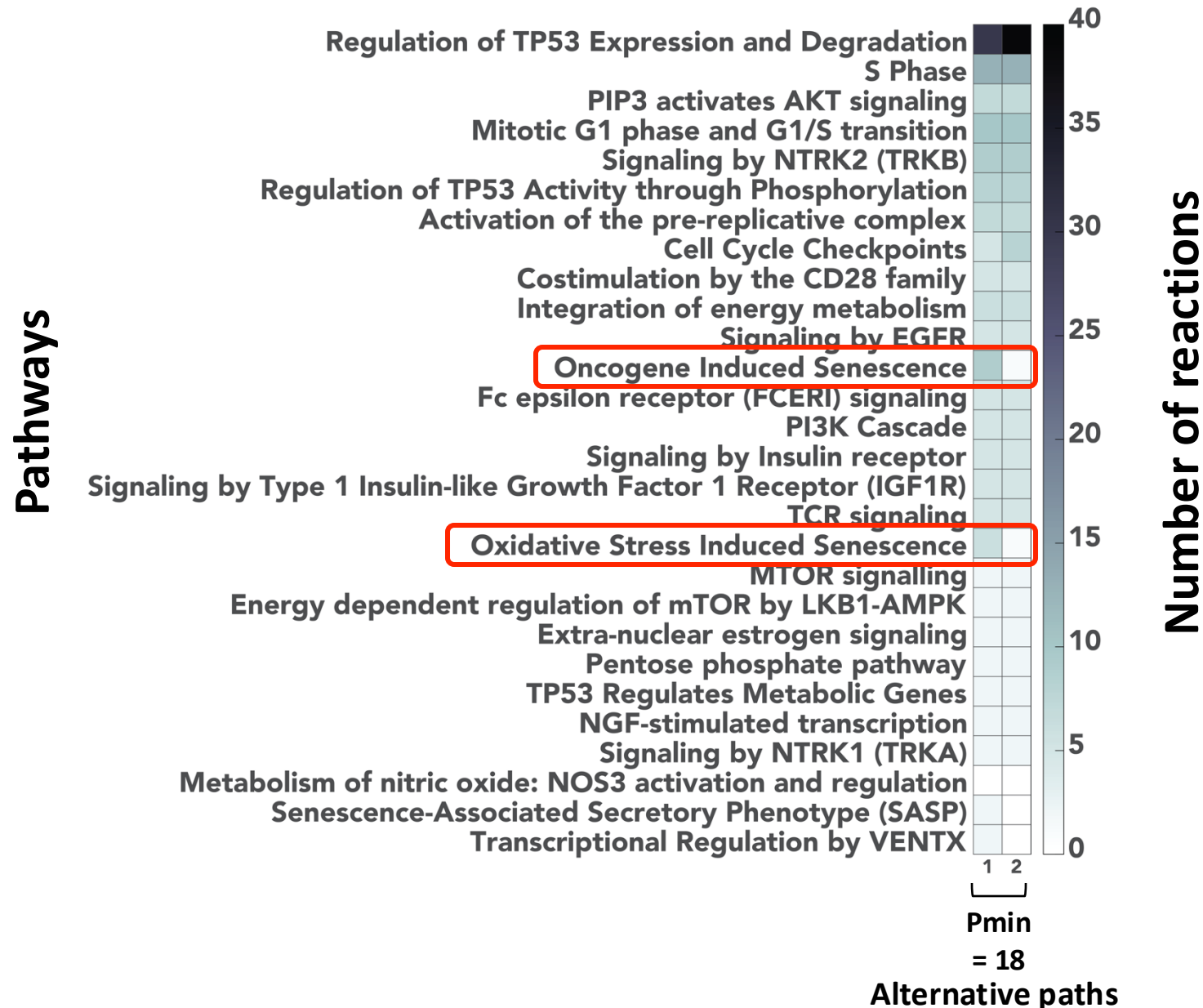
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- Metabolism of nitric oxide does not participate in this path

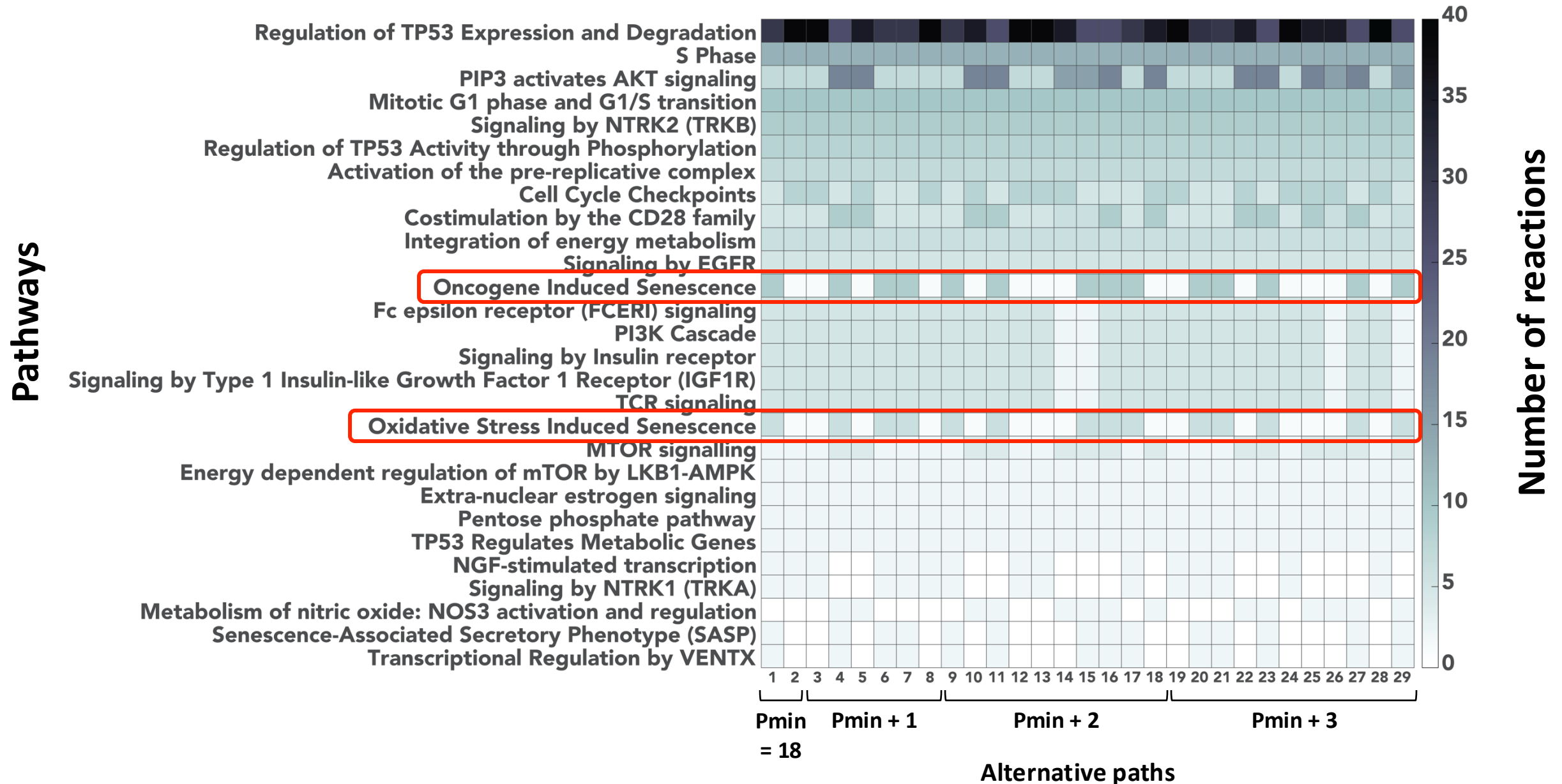
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BDNF to Activated AMPK heterotrimer



