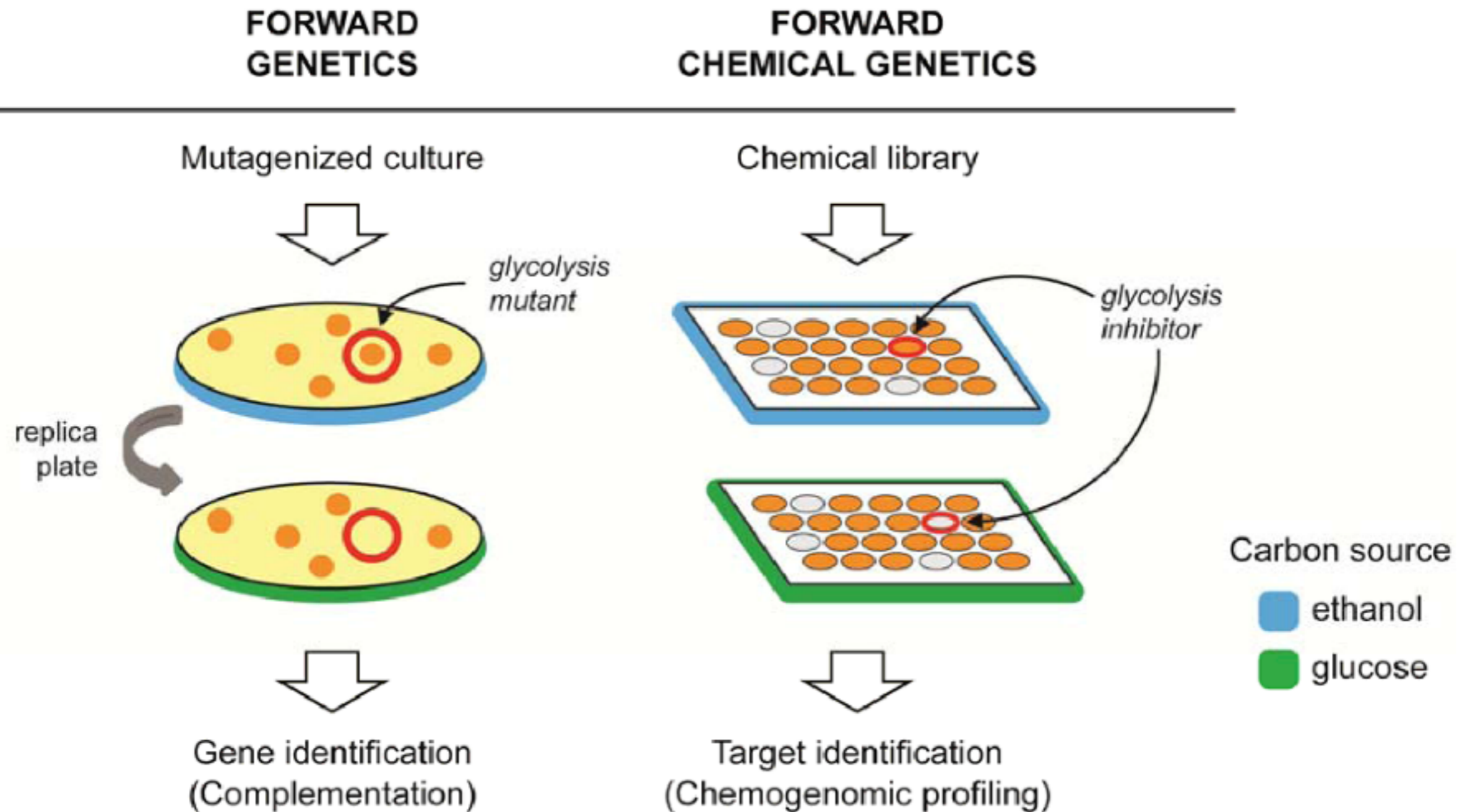


Single-cell biology

Week 6 .

**Sc-Based Perturbation Screenings
Spatial Omics techniques**

sc-Based Perturb screenings

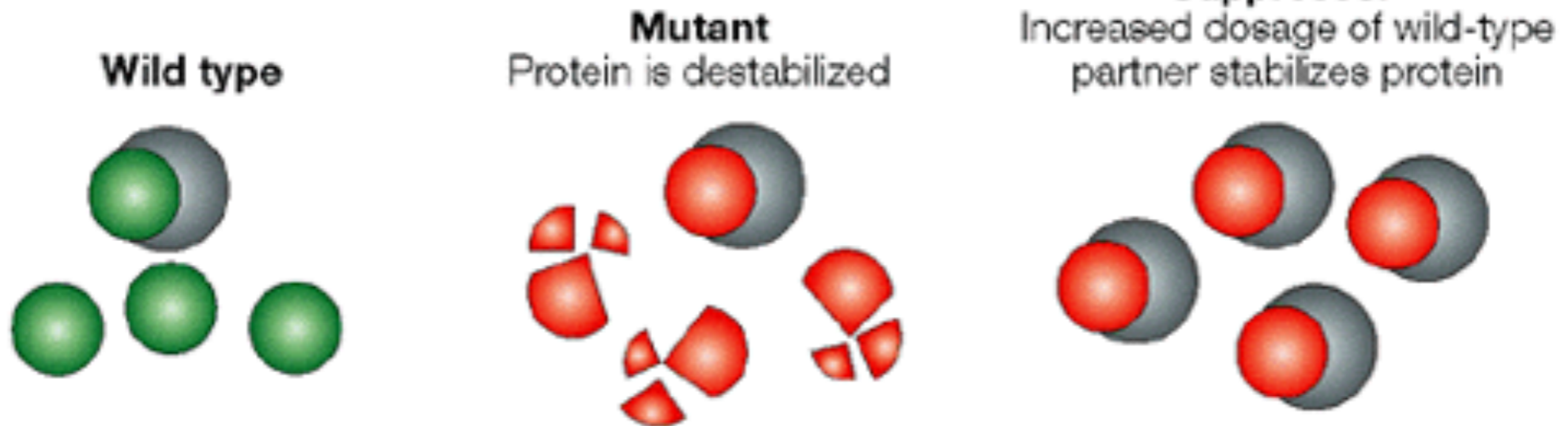


Forsburg 2001 Nat Rev in Genet

genetics offers unique tools for discovering gene function

sc-Based Perturb screenings

Dosage suppressor: rescues in high copy

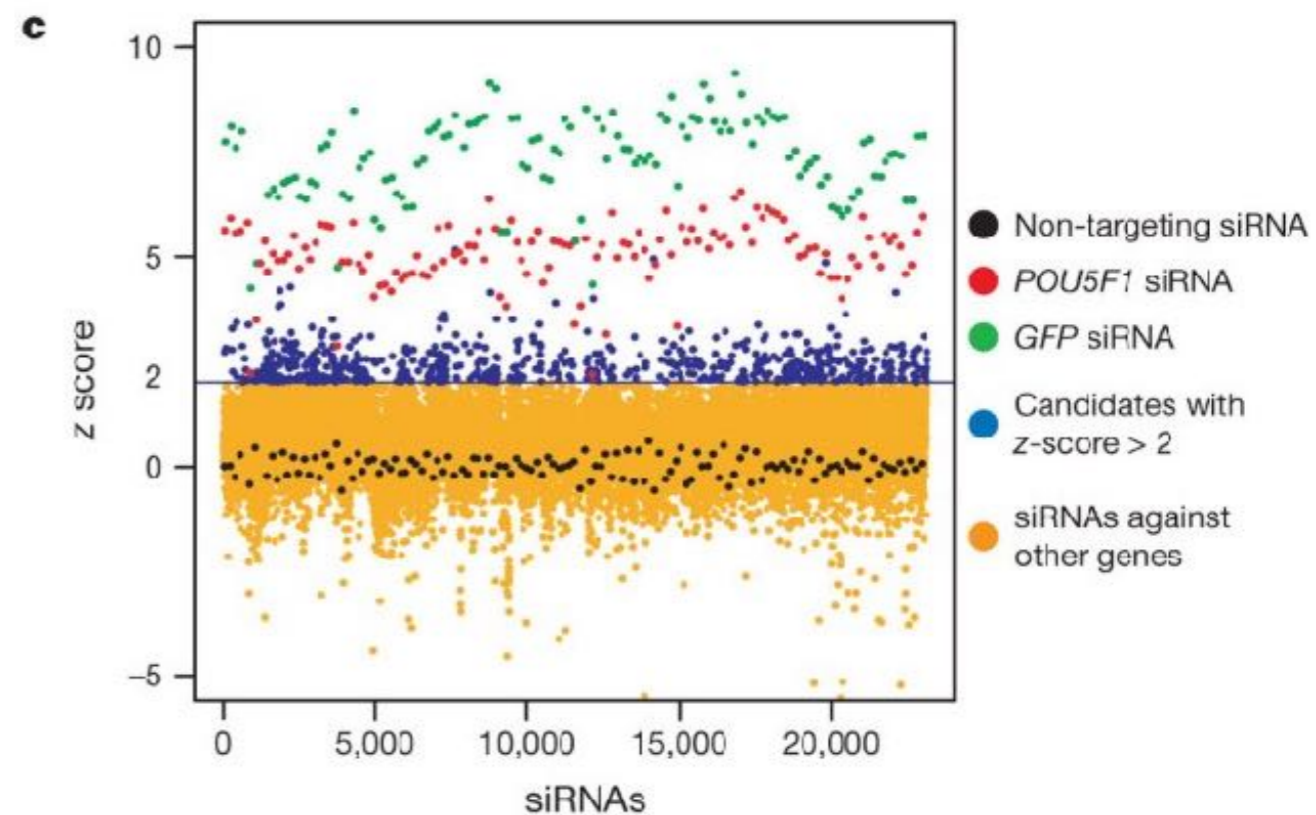
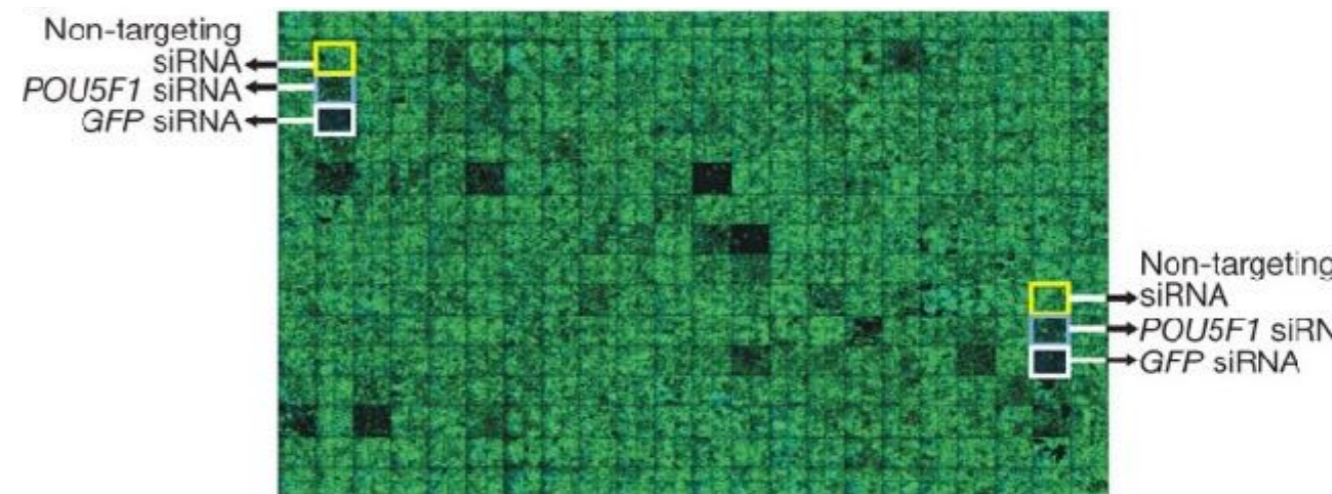
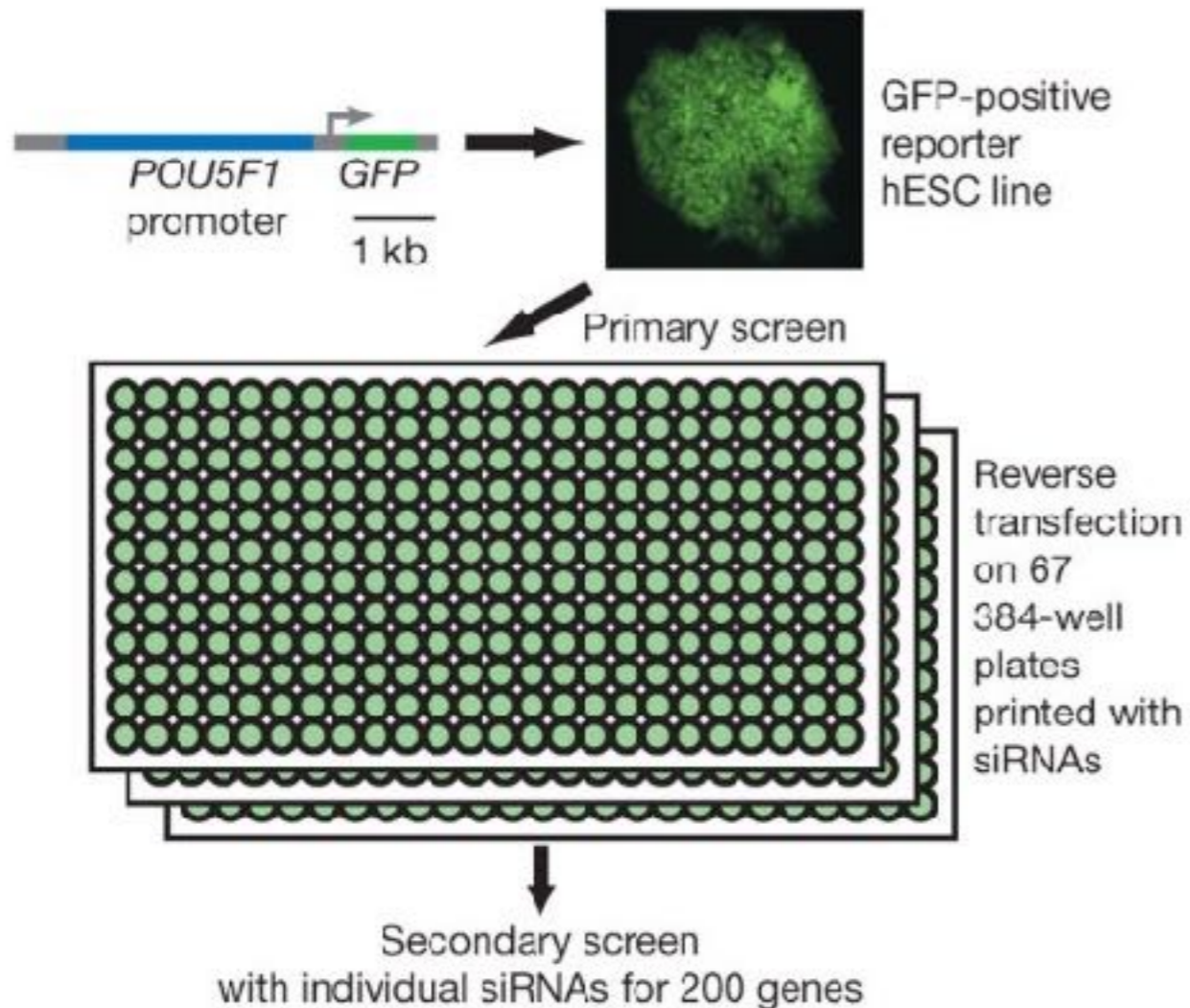


Interaction suppressor: allele specific, gene specific




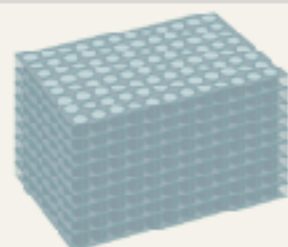






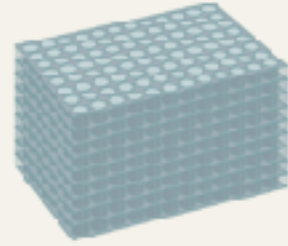

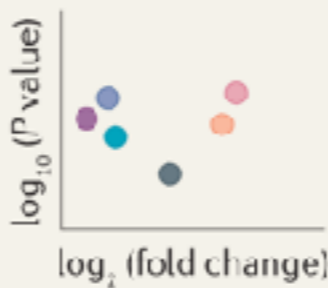









Forsburg 2001 Nat Rev in Genet

sc-Based Perturb screenings



genetic screens are often focused to the assessment of one parameter

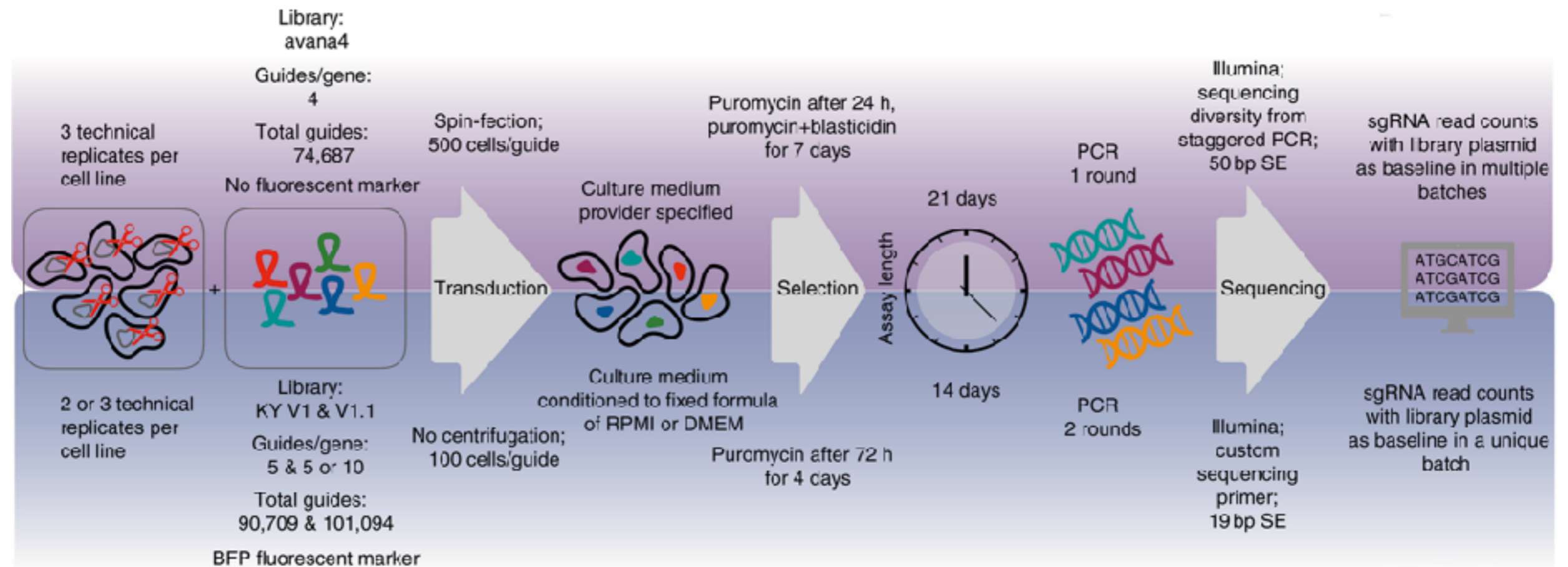
sc-Based Perturb screenings

	Pooled CRISPR screen	Arrayed CRISPR screen																				
Model Cell lines, primary cells, organoids, model organisms (in vivo screens)	Cas9-expressing cells in bulk 	Cas9-expressing cells in separate wells 																				
Perturbation CRISPR knockout, interference, activation, base editing, prime editing	CRISPR gRNA library  → Cells perturbed in bulk 	Cells perturbed in separate wells  → 																				
Challenge Cell survival and proliferation, drug treatment and resistance, virus/pathogen infection, metabolic challenges	 → Cells challenged in bulk (for example with a drug) 	Cells challenged in separate wells (for example with a drug) <div><div>A</div><div>B</div><div>C</div><div>D</div></div> → 																				
Read-out Sequencing-based counting of gRNA frequencies, single-cell RNA sequencing, multi-omics profiling, imaging	 → gRNA sequencing → gRNA enrichment/depletion 	Molecular phenotyping <table><tr><th>A</th><th>B</th><th>C</th><th>D</th><th>Drugs</th></tr><tr><td colspan="4"></td><td>Knockouts</td></tr><tr><td colspan="4"></td><td>Genes</td></tr><tr><td colspan="4"></td><td></td></tr></table>	A	B	C	D	Drugs					Knockouts					Genes					
A	B	C	D	Drugs																		
				Knockouts																		
				Genes																		
																						

