

THE DEVELOPMENT OF COX2 INHIBITORS

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Aspirin, arguably the world's favourite drug, has been around since the late nineteenth century, but it wasn't until the late 1970s that its ability to inhibit prostaglandin production by the cyclooxygenase enzyme was identified as the basis of its therapeutic action. Early hints of a second form of the cyclooxygenase that was differentially sensitive to other aspirin-like drugs ultimately ushered in an exciting era of drug discovery, culminating in the introduction of an entirely new generation of anti-inflammatories. This article reviews the story of this discovery and looks at the future of cyclooxygenase pharmacology.

CASE HISTORY

ISOZYME (ISOENZYME)
One of several forms of an enzyme in an individual or population that catalyse the same reaction but differ from each other in such properties as substrate affinity and maximum rates of enzyme-substrate reaction.

ANTIPYRETIC
Describes the fever-suppressive activity of a drug.

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The league table listing the 'top 20' drugs includes rofecoxib (Vioxx) and celecoxib (Celebrex), two inhibitors of the prostaglandin-forming cyclooxygenase (COX) (FIG. 1), which between them commanded sales in excess of US \$4 billion in the year 2000 (REF. 1). This statistic might seem surprising — after all, the therapeutic use of COX inhibitors has a venerable history dating back to the introduction of aspirin in 1898 (or even earlier if the use of salicylate-containing plant extracts is included) and, since then, the field has a record of almost continuous development. The 1940s saw the introduction of phenylbutazone, the fenamates appeared in the 1950s, indomethacin in the 1960s, the propionates in the 1970s and the oxicams in the 1980s. With such a long record of drug discovery in the area and such a vast range of drugs to choose from, one might be forgiven for thinking that the wellspring of chemical innovation that nurtured the field so efficiently over the past century must have long since run dry — or at least begun to falter — and that the current market would have little room for new versions of what would seem to be a rather tired and well-worn formula.

However, the discovery in the early 1990s of a second COX ISOZYME revitalized the field and stimulated a hunt for new and selective isoform inhibitors. This culminated in the introduction of Vioxx and Celebrex in Europe and North America within a mere ten years, and, at the same time, brought a fresh perspective on the

unusual therapeutic profile of several existing non-steroidal anti-inflammatory drugs (NSAIDs). But we are getting ahead of ourselves — how did this idea of the second isoform come about in the first place and what are the therapeutic advantages of these new inhibitors over the many older drugs which, after all, have seen valiant clinical service over the decades? To answer these questions, we must return to the 1970s when much of the seminal work on COX inhibition was published.

Early work

Although the NSAIDs do not reverse the course of systemic diseases such as arthritis, they form the mainstay symptomatic treatment of many inflammatory disorders and soft-tissue injuries. Approximately 50 NSAID preparations are listed in *Monthly Index of Medical Specialities* and, as a class, these are among the most commonly prescribed drugs. In the United Kingdom, for example, recent data (1999) indicate that 18.5 million prescriptions are written for NSAIDs each year at a cost of £170 million — and this figure doesn't take into account the over-the-counter sales, which are considerable. Aspirin itself is still consumed in prodigious amounts around the world and new uses are continually being found for this drug.

NSAIDs are sometimes known as the aspirin-like drugs because they have an activity profile that is broadly similar to that of aspirin. That is, they all possess

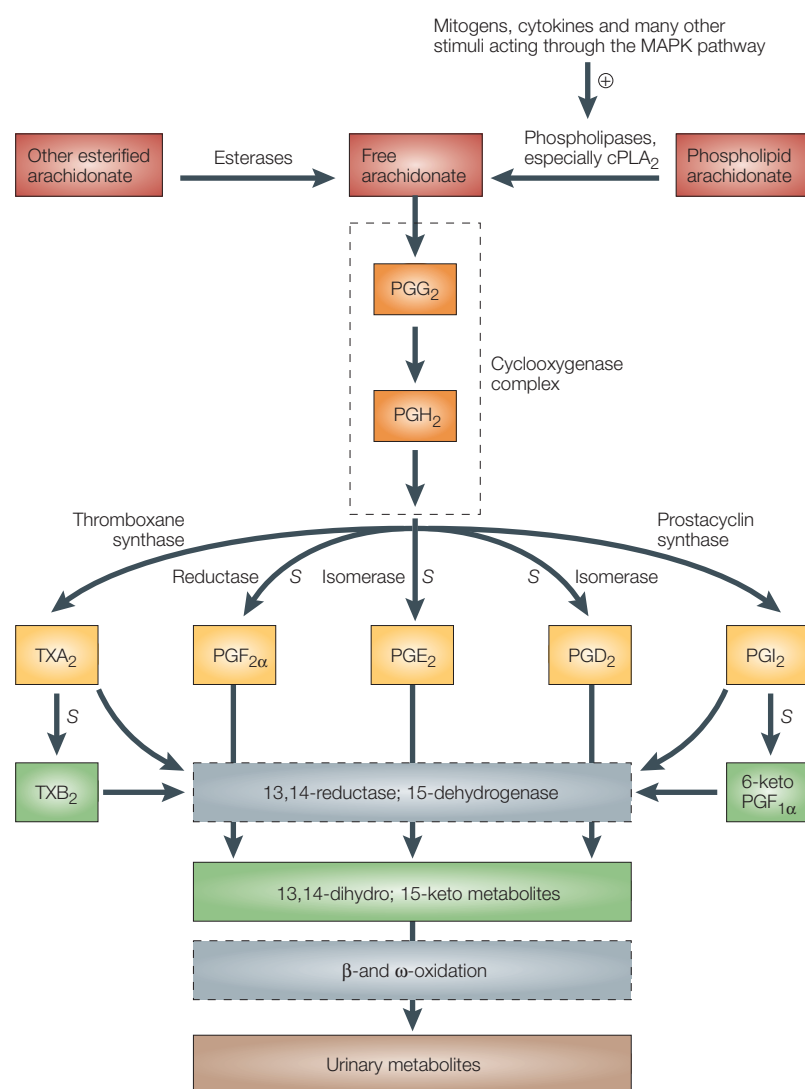


Figure 1 | An overview of prostaglandin synthesis and metabolism. In theory, free fatty acids such as arachidonate can be formed from several sources, although phospholipid-bound arachidonate is probably the most significant pool. Phospholipases, especially cytosolic phospholipase A_2 (cPLA $_2$), are highly-regulated enzymes (by the MAP kinase (MAPK) pathway) that liberate arachidonate, which is then transformed by the cyclooxygenase (COX) complex. The mechanism of this reaction is complex — 2 moles of molecular oxygen are introduced sequentially by a lipoxygenase reaction, followed by a COX reaction. This generates PGG $_2$, a 15-hydro-peroxide prostaglandin that is reduced to PGH $_2$, the corresponding hydroxy product. Both of these intermediates are short-lived but may have independent biological activity. However, in most cases it is not yet clear whether this independent activity is significant *in vivo*. A further battery of enzymes transform PGH $_2$ into a variety of products. Some of these enzymes, such as thromboxane (TX) synthase and prostacyclin (PGI $_2$) synthase, show marked tissue localization (for example, TX synthase in platelets and PGI $_2$ synthase in vascular endothelium). Other enzymes, such as the endoperoxide reductases and isomerases, are quite widely distributed, although in some cases they can be induced following inflammatory stimuli. *In vitro* at least, the endoperoxides PGG $_2$ and PGH $_2$ can also decay spontaneously (S) to PGF $_{2\alpha}$, PGE $_2$ and PGD $_2$. The evanescent products, TXA $_2$ and PGI $_2$, decay spontaneously to their respective inactive metabolites, TXB $_2$ and 6-keto PGF $_{1\alpha}$. The biological activity of the other prostaglandins is curtailed following uptake into cells, by a series of metabolic enzymes that are present in some tissues (for example, the lung) at high concentrations. Inactive metabolites of these prostanoids undergo carbon chain shortening (especially in the liver) prior to secretion in the urine, in which estimates of total body prostaglandin turnover can be made by selectively monitoring these products. The most significant products from the point of view of this review are PGE $_2$, because of its importance in inflammation, fever and pain; PGI $_2$ because of its anti-aggregatory action and possible role in hyperalgesia, and TXA $_2$ because of its important role in platelet aggregation. Colour code: red, precursors; orange, intermediates; yellow, 'primary' prostaglandins that mediate most of the biological activity of this system; green, inactive or largely inactive metabolites; brown, end metabolites that are excreted primarily in the urine.

analgesic, anti-inflammatory and ANTIPYRETIC properties to some degree, and produce characteristic side effects, including gastric intolerance and depression of blood clotting through inhibitory action on platelet function. As a group, the NSAIDs are structurally diverse, with most (but not all) being carboxylic acids (FIG. 2). The main question, from the pharmacologist's point of view, was how these apparently disparate therapeutic and side effects were mechanistically linked.

There were several early suggestions (REFS 2,3), but the real breakthrough came in 1971. Vane tells us⁴ that the idea that the aspirin-like drugs blocked the conversion of substrate arachidonic acid to prostaglandins came to him while reviewing experiments in which aspirin blocked the release of 'rabbit aorta contracting substance' (RCS) from guinea-pig and dog lung. Believing that RCS was an intermediate in prostaglandin synthesis, he wrote "...a logical corollary was that aspirin might well be blocking the synthesis of prostaglandins". A quartet of papers appeared that year from Vane and members of his group, showing that aspirin itself, indomethacin and (less effectively) sodium salicylate blocked prostaglandin synthesis in a cell-free system⁵ and in isolated perfused spleen of dogs⁶. In humans, therapeutic doses of aspirin taken by volunteers reduced prostaglandin generation by aggregating platelets *ex vivo*⁷ or in seminal plasma samples collected during the course of the treatment⁸. The overall message was clear — at least some NSAIDs were able to prevent the generation of prostaglandins by direct action on the COX enzyme, and did so in humans in clinical doses. But how was this linked to their therapeutic actions?

The late 1960s and early 1970s had seen an explosion of interest in the biology of the prostaglandins. It had already been shown that prostaglandins are generated during platelet aggregation to produce fever, HYPERALGESIA and inflammation (reviewed by Willis⁹). Prostaglandins had also been detected in gastric mucosa and been shown to inhibit ulcer formation in rodents¹⁰. In other words, the ability of NSAIDs to block COX provided the much sought after link between the therapeutic and side effects of these drugs.

Over the next couple of years, several other important findings emerged. Significantly, it was shown that the entire gamut of NSAIDs inhibit COX at concentrations well within their therapeutic plasma range and that the overall order of potency corresponded to their therapeutic activity¹¹. Other types of anti-inflammatories, such as the glucocorticoids and the so-called disease-modifying drugs, were inactive in these cell-free assays, providing further evidence for the specificity of the effect. Using ENANTIOMERIC pairs of NSAIDs, such as naproxen¹², an exquisite correlation was observed between the anti-inflammatory and anti-COX activity, and many more studies confirmed the notion that this is a fundamental mechanism of this class of drug (reviewed in REF 13).

By 1974, Vane's concept was firmly established. Researchers were quick to seize upon the fact that NSAIDs could be used to probe the functions of prostanoids in physiology and pathology, thereby opening an entirely new chapter in eicosanoid research.

