

Linkage between dynamics and catalysis in a thermophilic-mesophilic enzyme pair

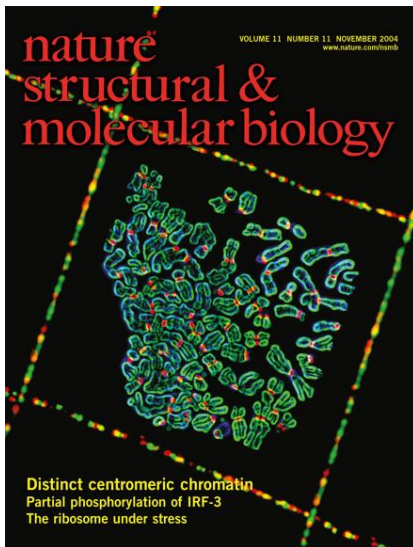
Vera Dias Gomes
Coralie Charpié
Coralie Reuse
Cédric Bärocher

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Linkage between dynamics and catalysis in a
thermophilic-mesophilic enzyme pair

Magnus Wolf-Watz, Vu Thai, Katherine Henzler-Wildman, Georgia Hadjipavlou, Elan Z Eisenmesser &
Dorothee Kern

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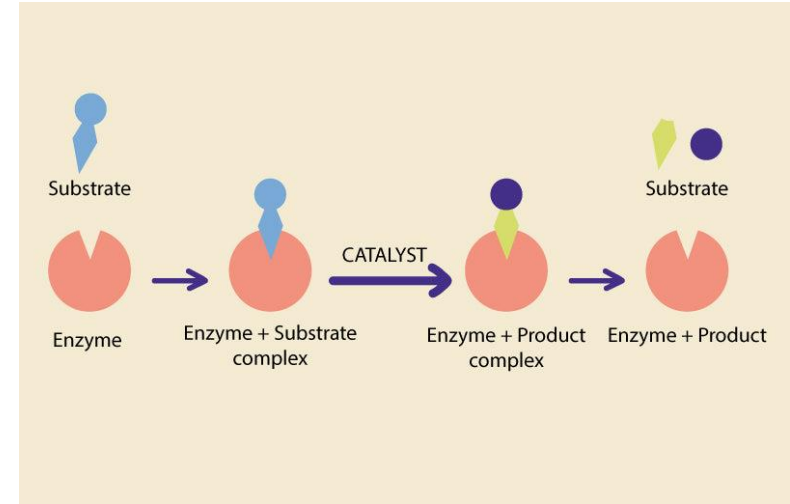


Introduction

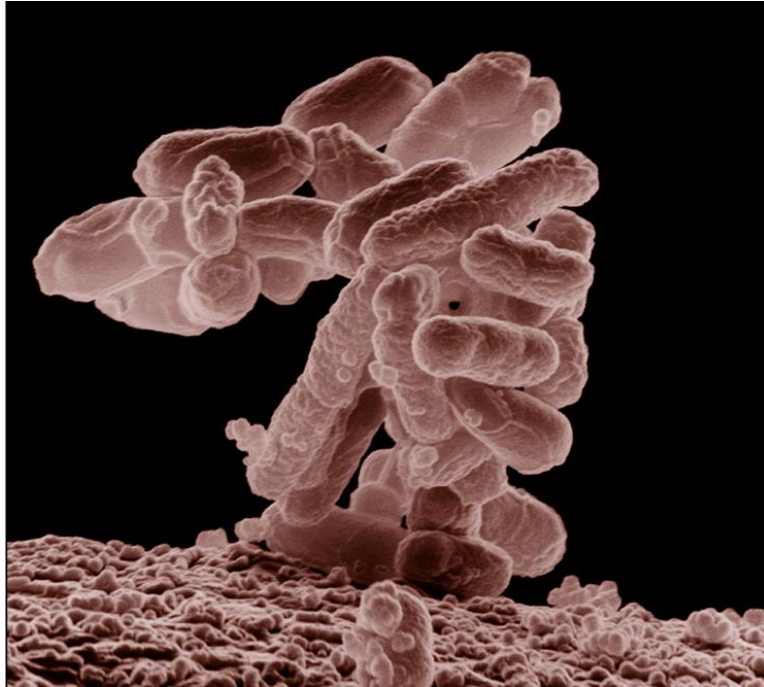
"Linkage between dynamics and catalysis in a thermophilic-mesophilic enzyme pair"