Bock et al. 2022 Nat Rev Met Prim

sc-Based Perturb screenings

Broad Institute



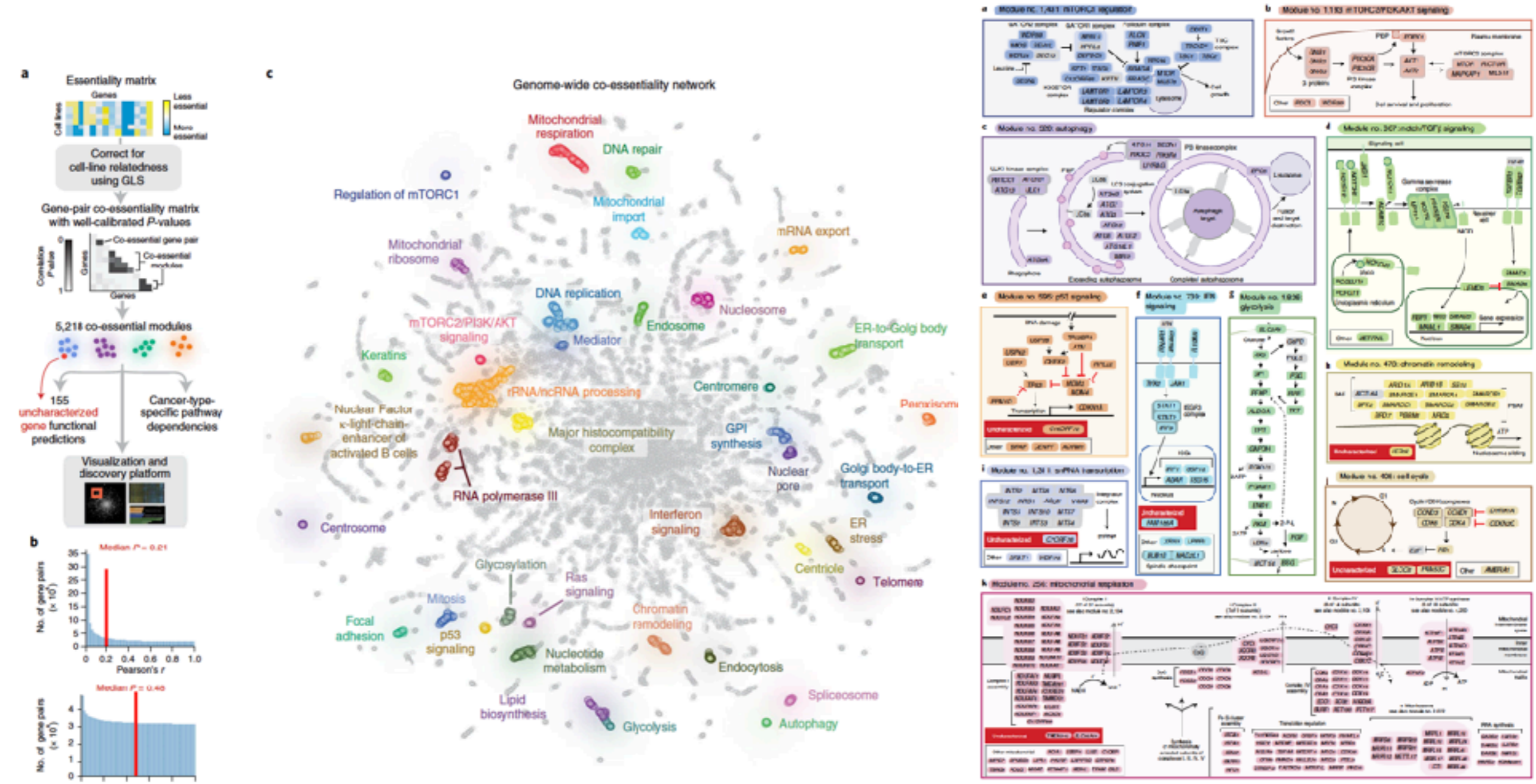
Sanger Institute

Tsherniak et al. 2017 Cell

Creating a genome-scale catalog of genetic vulnerabilities

A complete map of the vulnerabilities of cancer cell models is a key first step towards identifying therapeutics leads. Therefore, researchers are using genome-wide CRISPR loss-of-function screens to systematically identify essential genes across hundreds of human cancers

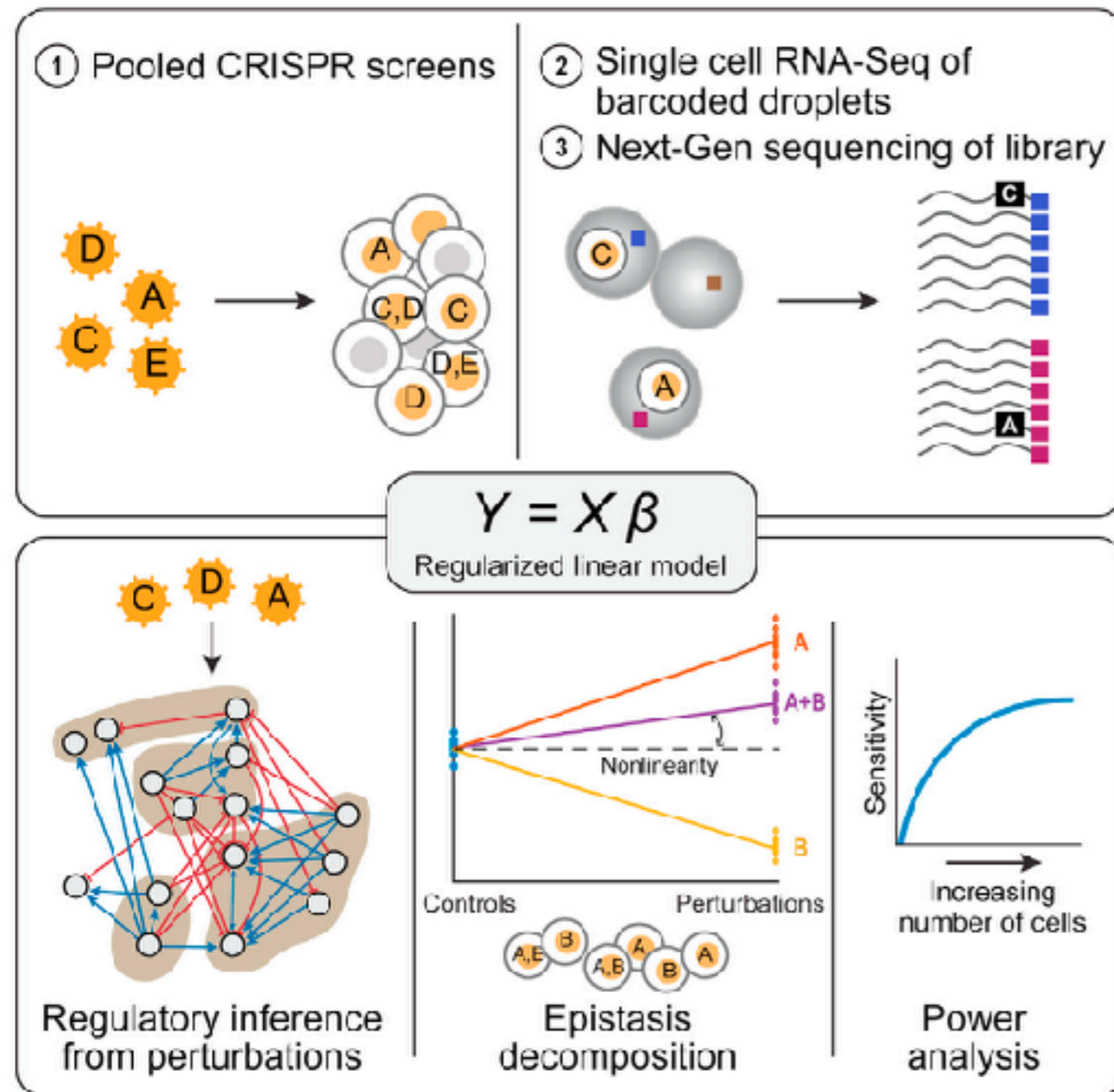
sc-Based Perturb screenings



Wainberg et al. 2021 Nat Genetics

Dep_Maps allow the inference of co-functional gene networks (with no priors)

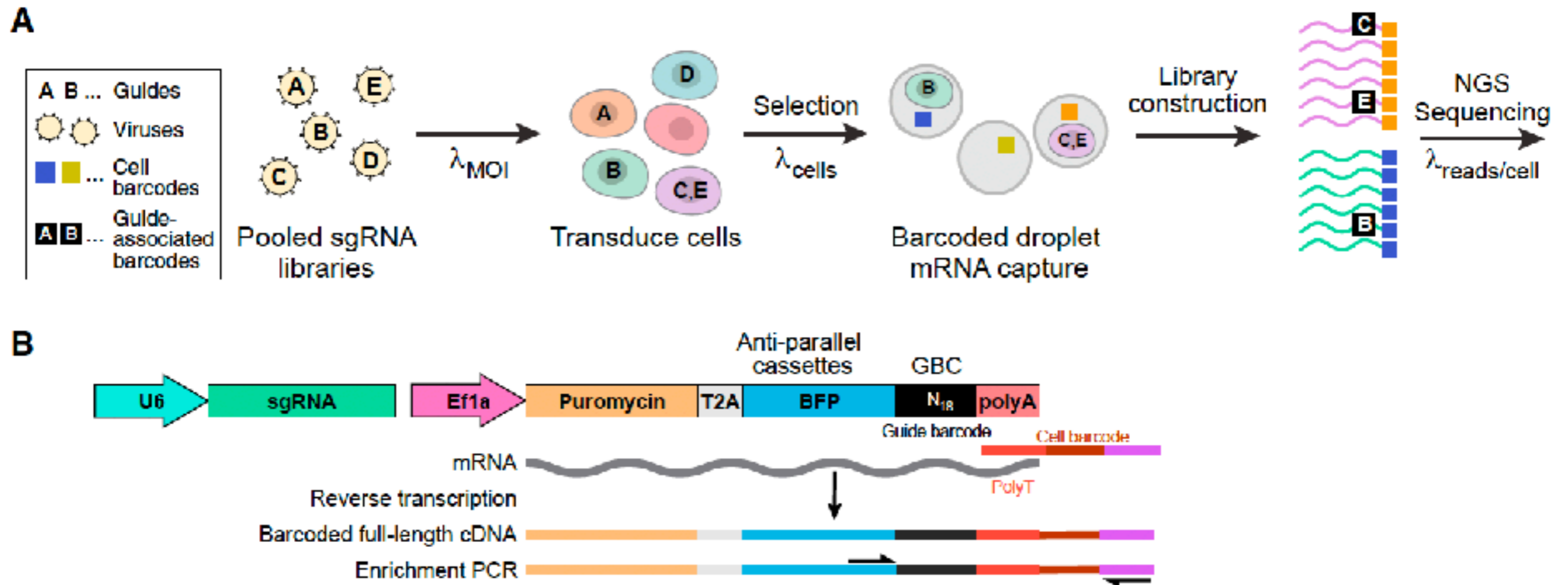
sc-Based Perturb screenings



Perturb-seq (also known as **CRISP-seq** and **CROP-seq**) refers to a high-throughput method of performing scRNA-seq on pooled genetic perturbation screens.

Perturb-seq combines multiplexed CRISPR mediated gene inactivations with single cell RNA sequencing to assess comprehensive gene expression phenotypes for each perturbation. Inferring a gene's function by applying genetic perturbations to KD or KD a gene and studying the resulting phenotype is known as reverse genetics. Perturb-seq is a reverse genetics approach that allows for the investigation of phenotypes at the level of the transcriptome, to elucidate gene functions in many cells, in a massively parallel fashion.

sc-Based Perturb screenings



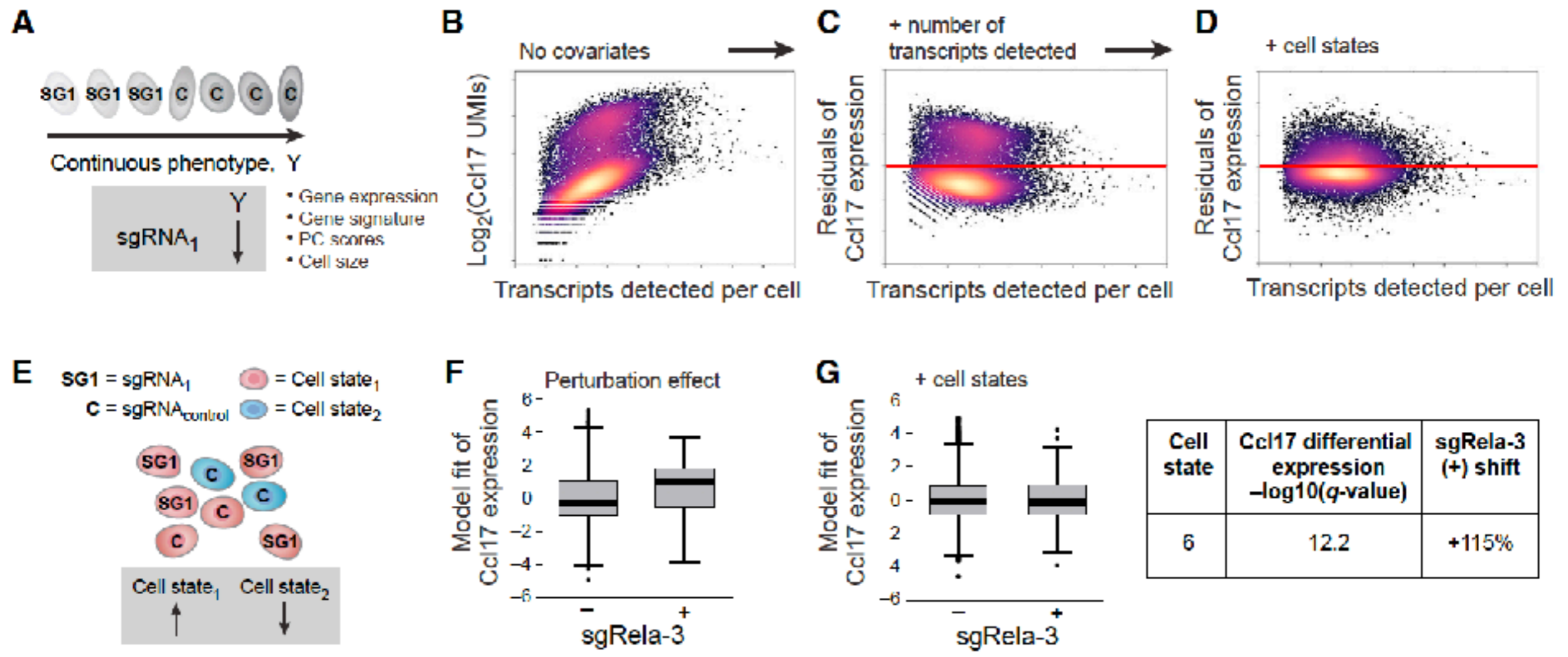
Dixit et al. 2016 Cell

Perturb-seq combines a pooled CRISPR screen with scRNA-seq by encoding the identity of the perturbation on an expressed guide barcode (GBC)

so each cell has 2 barcodes one for cell identity (CBC) and the other for perturbation identity (GBC)

sc-Based Perturb screenings

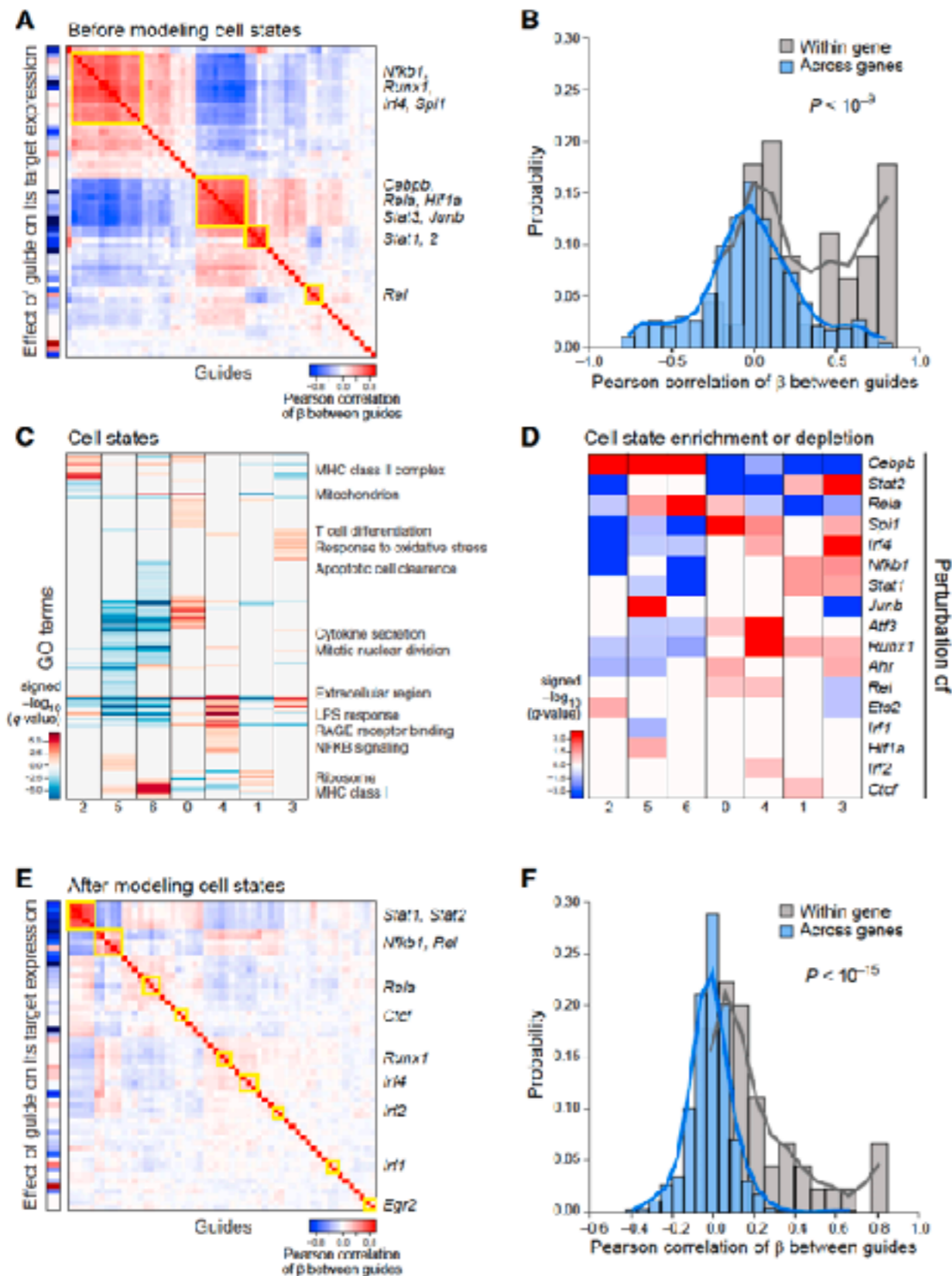
Bone marrow-derived dendritic cells (BMDCs), they targeted 24 transcription factors (TFs) in ctrl and LPS treated cells



