

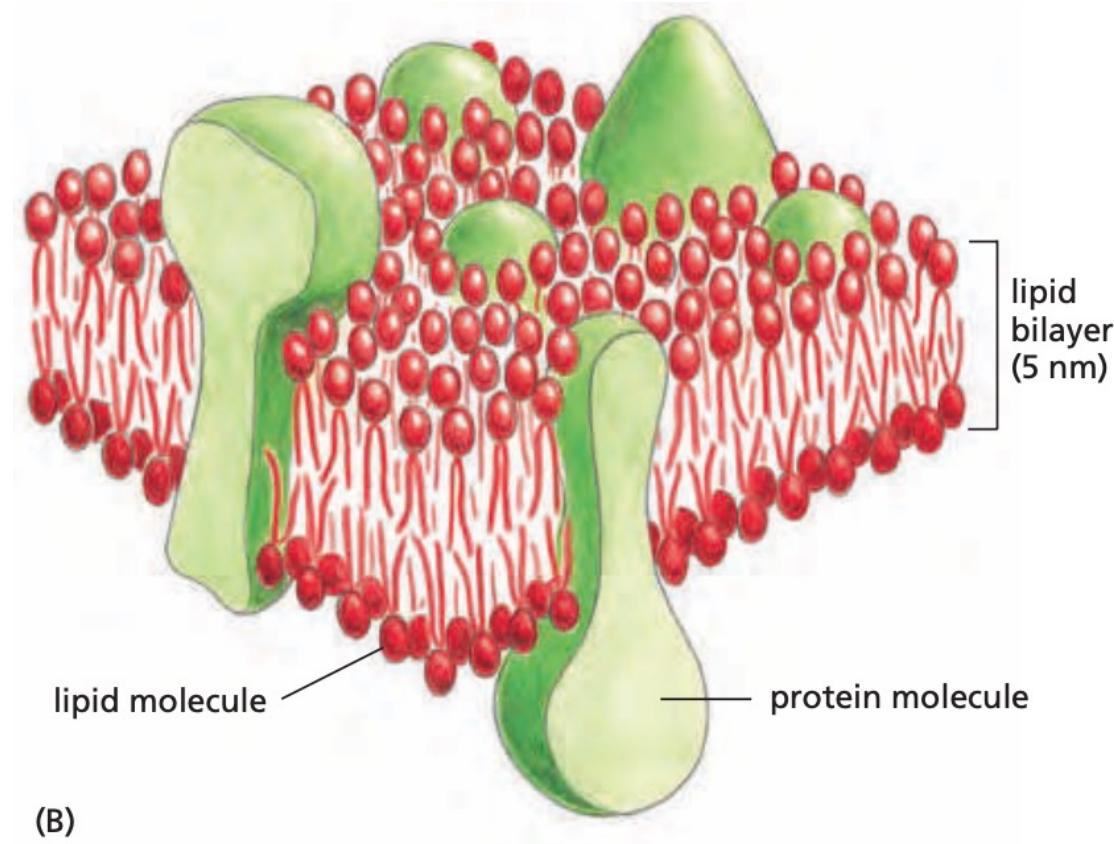
# **Molecular biology of the cell**

## **BIO 207**

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# Last weeks: Discussed the membrane structure



# Topic of the day: membrane transport

Membrane Transport of Small  
Molecules and the Electrical  
Properties of Membranes

CHAPTER  
11

IN THIS CHAPTER

PRINCIPLES OF MEMBRANE  
TRANSPORT

TRANSPORTERS AND ACTIVE  
MEMBRANE TRANSPORT

CHANNELS AND THE  
ELECTRICAL PROPERTIES OF  
MEMBRANES

# Topic of the day: membrane transport

Proteins involved in membrane transport

Active vs Passive transport

ATP-driven pumps and  $\text{Na}^+/\text{K}^+$  ATPase - ABC transporters

Channels

- Aquaporin

Introduction to ion channels

- Ion selectivity

- Ion current measurements

Examples of the importance of these transporters

# ion concentrations inside and outside a cell

**TABLE 11–1** A Comparison of Inorganic Ion Concentrations Inside and Outside a Typical Mammalian Cell\*

Component	Cytoplasmic concentration (mM)	Extracellular concentration (mM)
<b>Cations</b>		
Na <sup>+</sup>	5–15	145
K <sup>+</sup>	140	5
Mg <sup>2+</sup>	0.5	1–2
Ca <sup>2+</sup>	10 <sup>−4</sup>	1–2
H <sup>+</sup>	$7 \times 10^{-5}$ (10 <sup>−7.2</sup> M or pH 7.2)	$4 \times 10^{-5}$ (10 <sup>−7.4</sup> M or pH 7.4)
<b>Anions</b>		
Cl <sup>−</sup>	5–15	110

\*The cell must contain equal quantities of positive and negative charges (that is, it must be electrically neutral). Thus, in addition to Cl<sup>−</sup>, the cell contains many other anions not listed in this table; in fact, most cell constituents are negatively charged (HCO<sub>3</sub><sup>−</sup>, PO<sub>4</sub><sup>3−</sup>, nucleic acids, metabolites carrying phosphate and carboxyl groups, etc.). The concentrations of Ca<sup>2+</sup> and Mg<sup>2+</sup> given are for the free ions: although there is a total of about 20 mM Mg<sup>2+</sup> and 1–2 mM Ca<sup>2+</sup> in cells, both ions are mostly bound to other substances (such as proteins, free nucleotides, RNA, etc.) and, for Ca<sup>2+</sup>, stored within various organelles.

# The lipid bilayer is impermeable

- The lipid bilayer of cell membranes is a barrier that prevents water-soluble molecules and small ions to pass through.
- This allows to have different concentrations of solutes in and out. - Cells use specialized transmembrane proteins for transport

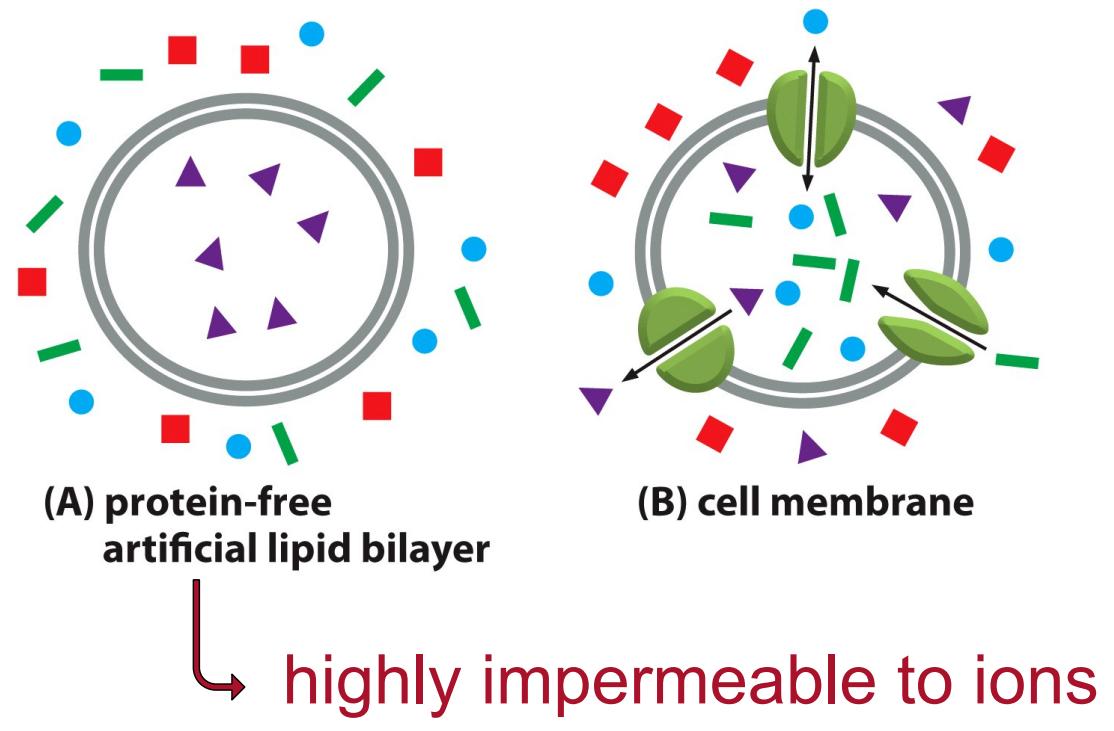
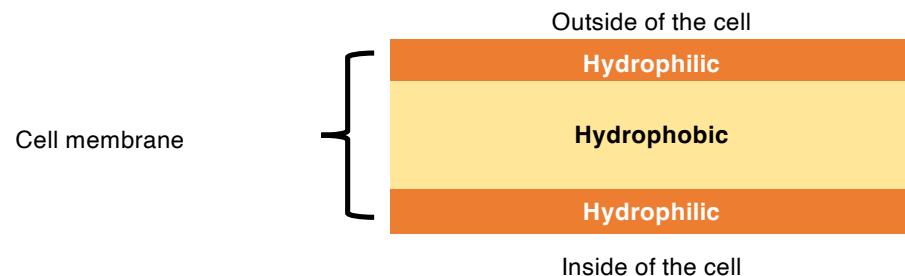
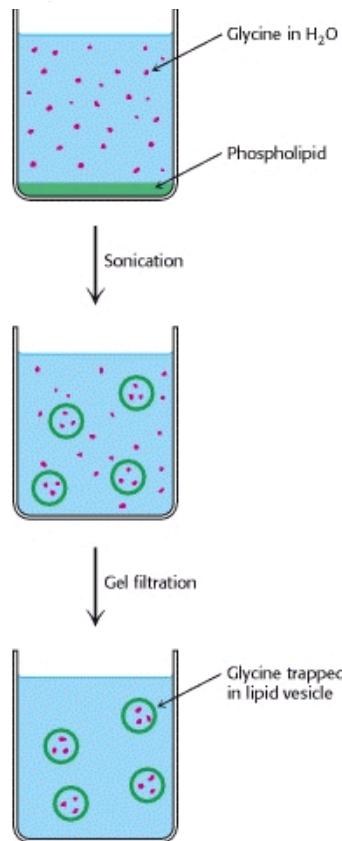


Figure 12-1 *Essential Cell Biology* (© Garland Science 2010)

# Using a synthetic lipid bilayer one can study permeability for different classes of molecules.



## Basic chemical principles for this experiment

Phospholipids form liposomes with an amphiphilic nature

Molecules will diffuse over the lipid bilayer down its concentration gradient

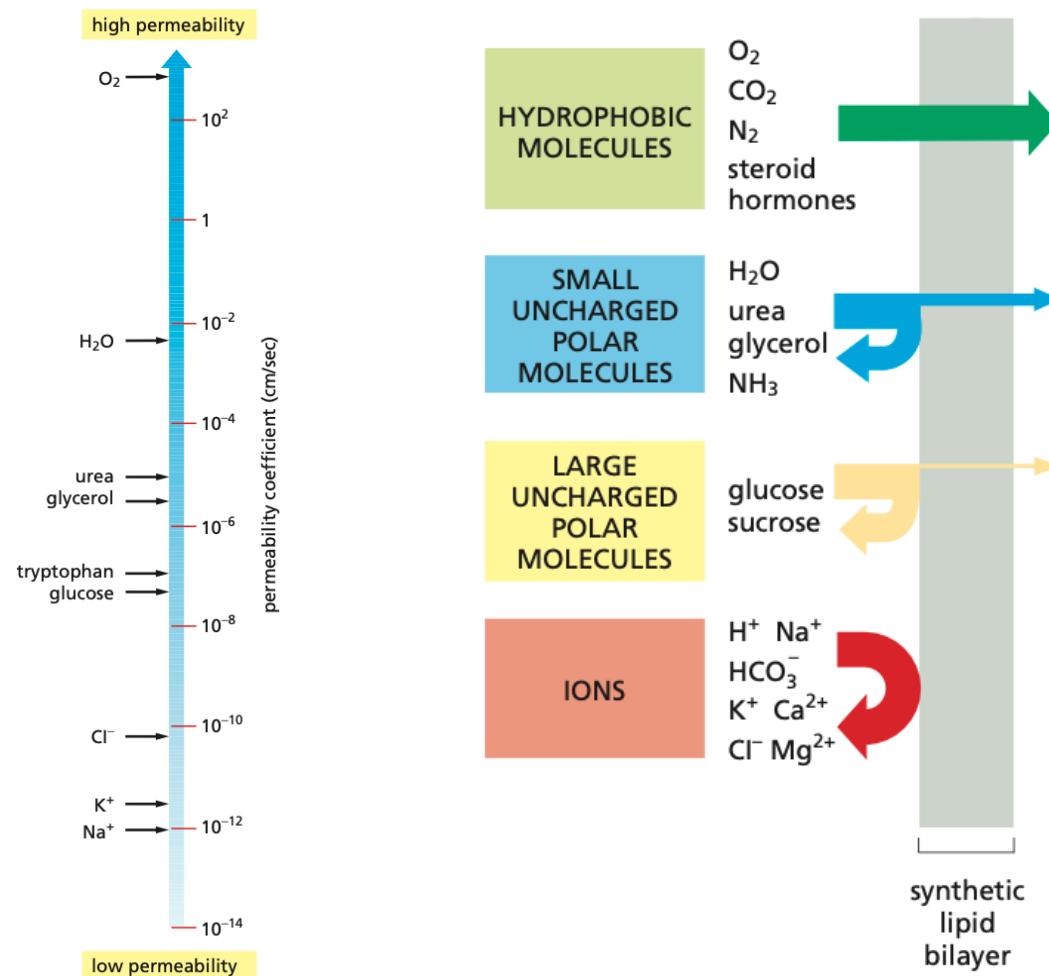
High concentration -> Low concentration

The features of the compound tested will define diffusion rate

Stryer, Biochemistry, 5<sup>th</sup> edition

Figure 12.13 Preparation of Glycine-Containing Liposome

# Using a synthetic lipid bilayer one can study permeability for different classes of molecules.

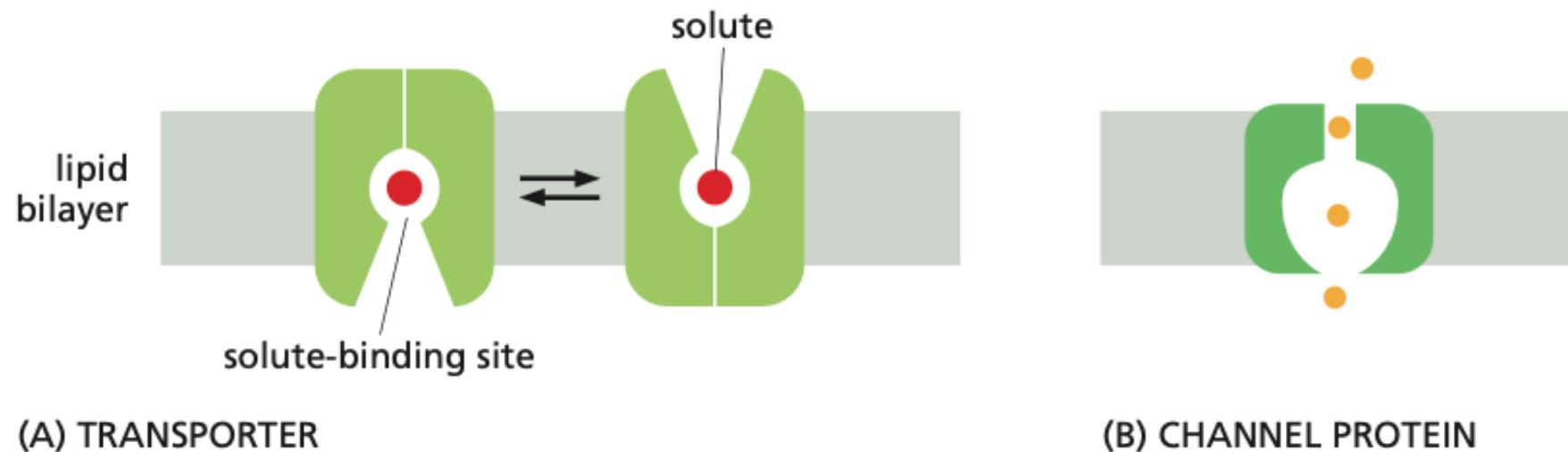


Using this information, we can predict which groups of molecules require membrane transport

**Molecules can be transported across the membrane by proteins**

**What are the types and features of these proteins?**

# Transporters and channel proteins



Transporters are also called carriers or permeases

# Channel proteins

- Form hydrophilic, often highly selective pores
- No change in conformation
- The pores can usually open and close rapidly
- No energy coupled, only passive transport

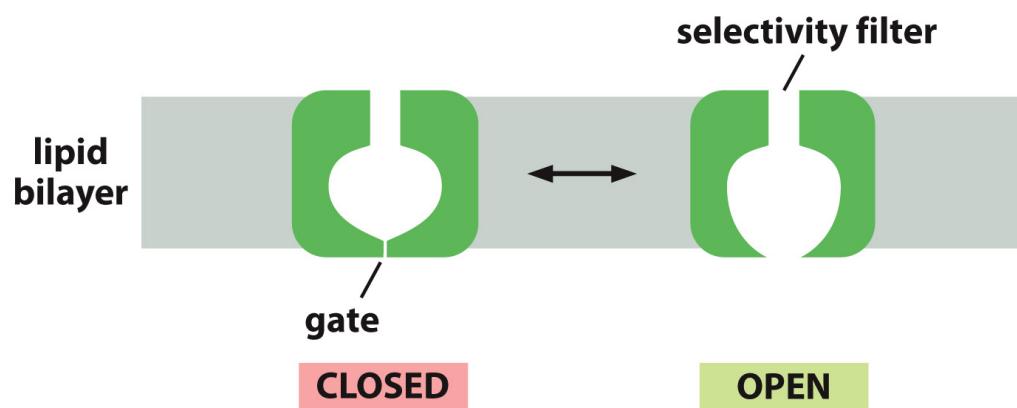


Figure 11-21 Molecular Biology of the Cell 6e (© Garland Science 2015)

# Transporters and channel proteins

	transporter	channel
<b>Interaction with the solute</b>	strong	weak
<b>Conformational change</b>	yes	no
<b>Transport speed</b>	slow	fast
<b>Transport type</b>	active or passive	passive