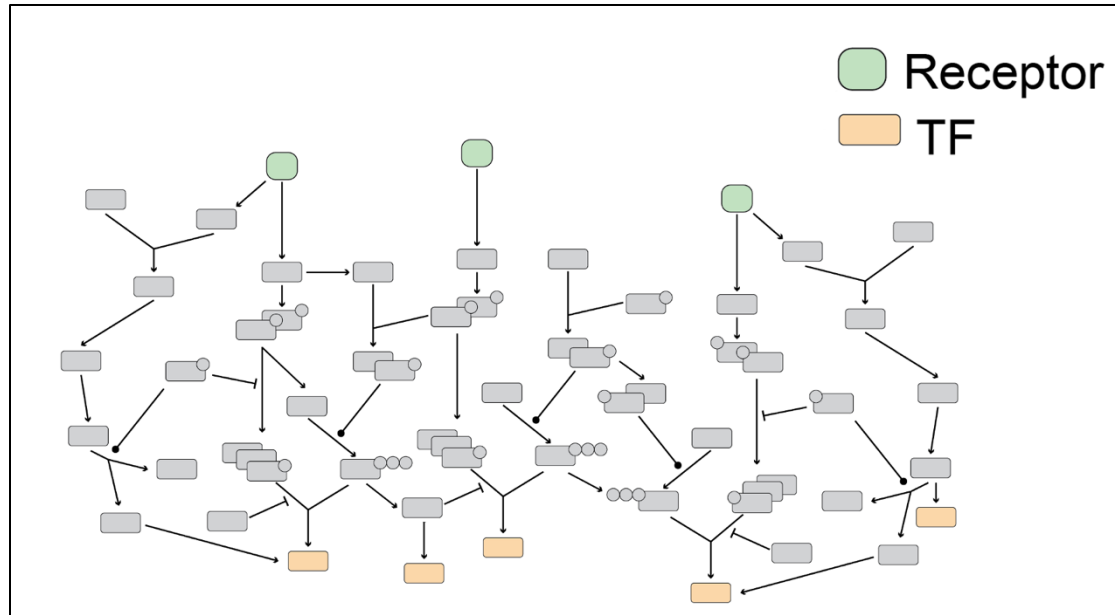
Studying BDNF connection to AMPK

BDNF to Activated AMPK heterotrimer



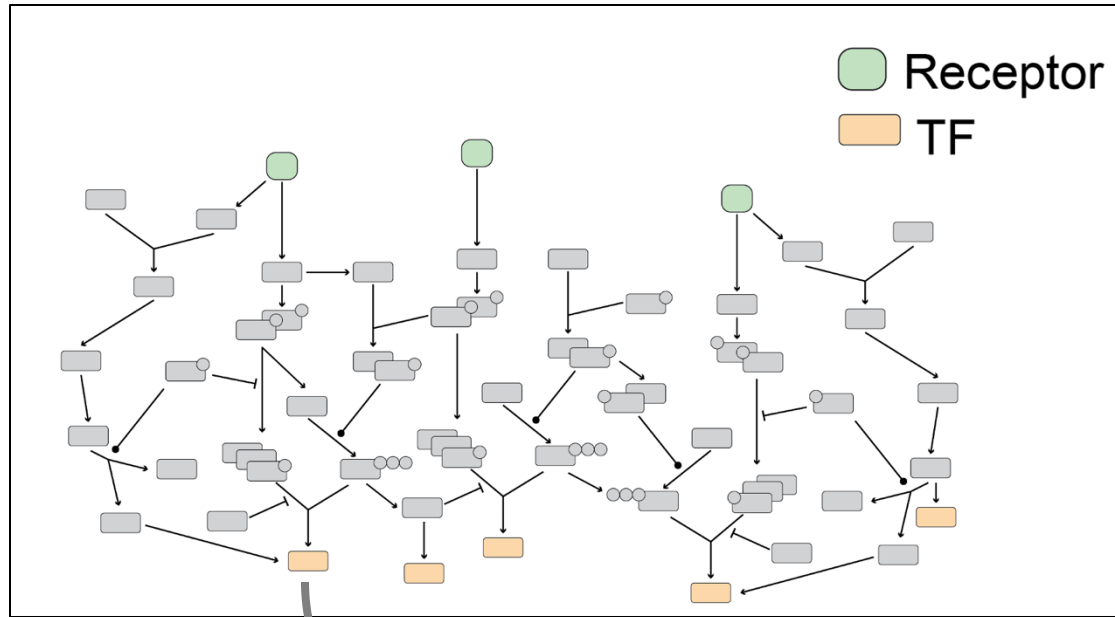
Connecting signaling to metabolism

Signaling network

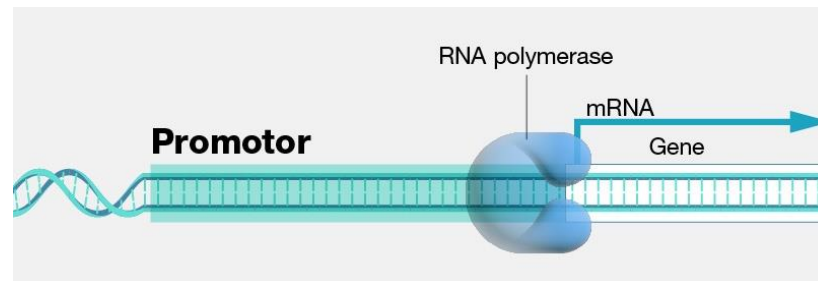


Connecting signaling to metabolism

Signaling network

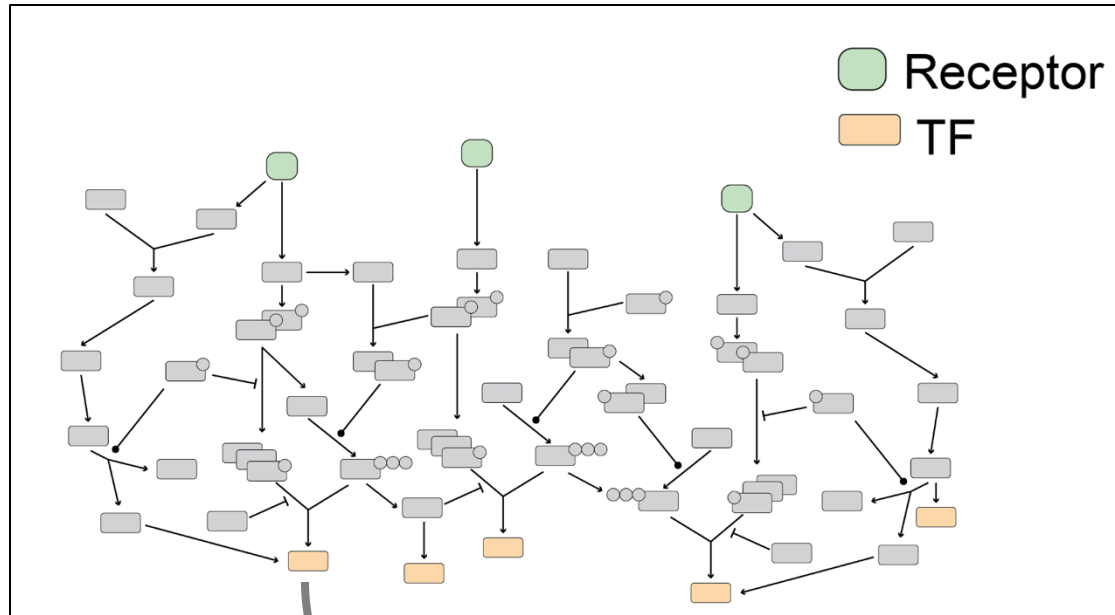


Gene expression

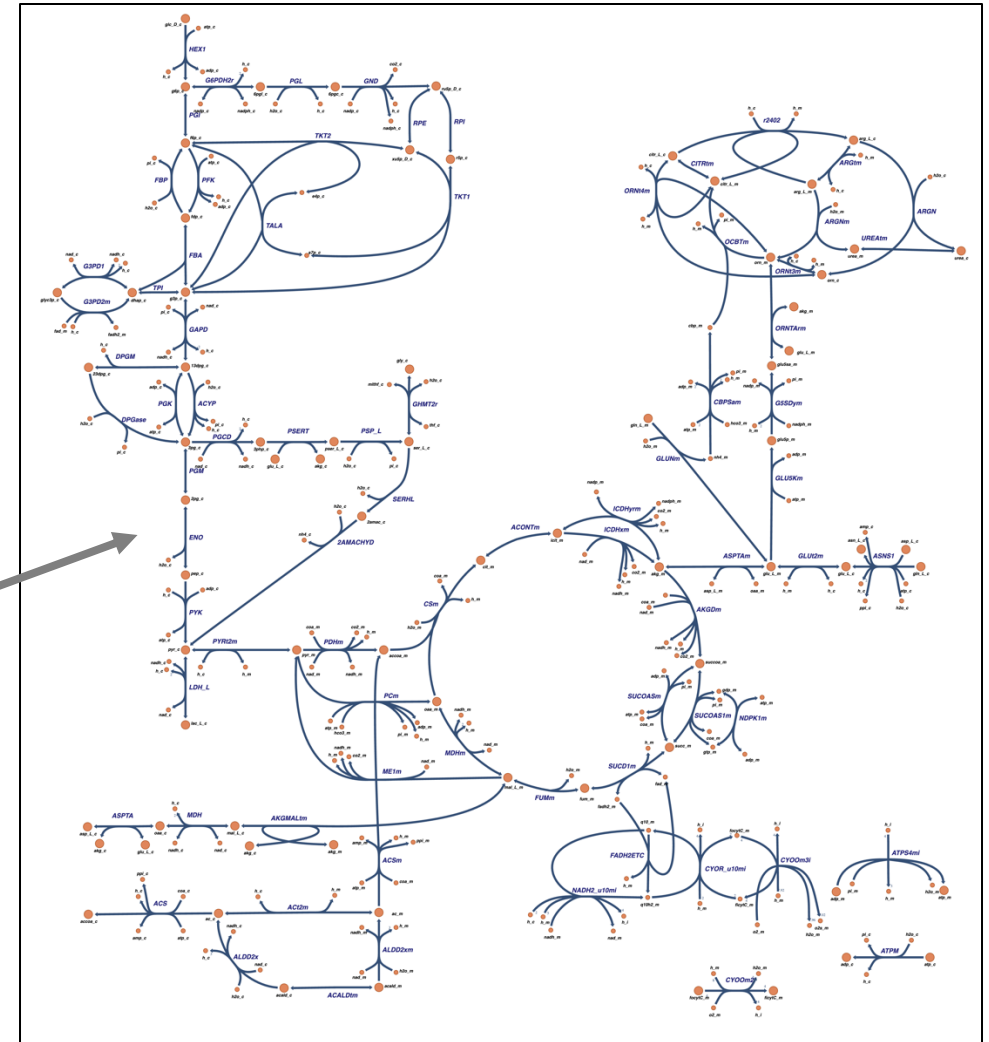


Connecting signaling to metabolism

Signaling network

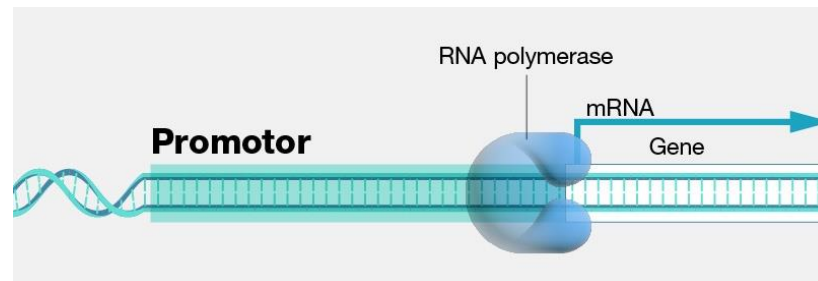


Metabolic network



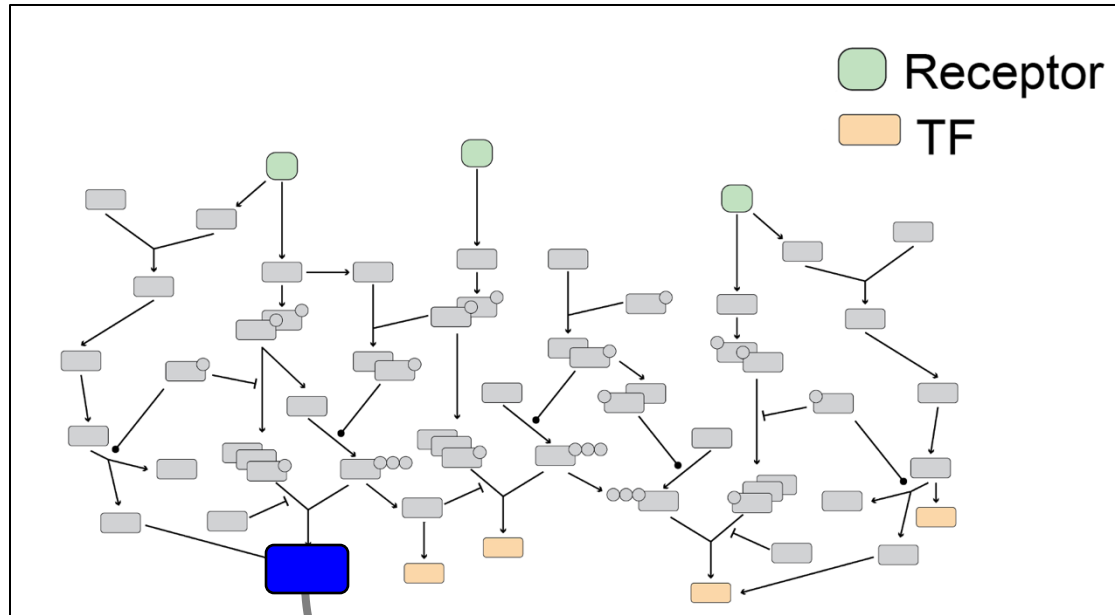
*** 115 TFs in Recon3D**

Gene expression

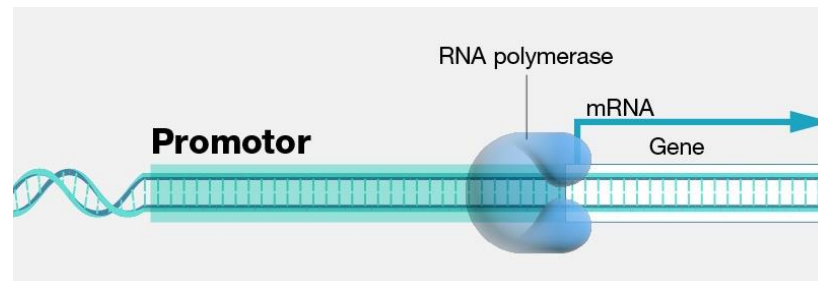


Connecting signaling to metabolism

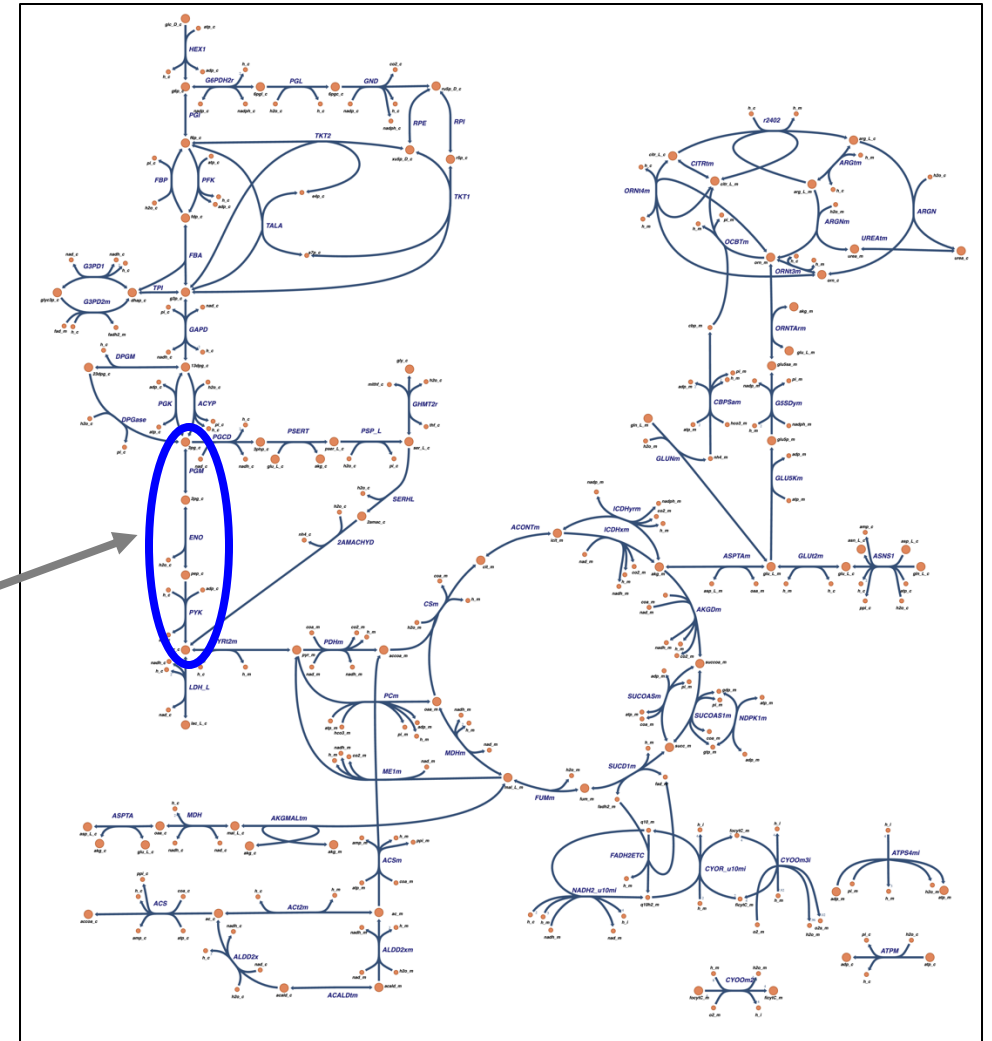
Signaling network



Gene expression

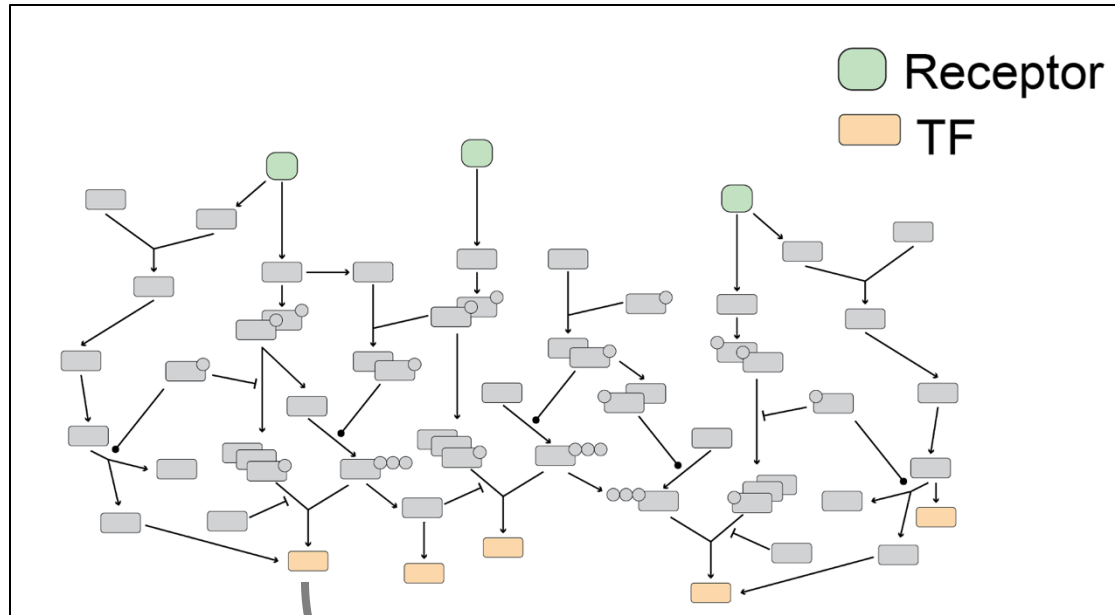


Metabolic network

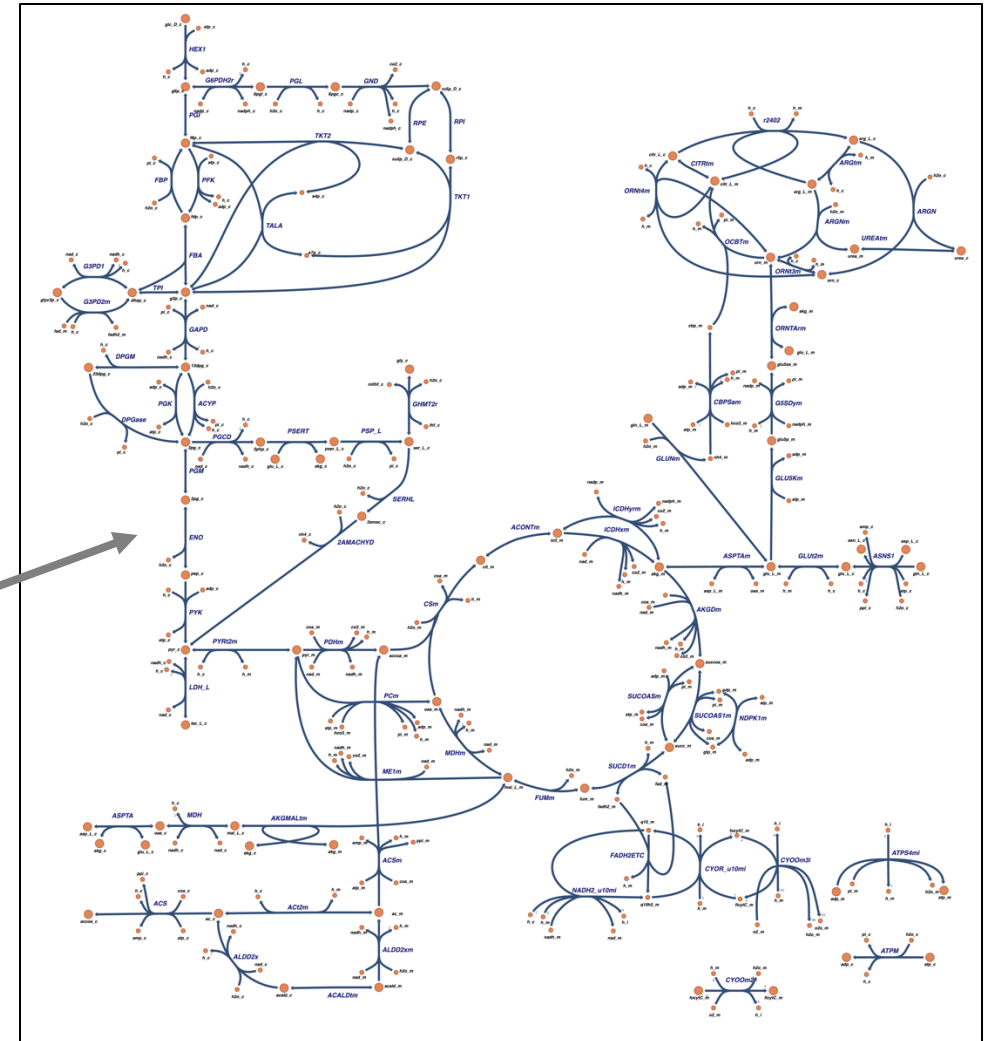


Connecting signaling to metabolism

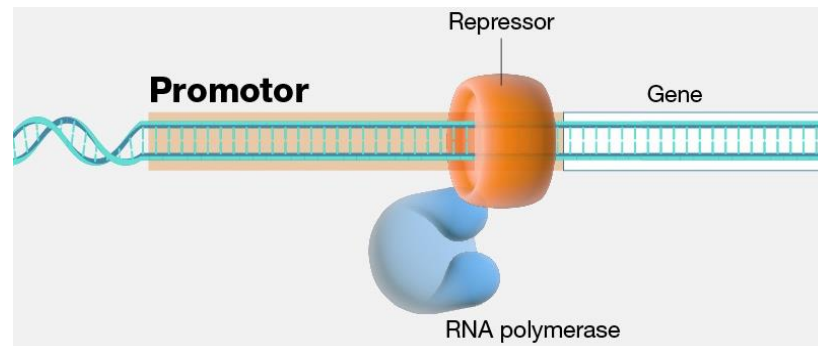
Signaling network



Metabolic network

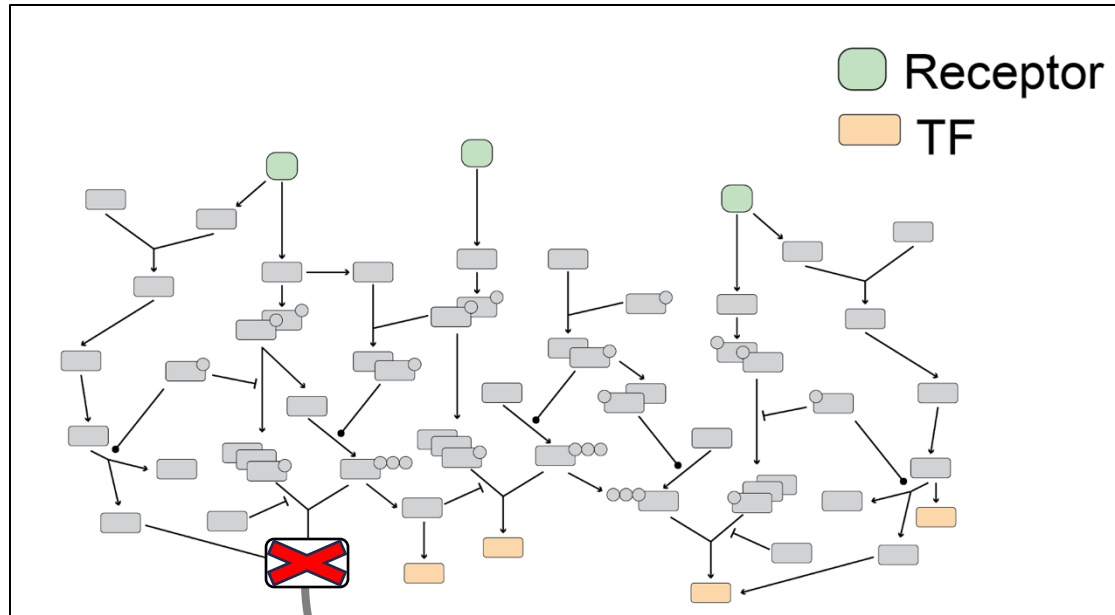


Gene expression

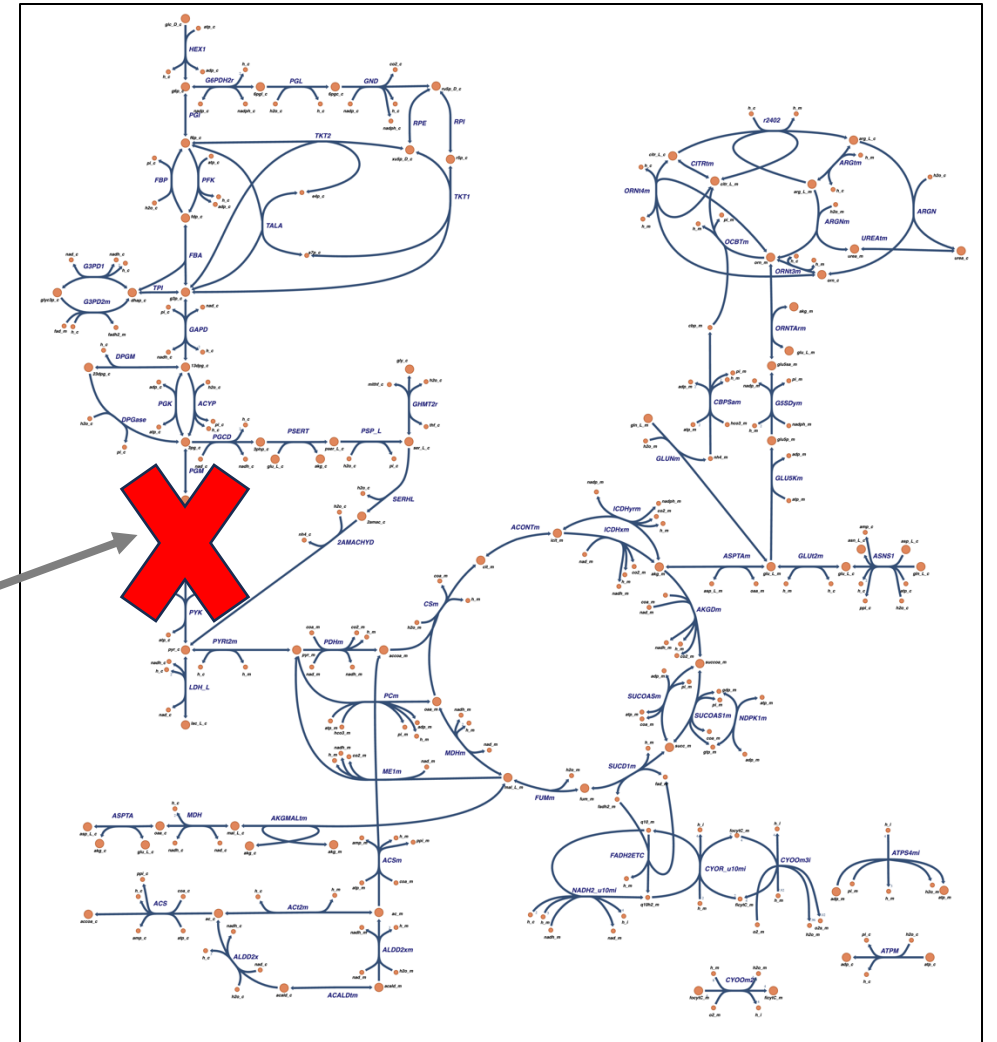


Connecting signaling to metabolism

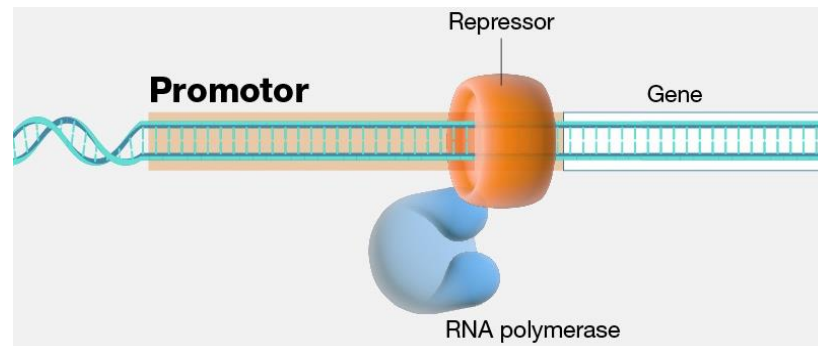
Signaling network



Metabolic network



Gene expression



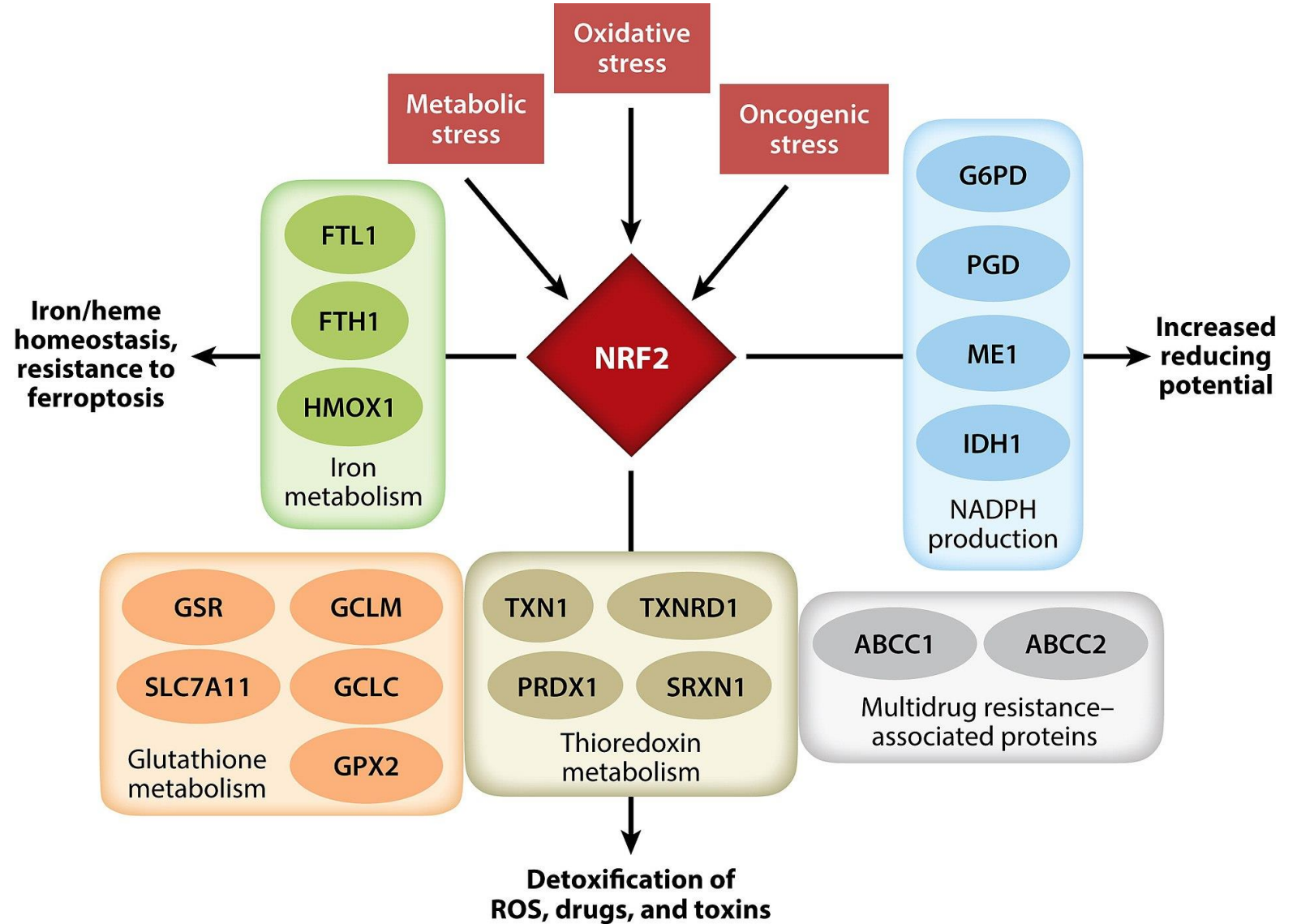
From ligand to transcription factor

From BDNF to NFE2L2 (NRF2) transcription factor

Transcription factor	Gene