Dixit et al. 2016 Cell

Allows to discriminate effects on cell state proportions from those on the expression of a given gene

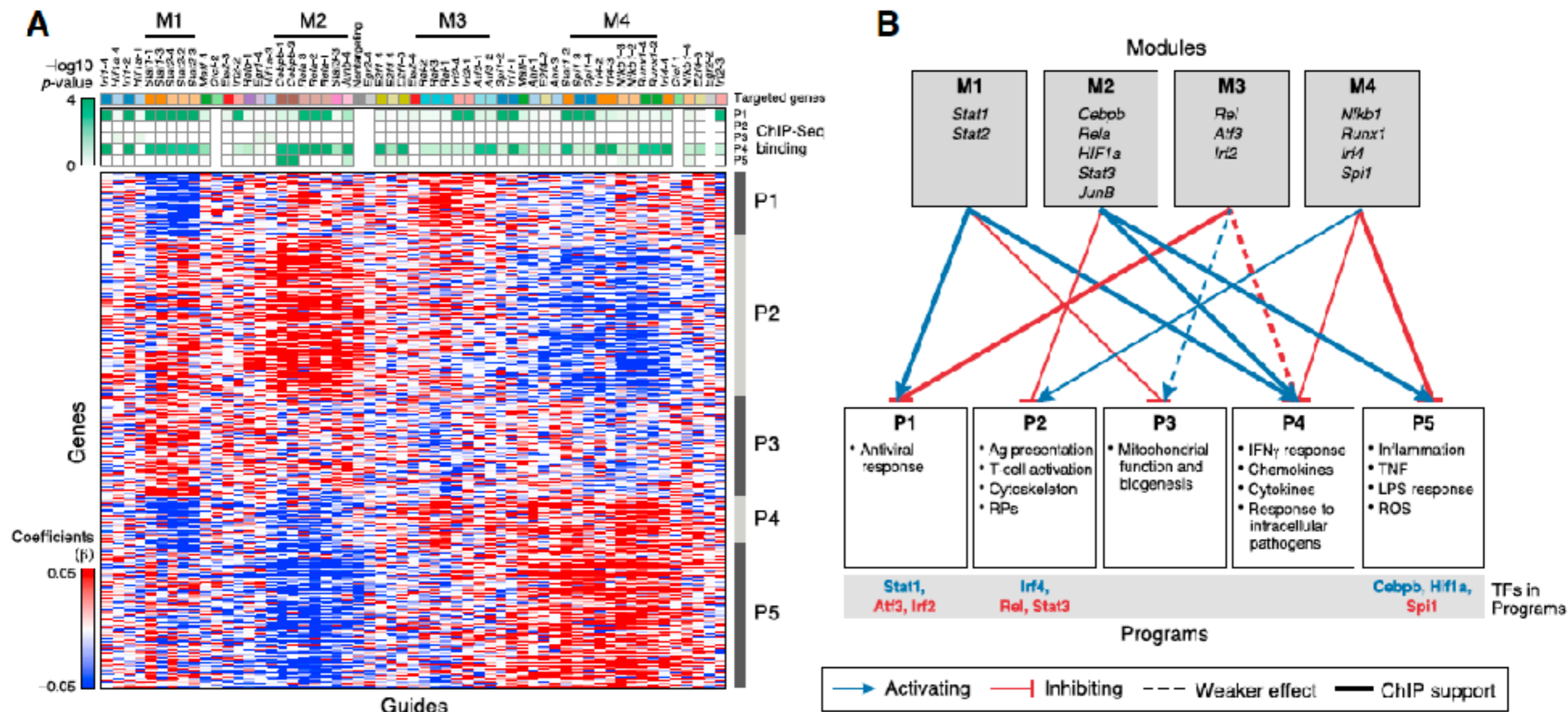
sc-Based Perturb screenings



Authors used the regulatory effects of each perturbed TF on each gene, to group TFs into modules by their similar regulatory effects and to group genes into programs by how they are affected by the perturbations.

Note this structure is largely dictated by effects on cell states

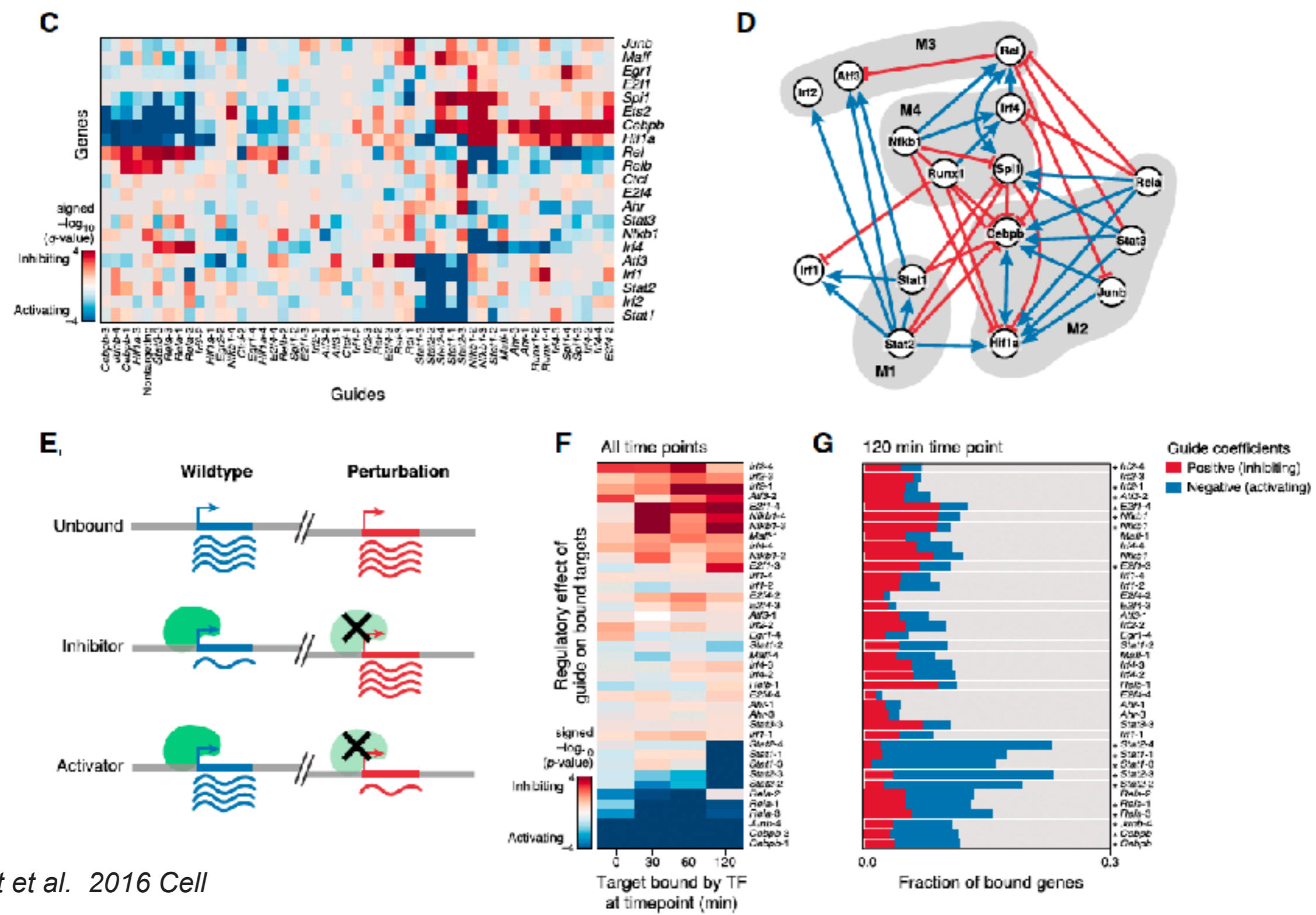
sc-Based Perturb screenings



Dixit et al. 2016 Cell

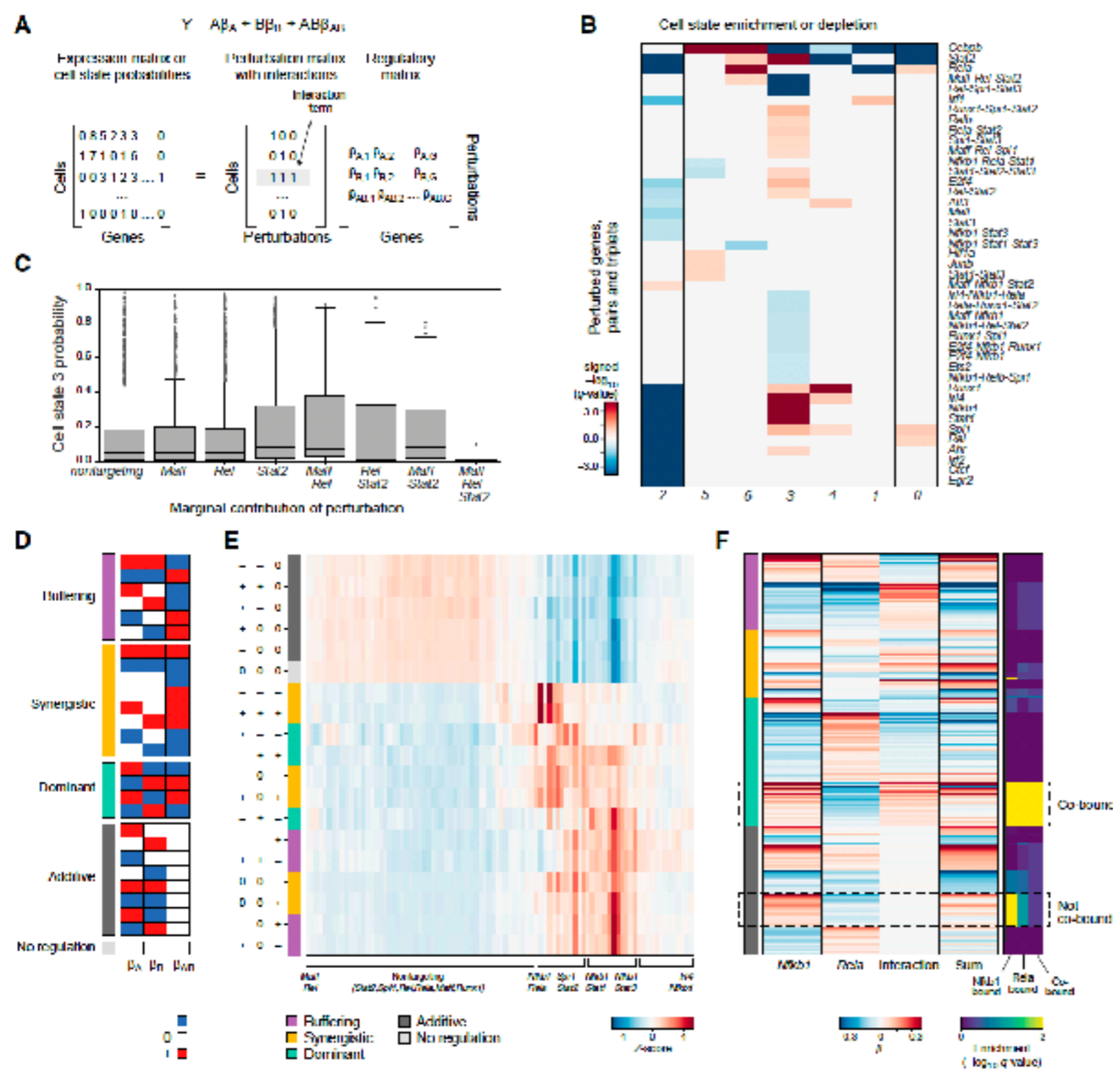
The Genetic Circuit Is Supported by TF Binding Profiles

sc-Based Perturb screenings



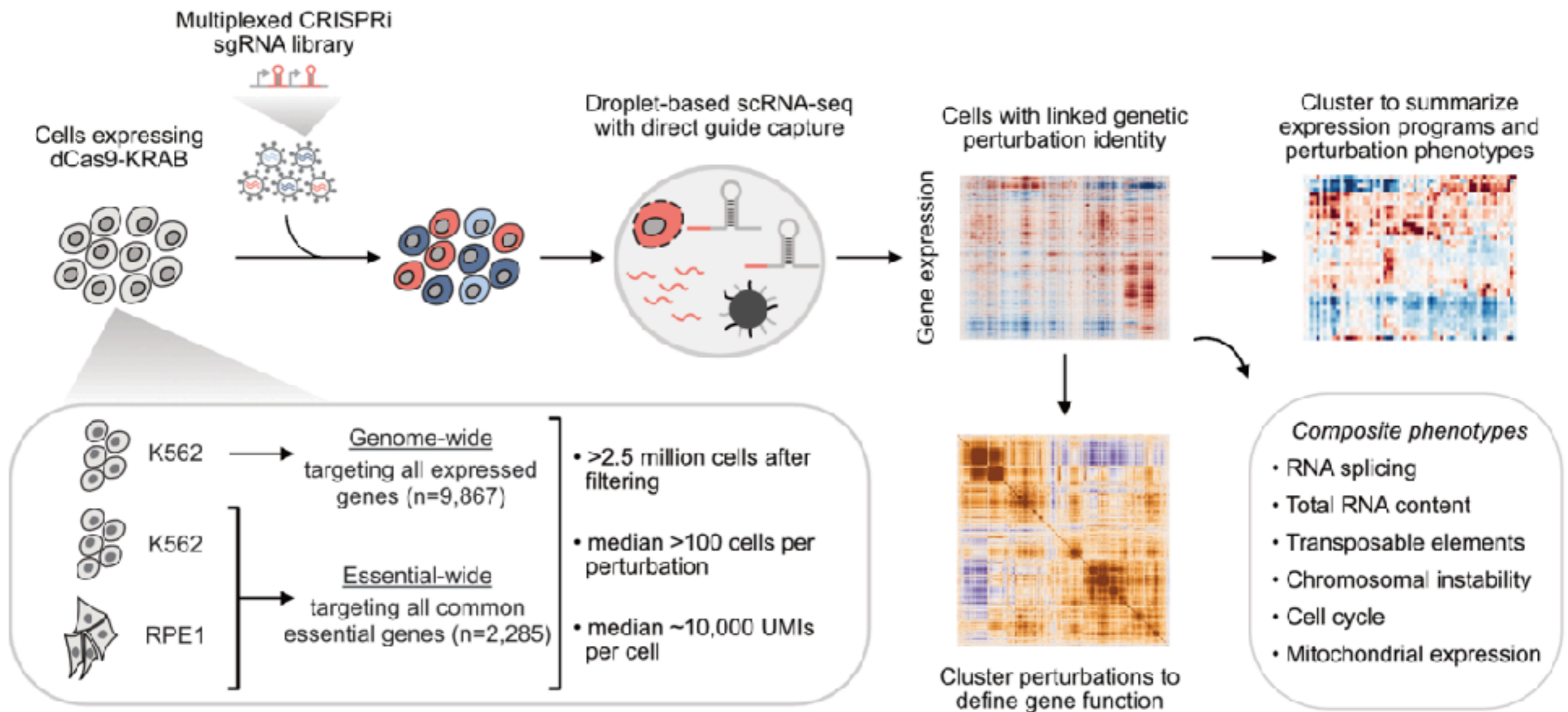
Dixit et al. 2016 Cell

sc-Based Perturb screenings



Genetic Interactions between TFs in BMDCs can be inferred by looking at double or triple infected cells