Introduction – Main issue

- What are main issues involved in enzymatic catalysis?
- 3D structure of enzymes
 - Structural biology
- Internal dynamics
 - Poorly understood



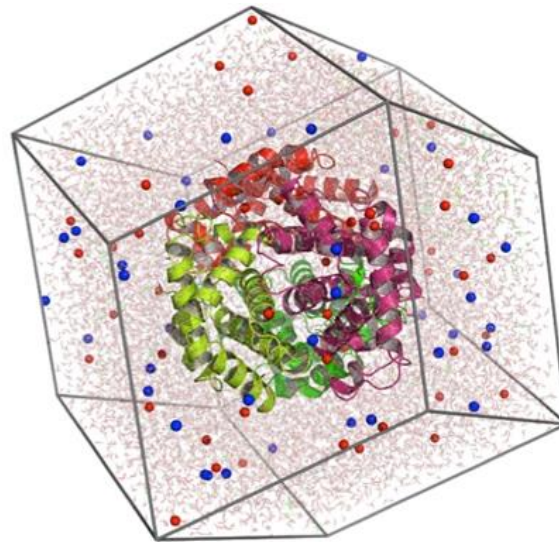
Introduction – definition



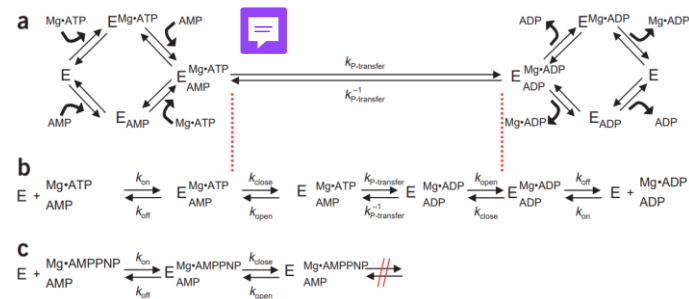
- **Thermophile:** organism living at high temperatures.
 - contains very stable enzymes
- **Mesophilic :** organisms living in more moderate conditions
 - Better catalytic activity



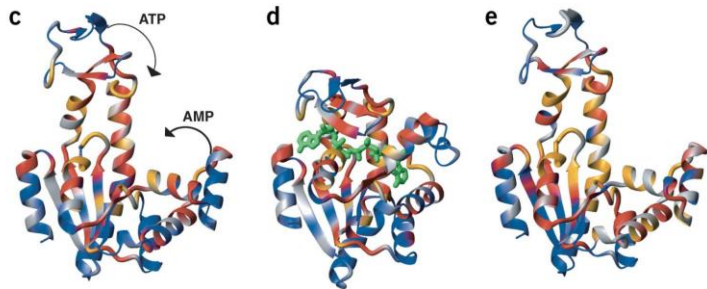
- The low activity of thermophilic enzymes at room temperature could be explained by reduced molecular dynamics
- Great stability → less flexibility
 - slow down of certain internal movements, essential for catalysis.



