

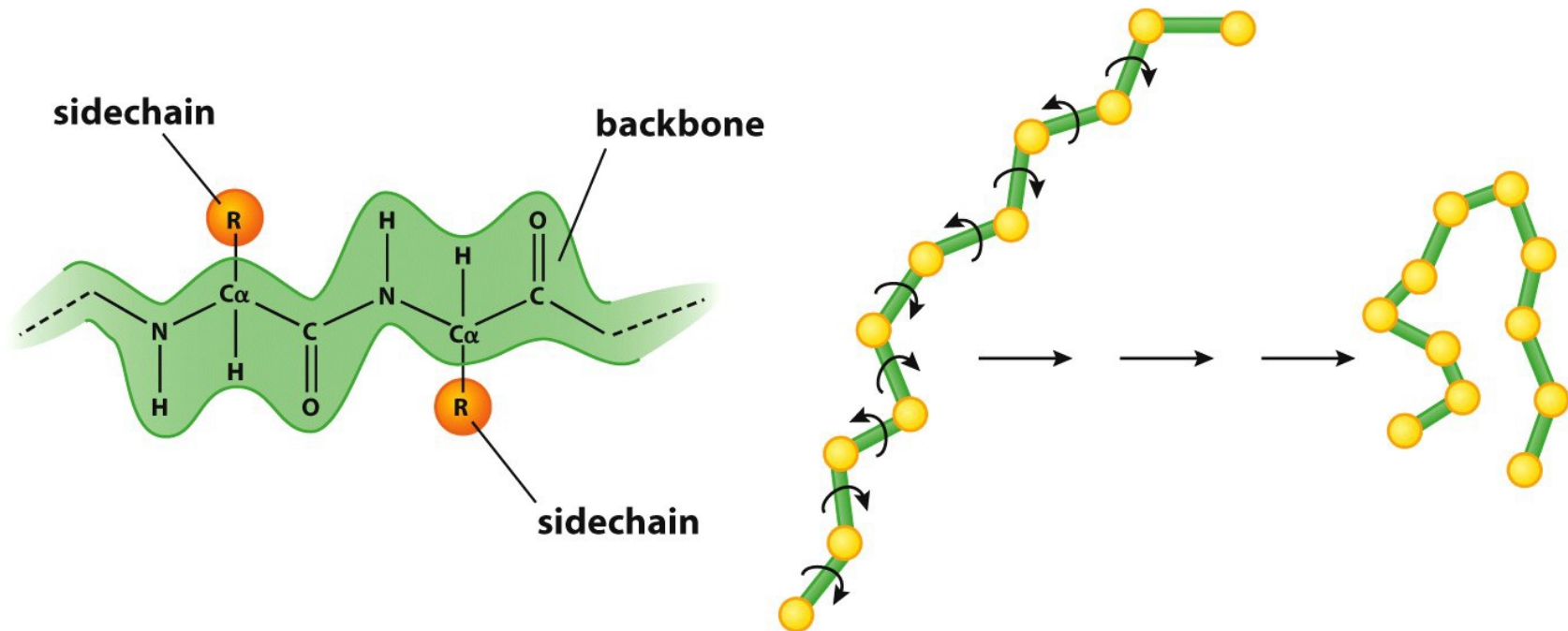
# Searching

## efficient sampling of protein conformations

1. conformational degrees of freedom

2. search methods

Protein folding involves rotations of the peptide backbone

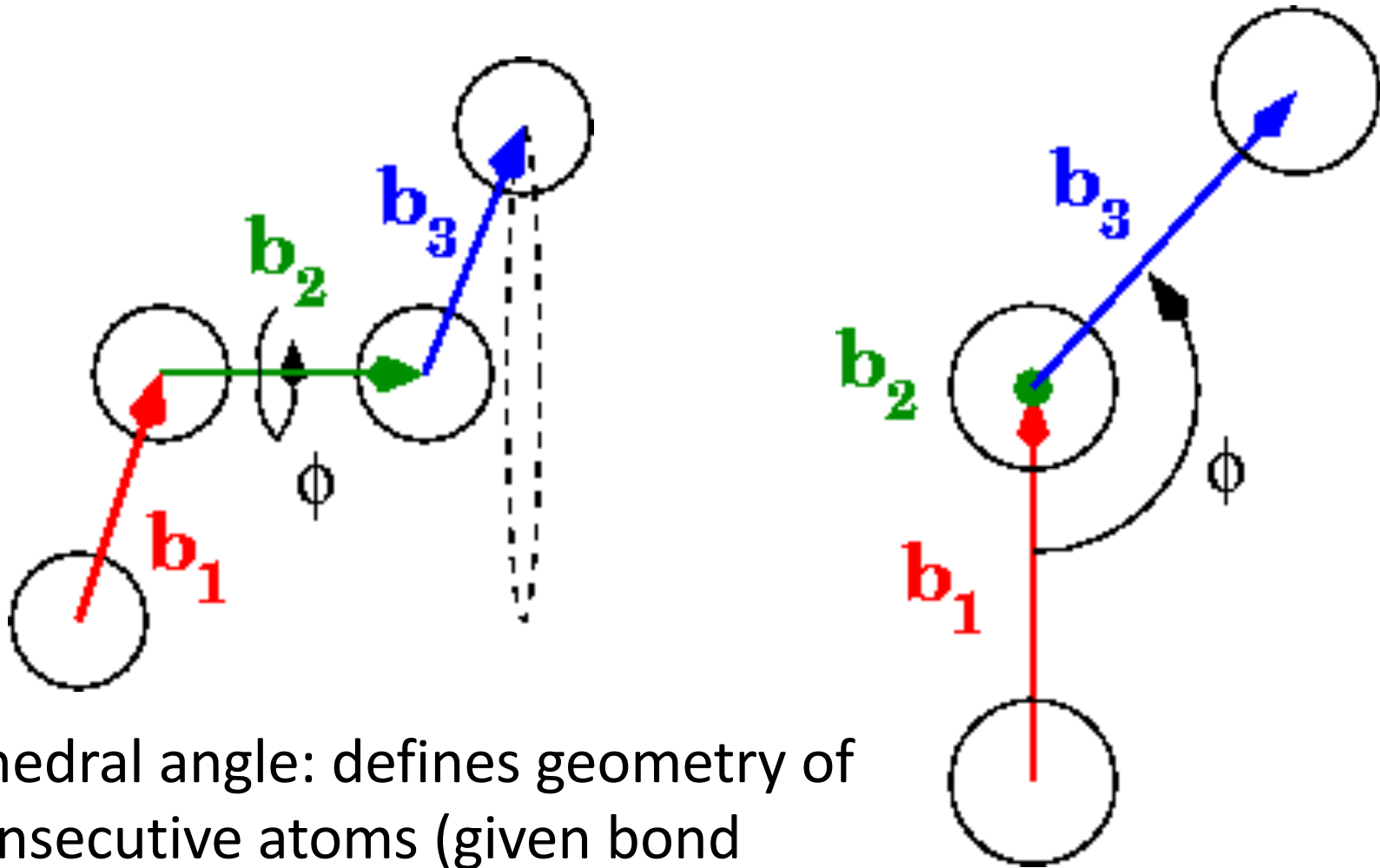


## Approximation:

# Peptide bond length and angles do not change

# Peptide dihedral angles define the structure

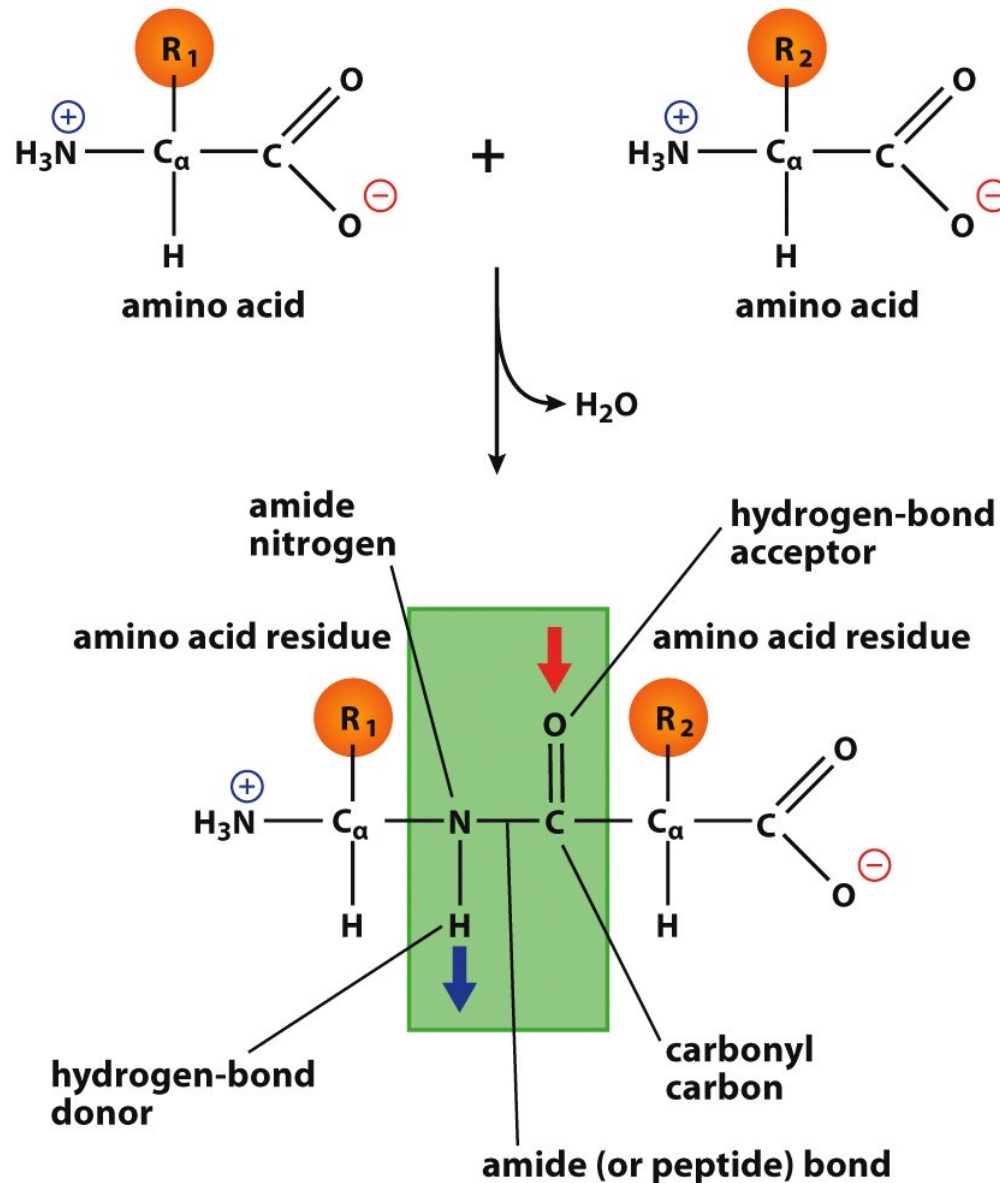
# Dihedral angles



- Dihedral angle: defines geometry of 4 consecutive atoms (given bond lengths and angles)