sc-Based Perturb screenings



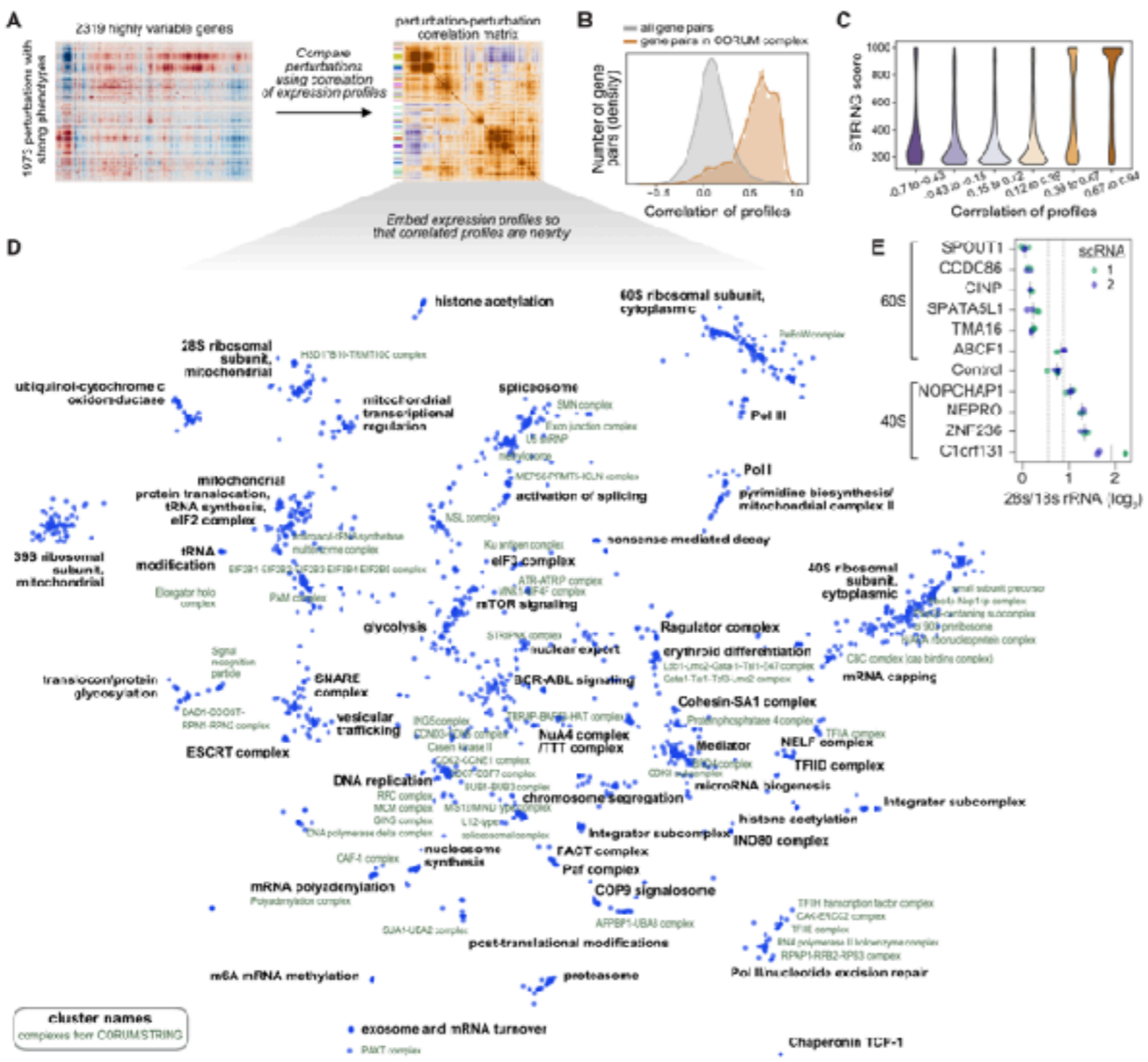
Replogle et al. 2021 Biorxiv

Whole Genome Perturb Seq

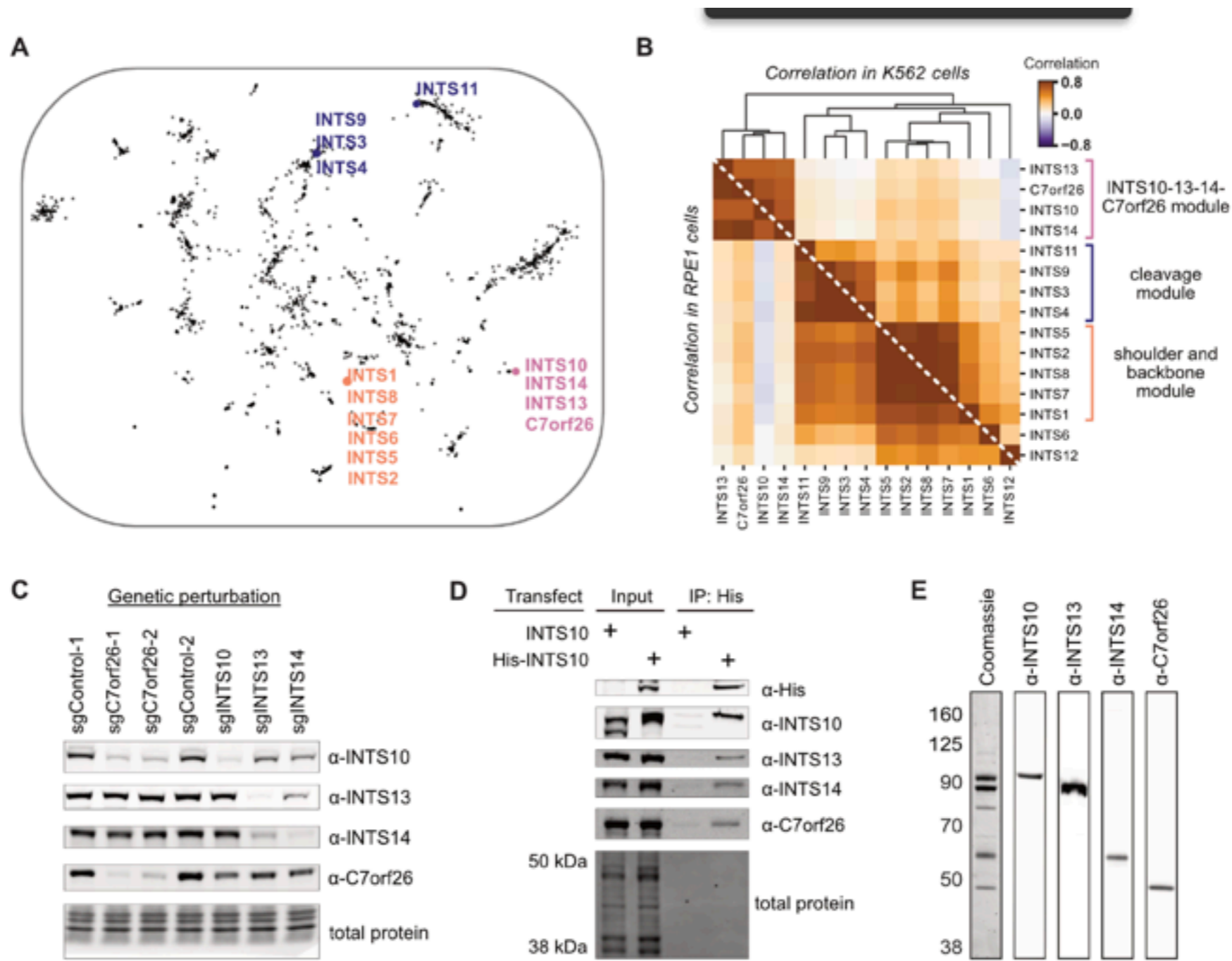
Whole Genome Perturb Seq

Whole genome
Perturb_seq allows
the inference of co-
functional gene
networks active in a
give cell type looking
at perturbation
correlation only

(with no priors)



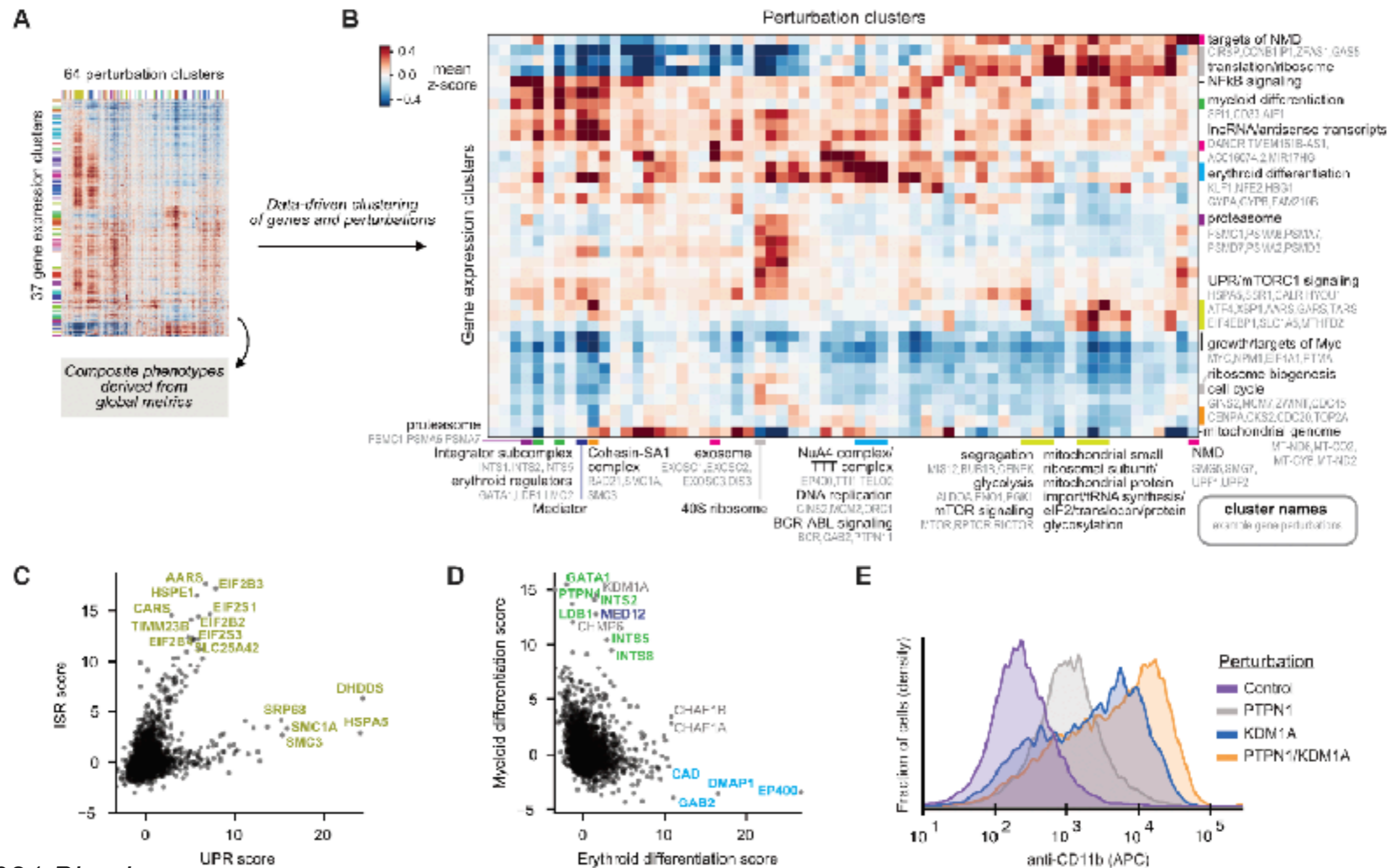
Whole Genome Perturb Seq



Replogle et al. 2021 Biorxiv

Whole Genome Perturb Seq

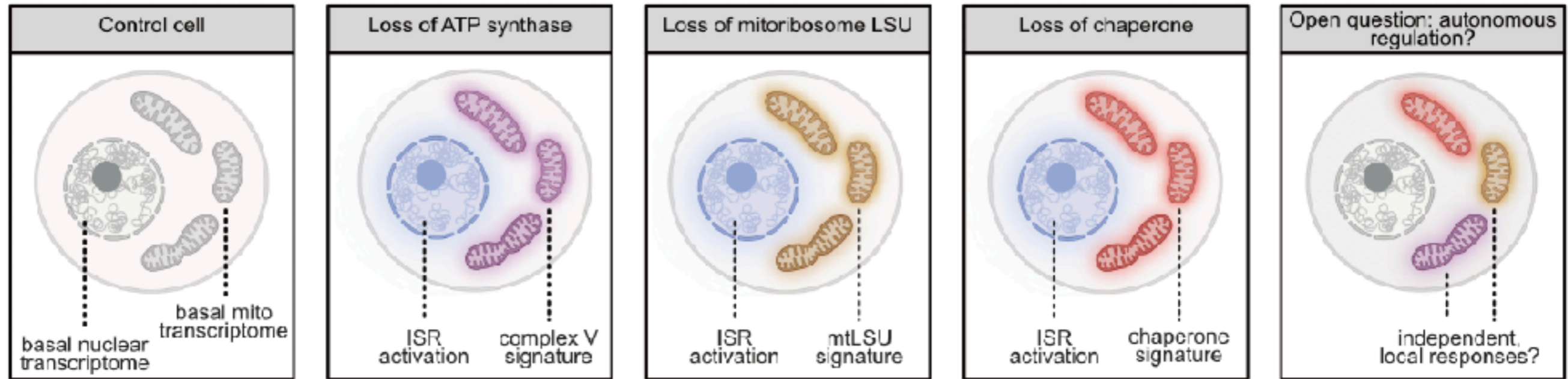
Summarizing genotype-phenotype relationships with Perturb-seq



Replogle et al. 2021 Biorxiv

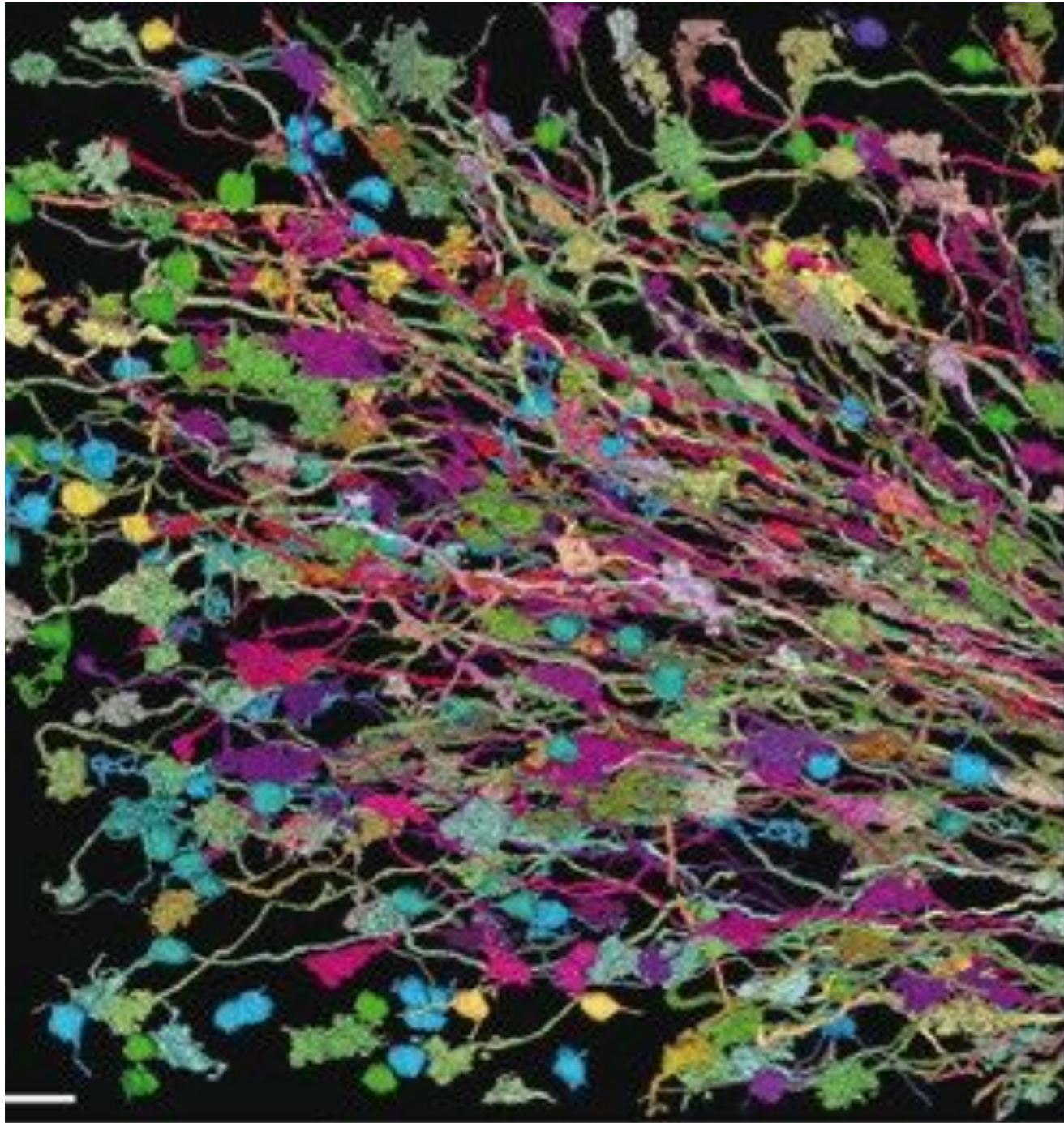
Data-driven definition of transcriptional programs

sc-Based Perturb screenings



Replogle et al. 2021 Biorxiv

Spatial Omics Techniques



Single-cell analyses, especially RNA sequencing and other genomics modalities, have been transformative in revealing new biology.

However, these approaches fail to provide a complete picture of biological processes, as contextual information on cellular location is lost.

New technologies leveraging multiplexed fluorescence, DNA, RNA and isotope labeling enable the detection of tens to thousands molecular biomarkers within their native spatial context.

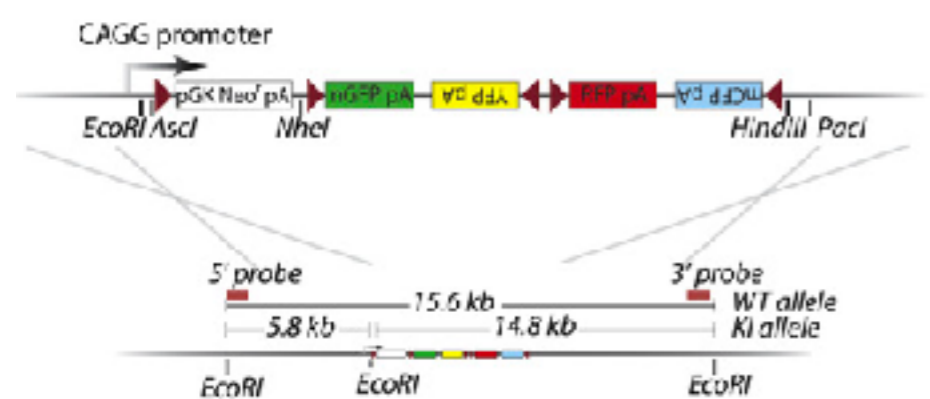
Spatial Omics Techniques

- (1) optical barcoding methods for tracking cell subclones
- (2) spatial proteomics methods
- (3) spatial transcriptomic methods
- (4) spatial metabolomics methods
- (5) computational integration of these modalities