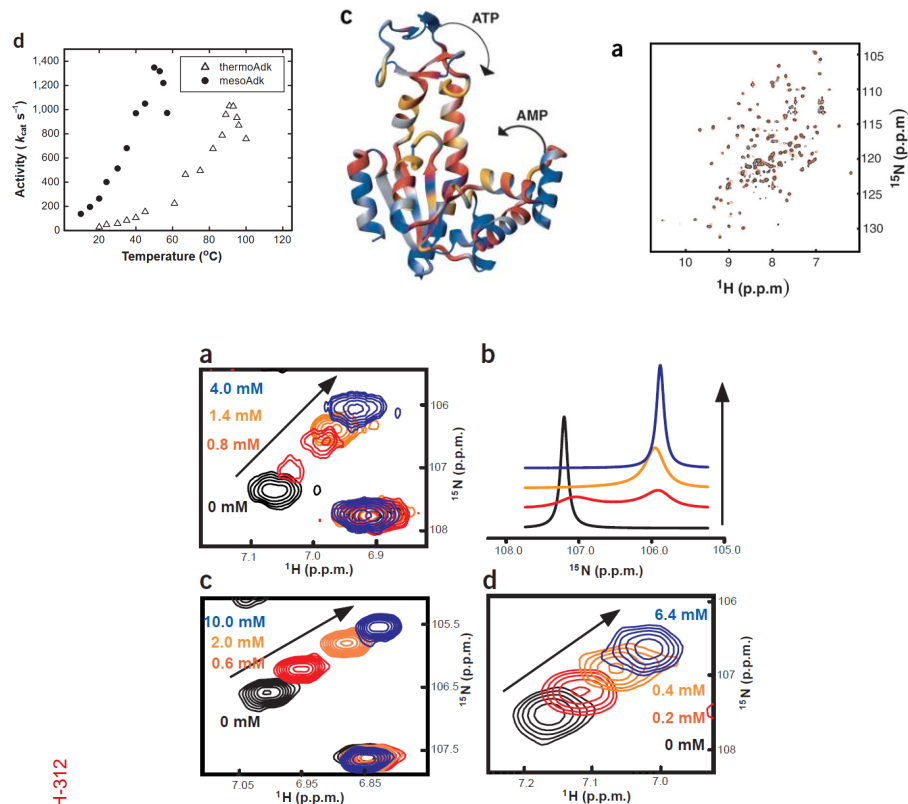
- Two homologs of the same enzyme, adenylate kinase (Adk)
 - **mesoAdk**, → mesophilic *Escherichia coli*,
 - **thermoAdk**, → thermophilic *Aquifex aeolicus*.
- Conversion of ATP and AMP into two molecules of ADP.



Introduction – approach



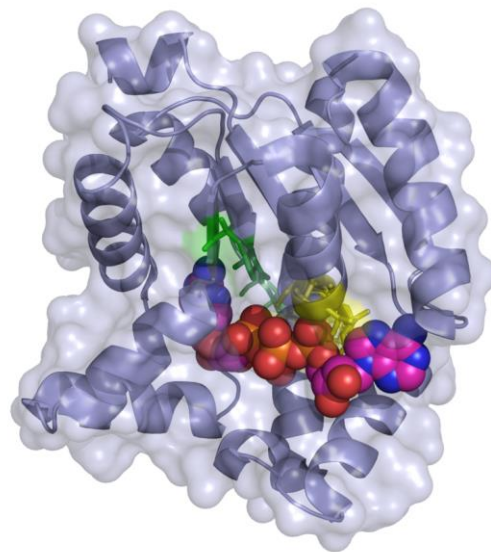
- Measurements of enzyme activity at different temperatures
- NMR measurements of dynamics
- Link the dynamic properties of the proteins to their catalytic capacity



Experimental techniques and Results

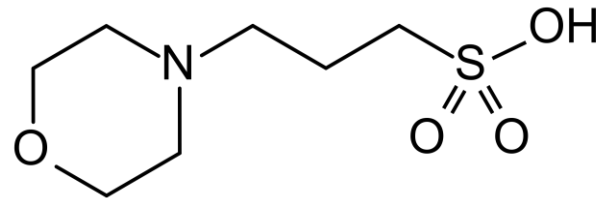
Study the catalytic activity and dynamics of the enzymes

- MesoAdk and thermoAdk recombinantly expressed in E.coli.
 - Purified by chromatography.
- NMR : ^{15}N -labeling, MOPS buffer.
 - Detect backbone amides.
- Preparation set up to mimic the conditions needed to monitor dynamics during catalysis.



Top : adenylate kinase (Adk)

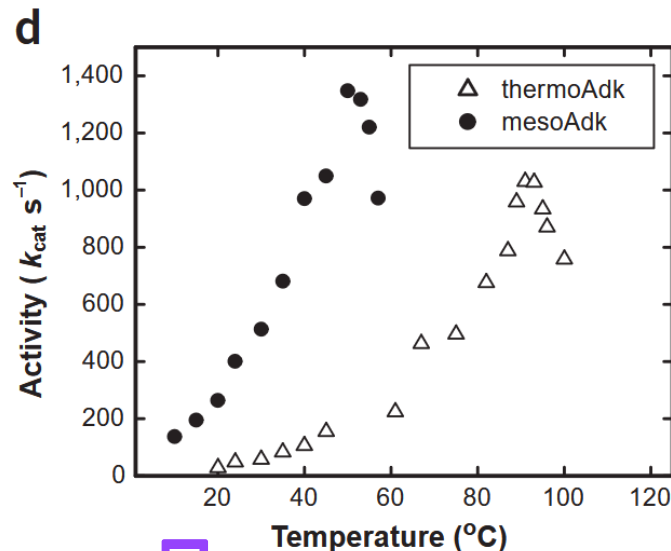
Bottom : MOPS



Coupled continuous spectroscopy assay

- Goal : compare how catalytic activity changes with temperature for the mesophilic Adk (mesoAdk) and the thermophilic one (thermoAdk).
- Tracks either ADP or ATP formation over time, by continuously monitoring the reaction.
- Coupling enzymes and nucleotides degrade at above 45°C...
 - couldn't use this continuous method for everything.