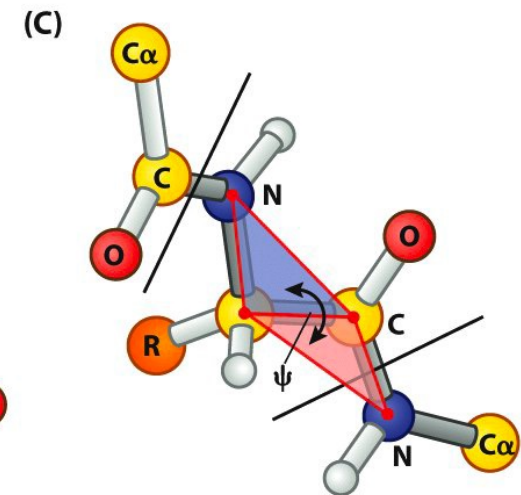
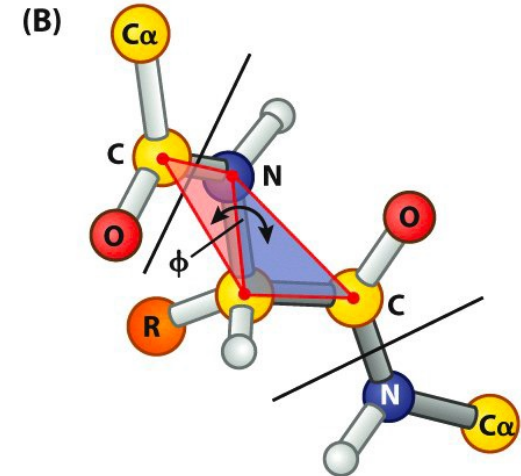
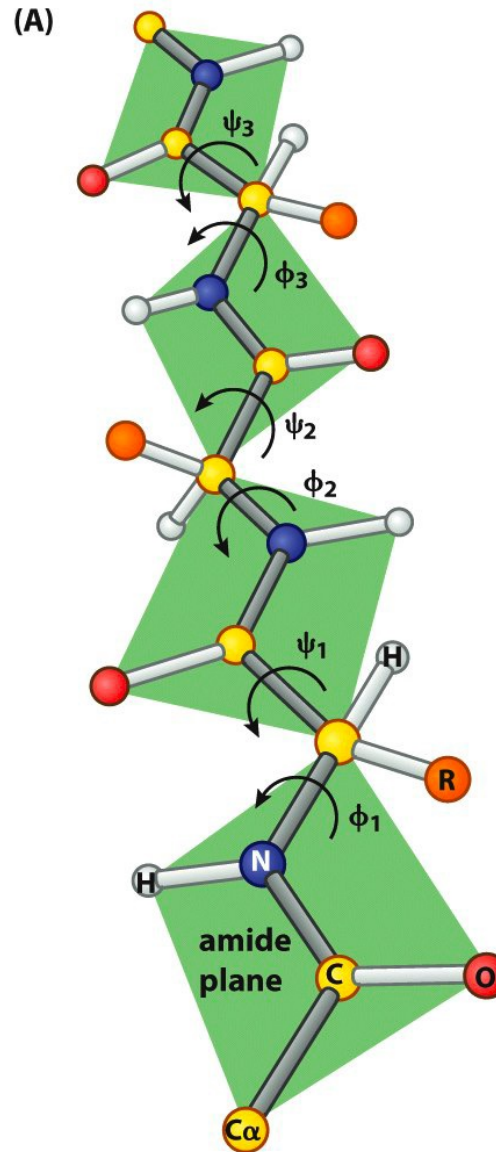
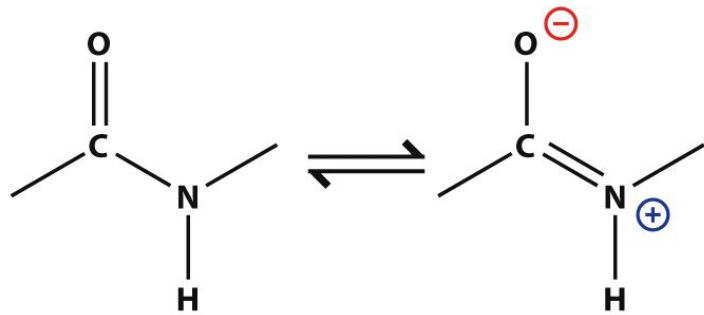
*From wikipedia*

# Formation of a peptide bond



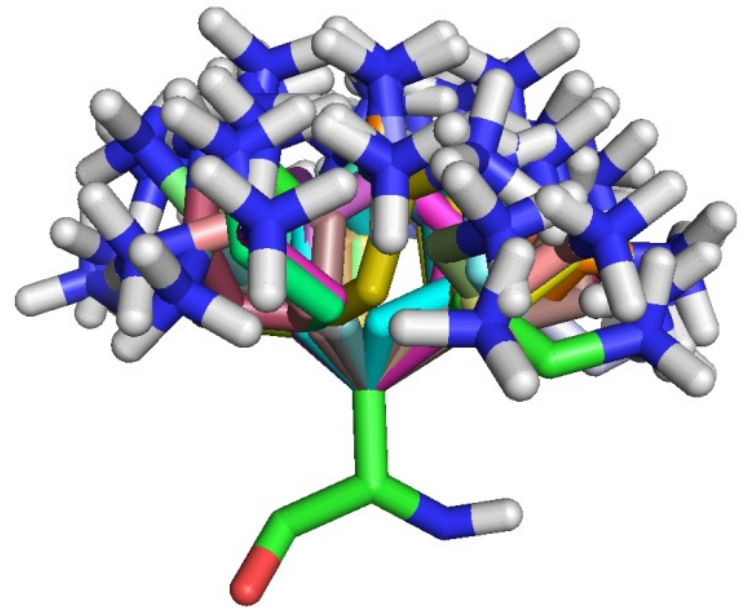
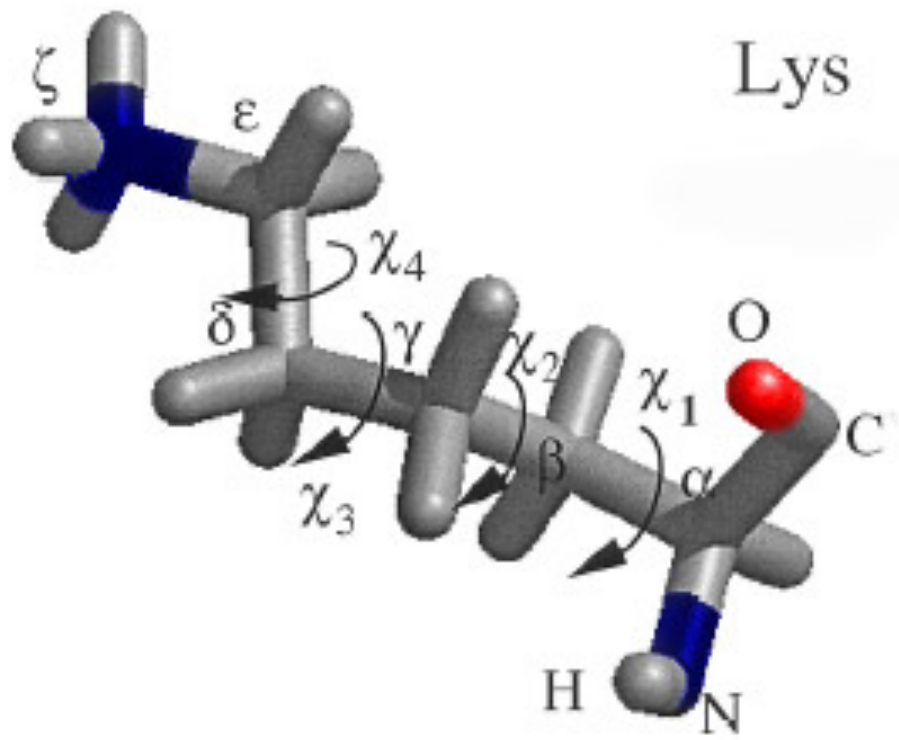
# Dihedral angles $\Phi$ and $\Psi$ define backbone conformation