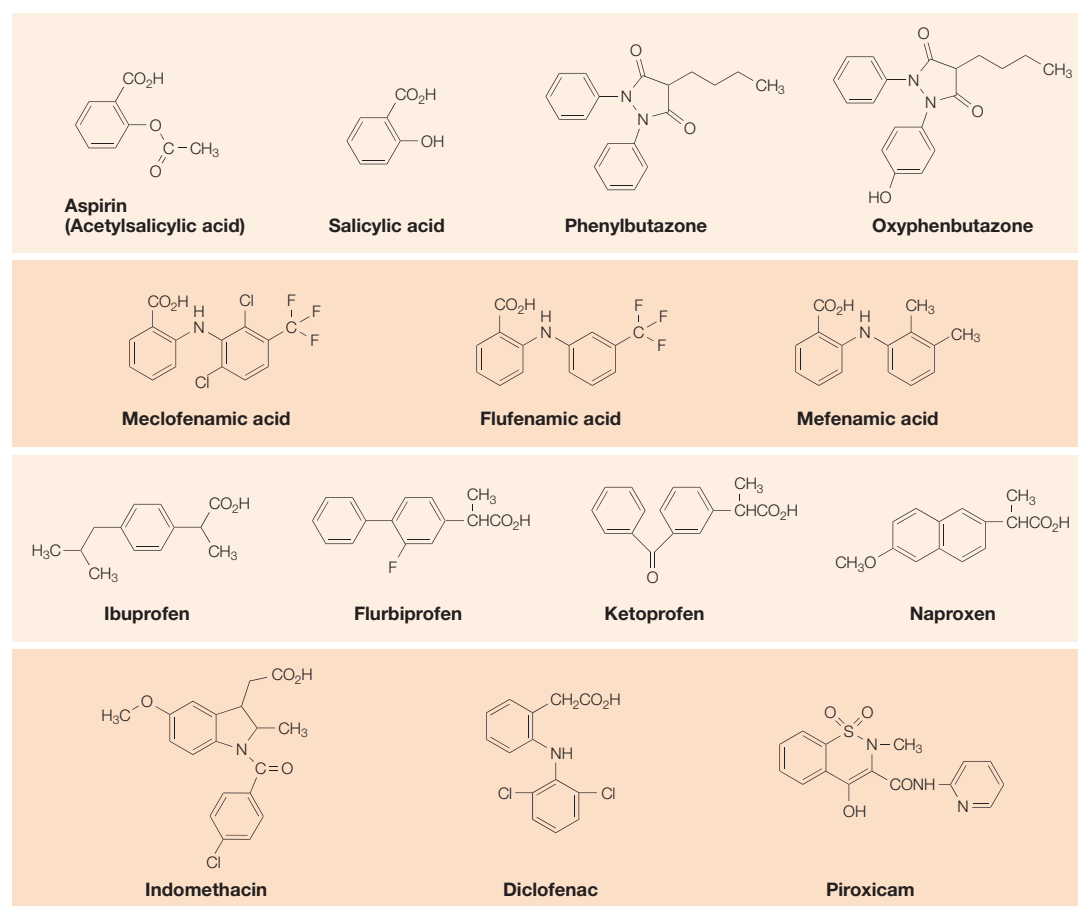


Figure 2 | **Chemical structures of NSAIDs and related compounds.** Structures of some 'classical' NSAIDs, including representative salicylates, pyrazolones, fenamates, propionates, oxicams and indomethacin. Note the general presence of a carboxylic-acid moiety.

Parenthetically, one might add that the pharmaceutical industry now possessed, probably for the first time, a simple and robust *in vitro* technique to screen compounds for putative anti-inflammatory activity. This in itself was a significant advance, and the number of chemical abstracts dealing with potential inhibitors of the COX enzyme rose markedly, with more than 2,500 per year recorded within a decade of these ideas taking hold.

But right from the beginning of the story, several anomalies were noted. The most relevant concerns paracetamol (acetaminophen) — another hugely popular drug that was also introduced in the 1890s (although it was not in common use until some 40 years later). As for all other aspirin-like drugs, paracetamol possessed antipyretic and analgesic activity but, unlike most, had little anti-inflammatory activity and caused virtually no gastric or platelet side effects. Apparently in accord with its therapeutic profile, paracetamol was found to have a different pattern of inhibitory activity, being more effective against brain COX than enzyme prepared from peripheral tissues such as the spleen¹⁴. At the time, this was put forward as a putative explanation for the selectivity of its therapeutic action, and the idea that there are several forms of the enzyme was formulated. Wide

variations in the inhibitory potency of indomethacin against COX enzymes prepared from a range of tissues was subsequently reported¹⁵ and the isoenzyme idea was further elaborated in several reviews^{13,16}.

The development of the field

Despite the early indications for alternative forms of COX, little concrete evidence emerged to support this idea for several years. The structural features of COX, which has dual hydroperoxidase and cyclooxygenase activity¹⁷, were not well understood. With the notable exception of bovine or ovine seminal vesicles, COX is usually expressed in tissues in low abundance. As a dimeric membrane-bound protein, it posed many challenges to purification and was not sequenced until 1988 (REFS 18,19). Other factors complicating the interpretation of potency differences included the wide variations in assay conditions that were used by different groups — which is still a problem today — as well as differences in the kinetics of the COX inhibitors, some of which produced 'competitive reversible' inhibition, whereas others exhibited unusual inhibitory effects such as the 'competitive non-reversible' mechanism that is observed in the case of indomethacin²⁰. The discovery^{21,22} that, alone among the group, aspirin

HYPERALGESIA

An abnormal state of increased sensitivity to painful stimuli.

ENANTIOMERS

A pair of compounds whose molecular structures are mirror images of each other.

irreversibly acetylated a serine residue (Ser530) within the COX active site to produce its effects, reinforced the opinion held by many workers at that time that the NSAIDs were a chemically heterogeneous group of drugs with widely differing modes of inhibitory action. In short, it seemed at least possible that some of these differences in inhibitor potency could be the result of factors other than the presence of isozymes.

Meanwhile, other apparently unrelated investigations were to provide the backdrop for future dramatic revelations. Using a model of rabbit kidney inflammation induced by ligation of the urethra, Needleman's group observed²³ that the affected, but not the CONTRALATERAL, organ unexpectedly developed an enormous capacity to generate prostaglandins. In the following year²⁴, the group showed that this increase was due to *de novo* synthesis of fresh enzyme, although no suggestion was made at this time that the new enzyme was a variant form. Nevertheless, this theme was pursued by the Needleman group over the next few years and, in 1982, a paper appeared that suggested the presence of two distinct forms of COX in brain tissue with differing sensitivities to indomethacin²⁵. Other studies in gastrointestinal tissue were also supportive of the selectivity of action of NSAIDs in different tissues²⁶.

The problem was taken up again by several laboratories towards the end of the 1980s. Treatment of vascular smooth muscle cells with epidermal growth factor (EGF)²⁷ or fibroblasts, and monocytes with pro-inflammatory stimuli such as interleukin-1^{28,29} or lipopolysaccharide³⁰, induced COX mRNA and *de novo* synthesis of enzyme which, when partially sequenced, seemed to be identical to the bovine seminal vesicle COX. In one paper³¹, Needleman's group wrote "Clearly, those putative enzyme pools may arise as different gene products, possibly through the expression of different COX genes..." Interestingly, glucocorticoids inhibited the induction of this new COX protein without altering the amount of enzyme that was constitutively present in cells³². Elsewhere, further evidence emerged indicating different forms of the enzyme. For example, Wong and Richards, using immunological techniques, reported two isoforms in the rat ovary, one of which was regulated by hormones³³.

The discovery of COX2

Paradoxically, the next significant step forward came in 1991 from workers in an entirely different field. While investigating the expression of early-response genes in fibroblasts transformed with Rous sarcoma virus, Simmons and his colleagues³⁴ identified a novel mRNA transcript that coded for a protein that had a high sequence similarity, but was not identical, to the seminal vesicle COX enzyme. The suggestion was that a COX isozyme had been discovered. Independent and simultaneous confirmation of this exciting finding came from the laboratory of Herschman and colleagues, who discovered a novel cDNA species encoding a protein with a predicted structure similar to COX1 while studying phorbol-ester-induced genes in Swiss 3T3 cells³⁵. The same laboratory subsequently showed that this gene

product was indeed a novel COX³⁶ and that its induction was inhibited by dexamethasone³⁷. Similarly indicative findings were also reported in mouse fibroblasts³⁸, cultured rat mesangial cells³⁹, RAW 264.7 cells⁴⁰, rat alveolar macrophages^{41,42}, the ovary^{43,44} and other cell types⁴⁵.

But was this enzyme physiologically relevant? Needleman's group conclusively identified their inflammation-inducible form of COX as the species that both Simmons and Herschman had cloned³². As the evidence for the relevance of the two enzymes mounted, they were renamed COX1, referring to the original enzyme isolated from seminal vesicles and subsequently found to be distributed almost ubiquitously, and COX2, denoting the 'inducible' form of the enzyme (although it was expressed basally in the brain⁴⁶). The two genes had a different chromosomal organization in rodents⁴⁷ and humans⁴⁸, and COX1/COX2 mRNA was differentially expressed⁴⁹ in human tissues. Promoter analysis confirmed a fundamental difference between the two isozymes, with the COX2 promoter containing elements strongly reminiscent of those genes that are switched on during cellular stress and switched off by glucocorticoids⁵⁰, whereas COX1 had the appearance of a 'HOUSEKEEPING' GENE. Histological and other studies confirmed this apparent division of labour between the two enzymes, and COX1 seemed to be the predominant isoform in healthy gastrointestinal tissue from rat, dog and monkey⁵¹.

These facts begged a key question — if COX2 was the predominant inflammatory species, shouldn't this be the target for NSAIDs? If this was so, the corollary was surely that COX2 inhibition was the true therapeutic modality of NSAIDs, whereas COX1 inhibition might be the cause of side effects such as gastric irritation and depression of platelet aggregation. This was the notion put forward by more than one group^{52–54}, which came to be known in a rather Orwellian way as the 'COX2-bad:COX1-good' hypothesis. If these propositions were true, then a selective COX2 inhibitor would be an ideal drug, possessing the anti-inflammatory action but lacking the gastric and other side effects. But would it be possible to find or design such a drug?

Some encouragement for this idea could be derived from publications of the day^{52,53}. Most of the non-steroidal drugs available at that time actually inhibited both enzymes to a greater or lesser degree, but some selectivity of action was seen with experimental drugs such as 6-MNA and BF389. But other data that subsequently emerged from transgenic approaches were less encouraging. *Cox1*-null (REF. 55) and *Cox2*-null (REF. 56) mice did not behave exactly as expected — *Cox1*-deficient mice did not have spontaneous stomach ulcers and, whereas the ulceration caused by indomethacin in these animals was less than in the wild types, it was still very much in evidence. Again contrary to theoretical predictions, homozygous mutant mice lacking *Cox2* showed a normal (albeit acute) inflammatory response when treated with several agonists, apparently contradicting the idea that *Cox2* was the predominant enzyme in

CONTRALATERAL
On or affecting the opposite side.

HOUSEKEEPING GENE
A gene that is usually expressed at a fairly constant rate in cells as it subserves some constant physiological requirement. By comparison, an inducible gene is one that normally appears at a very specific time and in response to a specific stimulus.