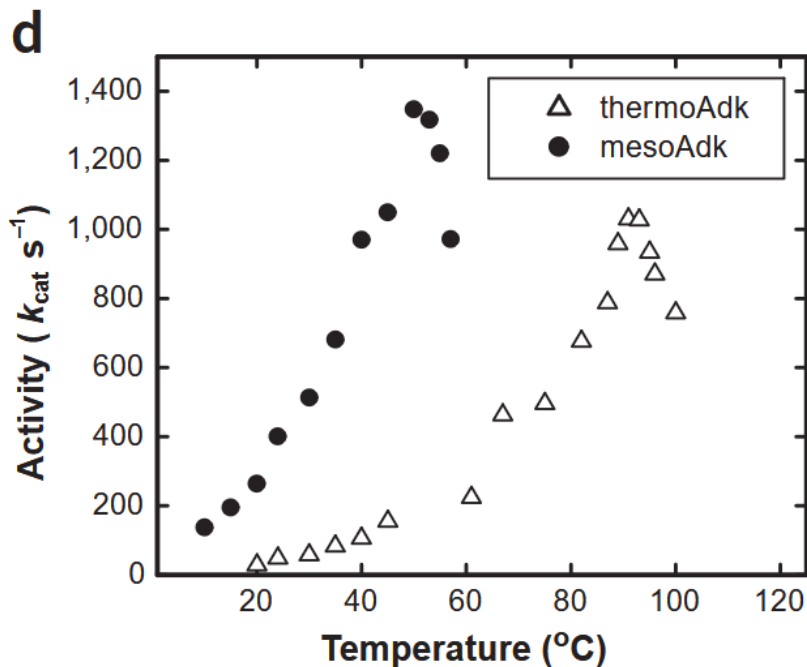
d : temperature dependence of turnover numbers in the direction of ADP formation



Activities of mesoAdk and thermoAdk as a function of temperature

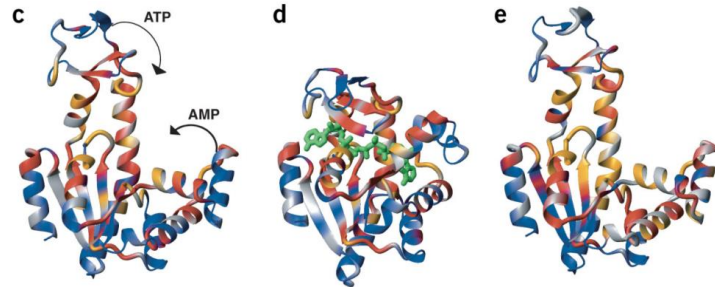
- Comparison of catalytic activities at different temperatures
- Quantification of kinetics parameters
 - Similar shapes
 - Optimal growth temperature
 - Same profiles for reverse process

d : temperature dependence of turnover numbers in the direction of ADP formation

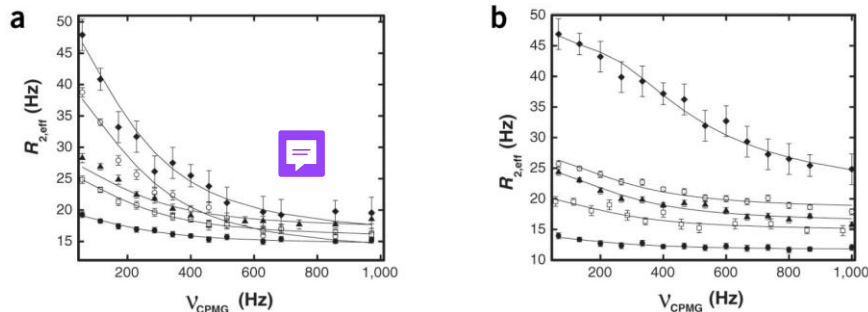


^{15}N NMR relaxation dispersion experiments

- To detect how the protein moves between two conformations (open/closed).
- CPMG (Carr-Purcell-Meiboom-Gill) relaxation dispersion.
 - Good for two-state exchange.
- At 20°C : mesoAdk is active.
- Dynamics captured reflect the phosphotransfer step.