The peptide bond is **planar** and **polar**

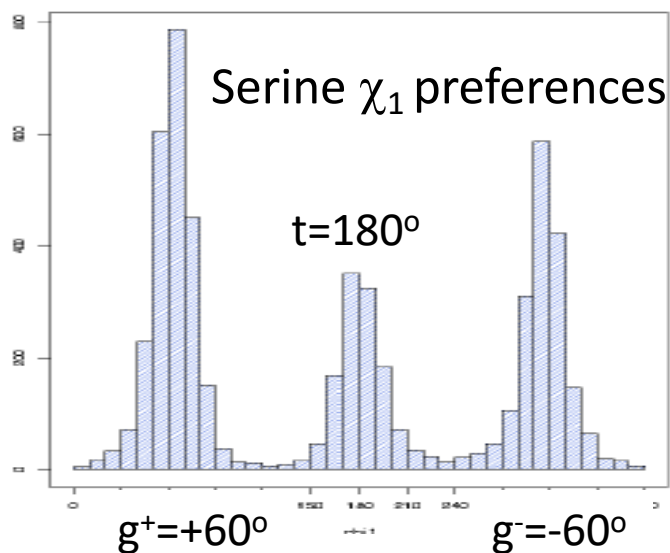
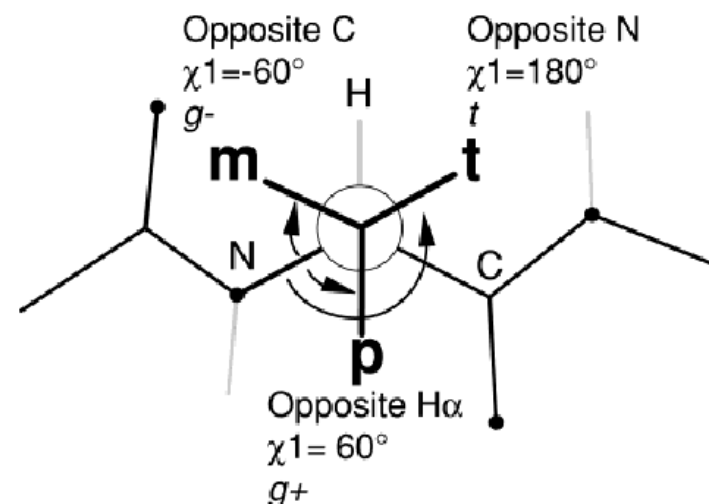
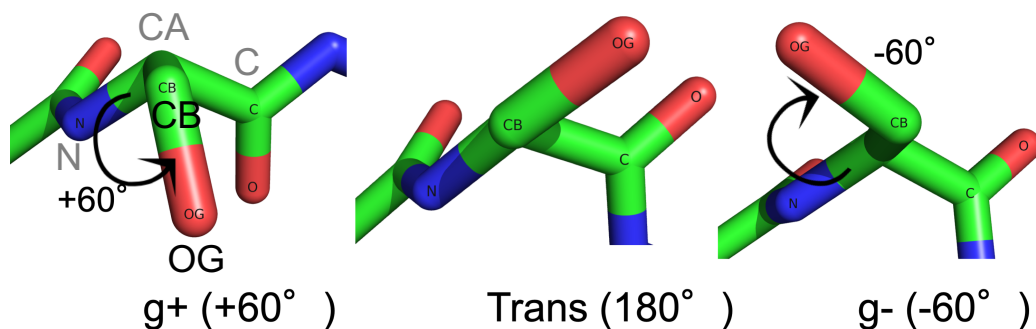


# Amino acid side chains can adopt multiple conformations

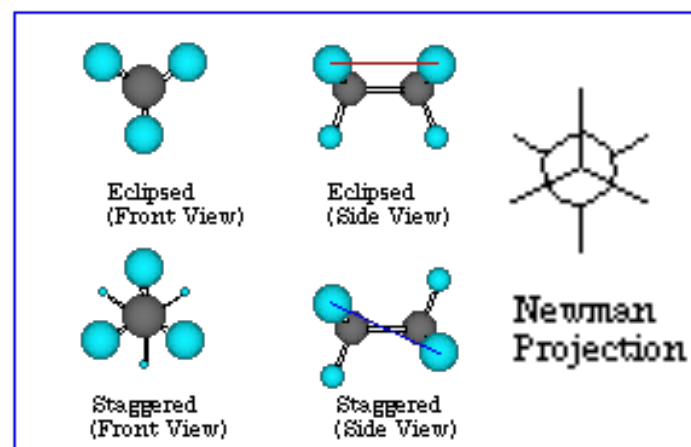
Dihedral angles  $\chi_1-\chi_4$  define side chain geometry



# Side chains assume discrete conformations

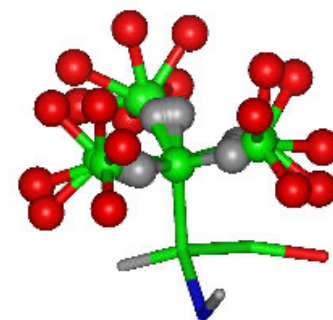


Staggered conformations minimize collision with neighboring atoms



# Rotamer libraries contain preferred conformations

Rotamer: discrete side chain conformation defined by  $\chi_1$ – $\chi_4$



**Table 1**

**Published rotamer libraries.**

Authors	Year	Type of library	Number of proteins in library	Resolution (Å)
Chandrasekaran and Ramachandran [2]	1970	BBIND	3	NA
Janin <i>et al.</i> [4]	1978	BBIND, SSDEP	19	2.5
Bhat <i>et al.</i> [3]	1979	BBIND	23	NA
James and Sielecki [5]	1983	BBIND	5	1.8, R-factor <0.15
Benedetti <i>et al.</i> [6]	1983	BBIND	238 peptides	R-factor <0.10
Ponder and Richards [7]	1987	BBIND	19	2.0
McGregor <i>et al.</i> [8]	1987	SSDEP	61	2.0
Tuffery <i>et al.</i> [9]	1991	BBIND	53	2.0
Dunbrack and Karplus [10]	1993	BBIND, BBDEP	132	2.0
Schrauber <i>et al.</i> [11]	1993	BBIND, SSDEP	70	2.0
Kono and Doi [12]	1996	BBIND	103	NA
De Maeyer <i>et al.</i> [13]	1995	BBIND	19	2.0
Dunbrack and Cohen [14]	1997–2002	BBIND, BBDEP	850*	1.7
Lovell <i>et al.</i> [15]	2000	BBIND, SSDEP	240	1.7
Shapovalov and Dunbrack*	2011	BBDEP	3854	1.8

\* Shapovalov & Dunbrack, *Structure* 2011

Dunbrack, 2002



# Representative rotamer libraries are surprisingly small

Ponder & Richards, 1987:  
Analysis of ~20 proteins  
(~2000 side chains)

**67** rotamers can adequately  
represent side chain  
conformations (for 17/20aa)

Side-chain angles		$\chi_1$	$\chi_2$	$\chi_3$	$\chi_4$				Atom position fixed by
Residue	Atom	$\alpha$	$\beta$	$\gamma$	$\delta$	$\epsilon$	$\zeta$	$\eta$	
Gly		*							Main chain
Ala		•							
Pro		•	•	•	•				
Ser		•	•	•	•	•			$\chi_1$
Cys		•	•	•	•	•			
Thr		•	•	•	•	•			
Val		•	•	•	•	•			
Ile		•	•	•	•	•	•		$\chi_1$ and $\chi_2$
Leu		•	•	•	•	•	•		
Asp		•	•	•	•	•	•	•	
Asn		•	•	•	•	•	•	•	
His		•	•	•	•	•	•	•	
Phe		•	•	•	•	•	•	•	
Tyr		•	•	•	•	•	•	•	
Trp		•	•	•	•	•	•	•	
Met		•	•	•	•	•	•	•	$\chi_1, \chi_2$ and $\chi_3$
Glu		•	•	•	•	•	•	•	
Gln		•	•	•	•	•	•	•	
Lys		•	•	•	•	•	•	•	$\chi_1, \chi_2,$ $\chi_3, \chi_4$
Arg		•	•	•	•	•	•	•	

## Isoleucine

— t	42	45.2	—60.9 (7.5)	168.7 (11.6)
— —	17	18.3	—59.6 (9.6)	—64.1 (14.3)
+ t	15	16.1	61.7 (5.0)	163.8 (16.4)
t t	12	12.9	—166.6 (10.1)	166.0 (8.9)
t +	3	3.2	—174.8 (24.9)	72.1 (10.5)
Other	4	4.3		

# Searching

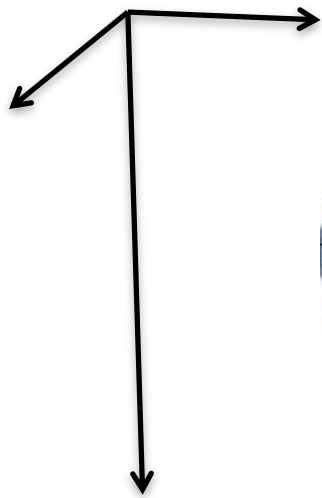
## efficient sampling of protein conformations

