



## NEUROSCIENCE

# Hormone-mediated neural remodeling orchestrates parenting onset during pregnancy

Rachida Ammari<sup>1†</sup>, Francesco Monaca<sup>1†</sup>, Mingran Cao<sup>1</sup>, Estelle Nassar<sup>1</sup>, Patty Wai<sup>1</sup>, Nicholas A. Del Grosso<sup>1</sup>, Matthew Lee<sup>1</sup>, Neven Borak<sup>1</sup>, Deborah Schneider-Luftman<sup>2</sup>, Johannes Kohl<sup>1\*</sup>

During pregnancy, physiological adaptations prepare the female body for the challenges of motherhood. Becoming a parent also requires behavioral adaptations. Such adaptations can occur as early as during pregnancy, but how pregnancy hormones remodel parenting circuits to instruct preparatory behavioral changes remains unknown. We found that action of estradiol and progesterone on galanin (Gal)-expressing neurons in the mouse medial preoptic area (MPOA) is critical for pregnancy-induced parental behavior. Whereas estradiol silences MPOA<sup>Gal</sup> neurons and paradoxically increases their excitability, progesterone permanently rewires this circuit node by promoting dendritic spine formation and recruitment of excitatory synaptic inputs. This MPOA<sup>Gal</sup>-specific neural remodeling sparsens population activity *in vivo* and results in persistently stronger, more selective responses to pup stimuli. Pregnancy hormones thus remodel parenting circuits in anticipation of future behavioral need.

**M**otherhood is associated with pronounced behavioral changes in many species, such as altered feeding routines and increased aggressivity (1–9). These adaptations are typically attributed to the hormonal changes associated with giving birth (parturition), which have been hypothesized to activate or prime parental circuits (10, 11). One of the most notable differences between sexually inexperienced (virgin) females and mothers is their infant-directed behaviors: Whereas virgins typically avoid infants or exhibit low levels of parental behavior, mothers are highly parental (12–14). Classical studies in rats have found increased maternal responsiveness during pregnancy (9, 15, 16). This even occurs in females undergoing a caesarean section during mid- or late pregnancy and persists for weeks (15, 17–20). Correspondingly, parental behavior can be elicited in virgin rats by mimicking the hormonal changes of pregnancy (21–27), which include drastic rises in the levels of estradiol (E2) and progesterone (P4) (fig. S1A). These observations indicate hormone-mediated, preparatory neural adaptations to infant-directed behavior during pregnancy. However, despite the identification of numerous forms of pregnancy-associated neural plasticity (1, 28, 29), it remains unknown how pregnancy hormones affect parenting circuits to mediate changes in infant-directed behavior.

## Hormone-dependent, long-lasting changes to pup-directed behavior in pregnancy

Whereas virgin female rats and wild house mice typically ignore or attack pups, respectively,

virgin female laboratory mice often exhibit hormone-independent, spontaneous parental behavior (9). We first investigated when and how pup interactions change during pregnancy. We exposed female mice to pups at regular intervals before, during, and after pregnancy and scored their behaviors (Fig. 1A, *Preg*). Most aspects of parental behavior were affected by pregnancy (Fig. 1, D to G, and fig. S1, B to J), and this was particularly pronounced in late pregnant females [day 18 (D18): All D18 females retrieved pups with short latency (D18,  $39.7 \pm 10.8$  s; virgins,  $477.9 \pm 143.3$  s; Fig. 1, D to G), crouched above pups ( $17.3 \pm 3.5\%$  of assay duration), and spent most of their time in the nest (fig. S1, B and C). In addition to individual aspects of parenting, pregnancy affected behavioral sequences: Whereas D18 females performed sequences of retrieval, crouching, nest building, and grooming, virgins engaged in repetitive sniffing–grooming–nest entering episodes (Fig. 1H). The increased parental performance of D18 females could be a result of hormonal effects and/or frequent pup exposure (30). We therefore assessed pup interactions in females that were exposed to pups only as virgins and at D18 (Fig. 1B, *Dual*) and in ovariectomized females (Fig. 1C, *OVX*) (31). Pup retrieval, crouching, and time in nest differed between virgins (Vir) and D18 females in the *Preg* and *Dual* groups. By contrast, such differences were not present in the *OVX* group over similar time points and are thus primarily affected by pregnancy hormones (Fig. 1, D to G, and fig. S1, B to E). These behaviors were also up-regulated in females that were exposed to pups only once, at D18, which illustrates that the pregnancy-induced onset of parenting does not require any previous pup exposure (fig. S1, N to S). The hormonal milieu of pregnancy thus leads to an onset of specific parental behaviors in mice, and these behavioral changes are maximal in late pregnancy. Most behavioral changes

persisted until at least 1 month after parturition (D50) (Fig. 1, D and F, and fig. S1D), when hormone levels have returned to baseline (fig. S1A). These adaptations thus likely result from long-lasting remodeling of the brain by pregnancy hormones.

## Hormone action on MPOA<sup>Gal</sup> neurons is critical for parenting onset

Parenting is controlled by brain-wide circuits (32–36), several elements of which might be affected by hormones. In particular, the medial preoptic area (MPOA)—which is critical for parental behavior—has been shown to be a hormonal target (37–39). Parental behavior can be induced in virgin female rats by administration of the hormones E2, P4, prolactin, and oxytocin (21–26, 40, 41), and global knockout (KO) of their canonical receptors impairs parenting (42–45). Because combined systemic administration of E2 and P4 is most effective in triggering parenting onset (12), the underlying neural substrates are likely sensitive to both hormones. E2 and P4 can permanently modulate neuronal function through intracellular receptors that act as transcription factors (46–49). We focused on the intracellular estrogen receptor 1 (Esr1) and progesterone receptor (PR) because they are critical for parental behavior (43, 50–52) and because the long-lasting nature of pregnancy-induced behavioral changes implicates gene expression-dependent forms of plasticity. Using single-molecule fluorescence *in situ* hybridization (smFISH) from hypothalamic brain sections of virgins and D18 females, we found that Esr1-PR coexpressing neurons (as well as neurons expressing either receptor) were most enriched in MPOA subregions (fig. S2, A to F). Prolactin receptor, but not oxytocin receptor, expression was similarly enriched in the MPOA (fig. S2, G and H).