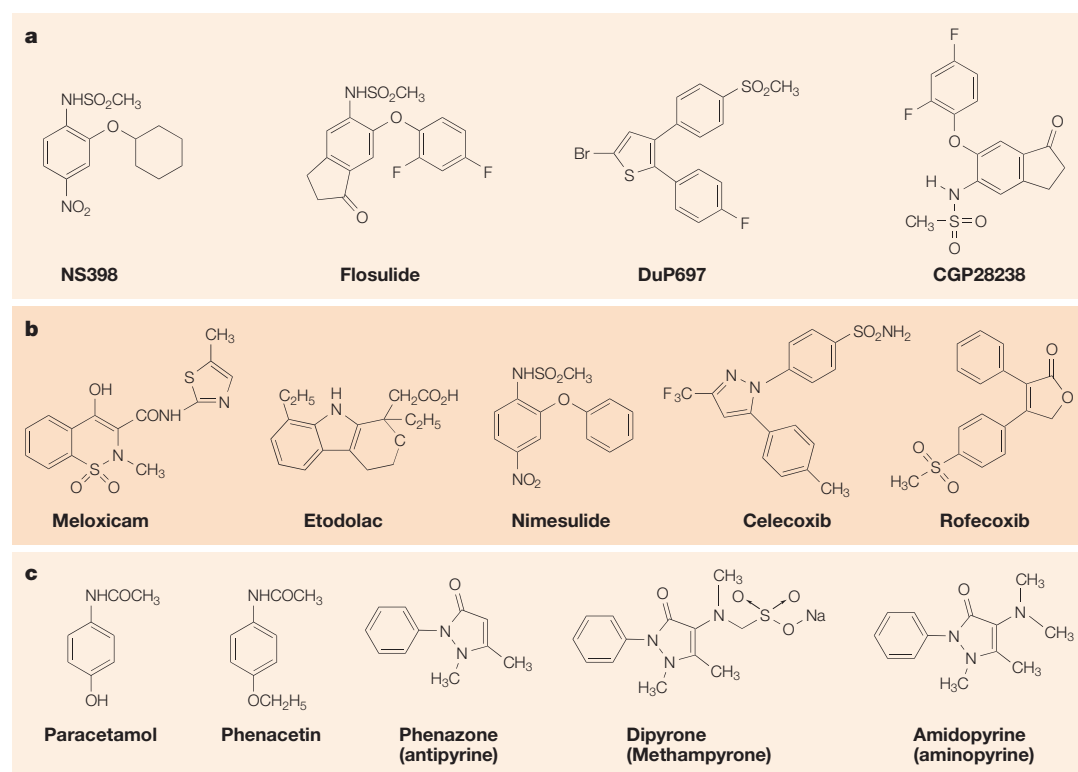


Figure 3 | **Chemical structures of NSAIDs and related compounds.** **a** | Structures of DuP697, NS398 and other similar compounds. **b** | Selective COX2 inhibitors that were discovered as a result of a search for selective isoform inhibitors (celecoxib and rofecoxib) or that were 'revealed' as being COX2 selective (meloxicam, etodolac and nimesulide). **c** | Structures of some compounds that are more effective inhibitors of COX3 according to Simmons¹²⁴.

inflammation⁵⁶. From the medicinal chemist's point of view, there was also another depressing fact — the sequence of the catalytic domains of the two isozymes was so similar that the prospect of finding a specific inhibitor seemed remote. Fortunately, this perception was soon to be changed in a rather dramatic way.

'Sleepers'

DuP697 was a drug reported in 1990 by the Dupont Company to be an effective anti-inflammatory agent with reduced ulcerogenic properties³⁷. Significantly, DuP697 showed only feeble activity in *in vitro* assays of COX using seminal vesicle or rat kidney preparations (known to predominantly contain COX1), but was more effective against rat brain prostanoid synthesis. The authors originally attributed the gastrointestinal safety of this compound to its chemical structure (a non-acidic thiophene) (FIG. 3a), which was presumed to have different pharmacokinetic properties to other COX inhibitors, most of which were carboxylic acids. Reports of other compounds with similar properties were published later, including some experimental compounds such as NS398, flosulide and CGP28238 (REF. 58) (FIG. 3a). However, in light of the discovery of COX2, it was not long before alert pharmacologists realized that these drugs might have this unusual behaviour because they acted predominately on the COX2 isozyme.

The moment of this Damascene revelation cannot be pinpointed precisely but, in the context of future drug development, two 'prostaglandin' meetings in 1992 (in Keystone, Colorado, in January, and Montreal in July) seem to have been of particular significance. At the first meeting, the Dupont Group presented evidence that gastric tolerance to DuP697 might be accounted for by differential inhibition of COX enzymes. At the second meeting, workers from Taisho described similar data with another structurally unrelated compound, NS398 (REFS 59,60). These meetings were attended by many industrial scientists, and it seems clear from published accounts that these events initiated — or at least greatly accelerated — many in-house 'COX2' programmes. It is also clear that the structures of DuP697 and NS398 were crucial starting points in the hunt for new selective inhibitors. Two companies who capitalized on these leads were Searle Monsanto (now Pharmacia) and Merck. The former focused on sulphonamide-substituted 1,5-diaryl pyrazole compounds, whereas Merck scientists settled on a series of methylsulphonylphenyl compounds.

While medicinal chemistry programmes were being developed, the field continued to produce data that were, on the whole, supportive to the COX1/COX2 concept. For example, COX2 was found in human SYNOVIAL TISSUE taken from patients with rheumatoid arthritis, and the 'inducibility' of this enzyme by cytokines was

SYNOVIAL TISSUE
The tissues that surround
the joints.

confirmed^{61,62}. Support for the COX1/COX2 concept also came from pharmacological studies. Masferrer *et al.*⁶³ showed that the onset of inflammation in the rat air pouch correlated with the appearance of COX2 in the lesion, and that NS398 blocked the production of prostaglandins at this site without causing intestinal lesions or affecting gastric prostaglandin synthesis. Similar results were seen in various inflammatory models with an early Searle Monsanto selective inhibitor SC58125 (REFS 64,65). A number of other useful, selective COX2 inhibitors were soon described in the literature — SC558 (a celecoxib prototype) showed good efficacy in rodent models of inflammation, fever and pain, whereas the closely related SC560, a selective COX1 inhibitor, was ineffective⁶⁶. Celecoxib itself (then known as SC58635) reversed CARRAGEENAN-induced hyperalgesia and local prostaglandin production⁶⁷ in rats, and a related compound was active intrathecally⁶⁸. Recombinant COX1 and COX2 had by now been prepared, and the selectivity of DuP697 and NS398 had been confirmed with *in vitro* assay systems using these enzymes^{69,70}.

The solution of the crystal structures of COX1 in 1994 (REF. 71) and COX2 in 1996 (REF. 72) made a substantial impact on the field. In the former paper, attention was drawn to the crucial roles of arginine 120 (Arg120, which interacted with the carboxyl group of both substrate and inhibitors) and tyrosine 355 (Tyr355) in determining the stereospecificity of NSAID binding (FIGS 4, 5). In the latter paper, the structure of COX2 was determined in the presence of several inhibitors, enabling the authors to deduce the conformational and other changes required for selective inhibition. Despite the great similarity in the sequence data, detailed examination of the structure of the catalytic sites revealed the substrate binding 'channel' in the two enzymes to be quite different. A single amino-acid change, from the comparatively bulky isoleucine (Ile) in COX1 to valine at position 523 in COX2 (equivalent to position 509 in COX1), and the conformational changes that this produced resulted in enhanced access to a 'side pocket' that allowed the binding of COX2-specific inhibitors by providing a docking site for the bulky phenylsulphonamide residue of drugs such as SC558 (FIG. 4). In an elegant demonstration of how crucial this single change was, Gierse *et al.*⁷³ showed that mutation of this residue in COX2 back to Ile largely prevented selective inhibitors such as SC58125, SC236 and NS398 from working. Further work on the structural basis for inhibition of both isoforms delineated the role of Arg106 in the binding of substrate and certain NSAIDs⁷⁴, as well as the role of Tyr355 located at the entrance of the active site of COX2 (REF. 75).

These structural data also helped explain differences in the inhibitory kinetics of COX1 and COX2 with drugs such as DuP697 and NS398 (REF. 76). There are, as stated above, several distinct mechanisms by which COX1 inhibitors can inhibit the enzyme, but many are of the competitive reversible type. By contrast, the COX2 inhibitors are irreversible, time-dependent (in the context of enzyme kinetics) inhibitors, partly as a result of the binding of the sulphonamide (or related)

moiety into the enzyme 'side pocket'. In an analysis of the kinetic behaviour of several COX inhibitors, Gierse *et al.*⁷⁷ subsequently discerned four separate modes of enzyme inhibition, ranging from the simple competitive inhibition of drugs such as ibuprofen, through the 'weak binding, time-dependent' mechanism of naproxen and the oxicams and the 'tight binding, time-dependent' inhibition of indomethacin, to the covalent modification produced by aspirin.

The development of the 'coxibs'

Encouraged by the 'concept testing' experiments with selective inhibitors, and armed with several solid leads and a clear idea of the nature of the binding site, development of this field was rapid. Celecoxib arose from the Searle Monsanto programme and showed marked selectivity for COX2 *in vitro*⁷⁸. Preclinical studies with this compound revealed that the drug had good efficacy in rodent models of inflammation, fever and pain. Early human studies confirmed its effectiveness in the treatment of osteoarthritis, rheumatoid arthritis and post-surgical pain when tested in comparison with a placebo or with comparator NSAIDs such as naproxen, ibuprofen and diclofenac. Crucially, there was also confirmation of the reduced incidence of platelet and gastrointestinal side effects (reviewed by Lefkowitz⁷⁹), and celecoxib was subsequently licensed in the United Kingdom for osteoarthritis and rheumatoid arthritis in 2000.

From Merck's methylsulphonylphenyl series came MK0966, later named rofecoxib. Once again, a series of preclinical studies confirmed the efficacy and gastrointestinal safety of this compound in rodent models of inflammation, and early human studies established its clinical utility in fever and pain (dental, DYSMENORRHOEA and post-operative models) when compared with standard NSAIDs such as ibuprofen or naproxen (reviewed by Morrison⁸⁰). In osteoarthritis and rheumatoid arthritis, rofecoxib proved superior to placebo and comparable in efficacy to standard doses of other NSAIDs such as diclofenac. Rofecoxib was also found to have greater gastrointestinal safety and not to affect platelet aggregation — it was licensed in the United Kingdom for osteoarthritis in 1999 and rheumatoid arthritis in 2001.

But it was not just the quest for new drugs that was stimulated by the discovery of COX2. The enolcarboxamide meloxicam (Boehringer Ingelheim), as well as the tetrahydropyranoindole etodolac (Wyeth/Shire), were drugs already under development at the time that the COX2 story 'broke', whereas a sulphonanilide drug, nimesulide (Helsinn), had been marketed in Europe since 1985 (FIG. 3b). In each case, clinical and experimental evidence already indicated that these agents were different from the other NSAIDs, especially in terms of their good gastrointestinal tolerance. Pharmacologists could now view these anomalous data with a vision greatly sharpened by the emerging COX2 concept, and these drugs turned out to be effective COX2 inhibitors^{81–83}. In an authoritative survey, Warner *et al.*⁸⁴ found, for example, that meloxicam and etodolac showed almost the same order of selectivity for COX1/COX2 as some of the newer agents.

CARRAGEENAN

A sulphated cell-wall polysaccharide that is found in certain red algae, which contains repeating sulphated disaccharides of galactose, and sometimes anhydrogalactose, and is used to induce an inflammatory lesion when injected into experimental animals.

DYSMENORRHOEA

Painful menstruation, often associated with nausea, vomiting, headache and faintness. It is thought to be related to excessive prostaglandin production.

