



BIO-463

Genomics and bioinformatics

Lecture 5:

Population genetics: mutations, selection & drift

Prof. Anne-Florence Bitbol

EPFL

Schedule of this class

Lecture 1: Feb 18

Lecture 2: Feb 25

Lecture 3: March 4

Lecture 4: March 11

Lecture 5: March 18 – Assignment 1 available on March 20

Lecture 6: March 25 – Problem class devoted to **assignment 1**; deadline on **March 28**

Lecture 7: April 1

Lecture 8: April 8

Lecture 9: April 15 – Assignment 2 available on April 18

Lecture 10: April 29 – Problem class devoted to **assignment 2**; deadline on **May 2**

Lecture 11: May 6 – Mini-projects available on April 28; choose yours by May 6

Lecture 12: May 13

Lecture 13: May 20

Lecture 14: May 27 – Mini-project deadline on **May 30**

Information about the first assignment

- Assignment released on March 20, problem class on March 25, deadline on March 28

This assignment is a graded problem set, and will count for **25% of your final grade**

You can discuss with TAs and with fellow students about the problem set, but in the end, you should hand in a **personal solution**

Detected plagiarism will result in a reduction of your grade

The expected language is R, as in all the class BIO-463 – some R functions and libraries may be recommended in the problems

Please hand in your solution in two files:

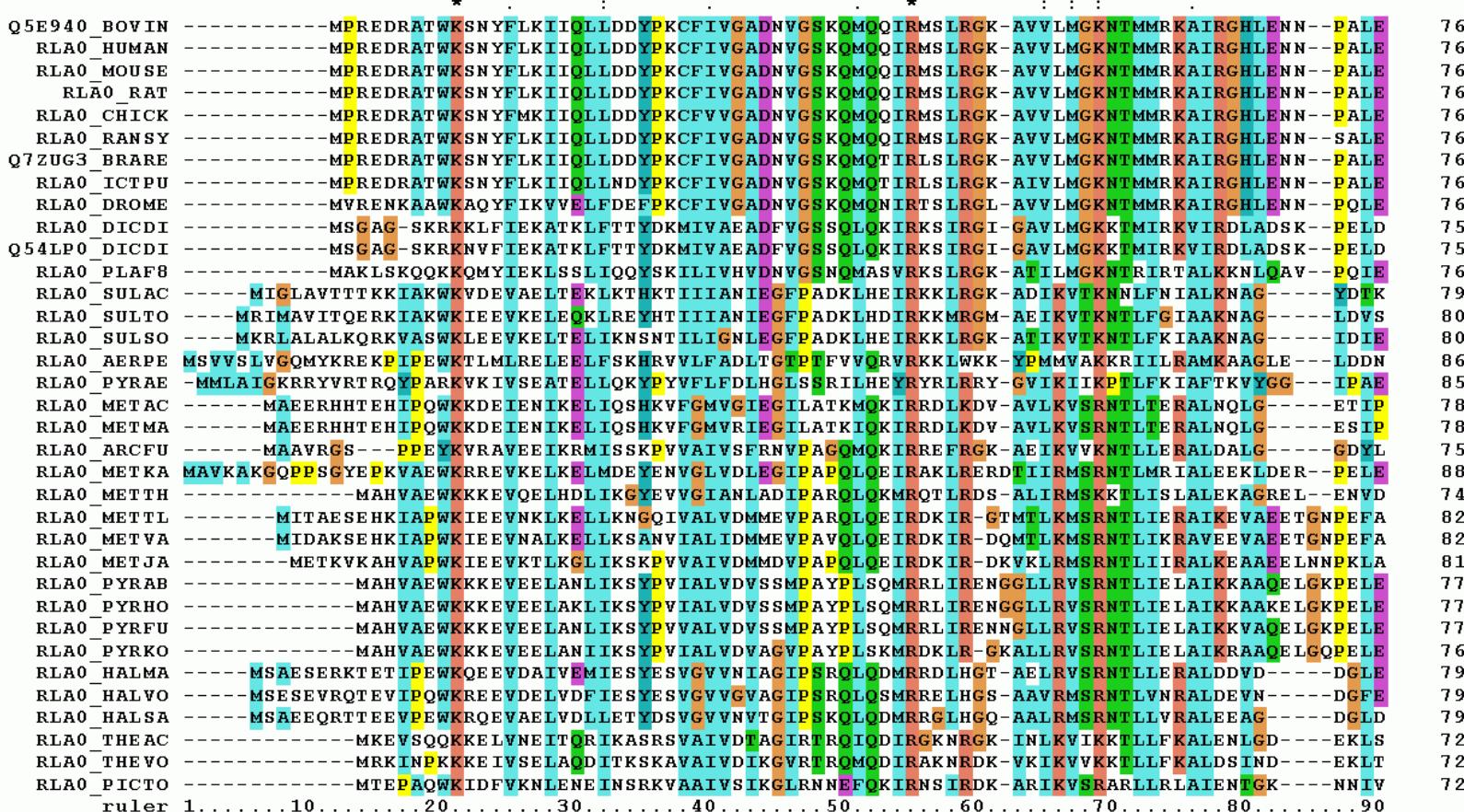
- one should be your **source file**, with the following format: **.Rmd, .qmd**
- the other one should be the **html file** deriving from your source file

You will have to hand in your solution **via Moodle by Friday March 28**

Reminder: sequence data

Multiple sequence alignments

Focus on amino-acid sequences of proteins (translated from the coding part of genomes)



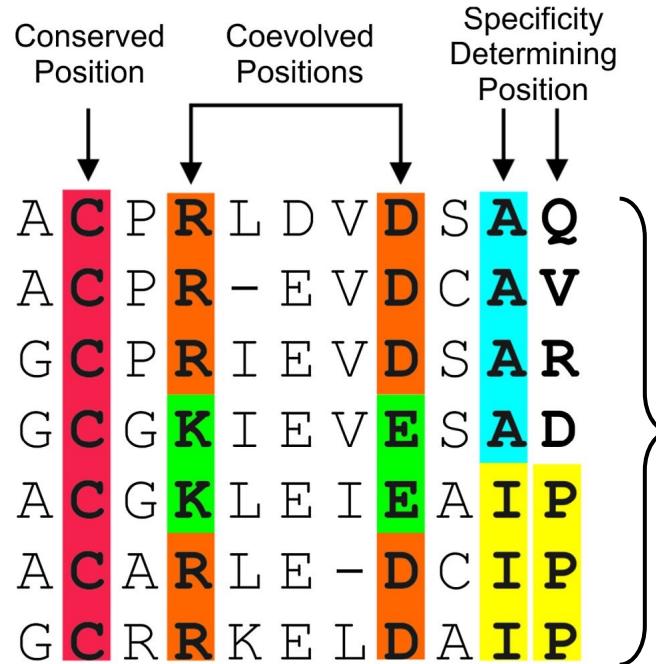
Acidic ribosomal protein P0 (first 90 positions) from several organisms

Row = sequence
Column = site
(given position in 3D structure)

Colors = level of conservation

Reminder: sequence data

▪ Multiple sequence alignments



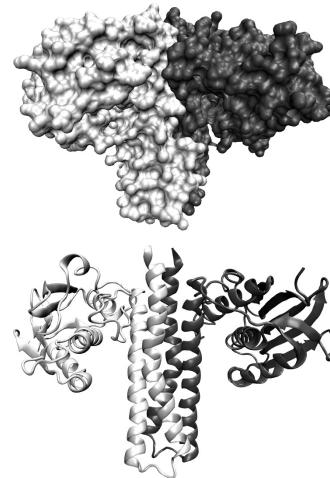
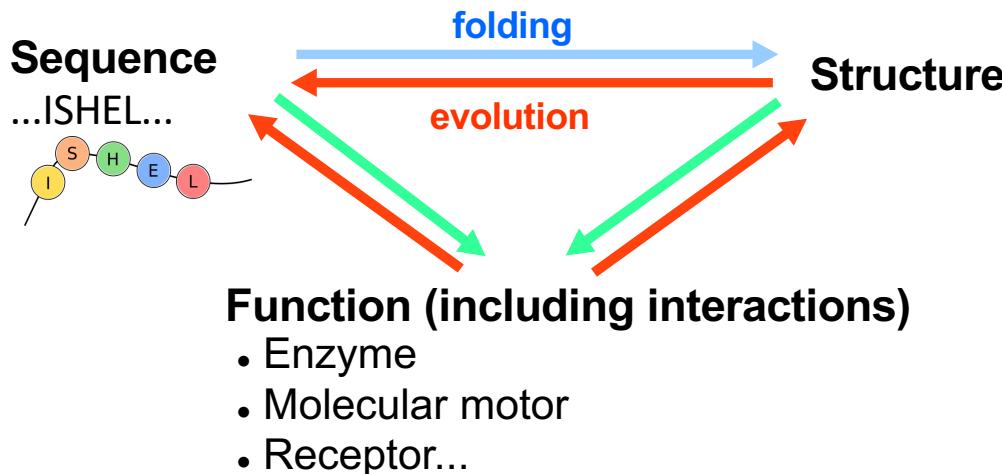
Multiple sequence alignment
of homologous protein sequences:
same ancestry,
similar function,
similar 3D structure