1. conformational degrees of freedom

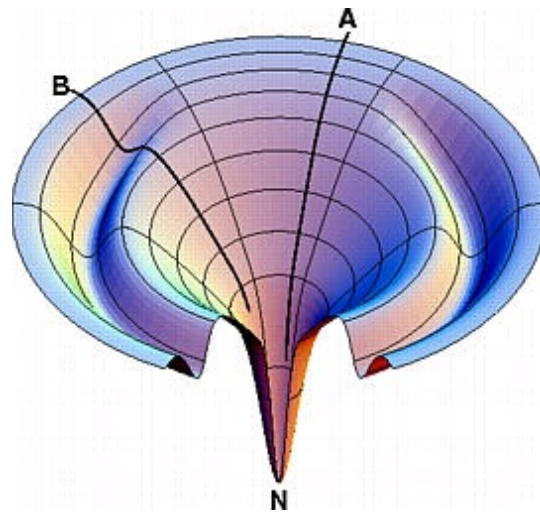
2. search methods

# Characteristics of the Protein Conformational Energy Landscape

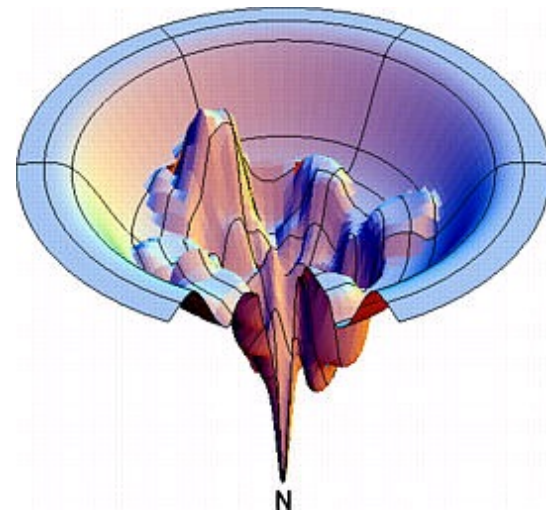
space of conformations



energy

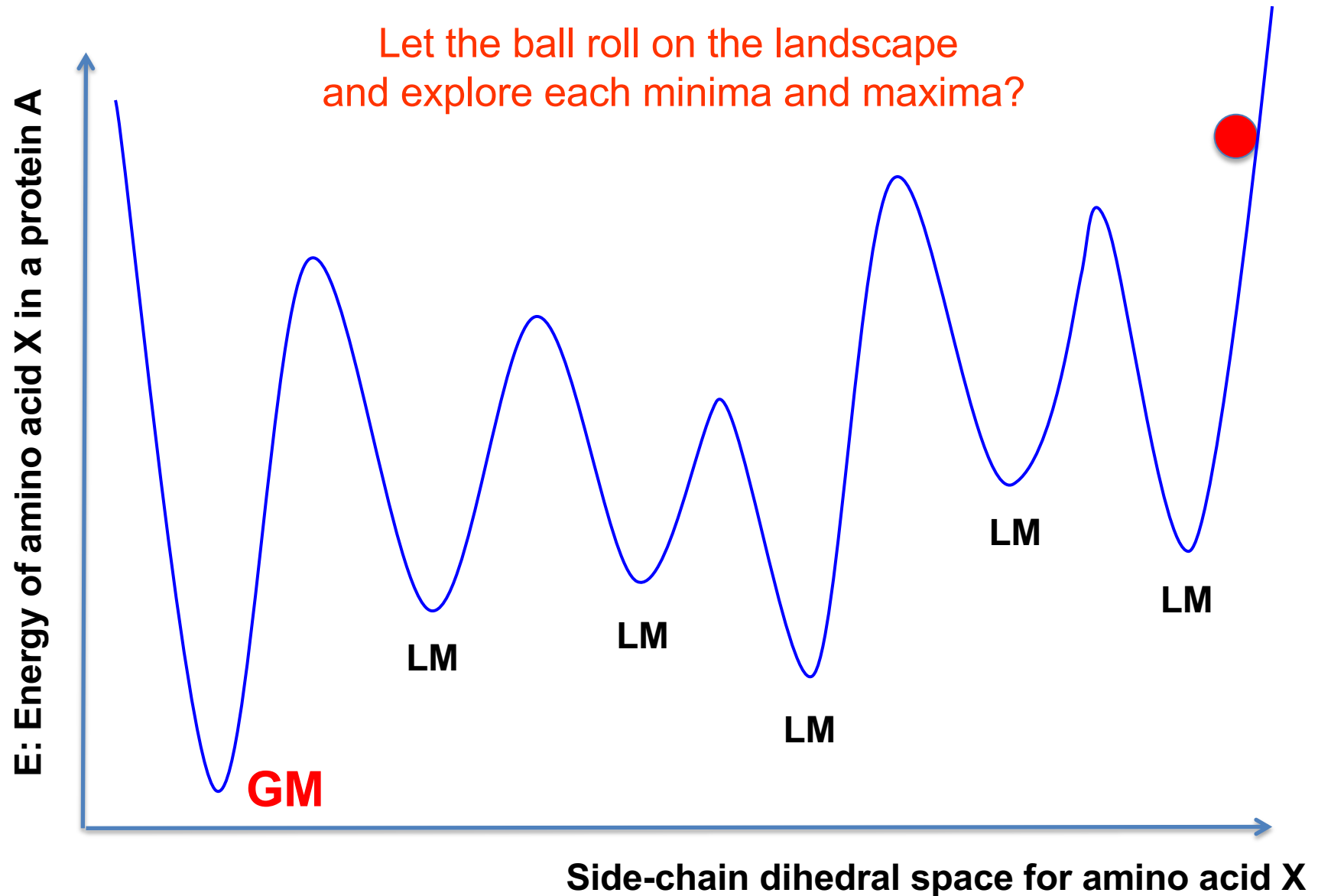


smooth?

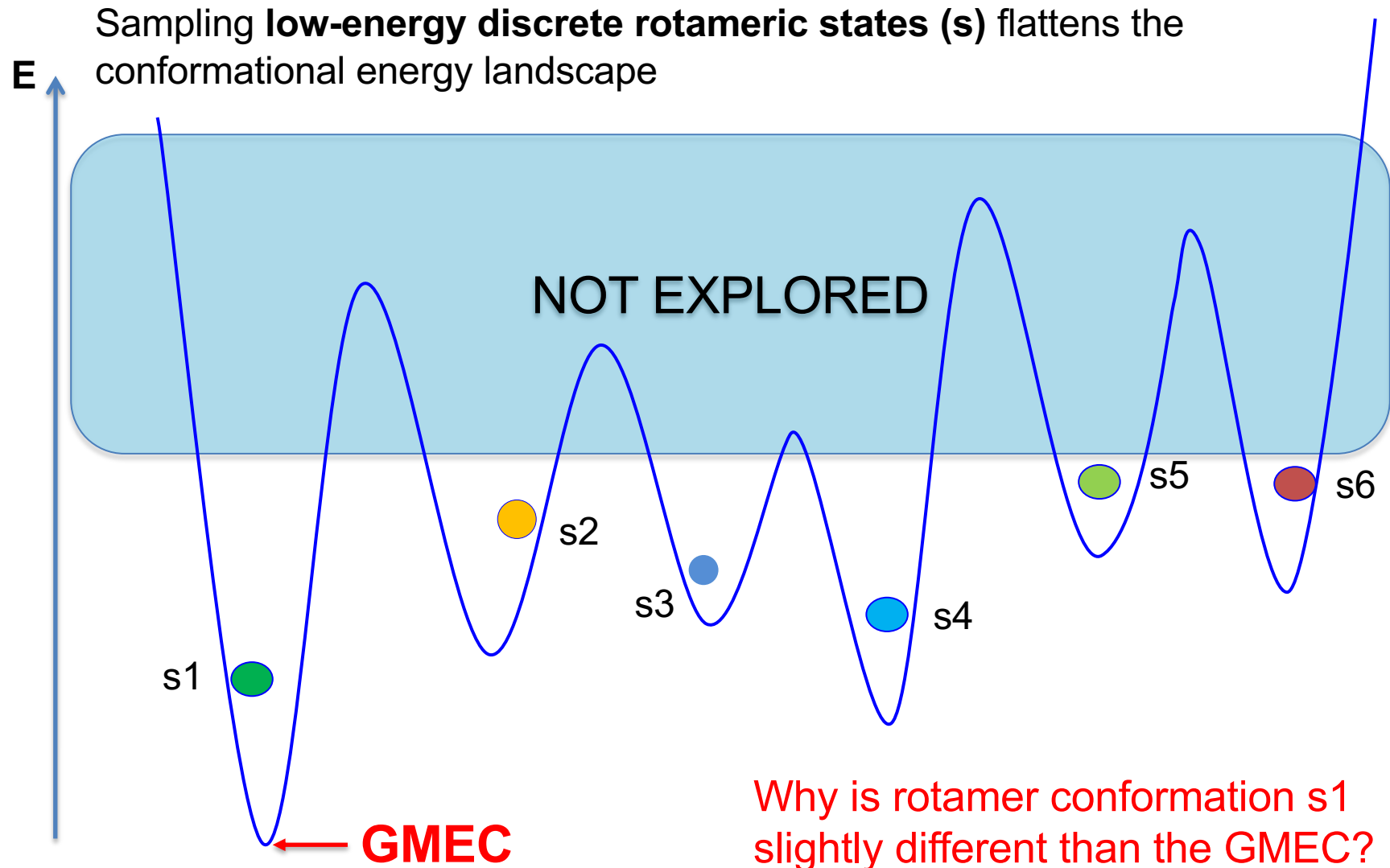


rugged?

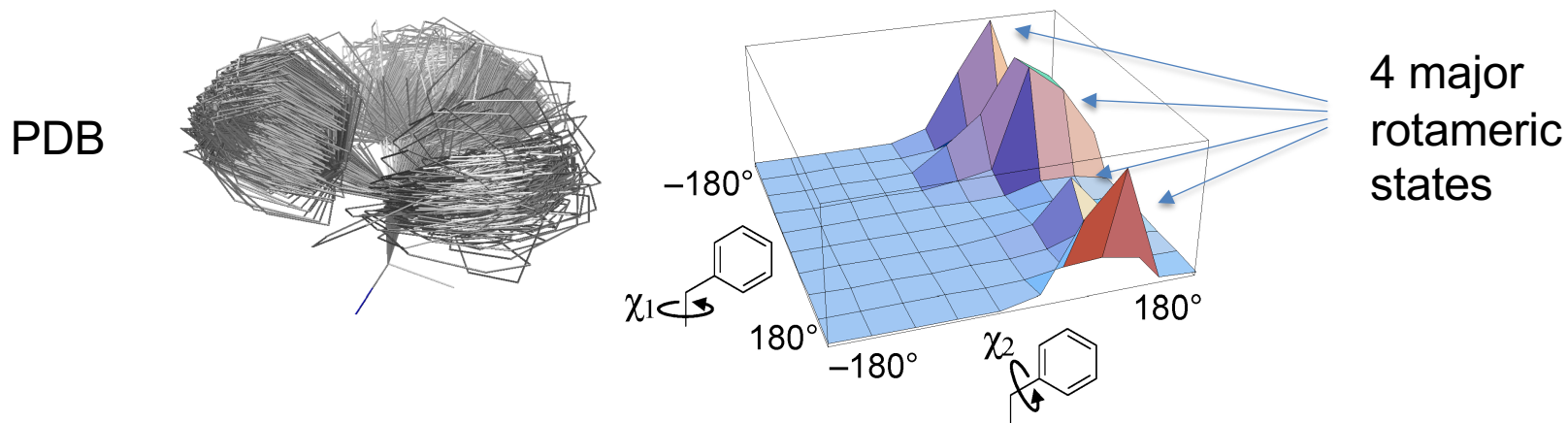
# The Problem: Find Local (LM) and Global Minima (GM) on a Rugged One Dimensional Surface