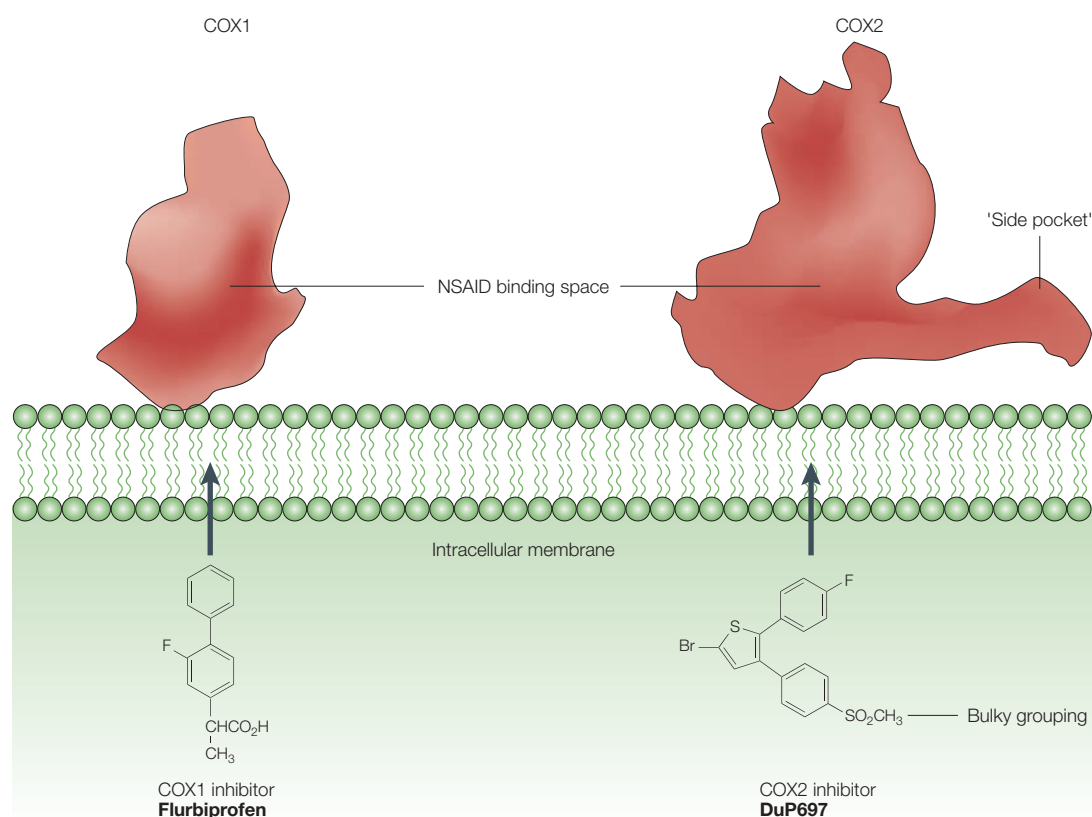


Figure 4 | **Comparison of the NSAID binding sites of COX1 and COX2 after Browner.** Schematic cartoon (modified from REF. 142), showing the differences in the NSAID binding sites of COX1 and COX2. Note that the COX2 binding site is more accommodating and is characterized by a 'side pocket' that can accommodate bulky groups such as the methyl sulphonyl moiety of DuP697. COX, cyclooxygenase; NSAID, non-steroidal anti-inflammatory drug.

The COX1/COX2 concept today

How does this very influential idea stack up today in the light of the results that have been published on the actions of the coxibs and other selective inhibitors of the COX2 isoform? Like all ideas, it has seen some modifications over the passage of time. At the tissue level there is an awareness that COX2 is more widespread in 'normal' tissue than was at first suspected and that it is not necessarily restricted to inflammatory sites. Rat brain has a large amount of the enzyme under 'resting' conditions⁸⁵, with further COX2 induced at separate sites following the administration of lipopolysaccharide⁸⁶. Certain other tissues around the body, including the MACULA Densa region of the kidney⁸⁷, are also rich sources of the enzyme under 'normal' conditions and, probably because of this, COX2 inhibitors can cause hypertension and fluid retention in some patients⁸⁸. Conversely, it has become apparent that COX1 can also be regulated under some circumstances⁸⁹.

The central idea that it is the inhibition of COX2 that is responsible for the therapeutic actions of the drugs, whereas COX1 inhibition is responsible for the side effects, is still valid, although there are some caveats. For example, there have also been well-founded reports that COX2 is present in the marginal tissue of healing ulcers and that COX2 products might contribute to the resolution of inflammation in the

gastrointestinal tract^{90–93} and elsewhere⁹⁴. This implies that inhibition of the healing response in various tissues might be an unwelcome side effect of these drugs. Unexpectedly, celecoxib (as well as ibuprofen, a mixed inhibitor) reduced the generation of the protective eicosanoid prostacyclin in humans, giving rise to a concern about unwanted cardiovascular side effects⁹⁵. Taking another tack entirely, Wallace and his colleagues⁹⁶ have argued convincingly that inhibition of one isoform in the gastrointestinal tract can be compensated for if the other isoform is not inhibited, and that it is only mixed inhibitors of the COX isoforms that give rise to gastrointestinal damage. Interestingly, the administration of COX1 inhibitors leads to the rapid induction of COX2 in the gastric mucosa (presumably a protective response), perhaps explaining why both need to be inhibited to produce damage⁹⁷.

Despite all these qualifications, the hypothesis has taken us a long way forward. It has led to the discovery of a family of drugs that are better tolerated than the older NSAIDs, our knowledge of the action of COX inhibitors has been greatly fortified and the advent of COX2-selective drugs has enabled us to discern the role of this enzyme in physiological or pathological processes. There have, of course, been new technical and other problems to solve. The question of the correct way of assessing and expressing the selectivity of

MACULA Densa
Part of the juxtaglomerular apparatus of the kidney that is important in sensing changes in blood pressure.

COX1/COX2 inhibitors is one that has exercised many investigators and been responsible for the appearance of much confusing data in the literature. Novel COX assay systems had to be devised and the potential pitfalls, such as variations in the amount of substrate, time of pre-incubation and other factors that are likely to influence the action of putative inhibitors, have been highlighted by several authors^{98,99}. Many members of the scientific and clinical community have now settled on the 'whole blood assay'¹⁰⁰ that was originally described by Patrignani *et al.* in 1994 (REF 101) as a standard technique. No assay is without its irritations, but this method has the great advantage of measuring the relative inhibitory activity in one sample of blood from the relevant species (usually human) and of taking into account factors such as plasma protein binding, which are usually ignored. The new data have also forced us to review the way in which we classify NSAIDs. There have been various suggestions^{102,103} for new nomenclature or classification of these drugs. Using a modification of Patrignani's method, Warner *et al.*⁸⁴ produced a definitive survey of most of the COX1/COX2 inhibitors available in 1999 and proposed an extremely useful classification system that has been widely adopted (TABLE 1).

From the clinical point of view, the results of two major prospective studies — the VIGOR (Vioxx Gastrointestinal Outcomes Study)¹⁰⁴ and the CLASS (Celecoxib Long Term Arthritis Safety Study)¹⁰⁵ — compared gastrointestinal safety records of rofecoxib with those of naproxen, and celecoxib to ibuprofen and diclofenac. Despite some similarities, the two trials were executed somewhat differently. In the former study using a cohort of rheumatoid arthritis patients, the concomitant use of other NSAIDs was not allowed, but the osteoarthritis cohort in the CLASS study were allowed to take low-dose aspirin in addition to celecoxib. In the completed VIGOR study, the incidence of PUBs (perforations, ulcers and bleeding) in subjects taking rofecoxib was less than half that in those using naproxen. Data from the first six months of the CLASS study did not show any difference in the incidence of the primary end point (the appearance of complicated ulcers), although the incidence of the secondary end point (symptomatic ulcers) was significantly reduced when compared with ibuprofen, but not when compared with diclofenac. One-year data from the CLASS study showed some benefit

compared with ibuprofen, but not when aspirin was allowed. Interestingly, patients entered into the VIGOR study showed a slightly higher incidence of cardiovascular events, indicating that the loss of the antiplatelet (COX1) effect of NSAIDs might be clinically disadvantageous to a small proportion of patients, even though concomitant therapy vitiates the beneficial gastrointestinal profile. Overall, the results of these two long-term studies might be said to support the COX1/COX2 concept while at the same time highlighting potential cardiovascular risks to some patients. The clinical implications of this and other aspects of the pharmacology of COX2 inhibitors have been well reviewed by FitzGerald and Patrono¹⁰⁶.

Lessons for drug discovery

So what lessons can we draw from this story? There are many ways of discovering new drugs, but it is clear that the rapid and successful discovery and marketing of celecoxib and rofecoxib owed little to the high-tech approach that most feel is the way forward for the pharmaceutical industry in this post-genomic era. It can be argued, with some justice, that this was a unique case, in that several potential lead compounds were already apparent from the literature. But at the very least, the episode shows that highly successful drugs can still be discovered (and substantial sales achieved) by organizations that are flexible in their approach to finding new medicines. It stresses the key role of the skilled and alert scientist backed by management with the vision and resources to exploit such findings as they arise.

The manner in which COX2 was identified also reminds us that we seldom know where the cutting edge of our field is at any one time. It highlights the extraordinary value of the anomalous observation and warns us to be wary of ignoring compounds that have therapeutic profiles that do not quite 'fit'. Molecular biology was key in the discovery and characterization of the COX2 isozyme, but the outcome of the initial transgenic 'hypothesis testing' studies were, perhaps, not so clear, and those who rely heavily on transgenic disease models would be right to regard this as a cautionary tale. The other point that emerges clearly is the significance of structural data, with its important lesson that even small and apparently unimportant changes in sequence might make a huge difference to the pharmacology of the enzyme.

Table 1 | **A classification of NSAIDs according to Warner *et al.*⁸⁴**

Class	Properties	Examples
Group 1	NSAIDs that can completely inhibit both COX1 and COX2 but have little selectivity	Aspirin, diclofenac, fenoprofen, flurbiprofen, indomethacin, ibuprofen, ketoprofen, mefenamic acid, naproxen, piroxicam, sulindac sulphide
Group 2	NSAIDs that inhibit COX2 with a 5–50-fold selectivity	Celecoxib, etodolac, meloxicam
Group 3	NSAIDs that inhibit COX2 with a >50-fold selectivity	Rofecoxib
Group 4	NSAIDs that are weak inhibitors of both isoforms	5-amino salicylic acid, diflunisal, sodium salicylate, nabumetone, sulphasalazine

Adapted from REF. 84. COX, cyclooxygenase; NSAID, non-steroidal anti-inflammatory drug.

IC₅₀ VALUE
The concentration of a drug or substance that inhibits a particular effect by 50%.

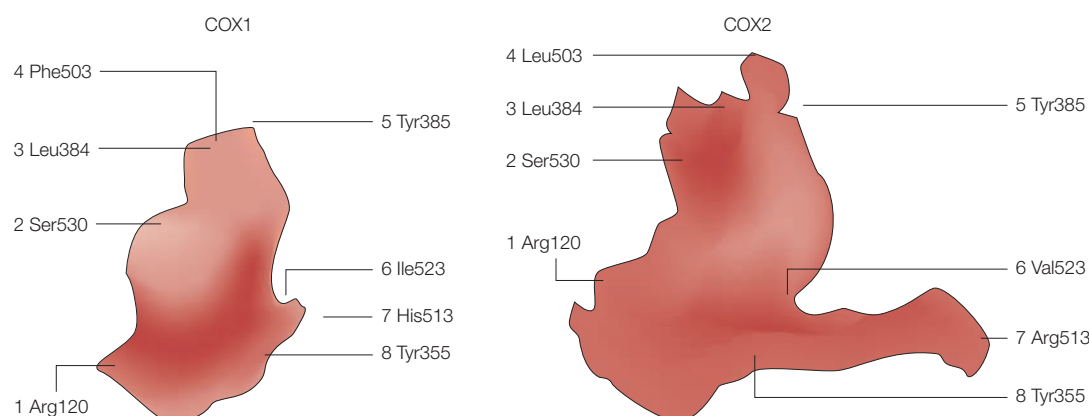


Figure 5 | Some key residues in COX1 and COX2. The left-hand panel shows a schematic diagram of the NSAID-binding site in COX1. The highly conserved residues Arg120 and Tyr355 (1 and 8) stabilize the carboxylate group that is present in most NSAIDs, whereas the aromatic ring structures are accommodated within the largely hydrophobic binding channel and often about onto the highly conserved Tyr385 (5). Tyr385 is close to the peroxidase site that forms a tyrosyl radical that is crucial to the introduction of molecular oxygen into the arachidonic acid substrate. Ser530 (2) is the residue that is acetylated by aspirin. Note the presence of the relatively bulky Ile523 (6) and the presence of Leu384 (3) in proximity to Phe503 (4), and the presence of His513 (7). The right-hand panel depicts the COX2 binding site. The highly conserved residues Arg120 and Tyr355 (1 and 8) are present as before, as is Tyr385 (5) and Ser530 (2). However, residue 4 now becomes Leu503 which, being less bulky, is not packed as tightly. This allows expansion of the available space at the top of the channel. Residue 6 becomes Val523 in COX2, which allows opening of the side pocket. The side pocket can completely accommodate the sulphonamide or analogous group of the COX2 inhibitors that are stabilized by hydrogen bonding with residue 7, which becomes Arg513 in COX2. Overall, the available space in the COX2 binding pocket is more than 25% greater than in the COX1 binding site. This is mainly due to the side pocket and the increase in available space at the top of the channel. Adapted from REFS 72, 143. For clarity, an equivalent numbering system has been used for both COX enzymes. COX, cyclooxygenase; NSAID, non-steroidal anti-inflammatory drug.