p-NFE2L2 [n]	GSR
	GSTA3
	HMOX1
	IDH1
	ME1
	PRDX1
	SRXN1
	TALDO1
	TKT
p-AcK-NFE2L2:CREBBP, EP300 [n]	G6PD
	GCLC
	GCLM
	GSTA1
	NQO1
	PGD
	SLC7A11
	TXNRD1
p-NFE2L2:MAFK [n]	HMOX1

From ligand to transcription factor

From BDNF to NFE2L (NRF2) transcription factor



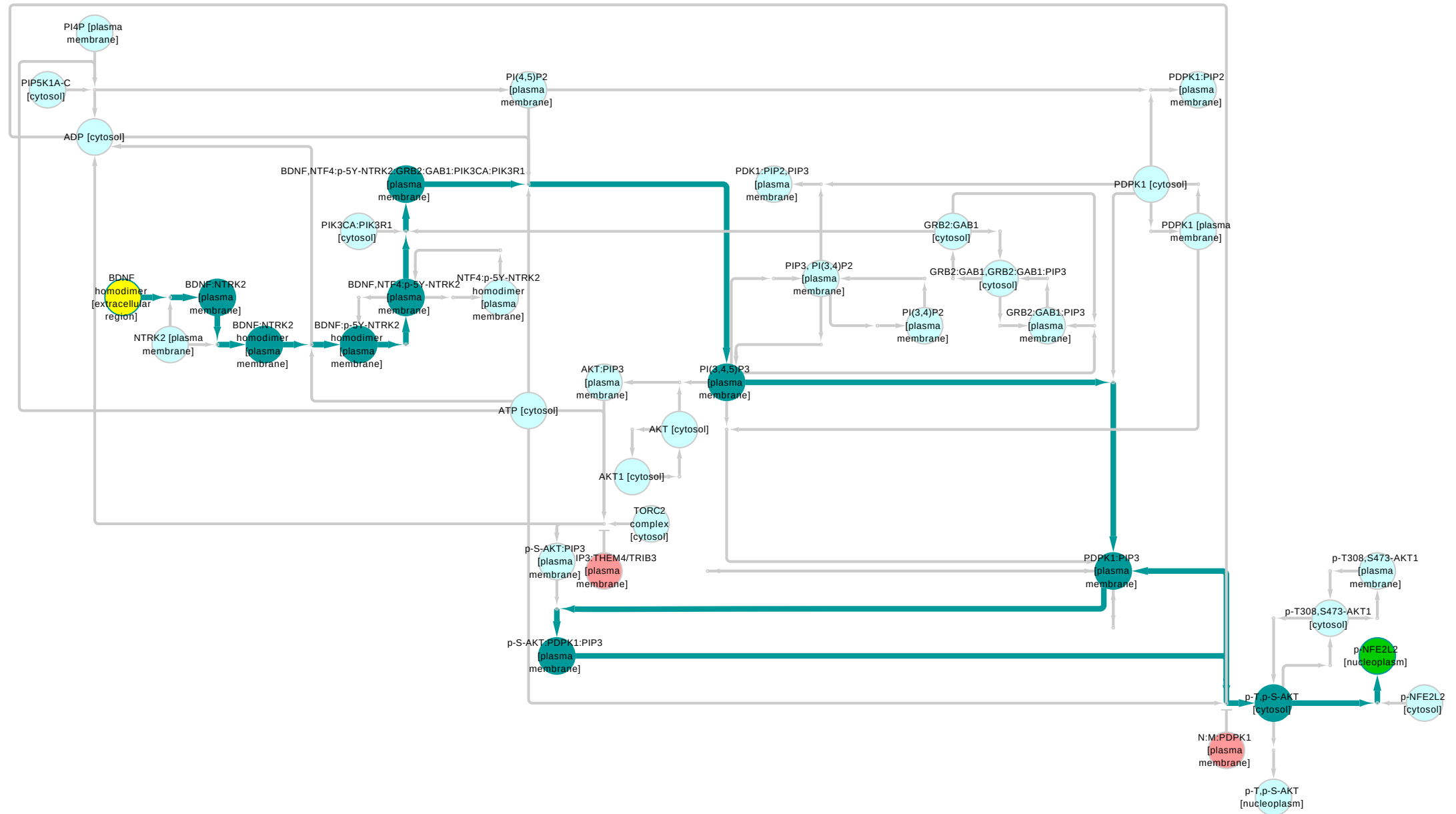
From ligand to transcription factor

Start	Target	Result	Min path
BDNF homodimer [e]	p-NFE2L2 [n]	YES	10
BDNF homodimer [e]	p-AcK-NFE2L2:CREBBP, EP300 [n]	YES	12
BDNF homodimer [e]	p-NFE2L2:MAFK [n]	YES	11

From ligand to transcription factor

Start	Target	Result	Min path
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From ligand to transcription factor



Perspectives

- **Explore signaling-metabolism interaction**

Perspectives

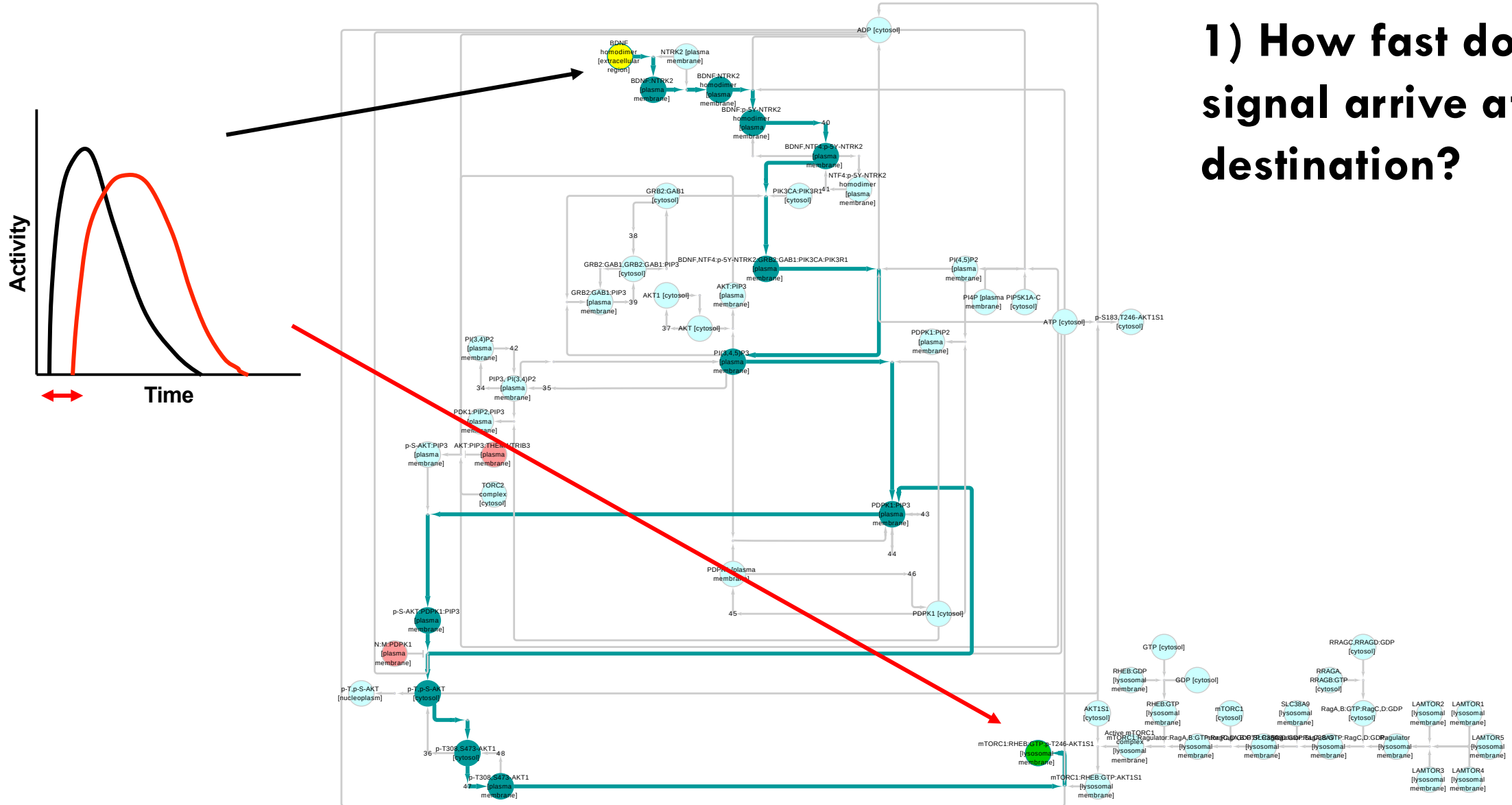
- **Explore signaling-metabolism interaction**
- **Expand multi-omics data integration
(condition-specific signaling models, information
flows deregulation)**

Perspectives

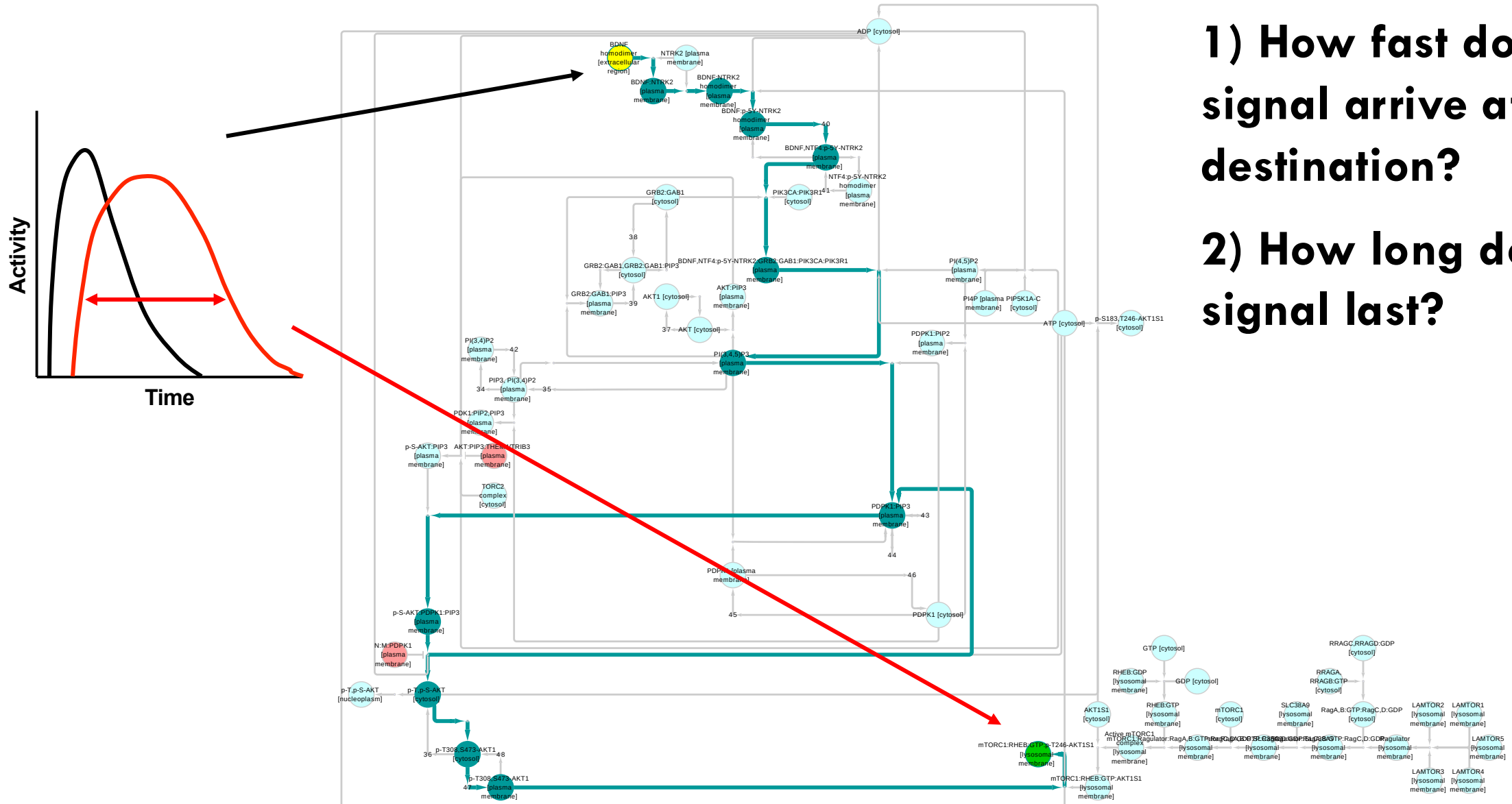
- **Explore signaling-metabolism interaction**
- **Expand multi-omics data integration
(condition-specific signaling models, information
flows deregulation)**
- **Create dynamical signaling models**

Perspectives

Create dynamical signaling models



Create dynamical signaling models

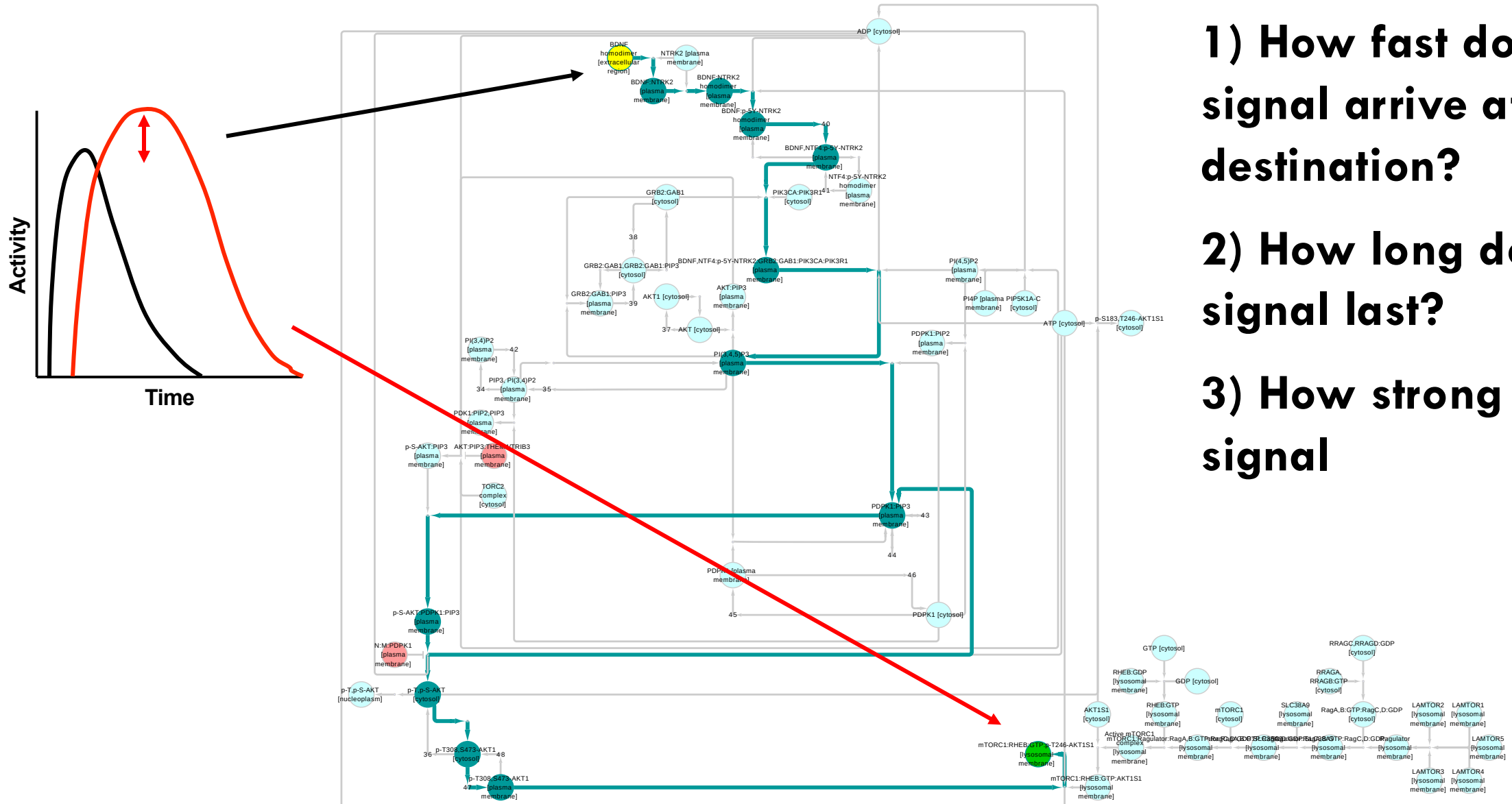


1) How fast does the signal arrive at its destination?