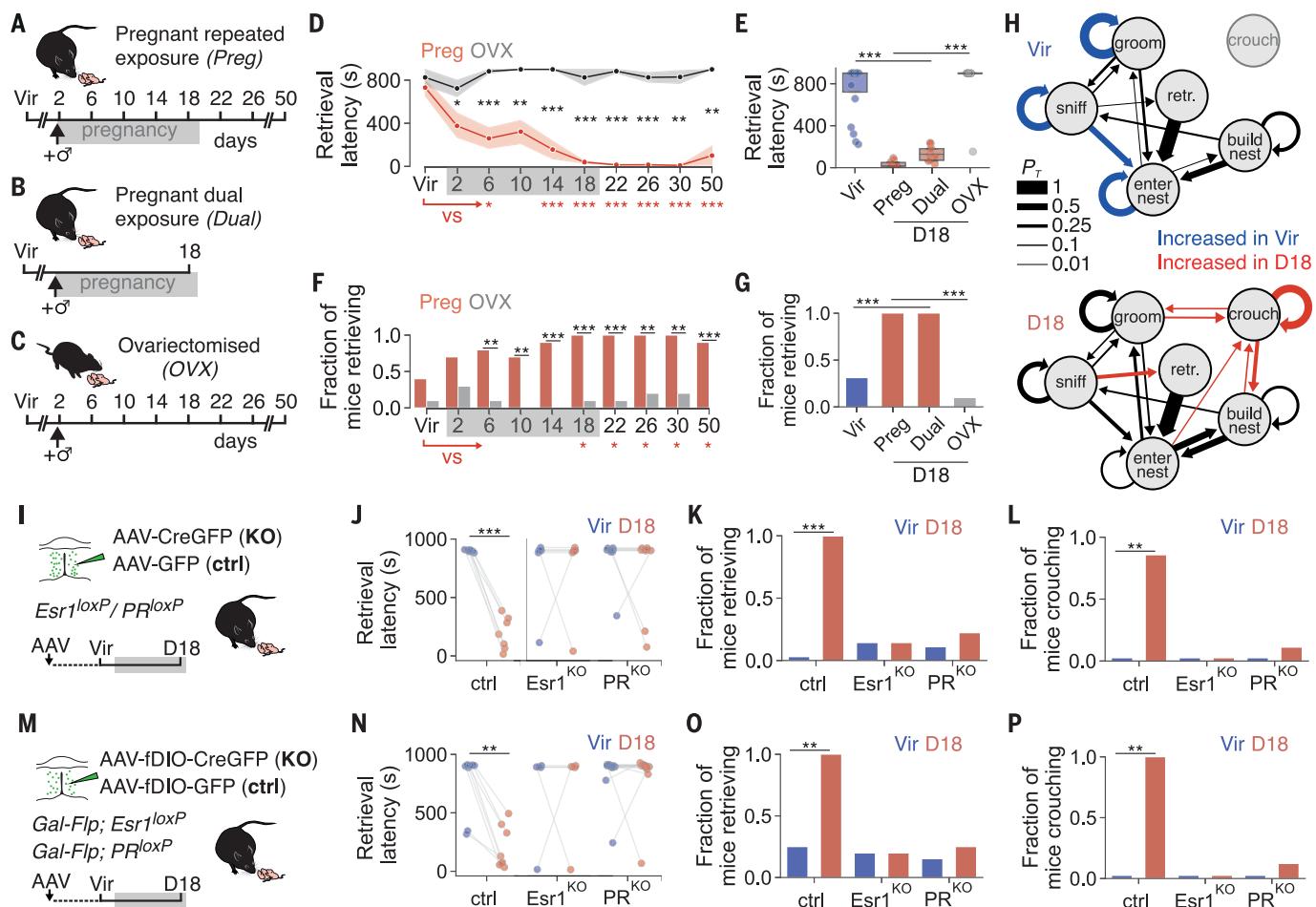
To determine whether Esr1 or PR expression in the MPOA is required for parenting onset during pregnancy, mice carrying floxed receptor alleles (materials and methods) (53) were injected into the MPOA with an adenovirus-associated virus (AAV)—expressing Cre recombinase (Fig. 1I). This resulted in local receptor KO, whereas injection of a green fluorescent protein (GFP)-expressing control AAV did not affect receptor expression (fig. S3, A to H). MPOA-specific ablation of either Esr1 or PR had no effect on pup interactions in virgins but completely blocked the pregnancy-induced up-regulation of pup retrieval, crouching, and nest time at D18 (Fig. 1, J to L, and fig. S4, A to C). By contrast, parental behaviors were normally up-regulated at D18 in animals injected with control AAVs (Fig. 1, J to L, and fig. S4, A to D).

Several overlapping populations of MPOA neurons are involved in the control of parenting, with galanin-expressing (MPOA<sup>Gal</sup>) neurons being critical for this behavior (33–35, 54–56).

<sup>1</sup>State-Dependent Neural Processing Laboratory, The Francis Crick Institute, London NW1 1AT, UK. <sup>2</sup>Bioinformatics and Biostatistics Science Technology Platform, The Francis Crick Institute, London NW1 1AT, UK.

\*Corresponding author. Email: [jonny.kohl@crick.ac.uk](mailto:jonny.kohl@crick.ac.uk)

†These authors contributed equally to this work.