Teppa et al 2012

Special sites (e.g. highly conserved ones): signature of natural selection on these sites
We only observe sequences that have survived natural selection

Protein sequences and natural selection

■ Evolution of proteins



Mutations act on sequences
BUT
selection acts on function

- Heteropolymers made of 20 types of amino-acids (monomers) → $\sim 20^{100}$ possible proteins
- A given natural protein folds into a compact and (almost) unique 3D **structure**
- It has specific **interactions** with other molecules → **function**
- Experiment: random proteins do not fold properly **Socolich et al. (2005)**

→ Natural proteins are special, due to natural selection for folding and function

Protein sequences and natural selection

■ A way to detect selection: dN/dS

The genetic code has some redundancies:

		Second letter					
		U	C	A	G		
First letter	U	UUU UUC UUA UUG	Phenyl-alanine Serine	UAU UAC UAA UAG	Tyrosine Stop codon Stop codon	UGU UGC UGA UGG	Cysteine Stop codon Tryptophan
	C	CUU CUC CUA CUG	Leucine	CCU CCC CCA CCG	Proline	CAU CAC CAA CAG	Histidine Glutamine
	A	AUU AUC AUA AUG	Isoleucine Methionine; start codon	ACU ACC ACA ACG	Threonine	AAU AAC AAA AAG	Asparagine Lysine
	G	GUU GUC GUA GUG	Valine	GCU GCC GCA GCG	Alanine	GAU GAC GAA GAG	Aspartic acid Glutamic acid
						GGU GGC GGA GGG	Glycine
		Third letter					
		U	C	A	G		

Some mutations are synonymous
→ they do not impact the protein sequence

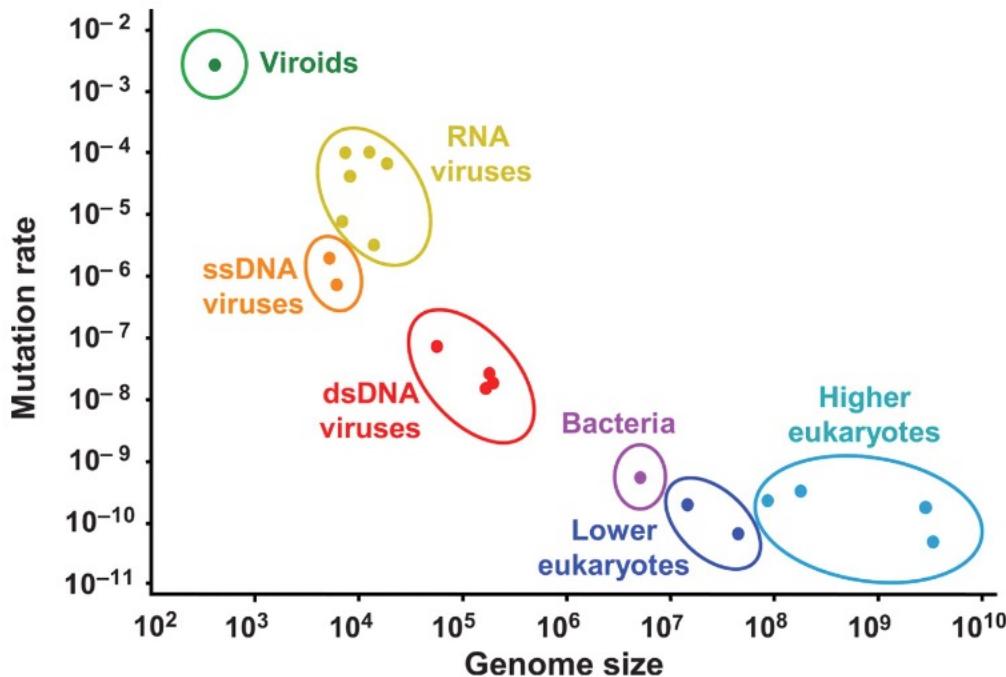
Selection can be assessed by comparing
the rate of non-synonymous and
synonymous mutations
Known as dN/dS (or Ka/Ks or ω)

dN/dS < 1 → selection to stay the same
dN/dS = 1 → no selection
dN/dS > 1 → selection to change

Mutations

▪ How frequent are mutations?

Mutation rates can be measured by the fluctuation test, inspired by the Luria-Delbrück experiment
They can also be measured by sequencing



Mutation probabilities per base pair per replication (substitutions only)

Viruses have high mutation probabilities ($\sim 10^{-5}$)

Bacteria and eukaryotes have lower ones ($\sim 10^{-9}$)
Proofreading and error correction mechanisms allow to reach such low values

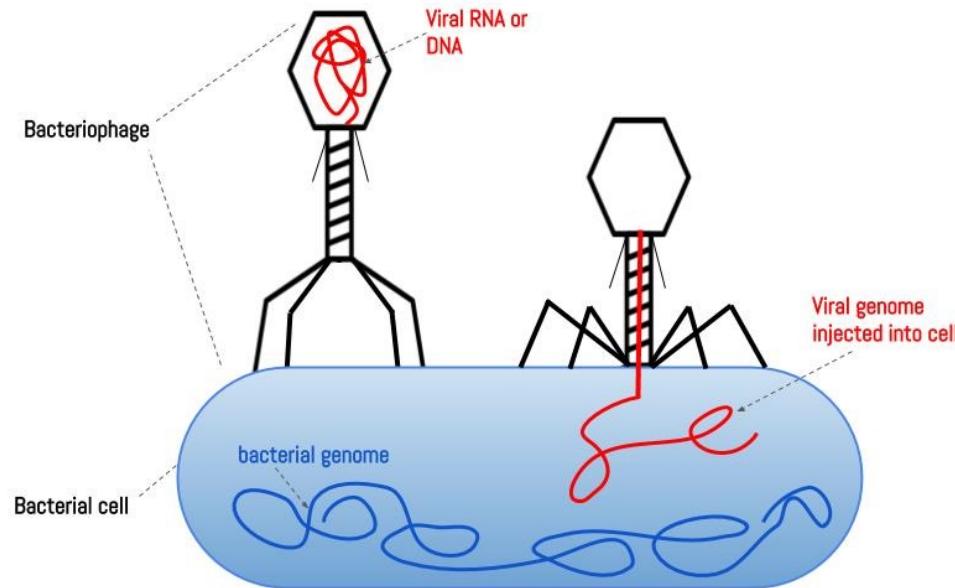
Mutations: Luria-Delbrück experiment

■ Luria-Delbrück experiment (1943)

Phage and bacteria
(phage T1, obligately
lytic virus of *E. coli*)

By random mutations, bacteria can
develop resistance to phage infection

These mutants survive exposure
to phage

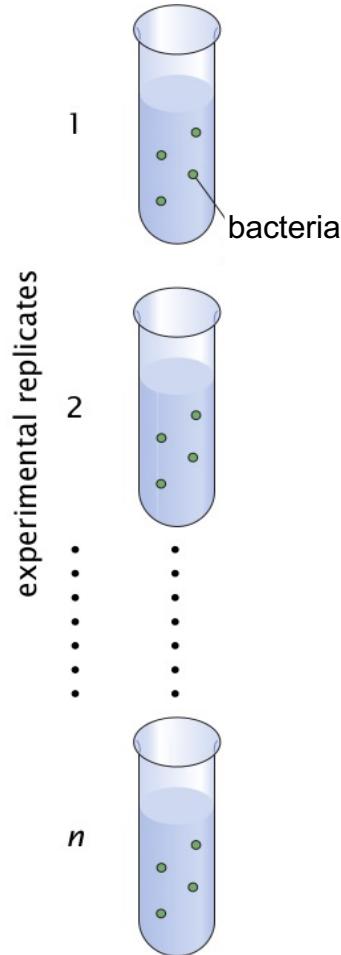


- Counting the bacteria that survive phage and using inference based on the **probability distribution** of the number of phage-resistant bacteria