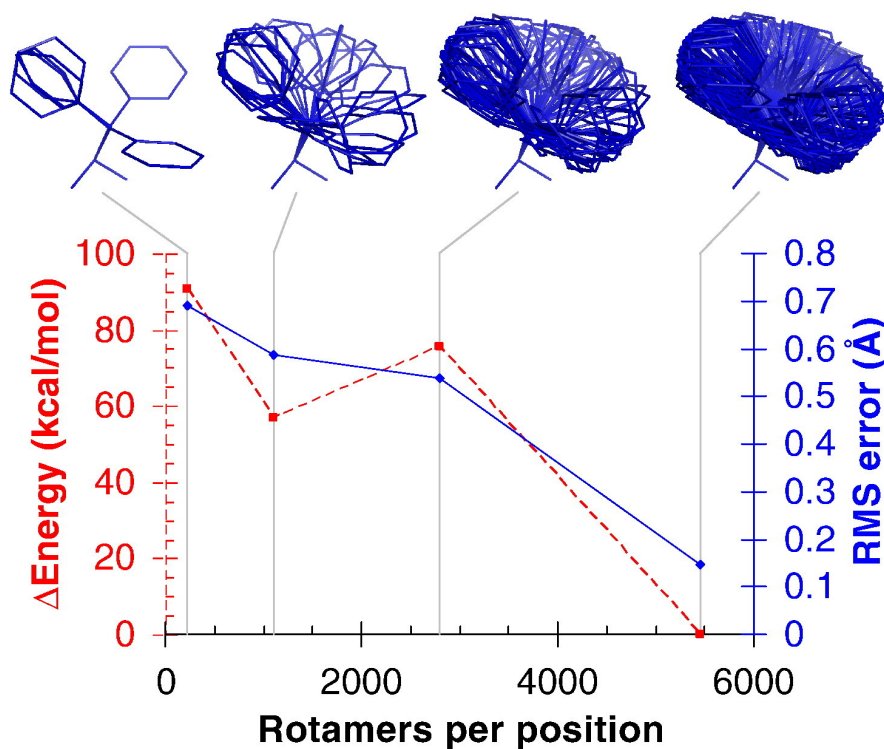
# Sampling low-energy discrete rotameric states simplifies the search in conformational space and allows rapid exploration of local minima



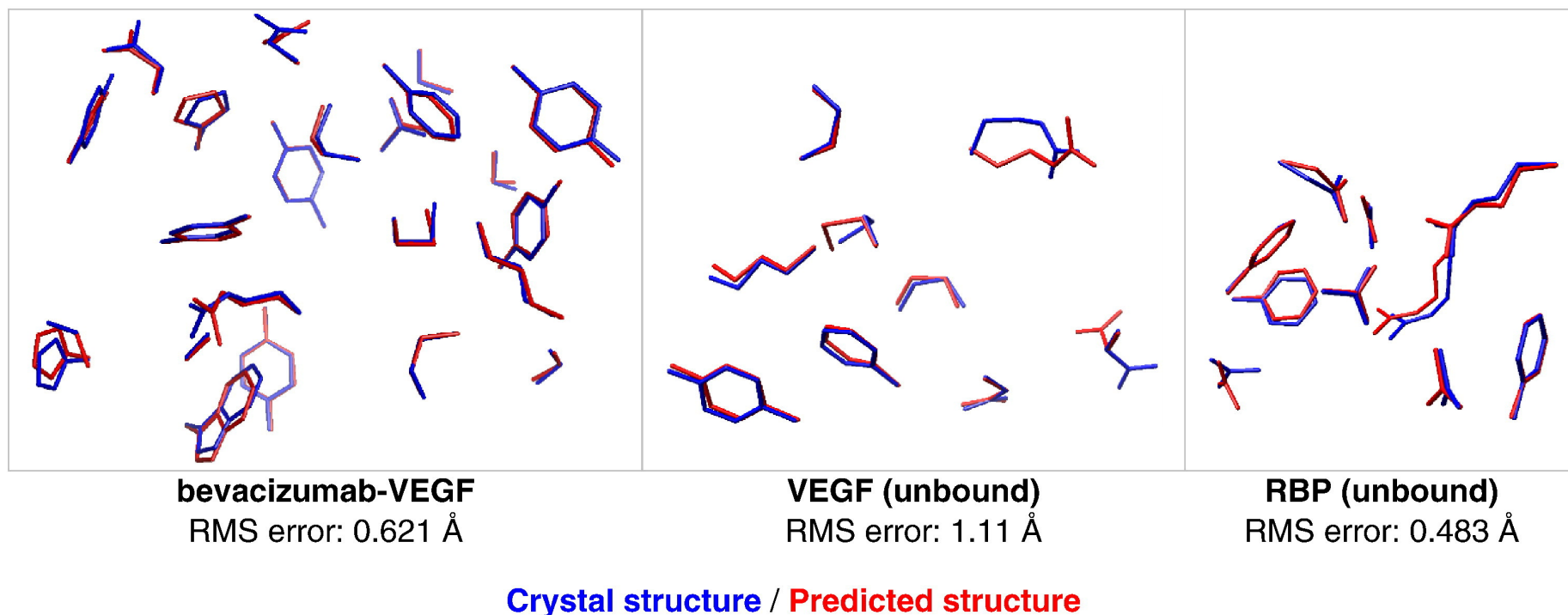
# Rotamer conformation accuracy



Effect of the resolution of the rotamer library on the conformation prediction accuracy

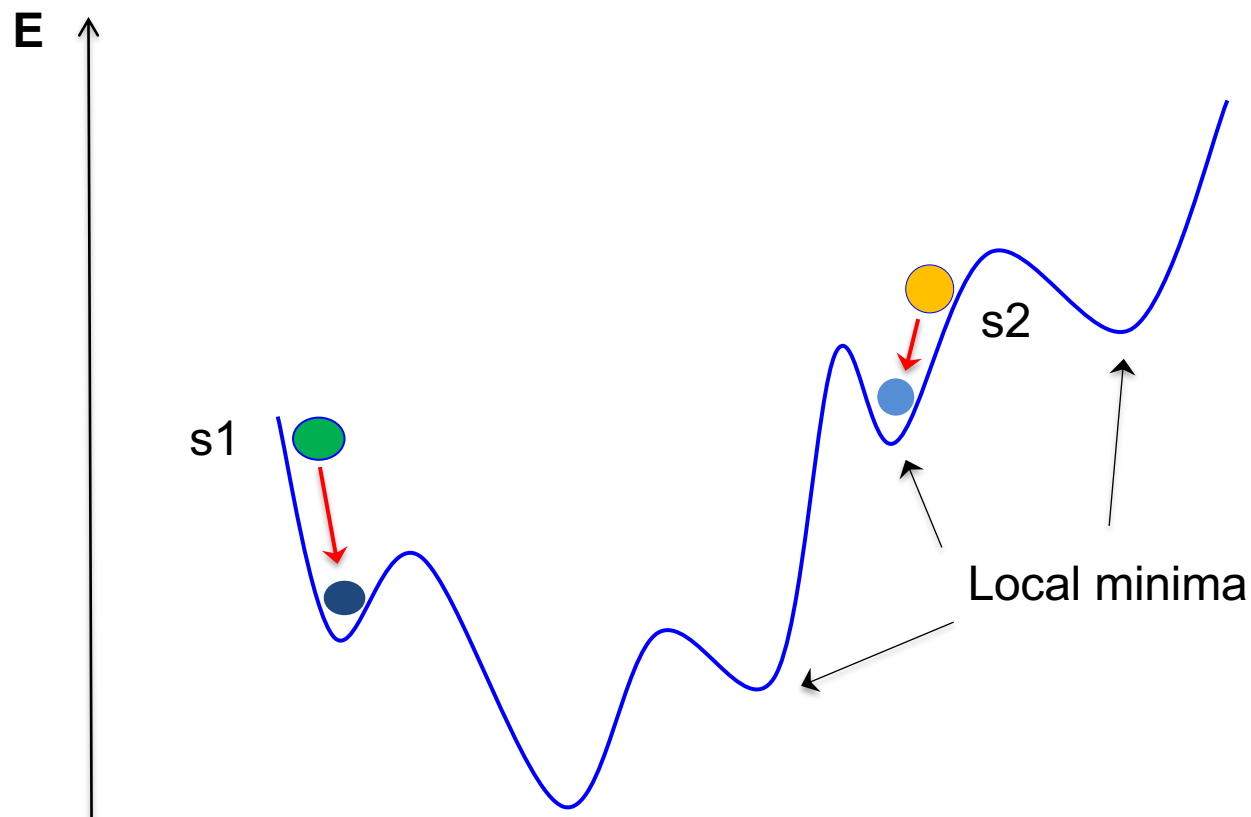


# Rotamers are an approximation to the true side-chain conformation



## Sampling method: Finding near-by local minima

- Derivative-based methods (Gradient Descent, Newton's method, DFP) are excellent at **finding** near-by local minima



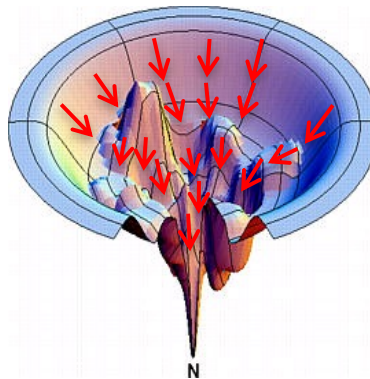


# Gradients and Hessians

Gradients and Hessians generalize the first and second derivatives (respectively) of multi-variate scalar functions (= functions from vectors to scalars)

$$\textbf{Energy} = f(\mathbf{x}_1, \mathbf{y}_1, \mathbf{z}_1, \dots, \mathbf{x}_n, \mathbf{y}_n, \mathbf{z}_n)$$

$$\bar{\nabla}_i = \frac{\partial E}{\partial \vec{r}_i} = \begin{pmatrix} \frac{\partial E}{\partial x_i} \\ \frac{\partial E}{\partial y_i} \\ \frac{\partial E}{\partial z_i} \end{pmatrix}$$