**Fig. 1. Hormone action on MPOA<sup>Gal</sup> neurons is critical for pregnancy-induced onset of parental behavior.** (A to C) Testing pup-directed behavior in repeatedly pup-exposed pregnant females (Preg,  $n = 10$ ) (A), pregnant females exposed to pups twice (Dual,  $n = 9$ ) (B), and repeatedly pup-exposed ovariectomized females (OVX,  $n = 10$ ) (C). Day of pregnancy [(A) and (B)] or relative to pairing with male (C) are shown. (D and F) Parental behaviors in Preg group. Within-group [Preg, virgin (Vir) versus each subsequent time point; red asterisks] and between-group (Preg versus OVX; black asterisks) comparisons are shown. (E and G) Comparison of Vir and D18 time points across groups. Note that virgins from Preg and Dual groups are pooled (fig. S1M). (H) Behavioral state transition diagrams for Vir and D18 females (Preg,

$n = 10$ ). Average transition probabilities ( $P_T$ ) between behaviors are shown, and differences between Vir and D18 females are highlighted if  $P < 0.05$  ( $U$  test; see materials and methods). retr., retrieval. (I) AAV-mediated ablation (KO) of Esr1 or PR in MPOA and control (ctrl). (J to L) Effects of MPOA-wide KO of Esr1 or PR on pup-directed behaviors ( $n = 7$ , 8, and 9 mice). (M) KO of Esr1 or PR in MPOA<sup>Gal</sup> neurons. (N to P) Effects of MPOA<sup>Gal</sup>-specific KO of Esr1 or PR on pup-directed behaviors ( $n = 8$ , 5, and 13 mice). Statistics by Kaplan-Meier survival analysis with log rank test in (D), (E), (J), and (N) and by Fisher's exact test with Benjamini-Hochberg adjustment for multiple comparisons in (F), (G), (K), (L), (O), and (P). Shaded area in (D) is SEM. \*\*\* $P < 0.001$ ; \*\* $P < 0.01$ ; \* $P < 0.05$ .

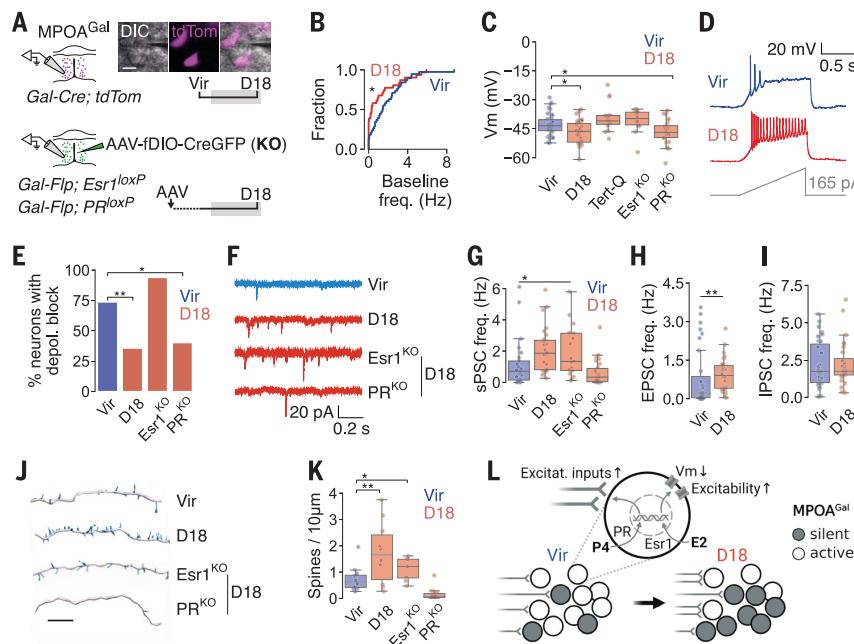
MPOA<sup>Gal</sup> neurons, most of which express Esr1 and PR (fig. S3, J and K), constitute ~20% of MPOA neurons (35). To determine whether hormonal sensitivity of this subpopulation is necessary for pregnancy-induced behavioral adaptations, we made a knock-in mouse line expressing Flp recombinase in Gal neurons (fig. S3I and materials and methods) and crossed this allele into mice with floxed receptor genes. AAV-mediated ablation of either Esr1 or PR in MPOA<sup>Gal</sup> neurons fully recapitulated the effects observed after MPOA-wide receptor KO (Fig. 1, M to P, and fig. S4, E to H). By contrast, pup contact latency—a parameter not modulated by pregnancy—was not affected by this manipulation (fig. S4I). The parental behaviors of these receptor-

ablated animals remained impaired even after giving birth (D22; fig. S5), which indicates that the lack of hormone-mediated behavioral preparations during pregnancy cannot be compensated for by the subsequent endocrine events of parturition. Direct action of E2 and P4 on MPOA<sup>Gal</sup> neurons through their intracellular hormone receptors is therefore necessary for pregnancy-mediated increases in parental behavior.

#### Long-lasting hormonal remodeling of MPOA<sup>Gal</sup> neurons during pregnancy

We next investigated how pregnancy affects MPOA<sup>Gal</sup> neurons and performed patch clamp recordings in brain slices from virgins and D18 females (Fig. 2A, top). Recorded neurons were

neurobiotin-filled and reconstructed to assess morphological changes. MPOA<sup>Gal</sup> neurons exhibited lower baseline firing and resting membrane potential in late pregnancy (Fig. 2, B and C, and supplementary text), with a significantly higher proportion of silent neurons at D18 (fig. S6, A and B). This silencing was abolished by Tertiapin-Q (TQ) and might thus be mediated by GIRK channels (Fig. 2C). At the same time, MPOA<sup>Gal</sup> neurons were more excitable at D18 and less frequently exhibited depolarization block (Fig. 2, D and E, and fig. S6K). We observed a reduction in action potential half-width at D18 (fig. S6E), which hints at increased function of delayed rectifier K<sup>+</sup> channels that repolarize neurons to permit sustained firing (57). These effects on neuronal membrane