→ Mutation rate estimate (for mutations giving resistance to phage)

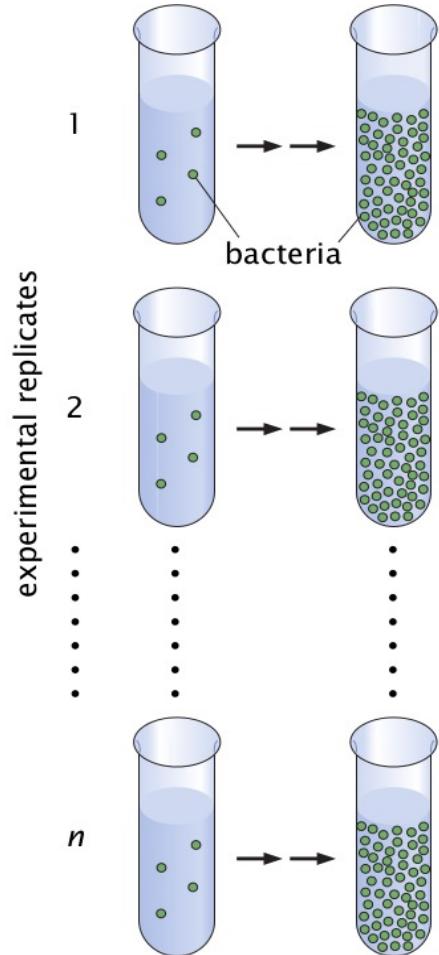
Mutations: Luria-Delbrück experiment

1. Prepare n separate identical cultures of the same bacteria



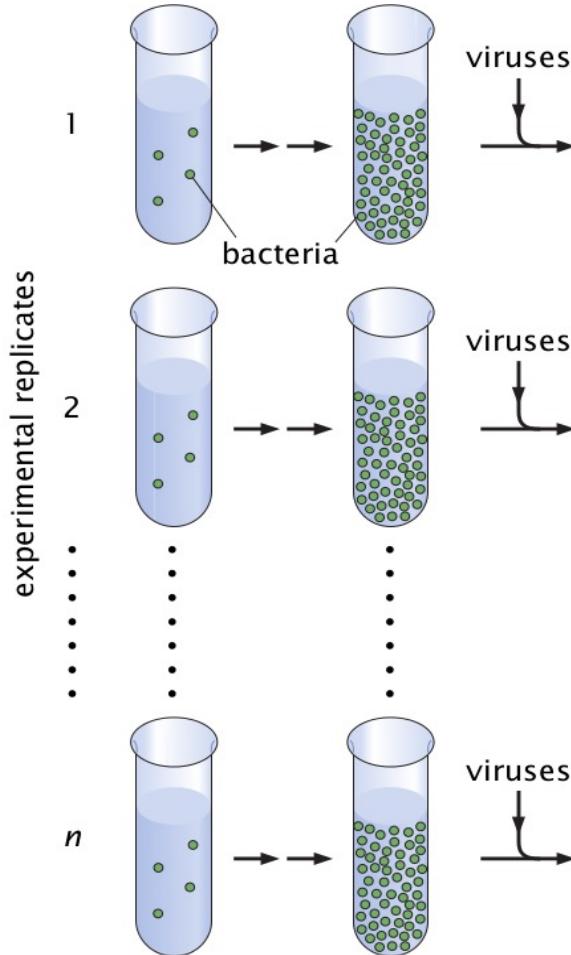
Mutations: Luria-Delbrück experiment

1. Prepare n separate identical cultures of the same bacteria
2. Let them grow



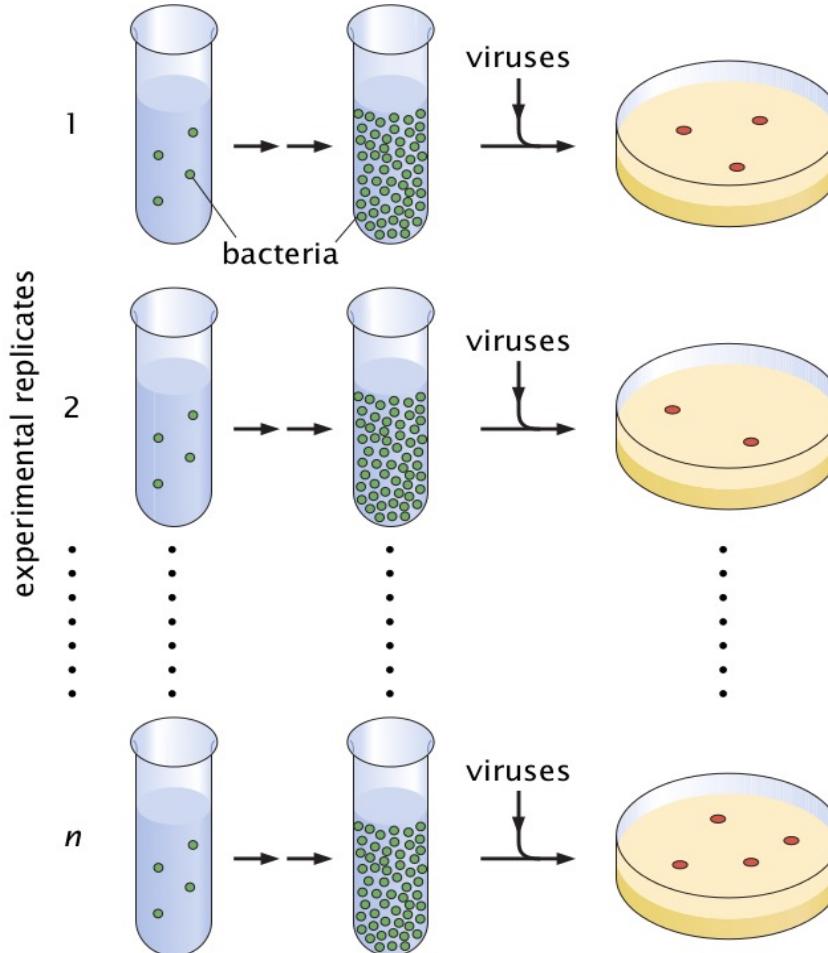
Mutations: Luria-Delbrück experiment

1. Prepare n separate identical cultures of the same bacteria
2. Let them grow
3. Add an excess of bacteriophage viruses (phage T1)
→ most bacteria die; only phage-resistant ones survive