Transporters are found in most or all biological membranes

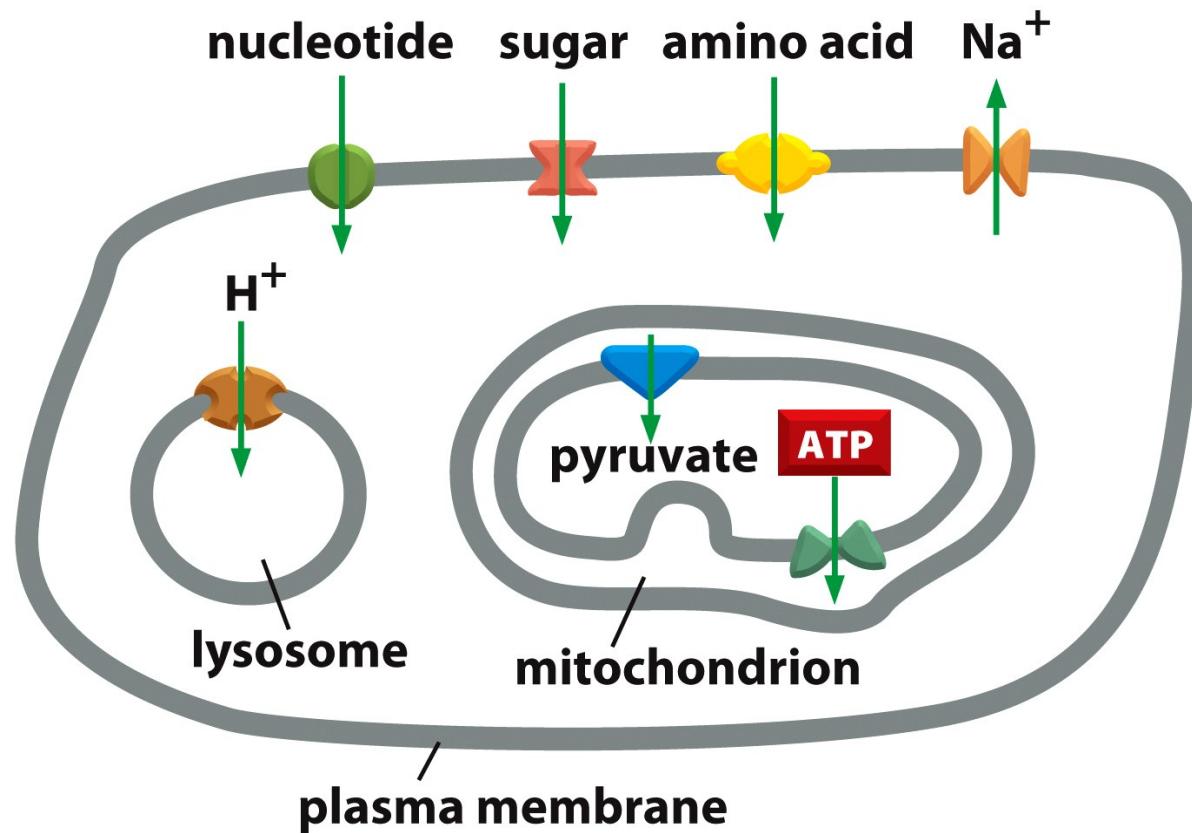
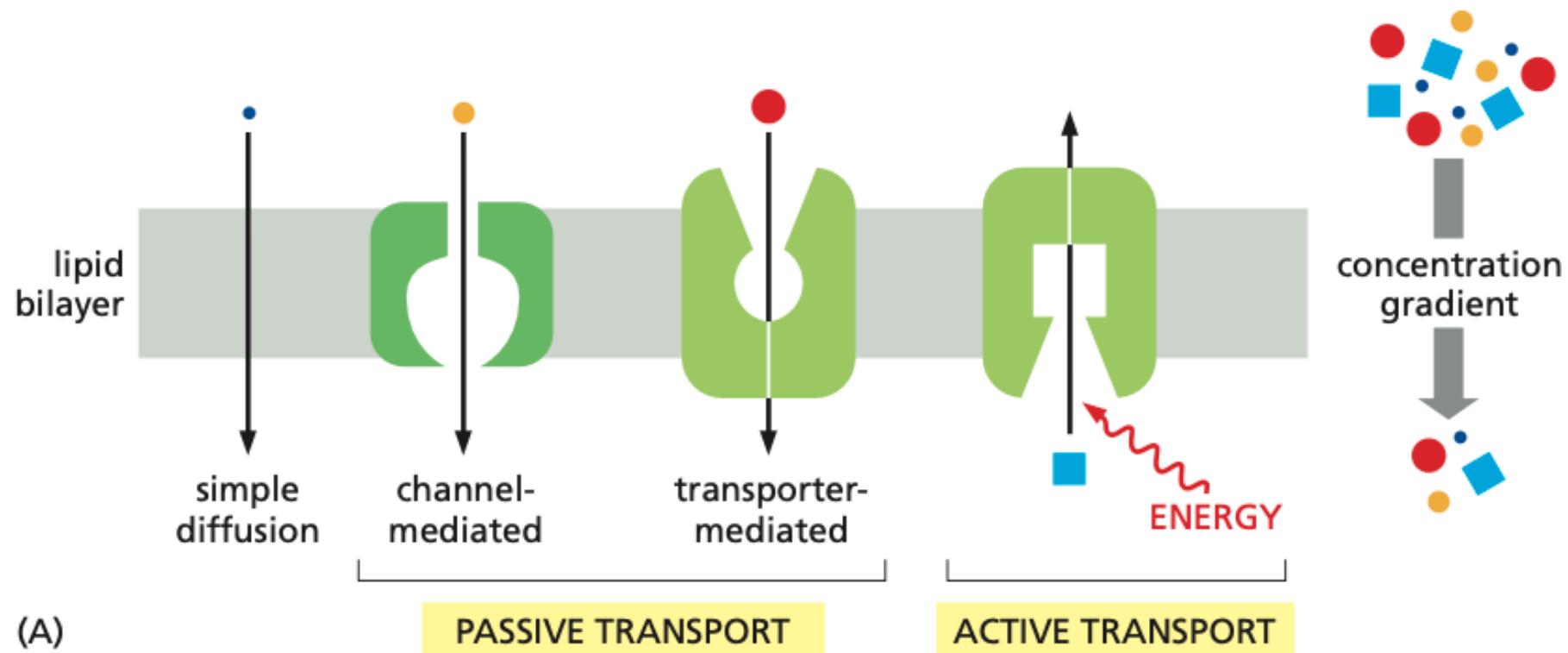


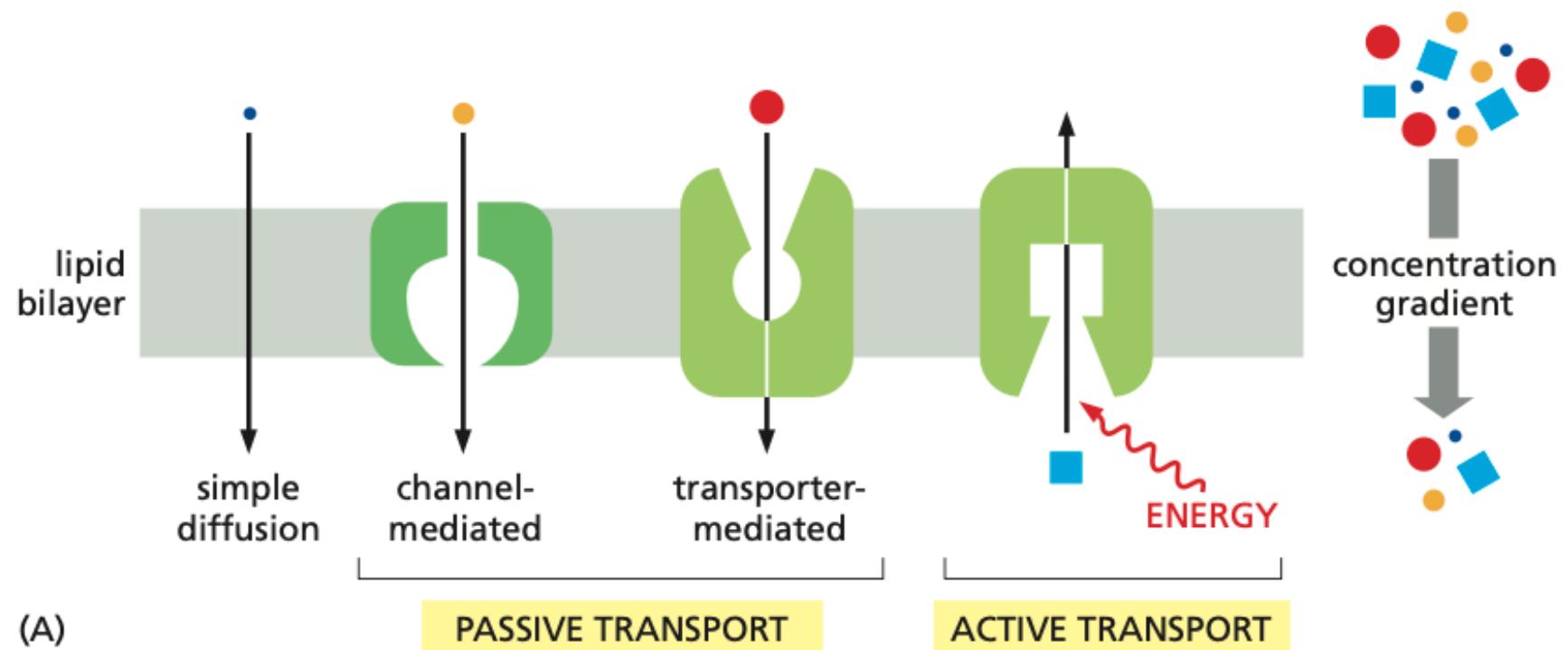
Figure 12-5 *Essential Cell Biology* (© Garland Science 2010)

# Passive versus Active transport



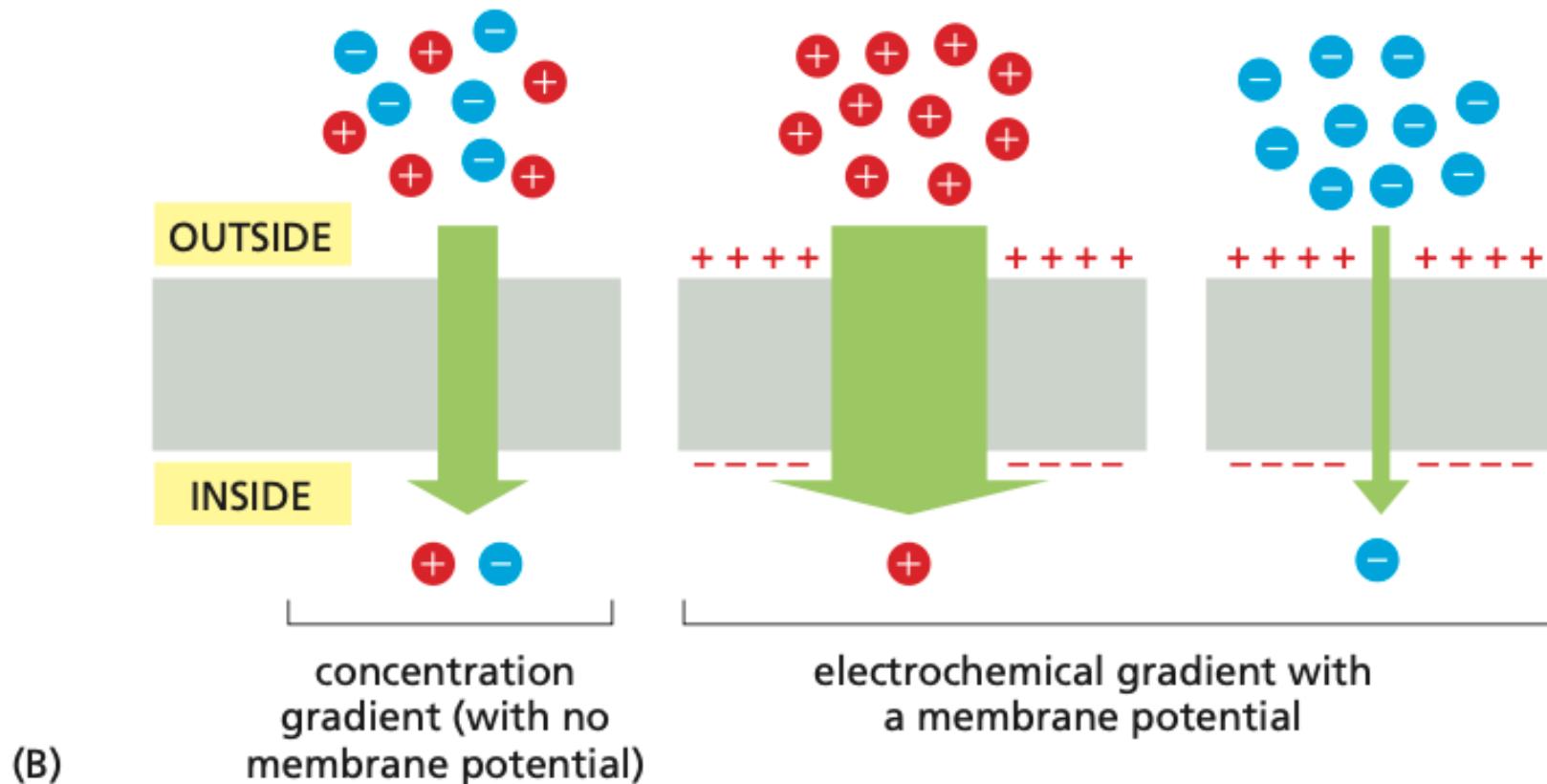
Passive transport relies on concentration gradients

# Passive versus Active transport

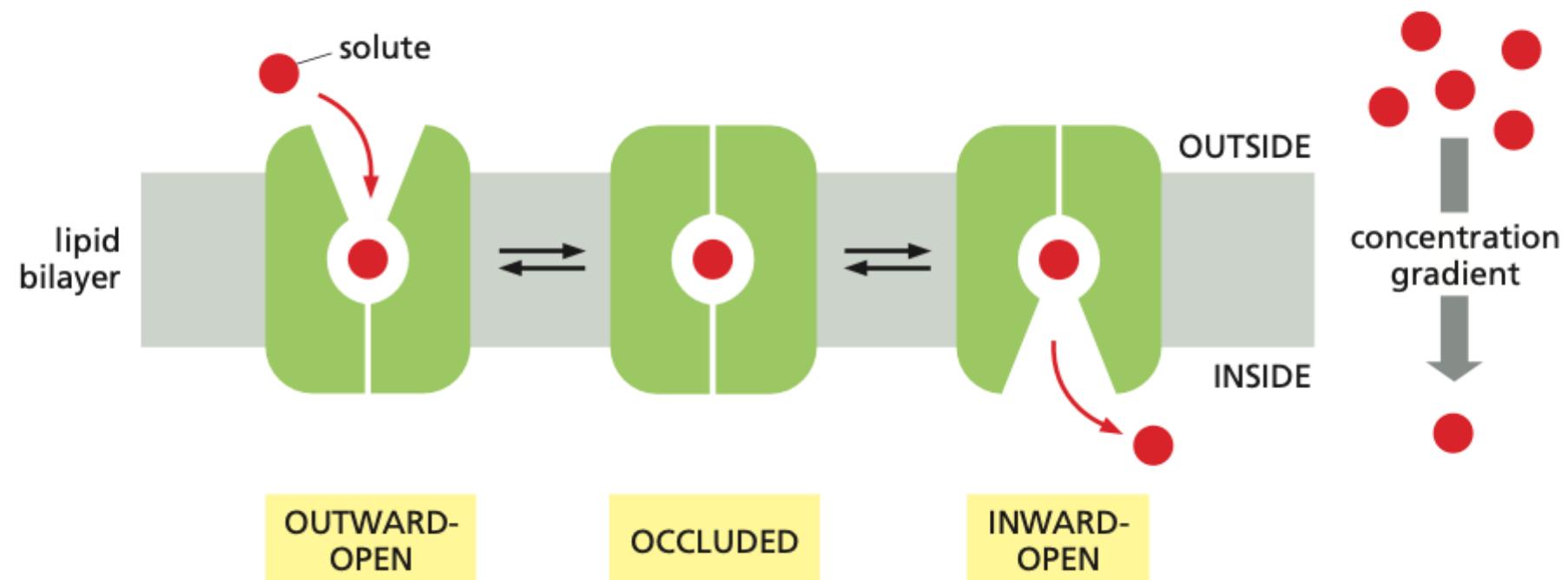


The basic structures and amino acid sequences of passive transporters and active transporters are very similar.

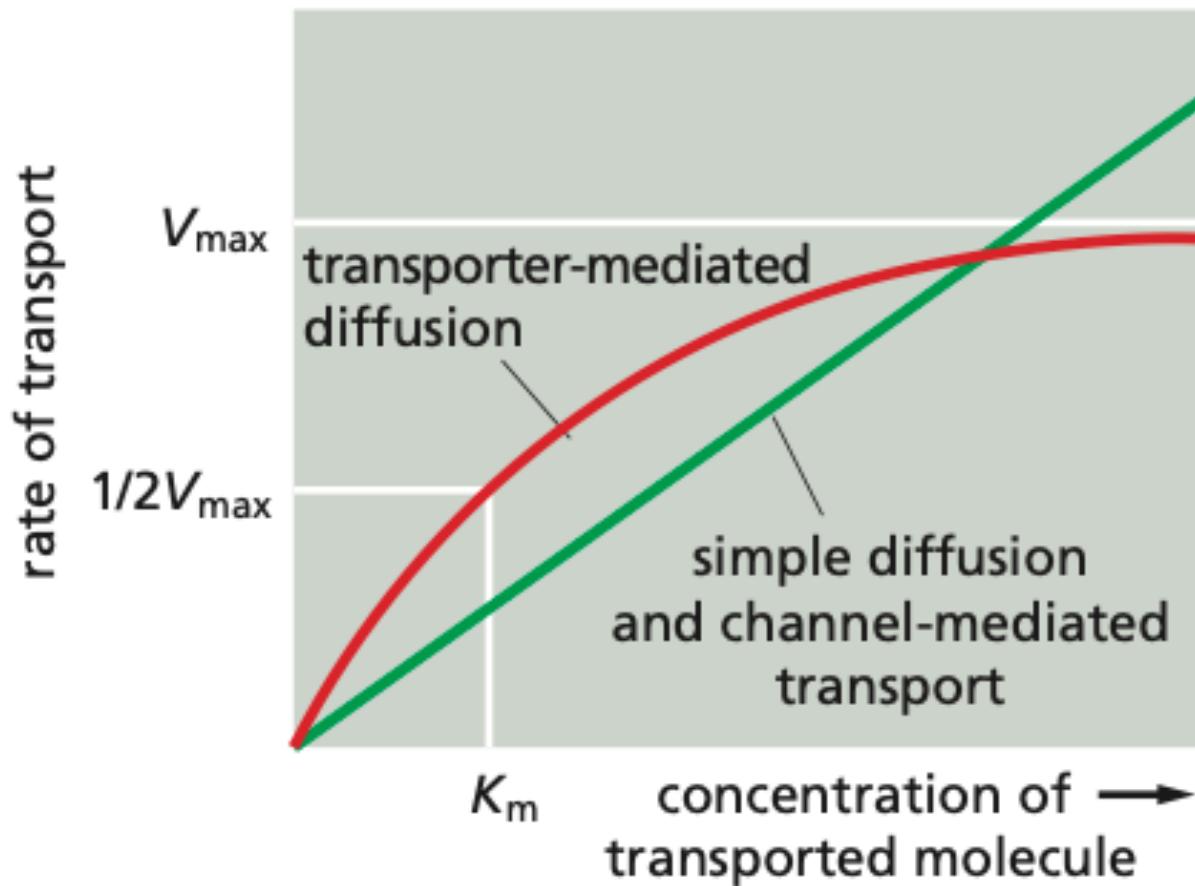
# Charge of the transported molecule influences transport