**Fig. 2. Hormonal remodeling of MPOA<sup>Gal</sup> neurons.** (A) Whole-cell recordings from wild-type (top) and receptor-deleted (KO) (bottom) MPOA<sup>Gal</sup> neurons. (B) Cumulative distribution of baseline firing frequency (Vir and D18; 33 and 21 cells from  $n = 15$  and 7 mice). (C) Resting membrane potential of control and receptor-deleted MPOA<sup>Gal</sup> neurons and recordings in presence of GIRQ channel blocker Tertiapin-Q (Tert-Q) (38, 32, 15, 18, and 26 cells from  $n = 15$ , 9, 3, 3, and 5 mice). (D) Example current clamp recording traces of cells with (Vir) and without (D18) depolarization block. (E) Percentage of neurons exhibiting depolarization block (34, 30, 18, and 25 cells from  $n = 15$ , 8, 3, and 5 mice). (F) Example voltage clamp recording traces with sPSCs. (G) sPSC frequency (21, 23, 18, and 26 cells from  $n = 9$ , 6, 3, and 5 mice). (H) and (I) Excitatory postsynaptic current (EPSC) (Vir and D18; 31 and 30 cells from  $n = 5$  and 4 mice) (H) and inhibitory postsynaptic current (IPSC) (31 and 28 cells from  $n = 5$  and 4 mice) (I) frequency. (J) Dendritic segments of MPOA<sup>Gal</sup> neurons with spines. (K) Spine density (14, 10, 8, and 15 cells from  $n = 10$ , 4, 3, and 4 mice). (L) Summary scheme for hormonal remodeling of MPOA<sup>Gal</sup> neurons. Statistics by  $U$  test in (B); one-way analysis of variance (ANOVA) with Dunnett's post hoc test in (C), (G), and (K); Fisher's exact test with Benjamini-Hochberg adjustment in (E); and K-S test in (H) and (I). Scale bars, 20  $\mu$ m (A) and 10  $\mu$ m (J). \*\*\* $P < 0.001$ ; \*\* $P < 0.01$ ; \* $P < 0.05$ .

properties were already apparent in midpregnancy (D10; fig. S6, B to E) and were linked because 80% of silent MPOA<sup>Gal</sup> neurons also did not exhibit depolarization block at D18 (fig. S6O). MPOA<sup>Gal</sup> silencing was not a result of increased inhibitory synaptic inputs: Although these neurons received more spontaneous postsynaptic currents (sPSCs) at D18, this was because of an increase in excitatory inputs (Fig. 2, F to I, and fig. S6M), which predominantly targeted spontaneously active—i.e., nonsilenced—neurons (fig. S6N). Correspondingly, MPOA<sup>Gal</sup> neurons had more dendritic spines at D18 (Fig. 2, J and K). This remodeling of synaptic inputs was also already detectable at D10 (fig. S6, I and J). We did not observe changes to sPSC amplitude and dynamics (fig. S6L) or to dendritic complexity and somatic volume (fig. S6, P and Q) (58). Pregnancy therefore reduces the baseline activity of MPOA<sup>Gal</sup> neurons while increasing their excitability and promoting the recruitment of excitatory synaptic inputs. Pregnancy did not

have equivalent effects on Gal-negative MPOA neurons, which highlights the specificity of MPOA<sup>Gal</sup> neuronal remodeling (fig. S7).

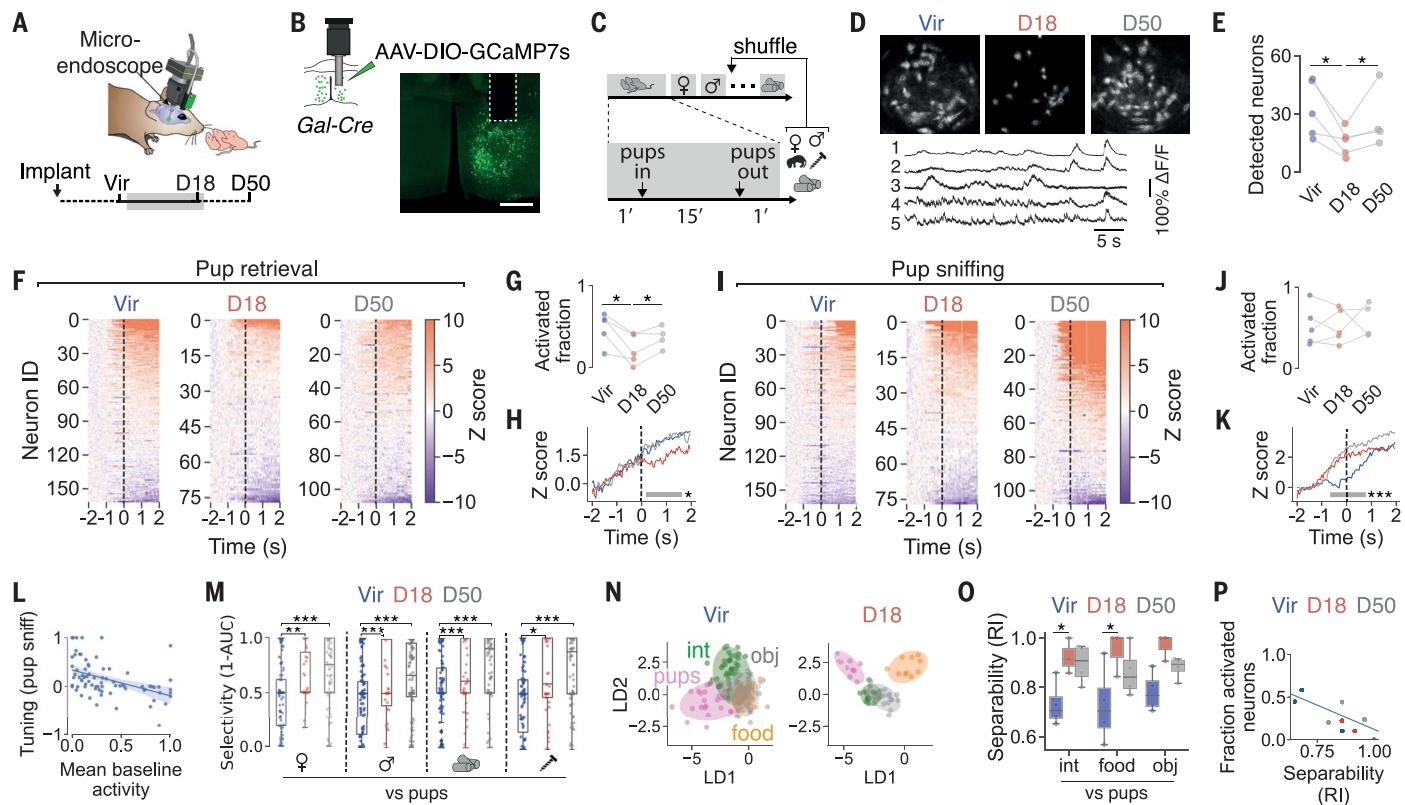
To address whether these biophysical and morphological changes were the result of direct hormonal action, we recorded from MPOA<sup>Gal</sup> neurons in which Esr1 or PR were deleted (Fig. 2A, bottom). Ablation of these receptors returned specific, nonoverlapping aspects of D18 neuronal physiology to a virgin-like state: Esr1 deletion specifically prevented pregnancy-induced silencing and changes to excitability (Fig. 2, C and E) but did not affect synaptic inputs and spine density (Fig. 2, F, G, J, and K). By contrast, PR deletion selectively abolished the up-regulation of synaptic inputs and spine density (Fig. 2, G and K) without affecting membrane properties (Fig. 2, C and E). Transduction with a control AAV had no effect (fig. S6, R to W). E2 and P4 therefore control discrete aspects of pregnancy-induced plasticity in MPOA<sup>Gal</sup> neurons: Whereas E2 tunes membrane potential and intrinsic excitability, P4