2) How long does the signal last?

Perspectives

Create dynamical signaling models



Acknowledgements



Dr. Maria Masid



David Liaskos



Dr. Omid Oftadeh

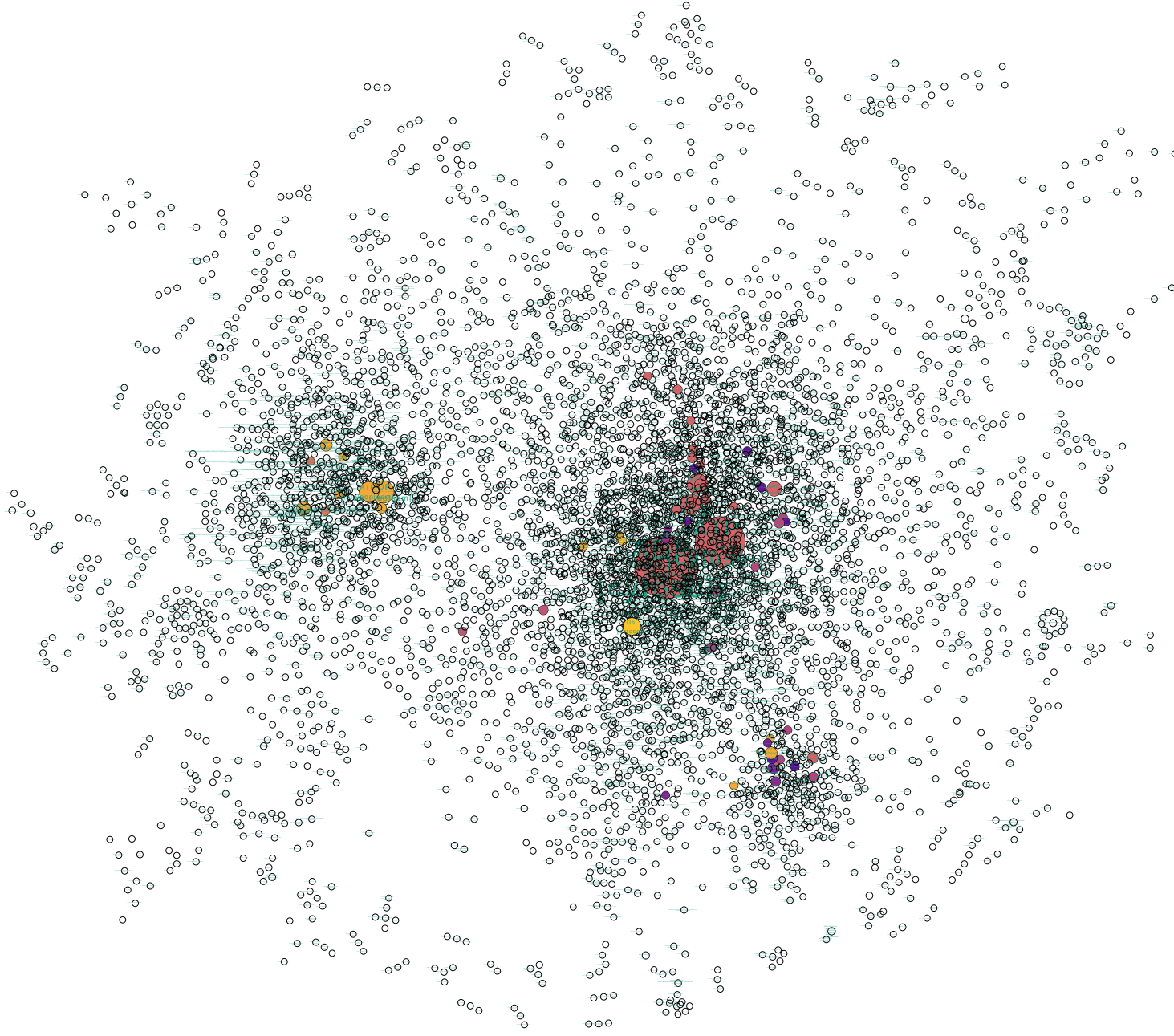


Margherita Tonini

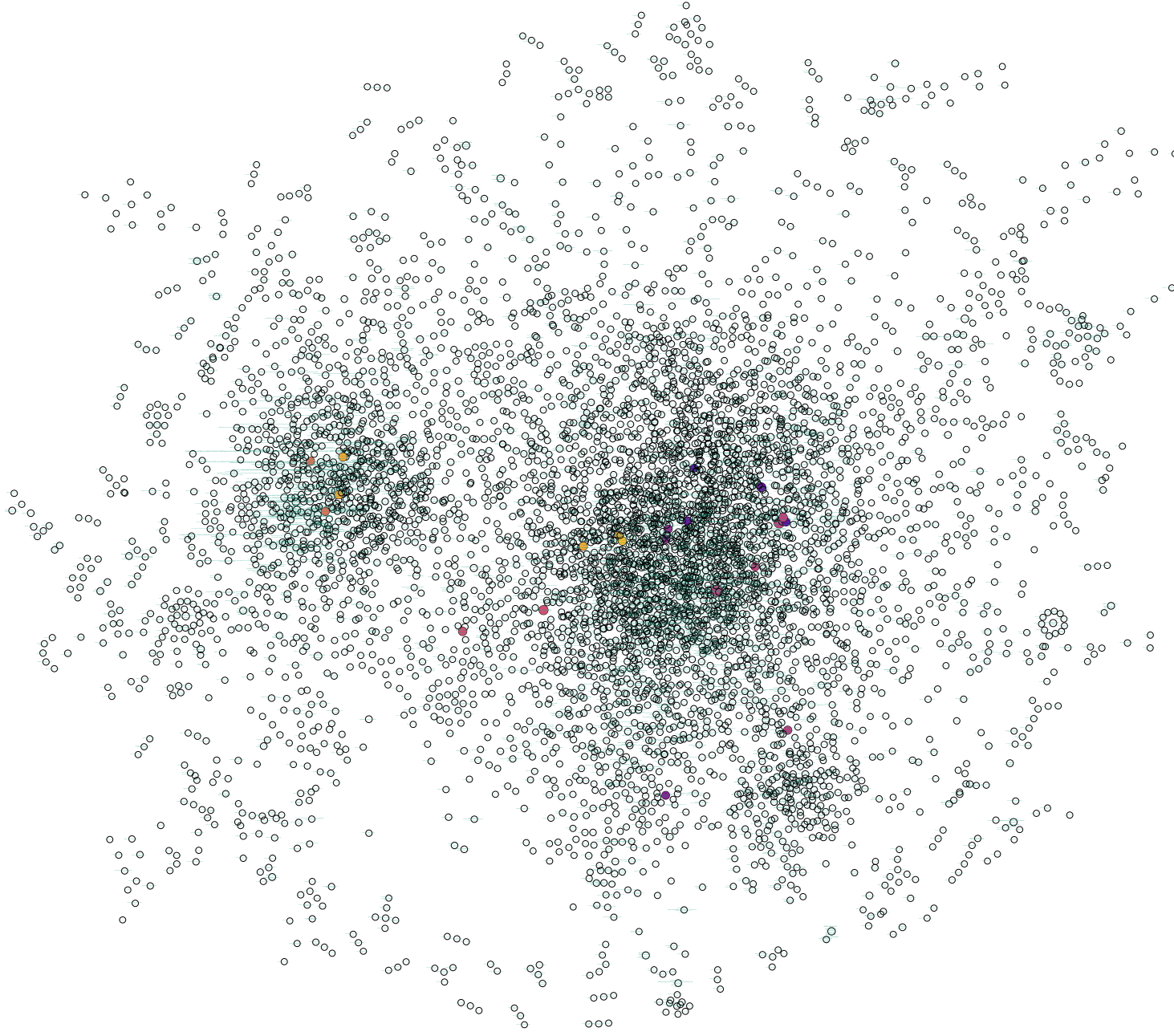


Questions

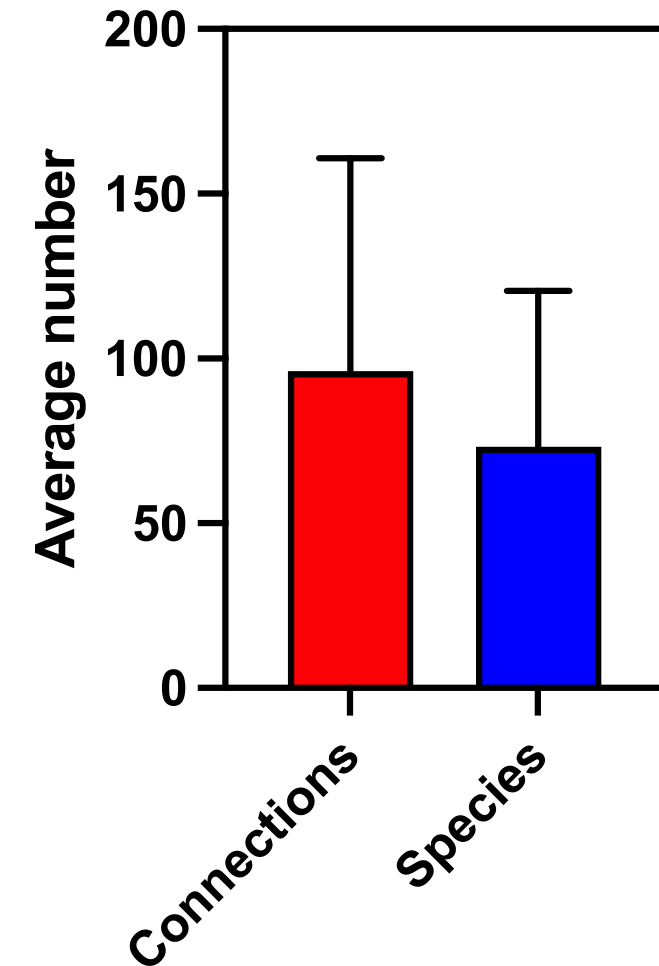
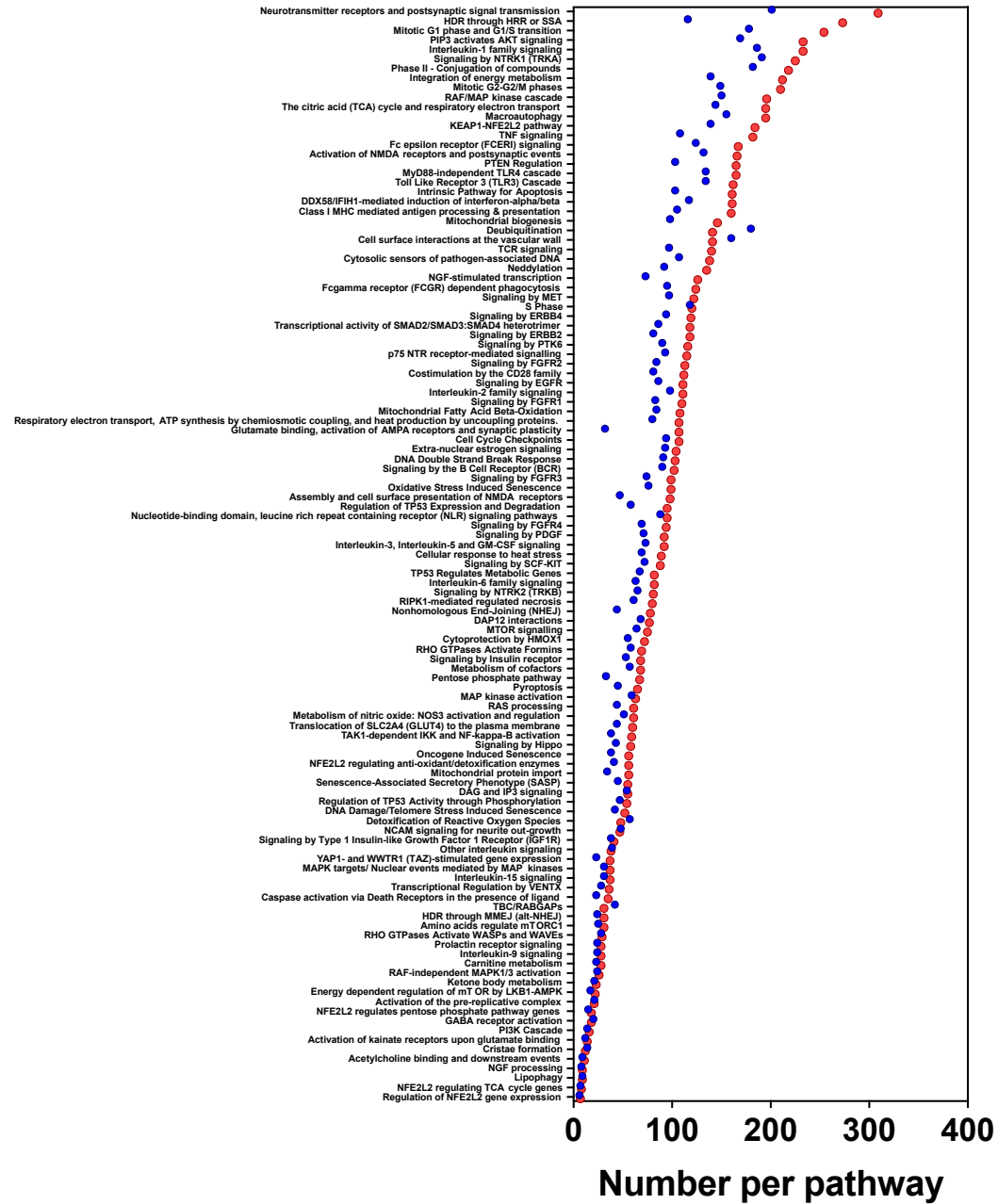
Co-factors identification and removal



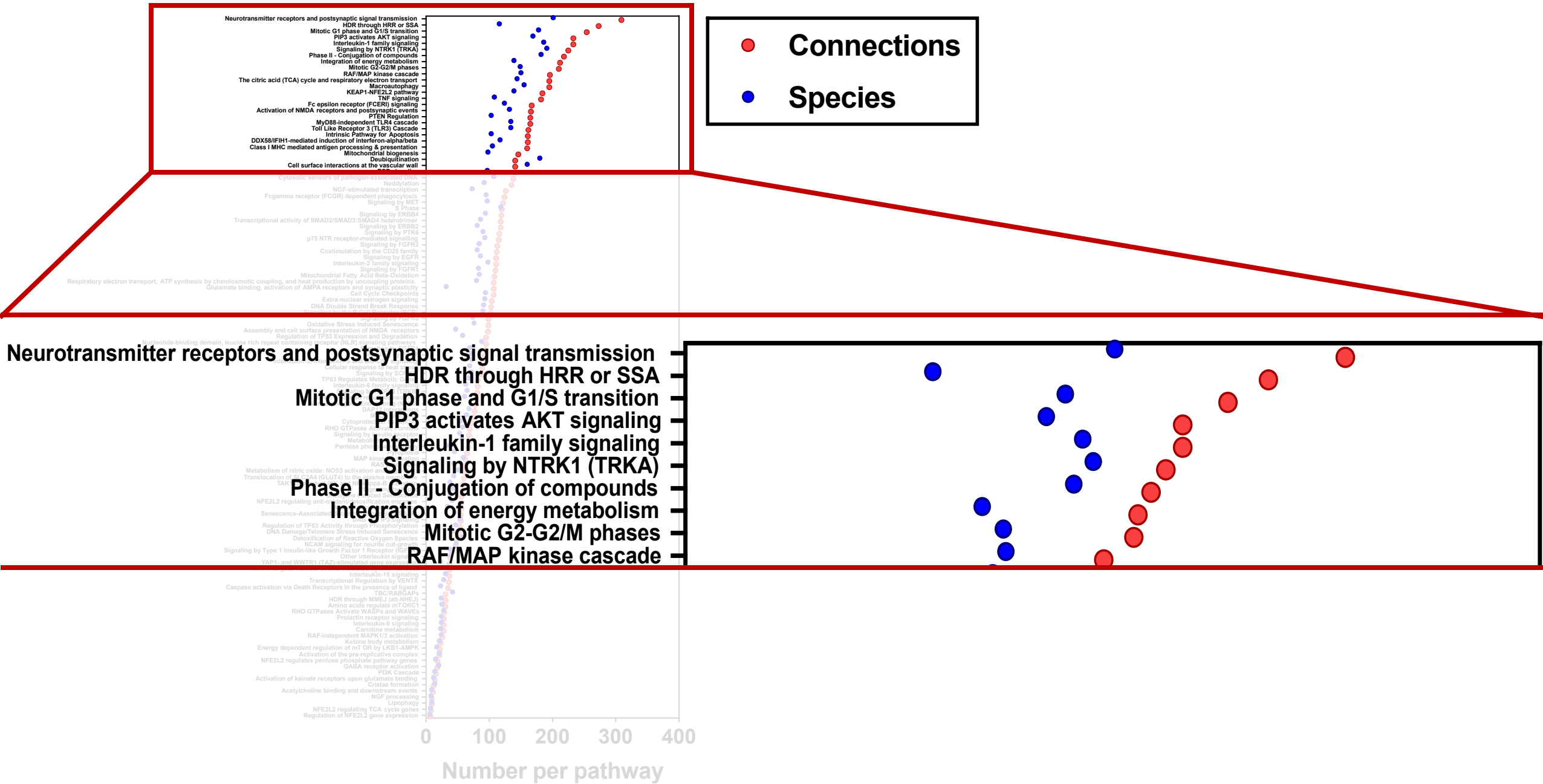
Co-factors identification and removal



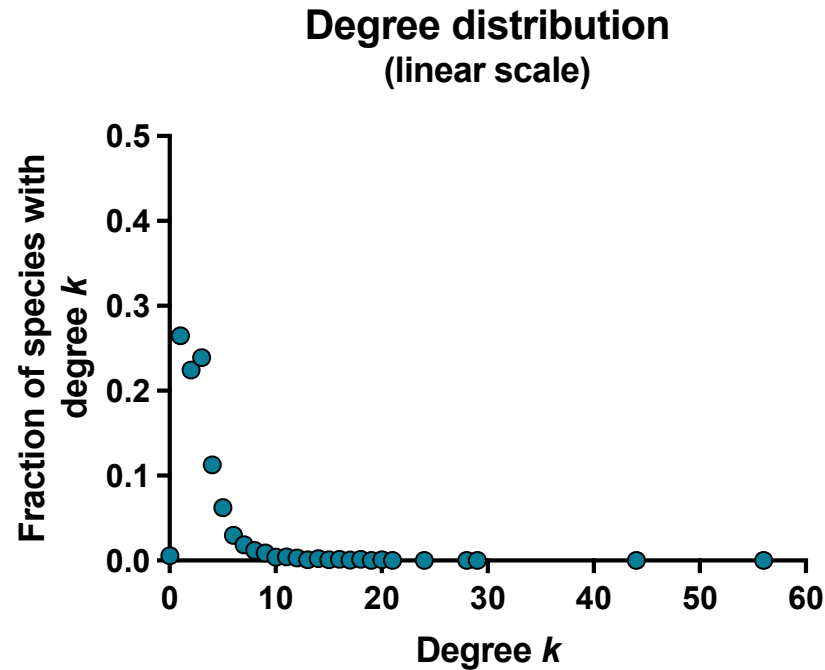
Analysis of the final network of selected pathways