# A model of how a conformational change in a transporter mediates the passive movement of a solute

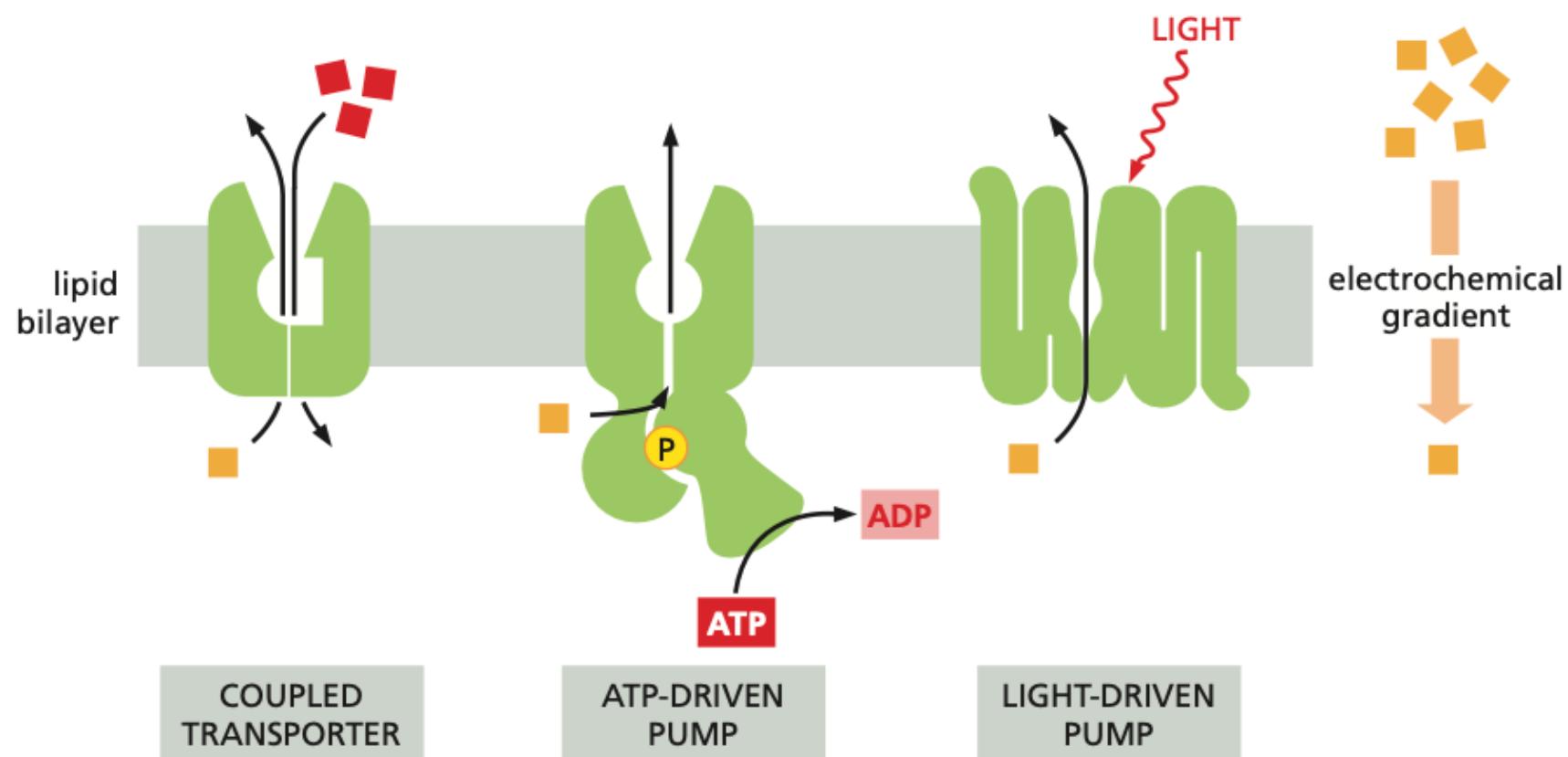


# The kinetics of transporters resemble an enzymatic reaction



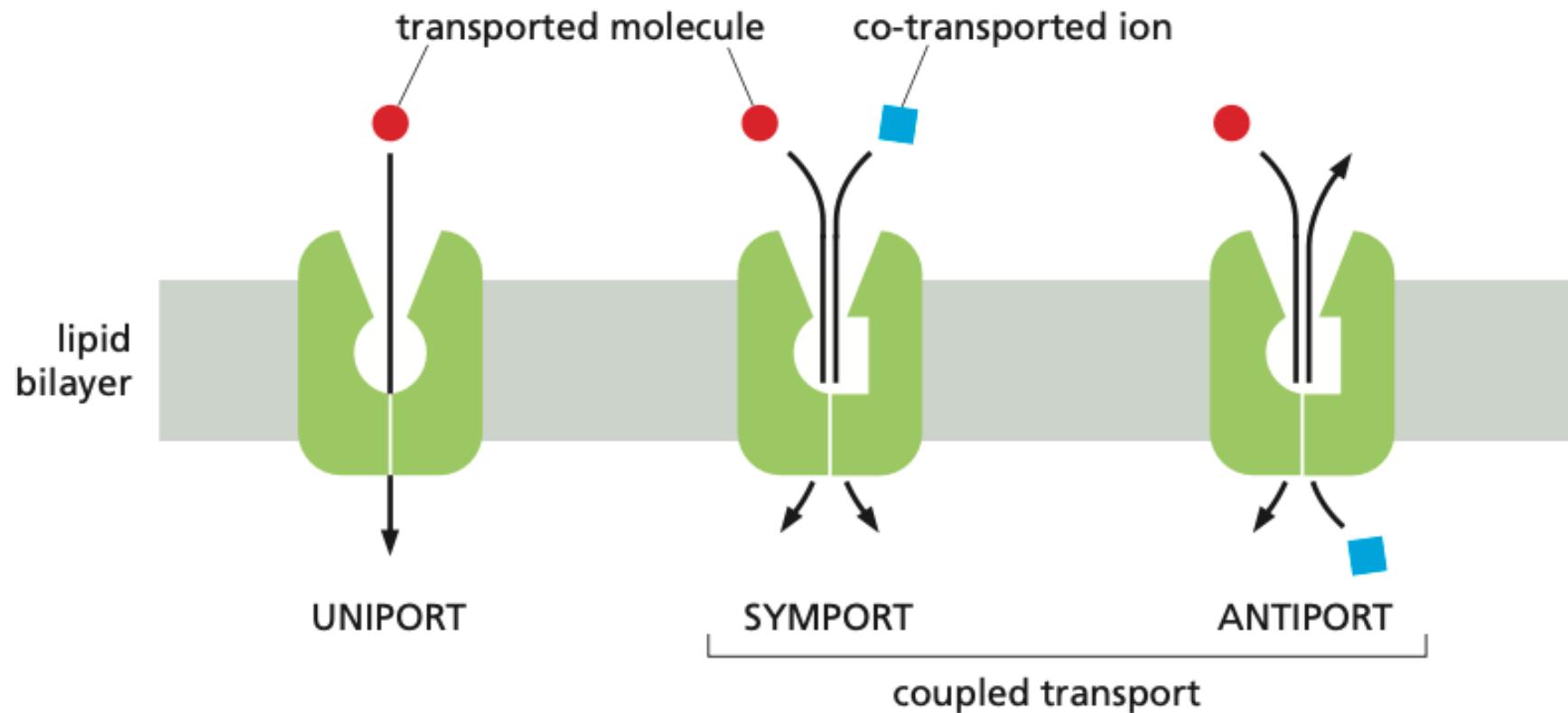
- Each carrier has a defined maximal flipping rate ( $V_{max}$ )
- $V_{max}$  is reached when the transporter is saturated
- The affinity ( $K_m$ ) equals the concentration of solute at  $1/2 V_{max}$

# Three ways of driving active transport

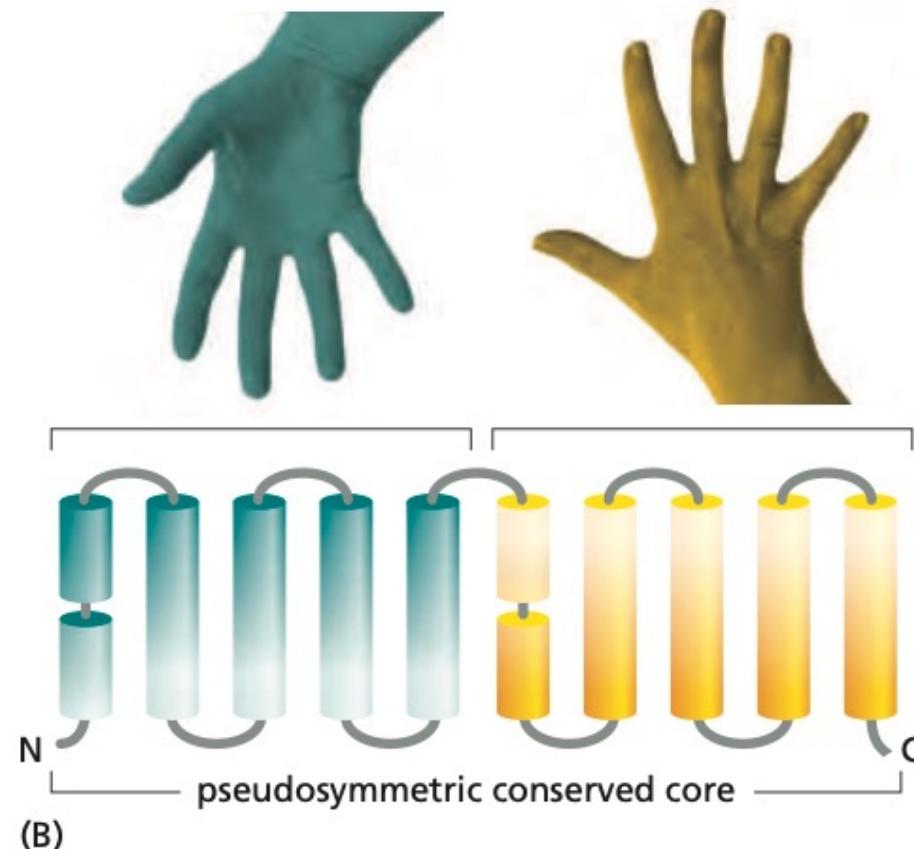
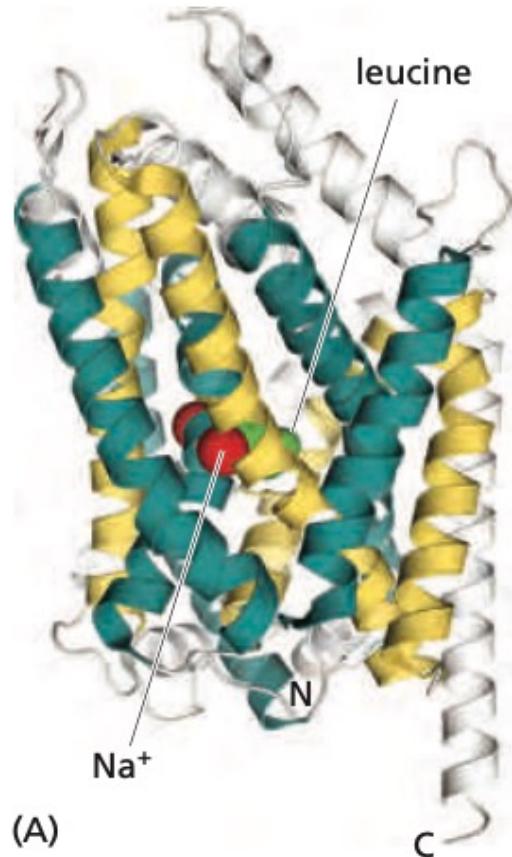


Active transport uses energy to drive transport against the electrochemical current

# Different modes of coupled transport. uniporters, symporters, and antiporters

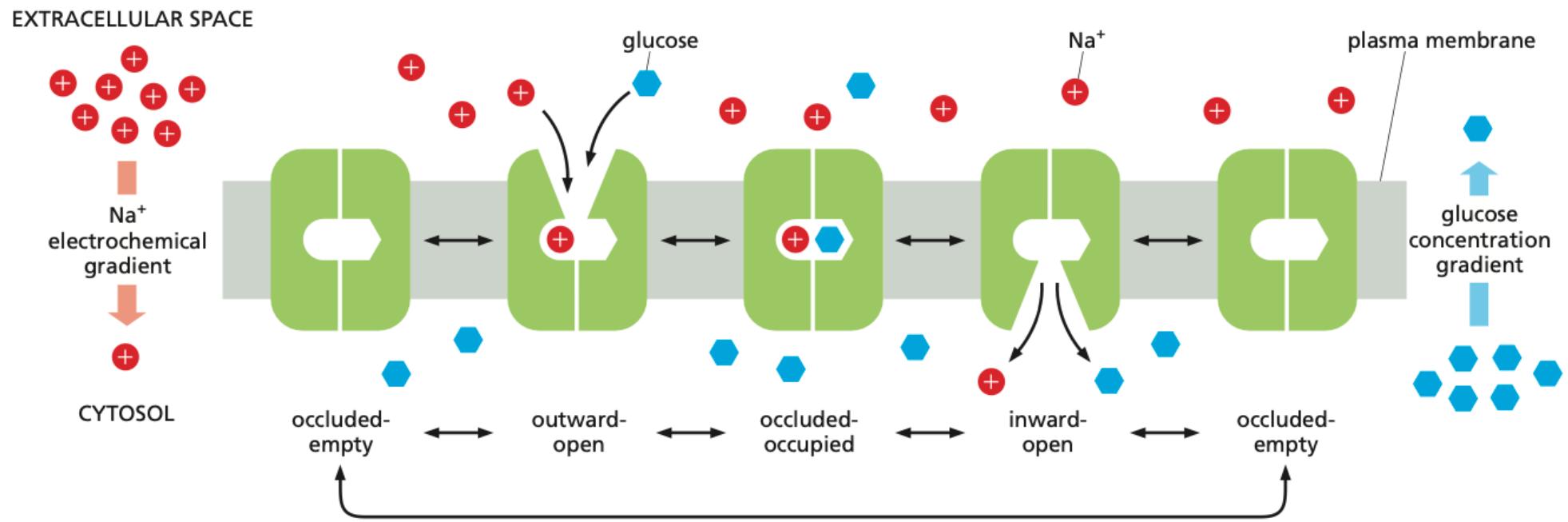


# Transporters are built from inverted repeats



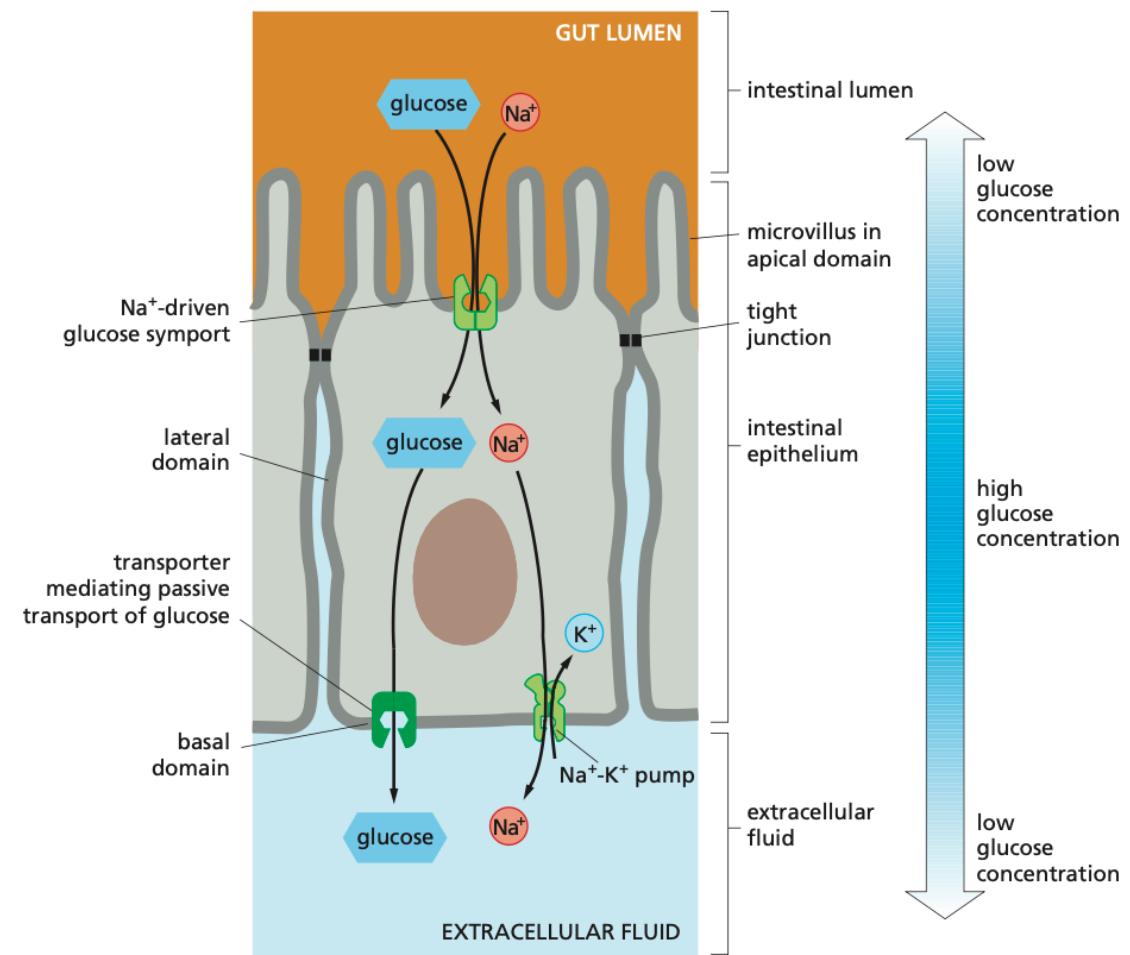
# **Active coupled transport driven by a ion concentration**