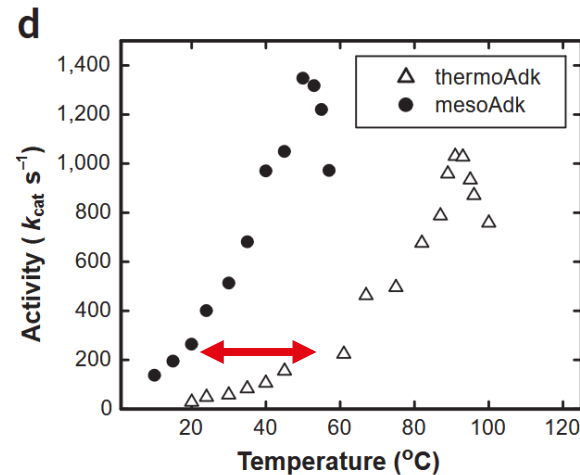
c : mesoAdk open state
d : mesoAdk closed state
e : thermoAdk open state



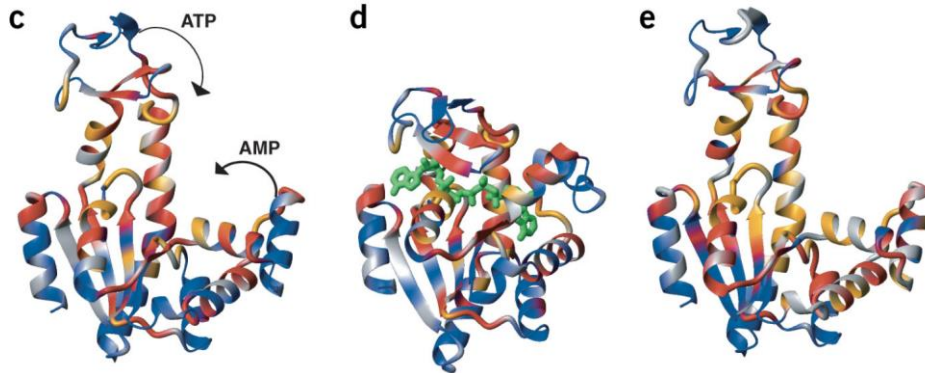
a : ^{15}N NMR relaxation dispersion, globally fitted, curve for mesoAdk.
b : ^{15}N NMR relaxation dispersion, globally fitted, curve for thermoAdk.

Backbone dynamics during turnover

- Measure of dynamic turnover using ^{15}N NMR relaxation techniques
 - 20°C for mesoAdk
 - 50°C for thermoAdk
- Phosphotransfer step measurement
 - Wide distribution of the dynamic hotspots
- Single exchange rate constant:
 - $1660 \pm 90 \text{ s}^{-1}$ for mesoAdk
 - $1615 \pm 100 \text{ s}^{-1}$ for thermoAdk at 20°C