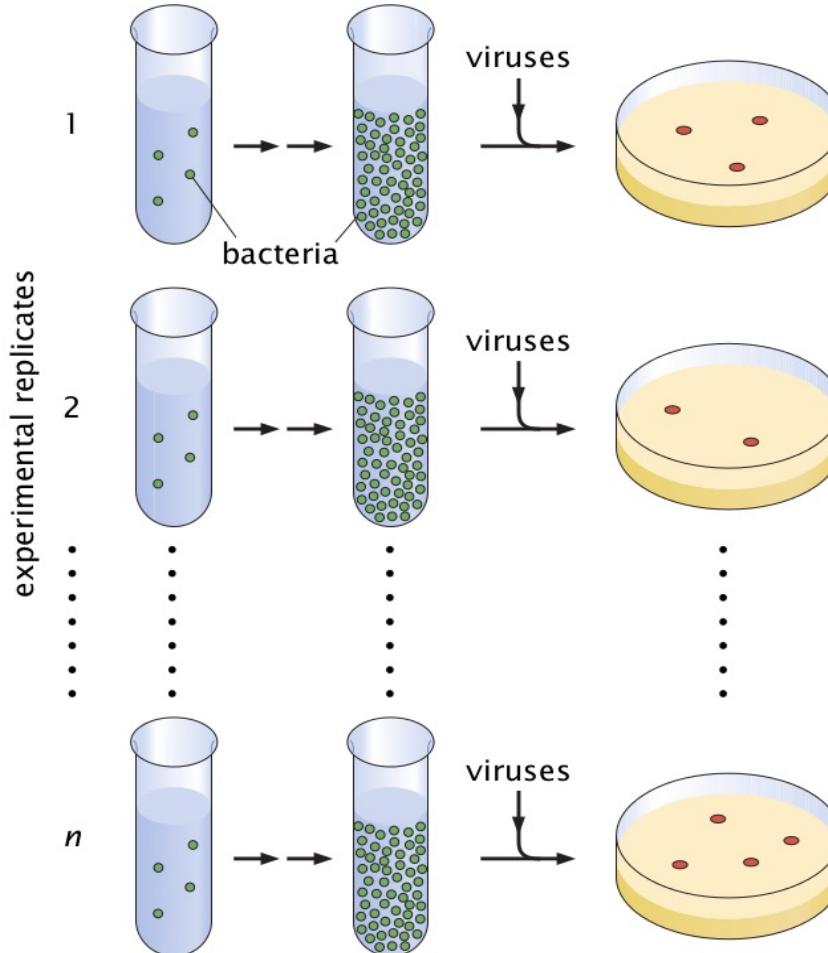
Mutations: Luria-Delbrück experiment

1. Prepare n separate identical cultures of the same bacteria
2. Let them grow
3. Add an excess of bacteriophage viruses (phage T1)
→ most bacteria die; only phage-resistant ones survive
4. To count the survivors, plate each culture separately → each survivor forms a colony



Mutations: Luria-Delbrück experiment

1. Prepare n separate identical cultures of the same bacteria
2. Let them grow
3. Add an excess of bacteriophage viruses (phage T1)
→ most bacteria die; only phage-resistant ones survive
4. To count the survivors, plate each culture separately → each survivor forms a colony
5. Count the number m of colonies growing in each plate → get n values of m



Fitness effects of mutations

- Different measurements – most mutations are deleterious

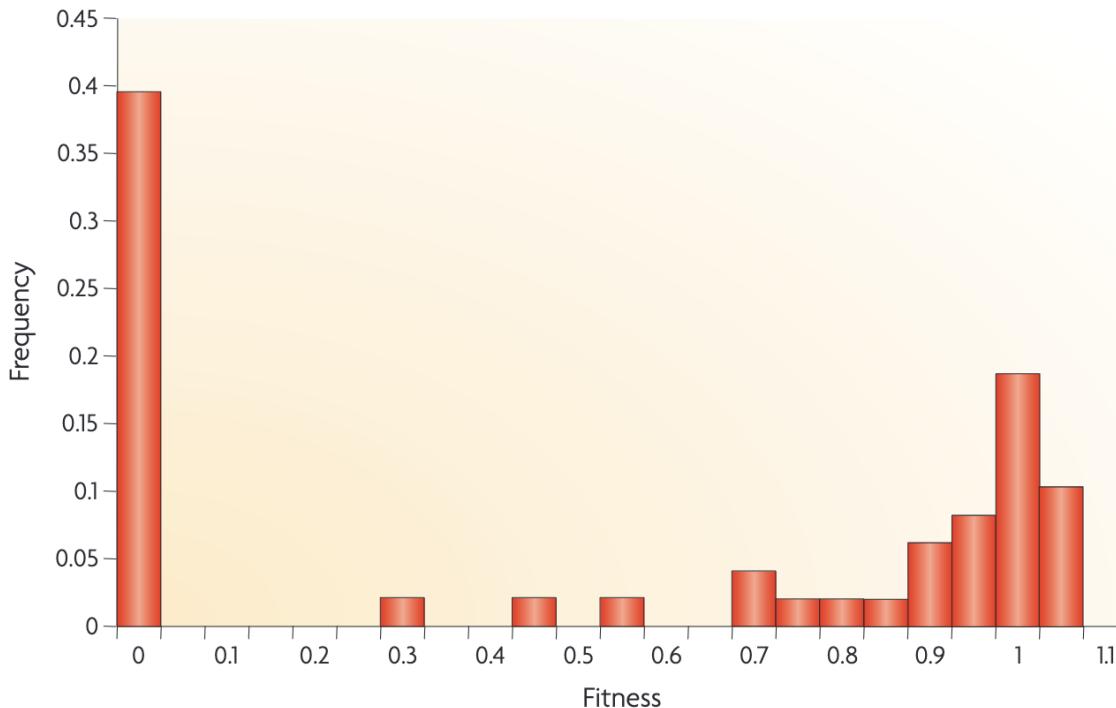


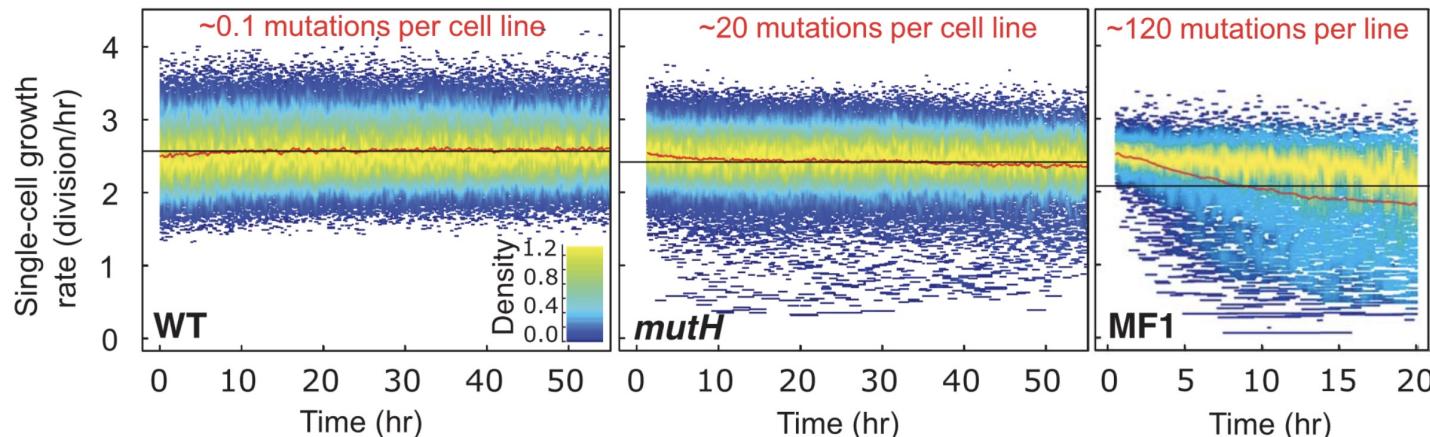
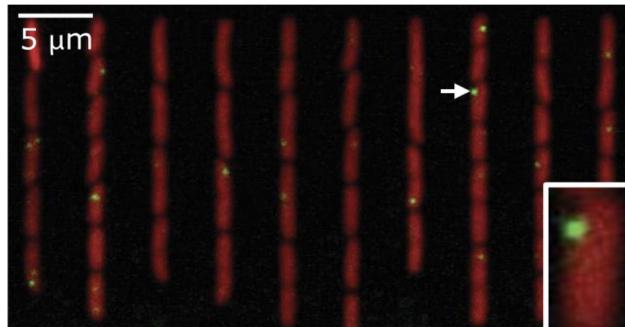
Figure 1 | The distribution of fitness effects of random mutations in vesicular stomatitis virus. In this experiment, random mutations were introduced into the virus, and the fitnesses of the mutants were compared against the unmutated wild type. A fitness of less than one indicates that the mutant was less fit than the wild type, so the mutation was deleterious. A fitness of zero indicates that no mutated progeny were recovered, and that the mutation was therefore lethal. Data from REF. 15.