mediates the recruitment of additional excitatory synaptic inputs (Fig. 2L). To assess how long-lasting these changes were, we recorded from MPOA<sup>Gal</sup> neurons in mothers shortly after parturition (D22) and at D50, when pregnancy- and parturition-associated hormone levels have returned to baseline (fig. S8A). MPOA<sup>Gal</sup> resting membrane potential and firing frequency remained reduced at D22 and only returned to virgin-like levels at D50 (fig. S8, B and C), whereas neuronal excitability reverted immediately after parturition (fig. S8, E and F). By contrast, synaptic inputs and spine density showed a long-lasting up-regulation (fig. S8, G and H). Similar to the lasting behavioral effects of receptor ablation, its physiological effects persisted in mothers (fig. S8, I to K). These observations suggest that pregnancy hormones permanently alter the circuit integration of MPOA<sup>Gal</sup> neurons, thereby providing a cellular substrate for the long-lasting behavioral effects of pregnancy.

### Reorganization of MPOA<sup>Gal</sup> neuronal and neural population activity during pregnancy

We next investigated the effects of pregnancy on MPOA<sup>Gal</sup> neural activity in vivo. We performed longitudinal, cellular-resolution calcium imaging from MPOA<sup>Gal</sup> neurons in females exposed to pups and a set of social and nonsocial stimuli (Fig. 3, A to C, and fig. S9A) (59). Consistent with the silencing observed in our slice physiology recordings, the number of detectable (nonsilent) MPOA<sup>Gal</sup> neurons was significantly reduced at D18 in vivo (Fig. 3, D and E). This reduction was not the result of a decline in the number of GCaMP-expressing MPOA<sup>Gal</sup> neurons over time or increased calcium buffering by rising GCaMP levels because it was reversible (Fig. 3E) and did not occur in virgin females recorded at identical time points (fig. S9B). It also did not result from potential shifts in the recording plane because we observed this effect when imaging ex vivo (fig. S9, E and F). Finally, the number of detected neurons was not significantly decreased in MPOA-wide recordings (fig. S9, C and D). Pregnancy-induced silencing therefore preferentially occurs in MPOA<sup>Gal</sup> neurons, consistent with our electrophysiological findings in brain slices (fig. S7, A to E).

The fraction of neurons activated during pup retrieval and pup grooming decreased at D18 (Fig. 3, F and G, and fig. S10A). By contrast, similar fractions of MPOA<sup>Gal</sup> neurons were active during pup sniffing in virgins and at D18, but their responses occurred with shorter latency at D18 and D50 (Fig. 3, I to K), which indicates a higher excitability of MPOA<sup>Gal</sup> neurons to pup stimuli during and after pregnancy. Pregnancy therefore sparsens MPOA<sup>Gal</sup> population activity during parental actions and makes these neurons more excitable to pup stimuli. The baseline activity of individual



**Fig. 3. Reorganization of MPOA<sup>Gal</sup> population activity during pregnancy.**

(A) Recording setup for miniature microscope recordings. (B) *Gal-Cre* animals were injected into the MPOA with AAV-FLEX-GCaMP7s and implanted with a GRIN lens. GCaMP7s expression and GRIN lens position are shown. (C) Experimental design (see materials and methods). (D) Sample recording frames with detected neurons and example activity traces from a virgin. (E) Number of detected (nonsilent) neurons per animal ( $n = 5$  mice). (F and I) Temporal profile of MPOA<sup>Gal</sup> responses during pup retrieval (F) or sniffing (I) in virgins, at D18, and at D50 (162, 77, and 93 neurons from  $n = 5$  mice). Dashed lines indicate action onset. Order was based on hierarchical clustering sorted by mean cluster response onset. (G and J) Fraction of neurons with positive evoked response during pup retrieval (G) or sniffing (J) ( $n = 5$  mice). (H and K) Averaged Z score for neurons activated during pup retrieval in virgins, at D18, and at D50 (115, 41, and 63 neurons from 5, 5, and 4 mice) (H) or sniffing (122, 51, and 86 neurons from 5, 5, and 4 mice) (K). Statistics by two-way ANOVA with Tukey post hoc test, and gray bars indicate periods of significant difference for Vir versus D18 and Vir versus D50. (L) Correlation