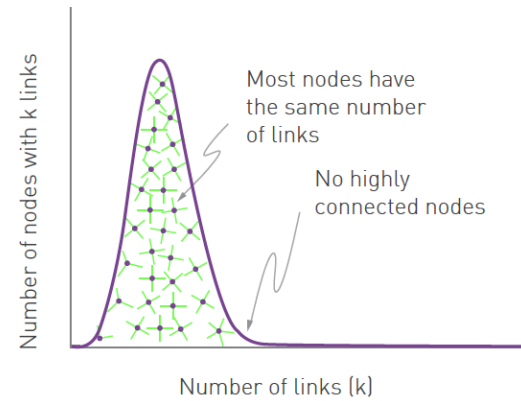
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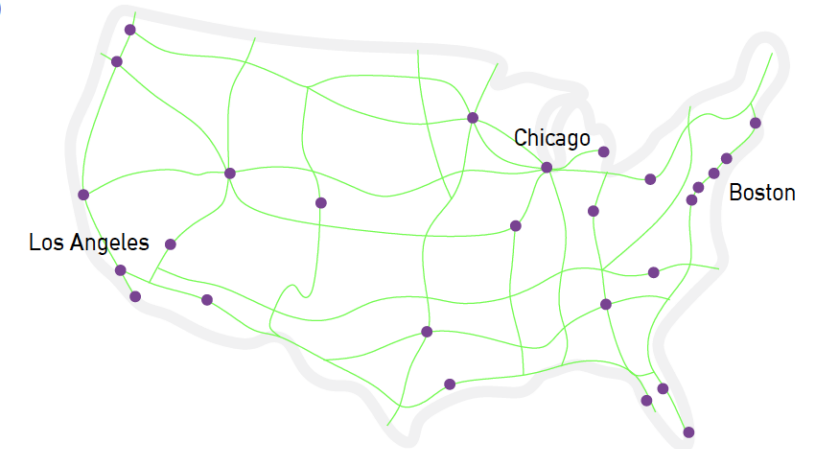
Analysis of the final network of selected pathways



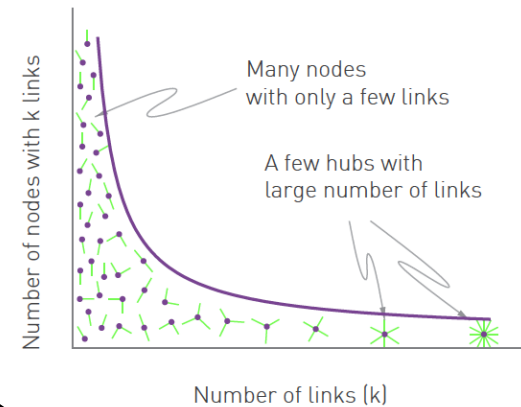
(a) POISSON



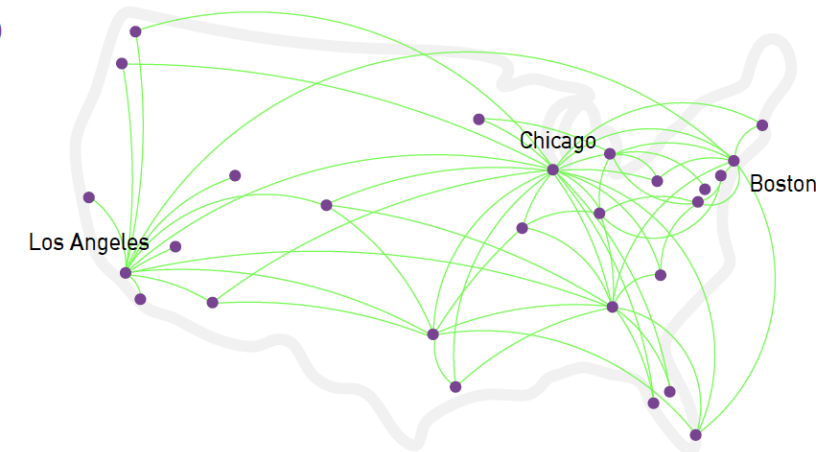
(b)



(c) POWER LAW



(d)

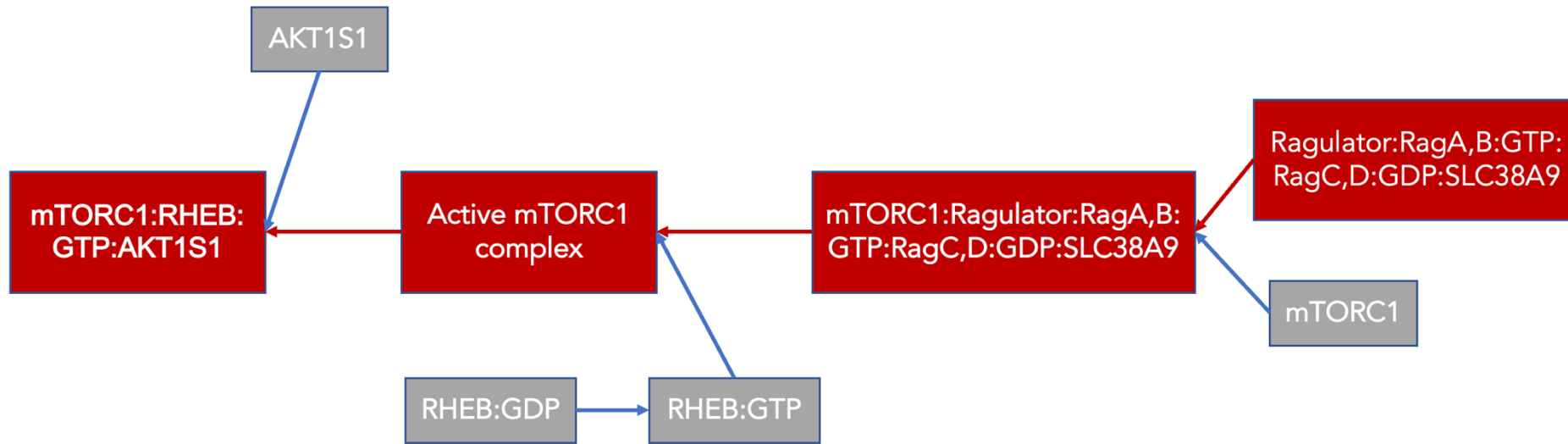


Erdos, Paul, and Renyi, Alfred. "On random graphs I", *Publ. Math. Debrecen*, 1959

Barabási, Albert-László, and Réka Albert. "Emergence of Scaling in Random Networks." *Science*, 1999

Barabási, Albert-László. *Network Science*. United Kingdom: Cambridge University Press, 2016

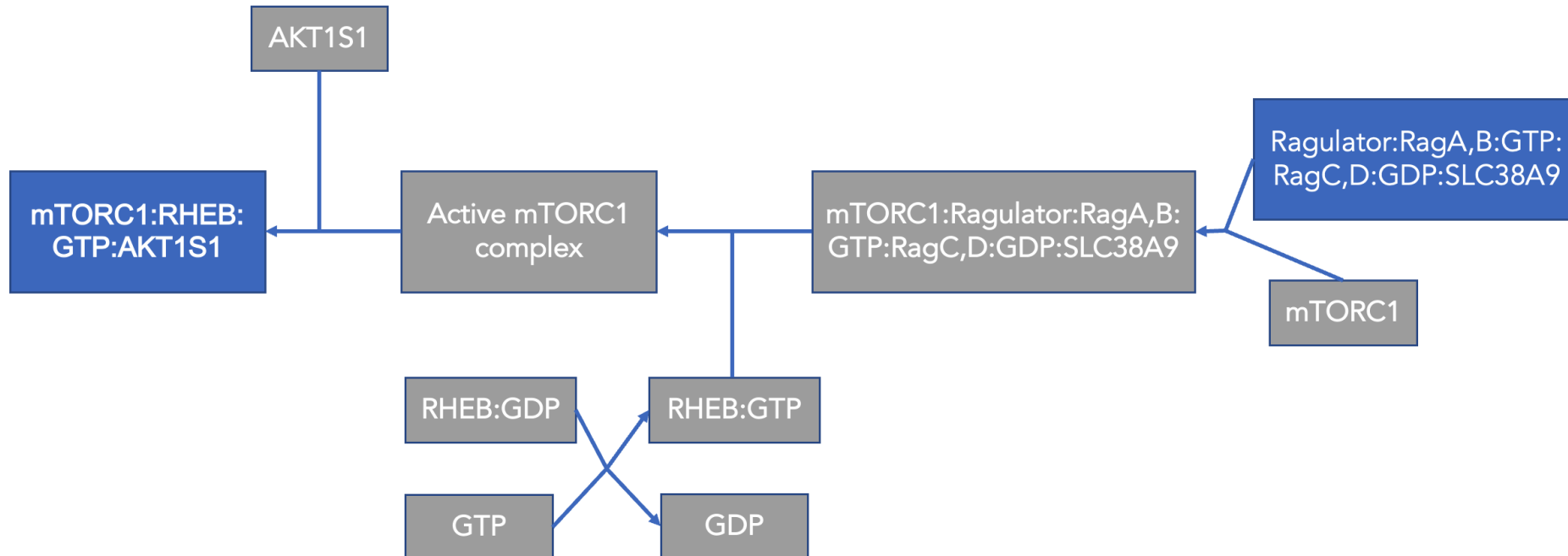
Method for generating balanced signaling networks



8 species
7 connections

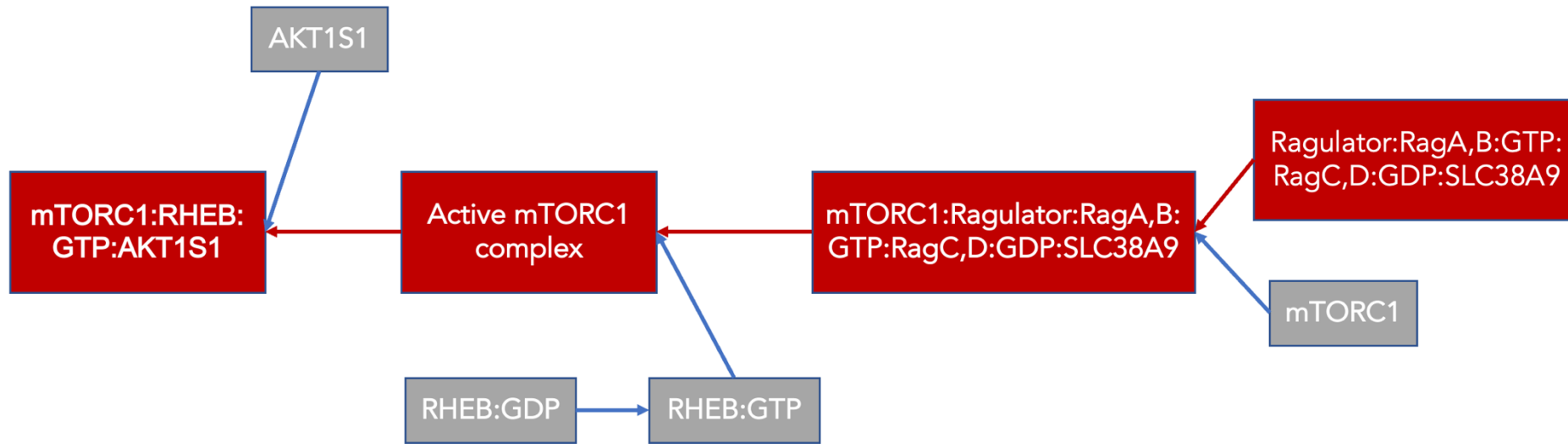
Minimal path

4 species
3 connections



10 species
4 reactions

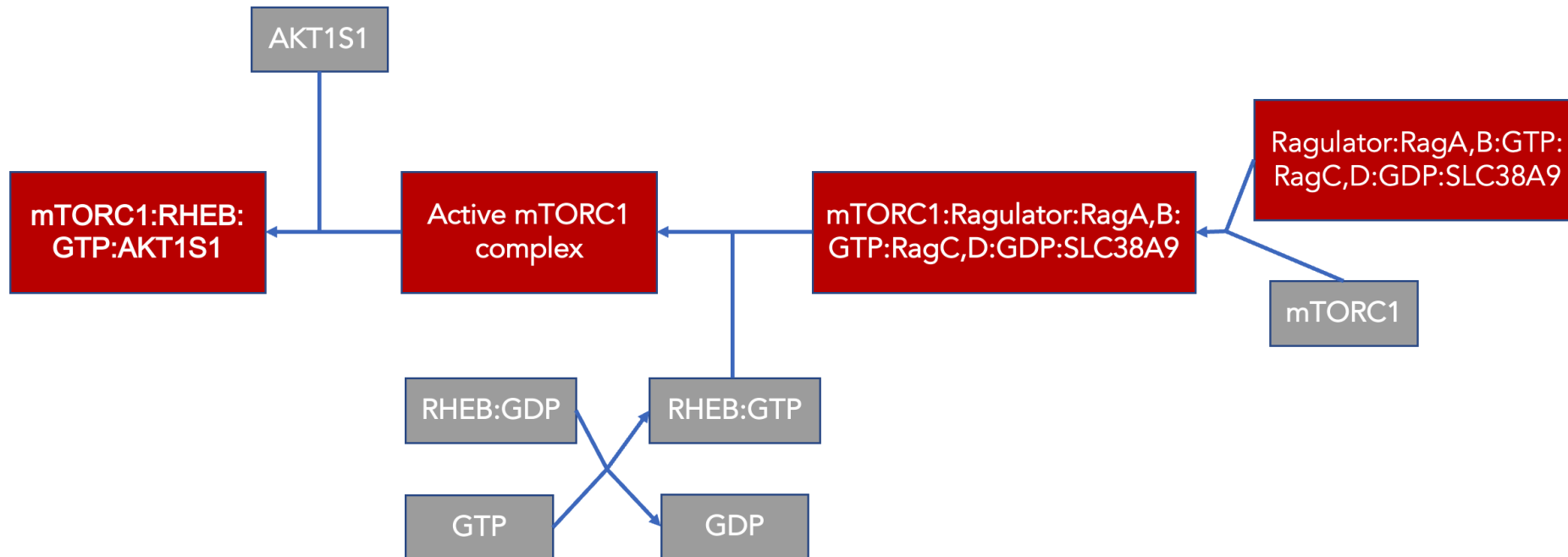
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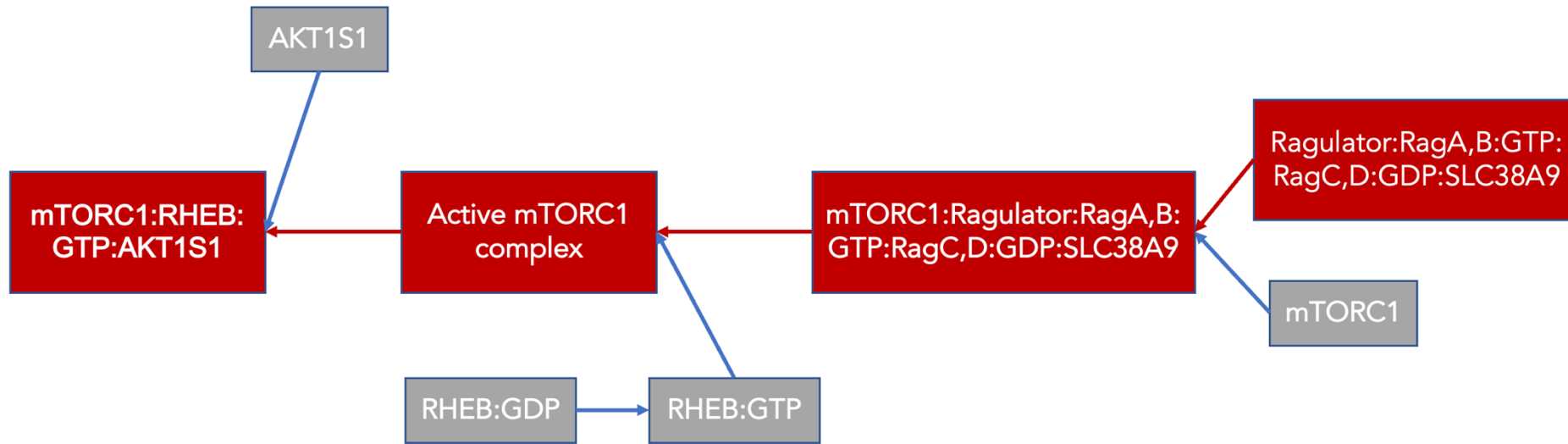
Minimal path