Sanjuan et al, 2004

Fitness effects of mutations

- Different measurements – most mutations are deleterious

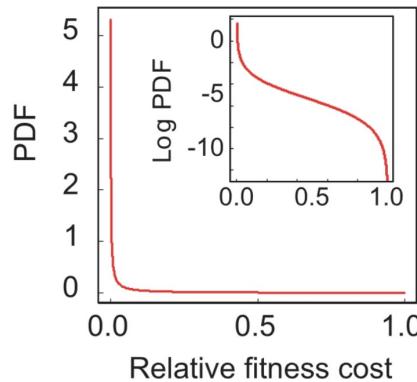
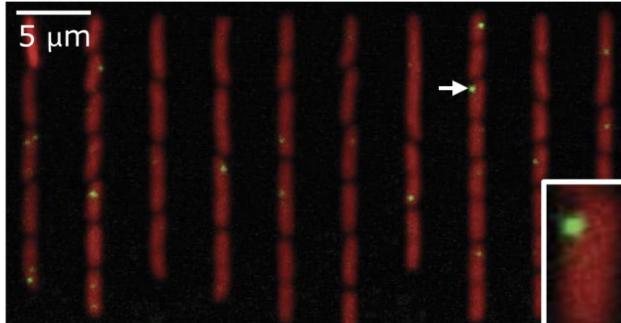
Mutation accumulation in *E. coli* in a microfluidic mother machine – [Robert et al, 2018](#)



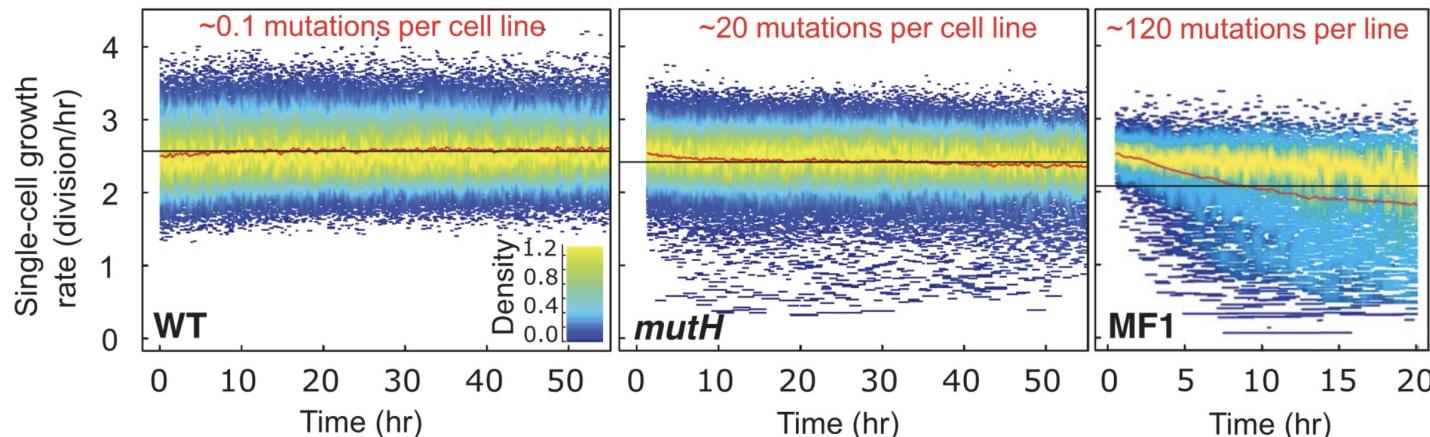
Fitness effects of mutations

- Different measurements – most mutations are deleterious

Mutation accumulation in *E. coli* in a microfluidic mother machine – [Robert et al, 2018](#)



Mean relative fitness cost:
 3.1×10^{-3}
(+ 1% lethal mutations)



Large population: natural selection

Let us focus on the fate of one mutation: does it spread in the population?

▪ Deterministic description for large populations

Consider a population in exponential growth, with 2 types:

$$\begin{cases} \frac{dA}{dt} = (1+s)A, \\ \frac{dB}{dt} = B, \end{cases}$$

which gives $\frac{dx}{dt} = sx(1-x)$

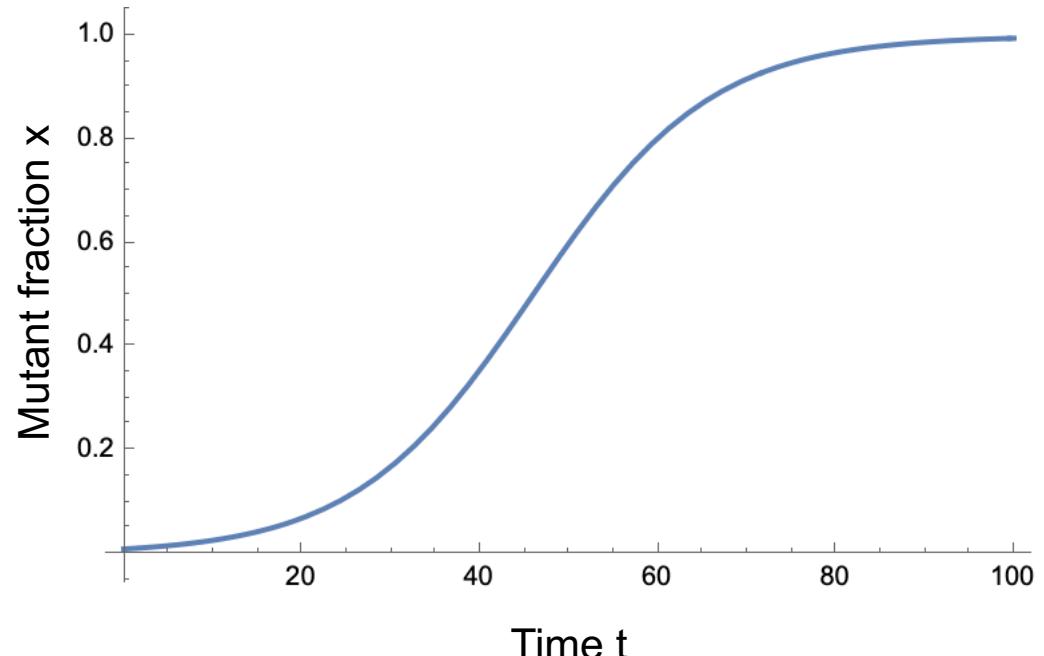
Solution: $x(t) = \frac{x_0 e^{st}}{1 + x_0(e^{st} - 1)}$

If $s > 0$, mutant fraction grows toward a limit of 1 for large t (but does not reach it)

→ Natural selection: fitter type dominates (no coexistence at fixed x)

Example: $x_0=0.01$, $s=0.1$

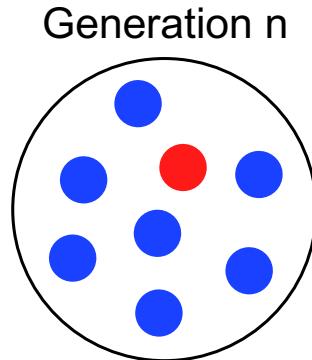
- B wild-types, division rate 1 (sets time unit)
- A mutants, division rate 1+s



Finite population: genetic drift

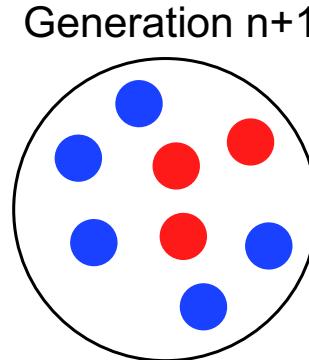
What is the fate of a new mutation appearing in a finite-size population of haploid and asexual microorganisms (e.g. bacteria)?
First consider neutral mutants (no natural selection)