# Mechanism of glucose transport fuelled by a $\text{Na}^+$ gradient.



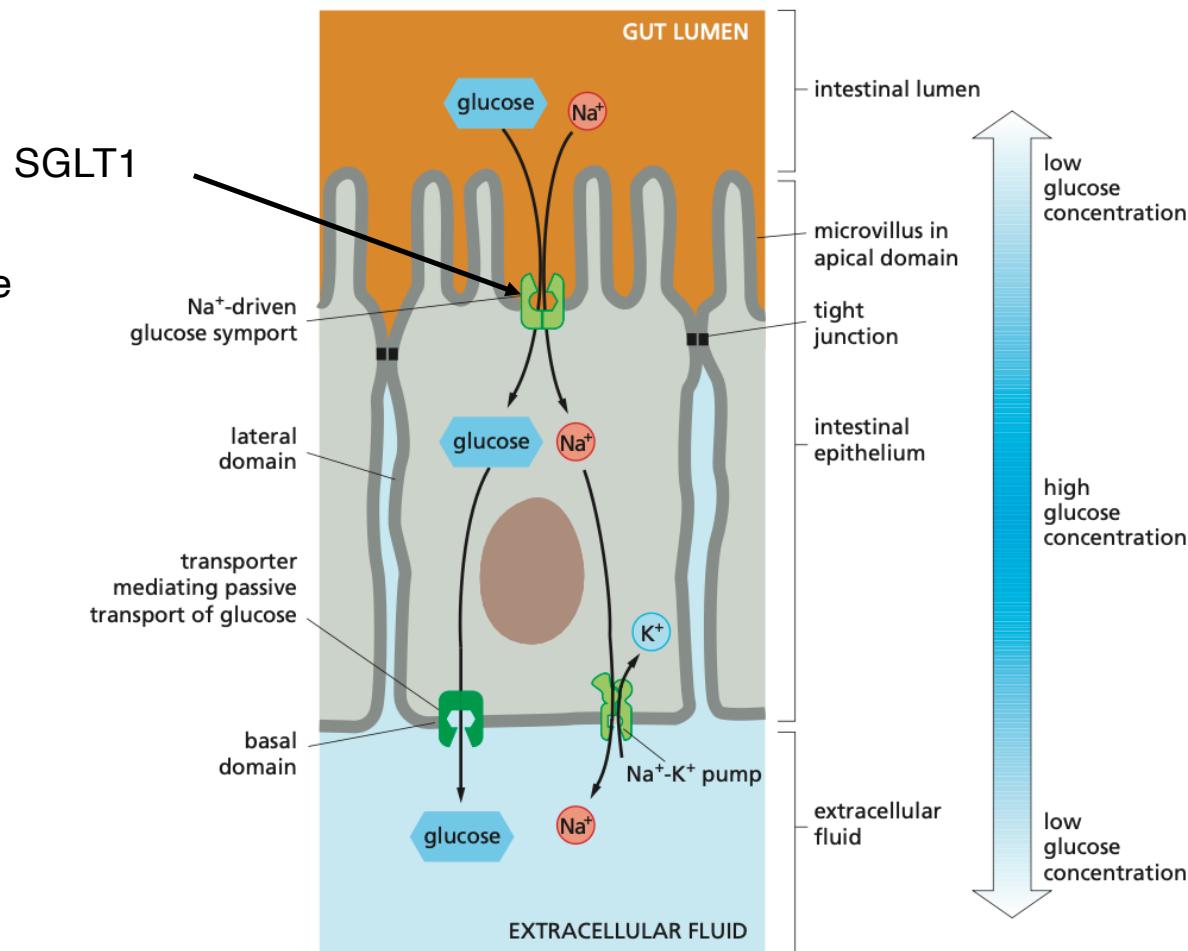
These glucose transporters are called the sodium glucose linked transport (SGLT) family of transporters. There are several with SGLT1 and SGLT2 the most well known

# Glucose absorption in the small intestine

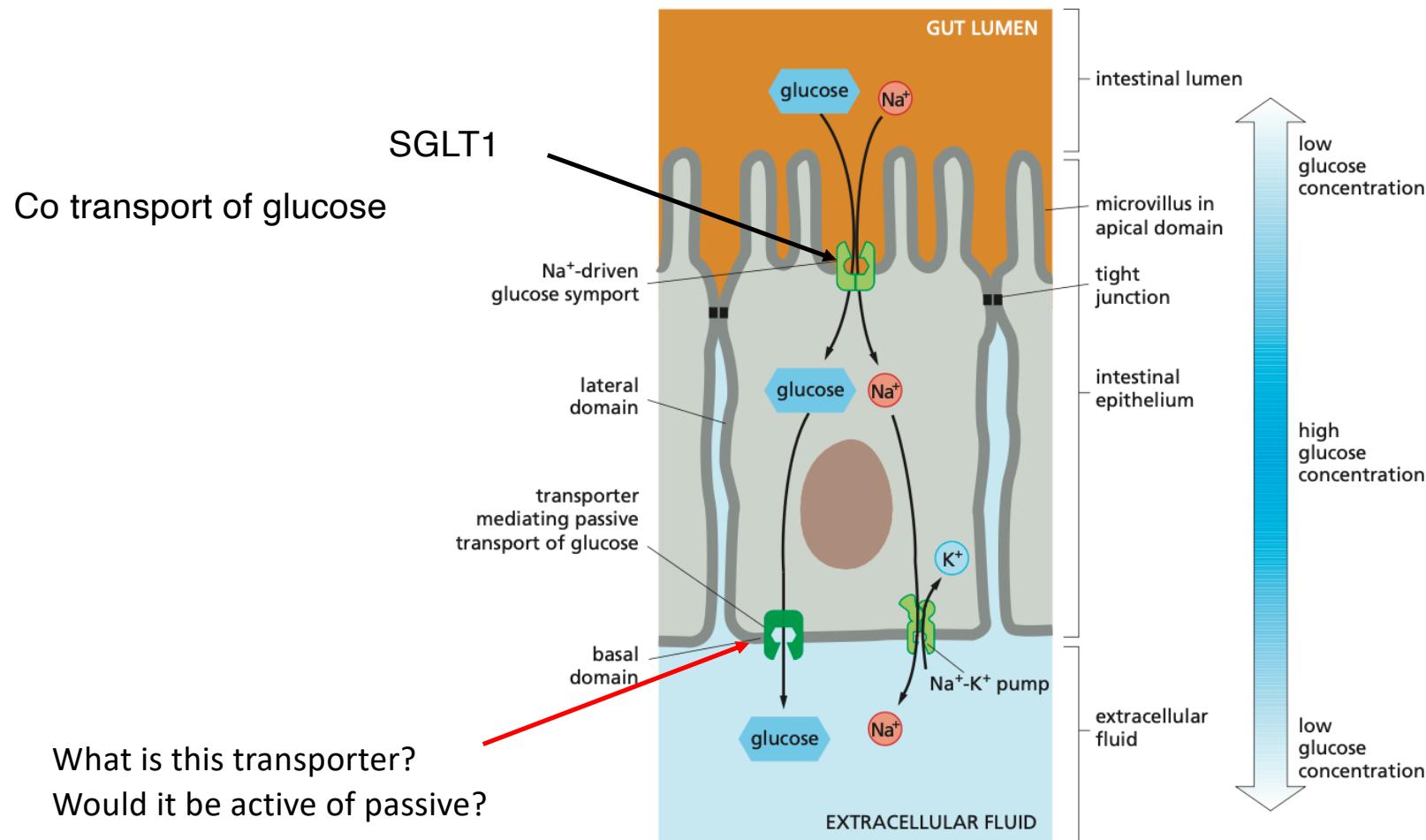


# Transcellular transport

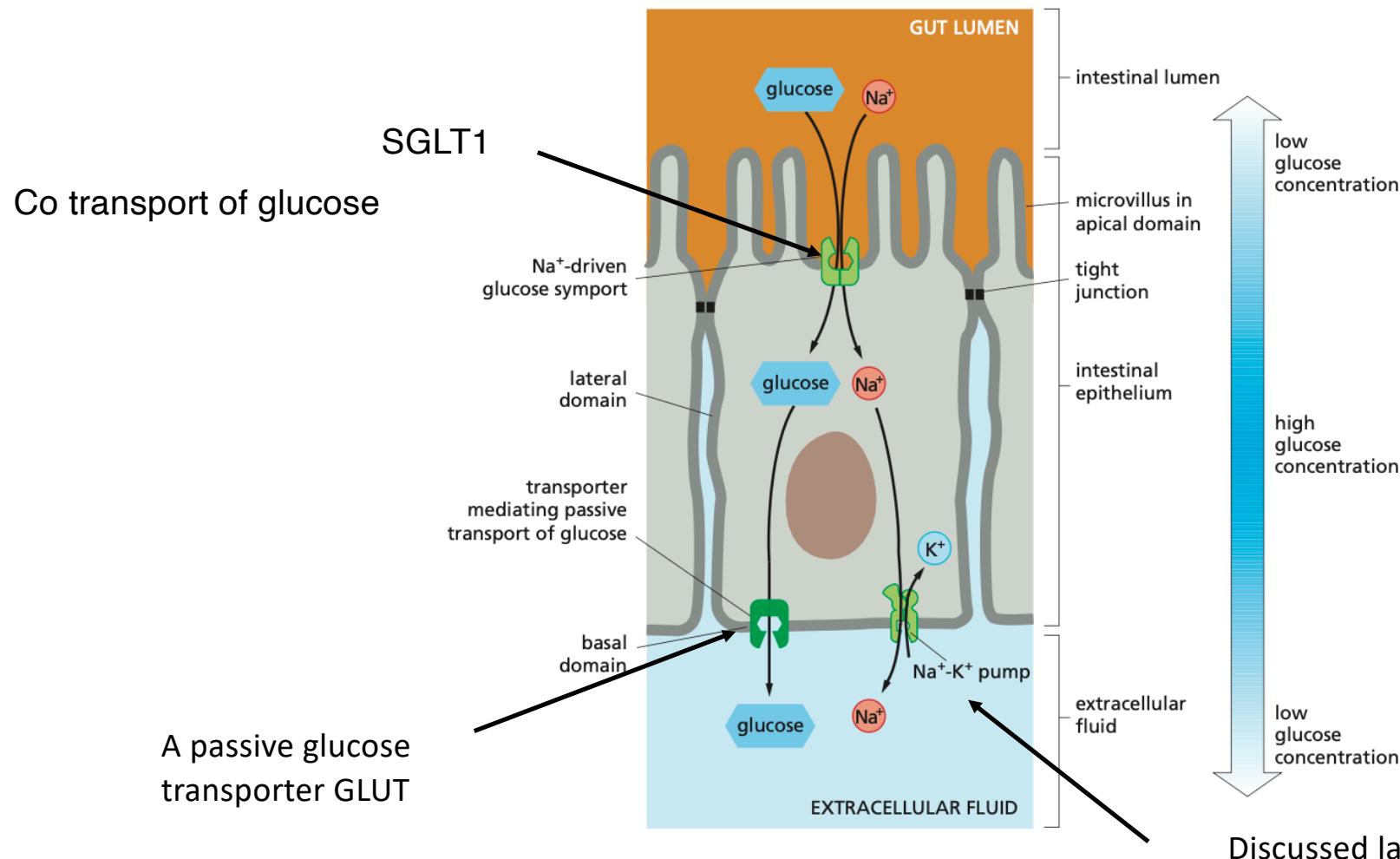
Co transport of glucose



# Transcellular transport



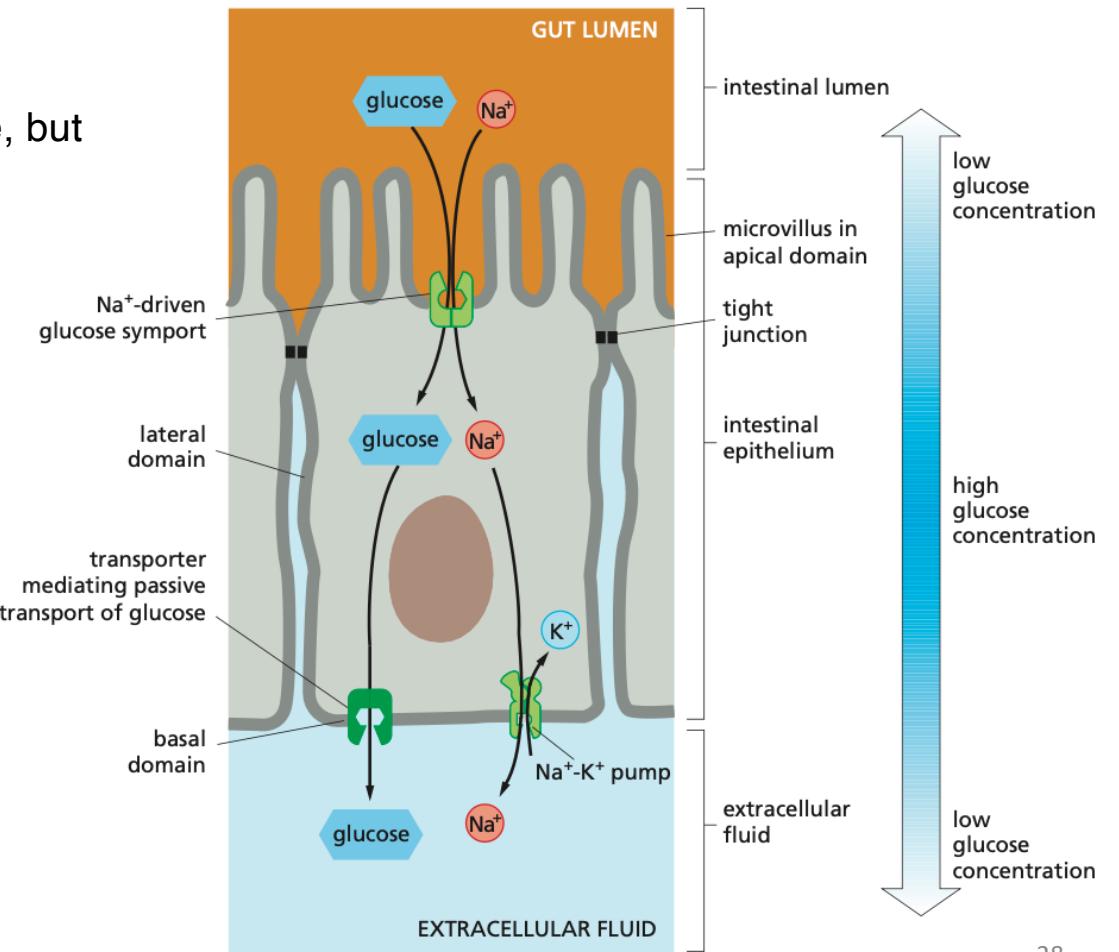
# Transcellular transport



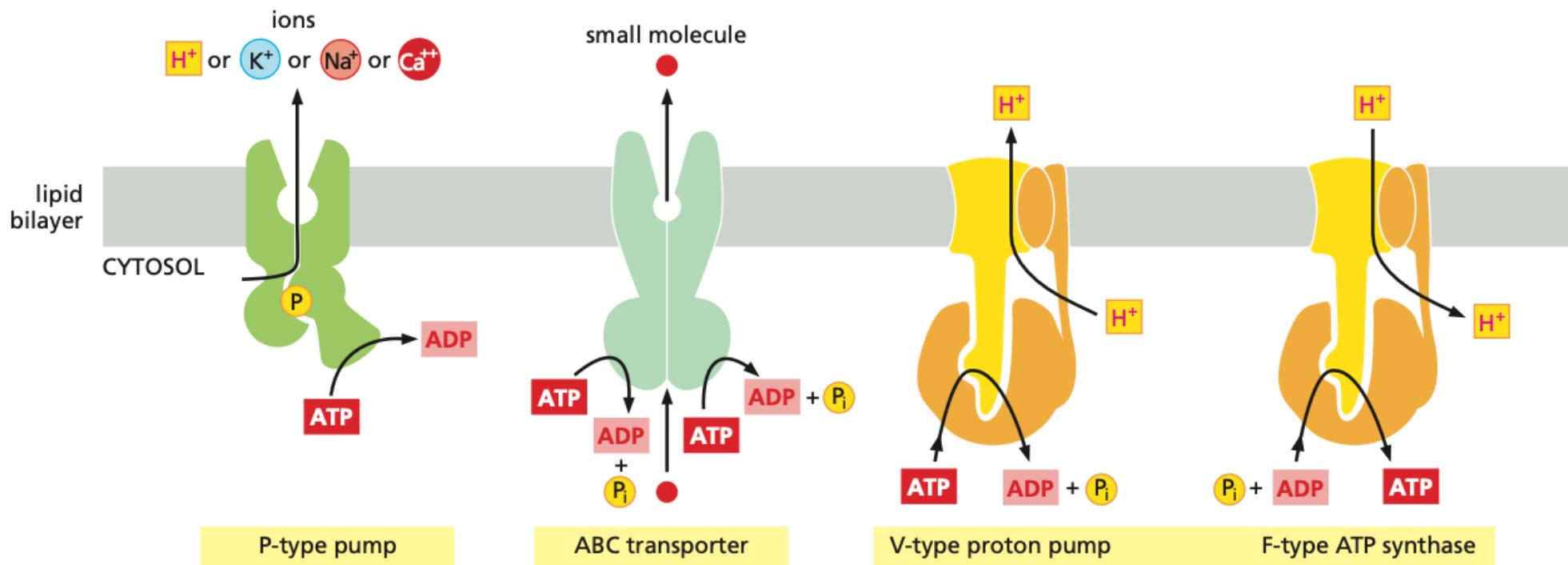
# Transcellular transport

There are 14 members in the GLUT family.

- Transport different substrates. Not only glucose, but fructose (GLUT5), urate (GLUT9).
- Have different  $K_m$  and  $V_{max}$
- Are expressed in different tissues



# Three types of ATP driven pumps



**Figure 11–12** Three types of ATP-driven pumps. Like any enzyme, all ATP-driven pumps can work in either direction, depending on the electrochemical gradients of their solutes and the ATP/ADP ratio. When the ATP/ADP ratio is high, they hydrolyze ATP; when the ATP/ADP ratio is low, they can synthesize ATP. The F-type ATPase in mitochondria normally works in this “reverse” mode to make most of the cell’s ATP.