between normalized tuning index for responses to pup sniffing and normalized mean baseline activities at D18 (coefficient of determination  $r^2 = 0.202$ ;  $P < 0.001$ ). (M) Selectivity of chemoinvestigation-associated responses for indicated stimulus pairs at Vir, D18, and D50 (142, 35, and 108 cells from  $n = 4$ , 3, and 4 mice) compared with pups. A selectivity score of 1 means the neuron is only activated during pup sniffing, a score of 0 means selective activation during sniffing of other stimulus, and a score of 0.5 equals a nonselective response (see materials and methods). (N) Example MPOA<sup>Gal</sup> neuronal activity at Vir and D18 during object investigation in LDA space (int, intruder; obj, object). Temporal bins were used as features. Ellipsoids represent 95% confidence area of neuronal activity to each stimulus. (O) Separability of indicated stimulus combinations by the MPOA<sup>Gal</sup> population [Rand Index (RI),  $n = 4$ , 3, and 4 mice]. (P) Correlation between separability (pup versus intruder) and activated fraction of neurons during pup retrieval ( $r^2 = 0.56$ ;  $P < 0.001$ ). Statistics by paired *t* tests in (E), (G), and (J); mixed linear model with mouse ID as group in (M); linear regression in (L) and (P); and unpaired *t* tests in (O). Scale bar in (B), 500  $\mu$ m. \*\*\* $P < 0.001$ ; \*\* $P < 0.01$ ; \* $P < 0.05$ .

MPOA<sup>Gal</sup> neurons was negatively correlated with their tuning to pup stimuli at D18, thereby linking neuronal silencing to stronger pup-evoked responses (Fig. 3L). To address pregnancy-induced differences in how MPOA<sup>Gal</sup> neurons represent pup stimuli, we examined their activity patterns during chemoinvestigation of pups and other stimuli (Fig. 3C). MPOA<sup>Gal</sup> neuronal stimulus selectivity for, and response strength to, pups increased in late pregnancy (Fig. 3M and fig. S11, A to E). Similarly, although linear discriminant analysis (LDA) could not separate pup representations in MPOA<sup>Gal</sup> neurons well from those of other stimuli in virgins, separability of pup representations from those of other stimuli was en-

hanced at D18 (Fig. 3, N and O). Stimulus separability was positively correlated with population sparsening, thereby linking improved pup representations with effective encoding of parental actions (Fig. 3P). At D50, the numbers of spontaneously active and retrieval-activated neurons had largely returned to virgin levels (Fig. 3, E and G), mirroring our findings in brain slices (fig. S8, B to D). By contrast, pup stimulus selectivity and separability showed a long-lasting increase (Fig. 3, M to O). These findings demonstrate that pregnancy leads to a pronounced sparsening of spontaneous and parenting-associated activity in MPOA<sup>Gal</sup> neurons and to increased selectivity for infant stimuli.

## Discussion

Considerable progress has been made in uncovering the functional architecture of parenting circuits (9, 32–36), but little is known about how hormones alter these circuits to ensure state-dependent behavioral flexibility. We discovered that pregnancy hormone action on MPOA<sup>Gal</sup> neurons—a hub in parenting circuits—is critical to instruct a preparatory change in infant-directed behavior. The ovarian hormones E2 and P4 each control distinct aspects of pregnancy-induced neural remodeling: Whereas E2 transiently silences MPOA<sup>Gal</sup> neurons and increases their excitability, P4 permanently remodels this circuit element by recruiting synaptic inputs. This results in sparsened population

activity during parental behavior and in potentiated, more-selective responses to pup stimuli. We propose that the resulting increase in signal-to-noise ratio, both in individual neurons and at the population level, enables more efficient encoding of parental motor actions by MPOA<sup>Gal</sup> neurons. Population sparsening through silencing might contribute to setting up the circuit for efficient parental behavior by selectively recruiting inputs onto active MPOA<sup>Gal</sup> neurons during pregnancy. Once rewired, this circuit could then drive robust parenting in response to pup cues, whereas release from silencing during the postpartum period might allow for recruitment of these neurons during nonparental social interactions.

The long-lasting, P4-mediated remodeling of MPOA<sup>Gal</sup> synaptic inputs provides a cellular correlate for the long-lasting behavioral changes that we observe. Although parturition-associated hormonal changes and subsequent maternal experience cannot compensate for lack of hormonal remodeling during pregnancy, these factors might normally contribute to long-term enhancement of maternal behavior (maternal memory) (9). Repeated and/or prolonged co-housing of virgins with pups results in elevated levels of parenting through sensitization (60, 61), and parental care can be socially transmitted by mothers (62). It is unclear whether these paradigms result in similar neuronal changes. Ablating MPOA<sup>Gal</sup> neurons or making them hormone-insensitive both abolish pup retrieval, but optogenetic activation of these neurons elicits pup grooming in virgins (35). Although it remains unknown which neuronal ensembles are recruited by artificial, acute stimulation, they seem to differ from the sparse populations that drive robust parenting in late pregnancy.

E2 silences MPOA<sup>Gal</sup> neurons beyond parturition, presumably by up-regulating GIRK channel expression, whereas the more transient increases in excitability are likely a result of potentiated function of delayed rectifier K<sup>+</sup> channels (57). The identity of the additional excitatory inputs recruited by P4 remains unknown. They might constitute long-range afferents conveying pup sensory information because most local MPOA neurons are GABAergic [ $\gamma$ -aminobutyric acid (GABA)] (54). Future work will characterize the identity and functional role of the cellular pathways targeted by Esr1 and PR. Coexpression of these receptors is not unique to Gal-expressing MPOA neurons (fig. S2C). We hypothesize that permissive chromatin states in these neurons allow for cell type-specific, hormonally induced target gene expression. MPOA<sup>Gal</sup> neurons form molecularly distinct subpopulations (54) that might be differentially affected by pregnancy hormones. We also expect MPOA<sup>Gal</sup> and other neurons in parenting circuits to be sensitive to additional pregnancy hormones, such as prolactin, placental lactogens, allopregnanolone,

and oxytocin. Prolactin for instance, which acts on the MPOA in early pregnancy to reduce physical activity (63), might also contribute to early changes in pup-directed behavior (Fig. 1, D and F).