▪ Population with finite and constant size N : Wright-Fisher model



$$k_n=1$$
$$x_n=k_n/N=1/8$$

Binomial sampling



$$k_{n+1}=3$$
$$x_{n+1}=k_{n+1}/N=3/8$$

Non-overlapping generations
 k_n : number of mutants in generation n

Next generation formed by binomial sampling

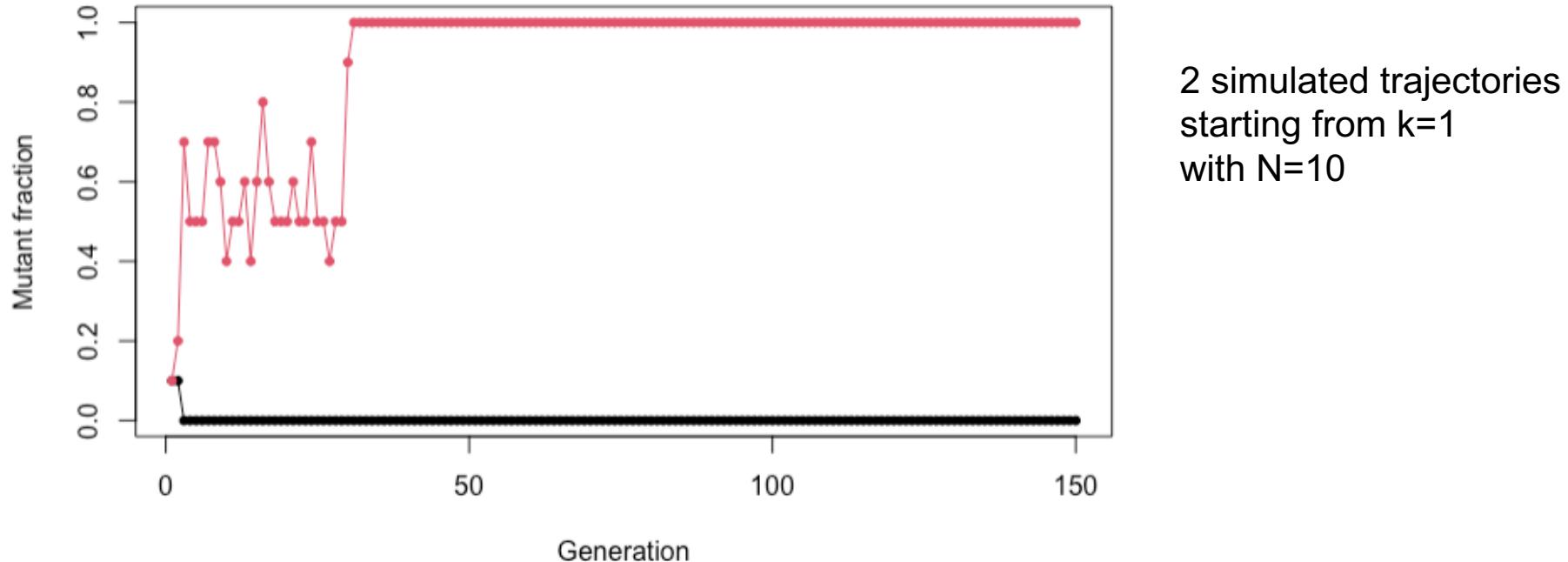
$$P(k_{n+1}) = \binom{N}{k_{n+1}} (x_n)^{k_{n+1}} (1 - x_n)^{N - k_{n+1}}$$

Remark: other model: Moran model (one individual dies and one divides at each time step)

Finite population: genetic drift

- Population with finite and constant size N : Wright-Fisher model

Fraction of mutants over time (generation after generation): random walk



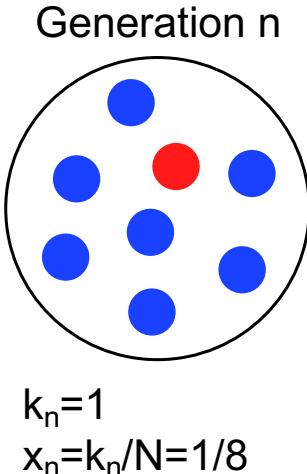
After a large number of generations, mutants either take over (fix) or disappear (go extinct)
This is due to finite-size fluctuations, called genetic drift
Fixation probability starting from 1 mutant: $1/N$, by symmetry

Finite population: genetic drift and selection

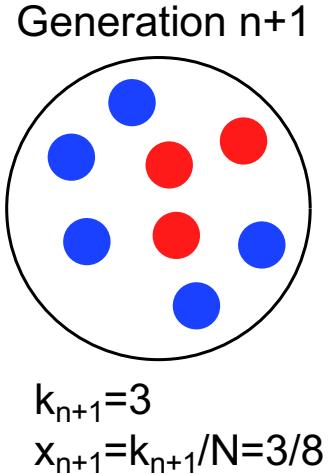
What happens if there is natural selection in a finite-size population?

Assume that mutants are more (or less) likely than wild-types to contribute to the next generation
Encode this in *fitnesses*: 1 for wild-types (reference), 1+s for mutants

Population with finite and constant size N : Wright-Fisher model



Binomial sampling



Non-overlapping generations
 k_n : number of mutants in generation n

Next generation formed by binomial sampling

$$P(k_{n+1}) = \binom{N}{k_{n+1}} (x'_n)^{k_{n+1}} (1 - x'_n)^{N - k_{n+1}}$$

$$x'_n = \frac{(1+s)x_n}{(1+s)x_n + 1 - x_n} = \frac{(1+s)x_n}{1 + sx_n}$$

What is the probability that a mutant fixes, if it has a selective advantage s?

Finite population: genetic drift and selection

Population with finite and constant size N : Wright-Fisher model

Fixation probability starting from one mutant (1): branching process

Focus on the first sampling step, from initial state (called generation 1) to next generation (generation 2)

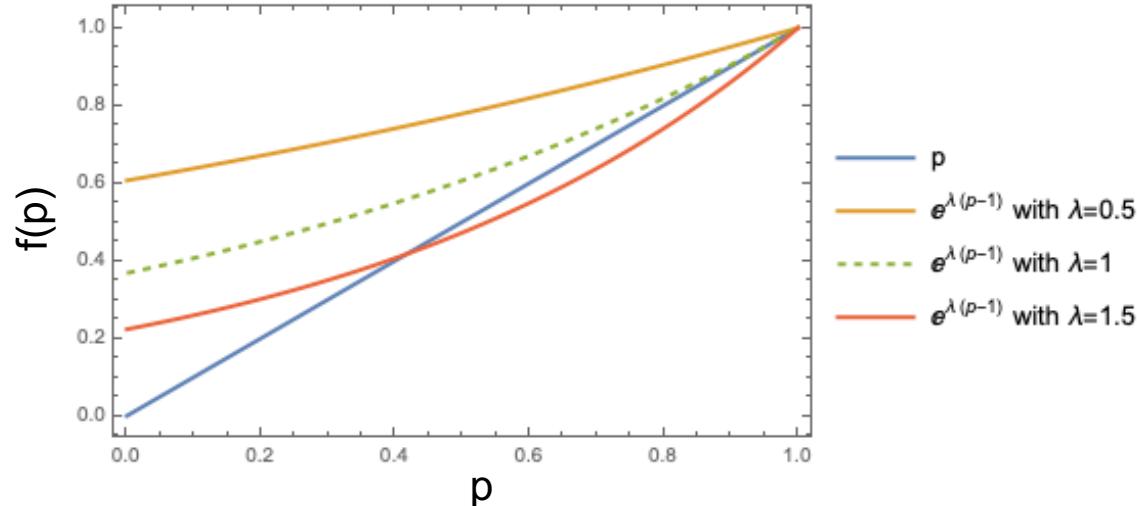
Poisson approximation: $P(k_2) = \binom{N}{k_2} (x'_1)^{k_2} (1 - x'_1)^{N - k_2} \approx \frac{\lambda^{k_2}}{k_2!} e^{-\lambda}$, with $\lambda = Nx'_1$

$N \gg 1$, $x'_1 \ll 1$ and Nx'_1 is of order 1

after growth, starting from $x_1 = 1/N \ll 1$

Assuming that all mutant lineages are independent, the probability of extinction p of the mutant satisfies:

$$p = \exp[\lambda(p - 1)]$$



Finite population: genetic drift and selection

Population with finite and constant size N : Wright-Fisher model

Fixation probability starting from one mutant (1): branching process

Focus on the first sampling step, from initial state (called generation 1) to next generation (generation 2)

Poisson approximation: $P(k_2) = \binom{N}{k_2} (x'_1)^{k_2} (1 - x'_1)^{N - k_2} \approx \frac{\lambda^{k_2}}{k_2!} e^{-\lambda}$, with $\lambda = Nx'_1$

$N \gg 1$, $x'_1 \ll 1$ and Nx'_1 is of order 1

after growth, starting from $x_1 = 1/N \ll 1$

Assuming that all mutant lineages are independent, the probability of extinction p of the mutant satisfies:

$$p = \exp[\lambda(p - 1)]$$

If $\lambda < 1$ i.e. $s < 0$, or if $\lambda = 1$ i.e. $s = 0$, the only solution is $p = 1$: extinction is certain

If $\lambda > 1$ i.e. $s > 0$, another solution exists

Strategy to solve this equation and to obtain p if $s > 0$: perform an expansion for small s

Starting from one mutant: $x_1 = 1/N \ll 1 \rightarrow \lambda = Nx'_1 = N \frac{(1+s)x_1}{1+sx_1} = \frac{1+s}{1+s/N} = 1 + s - \frac{s}{N} + O\left(\frac{s^2}{N^2}\right)$

Assume $|s| \ll 1$ and $N|s| \gg 1 \rightarrow \lambda = 1 + s + o(s^2)$

Then, to first order in $s > 0$, we obtain $p = 1 - 2s$, meaning that the probability of mutant fixation is $1 - p = 2s$ $\ll 1$

Finite population: genetic drift and selection

Population with finite and constant size N : Wright-Fisher model

Fixation probability starting from one mutant (2): diffusion approximation

The branching process makes strong approximations, in particular $N \gg 1$: neglects finite population size
It gives a fixation probability 0 for $s=0$ – but we know that it is actually $1/N$...