*Gradient*

$$\mathbf{h}_{ij} = \frac{\partial^2 E}{\partial \vec{r}_i \partial \vec{r}_j} = \begin{pmatrix} \frac{\partial^2 E}{\partial x_i \partial x_j} & \frac{\partial^2 E}{\partial x_i \partial y_j} & \frac{\partial^2 E}{\partial x_i \partial z_j} \\ \frac{\partial^2 E}{\partial y_i \partial x_j} & \frac{\partial^2 E}{\partial y_i \partial y_j} & \frac{\partial^2 E}{\partial y_i \partial z_j} \\ \frac{\partial^2 E}{\partial z_i \partial x_j} & \frac{\partial^2 E}{\partial z_i \partial y_j} & \frac{\partial^2 E}{\partial z_i \partial z_j} \end{pmatrix}$$

*Hessian*

# Analytical Energy Gradient

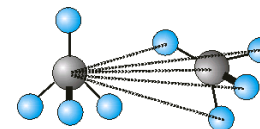
## (i) Cartesian Coordinates

$$E = f(x_1, y_1, z_1, \dots, x_n, y_n, z_n)$$

$$\vec{\nabla} = \left( \frac{\partial E}{\partial x_1} \frac{\partial E}{\partial y_1} \frac{\partial E}{\partial z_1} \dots \frac{\partial E}{\partial x_n} \frac{\partial E}{\partial y_n} \frac{\partial E}{\partial z_n} \right)$$

### Example:

Van der-Waals energy between pairs of atoms –  $O(n^2)$  pairs:

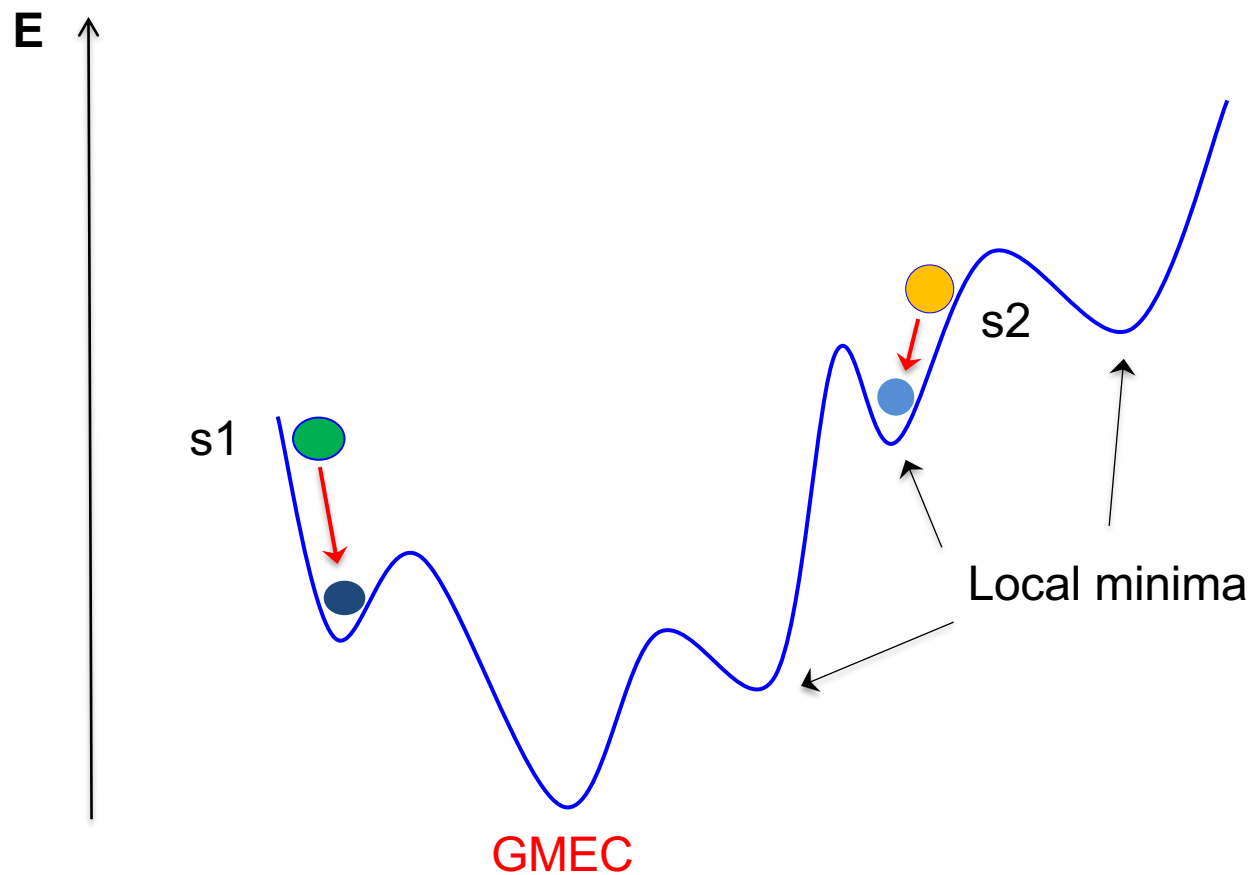


$$E_{vdW} = \sum_{i,j} \frac{A}{\bar{R}_{ij}^{12}} - \frac{B}{\bar{R}_{ij}^6} \quad \Rightarrow \quad \frac{\partial E_{vdW}}{\partial R_{ij}} = \frac{-12A}{R_{ij}^{13}} + \frac{6B}{R_{ij}^7}$$

$$R_{ij} = \sqrt{(x_i - x_j)^2 + (y_i - y_j)^2 + (z_i - z_j)^2}$$

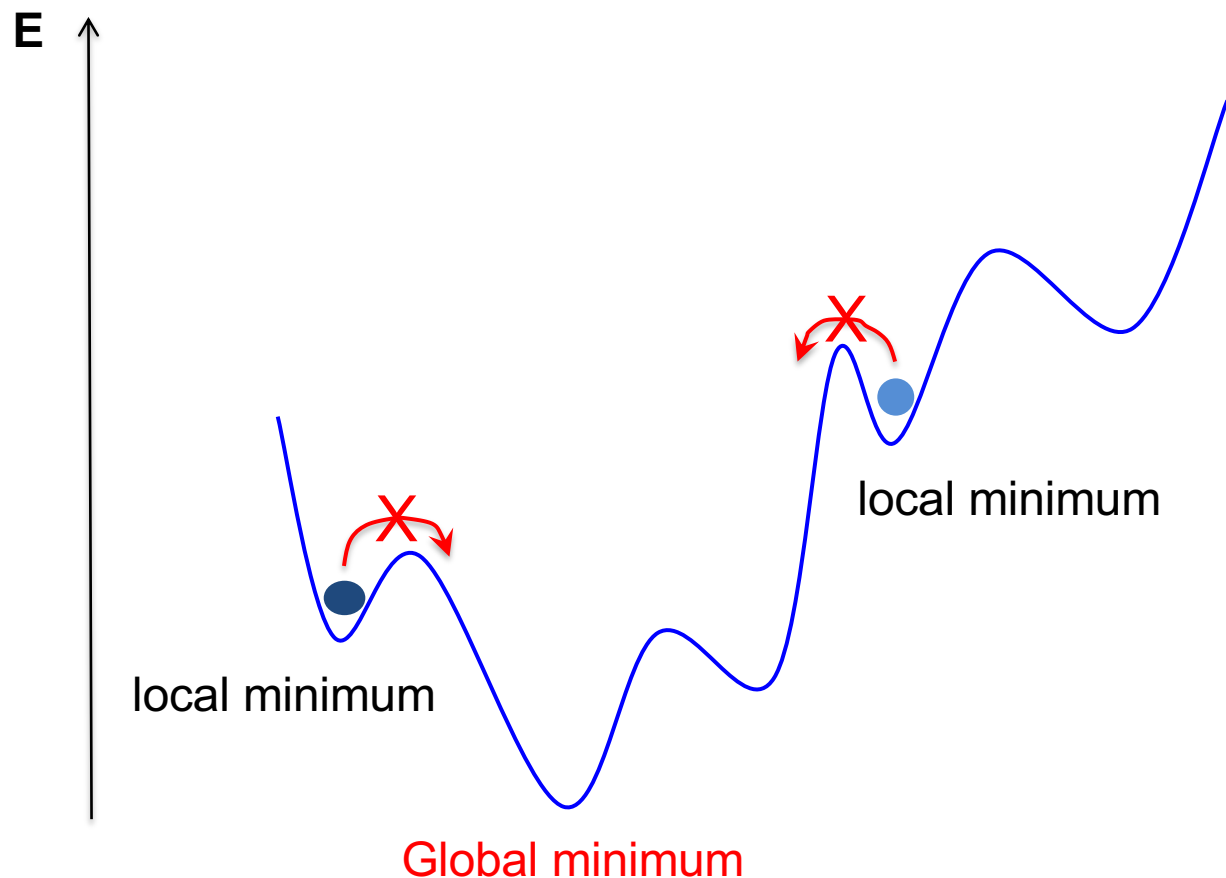
# Sampling method: Finding near-by local minima

How can we now find the GMEC?