Unlike laboratory mice, the majority of wild virgin female mice exhibit infanticide (64). Our work provides mechanisms through which hormones might act in parental circuits of wild mice and other species that critically depend on endocrine changes for the onset of short-latency maternal behavior, such as rats, rabbits, and sheep (9). The neural activity changes observed in this work—i.e., population sparsening and increased stimulus selectivity and discriminability—are reminiscent of changes that occur during critical periods in the developing brain (65). Our work therefore suggests that pregnancy hormones open a window of adult plasticity during which neural remodeling orchestrates behavioral adaptations for the future challenges of motherhood.

## REFERENCES AND NOTES

1. P. J. Brunton, J. A. Russell, *Nat. Rev. Neurosci.* **9**, 11–25 (2008).
2. E. N. Makarova, E. D. Kochubei, N. M. Bazhan, *Neurosci. Behav. Physiol.* **40**, 263–267 (2010).
3. M. A. Mann, C. Konen, B. Svare, *Horm. Behav.* **18**, 140–160 (1984).
4. J. A. Russell, A. J. Douglas, C. D. Ingram, *Prog. Brain Res.* **133**, 1–38 (2001).
5. M. C. Buholt-Averseng, J. Goyens, *Aggress. Behav.* **7**, 111–121 (1981).
6. R. Haddad-Tóvolli et al., *Nat. Metab.* **4**, 424–434 (2022).
7. R. D. Lisk, *Anim. Behav.* **19**, 606–610 (1971).
8. T. Topilko et al., *Neuron* **110**, 1385–1399.e8 (2022).
9. M. Numan, *The Parental Brain: Mechanisms, Development, and Evolution* (Oxford Univ. Press, 2020).
10. M. Numan, B. Woodside, *Behav. Neurosci.* **124**, 715–741 (2010).
11. M. Numan, L. J. Young, *Horm. Behav.* **77**, 98–112 (2016).
12. J. S. Rosenblatt, A. D. Mayer, A. L. Giordano, *Psychoneuroendocrinology* **13**, 29–46 (1988).
13. A. S. Fleming, J. S. Rosenblatt, *J. Comp. Physiol. Psychol.* **86**, 957–972 (1974).
14. M. Numan, T. R. Insel, *The Neurobiology of Parental Behavior*, vol. 1 of *Hormones, Brain, and Behavior*, G. F. Ball, J. Balthazard, R. J. Nelson, Eds. (Springer, 2003).
15. J. S. Rosenblatt, *Am. J. Orthopsychiatry* **39**, 36–56 (1969).
16. B. M. Slotnick, M. L. Carpenter, R. Fusco, *Horm. Behav.* **4**, 53–59 (1973).
17. R. S. Bridges, *Physiol. Behav.* **18**, 487–490 (1977).
18. J. Labriola, *Proc. Soc. Exp. Biol. Med.* **83**, 556–557 (1953).
19. H. Moltz, D. Robbins, M. Parks, *J. Comp. Physiol. Psychol.* **61**, 455–460 (1966).
20. B. P. Wiesner, N. M. Sheard, *Maternal Behavior in the Rat* (Oliver and Boyd, 1933).
21. R. S. Bridges, M. Numan, P. M. Ronsheim, P. E. Mann, C. E. Lupini, *Proc. Natl. Acad. Sci. U.S.A.* **87**, 8003–8007 (1990).
22. A. D. Mayer, H. B. Ahdieh, J. S. Rosenblatt, *Horm. Behav.* **24**, 152–173 (1990).
23. H. Moltz, M. Lubin, M. Leon, M. Numan, *Physiol. Behav.* **5**, 1373–1377 (1970).
24. C. A. Pedersen, J. A. Ascher, Y. L. Monroe, A. J. Prange Jr., *Science* **216**, 648–650 (1982).
25. O. Riddle, E. L. Lahr, R. W. Bates, *Exp. Biol. Med.* **32**, 730–734 (1935).
26. J. S. Rosenblatt, A. Olufowobi, H. I. Siegel, *Horm. Behav.* **33**, 104–114 (1998).
27. J. Terkel, J. S. Rosenblatt, *J. Comp. Physiol. Psychol.* **65**, 479–482 (1968).
28. K. M. Hillerer, V. R. Jacobs, T. Fischer, L. Aigner, *Neural Plast.* **2014**, 574159 (2014).
29. N. Zilkha, N. Scott, T. Kimchi, *Annu. Rev. Neurosci.* **40**, 273–305 (2017).
30. B. J. Marlin, M. Mitre, J. A. D'Amour, M. V. Chao, R. C. Froemke, *Nature* **520**, 499–504 (2015).
31. P. M. Wise, A. Ratner, *Neuroendocrinology* **30**, 15–19 (1980).
32. P. B. Chen et al., *Cell* **176**, 1206–1221.e18 (2019).
33. Y.-Y. Fang, T. Yamaguchi, S. C. Song, N. X. Tritsch, D. Lin, *Neuron* **98**, 192–207.e10 (2018).
34. J. Kohl et al., *Nature* **556**, 326–331 (2018).
35. Z. Wu, A. E. Autry, J. F. Bergan, M. Watabe-Uchida, C. G. Dulac, *Nature* **509**, 325–330 (2014).
36. S. Stagkourakis et al., *Cell* **182**, 960–975.e15 (2020).
37. M. Numan, J. S. Rosenblatt, B. R. Komisaruk, *J. Comp. Physiol. Psychol.* **91**, 146–164 (1977).
38. S. E. Fahrbach, D. W. Pfaff, *Horm. Behav.* **20**, 354–363 (1986).
39. Y. Tsuneoka et al., *J. Comp. Neurol.* **521**, 1633–1663 (2013).
40. R. S. Bridges, P. M. Ronsheim, *Endocrinology* **126**, 837–848 (1990).
41. A. D. Mayer, M. A. Monroy, J. S. Rosenblatt, *Horm. Behav.* **24**, 342–364 (1990).
42. B. K. Lucas, C. J. Ormandy, N. Binart, R. S. Bridges, P. A. Kelly, *Endocrinology* **139**, 4102–4107 (1998).
43. S. Ogawa et al., *Endocrinology* **139**, 5070–5081 (1998).
44. M. E. Rich, E. J. deCárdenes, H.-J. Lee, H. K. Caldwell, *PLOS ONE* **9**, e98839 (2014).
45. Y. Takayanagi et al., *Proc. Natl. Acad. Sci. U.S.A.* **102**, 16096–16101 (2005).
46. E. R. Levin, *Trends Endocrinol. Metab.* **20**, 477–482 (2009).
47. N. Z. Lu et al., *Pharmacol. Rev.* **58**, 782–797 (2006).
48. P. Thomas, Y. Pang, *Neuroendocrinology* **96**, 162–171 (2012).
49. C. D. Toran-Allerand et al., *J. Neurosci.* **22**, 8391–8401 (2002).
50. R. D. Brinton et al., *Front. Neuroendocrinol.* **29**, 313–339 (2008).
51. A. C. Ribeiro et al., *Proc. Natl. Acad. Sci. U.S.A.* **109**, 16324–16329 (2012).
52. M. Numan et al., *Brain Res.* **830**, 358–371 (1999).
53. P. Antonson, Y. Omoto, P. Humire, J.-A. Gustafsson, *Biochem. Biophys. Res. Commun.* **424**, 710–716 (2012).
54. J. R. Moffitt et al., *Science* **362**, eaau5324 (2018).
55. Y.-C. Wei et al., *Nat. Commun.* **9**, 279 (2018).
56. C. Yoshihara et al., *Cell Rep.* **35**, 109204 (2021).
57. J. A. McHenry et al., *Nat. Neurosci.* **20**, 449–458 (2017).
58. L. Keyser-Marcus et al., *Brain Res. Bull.* **55**, 737–745 (2001).
59. K. K. Ghosh et al., *Nat. Methods* **8**, 871–878 (2011).
60. J. S. Rosenblatt, *Science* **156**, 1512–1514 (1967).
61. D. S. Slotenberg, E. F. Rissman, *J. Neuroendocrinol.* **23**, 345–354 (2011).
62. I. Carcea et al., *Nature* **596**, 553–557 (2021).
63. S. R. Ladyman, K. M. Carter, M. L. Gillett, Z. K. Aung, D. R. Grattan, *elife* **10**, e62260 (2021).
64. M. M. McCarthy, F. S. vom Saal, *Physiol. Behav.* **35**, 843–849 (1985).
65. G. B. Smith et al., *Nat. Neurosci.* **18**, 252–261 (2015).
66. J. Kohl, AmmariMonaca2023, Figshare (The Francis Crick Institute, 2023); <https://doi.org/10.25418/crick.c.6706164.v1>.
67. J. Kohl, M. Cao, F. Monaca, Hormone-mediated neural remodeling orchestrates parenting onset during pregnancy, version v1, Zenodo (2023); <https://doi.org/10.5281/zenodo.8222741>.