Nevertheless, it takes into account the fact that the mutant starts in small numbers

Diffusion approximation: another, more precise, approximation

Assumes $|s| \ll 1$ and $N \gg 1$ but includes term in $1/N$, from binomial sampling variance $\Delta x_{n+1}^2 = \frac{x'_n(1-x'_n)}{N}$

It gives $\rho(1/N) = \frac{1 - e^{-2s}}{1 - e^{-2Ns}}$ for the mutant fixation probability ρ starting from one mutant ($x_1=1/N$)

In particular:

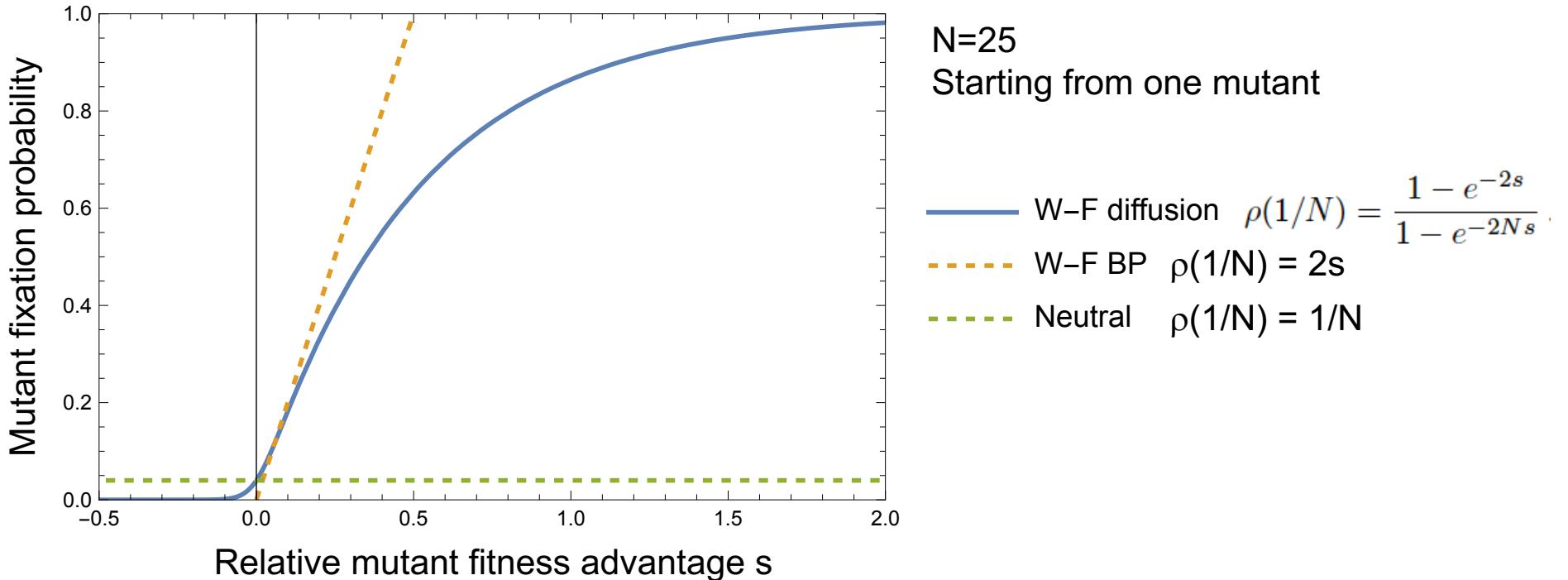
$$|s| \ll 1/N \longrightarrow \rho(1/N) = \frac{1 - (1 - 2s + O(s^2))}{1 - (1 - 2Ns + O(N^2s^2))} \sim \frac{1}{N} : \text{"effectively neutral" regime}$$

$$N \gg 1 \text{ while } s > 0 \text{ but } s \ll 1 \text{ and } Ns \gg 1 \longrightarrow \rho(1/N) = \frac{1 - (1 - 2s + O(s^2))}{1 - e^{-2Ns}} \sim 2s : \text{branching process regime}$$

$$N \gg 1 \text{ while } s < 0 \text{ but } |s| \ll 1 \text{ and } N|s| \gg 1 \longrightarrow \rho(1/N) = \frac{1 - (1 - 2s + O(s^2))}{1 - e^{-2Ns}} \sim -2se^{2Ns} \rightarrow 0 : \text{deleterious regime}$$

Finite population: genetic drift and selection

Population with finite and constant size N : Wright-Fisher model



Important scale: $N |s|$

- if $N |s| \ll 1$, effectively neutral regime
- if $N |s| \gg 1$, selective regime

Finite population: genetic drift and selection

Population with finite and constant size N: Wright-Fisher model

Fixation probability starting from a fraction x of mutants: diffusion approximation

$$\rho(x) = \frac{1 - e^{-2Nsx}}{1 - e^{-2Ns}}$$

Assume $s > 0$ and $Ns \gg 1$. Then, $\rho(x) \approx 1 - e^{-2Nsx}$

becomes small if $Nsx > 1$, i.e. $x > 1/(Ns)$

If a mutant with relative fitness advantage $s \gg 1/N$ reaches a fraction $x > 1/(Ns)$, **it is very likely to fix**

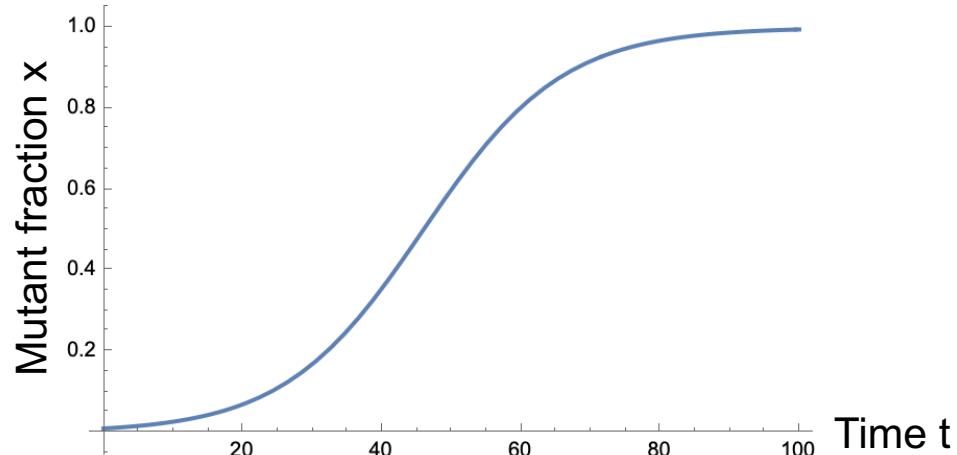
For beneficial mutants, extinctions happen early, when they are in small numbers, due to fluctuations

Fixation timescale: $\sim 1/s$ generations

Reminder: deterministic description:

This is OK if $N \gg 1$
and $s \gg 1/N$
and $x \gg 1/(Ns)$

Large population, sufficient selective advantage and sufficient mutant fraction



Finite population: genetic drift, selection and mutations

So far, we described the fate of one mutation appearing in a population
But other mutations may appear in the meantime: what is their effect?