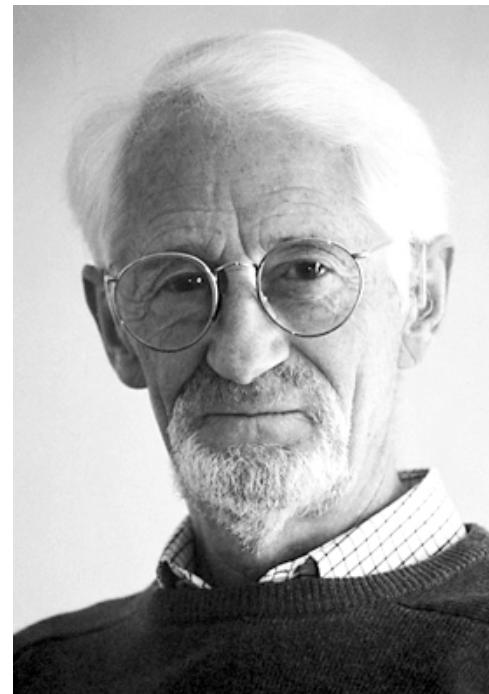
# Examples of ATP driven pumps

Transporter type	function
Na <sup>+</sup> /K <sup>+</sup> pump (P-type)	To maintain gradient of Na <sup>+</sup> and K <sup>+</sup>
Ca <sup>++</sup> ATPase (P-type)	To keep [Ca <sup>++</sup> ] low (for signaling)
Gastric H <sup>+</sup> /K <sup>+</sup> pump (P-type)	To secrete HCl

# The Na<sup>+</sup>/K<sup>+</sup> ATPase

Jens C. Skou

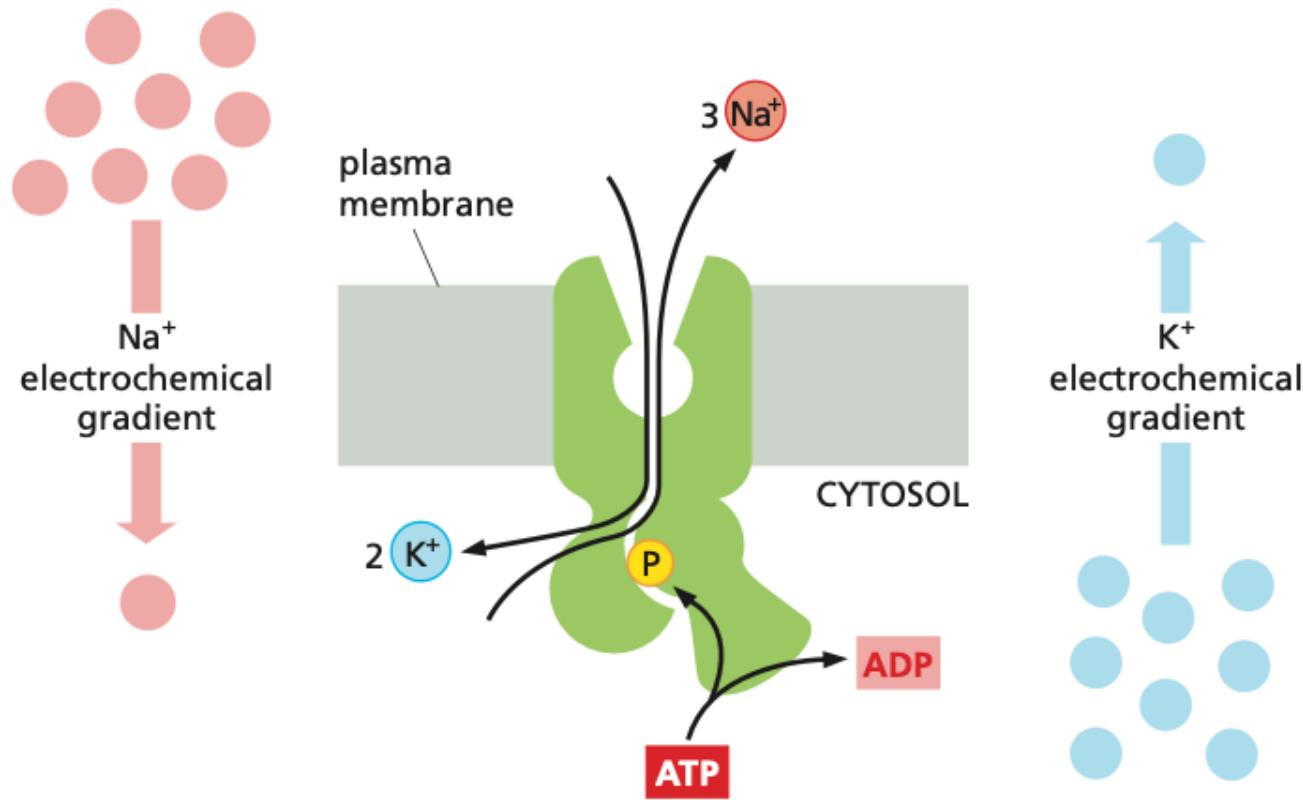
- ATP-driven antiporter  
P-type ATPase  
Maintains the Na<sup>+</sup> gradient important for:
- transport of nutrients into cells - osmotic balance
- 1/3 of the cell's energy is devoted to this pump!



Nobel prize in chemistry, 1997

*"for the first discovery of an ion-transporting enzyme, Na<sup>+</sup>, K<sup>+</sup> -ATPase"*

# The function of the Na<sup>+</sup>-K<sup>+</sup> pump



# The cycle of the $\text{Na}^+-\text{K}^+$ pump

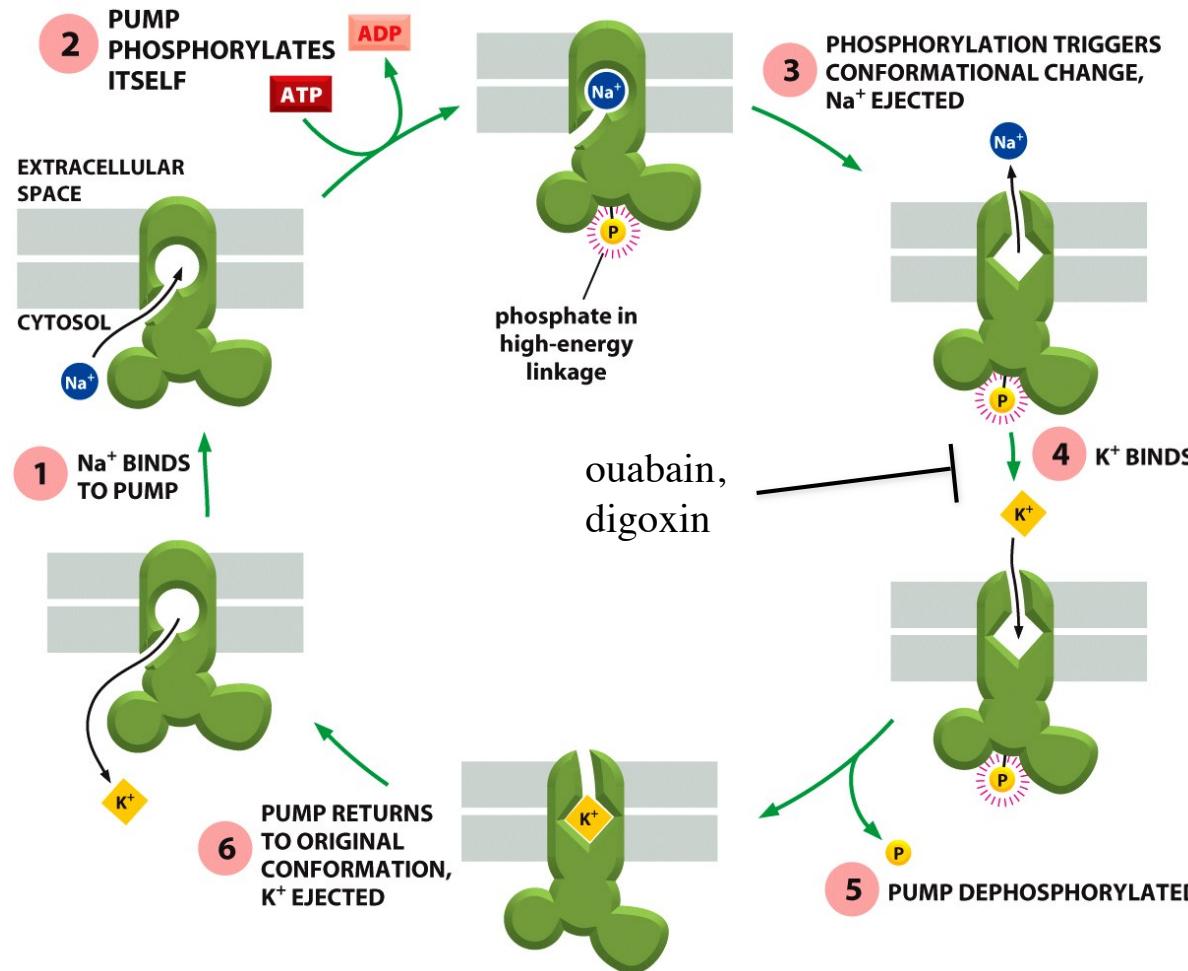


Figure 12-11 *Essential Cell Biology* (© Garland Science 2010)

# The Na<sup>+</sup>/K<sup>+</sup> ATPase is important to regulate cellular osmolarity

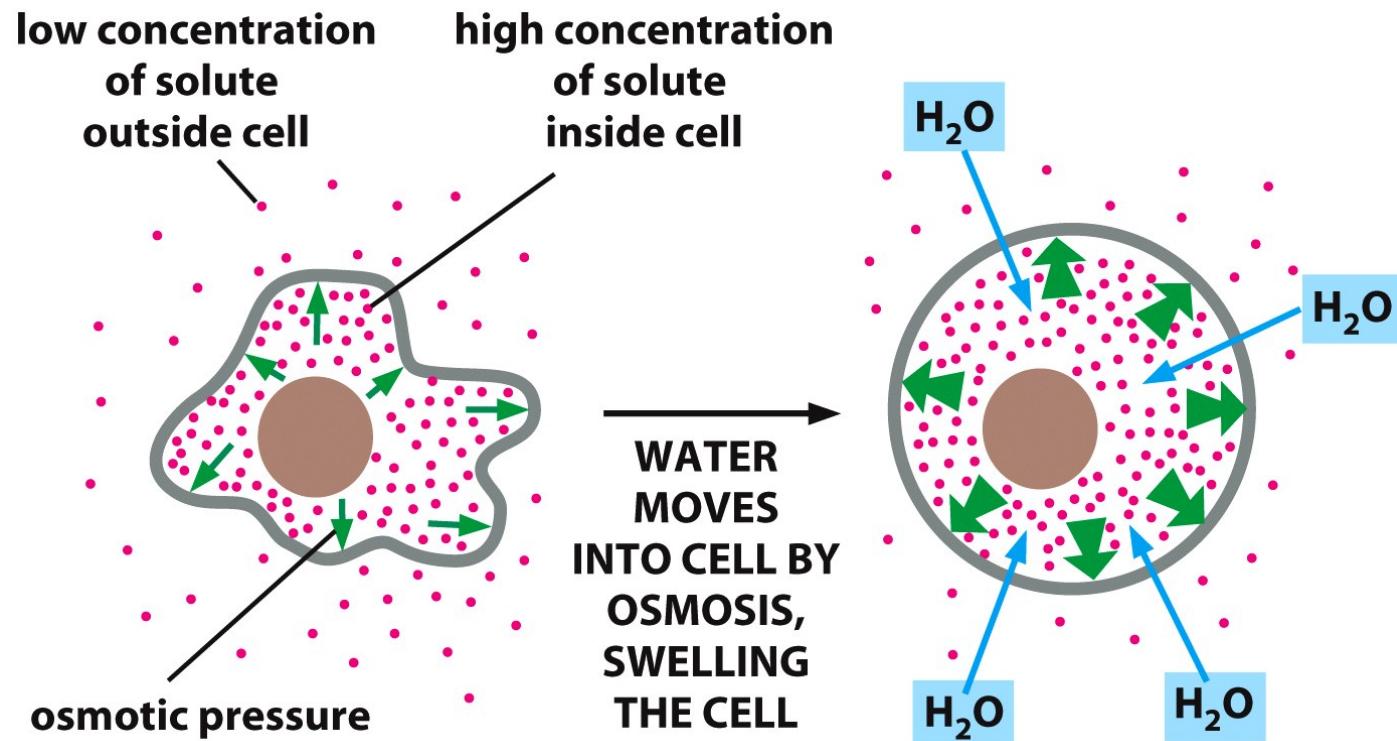


Figure 12-12 *Essential Cell Biology* (© Garland Science 2010)

# Osmolarity in red blood cells

- Plasma membrane has high permeability to  $\text{H}_2\text{O}$
- $\text{Na}^+/\text{K}^+$  pump is important to keep red blood cell volume

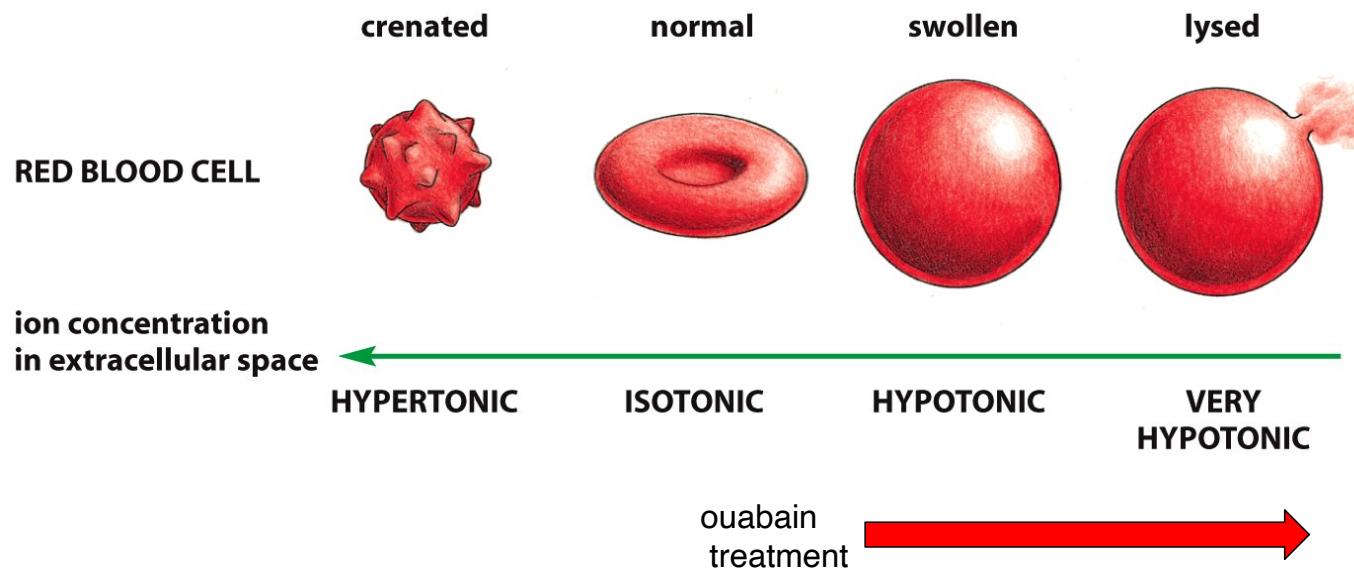
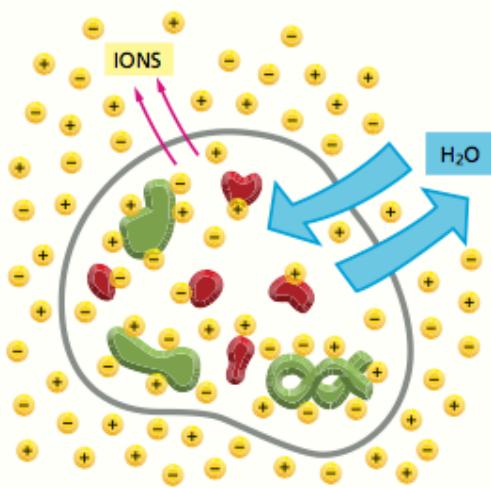


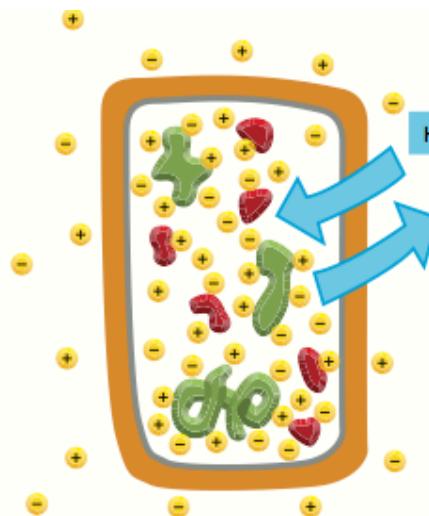
Figure 11-16 *Molecular Biology of the Cell* (© Garland Science 2008)

# Osmolarity across life

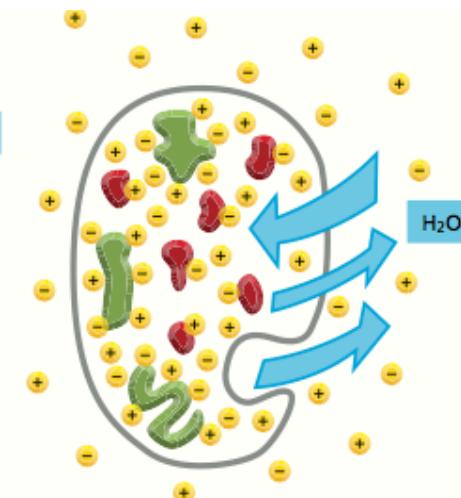
Animal cells and bacteria



Plant cells



Protozoa



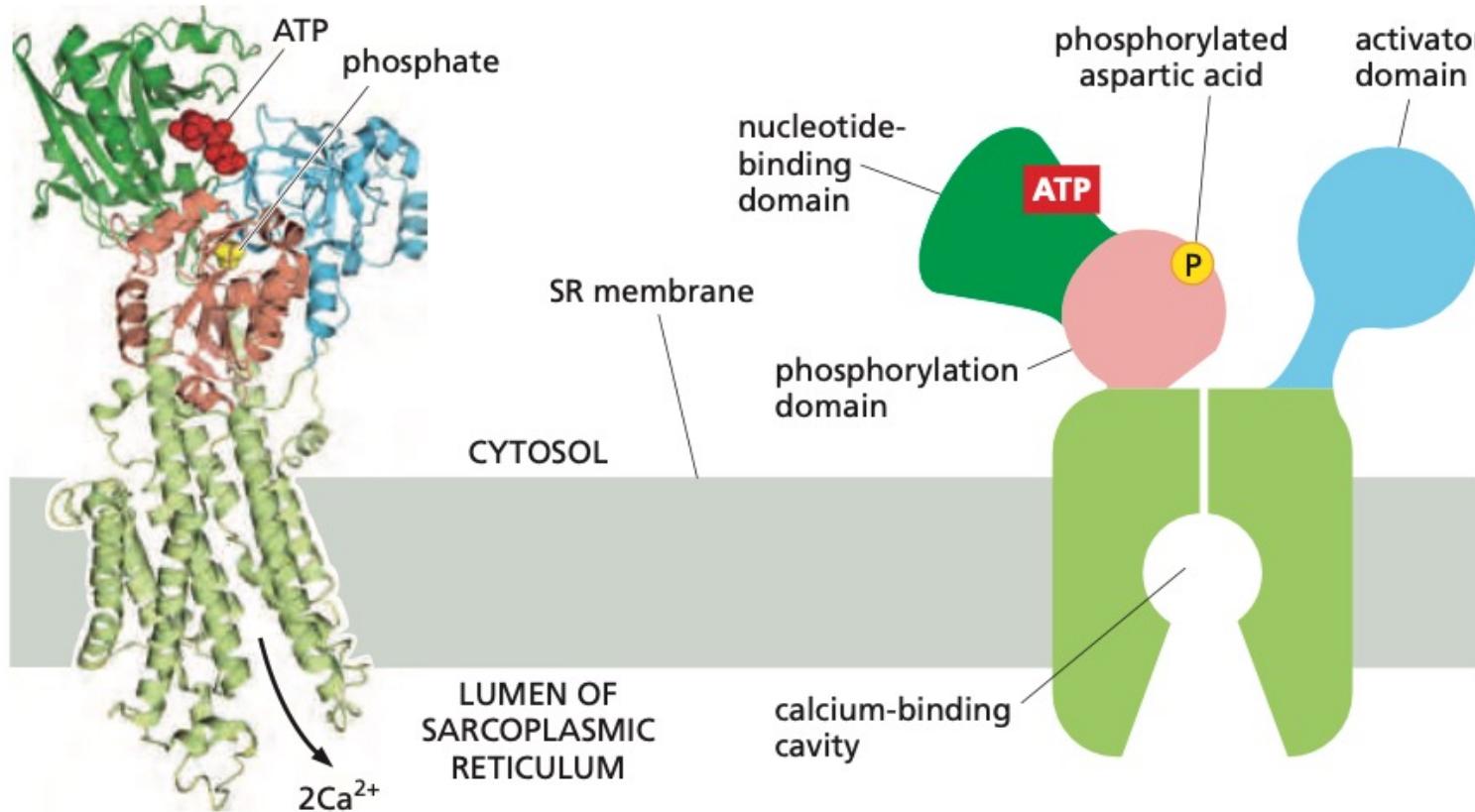
Active pumping out  
To have lower intracellular [ion], to  
compensate the excess of organic solutes