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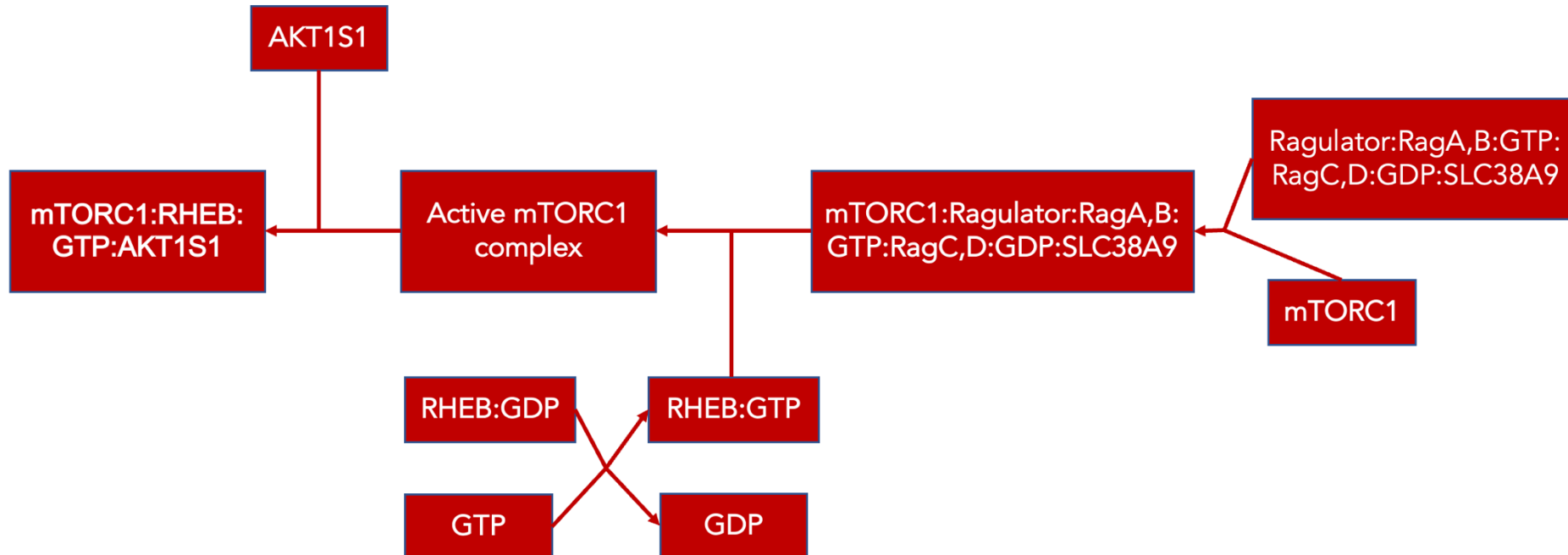
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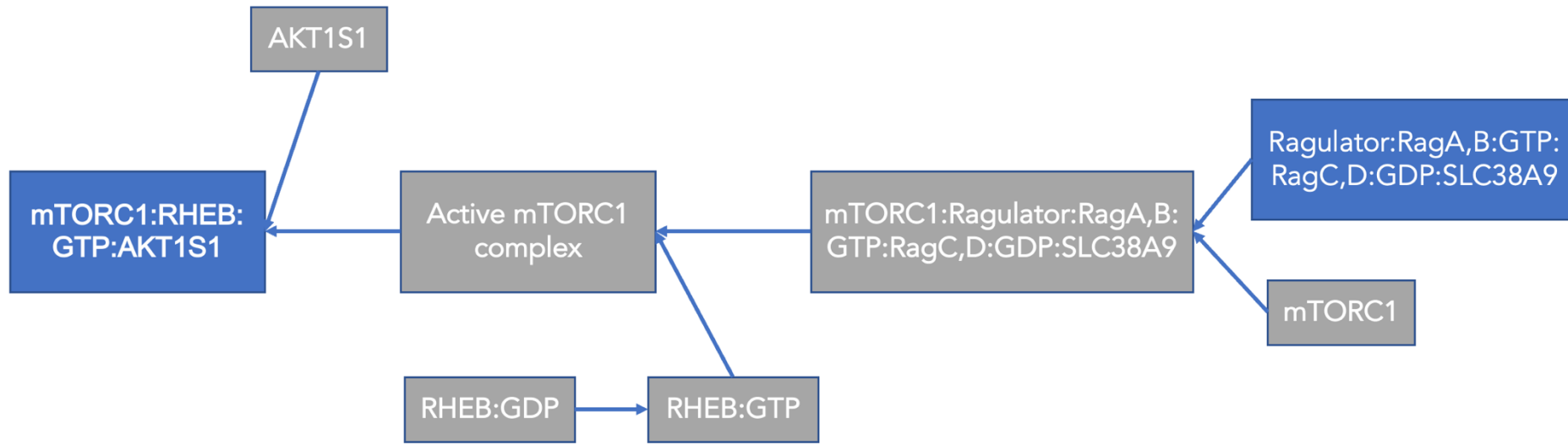


10 species
4 reactions

Balanced path

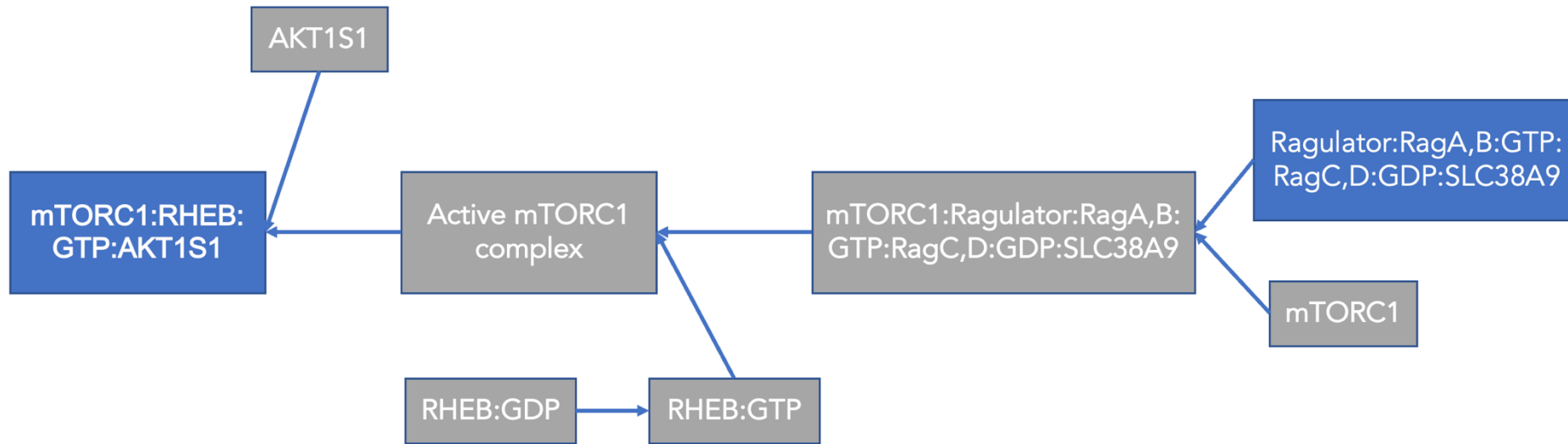
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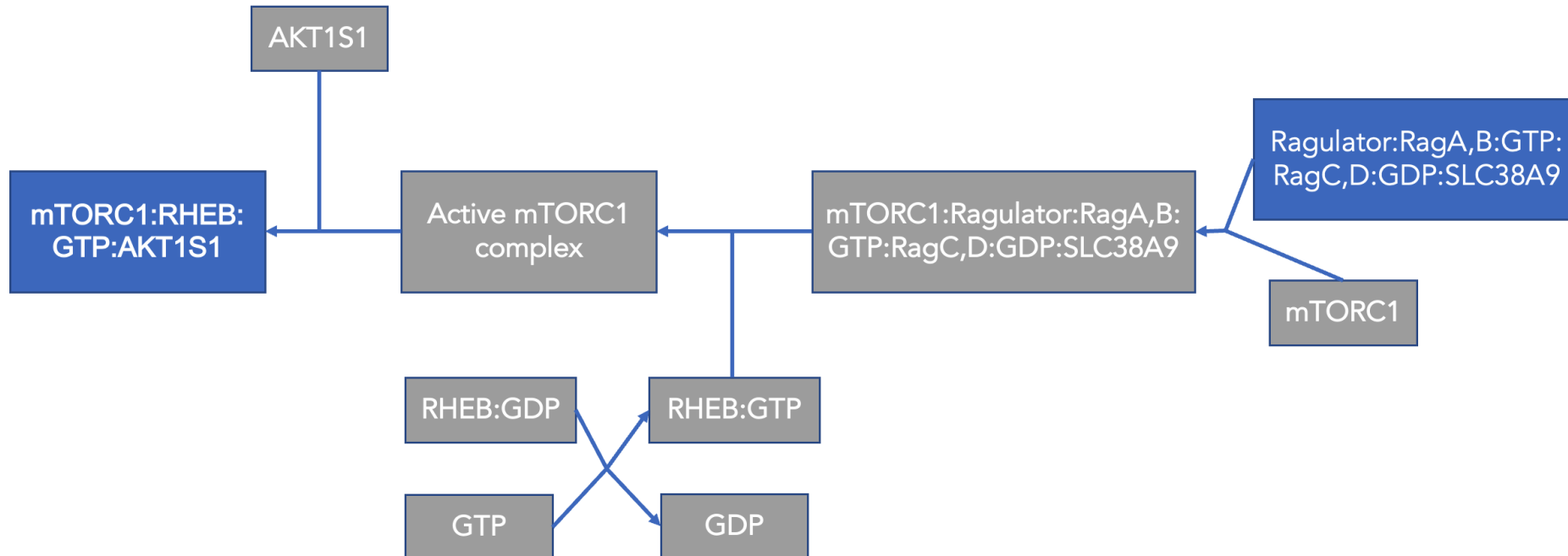


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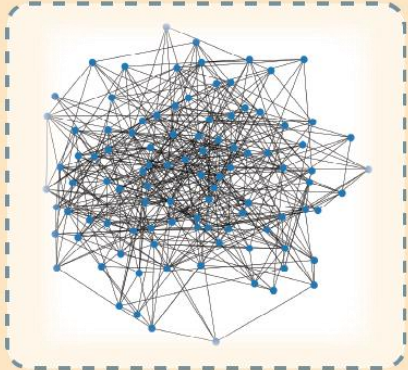
10 species
4 reactions