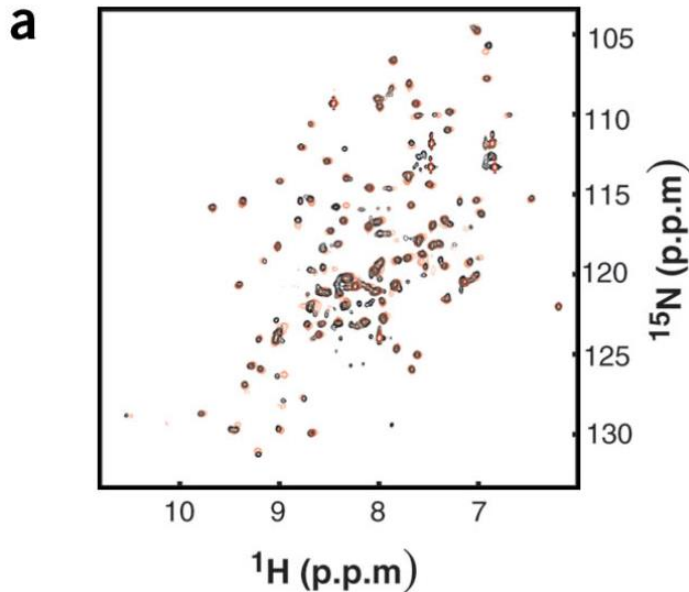


d : temperature dependence of turnover numbers in the direction of ADP formation



c : mesoAdk open state
d : mesoAdk closed state
e : thermoAdk open state

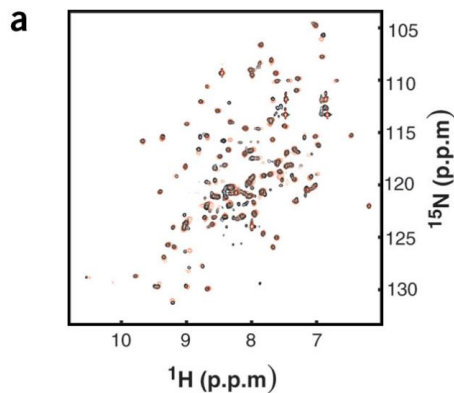
- Looks at protein backbone dynamics in the presence of AMP and AMPPNP (non-reactive ATP analog).
 - blocks the phosphotransfer.
- Test if the motions observed are linked to lid movement rather than the chemical reaction.



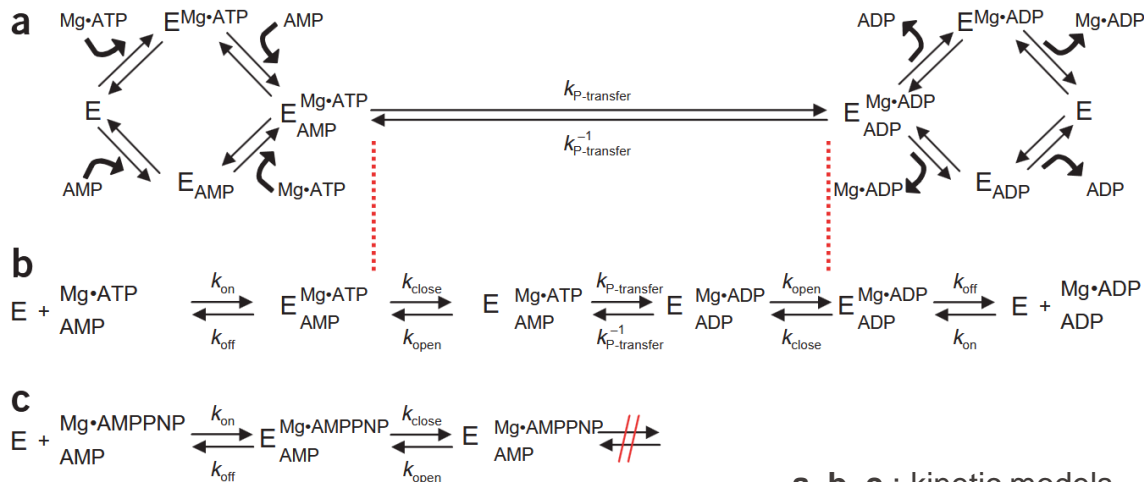
a : 2D ^1H - ^{15}N correlation spectra of thermoAdk

Backbone dynamics in the presence of AMP and AMPPNP

- Measure of enzyme dynamics of thermoAdk with and without turnover
 - lid-opening and closing is detected in relaxation experiments during catalysis
- Fitting of ^{15}N NMR
 - $k_{\text{ex}} = 1410 \pm 100 \text{ s}^{-1}$
 - same dynamic process with & without substrate