- Sequential fixation of mutations versus multiple mutations & clonal interference

Mutation probability μ per individual and generation → total mutation probability $N\mu$ per generation

Beneficial mutant with $s >> 1/N$ but $s << 1$:

Probability of fixation $\sim 2s$ → probability $2N\mu s$ per generation to have such a mutation that will then fix
Fixation timescale $\sim 1/s$ generations → if $2N\mu s \ll s$ i.e. $N\mu \ll 1$, no new mutation appears during fixation

Effectively neutral mutant with $|s| << 1/N$:

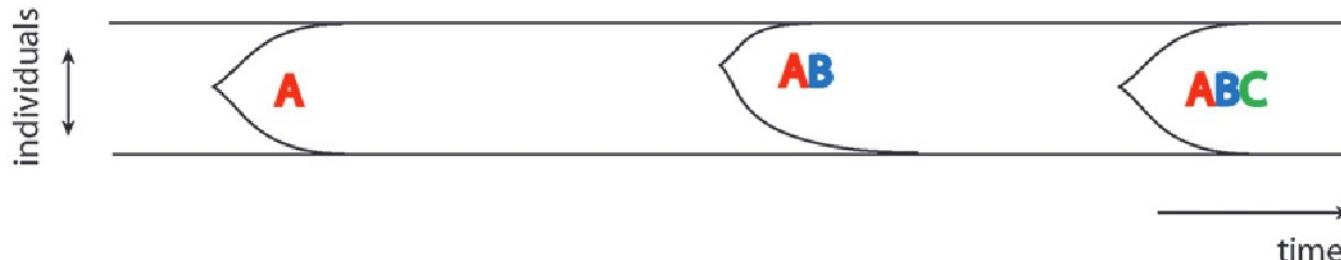
Probability of fixation $\sim 1/N$ → probability μ per generation to have such a mutation that will then fix
Fixation timescale $\sim N$ generations → if $N\mu \ll 1$, no new mutation appears during fixation (same as above)

→ If $N\mu \ll 1$, we can consider that mutations fix successively – it is fine to focus on one at a time
If $N\mu > 1$, new mutant lineages typically appear during the fixation process of a mutant
This is more complex (not described by our Wright-Fisher analysis)

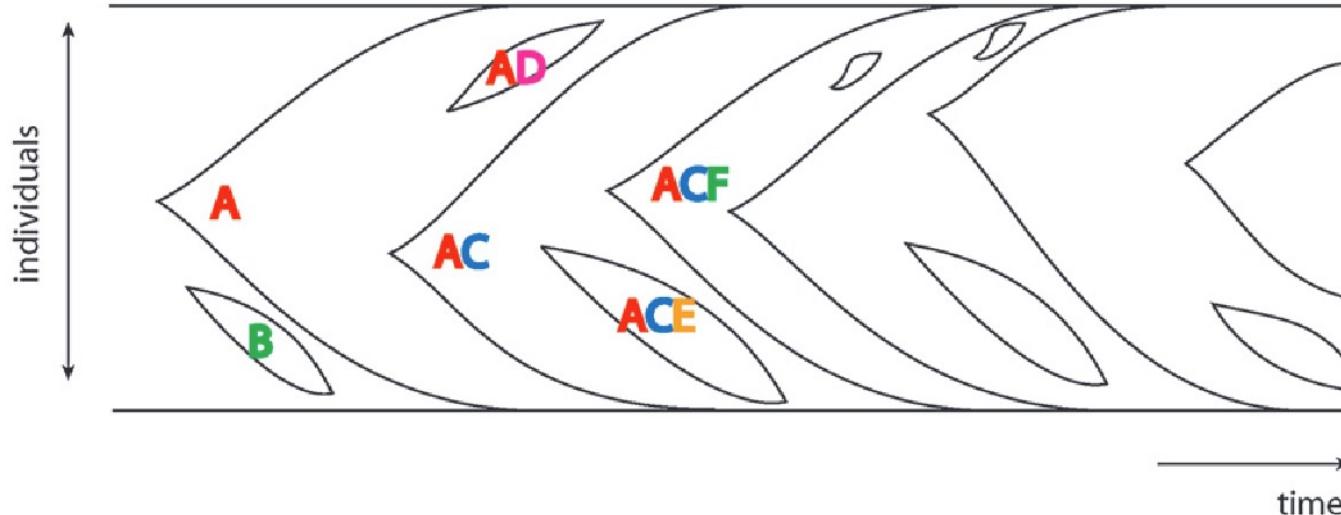
Finite population: genetic drift, selection and mutations

- Sequential fixation of mutations versus multiple mutations & clonal interference

Two different regimes – Desai and Fisher 2007



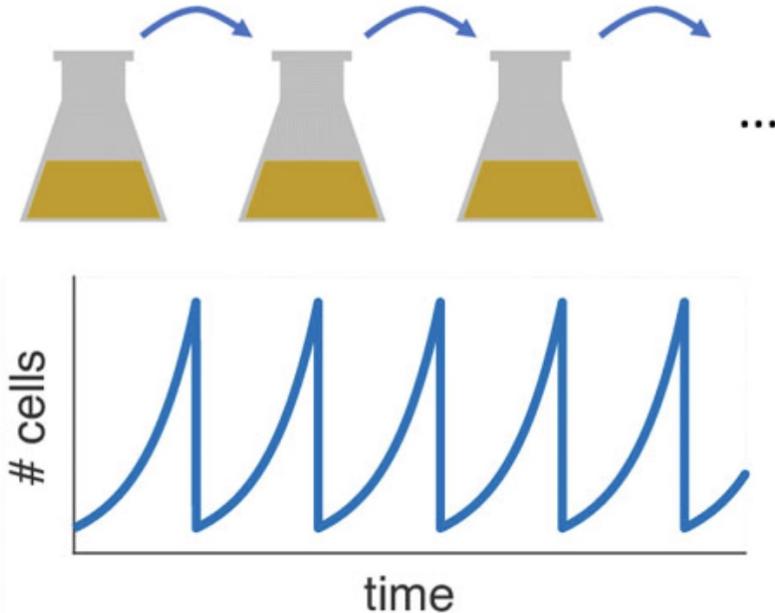
$N\mu < 1$: sequential fixation of mutations



$N\mu > 1$: multiple mutations growing in frequency at the same time
→ “clonal interference”: if two beneficial mutations that would fix if alone appear at similar times, they compete, and only one fixes

Link to evolution experiments

- Experimental protocol: serial transfer (or serial passage, or serial dilution)



1- Cells are placed in culture medium and grow (exponentially – may or may not reach stationary phase)

2- Periodically, a small volume is sampled and placed in new medium – the rest is discarded
→ bottleneck

Phases 1&2 are repeated

Assume that the bottleneck has constant size K

Link to evolution experiments

- Modeling serial transfer (or serial passage, or serial dilution)

1- **Growth phase** → deterministic exponential growth with no death, starting from K cells

Starting from mutant fraction $x_n = k_n/K$ at bottleneck n, the fraction after growth reads $x'_n = \frac{x_n e^{st}}{1 + x_n (e^{st} - 1)}$

Introducing $\sigma = e^{st} - 1$, we can write $x'_n = \frac{(1 + \sigma)x_n}{1 + \sigma x_n}$ → as in the Wright-Fisher model, with σ instead of s

2- **Transfer / bottleneck** → binomial sampling of K individuals from the grown population

Binomial sampling: $P(k_{n+1}) = \binom{K}{k_{n+1}} (x'_n)^{k_{n+1}} (1 - x'_n)^{K - k_{n+1}}$ → as in the Wright-Fisher model

where k_{n+1} is the number of mutants at bottleneck n+1

Mutant fixation probability: as in the Wright-Fisher model, the diffusion approximation gives

$$\rho(1/K) = \frac{1 - e^{-2\sigma}}{1 - e^{-2K\sigma}} = \frac{1 - e^{-2st}}{1 - e^{-2Kst}}$$

starting from one mutant at a bottleneck (fraction 1/K)

$K \gg 1, |\sigma| \ll 1$ $K \gg 1, |s|t \ll 1$

Summary: effects at play

■ Different effects so far

- **Mutations:**

- generate diversity
- most have small fitness effects, most are deleterious

- **Natural selection:**

- acts upon random mutations
- because of it, beneficial mutations tend to take over and fix

- **Genetic drift:**

- corresponds to stochastic fluctuations
- arises from finite population size (total population or mutant population)
- means that moderately beneficial mutations often do not fix – new mutations start in a single individual

■ Additional important effects

- Recombination (horizontal gene transfer; sexual reproduction)
- Interactions between mutations – not just additive fitness effects
- Specific interactions between individuals (beyond mere competition) – cooperativity, attacks...
- Spatial population structure, migrations and genetic flow
- Environmental variability