**Add co-factors
back to balance
our network.**

Study deregulation in signaling pathways

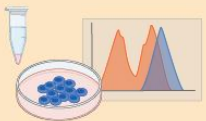
SIGMA Method



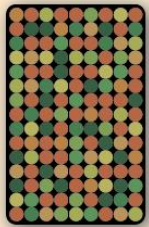
reactome

Signaling network

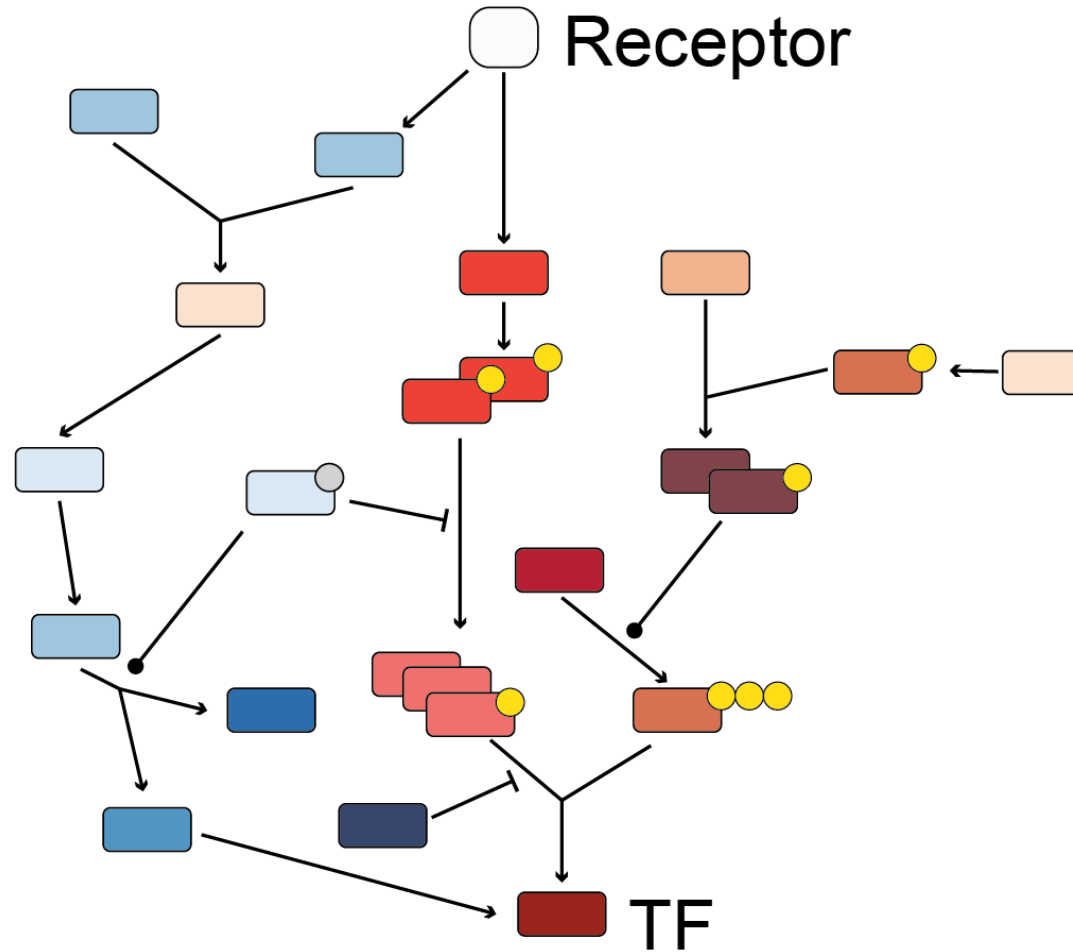
omics data



(sc)RNA-seq data

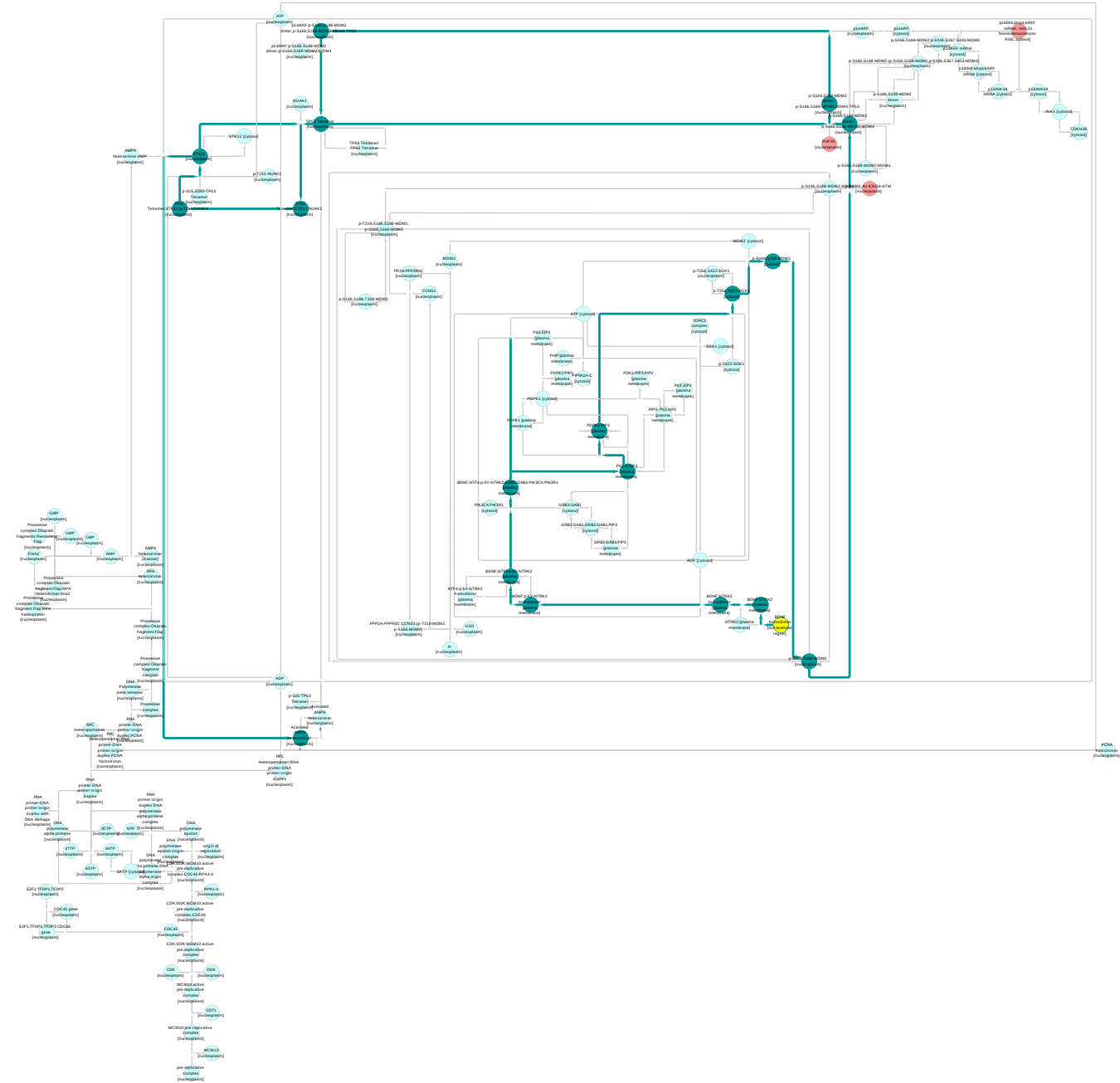


phosphoproteomics



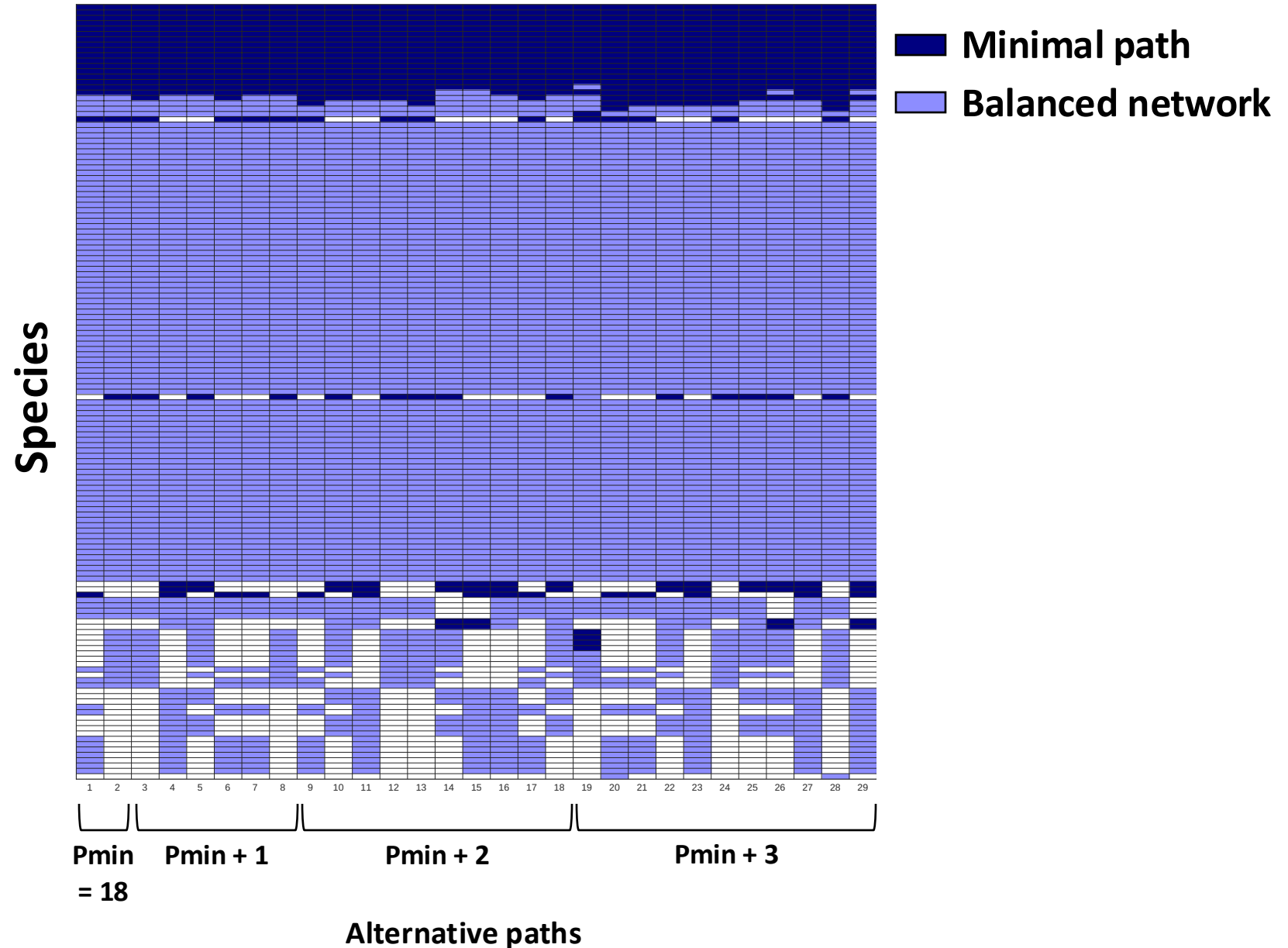
enrichment analysis

Studying BDNF connection to AMPK



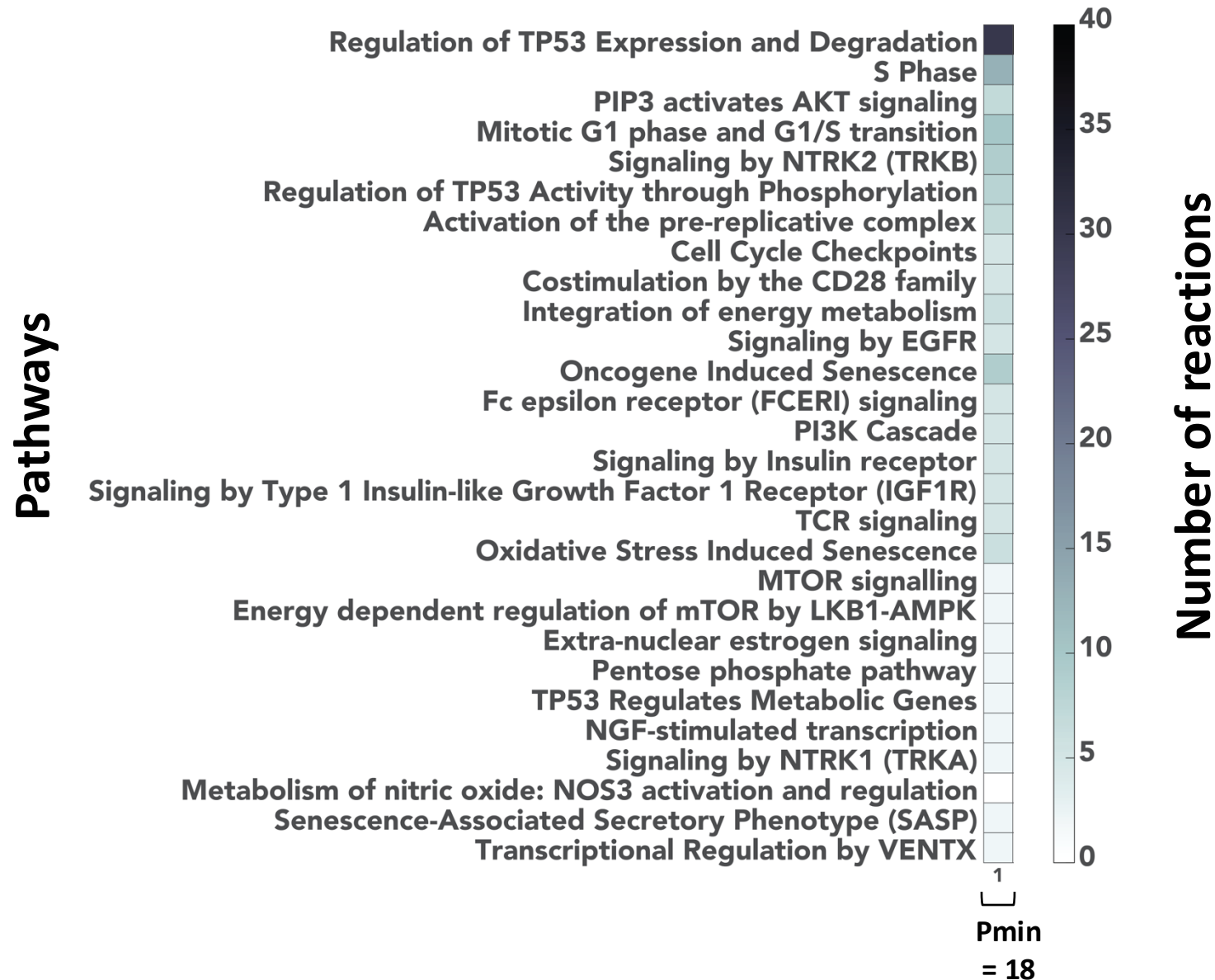
Studying BDNF connection to AMPK

BDNF to Activated AMPK heterotrimer



Studying BDNF connection to AMPK

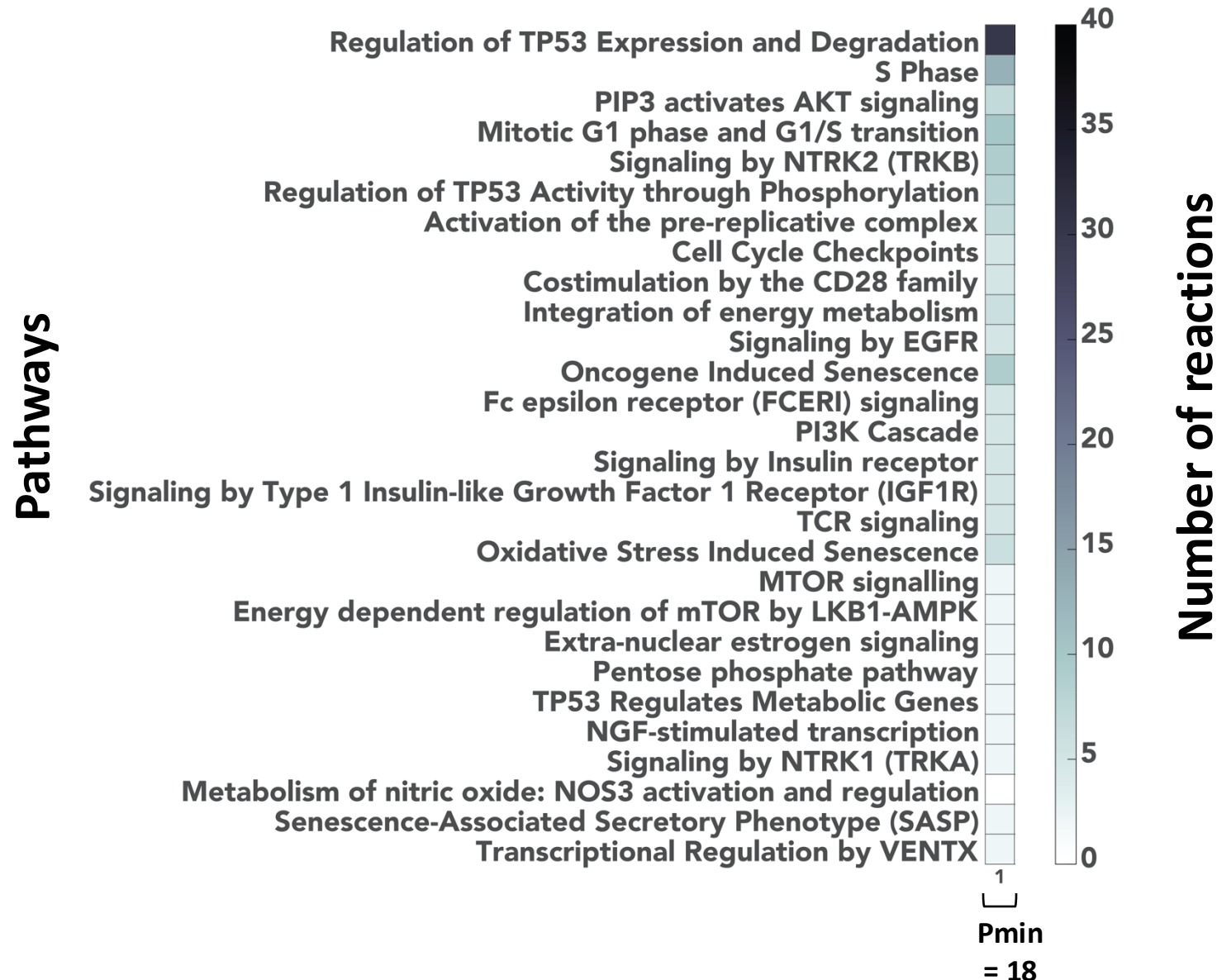
BDNF to Activated AMPK heterotrimer



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Studying BDNF connection to AMPK

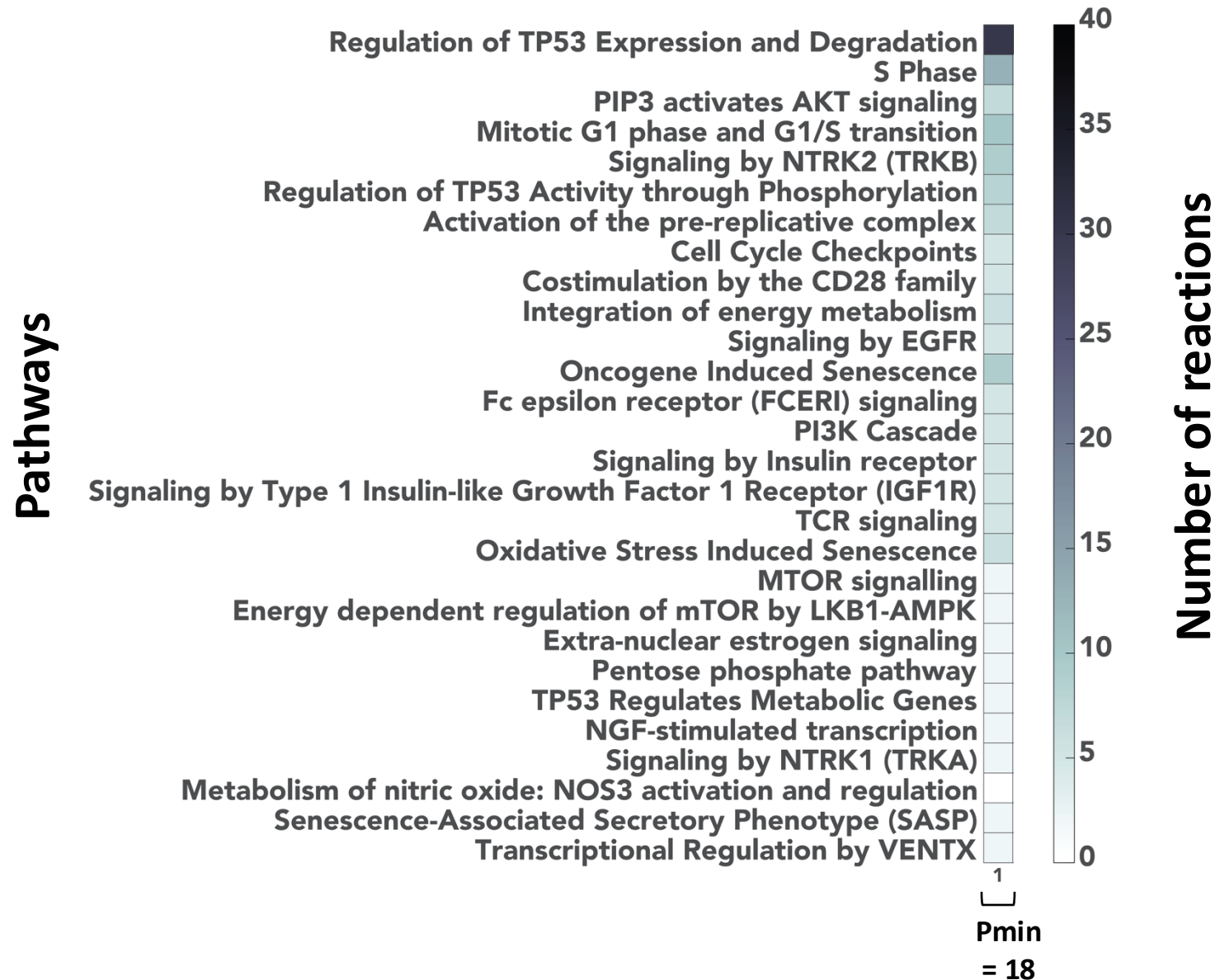
BDNF to Activated AMPK heterotrimer



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- Total of 104 reactions (including the peripheral)