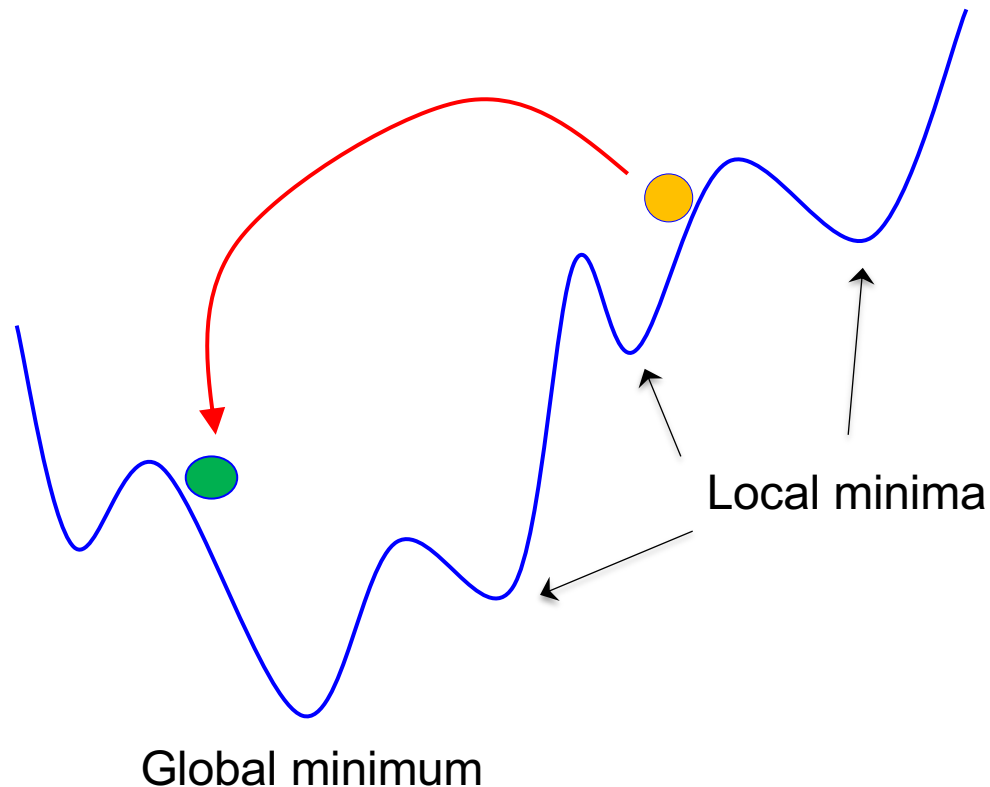
# Sampling method: Finding Global minima

- Derivative-based methods **cannot escape** near-by local minima



## Sampling method: jumping between discrete minima

- Several algorithms can be used to **jump** between discrete ***near-by local minima*** (e.g. Monte Carlo, Genetic algorithm, Self consistent Mean Field, Dead End Elimination)



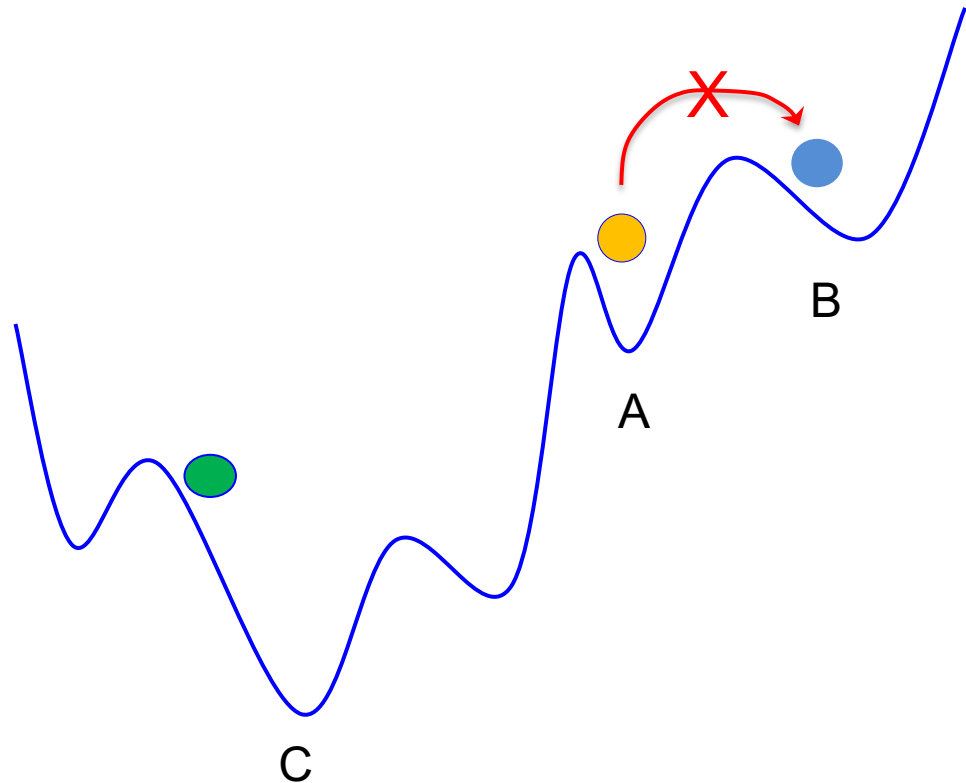
# Monte Carlo: stochastic sampling of discrete minima

Basic steps:

Random move:  $A \rightarrow B$

$E_B < E_A$  ?

No: move rejected



# Monte Carlo: stochastic sampling of discrete minima

Basic steps:

Random move: A→B

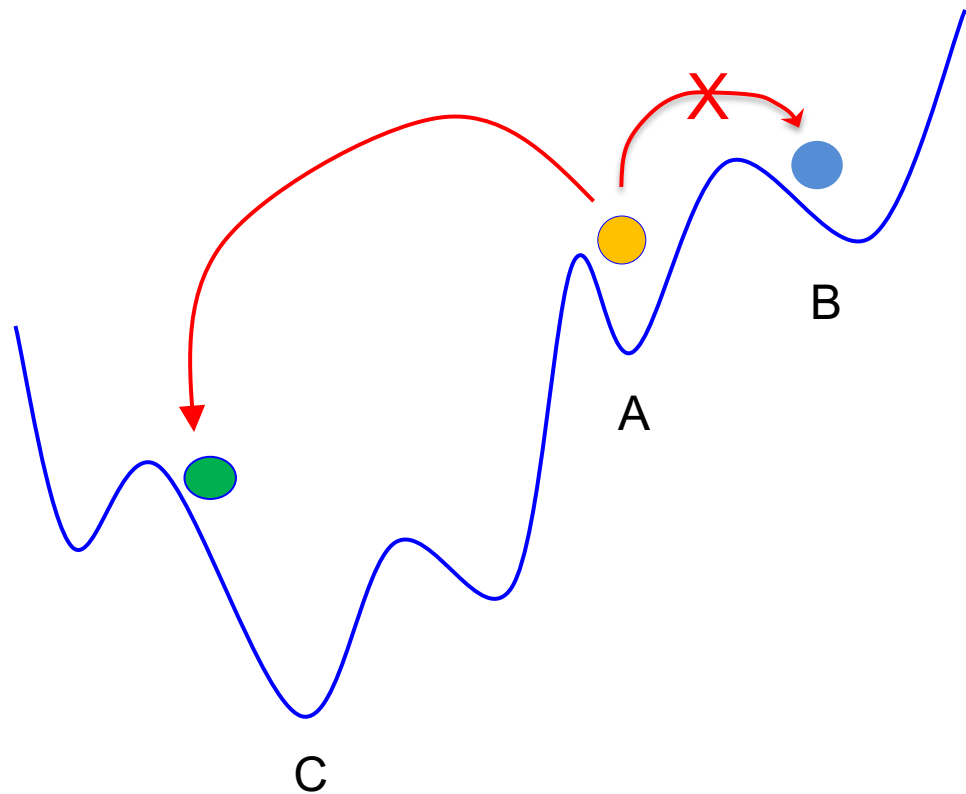
$E_B < E_A$  ?

No: move rejected

Random move: A→C

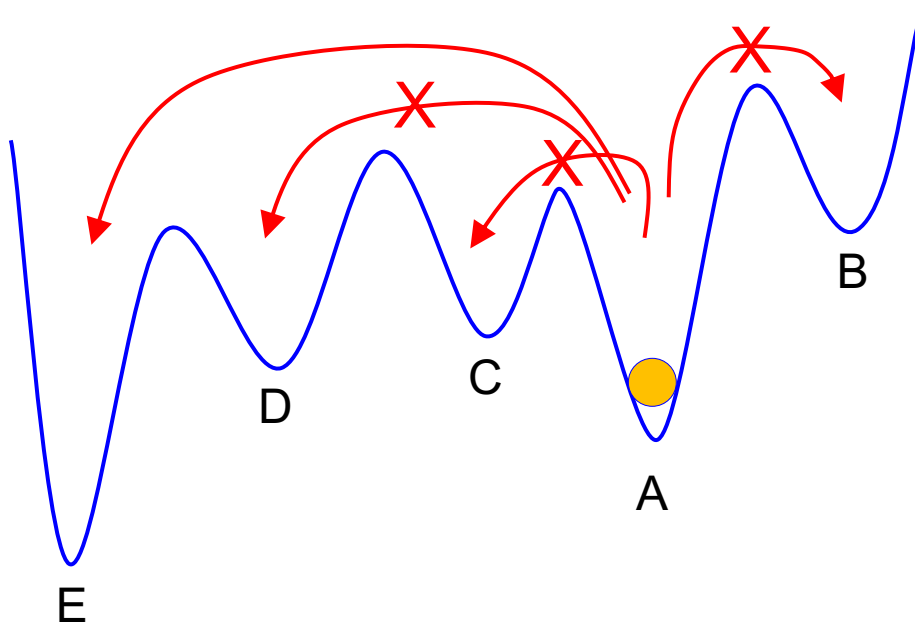
$E_C < E_A$  ?