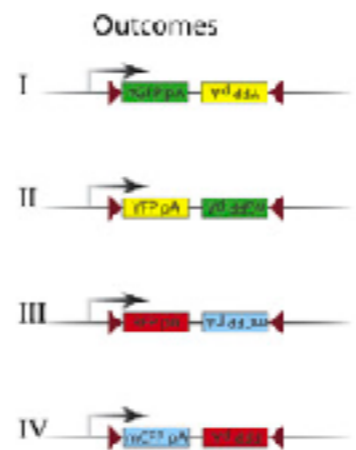
Spatial Omics Techniques

Tracking the spatiotemporal fate of live cells in their tissue context

Rosa26 locus in Mouse, Chr6

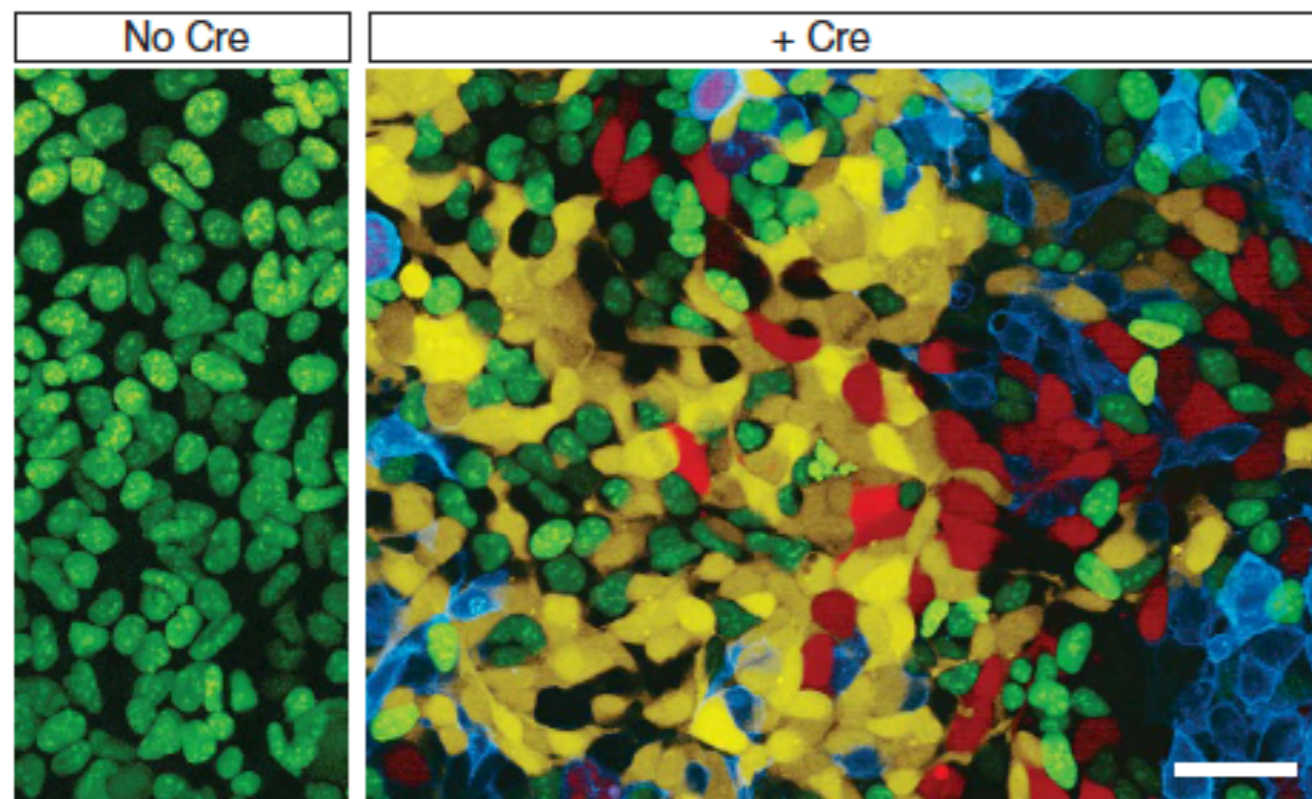


Cre recombination



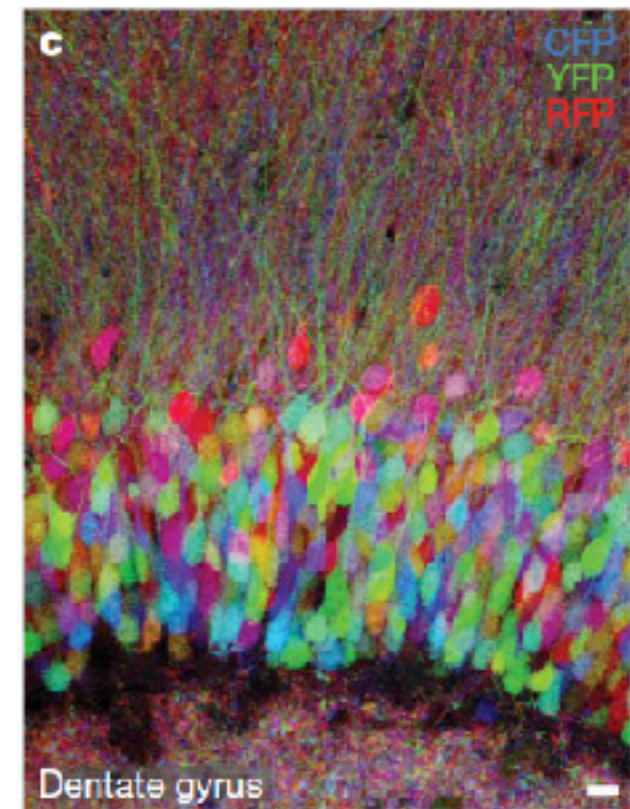
R26R-Confetti knock-in strategy.

Brainbow2.1 encoding four fluorescent proteins was inserted into the Rosa26 locus. Upstream, the strong CAGG promoter, a LoxP site, and a neomycin resistance roadblock cassette were inserted. Upon cre activation, the neomycin roadblock is excised, while the brainbow2.1 recombines in a random fashion to four possible outcomes.



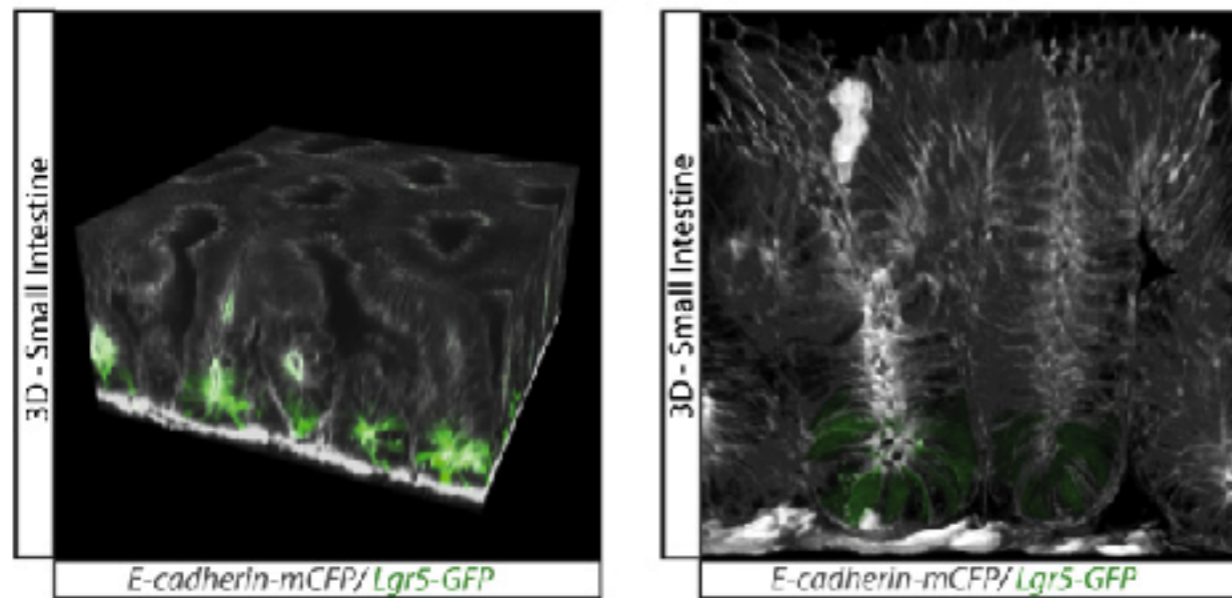
a XFP combinations

Outcome for each copy			Resulting colour
1	2	3	
C	C	C	Blue
C	C	Y	Light blue
C	Y	Y	Blue-green
Y	Y	Y	Green
Y	Y	R	Light green
Y	R	R	Orange
R	R	R	Red
R	R	C	Magenta
R	C	C	Purple
R	C	Y	Grey



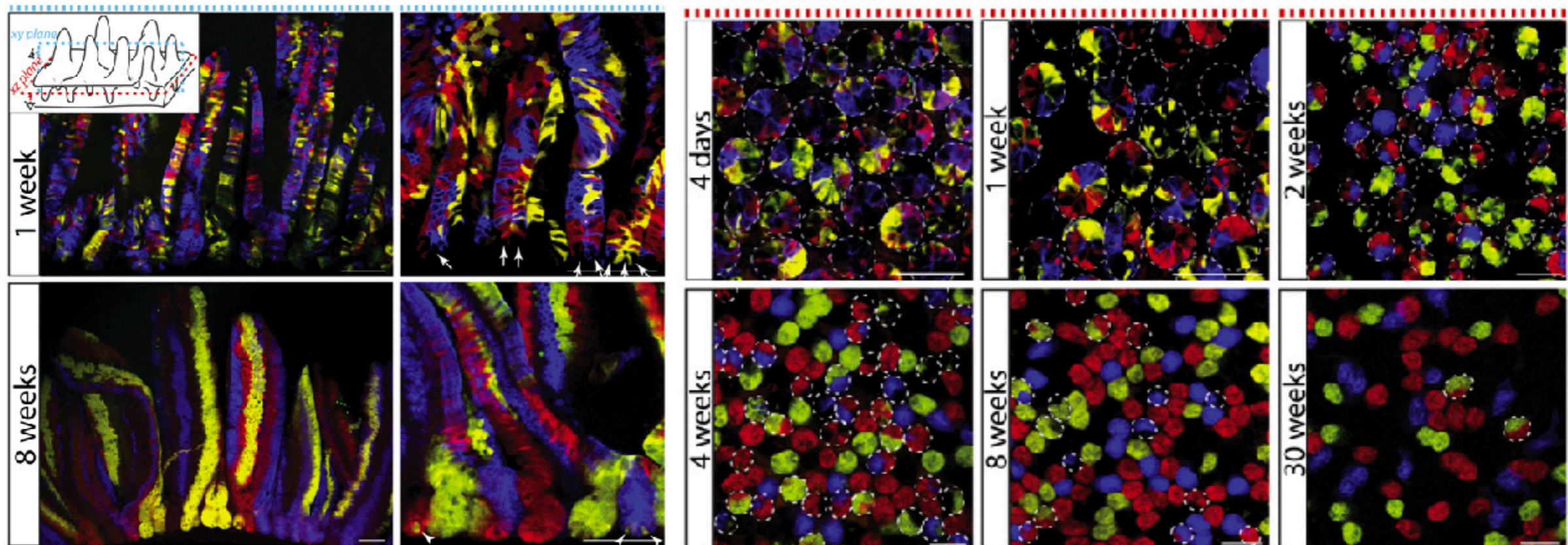
Spatial Omics Techniques

Tracking the spatiotemporal fate of live cells in their tissue context



Intestinal stem cells, characterized by high *Lgr5* expression, reside between Paneth cells at the small intestinal crypt base and divide every day.

The authors investigated the fate of stem cells in vivo.



Spatial Omics Techniques

increasing the number of detectable proteins in a spatial context

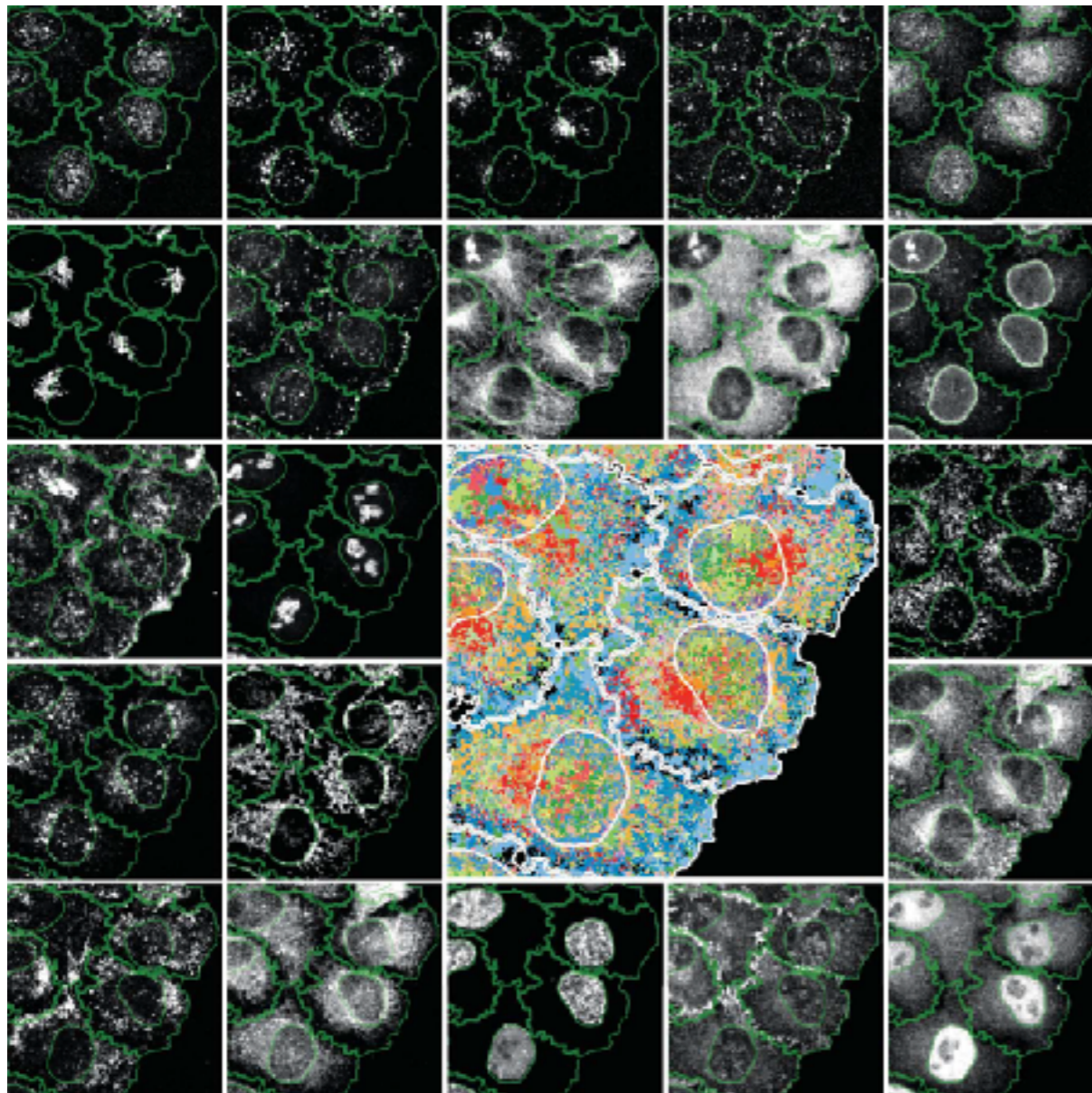
Spatial proteomics

					No. of targets	Tissue prep.
Iterative	mIHC				30	FFPE
	OPAI				10	FFPE
	CycIF				60	FFPE
Iterative (fluorescence)	REAdye, lease and REAdity				100 (100)	FFPE
	CODFX				60	FF* FFPE
	Immuno-SABER				10 (50)	Whole-mount FF* FFPE
	InSituPlex				10	FFPE
TOF-mass spectrometry	IMC				40 (100)	FF FFPE
	MIBI				40 (100)	FF FFPE
Sequencing	DSP				44 (100)	FF* FFPE

Spatial Proteomics

Spatial Omics Techniques

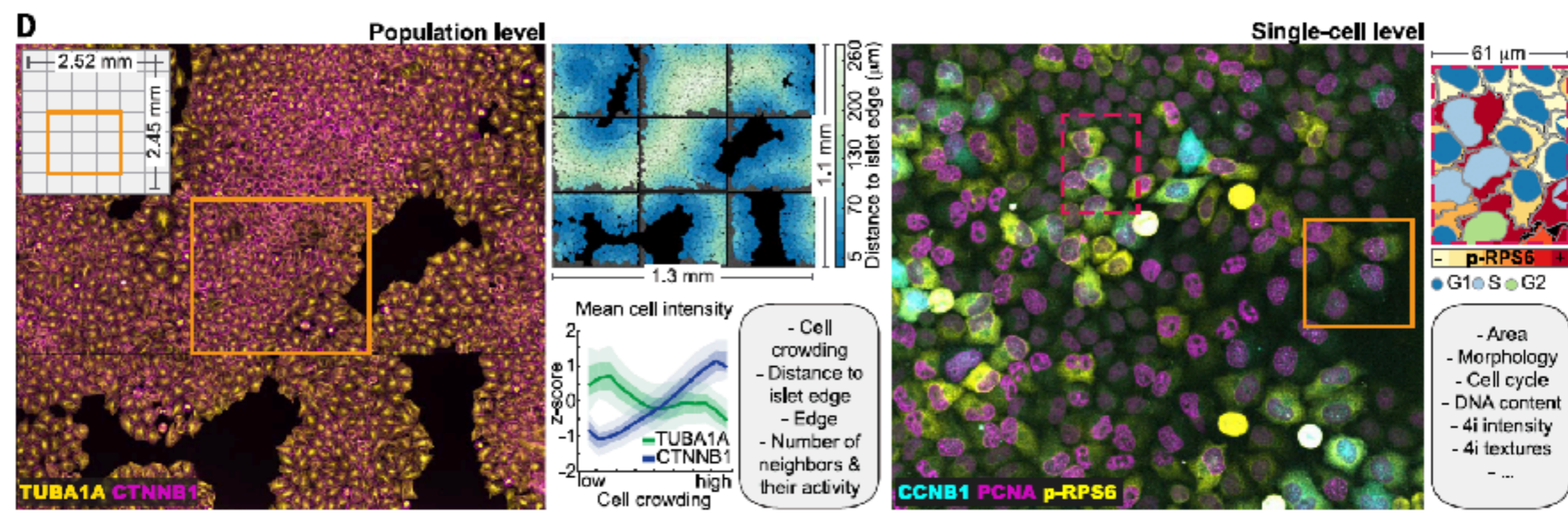
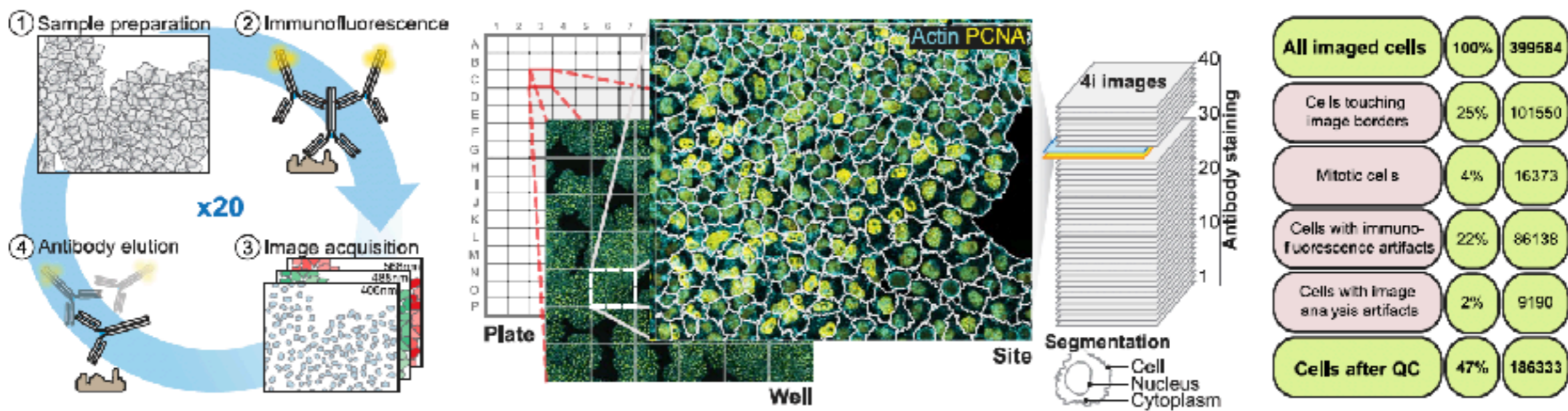
iterative staining (4i)



Iterative indirect immunofluorescence imaging (4i), 40-plex protein readouts from biological samples at high-throughput from the millimeter to the nanometer scale. This approach simultaneously captures properties apparent at the population, cellular, and subcellular levels, including microenvironment, cell shape, and cell cycle state. It also captures the detailed morphology of organelles, cytoskeletal structures, nuclear subcompartments, and the fate of signaling receptors in thousands of single cells in situ.