Can tolerate an osmotic  
difference, due to rigid wall

Extrude water from vacuoles

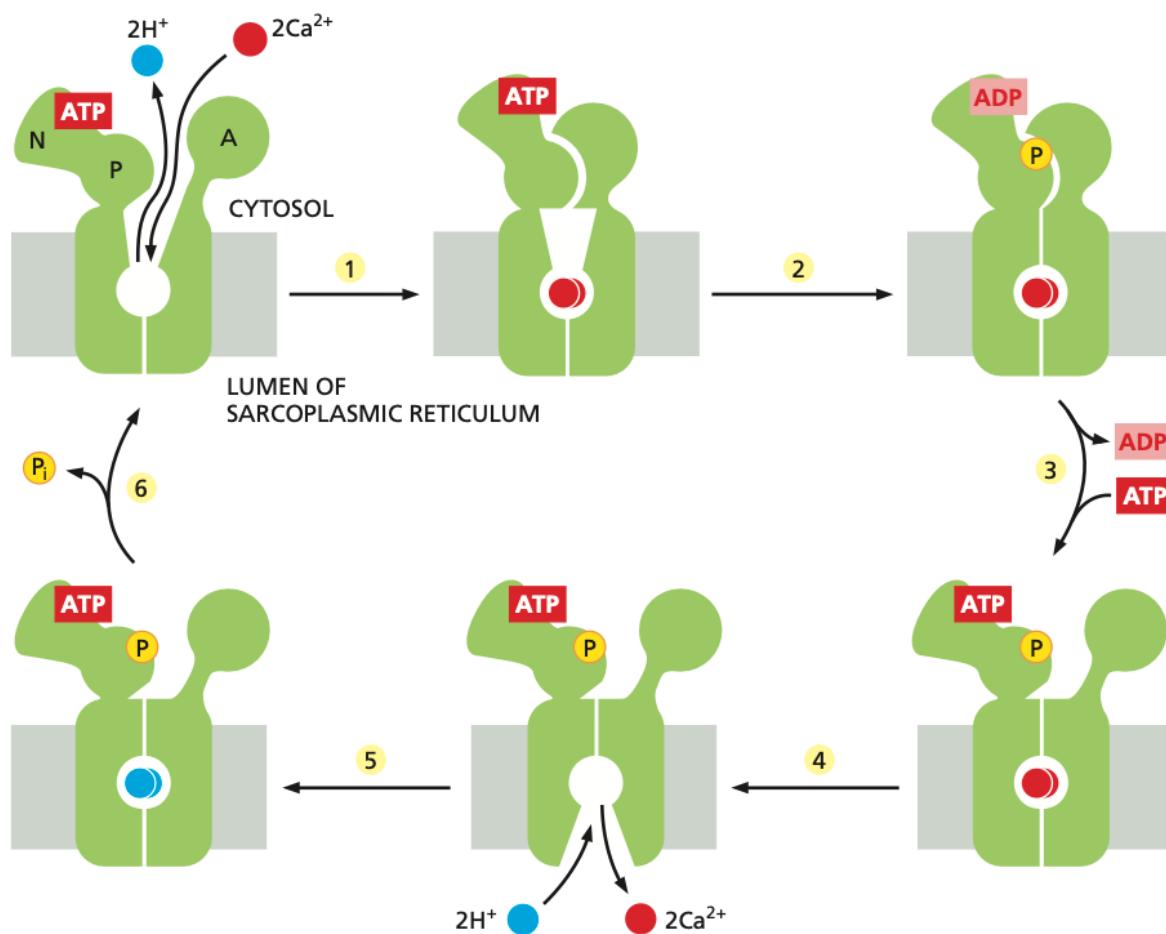
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# the sarcoplasmic reticulum $\text{Ca}^{2+}$ pump.



**Figure 11–13** The structure of the sarcoplasmic reticulum  $\text{Ca}^{2+}$  pump. The ribbon model (left), derived from x-ray crystallographic analyses, shows the pump in its phosphorylated, ATP-bound state. The three globular cytosolic domains of the pump—the nucleotide-binding domain (dark green), the activator domain (blue), and the phosphorylation domain (red), also shown schematically on the right—change conformation dramatically during the pumping cycle. These changes in turn alter the arrangement of the transmembrane helices, which allows the  $\text{Ca}^{2+}$  to be released from its binding cavity into the SR lumen (Movie 11.3). (PDB code: 3B9B.)

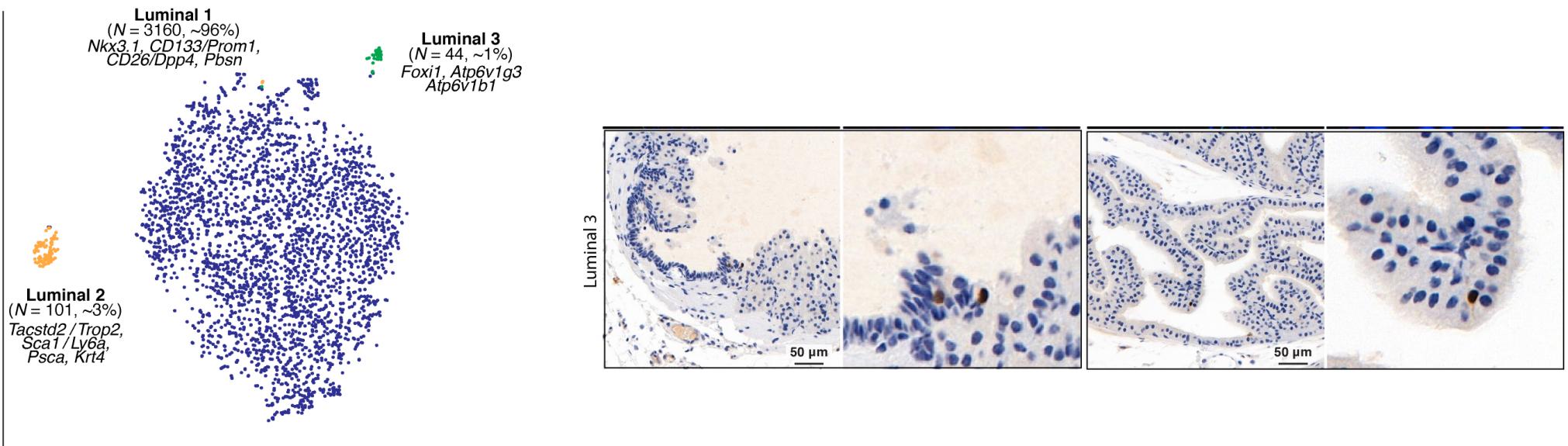
# The pumping cycle of the sarcoplasmic reticulum $\text{Ca}^{2+}$ pump.



**Figure 11-14** The pumping cycle of the sarcoplasmic reticulum  $\text{Ca}^{2+}$  pump. Ion pumping proceeds by a series of stepwise conformational changes in which movements of the pump's three cytosolic domains [the nucleotide-binding domain (N), the phosphorylation domain (P), and the activator domain (A)] are mechanically coupled to movements of the transmembrane  $\alpha$  helices. Helix movement opens and closes passageways through which  $\text{Ca}^{2+}$  enters from the cytosol and binds to the two centrally located  $\text{Ca}^{2+}$  binding sites. The two  $\text{Ca}^{2+}$  then exit into the SR lumen and are replaced by two  $\text{H}^{+}$ , which are transported in the opposite direction. The  $\text{Ca}^{2+}$ -dependent phosphorylation and  $\text{H}^{+}$ -dependent dephosphorylation of aspartic acid are universally conserved steps in the reaction cycle of all P-type pumps: they cause the conformational transitions to occur in an orderly manner, enabling the proteins to do useful work. (Adapted from C. Toyoshima et al., *Nature* 432:361–368, 2004 and J.V. Møller et al., *Q. Rev. Biophys.* 43:501–566, 2010.)

# Cell type Luminal 3 in the prostate (Karthaus lab)

A cell expressing vacuolar-type  $H^+$ -ATPase (V-ATPase) identified in the male prostate  
Regulates the pH of reproductive fluids  
Necessary for reproduction!

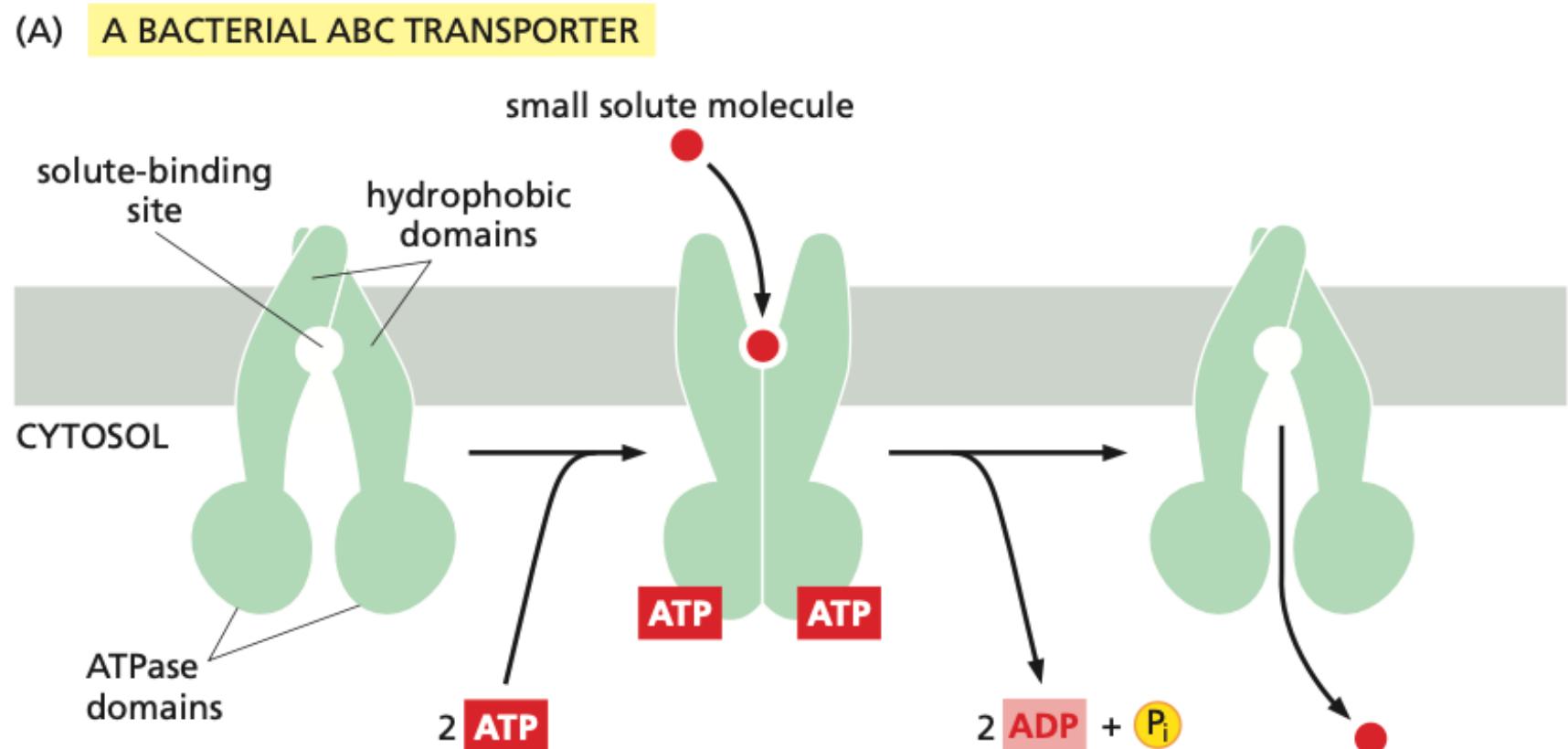


# **ABC transporters**

# ABC transporters

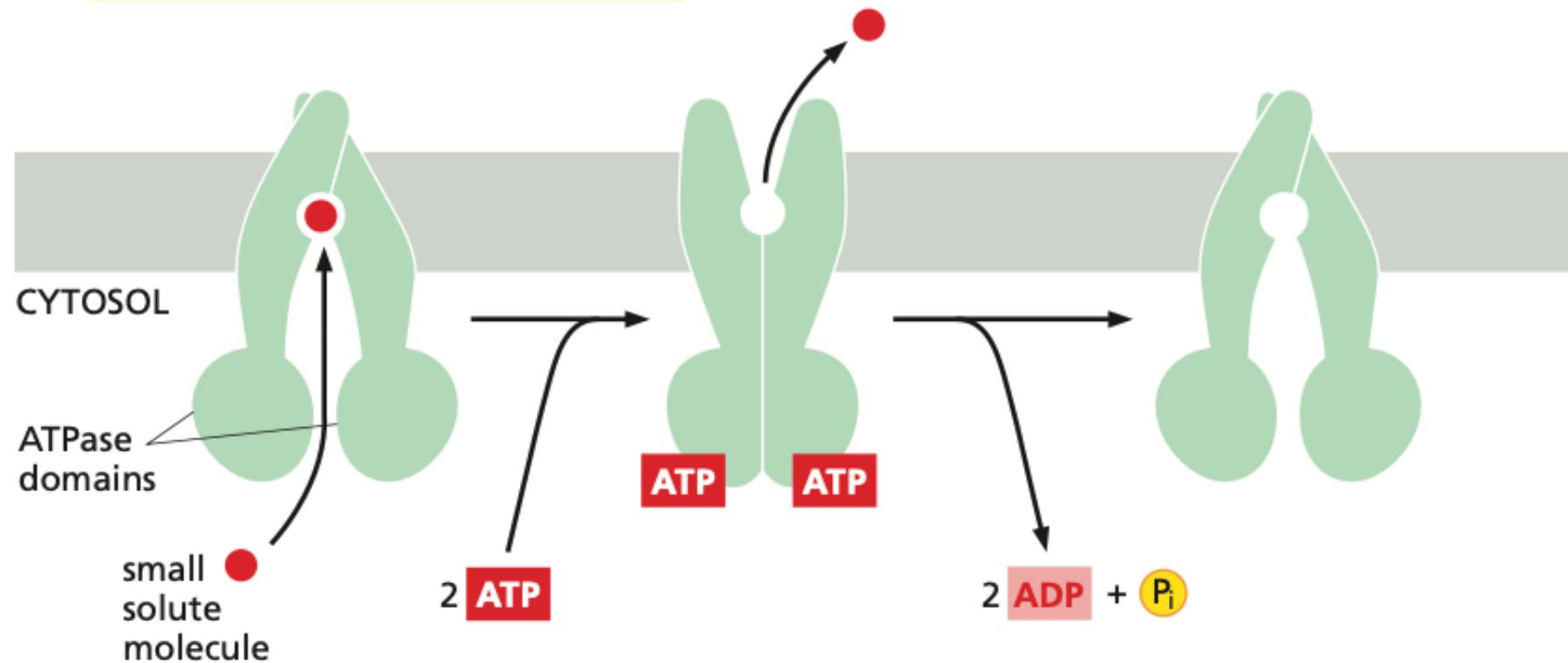
- Large family of membrane transport proteins (ex: 5% of *E. coli* genome). Exist in all phyla. 48 proteins in human.
- 2ATPase domains (ATP-binding cassettes) involved
- ATP binding leads to dimerization of the ATP-binding domains
- ATP hydrolysis leads to their dissociation
- Transport a wide variety of substrates: sugars, amino acids, drugs, antibiotics, toxins, lipids, peptides, nucleotides and more

# Small-molecule transport by typical ABC transporters

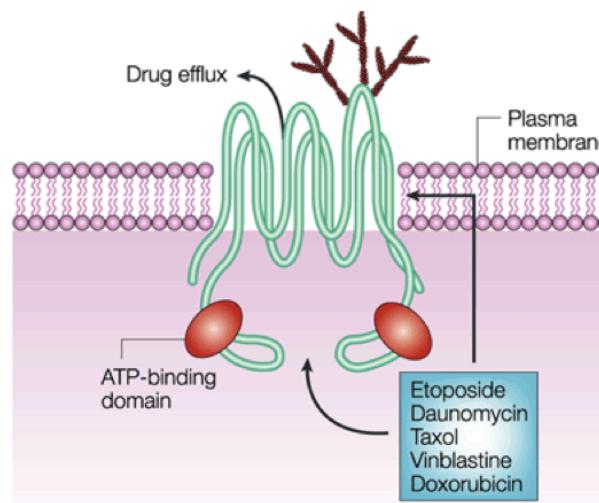


# Small-molecule transport by typical ABC transporters

(B) A EUKARYOTIC ABC TRANSPORTER



The multidrug resistance (MDR) efflux pumps.  
The expression of ABC drug transporters has been linked to  
chemotherapy resistance and poor prognosis.