## ACKNOWLEDGMENTS

We thank M. Strom for advice on and production of viruses and A. Schaefer, F. Iacaruso, P. Znamenskiy, F. Guillemot, J. Briscoe, Y. Isogai, L. Newman, and all members of the State-Dependent Neural Processing Laboratory for discussions and comments on the manuscript. We thank the animal facilities at the Francis Crick Institute for animal care and technical assistance as well as the Light Microscopy, Experimental Histopathology, Genetic Modification Services, and Bioinformatics science technology platforms; the Making Lab; and the Mechanical Workshop. **Funding:** This study received support from the Francis Crick Institute, core funding FC001153. The Francis Crick Institute receives its funding from Cancer Research UK, the UK Medical Research Council, and the Wellcome Trust (J.K.). This study was also supported by European Research Council starting grant ERC-2019-STG847873 (J.K.). NARSAD Young Investigator Award BB/V016946/1 (J.K.), and a Boehringer Ingelheim PhD fellowship (M.C.). **Author contributions:** Conceptualization: J.K., R.A., F.M., and M.C. Methodology: R.A., F.M., M.C., P.W., and J.K. Investigation: R.A., F.M., M.C., P.W., M.L., D.S.-L., and J.K. Formal analysis: R.A., F.M., M.C., E.N., N.A.D.G.,

N.B., D.S.-L., and J.K. Funding acquisition: J.K. and M.C. Supervision: J.K. Writing – original draft: J.K. Writing – review & editing: J.K., F.M., R.A., and M.C. **Competing interests:** The authors declare no competing interests. **Data and materials availability:** All data are available in the manuscript or the supplementary materials or are deposited at Figshare (66). Analysis code is available online (<https://github.com/FrancisCrickInstitute/AmmariMonaca2023>) (67). **License information:** Copyright © 2023 the authors, some rights reserved;

exclusive licensee American Association for the Advancement of Science. No claim to original US government works. <https://www.science.org/about/science-licenses-journal-article-reuse>

#### SUPPLEMENTARY MATERIALS

[science.org/doi/10.1126/science.adl0576](https://science.org/doi/10.1126/science.adl0576)  
Materials and Methods

Supplementary Text  
Figs. S1 to S11  
References (68–78)  
MDAR Reproducibility Checklist  
Movie S1

Submitted 11 May 2023; accepted 9 August 2023  
[10.1126/science.adl0576](https://science.org/doi/10.1126/science.adl0576)