The future of COX pharmacology

Are there any more 'anomalous' COX inhibitors out there and can we expect any further surprises from the COX system? Perhaps. We do not have to look far for drugs with unusual inhibitory mechanisms. In fact, the oldest NSAID of all must surely fall into this category. Clinically, salicylic acid is almost equipotent with aspirin as an anti-inflammatory drug but shows up with much lower IC_{50} values in all *in vitro* assays. The discovery of COX2 has not helped in this instance, as the drug is a weak inhibitor of both isoforms *in vitro*. A key observation relating to this matter was published in 1972 by Hamberg¹⁰⁷, who measured the excretion of prostaglandin metabolites in volunteers taking indomethacin, aspirin or salicylic acid. Indomethacin was the most potent, but both aspirin and salicylate (4×600 mg per day) had a similar maximum effect on prostaglandin output. Although the numbers were small, there was a tantalizing hint of a latency of effect with the latter drug, indicating a mode of action other than simple COX inhibition.

Several attempts have been made to solve the discrepancy between the potency of salicylic acid *in vitro* and *in vivo*. Early studies focused on the possibility that salicylate metabolites, such as the dihydroxy gentisic acid, were active inhibitors of COX¹³ — a notion that has received recent support¹⁰⁸. A further suggestion that has been widely canvassed is that salicylate inhibits transcription of COX genes by preventing phosphorylation of I κ B α (inhibitor α of nuclear factor- κ B (NF- κ B)), thereby inhibiting NF- κ B activation^{109,110}, or by inhibiting COX2 gene transcription¹¹¹. Tumour necrosis factor

(TNF)-stimulated inducible nitric oxide synthase (iNOS) gene expression and adhesion molecule expression in HUVECs¹⁰⁹ has been reported to be inhibited by both aspirin and sodium salicylate¹¹². However, this is by no means universally agreed as a potential mechanism for controlling COX¹¹³. Effects on other signalling systems, such as extracellular signal-regulated kinase and c-JUN N-terminal kinase in human neutrophils and fibroblasts have also been claimed^{114,115}, as well as other adenosine-dependent effects of these drugs that are apparently maintained in Nf κ b-null mice^{116,117}. What this all means mechanistically and whether any of these studies are truly relevant to therapeutic dosing is still not entirely clear. Ironically, it seems that although being the simplest from a structural viewpoint, the salicylates might turn out to be among the most complex drugs of all. The matter is still very relevant, for there is sound evidence from contemporary double-blind clinical trials that willow-bark extracts can ameliorate the pain of osteoarthritis¹¹⁸, and this effect is almost certainly due to conjugated salicylate in the preparation. It is an area that will certainly repay a further inspection.

Ironically, the discovery of COX2 also failed to directly solve the paracetamol mystery. As an inhibitor, this drug is weakly active against both purified preparations of COX1 and COX2, but the concentrations that are required for inhibitory activity are higher than one would expect to achieve *in vivo*. Some exciting findings from Simmons' laboratory^{119,120} sparked speculations^{120–123} that there might be other forms of the enzyme yet to be discovered. Simmons' team observed

APOPTOTIC
A term used to describe the state of programmed cell death.

Table 2 | The COX family: a summary of properties

Gene	Gene product	Tissue expression	Functions	Inhibitors	Comments
COX1	COX1	Constitutively expressed in most tissues	Platelet aggregation, GI protection, some pain, production of vascular prostacyclin	Most 'classical' NSAIDs; some selective inhibitors	First COX to be identified
COX1	COX3	Brain, heart and aorta; constitutive?	Pain perception?	Paracetamol, diclofenac, ibuprofen, dipyrrone, phenacetin, antipyrine	Few details presently known
COX1	pCOX1a	Brain	?	NA	Not catalytically active
COX1	pCOX1b	Brain	?	NA	Not catalytically active
COX2	COX2	Induced in many tissues by many stimuli including growth factors, cytokines, oxidative stress, brain hypoxia or seizures, and other forms of injury or stress. Constitutively present in brain, kidney and elsewhere	Inflammation, fever, some pain, parturition and renal function	Many NSAIDs, COX2-selective drugs such as the coxibs and others	
COX2	COX?	J774 cells during apoptosis	?	Paracetamol	Only studied in one system to date

Modified from REF. 143. COX, cyclooxygenase; GI, gastrointestinal; NA, not applicable; NSAID, non-steroidal anti-inflammatory drug.

the appearance of (what seem to be) isoforms of COX2 that are significantly more sensitive to paracetamol than lipopolysaccharide-induced COX2 in APOPTOTIC J774.2 cell lines, and have argued that this enzyme can exist in two catalytically distinct states.

Simmons' group have now progressed even further with this idea. While analysing COX expression in canine cerebral cortex, Simmons and colleagues noticed two distinct RNA species that hybridized with COX1 cDNA on Northern blots. They went on to describe the cloning, structure and expression of a variant form of COX1 — dubbed COX3 — together with two other species, partial (p) COX1a and pCOX1b¹²⁴. All of these species are derived from COX1 by an unusual splicing mechanism that retains intron 1 in the mature protein. In addition, pCOX1a has a deletion of amino acids 117–336 (corresponding to exons 5–8), whereas pCOX1b is identical to pCOX1a except that it lacks intron 1. All three proteins are glycosylated, but only COX3 has catalytic activity (TABLE 2).

COX3 therefore retains the principal features of the general and catalytic structures of COX1 and 2, but the presence of the translated intron 1 adds a further 30 amino acids to the amino terminus which, Simmons speculates, could alter the dimerization, folding or intracellular targeting of the enzyme. Reverse transcriptase polymerase chain reaction (RT-PCR) and other techniques indicate that COX3 is present mainly in the brain (where it is expressed at a level of approximately 5% of COX1), heart and aorta — a distribution that differs from that of COX1.

From the point of view of COX pharmacology, the most fascinating thing about COX3 is its unusual drug sensitivity. Using a canine cortex cDNA library and insect cell-expression systems, Simmons' group produced catalytically active COX3 and tested its sensitivity to several NSAIDs. They found that COX3 had enhanced sensitivity to paracetamol, diclofenac, ibuprofen and aspirin when compared with COX1 or COX2.

COX3 was also selectively sensitive to other antipyretic drugs, including antipyrine, aminopyrine, dipyrrone and phenacetin (FIG. 3c), which are largely inactive against other COX preparations^{13,124}.

We must now attempt to reconcile the sensitivity of this enzyme with the analgesic/antipyretic activity of these drugs. There is ample evidence for a role of both COX1 and 2 in various types of pain^{64,125,126}, but the question of antipyretic action is less clear. COX3 is derived from the same gene as COX1, but it is thought that COX2 is the main enzyme involved in the pathogenesis of fever, as highly selective COX2 inhibitors are effective antipyretics in man¹²⁷, whereas *Cox2*-null mice have a reduced pyretic response to lipopolysaccharide¹²⁸. One possibility is that, like COX1, alternative splicing of COX2 can give rise to other species that might also be sensitive to paracetamol. Simmons' previous work on diclofenac-inducible COX2 might point to this as a feasible mechanism.

While this concept is being developed and tested (and indeed other plausible explanations for the paracetamol enigma have been advanced (REFS 129,130)), we might expect further selective COX2 inhibitors to flow from the new medicinal chemistry opportunities that are afforded by the discoveries of the past decade. Already, Pharmacia and Merck have follow-up drugs (etoricoxib and valdecoxib) available in the clinic with apparently improved pharmacokinetics. Elsewhere, there have been reports of interesting new families of inhibitors, several of which are methylsulphonates or sulphonamides^{131–133}. In other cases, advantage was taken of the unique structural features of the COX2 binding pocket to design 'aspirin-like' molecules (for example, 2-acetoxyphenyl methyl sulphide) that preferentially acetylate the COX2 active site at the Ser516 residue^{134,135}. Ester and amide derivatization of indomethacin transforms this mixed COX inhibitor into a highly selective COX2 inhibitor^{136,137}, an approach that could presumably be used with other NSAIDs.

RT-PCR (Reverse transcription polymerase chain reaction). A technique that is used to amplify cDNA from an mRNA template using sequence-specific primers.

The story of COX pharmacology has already run for more than a century, but with so many innovative new leads and the recognition that COX2 inhibitors might be useful for treating diseases such as colon cancer^{138,139} and Alzheimer's^{140,141}, we might expect the field to continue

to flourish for many years to come. Undoubtedly, the recent discovery of COX3, and the prospect of more potent non-opioid analgesics, perhaps based on the structure of existing COX3 inhibitors, is likely to fuel another explosion of interest in COX pharmacology.