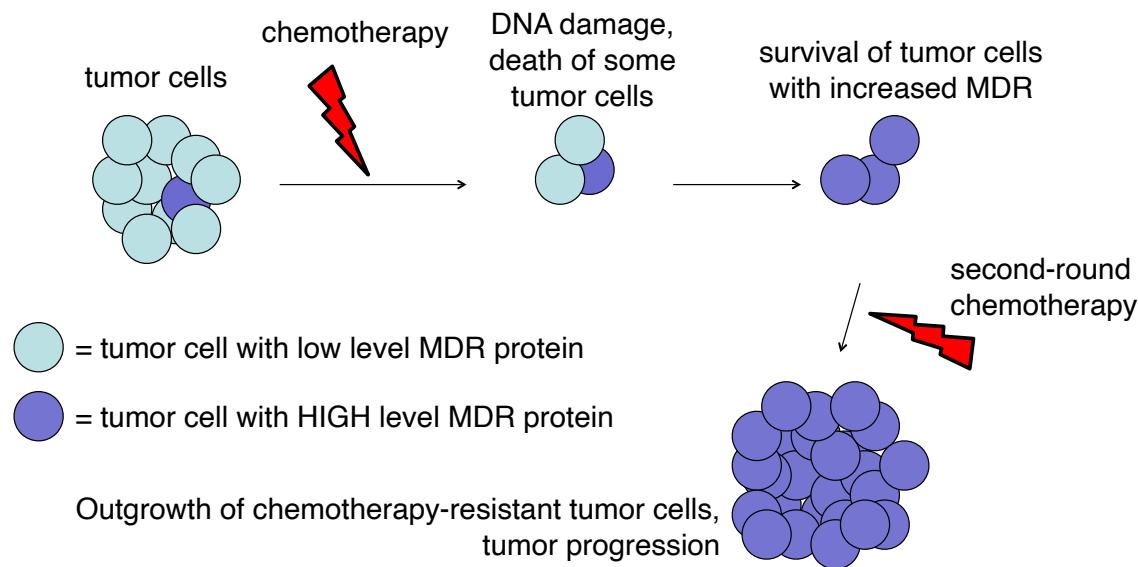


Nature Reviews | Cancer

Brian P. Sorrentino  
Nature Reviews Cancer 2, 431-441 (2002)

## Some eukaryotic ABC transporters promote resistance to chemotherapies

The multidrug resistance (MDR) proteins in cancer promote drug efflux



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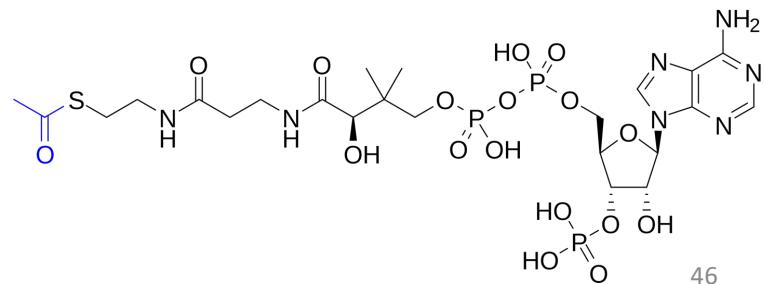
# An example of a transporter dependency in cancer



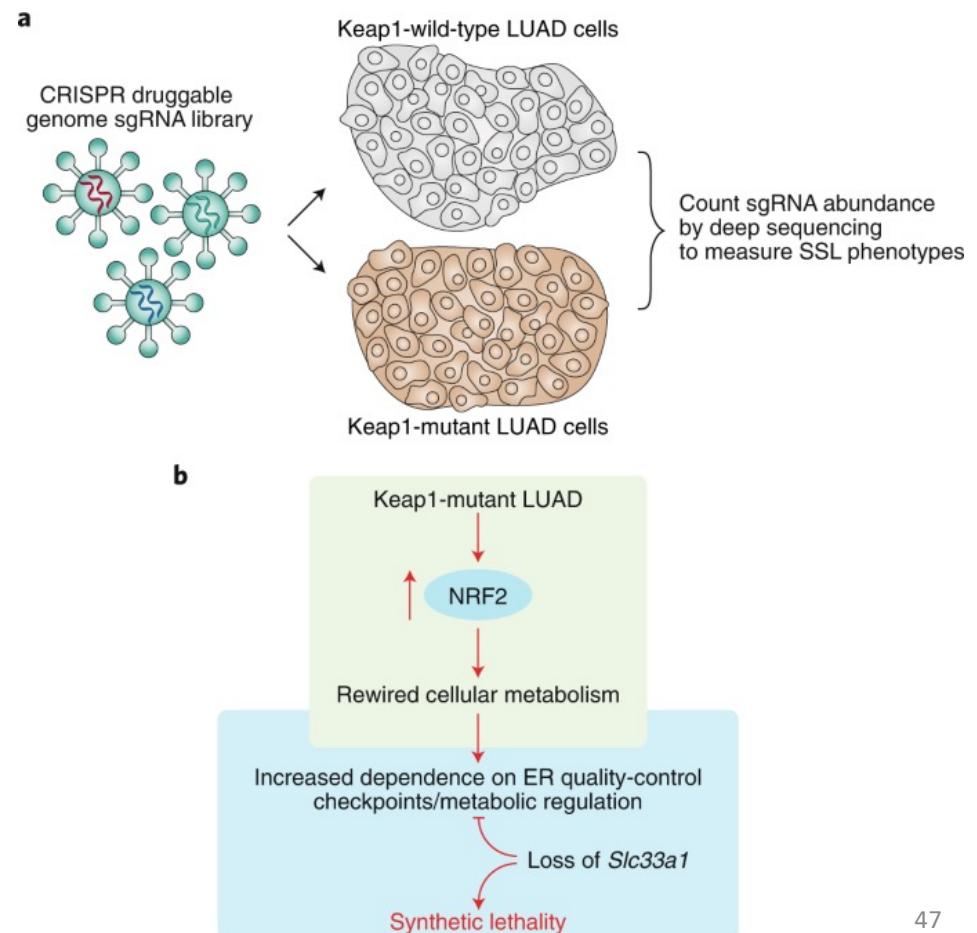
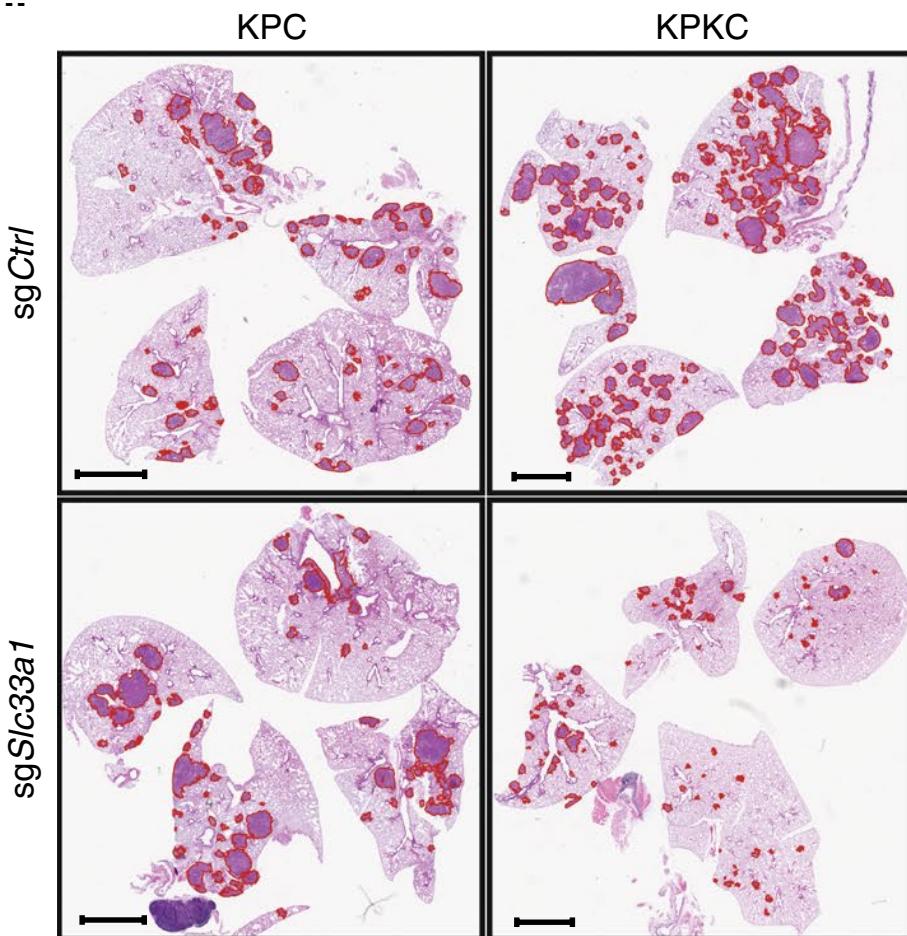
## Keap1 mutation renders lung adenocarcinomas dependent on *Slc33a1*

Rodrigo Romero  <sup>1,2,11</sup>, Francisco J. Sánchez-Rivera  <sup>1,2,9,11</sup>, Peter M. K. Westcott <sup>1</sup>, Kim L. Mercer <sup>1,3</sup>, Arjun Bhutkar <sup>1</sup>, Alexander Muir <sup>1,10</sup>, Tania J. González Robles <sup>1</sup>, Swanny Lamboy Rodríguez <sup>2</sup>, Laura Z. Liao  <sup>2</sup>, Sheng Rong Ng  <sup>1,2</sup>, Leanne Li <sup>1</sup>, Caterina I. Colón <sup>1</sup>, Santiago Naranjo <sup>1,2</sup>, Mary Clare Beytagh <sup>2</sup>, Caroline A. Lewis  <sup>4</sup>, Peggy P. Hsu <sup>1,5,6</sup>, Roderick T. Bronson <sup>7,8</sup>, Matthew G. Vander Heiden  <sup>1,2,6</sup> and Tyler Jacks  <sup>1,2,3</sup> 

**Acetyl-coenzyme A transporter 1** also known as **solute carrier family 33 member 1** (SLC33A1) is a [protein](#) that in humans is encoded by the SLC33A1 [gene](#).<sup>[5]</sup>

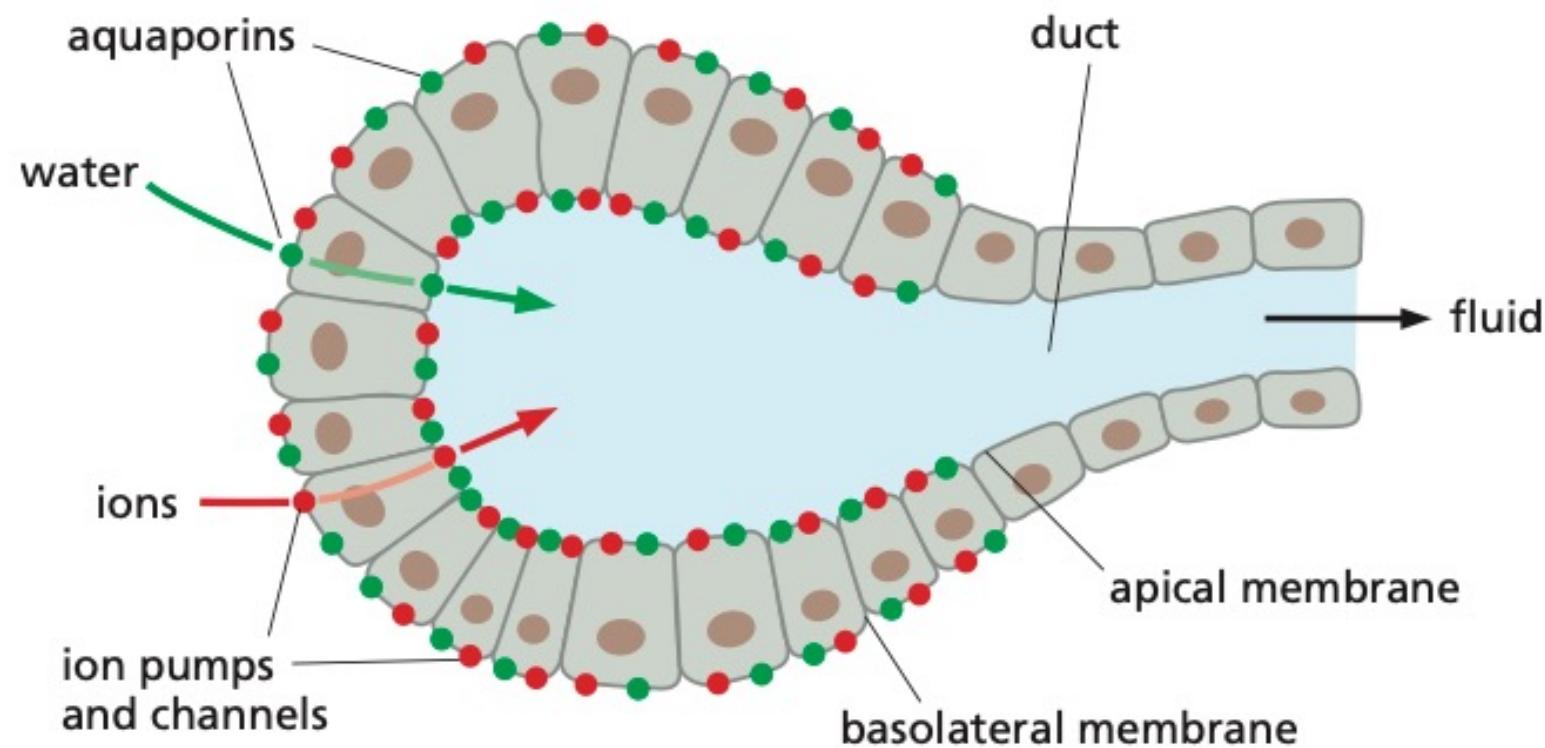


# An example of a transporter dependency in cancer



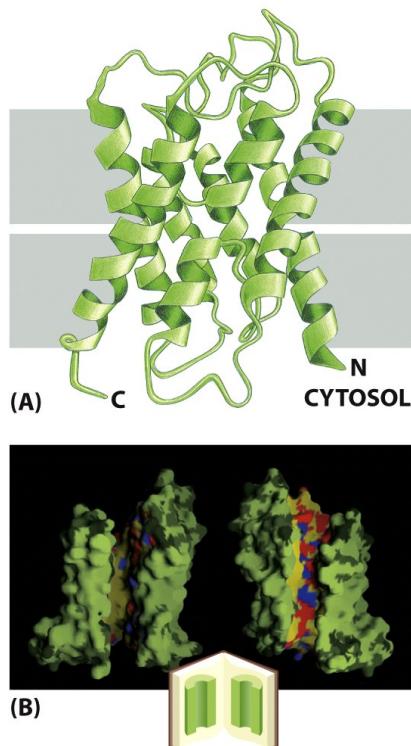
# **Aquaporins**

# The role of aquaporins in fluid secretion



# Aquaporins

There are thirteen known types of aquaporins in mammals, and six of these are located in the kidney  
Transport water at high rates  
Ions cannot pass



Pore is very thin  
↓  
Ions need to be dehydrated  
↓  
Energy loss not compensated because the hydrophobic wall of the pore cannot interact with dehydrated ion  
↓  
Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>++</sup>, Cl<sup>-</sup> excluded

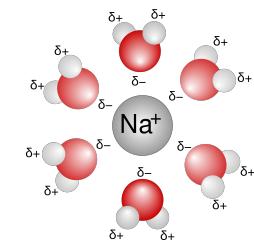
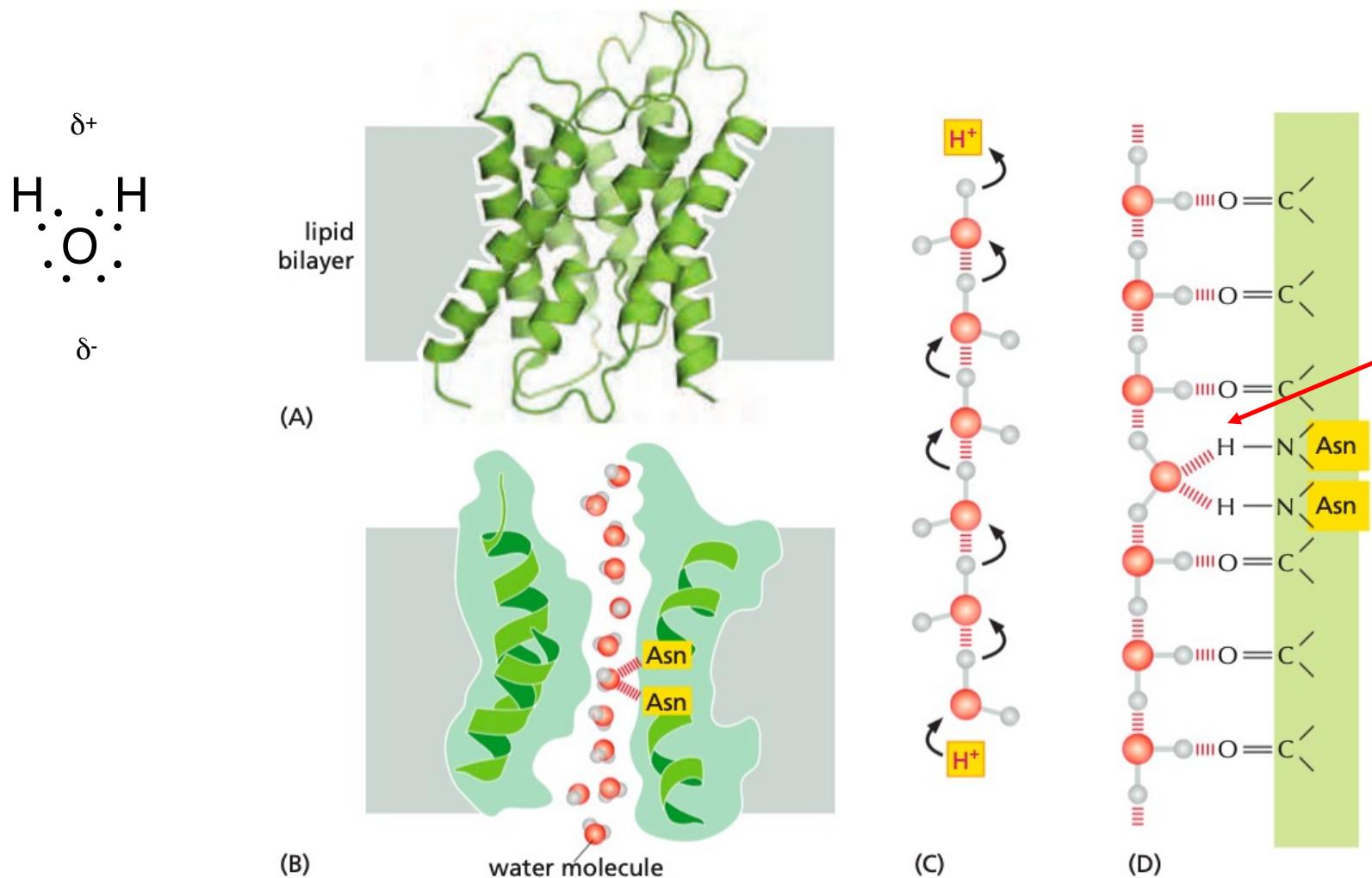


Figure 11-27 *Molecular Biology of the Cell* (© Garland Science 2008)