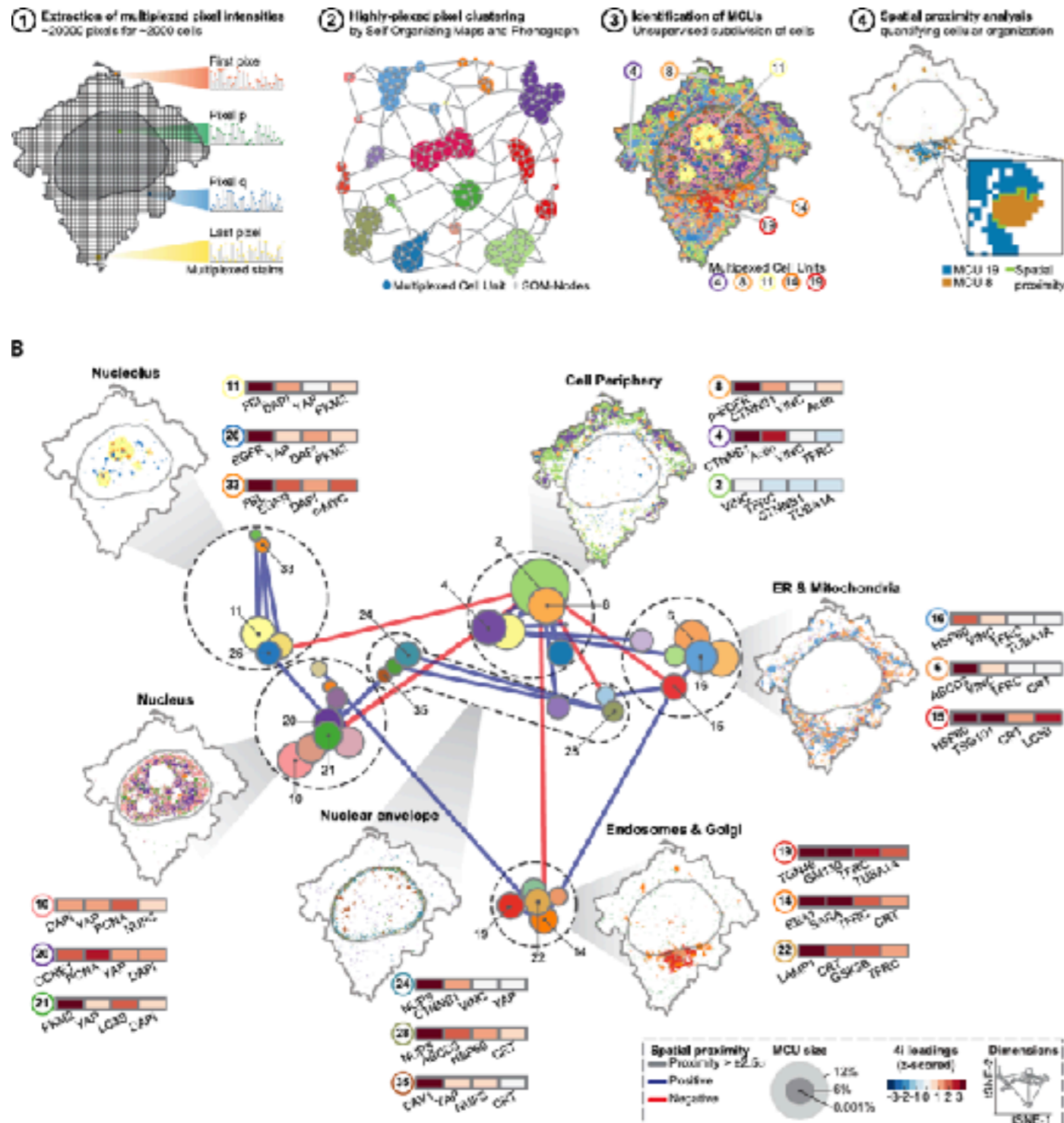
Spatial Omics Techniques

iterative staining (4i)



Spatial Omics Techniques

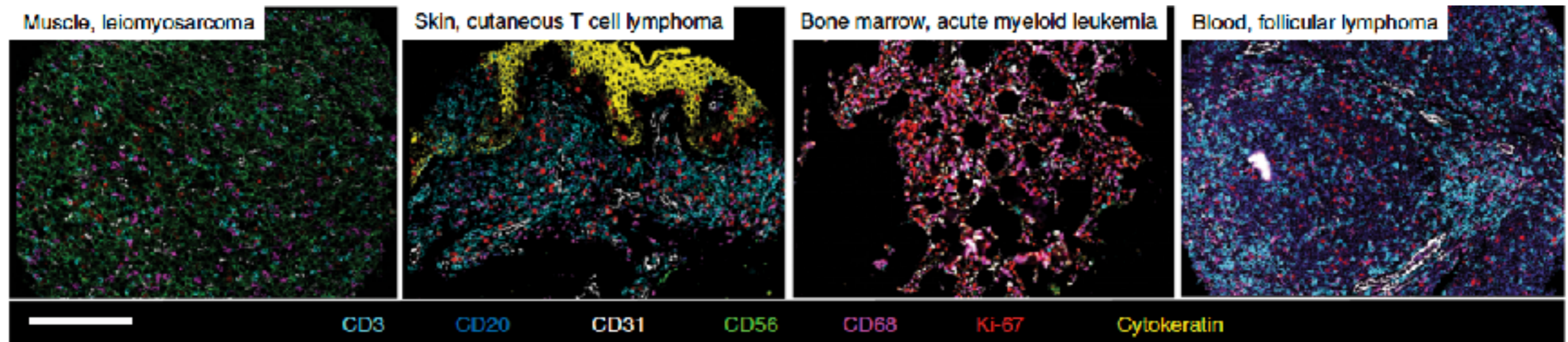
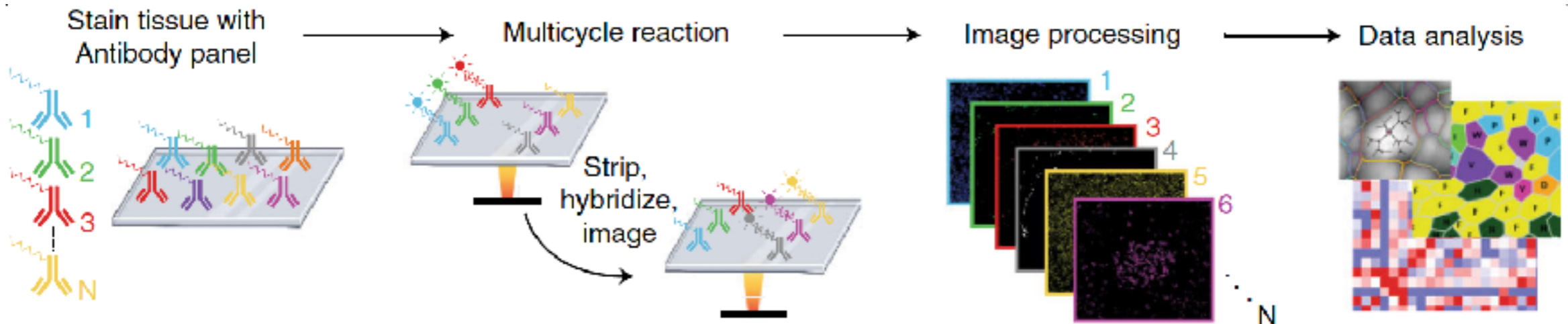
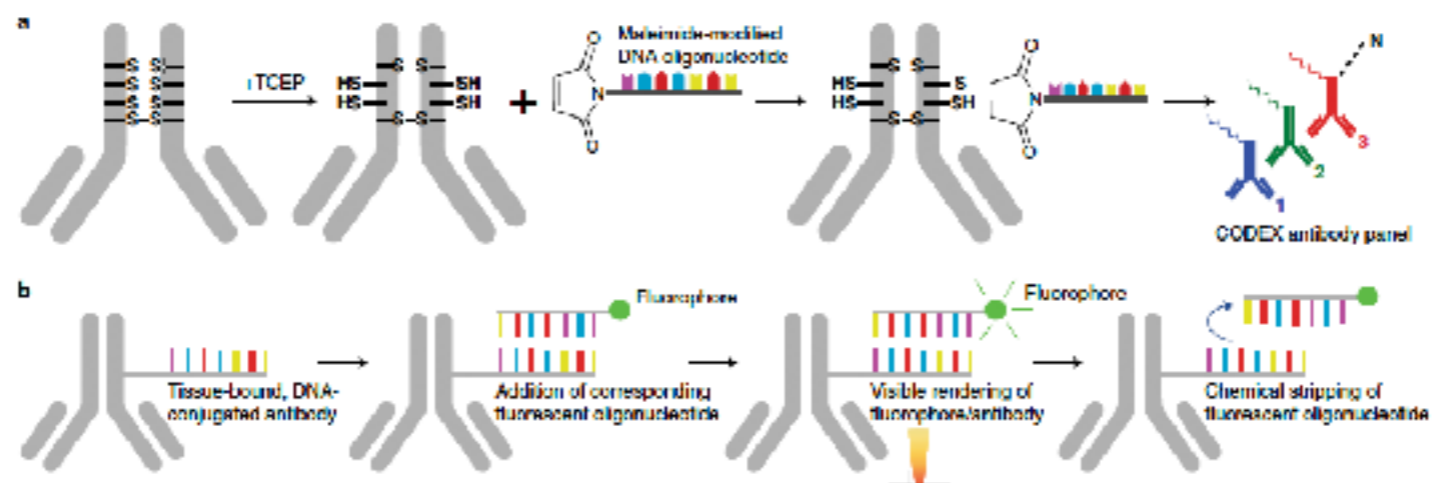
iterative staining (4i)



4i quantifies subcellular organization at high spatial detail in thousands of single cells based on multiplexed single-pixel profiles. 4i builds on a well-established highthroughput multivariate imaging platform

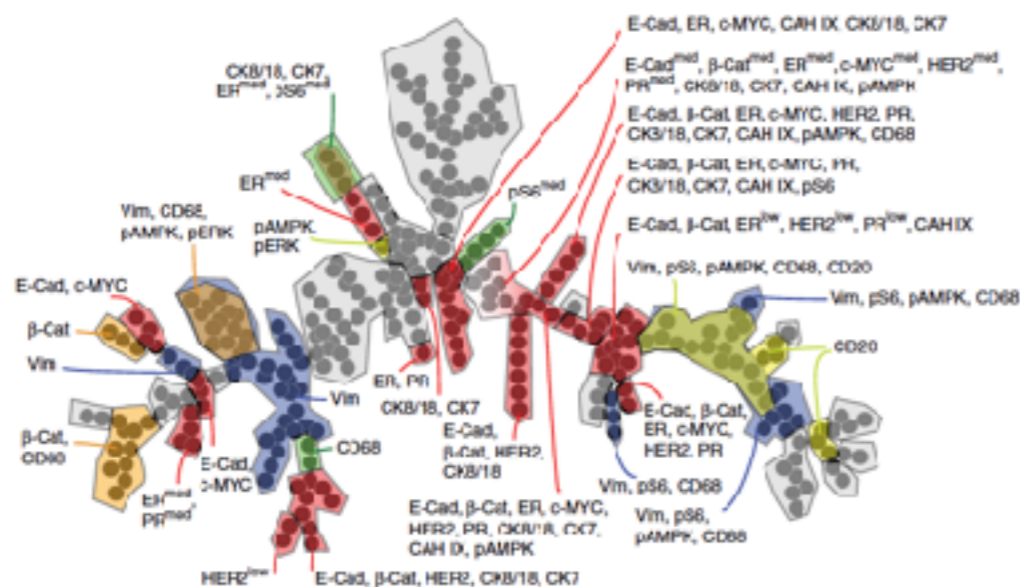
Spatial Omics Techniques

iterative staining CODEX



The flowchart illustrates the CyTOF workflow for single-cell proteomics, consisting of the following steps:

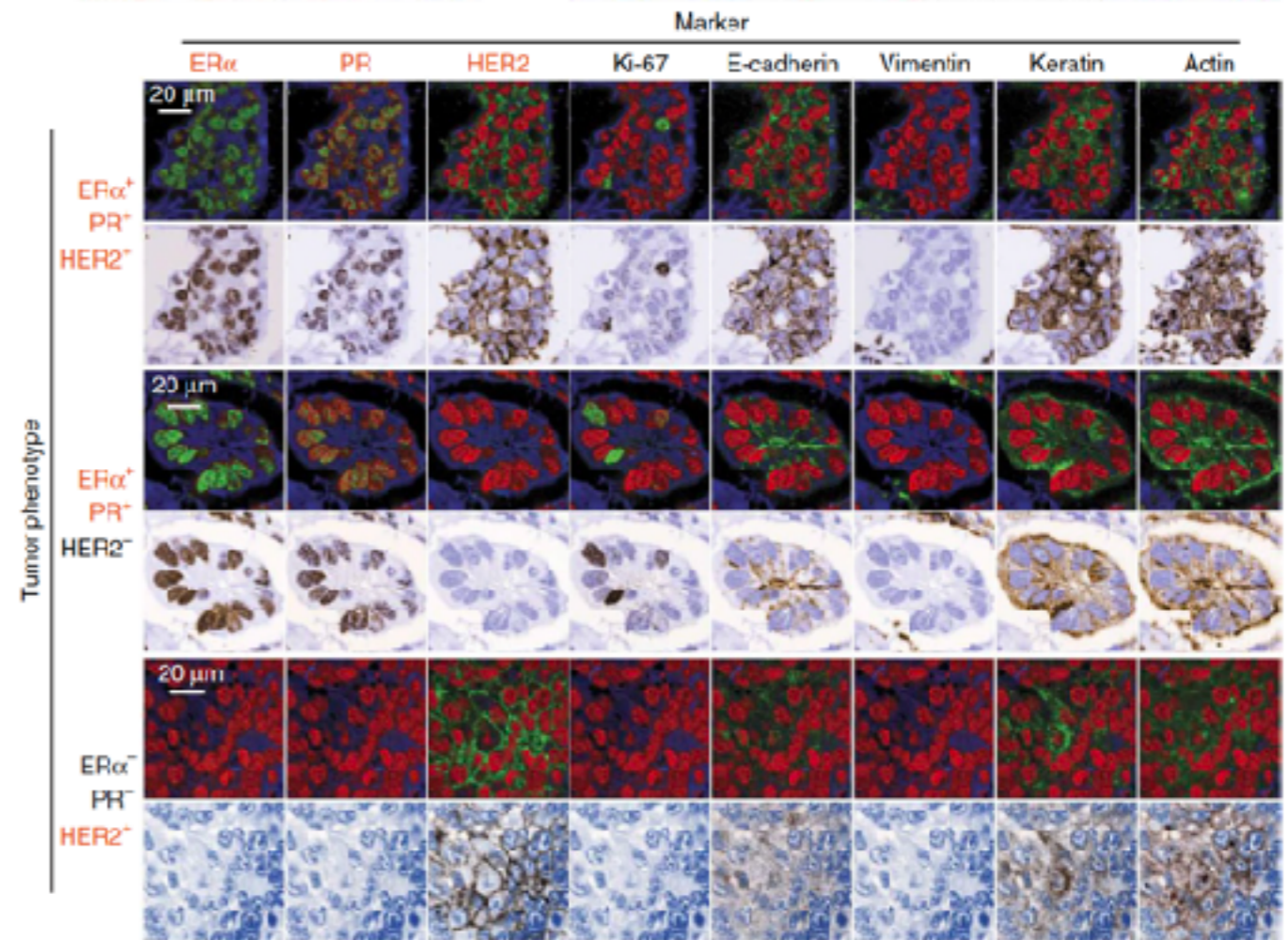
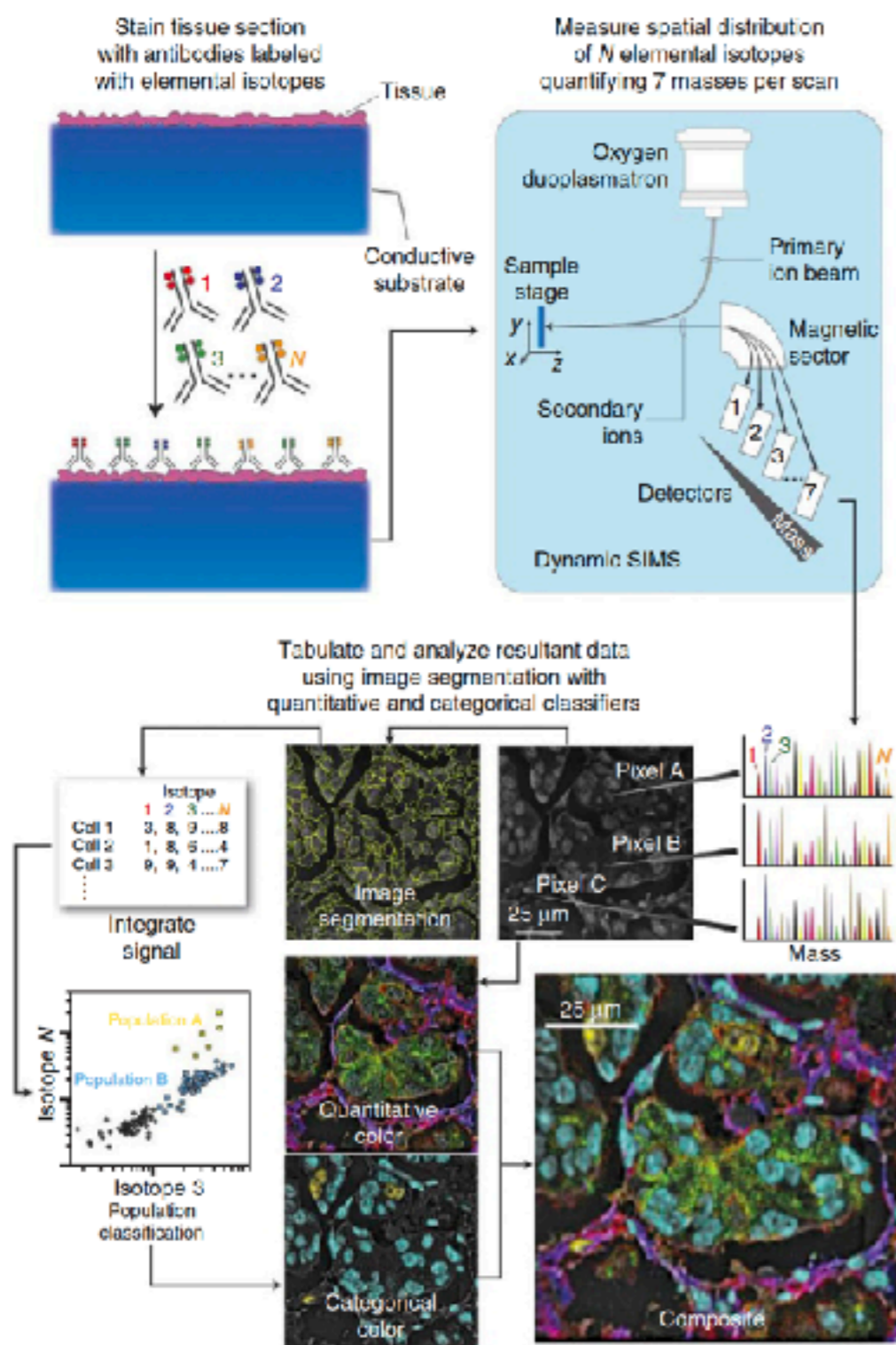
- Tissue or cell-line preprocessing**: The initial sample preparation stage.
- Marker staining with metal-labeled antibodies**: Cells are stained with antibodies labeled with different metals (represented by colored dots).
- Laser ablation coupled to mass cytometry**: A **UV laser** is used for ablation, and the released particles are analyzed by a **CyTOF mass cytometer**.
- Signal extraction of 32 measured markers**: The raw data is processed into 32 individual marker channels (shown as grayscale images).
- Data preprocessing and image assembly**: The individual channels are combined into a single multi-color image.
- Single-cell segmentation**: The assembled image is processed to identify and segment individual cells.
- Downstream data analysis**: The segmented data is used for biological analysis, such as identifying cell clusters (represented by a dendrogram and a cell diagram).