1. Analysts Rx Product Desk Reference (Merrill Lynch, London, 2001).
2. Smith, M. J. & Dawkins, P. D. Salicylate and enzymes. *J. Pharm. Pharmacol.* **23**, 729–744 (1971).
3. Collier, H. O. J. A pharmacological analysis of aspirin. *Adv. Pharmacol. Chemother.* **7**, 333–405 (1969).
4. Vane, J. R. Prostaglandins and the aspirin like drugs. *Hosp. Pract.* March, 61–71 (1972).
5. Vane, J. R. Inhibition of prostaglandin synthesis as a mechanism of action for aspirin-like drugs. *Nat. New Biol.* **231**, 232–235 (1971).
6. Ferreira, S. H., Moncada, S. & Vane, J. R. Indomethacin and aspirin abolish prostaglandin release from the spleen. *Nat. New Biol.* **231**, 237–239 (1971).
7. Smith, J. B. & Willis, A. L. Aspirin selectively inhibits prostaglandin production in human platelets. *Nat. New Biol.* **231**, 235–237 (1971).
8. Collier, J. G. & Flower, R. J. Effect of aspirin on human seminal prostaglandins. *Lancet* **2**, 852–853 (1971).
- References 5–8 are the original group of papers that showed that aspirin blocked the conversion of arachidonic acid to prostaglandins in cell-free systems, perfused organs and in vivo. These are the seminal papers on which the remainder of the COX story was built.**
9. Willis, A. L., Davison, P., Ramwell, P. W., Brocklehurst, W. E. & Smith, B. In *Release and Actions of Prostaglandins in Inflammation and Fever: Inhibition by Anti-inflammatory and Antipyretic Drugs* (eds P. W. R. & Pharris, B. B.) (Plenum, London, 1972).
10. Robert, A., Nezamis, J. E. & Phillips, J. P. Effect of prostaglandin E1 on gastric secretion and ulcer formation in the rat. *Gastroenterology* **55**, 481–487 (1968).
11. Flower, R., Gryglewski, R., Herbaczynska-Cedro, K. & Vane, J. R. Effects of anti-inflammatory drugs on prostaglandin biosynthesis. *Nat. New Biol.* **238**, 104–106 (1972).
- An important extension of the previous findings that showed that virtually all of the NSAIDs at that time had similar effects on the COX enzyme, that these effects occurred within the therapeutic plasma levels achieved in man, and that the anti-COX activity correlated well with the anti-inflammatory activity of these compounds. A paper that confirmed and greatly extended the original findings.**
12. Tomlinson, R. V. & Ringold, H. J. Relationship between inhibition of prostaglandin synthesis and drug efficacy: support for the current theory on mode of action of aspirin-like drugs. *Biochem. Biophys. Res. Commun.* **46**, 552–559 (1972).
13. Flower, R. J. Drugs which inhibit prostaglandin biosynthesis. *Pharmacol. Rev.* **26**, 33–67 (1974).
14. Flower, R. J. & Vane, J. R. Inhibition of prostaglandin synthetase in brain explains the anti-pyretic activity of paracetamol (4–acetamidophenol). *Nature* **240**, 410–411 (1972).
- A short paper that reported the differences in COX selectivity with regard to paracetamol (acetaminophen). This was the first indication of the degree of enzyme selectivity exhibited by COX inhibitors.**
15. Bhattacharjee, P. & Eakins, K. Inhibition of the PG synthetase system in ocular tissue by indomethacin. *Pharmacologist* **15**, 209–215 (1973).
16. Flower, R. J. & Vane, J. R. In *Prostaglandin Synthetase Inhibitors* (eds Robinson, H. J. & J. R. V.) 9–18 (Raven, New York, 1974).
17. Marnett, L. J., Rowlinson, S. W., Goodwin, D. C., Kalgutkar, A. S. & Lanzo, C. A. Arachidonic acid oxygenation by COX-1 and COX-2. Mechanisms of catalysis and inhibition. *J. Biol. Chem.* **274**, 22903–22906 (1999).
18. Merlie, J. P., Fagan, D., Mudd, J. & Needleman, P. Isolation and characterization of the complementary DNA for sheep seminal vesicle prostaglandin endoperoxide synthase (cyclooxygenase). *J. Biol. Chem.* **263**, 3550–3553 (1988).
19. DeWitt, D. L., el-Hariri, E. A. & Smith, W. L. Molecular cloning of prostaglandin G/H synthase. *Adv. Prostaglandin Thromboxane Leukot. Res.* **19**, 454–457 (1989).
- References 18 and 19 are important papers that dealt with the sequencing of COX1.**
20. Smith, W. L. & Lands, W. E. Stimulation and blockade of prostaglandin biosynthesis. *J. Biol. Chem.* **246**, 6700–6702 (1971).
21. Roth, G. J., Stanford, N. & Majerus, P. W. Acetylation of prostaglandin synthase by aspirin. *Proc. Natl Acad. Sci. USA* **72**, 3073–3076 (1975).
22. Roth, G. J. & Siok, C. J. Acetylation of the NH₂-terminal serine of prostaglandin synthetase by aspirin. *J. Biol. Chem.* **253**, 3782–3784 (1978).
- Another crucial discovery in the COX field is presented in references 21 and 22. Here, aspirin was shown to act by irreversibly acetylating a serine residue within the COX active site.**
23. Nishikawa, K., Morrison, A. & Needleman, P. Exaggerated prostaglandin biosynthesis and its influence on renal resistance in the isolated hydronephrotic rabbit kidney. *J. Clin. Invest.* **59**, 1143–1150 (1977).
24. Morrison, A. R., Moritz, H. & Needleman, P. Mechanism of enhanced renal prostaglandin biosynthesis in ureter obstruction. Role of *de novo* protein synthesis. *J. Biol. Chem.* **253**, 8210–8212 (1978).
- References 23 and 24 are papers that, in the light of subsequent discoveries, may be regarded as key developments showing increases in tissue COX levels in response to inflammation.**
25. Lysz, T. W. & Needleman, P. Evidence for two distinct forms of fatty acid cyclooxygenase in brain. *J. Neurochem.* **38**, 1111–1117 (1982).
26. Whittle, B. J., Higgs, G. A., Eakins, K. E., Moncada, S. & Vane, J. R. Selective inhibition of prostaglandin production in inflammatory exudates and gastric mucosa. *Nature* **284**, 271–273 (1980).
27. Pash, J. M. & Bailey, J. M. Inhibition by corticosteroids of epidermal growth factor-induced recovery of cyclooxygenase after aspirin inactivation. *FASEB J.* **2**, 2613–2618 (1988).
28. Raz, A., Wyche, A., Siegel, N. & Needleman, P. Regulation of fibroblast cyclooxygenase synthesis by interleukin-1. *J. Biol. Chem.* **263**, 3022–3028 (1988).
29. Raz, A., Wyche, A. & Needleman, P. Temporal and pharmacological division of fibroblast cyclooxygenase expression into transcriptional and translational phases. *Proc. Natl Acad. Sci. USA* **86**, 1657–1661 (1989).
30. Fu, J. Y., Masferrer, J. L., Seibert, K., Raz, A. & Needleman, P. The induction and suppression of prostaglandin H₂ synthase (cyclooxygenase) in human monocytes. *J. Biol. Chem.* **265**, 16737–16740 (1990).
31. Masferrer, J. L., Zweifel, B. S., Seibert, K. & Needleman, P. Selective regulation of cellular cyclooxygenase by dexamethasone and endotoxin in mice. *J. Clin. Invest.* **86**, 1375–1379 (1990).
- Another key paper in which the 'two enzyme' hypothesis was articulated.**
32. Masferrer, J. L., Seibert, K., Zweifel, B. & Needleman, P. Endogenous glucocorticoids regulate an inducible cyclooxygenase enzyme. *Proc. Natl Acad. Sci. USA* **89**, 3917–3921 (1992).
33. Wong, W. Y., DeWitt, D. L., Smith, W. L. & Richards, J. S. Rapid induction of prostaglandin endoperoxide synthase in rat preovulatory follicles by luteinizing hormone and cAMP is blocked by inhibitors of transcription and translation. *Mol. Endocrinol.* **3**, 1714–1723 (1989).
34. Xie, W. L., Chipman, J. G., Robertson, D. L., Erikson, R. L. & Simmons, D. L. Expression of a mitogen-responsive gene encoding prostaglandin synthase is regulated by mRNA splicing. *Proc. Natl Acad. Sci. USA* **88**, 2692–2696 (1991).
35. Kujubu, D. A., Fletcher, B. S., Varnum, B. C., Lim, R. W. & Herschman, H. R. TIS10, a phorbol ester tumor promoter-inducible mRNA from Swiss 3T3 cells, encodes a novel prostaglandin synthase/cyclooxygenase homologue. *J. Biol. Chem.* **266**, 12866–12872 (1991).
- References 34 and 35 are landmark papers — the conclusive identification by two laboratories of a novel gene that codes for an alternative form of COX.**
36. Fletcher, B. S., Kujubu, D. A., Perrin, D. M. & Herschman, H. R. Structure of the mitogen-inducible TIS10 gene and demonstration that the TIS10-encoded protein is a functional prostaglandin G/H synthase. *J. Biol. Chem.* **267**, 4338–4344 (1992).
37. Kujubu, D. A. & Herschman, H. R. Dexamethasone inhibits mitogen induction of the TIS10 prostaglandin synthase/cyclooxygenase gene. *J. Biol. Chem.* **267**, 7991–7994 (1992).
38. O'Banion, M. K., Sadowski, H. B., Winn, V. & Young, D. A. A serum- and glucocorticoid-regulated 4-kilobase mRNA encodes a cyclooxygenase-related protein. *J. Biol. Chem.* **266**, 23261–23267 (1991).
39. Simonson, M. S., Wolfe, J. A., Konieczkowski, M., Sedor, J. R. & Dunn, M. J. Regulation of prostaglandin endoperoxide synthase gene expression in cultured rat mesangial cells: induction by serum via a protein kinase-C-dependent mechanism. *Mol. Endocrinol.* **5**, 441–451 (1991).
40. Farber, J. M. A collection of mRNA species that are inducible in the RAW 264.7 mouse macrophage cell line by gamma interferon and other agents. *Mol. Cell Biol.* **12**, 1535–1545 (1992).
41. Lee, S. H. *et al.* Selective expression of mitogen-inducible cyclooxygenase in macrophages stimulated with lipopolysaccharide. *J. Biol. Chem.* **267**, 25934–25938 (1992).
42. O'Sullivan, M. G., Chilton, F. H., Huggins, E. M. Jr & McCall, C. E. Lipopolysaccharide priming of alveolar macrophages for enhanced synthesis of prostanoids involves induction of a novel prostaglandin H synthase. *J. Biol. Chem.* **267**, 14547–14550 (1992).
43. Sirosi, J., Simmons, D. L. & Richards, J. S. Hormonal regulation of messenger ribonucleic acid encoding a novel isoform of prostaglandin endoperoxide H synthase in rat preovulatory follicles. Induction *in vivo* and *in vitro*. *J. Biol. Chem.* **267**, 11586–11592 (1992).
44. Sirosi, J. & Richards, J. S. Purification and characterization of a novel, distinct isoform of prostaglandin endoperoxide synthase induced by human chorionic gonadotropin in granulosa cells of rat preovulatory follicles. *J. Biol. Chem.* **267**, 6382–6388 (1992).
45. Ryseck, R. P. *et al.* Identification of an immediate early gene, *pghs-B*, whose protein product has prostaglandin synthase/cyclooxygenase activity. *Cell Growth Differ.* **3**, 443–450 (1992).
46. Feng, L. *et al.* Cloning two isoforms of rat cyclooxygenase: differential regulation of their expression. *Arch. Biochem. Biophys.* **307**, 361–368 (1993).
47. Xie, W., Merrill, J. R., Bradshaw, W. S. & Simmons, D. L. Structural determination and promoter analysis of the chicken mitogen-inducible prostaglandin G/H synthase gene and genetic mapping of the murine homolog. *Arch. Biochem. Biophys.* **300**, 247–252 (1993).
48. Kosaka, T. *et al.* Characterization of the human gene (PTGS2) encoding prostaglandin endoperoxide synthase 2. *Eur. J. Biochem.* **221**, 889–897 (1994).
49. O'Neill, G. P. & Ford-Hutchinson, A. W. Expression of mRNA for cyclooxygenase-1 and cyclooxygenase-2 in human tissues. *FEBS Lett.* **330**, 156–160 (1993).
50. Evett, G. E., Xie, W., Chipman, J. G., Robertson, D. L. & Simmons, D. L. Prostaglandin G/H synthase isoenzyme 2 expression in fibroblasts: regulation by dexamethasone, mitogens, and oncogenes. *Arch. Biochem. Biophys.* **306**, 169–177 (1993).
51. Kargman, S. *et al.* Characterization of prostaglandin G/H synthase 1 and 2 in rat, dog, monkey, and human gastrointestinal tracts. *Gastroenterology* **111**, 445–454 (1996).
52. Mitchell, J. A., Akarasereenont, P., Thiemermann, C., Flower, R. J. & Vane, J. R. Selectivity of nonsteroidal anti-inflammatory drugs as inhibitors of constitutive and inducible cyclooxygenase. *Proc. Natl Acad. Sci. USA* **90**, 11693–11697 (1993).
53. Meade, E. A., Smith, W. L. & DeWitt, D. L. Differential inhibition of prostaglandin endoperoxide synthase (cyclooxygenase) isozymes by aspirin and other non-steroidal anti-inflammatory drugs. *J. Biol. Chem.* **268**, 6610–6614 (1993).