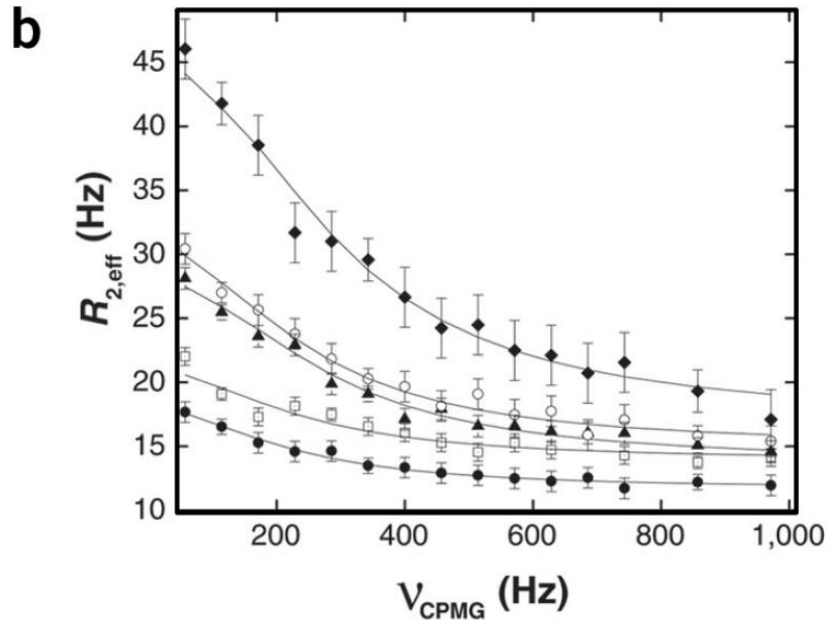
a : 2D ^1H - ^{15}N correlation spectra



a, b, c : kinetic models

Intermediate NMR time regime

- The exchange between open and closed states happens at a similar rate to the frequency difference between the two states in NMR.
- Estimate how fast the lid opens and closes.
 - measurements at two different magnetic field strengths.

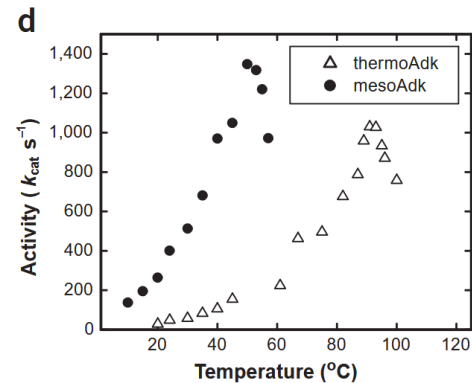


b : residues of 15N NMR relaxation dispersion profiles for thermoAdk (data globally fitted).

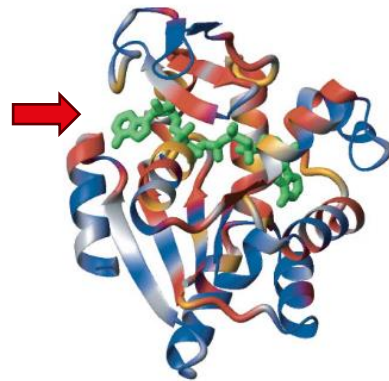
Determination of lid-closing and opening rates

- CPMG relaxation dispersion experiments

	thermoAdk	mesoAdk
k_{open}	$44 \pm 20 \text{ s}^{-1}$	$286 \pm 85 \text{ s}^{-1}$
k_{cat}	$30 \pm 10 \text{ s}^{-1}$	$263 \pm 30 \text{ s}^{-1}$
k_{close}	$1571 \pm 100 \text{ s}^{-1}$	$1374 \pm 110 \text{ s}^{-1}$

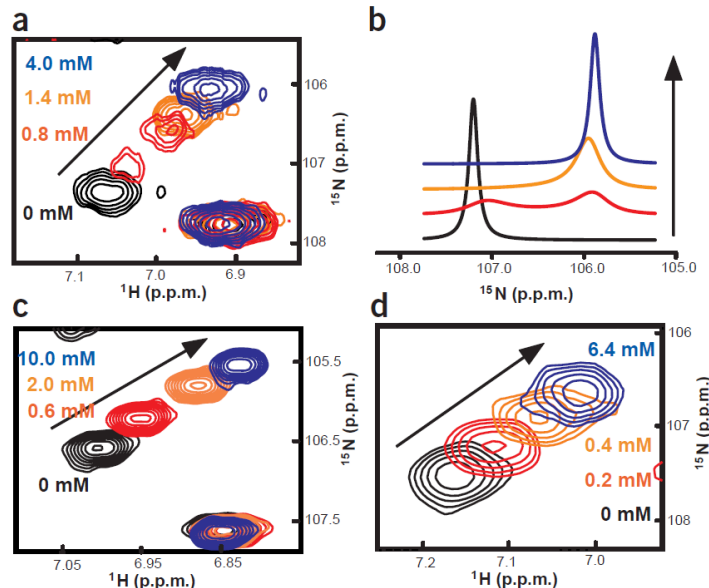


- Does the dynamic correspond to lid closure and opening?
 - similar chemical shifts during turnover and in the closed state
 - chemical shifts in relaxation dispersion and in open and close state are similar



Substrate titration experiments

- Gradually increased ligand concentration and monitored how the chemical shifts changed.
 - evidence for the slow dynamic process of thermoAdk
- 20°C : gradual shifts.
 - fast exchange.
- Intermediate ligand concentration: peak splitting.
 - slower opening of the lid.