Giesen et al Nat Met 2018

Spatial Omics Techniques

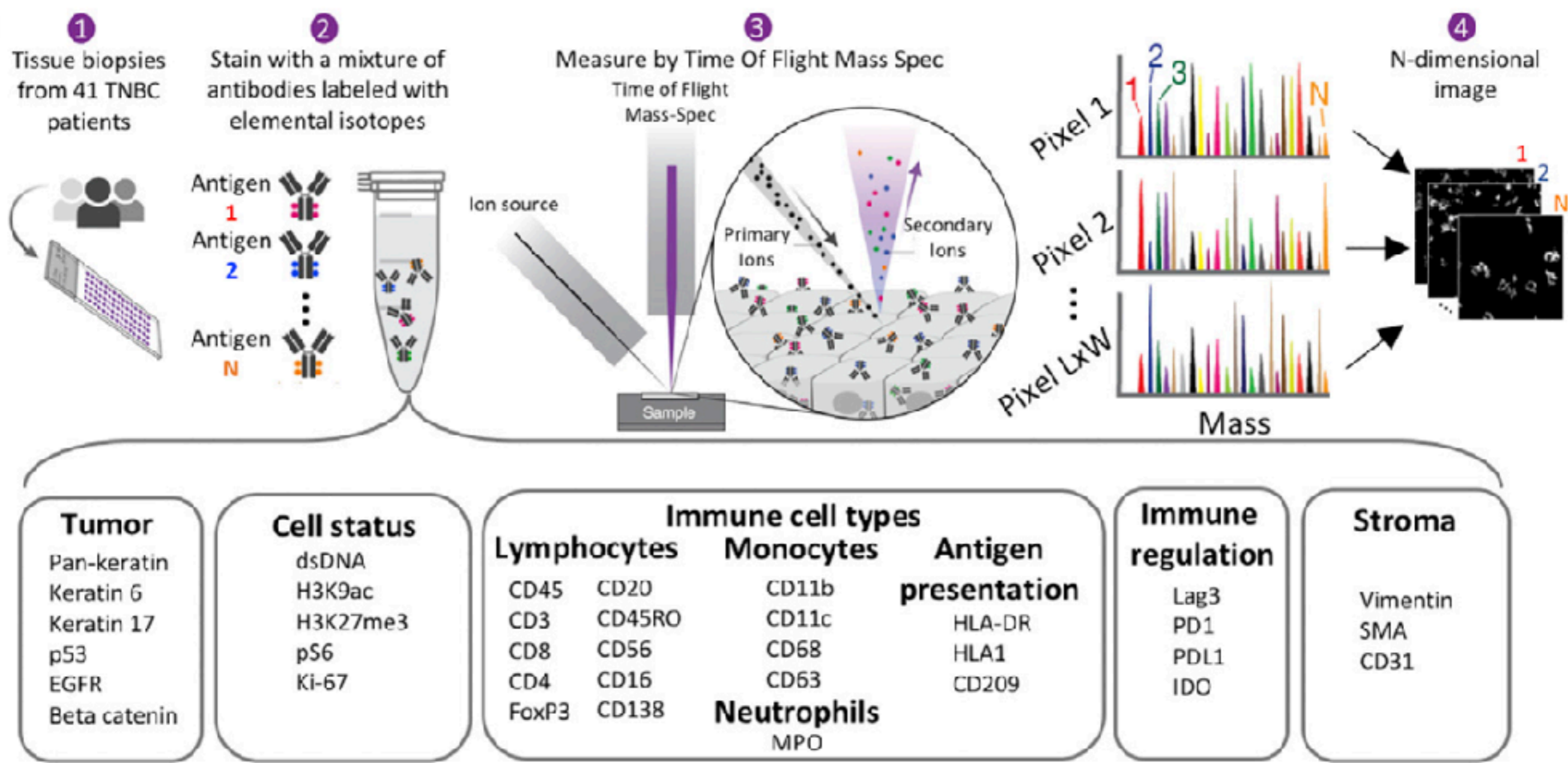
MS based methods - MIBI



multiplexed ion beam imaging (MIBI) can image upwards of 40 proteins in tissue. The primary ion beam is rastered across the tissue to generate secondary ions for detection by TOF mass spectrometry at defined spatial coordinates. In theory, the ion beam in MIBI can be reduced to a spot size of well below 500 nm in diameter

Spatial Omics Techniques

MS based methods - MIBI



Spatial Omics Techniques

FISH based methods

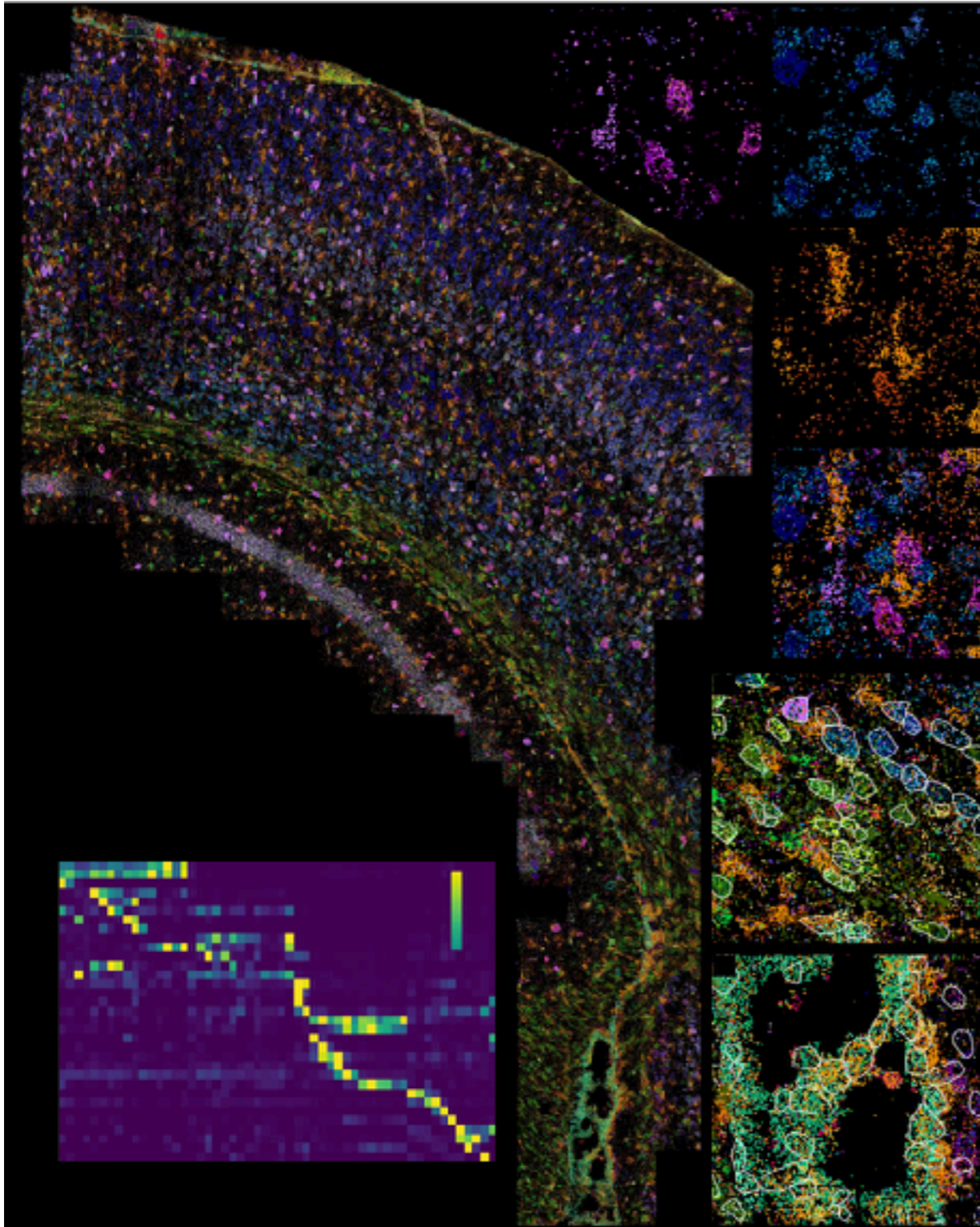
Spatial transcriptomics (FISH)

		Barcode	No. of targets	Tissue prep.				
smFISH		NA	<10	FF FFPE				
Spectral barcoding			32 (792)	NA				
Spatial barcoding			<10	NA				
	Round 1	Round 2	Round n					
osmFISH		Formamide		Formamide		NA	33	FF
MENFISH		Photobleach		Photobleach			10,000	FF
seqFISH		DNase		DNase			246	FF
seqFISH+		Formamide		Formamide			10,000	FF
RNAscope						NA	12	FF FFPE

Spatial Transcriptomics

Spatial Omics Techniques

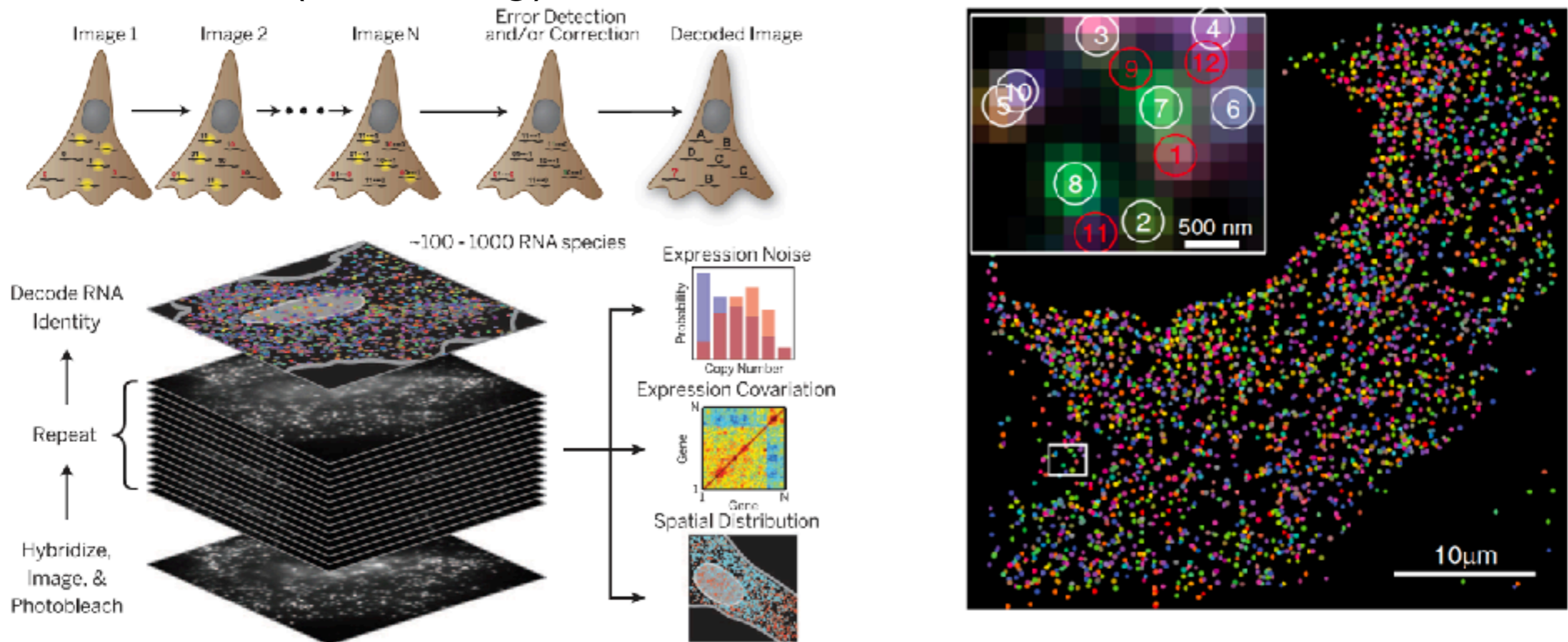
osmFISH



Sequential imaging can increase the multiplexing capacity of smFISH methods. Ouroboros single-molecule FISH (osmFISH) is a non-barcoded form of cyclic smFISH that targets transcripts in successive rounds of hybridization. Both osmFISH and other iterative fluorescence methods are efficient in the process of fluorophore stripping and quenching. While fluorophore brightness and tissue integrity may be marginally reduced with increasing numbers of hybridization rounds, samples generally maintain stability for an extended period of time.

Spatial Omics Techniques

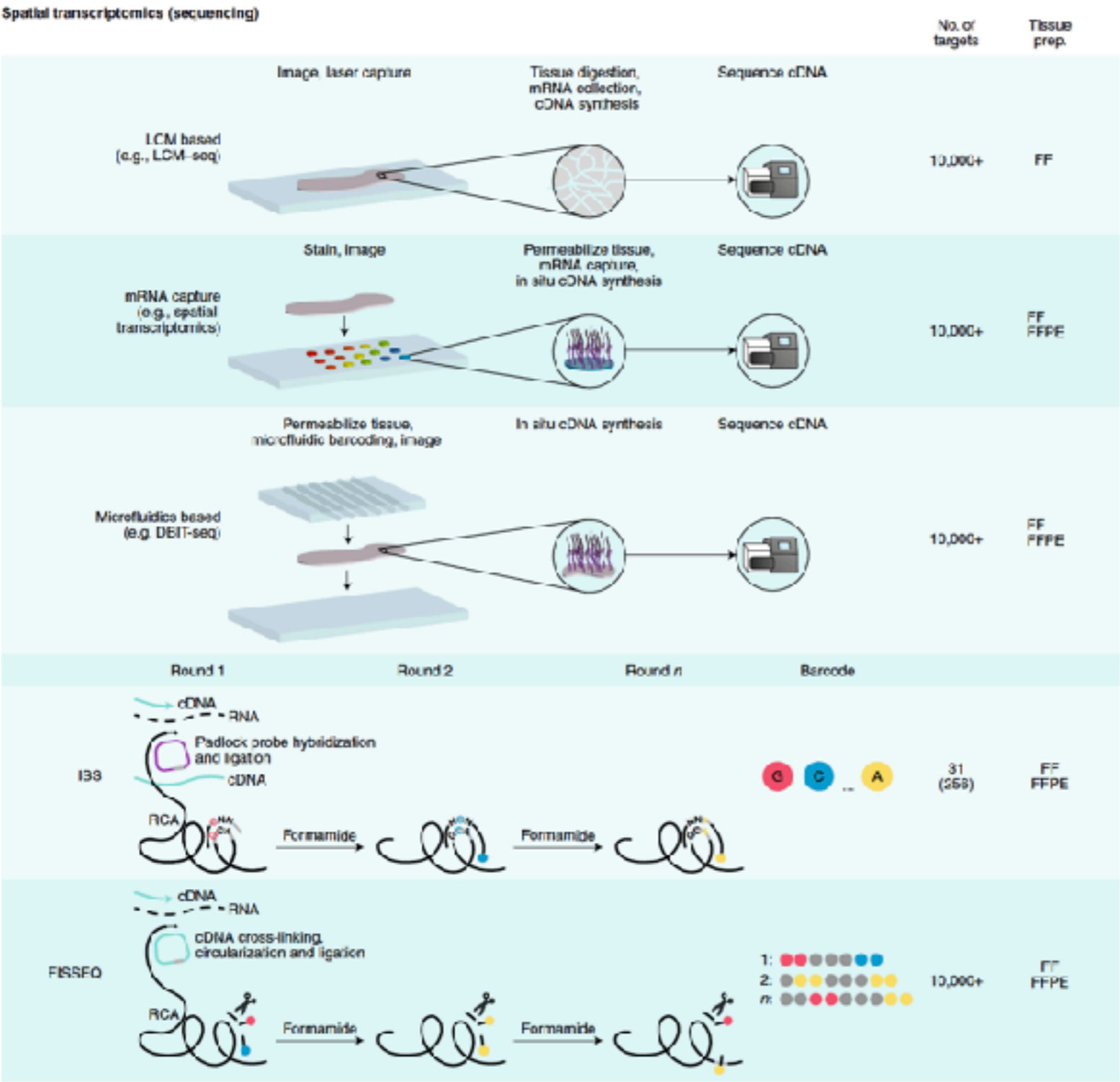
MERFISH (Barcoding)



MERFISH is a temporal barcoded smFISH method that measures **100–1,001** genes with high spatial resolution and detection efficiency. This is a single-molecule imaging approach that uses combinatorial labeling and sequential imaging with encoding schemes capable of detection and/or correction of errors. This highly multiplexed measurement of individual RNAs can be used to compute the gene expression profile and noise, covariation in expression among different genes, and spatial distribution of RNAs within single cells.