54. DeWitt, D. L., Meade, E. A. & Smith, W. L. PGH synthase isoenzyme selectivity: the potential for safer nonsteroidal anti-inflammatory drugs. *Am. J. Med.* **95**, S40–S44 (1993).
References 52–54 are a group of papers that first articulated the concept that it is inhibition of COX1 that produces the side effects of aspirin-like drugs, whereas inhibition of COX2 gives rise to the therapeutic effects.
55. Langenbach, R. *et al.* Prostaglandin synthase 1 gene disruption in mice reduces arachidonic acid-induced inflammation and indomethacin-induced gastric ulceration. *Cell* **83**, 483–492 (1995).
56. Dinchuk, J. E. *et al.* Renal abnormalities and an altered inflammatory response in mice lacking cyclooxygenase II. *Nature* **378**, 406–409 (1995).
References 55 and 56 are the original papers studying the effects of the absence of Cox1 and Cox2 using knockout mice.
57. Gans, K. R. *et al.* Anti-inflammatory and safety profile of DuP 697, a novel orally effective prostaglandin synthesis inhibitor. *J. Pharmacol. Exp. Ther.* **254**, 180–187 (1990).
Paper describing the pharmacology of DuP697, which turned out to have an extremely influential effect on the emerging COX2 field.
58. Klein, T., Nusing, R. M., Pfeilschifter, J. & Ullrich, V. Selective inhibition of cyclooxygenase 2. *Biochem. Pharmacol.* **48**, 1605–1610 (1994).
59. Futaki, N. *et al.* Selective inhibition of NS-398 on prostanoid production in inflamed tissue in rat carrageenan-air-pouch inflammation. *J. Pharm. Pharmacol.* **45**, 753–755 (1993).
60. Futaki, N. *et al.* NS-398, a new anti-inflammatory agent, selectively inhibits prostaglandin G/H synthase/cyclooxygenase (COX-2) activity *in vitro*. *Prostaglandins* **47**, 55–59 (1994).
61. Crofford, L. J. *et al.* Cyclooxygenase-1 and -2 expression in rheumatoid synovial tissues. Effects of interleukin-1 beta, phorbol ester, and corticosteroids. *J. Clin. Invest.* **93**, 1095–1101 (1994).
62. Geng, Y., Blanco, F. J., Cornelissen, M. & Lotz, M. Regulation of cyclooxygenase-2 expression in normal human articular chondrocytes. *J. Immunol.* **155**, 796–801 (1995).
63. Masferrer, J. L. *et al.* Selective inhibition of inducible cyclooxygenase 2 *in vivo* is antiinflammatory and nonulcerogenic. *Proc. Natl Acad. Sci. USA* **91**, 3228–3232 (1994).
64. Seibert, K. *et al.* Pharmacological and biochemical demonstration of the role of cyclooxygenase 2 in inflammation and pain. *Proc. Natl Acad. Sci. USA* **91**, 12013–12017 (1994).
65. Anderson, G. D. *et al.* Selective inhibition of cyclooxygenase (COX)-2 reverses inflammation and expression of COX-2 and interleukin 6 in rat adjuvant arthritis. *J. Clin. Invest.* **97**, 2672–2679 (1996).
66. Smith, C. J. *et al.* Pharmacological analysis of cyclooxygenase-1 in inflammation. *Proc. Natl Acad. Sci. USA* **95**, 13313–13318 (1998).
67. Zhang, Y., Shaffer, A., Portanova, J., Seibert, K. & Isakson, P. C. Inhibition of cyclooxygenase-2 rapidly reverses inflammatory hyperalgesia and prostaglandin E2 production. *J. Pharmacol. Exp. Ther.* **283**, 1069–1075 (1997).
68. Dirig, D. M., Isakson, P. C. & Yaksh, T. L. Effect of COX-1 and COX-2 inhibition on induction and maintenance of carrageenan-evoked thermal hyperalgesia in rats. *J. Pharmacol. Exp. Ther.* **285**, 1031–1038 (1998).
69. Gierse, J. K. *et al.* Expression and selective inhibition of the constitutive and inducible forms of human cyclo-oxygenase. *Biochem. J.* **305**, 479–484 (1995).
70. O'Neill, G. P. *et al.* Selective inhibitors of COX-2. *Agents Actions Suppl.* **46**, 159–168 (1995).
71. Picot, D., Loll, P. J. & Garavito, R. M. The X-ray crystal structure of the membrane protein prostaglandin H2 synthase-1. *Nature* **367**, 243–249 (1994).
72. Kurumbail, R. G. *et al.* Structural basis for selective inhibition of cyclooxygenase-2 by anti-inflammatory agents. *Nature* **384**, 644–648 (1996).
Solving the crystal structure of COX and COX2 (see references 71 and 72) had an enormous impact on the COX field and was vitally important in understanding the pharmacology of the enzyme.
73. Gierse, J. K. *et al.* A single amino acid difference between cyclooxygenase-1 (COX-1) and -2 (COX-2) reverses the selectivity of COX-2 specific inhibitors. *J. Biol. Chem.* **271**, 15810–15814 (1996).
An extremely elegant experiment demonstrating the crucial importance of a single amino acid change in the structure of COX2 that profoundly alters the architecture, and therefore the drug specificity, of the enzyme.
74. Greig, G. M. *et al.* The interaction of arginine 106 of human prostaglandin G/H synthase-2 with inhibitors is not a universal component of inhibition mediated by nonsteroidal anti-inflammatory drugs. *Mol. Pharmacol.* **52**, 829–838 (1997).
75. So, O. Y., Scarafia, L. E., Mak, A. Y., Callan, O. H. & Swinney, D. C. The dynamics of prostaglandin H synthases. Studies with prostaglandin h synthase 2 Y355F unmask mechanisms of time-dependent inhibition and allosteric activation. *J. Biol. Chem.* **273**, 5801–5807 (1998).
76. Copeland, R. A. *et al.* Mechanism of selective inhibition of the inducible isoform of prostaglandin G/H synthase. *Proc. Natl Acad. Sci. USA* **91**, 11202–11206 (1994).
77. Glaser, K. E., Koboldt, C. M., Walker, M. C., Seibert, K. & Isakson, P. C. Kinetic basis for selective inhibition of cyclo-oxygenases. *Biochem. J.* **339**, 607–614 (1999).
An elegant analysis of various types of COX inhibition.
78. Isakson, P., Kurumbail, R. G., Gierse, J. K., Seibert, K. & Maziasz, T. J. In *Therapeutic Roles of Selective Cox-2 Inhibitors* (eds Vane, J. R. & Botting, R. M.) 48–59 (William Harvey, London, 2001).
79. Lefkowitz, J. B., Verburg, K. M. & Geis, G. S. In *Therapeutic Roles of Selective Cox-2 Inhibitors* (eds Vane, J. R. & Botting, R. M.) 461–481 (William Harvey, London, 2001).
80. Morrison, B., Simon, T. J., DeTora, L. & Sperling, R. In *Therapeutic Roles of Selective Cox-2 Inhibitors* (eds Vane, J. R. & Botting, R. M.) 541–559 (William Harvey, London, 2001).
81. Glaser, K. *et al.* Etodolac selectively inhibits human prostaglandin G/H synthase 2 (PGHS-2) versus human PGHS-1. *Eur. J. Pharmacol.* **281**, 107–111 (1995).
82. Tavares, I. A., Bishai, P. M. & Bennett, A. Activity of nimesulide on constitutive and inducible cyclooxygenases. *Arzneimittelforschung* **45**, 1093–1095 (1995).
83. Vago, T., Bevilacqua, M. & Norbiato, G. Effect of nimesulide action time dependence on selectivity towards prostaglandin G/H synthase/cyclooxygenase activity. *Arzneimittelforschung* **45**, 1096–1098 (1995).
84. Warner, T. D. *et al.* Nonsteroid drug selectivities for cyclo-oxygenase-1 rather than cyclo-oxygenase-2 are associated with human gastrointestinal toxicity: a full *in vitro* analysis. *Proc. Natl Acad. Sci. USA* **96**, 7563–7568 (1999).
An authoritative survey that categorized all NSAIDs, including selective COX inhibitors, into different classes. A very useful reference.
85. Breder, C. D., Dewitt, D. & Kraig, R. P. Characterization of inducible cyclooxygenase in rat brain. *J. Comp. Neurol.* **355**, 296–315 (1995).
86. Cao, C., Matsumura, K., Yamagata, K. & Watanabe, Y. Induction by lipopolysaccharide of cyclooxygenase-2 mRNA in rat brain: its possible role in the febrile response. *Brain Res.* **697**, 187–196 (1995).
87. Harris, R. C. *et al.* Cyclooxygenase-2 is associated with the macula densa of rat kidney and increases with salt restriction. *J. Clin. Invest.* **94**, 2504–2510 (1994).
88. Whelton, A., White, W. B., Bello, A. E., Puma, J. A. & Fort, J. G. Effects of celecoxib and rofecoxib on blood pressure and edema in patients > or =65 years of age with systemic hypertension and osteoarthritis. *Am. J. Cardiol.* **90**, 959–963 (2002).
89. Smith, C. J., Morrow, J. D., Roberts, L. J., 2nd & Marnett, L. J. Induction of prostaglandin endoperoxide synthase-1 (COX-1) in a human promonocytic cell line by treatment with the differentiating agent TPA. *Adv. Exp. Med. Biol.* **400A**, 99–106 (1997).
90. McCarthy, D. M. Mechanisms of mucosal injury and healing: the role of non-steroidal anti-inflammatory drugs. *Scand. J. Gastroenterol. Suppl.* **208**, 24–29 (1995).
91. Wallace, J. L., Reuter, B. K., McKnight, W. & Bak, A. Selective inhibitors of cyclooxygenase-2: are they really effective, selective, and GI-safe? *J. Clin. Gastroenterol.* **27** Suppl 1, S28–S34 (1998).
92. Schmassmann, A. *et al.* Effects of inhibition of prostaglandin endoperoxide synthase-2 in chronic gastrointestinal ulcer models in rats. *Br. J. Pharmacol.* **123**, 795–804 (1998).
93. Bamba, H., Ota, S., Kato, A. & Matsuzaki, F. Nonsteroidal anti-inflammatory drugs may delay the repair of gastric mucosa by suppressing prostaglandin-mediated increase of hepatocyte growth factor production. *Biochem. Biophys. Res. Commun.* **245**, 567–571 (1998).
94. Appleton, I., Tomlinson, A., Mitchell, J. A. & Willoughby, D. A. Distribution of cyclooxygenase isoforms in murine chronic granulomatous inflammation. Implications for future anti-inflammatory therapy. *J. Pathol.* **176**, 413–420 (1995).
95. McAdam, B. F. *et al.* Systemic biosynthesis of prostacyclin by cyclooxygenase (COX)-2: the human pharmacology of a selective inhibitor of COX-2. *Proc. Natl Acad. Sci. USA* **96**, 272–277 (1999).
96. Wallace, J. L., McKnight, W., Reuter, B. K. & Vergnolle, N. NSAID-induced gastric damage in rats: requirement for inhibition of both cyclooxygenase 1 and 2. *Gastroenterology* **119**, 706–714 (2000).
An interesting paper that provides another interpretation for the lack of gastrointestinal side effects of COX2 inhibitors.
97. Fiorucci, S. *et al.* Cyclooxygenase-2-derived lipoxin A₂ increases gastric resistance to aspirin-induced damage. *Gastroenterology* **123**, 1598–1606 (2002).
98. Riendeau, D. *et al.* Comparison of the cyclooxygenase-1 inhibitory properties of nonsteroidal anti-inflammatory drugs (NSAIDs) and selective COX-2 inhibitors, using sensitive microsomal and platelet assays. *Can. J. Physiol. Pharmacol.* **75**, 1088–1095 (1997).
99. Lora, M., Denault, J. B., Leduc, R. & de Brum-Fernandes, A. J. Systematic pharmacological approach to the characterization of NSAIDs. *Prostaglandins Leukot. Essent. Fatty Acids* **59**, 55–62 (1998).
100. Brooks, P. *et al.* Interpreting the clinical significance of the differential inhibition of cyclooxygenase-1 and cyclo-oxygenase-2. *Rheumatology (Oxford)* **38**, 779–788 (1999).
The 'whole blood assay' that has been adopted as the gold standard for assessment of the inhibitory activity of COX.
101. Patrignani, P. *et al.* Biochemical and pharmacological characterization of the cyclooxygenase activity of human blood prostaglandin endoperoxide synthases. *J. Pharmacol. Exp. Ther.* **271**, 1705–1712 (1994).
102. Frolich, J. C. A classification of NSAIDs according to the relative inhibition of cyclooxygenase isoenzymes. *Trends Pharmacol. Sci.* **18**, 30–34 (1997).
103. Vane, J. R. & Warner, T. D. Nomenclature for COX-2 inhibitors. *Lancet* **356**, 1373–1374. (2000).
104. Bombardier, C. *et al.* Comparison of upper gastrointestinal toxicity of rofecoxib and naproxen in patients with rheumatoid arthritis. VIGOR Study Group. *N. Engl. J. Med.* **343**, 1520–1528, 2 p following 1528 (2000).
105. Silverstein, F. E. *et al.* Gastrointestinal toxicity with celecoxib vs nonsteroidal anti-inflammatory drugs for osteoarthritis and rheumatoid arthritis: the CLASS study: A randomized controlled trial. Celecoxib Long-term Arthritis Safety Study. *JAMA* **284**, 1247–1255 (2000).
References 104 and 105 report the outcome of the VIGOR and CLASS studies into the benefits of COX2 inhibitors over conventional therapy.
106. FitzGerald, G. A. & Patrono, C. The coxibs, selective inhibitors of cyclooxygenase-2. *N. Engl. J. Med.* **345**, 433–442 (2001).
An excellent analytical review of the advantages and disadvantages of COX2-selective inhibitors from a clinical perspective.
107. Hamberg, M. Inhibition of prostaglandin synthesis in man. *Biochem. Biophys. Res. Commun.* **49**, 720–726 (1972).
108. Hinz, B., Kraus, V., Pahl, A. & Brune, K. Salicylate metabolites inhibit cyclooxygenase-2-dependent prostaglandin E(2) synthesis in murine macrophages. *Biochem. Biophys. Res. Commun.* **274**, 197–202 (2000).
109. Pierce, J. W., Read, M. A., Ding, H., Lusinskas, F. W. & Collins, T. Salicylates inhibit I kappa B-alpha phosphorylation, endothelial-leukocyte adhesion molecule expression, and neutrophil transmigration. *J. Immunol.* **156**, 3961–3969 (1996).
110. Fernandez de Arriba, A. *et al.* Inhibition of cyclooxygenase-2 expression by 4-trifluoromethyl derivatives of salicylate, triflusal, and its deacetylated metabolite, 2-hydroxy-4-trifluoromethylbenzoic acid. *Mol. Pharmacol.* **55**, 753–760 (1999).
111. Xu, X. M. *et al.* Suppression of inducible cyclooxygenase 2 gene transcription by aspirin and sodium salicylate. *Proc. Natl Acad. Sci. USA* **96**, 5292–5297 (1999).
112. Farivar, R. S., Chobanian, A. V. & Brecher, P. Salicylate or aspirin inhibits the induction of the inducible nitric oxide synthase in rat cardiac fibroblasts. *Circ. Res.* **78**, 759–768 (1996).
113. Mitchell, J. A., Saunders, M., Barnes, P. J., Newton, R. & Belvisi, M. G. Sodium salicylate inhibits cyclo-oxygenase-2 activity independently of transcription factor (nuclear factor kappaB) activation: role of arachidonic acid. *Mol. Pharmacol.* **51**, 907–912 (1997).