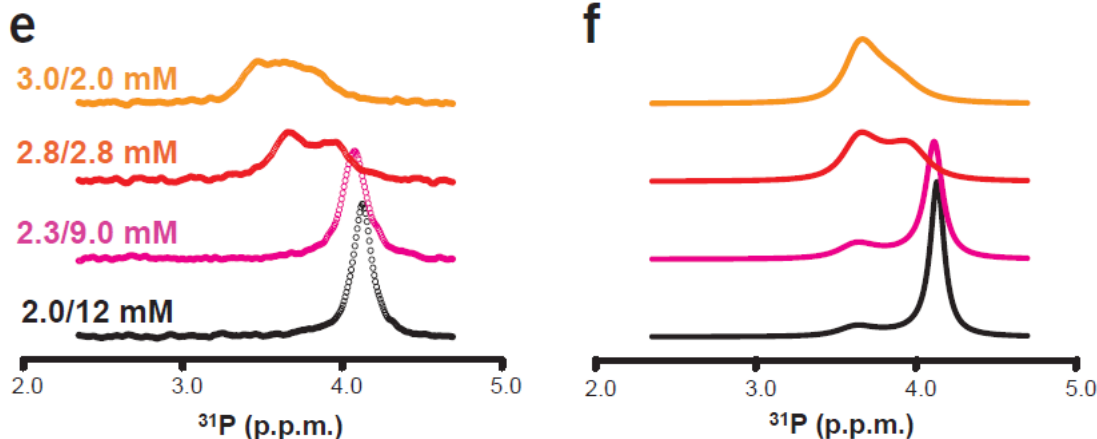
Chemical shifts of thermoAdk at :

- 20°C (a)
- 70°C (c)

b : lineshape simulation of a.

d : chemical shift of mesoAdk at 20°C.

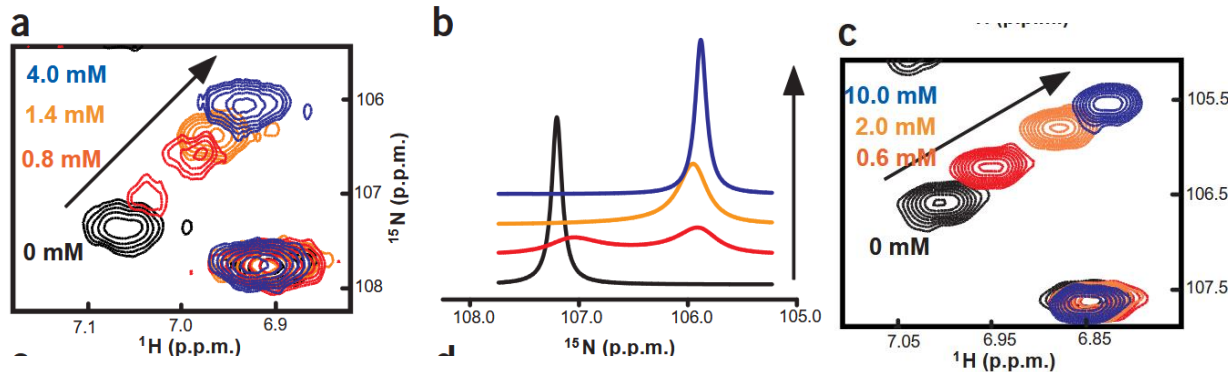
- To get more informations on the dynamics of thermoAdk at 20°C.



e : experimental ^{31}P lineshape of AMP, with increasing thermoAdk concentrations
f : theoretical ^{31}P lineshape of AMP, with increasing thermoAdk concentrations

Substrate-enzyme titration experiments

- Chemical shifts are due to conformational rearrangement



Chemical shifts of thermoAdk at :

- 20°C (a)
- 70°C (c)

b : lineshape simulation