Spatial Omics Techniques

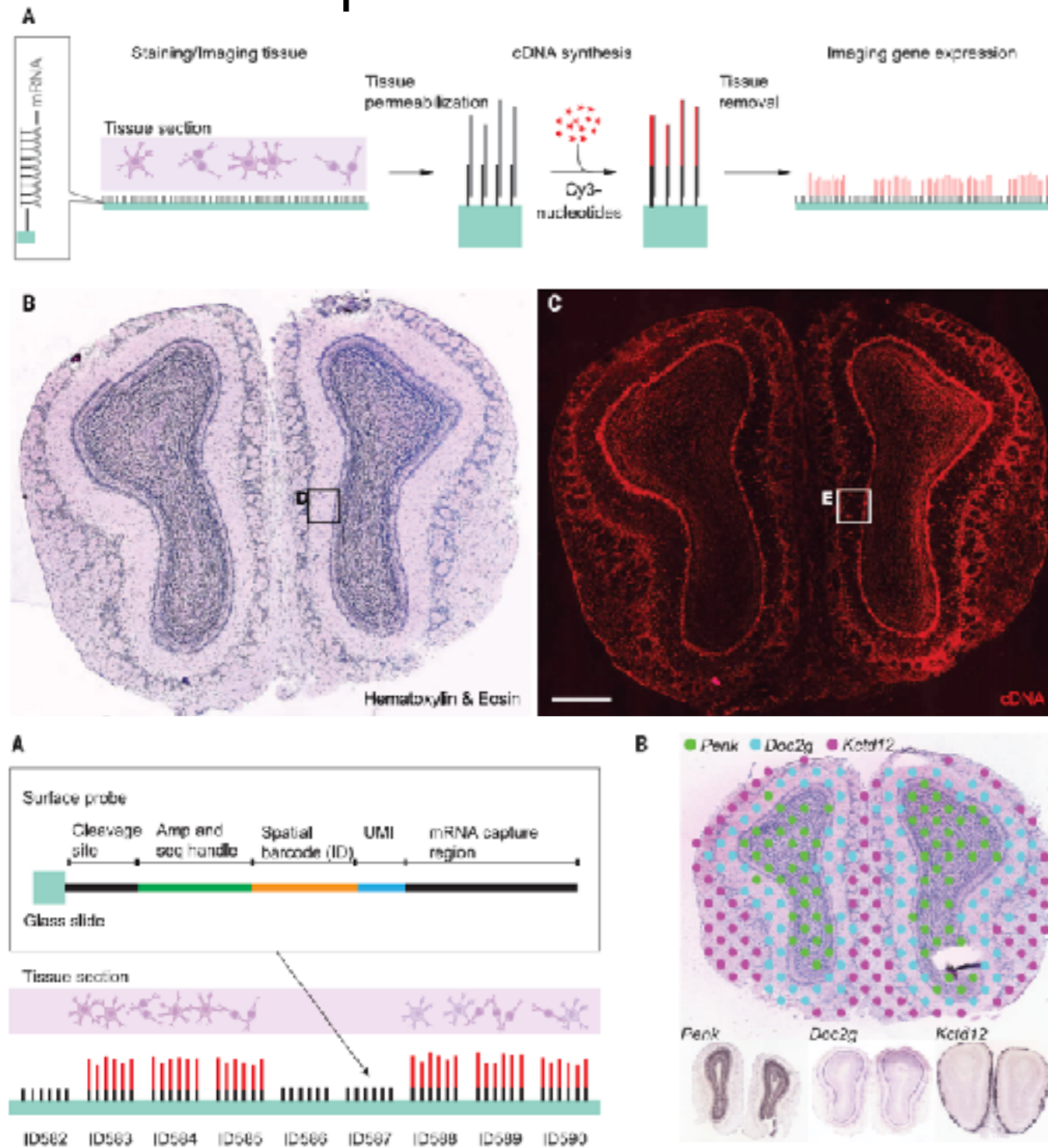
Sequencing methods



Spatial Transcriptomics

Spatial Omics Techniques

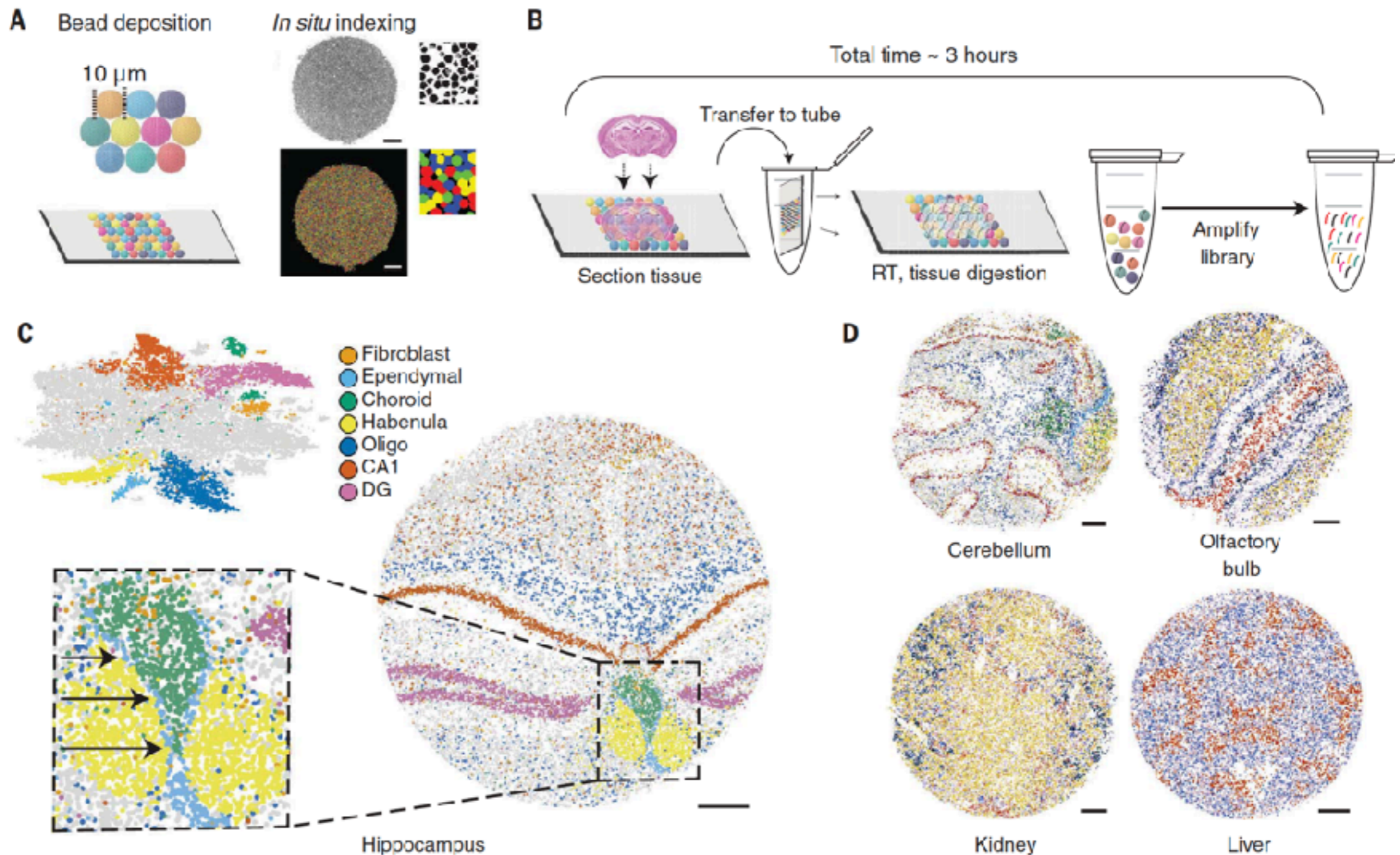
mRNA-Capture



The recent emergence of sequencing-based spatially resolved transcriptomics, as pioneered by 'spatial transcriptomics' has heralded the ability to determine the unbiased transcriptome of multiple regions in an entire tissue section. Here gridded areas of a slide contain or receive poly(dT) capture probes with identifier barcode sequences enabling later spatial assignment. After probes bind to the total mRNA present within the tissue, they are converted to cDNA, incorporating the spatial barcode. Gene expression is then traced back to a specific tissue location by way of the associated spatial barcode and through the use of fiducial markers on a corresponding histological image.

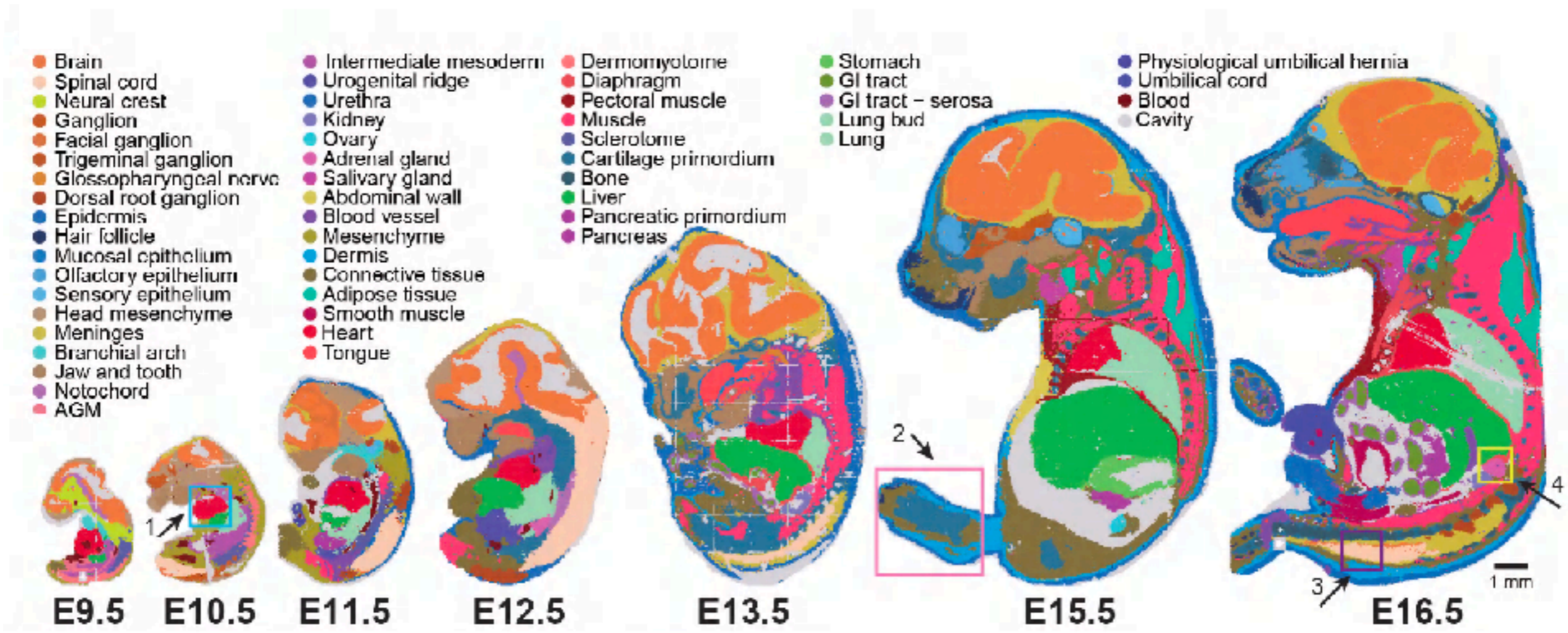
Spatial Omics Techniques

Slide-seq



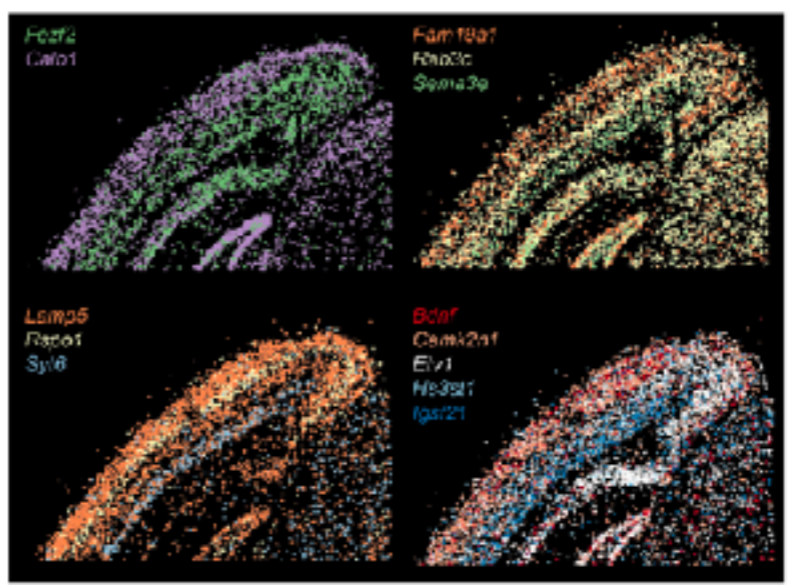
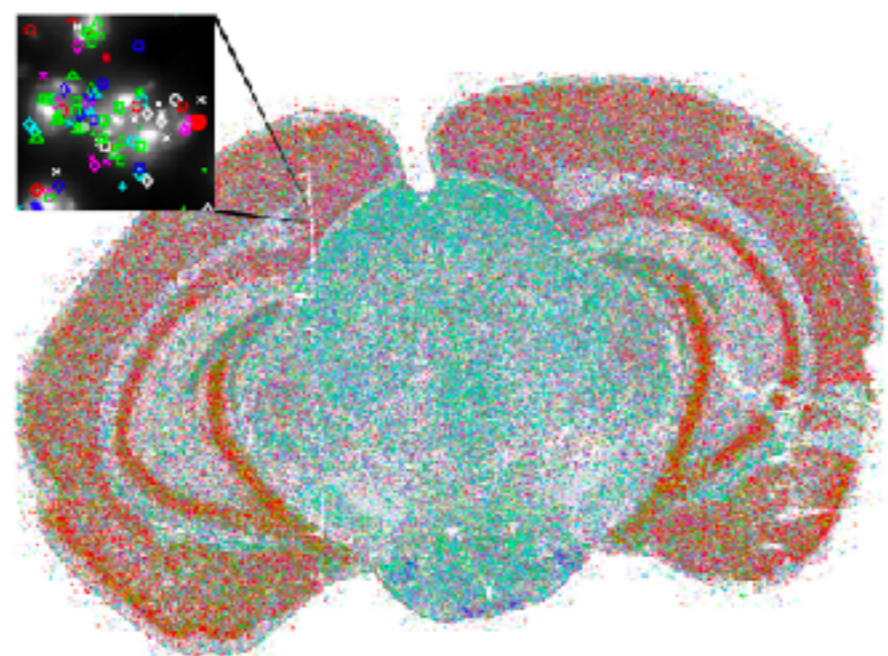
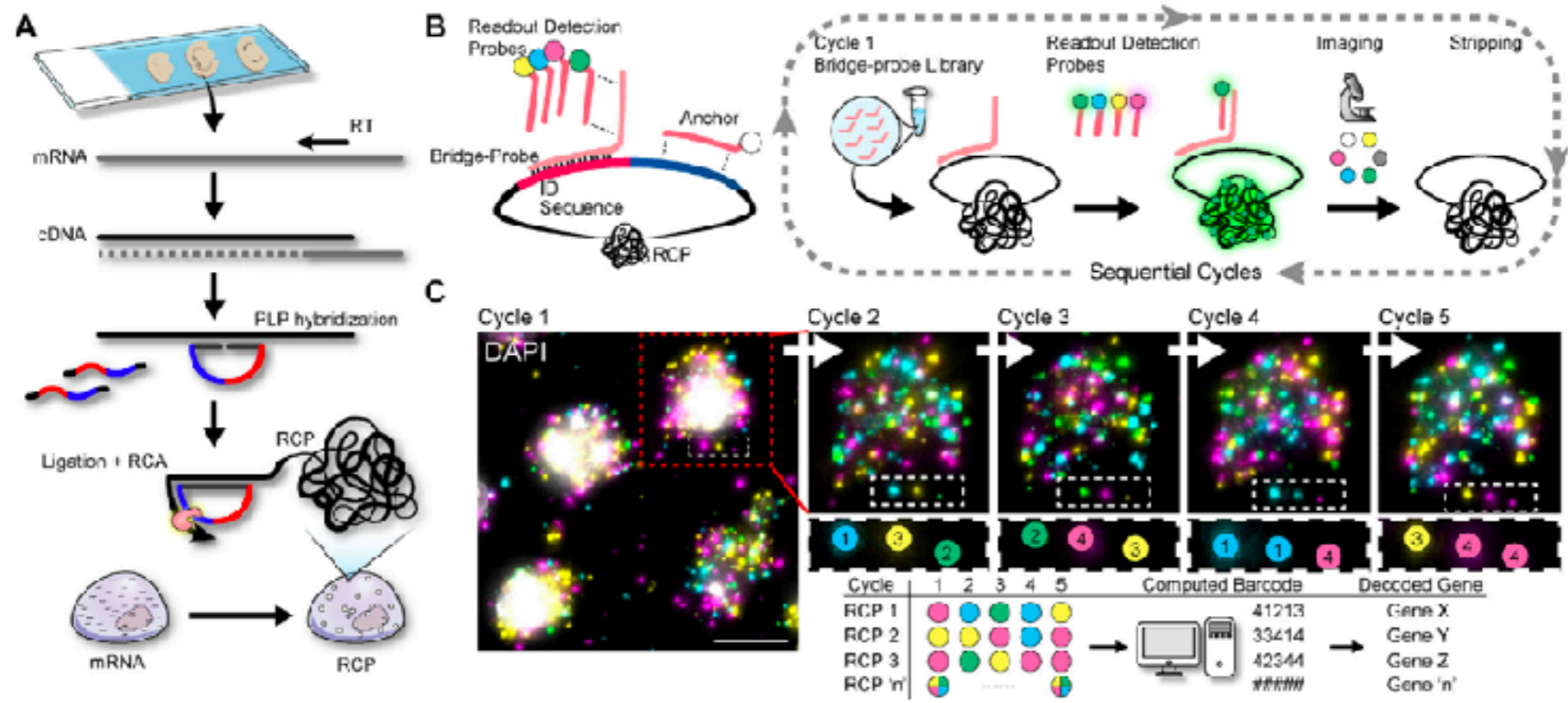
Spatial Omics Techniques

Stereo-seq



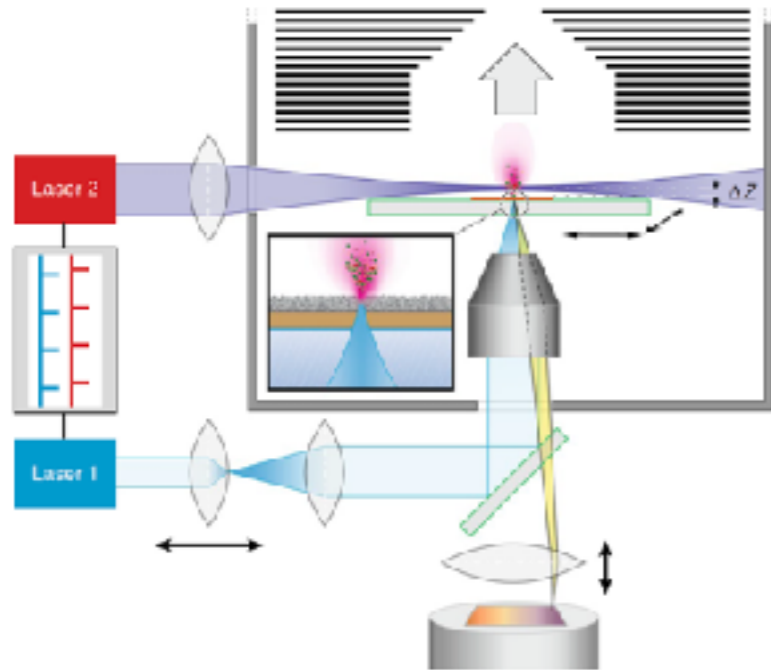
Spatial Omics Techniques

HybISS (Hybridization-based *in situ* sequencing)



Spatial Omics Techniques

MALDI-2-IMS



Matrix-assisted laser desorption–ionization mass spectrometry imaging in transmission-mode geometry (t-MALDI–MSI) can provide molecular information with a pixel size of 1 μm and smaller, which makes this label-free method highly interesting for characterizing the chemical composition of tissues and cells on a (sub)cellular level. laser-induced postionization (MALDI-2) increase the sensitivity of this technique to allow in situ single cell metabolomics