114. Pillinger, M. H. *et al.* Modes of action of aspirin-like drugs: salicylates inhibit erk activation and integrin-dependent neutrophil adhesion. *Proc. Natl Acad. Sci. USA* **95**, 14540–14545 (1998).
 115. Schwenger, P., Alpert, D., Skolnik, E. Y. & Vilecek, J. Cell-type-specific activation of c-Jun N-terminal kinase by salicylates. *J. Cell Physiol.* **179**, 109–114 (1999).
 116. Cronstein, B. N., Montesinos, M. C. & Weissmann, G. Sites of action for future therapy: an adenosine-dependent mechanism by which aspirin retains its anti-inflammatory activity in cyclooxygenase-2 and NFκB knockout mice. *Osteoarthritis Cartilage* **7**, 361–363 (1999).
 117. Cronstein, B. N., Montesinos, M. C. & Weissmann, G. Salicylates and sulfasalazine, but not glucocorticoids, inhibit leukocyte accumulation by an adenosine-dependent mechanism that is independent of inhibition of prostaglandin synthesis and p105 of NFκB. *Proc. Natl Acad. Sci. USA* **96**, 6377–6381 (1999).
 118. Chrubasik, S. *et al.* Treatment of low back pain exacerbations with willow bark extract: a randomized double-blind study. *Am. J. Med.* **109**, 9–14 (2000).
 119. Simmons, D. L., Botting, R. M., Robertson, P. M., Madsen, M. L. & Vane, J. R. Induction of an acetaminophen-sensitive cyclooxygenase with reduced sensitivity to nonsteroid anti-inflammatory drugs. *Proc. Natl Acad. Sci. USA* **96**, 3275–3280 (1999).
 120. Simmons, D. L., Wagner, D. & Westover, K. Nonsteroidal anti-inflammatory drugs, acetaminophen, cyclooxygenase 2, and fever. *Clin. Infect. Dis.* **31** Suppl 5, S211–S218 (2000).
 121. Flower, R. J. Non-steroidal anti-inflammatory drugs: back to the future. *Rheumatology (Oxford)* **38**, 693–696 (1999).
 122. Botting, R. M. Mechanism of action of acetaminophen: is there a cyclooxygenase 3? *Clin. Infect. Dis.* **31** Suppl 5, S202–S210 (2000).
 123. Willoughby, D. A., Moore, A. R. & Colville-Nash, P. R. COX-1, COX-2, and COX-3 and the future treatment of chronic inflammatory disease. *Lancet* **355**, 646–648 (2000).
 124. Chandrasekharan, N. V. *et al.* COX-3, a cyclooxygenase-1 variant inhibited by acetaminophen and other analgesic/antipyretic drugs: cloning, structure, and expression. *Proc. Natl Acad. Sci. USA* **99**, 13926–13931 (2002).
- References 119, 120 and 124 are new publications that indicate the existence of isoforms of COX2 and describe COX3, a novel COX enzyme formed by alternative splicing of COX1. These papers are set to have a profound influence on the future of COX pharmacology.**
125. Martinez, R. V. *et al.* Involvement of peripheral cyclooxygenase-1 and cyclooxygenase-2 in inflammatory pain. *J. Pharm. Pharmacol.* **54**, 405–412 (2002).
 126. Ochi, T., Motoyama, Y. & Goto, T. The analgesic effect profile of FR122047, a selective cyclooxygenase-1 inhibitor, in chemical nociceptive models. *Eur. J. Pharmacol.* **391**, 49–54 (2000).
 127. Schwartz, J. I. *et al.* Cyclooxygenase-2 inhibition by rofecoxib reverses naturally occurring fever in humans. *Clin. Pharmacol. Ther.* **65**, 653–660 (1999).
 128. Li, S. *et al.* The febrile response to lipopolysaccharide is blocked in cyclooxygenase-2(–/–), but not in cyclooxygenase-1(–/–) mice. *Brain Res.* **825**, 86–94 (1999).
 129. Boutaud, O., Aronoff, D. M., Richardson, J. H., Marnett, L. J. & Oates, J. A. Determinants of the cellular specificity of acetaminophen as an inhibitor of prostaglandin H(2) synthases. *Proc. Natl Acad. Sci. USA* **99**, 7130–7135 (2002).
 130. Ouellet, M. & Percival, M. D. Mechanism of acetaminophen inhibition of cyclooxygenase isoforms. *Arch. Biochem. Biophys.* **387**, 273–280 (2001).
 131. Huang, H. C. *et al.* Diarylspiro[2.4]heptenes as orally active, highly selective cyclooxygenase-2 inhibitors: synthesis and structure–activity relationships. *J. Med. Chem.* **39**, 253–266 (1996).
 132. Li, J. J. *et al.* Novel terphenyls as selective cyclooxygenase-2 inhibitors and orally active anti-inflammatory agents. *J. Med. Chem.* **39**, 1846–1856 (1996).
 133. Riendeau, D. *et al.* Biochemical and pharmacological profile of a tetrasubstituted furanone as a highly selective COX-2 inhibitor. *Br. J. Pharmacol.* **121**, 105–117 (1997).
 134. Kalgutkar, A. S. *et al.* Aspirin-like molecules that covalently inactivate cyclooxygenase-2. *Science* **280**, 1268–1270 (1998).
 135. Kalgutkar, A. S., Kozak, K. R., Crews, B. C., Hochgesang, G. P. Jr & Marnett, L. J. Covalent modification of cyclooxygenase-2 (COX-2) by 2-acetoxyphenyl alkyl sulfides, a new class of selective COX-2 inactivators. *J. Med. Chem.* **41**, 4800–4818 (1998).
 136. Kalgutkar, A. S. *et al.* Biochemically based design of cyclooxygenase-2 (COX-2) inhibitors: facile conversion of nonsteroidal anti-inflammatory drugs to potent and highly selective COX-2 inhibitors. *Proc. Natl Acad. Sci. USA* **97**, 925–930 (2000).
 137. Kalgutkar, A. S., Marnett, A. B., Crews, B. C., Remmel, R. P. & Marnett, L. J. Ester and amide derivatives of the nonsteroidal anti-inflammatory drug, indomethacin, as selective cyclooxygenase-2 inhibitors. *J. Med. Chem.* **43**, 2860–2870 (2000).
 138. DuBois, R. N., Giardiello, F. M. & Smalley, W. E. Nonsteroidal anti-inflammatory drugs, eicosanoids, and colorectal cancer prevention. *Gastroenterol. Clin. North Am.* **25**, 773–791 (1996).
 139. Steinbach, G. *et al.* The effect of celecoxib, a cyclooxygenase-2 inhibitor, in familial adenomatous polyposis. *N. Engl. J. Med.* **342**, 1946–1952 (2000).
 140. Stewart, W. F., Kavas, C., Corrada, M. & Metter, E. J. Risk of Alzheimer's disease and duration of NSAID use. *Neurology* **48**, 626–632 (1997).
 141. Pasinetti, G. M. & Aisen, P. S. Cyclooxygenase-2 expression is increased in frontal cortex of Alzheimer's disease brain. *Neuroscience* **87**, 319–324 (1998).
 142. Luong, C. *et al.* Flexibility of the NSAID binding site in the structure of human cyclooxygenase-2. *Nature Struct. Biol.* **3**, 927–933 (1996).
 143. Bazan, N. & Flower, R. Lipid signals in pain control. *Nature* **420**, 135–138 (2002).

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