Studying BDNF connection to AMPK

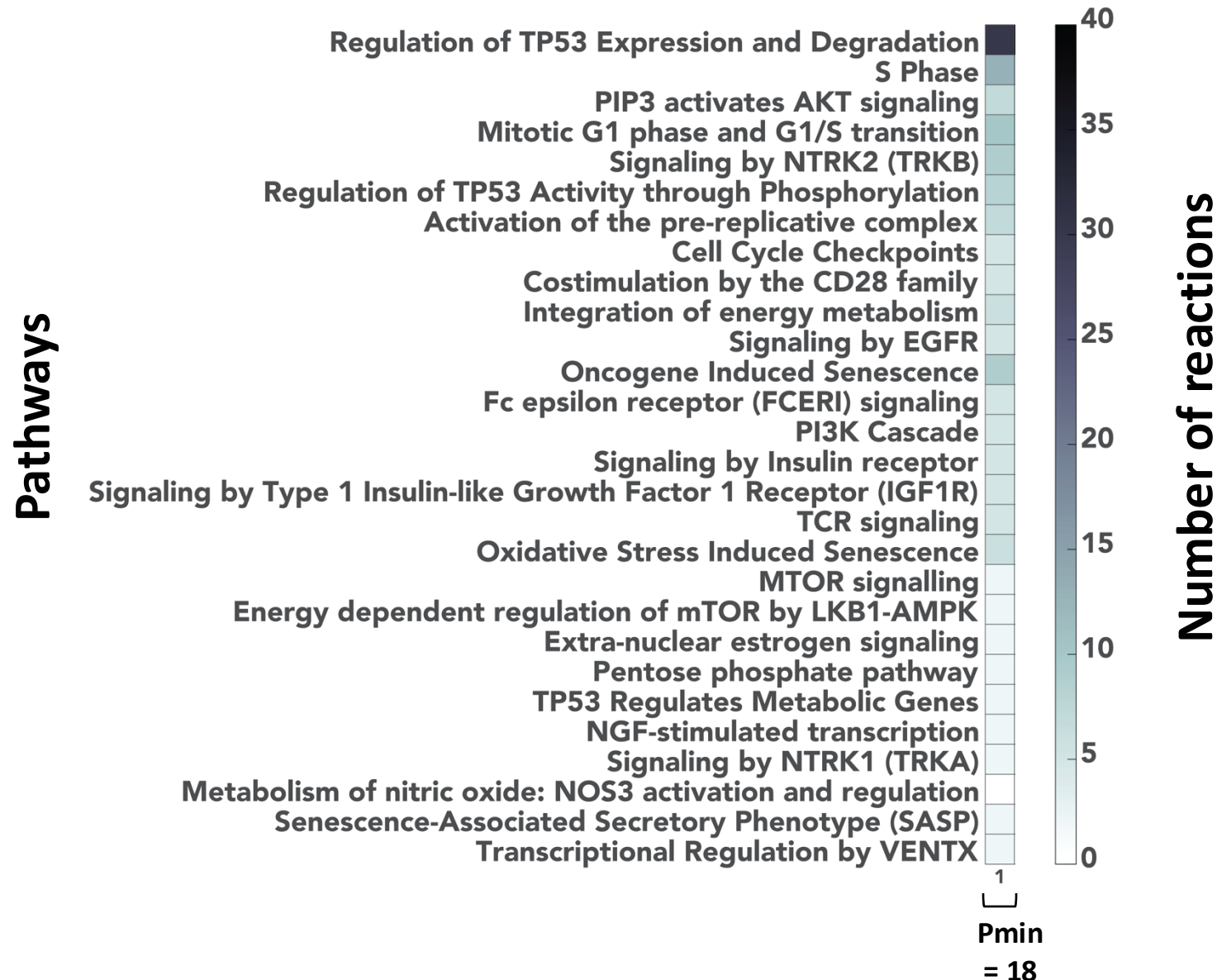
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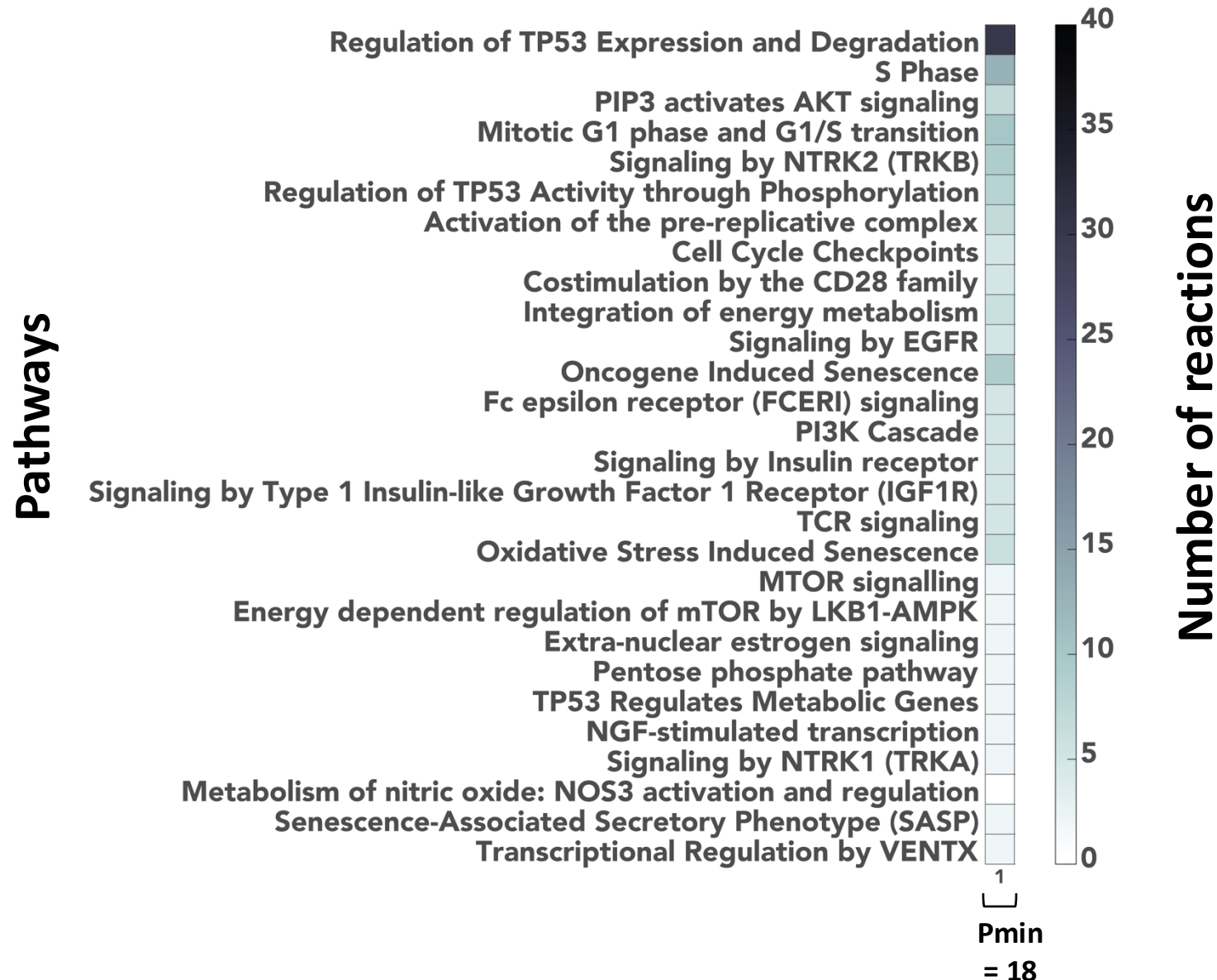
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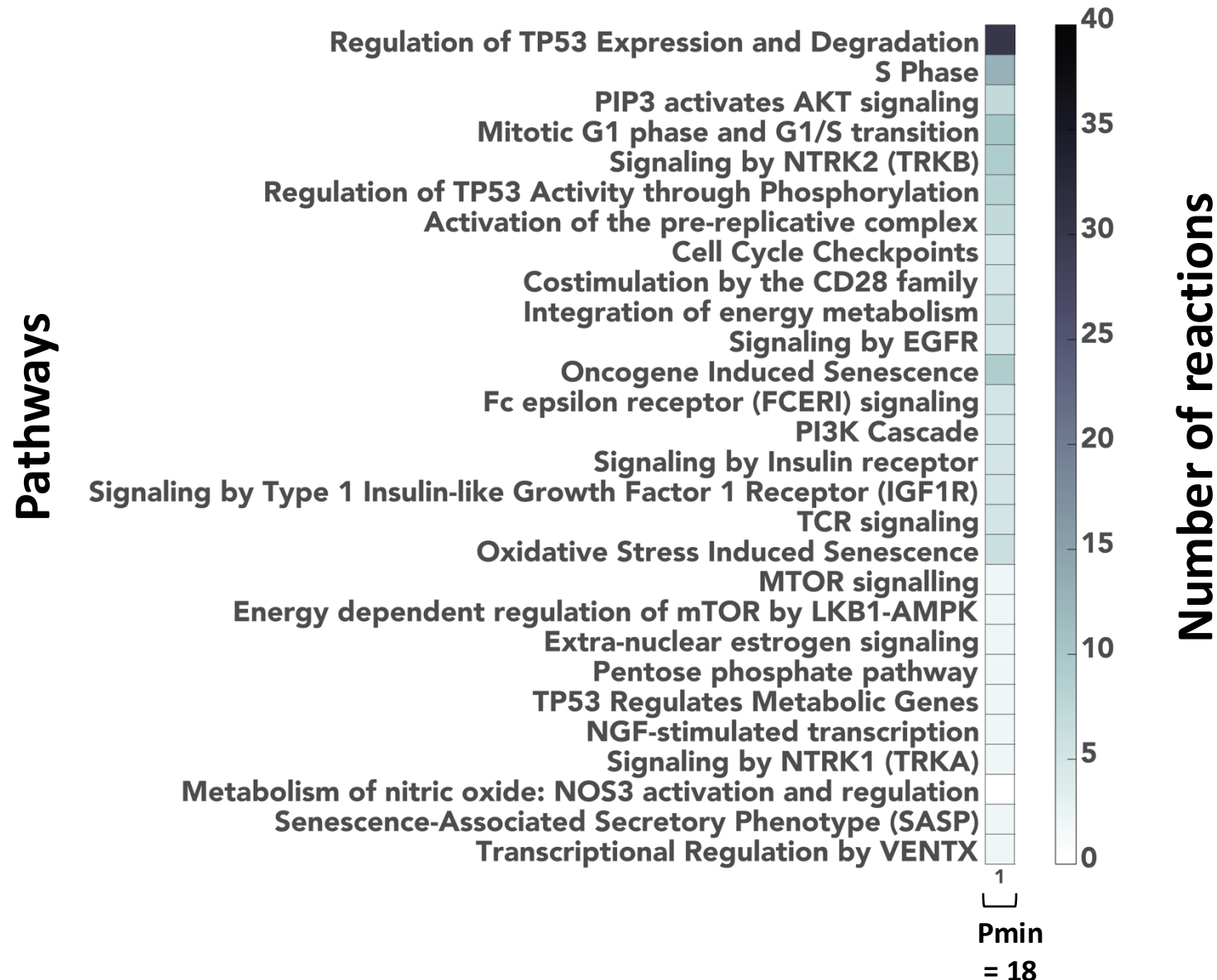
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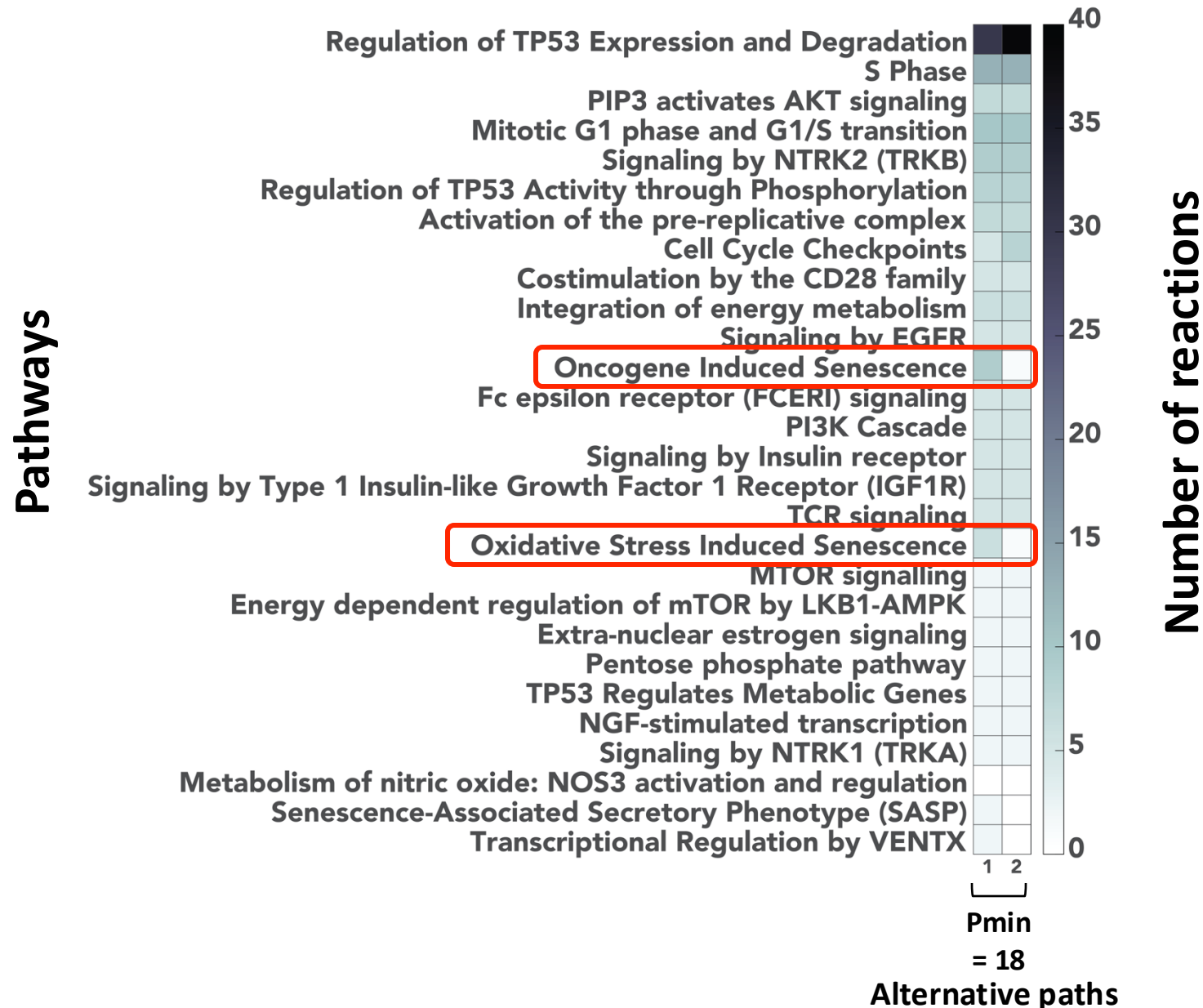
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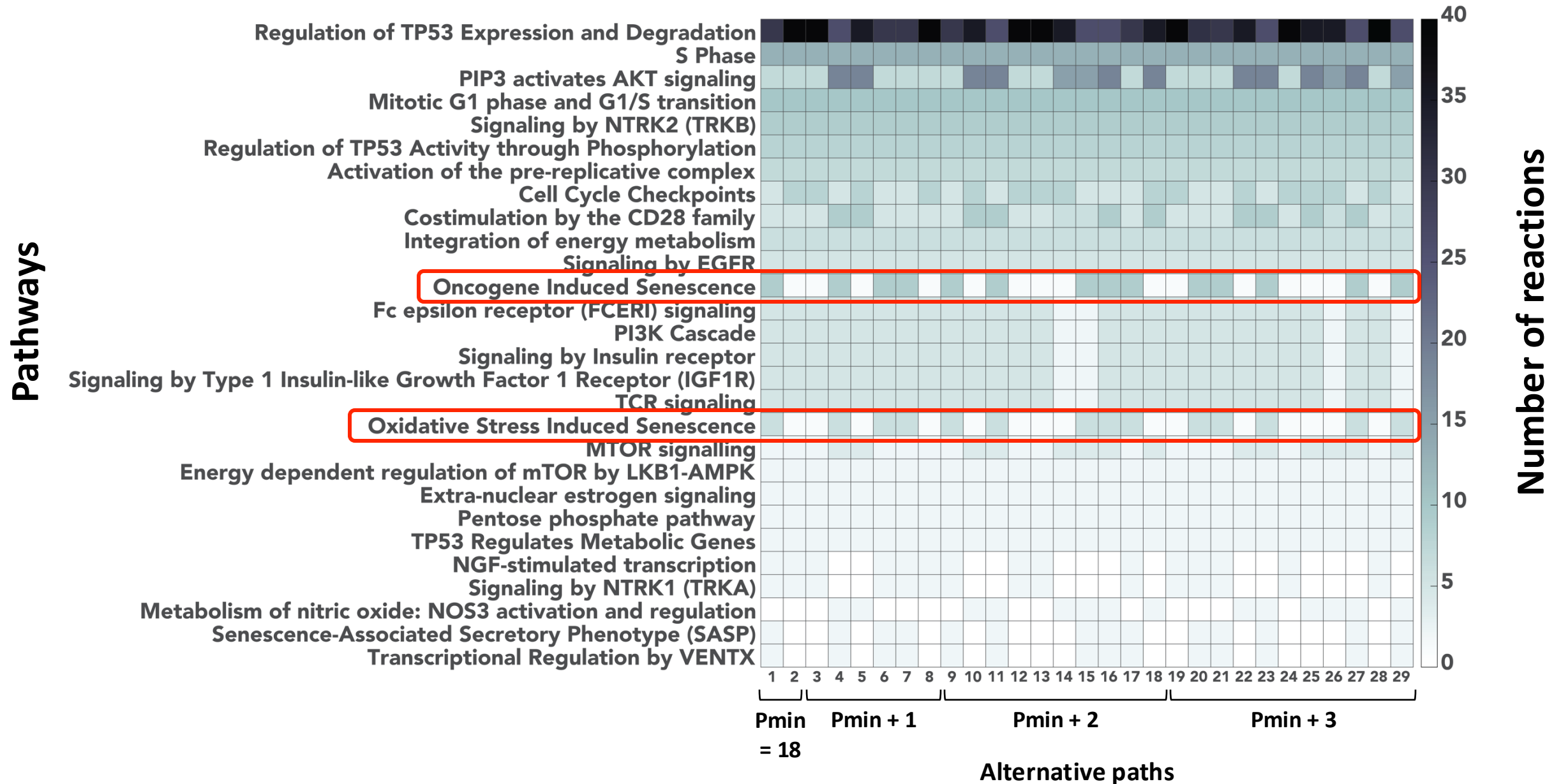
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Studying BDNF connection to AMPK

BDNF to Activated AMPK heterotrimer



BDNF to Activated AMPK heterotrimer



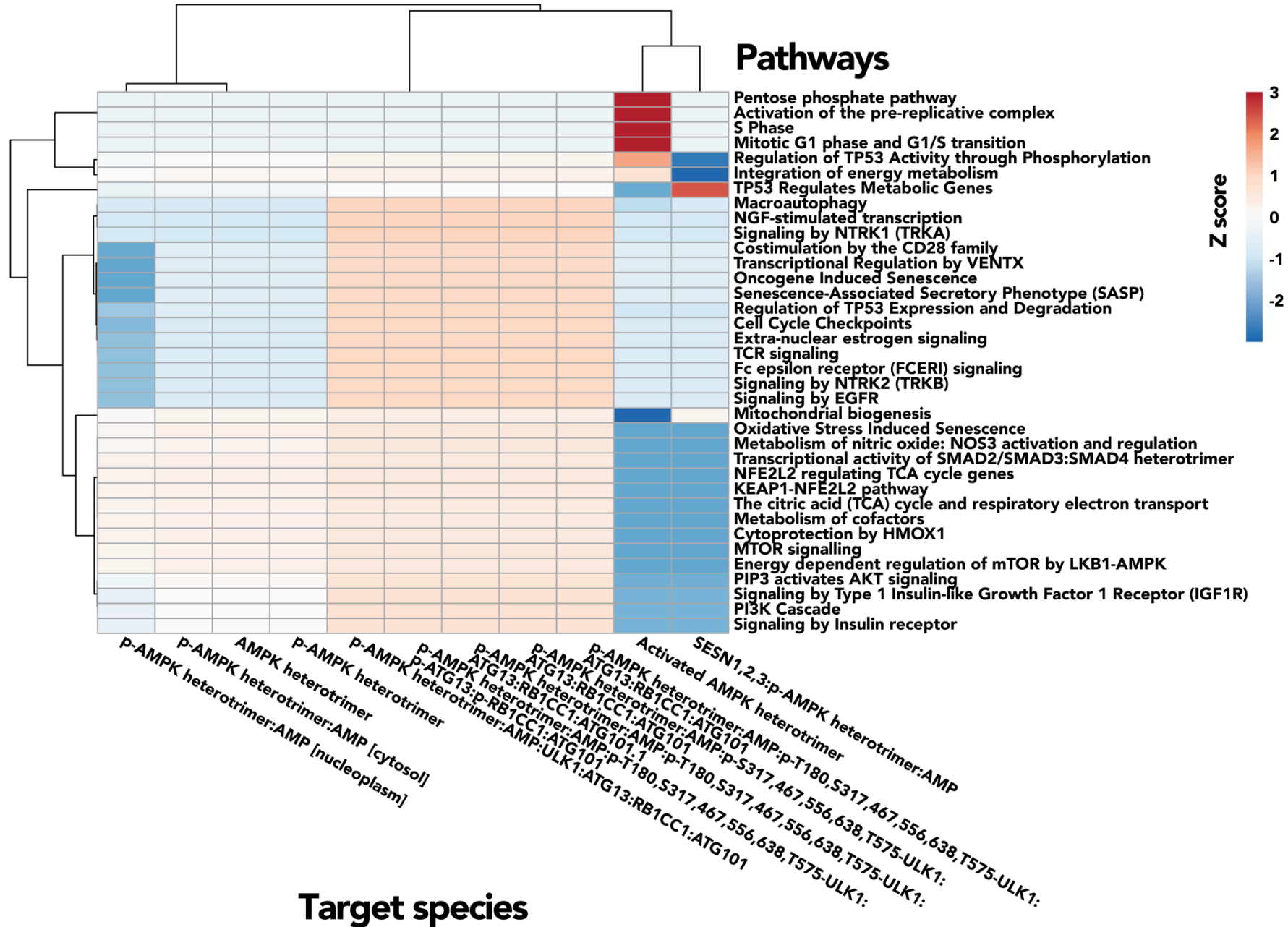
Pathways

- Pentose phosphate pathway
- Activation of the pre-replicative complex
- S Phase
- Mitotic G1 phase and G1/S transition
- Regulation of TP53 Activity through Phosphorylation
- Integration of energy metabolism
- TP53 Regulates Metabolic Genes
- Macroautophagy
- NGF-stimulated transcription
- Signaling by NTRK1 (TRKA)
- Costimulation by the CD28 family
- Transcriptional Regulation by VENTX
- Oncogene Induced Senescence
- Senescence-Associated Secretory Phenotype (SASP)
- Regulation of TP53 Expression and Degradation
- Cell Cycle Checkpoints
- Extra-nuclear estrogen signaling
- TCR signaling
- Fc epsilon receptor (FCER1) signaling
- Signaling by NTRK2 (TRKB)
- Signaling by EGFR
- Mitochondrial biogenesis
- Oxidative Stress Induced Senescence
- Metabolism of nitric oxide: NOS3 activation and regulation
- Transcriptional activity of SMAD2/SMAD3:SMAD4 heterotrimer
- NFE2L2 regulating TCA cycle genes
- KEAP1-NFE2L2 pathway
- The citric acid (TCA) cycle and respiratory electron transport
- Metabolism of cofactors
- Cytoprotection by HMOX1
- MTOR signalling
- Energy dependent regulation of mTOR by LKB1-AMPK
- PIP3 activates AKT signaling
- Signaling by Type 1 Insulin-like Growth Factor 1 Receptor (IGF1R)
- PI3K Cascade
- Signaling by Insulin receptor

Z score

Target species

- p-AMPK heterotrimer:AMP [nucleoplasm]
- p-AMPK heterotrimer:AMP [cytosol]
- AMPK heterotrimer
- p-AMPK heterotrimer
- p-AMPK heterotrimer
- p-ATG13:p-RB1CC1:ATG101
- p-AMPK heterotrimer:AMP
- p-ATG13:p-RB1CC1:ATG101
- p-AMPK heterotrimer:AMP
- p-T180,S317,467,556,638,T575-ULK1:
- Activated AMPK heterotrimer
- SESN1,2,3:p-AMPK heterotrimer



[illegible]