Yes: move accepted



# Metropolis Monte Carlo

Very low probability  
to move out of A  
and reach the  
Global Minimum E



Acceptance with probability  $P = e^{-\Delta E / k_B T}$  (Boltzmann weight),  $\Delta E = E_B - E_A$

$P = 1$  if  $E_B < E_A$

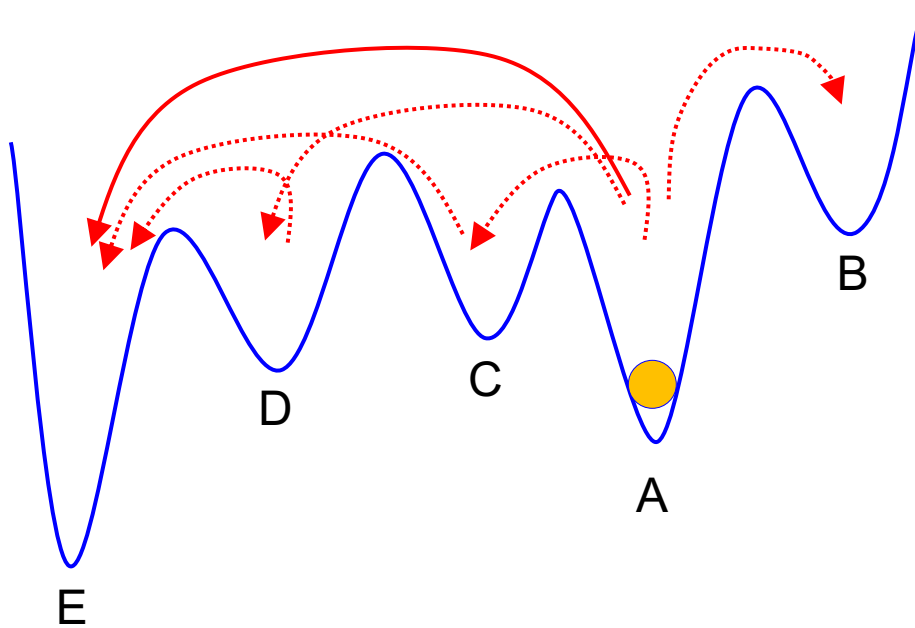
$P = 1$  if  $E_B > E_A$  &&  $e^{-\Delta E / k_B T} \geq rand(0,1)$

$P = 0$  if  $E_B > E_A$  &&  $e^{-\Delta E / k_B T} \leq rand(0,1)$



# Metropolis Monte Carlo

Probability  $P$   
to move out of A  
depends on the  
temperature  $T$



$$P = e^{-\Delta E / k_B T}$$

$$P = 1 \text{ if } E_B < E_A$$

$$P = 1 \text{ if } E_B > E_A \ \&\& \ e^{-\Delta E / k_B T} \geq rand(0,1)$$

$$P = 0 \text{ if } E_B > E_A \ \&\& \ e^{-\Delta E / k_B T} \leq rand(0,1)$$

# Metropolis Monte Carlo with Simulated Annealing

The Boltzman distribution depends on the *in-silico* temperature **T**:

- At low temperatures, we will get stuck in local minima (we will always get zero acceptance if the energy rises even slightly)
- At high temperatures, acceptance will always be 1 (always jump between conformations regardless of their energies).

$$P = 1 \text{ if } E_j > E_i \quad \&\& \quad e^{-\Delta E / k_B T} \geq rand(0,1)$$

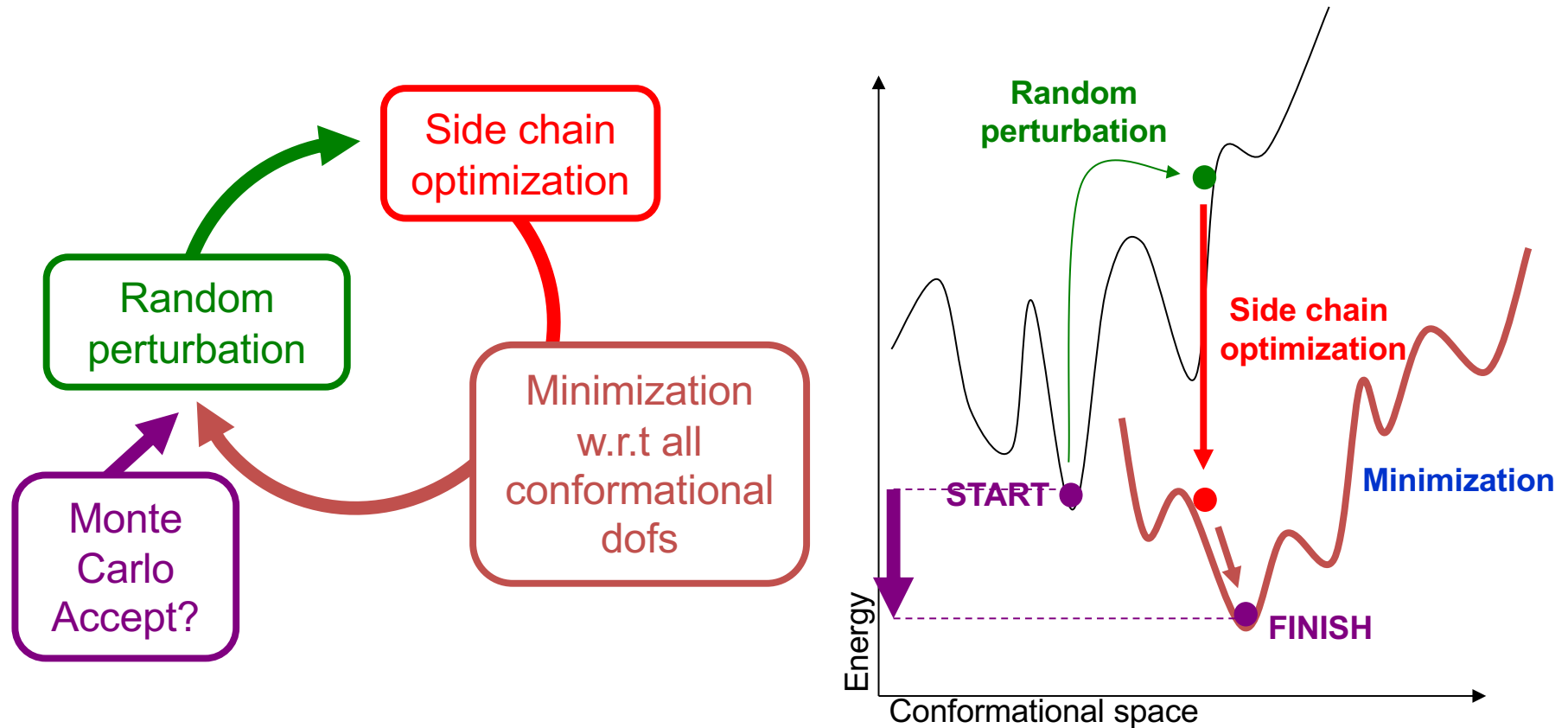
$$P = 0 \text{ if } E_j > E_i \quad \&\& \quad e^{-\Delta E / k_B T} \leq rand(0,1)$$

**In simulated annealing, we gradually decrease (“cool down”) the virtual temperature factor, until we converge to a minimum point**

# Basic design cycle step in Rosetta

Start: target structure

Random perturbation: amino acid substitution

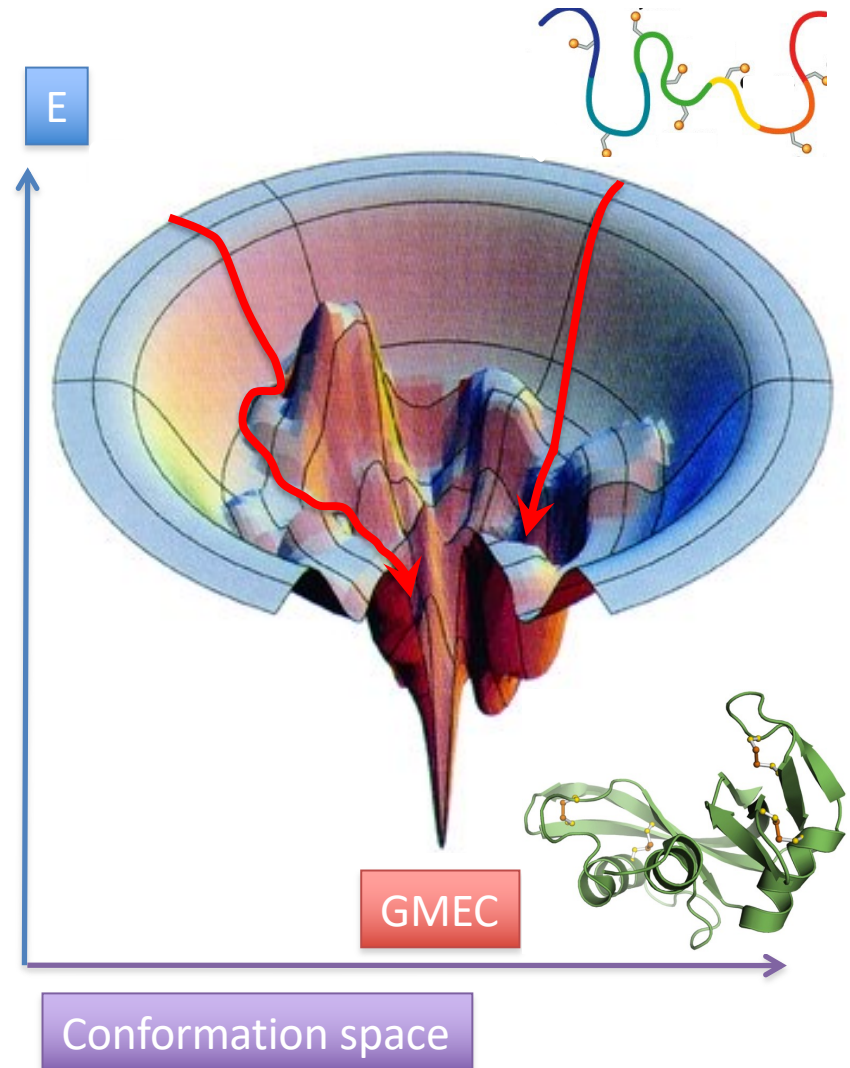


# Take home message

Computational protein design is challenging but made possible thanks to:

1. An **energy function** to rapidly rank sequences

2. An efficient **search technique** to find the GMEC



## Main points to remember

1. Protein design challenges: explore astronomically large space of possibilities

Approximations to accelerate physics-based protein design calculations based on:

2. Simple description physical interactions

3. Discretization of conformational space

4. Efficient continuous and stochastic search technique to find the GMEC