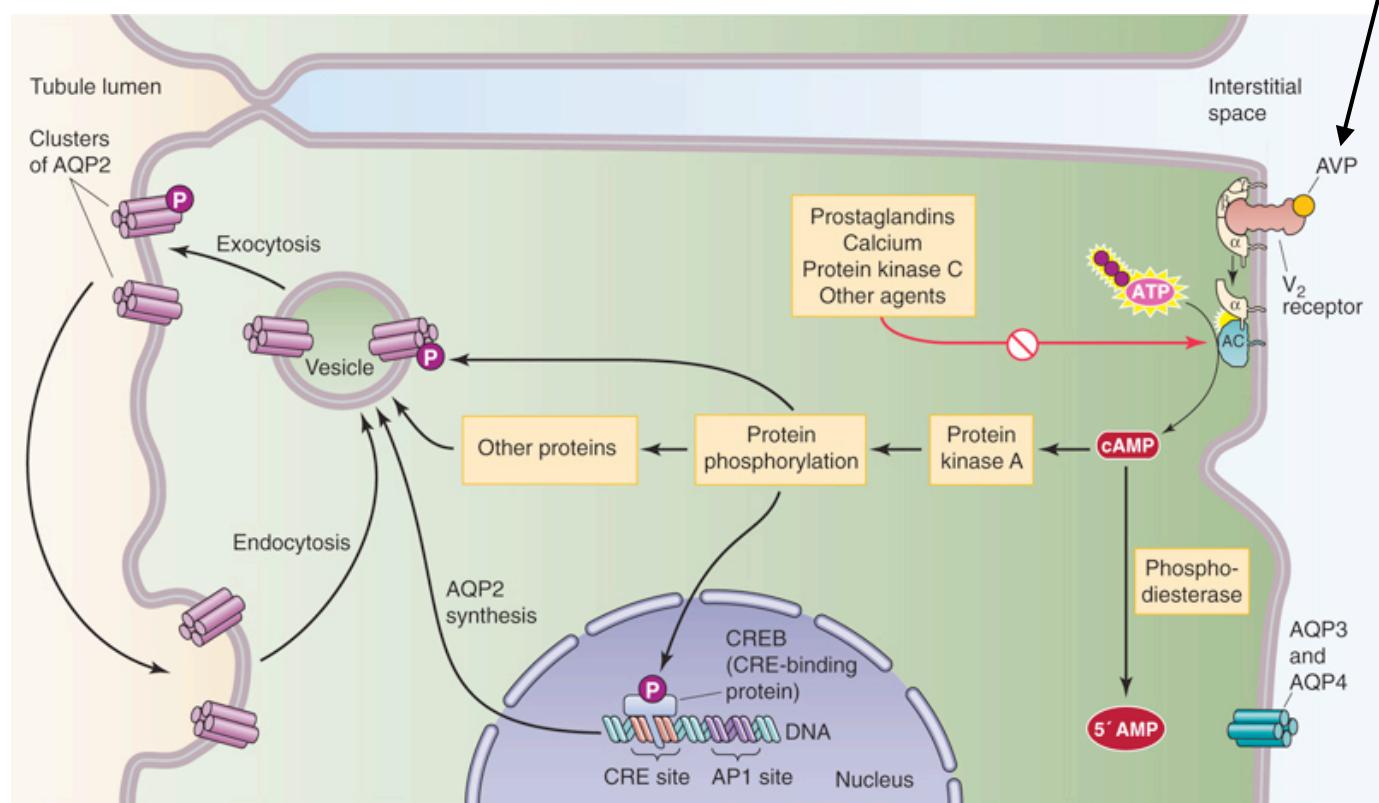
# The structure of aquaporins



# The response to dehydration

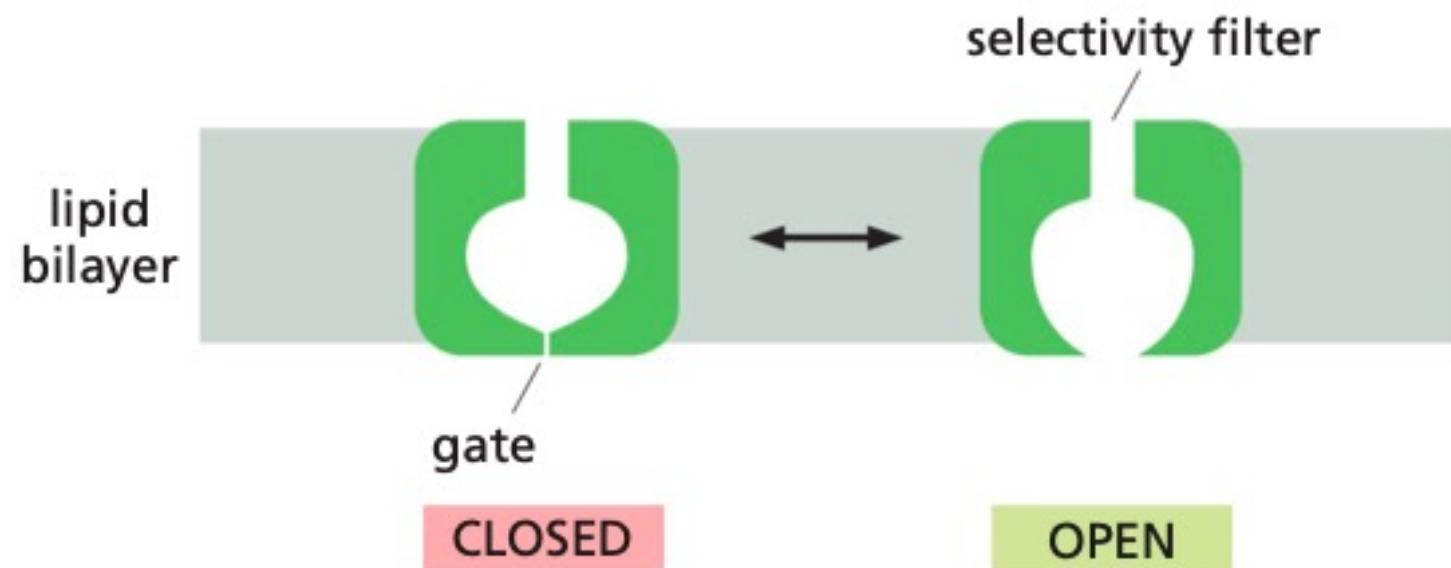
Effect of vasopressin on aquaporin-2 (AQP2) expression in kidney epithelial cells, in response to dehydration

hypothalamus: vasopressin



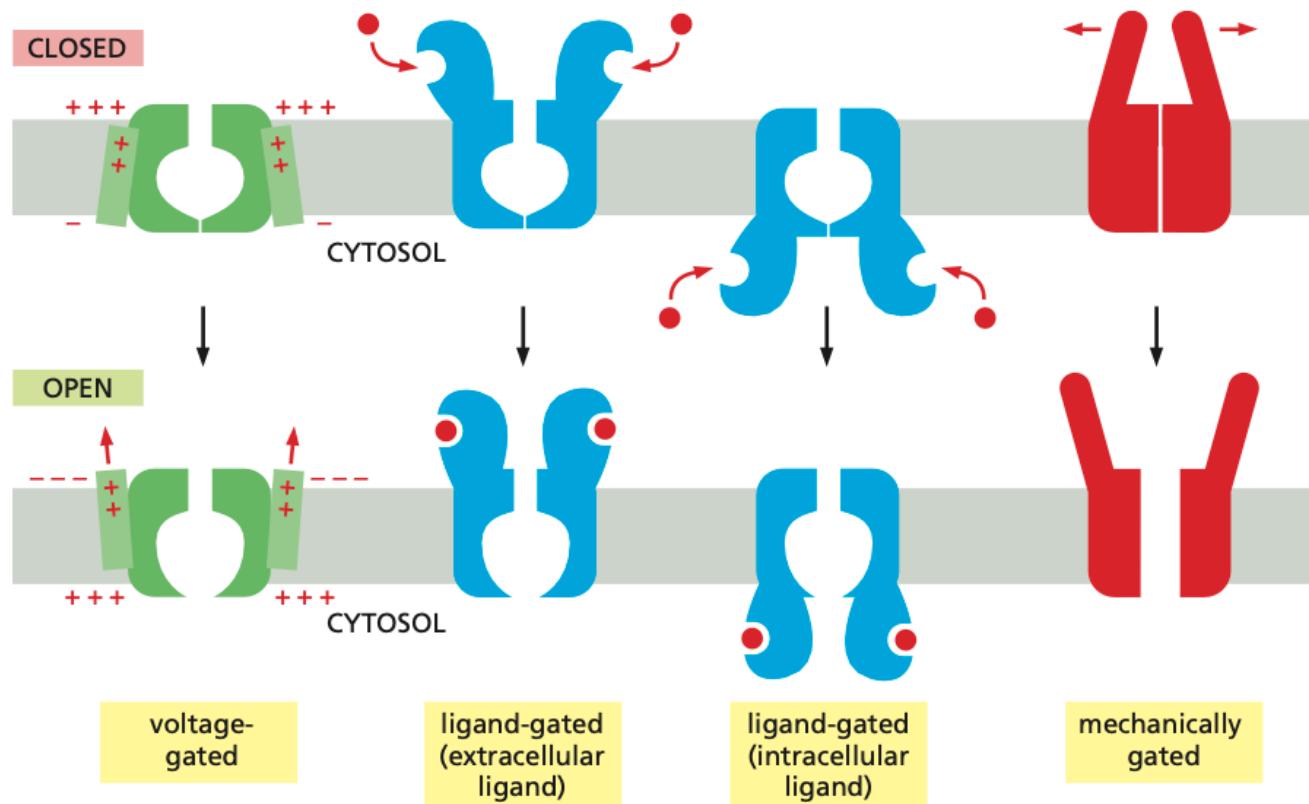
# **Ion channels**

# a typical ion channel, fluctuates between closed and open conformations



**Figure 11–21** A typical ion channel, which fluctuates between closed and open conformations. The ion channel shown here in cross section forms a pore across the lipid bilayer only in the “open” conformational state. The pore narrows to atomic dimensions in one region (the selectivity filter), where the ion selectivity of the channel is largely determined. Another region of the channel forms the gate. <sup>54</sup>

# The gating of ion channels can be regulated in many ways



**Figure 11–22** The gating of ion channels. This schematic drawing shows several kinds of stimuli that open ion channels. Mechanically gated channels often have cytoplasmic extensions (not shown) that link the channel to the cytoskeleton.

# Ion channel selectivity

## Ion channels

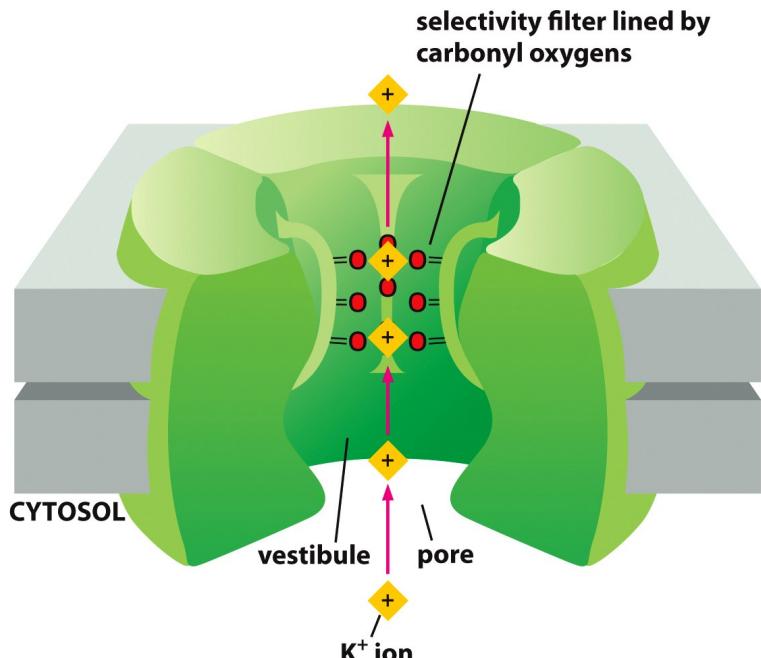


Figure 12-20 *Essential Cell Biology* (© Garland Science 2010)

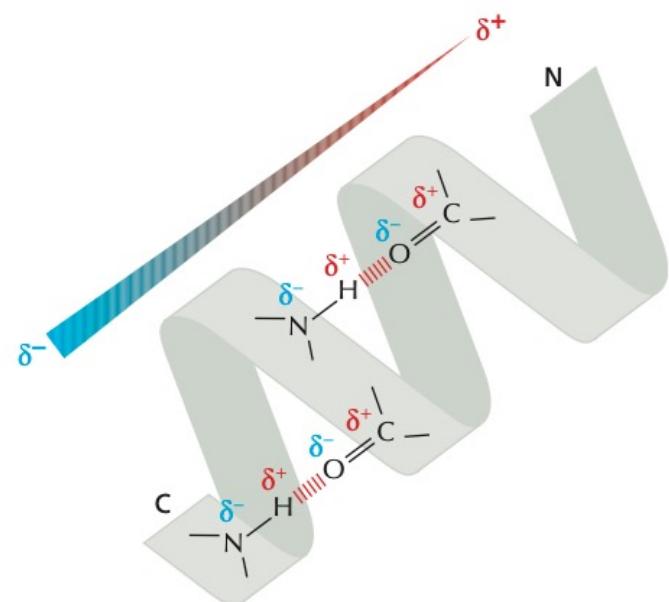
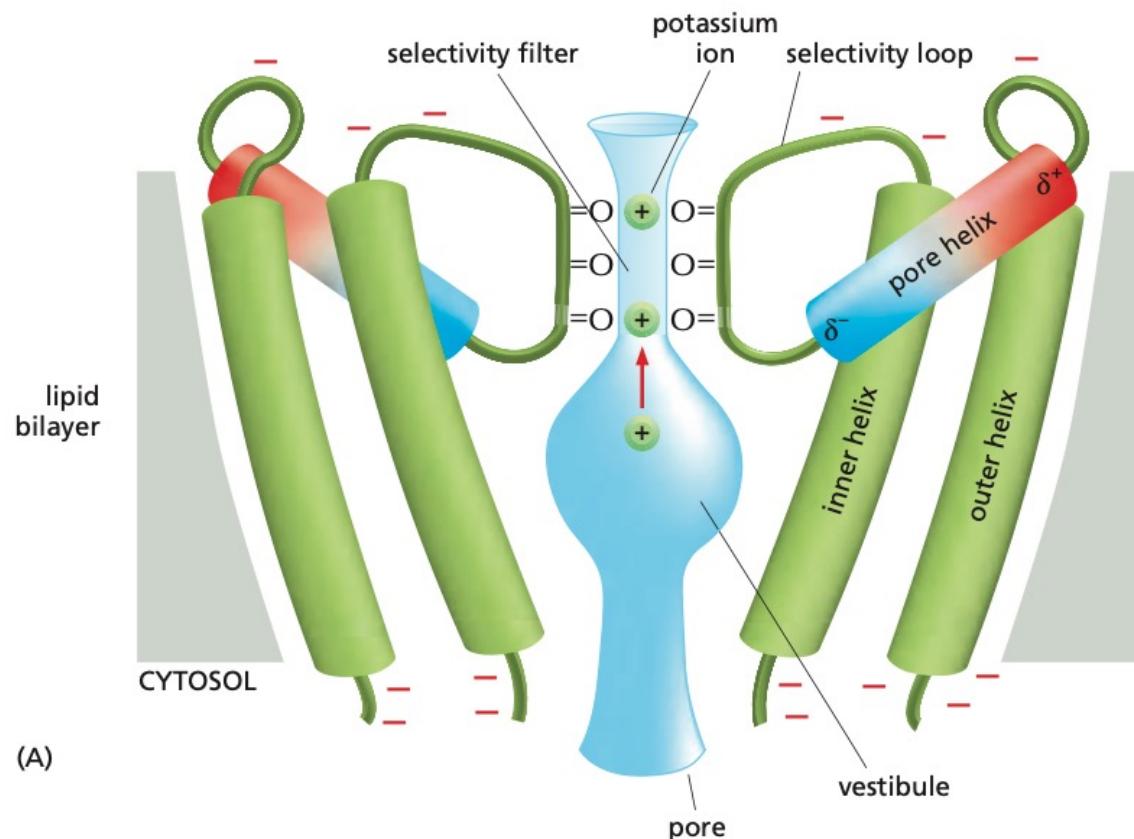
To ensure that only ions of appropriate size and charge can pass

The ions shed their associated H<sub>2</sub>O molecules to pass

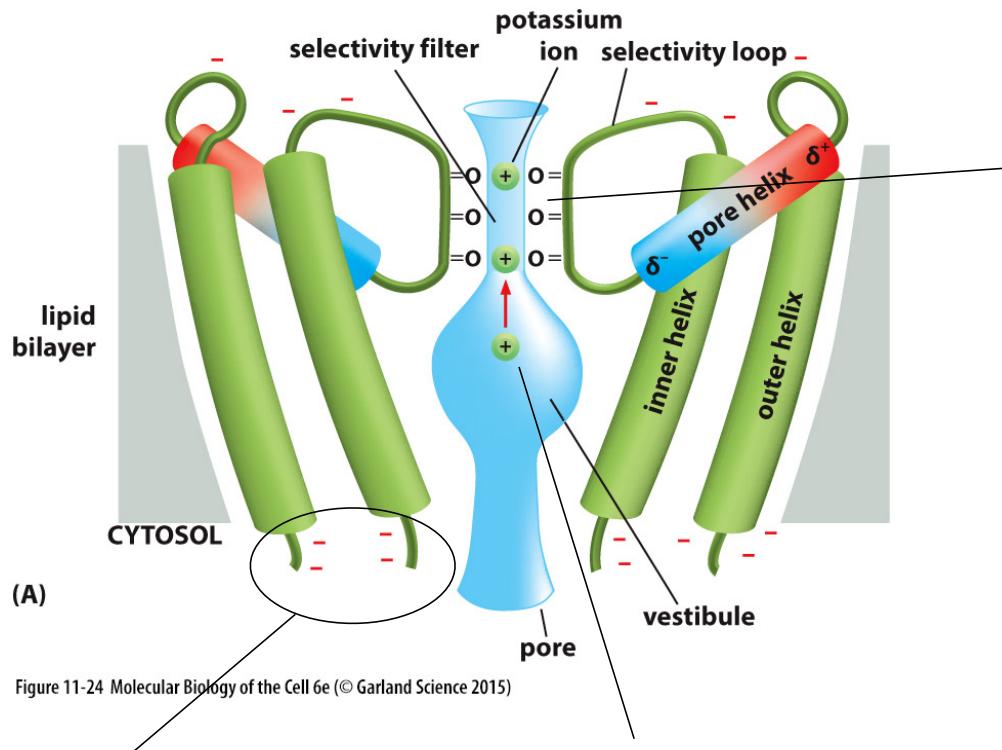
# Ion channel selectivity

Why don't  $K_+$  channels let  $Na_+$  pass through ? Not the pore size:  $Na_+$  is smaller.  
High affinity binding to  $K_+$  is not the answer. Otherwise speed would be greatly affected.  
The answer came from the crystal structure of a bacterial  $K_+$  channel

# The structure of a bacterial K<sup>+</sup> channel



# The structure of a bacterial K<sup>+</sup> channel

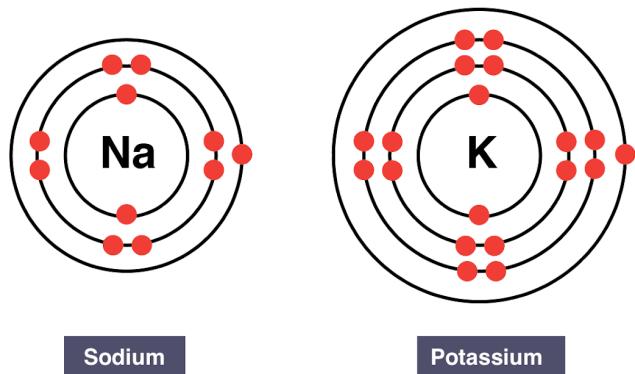


Cation attraction by negatively charged amino acids

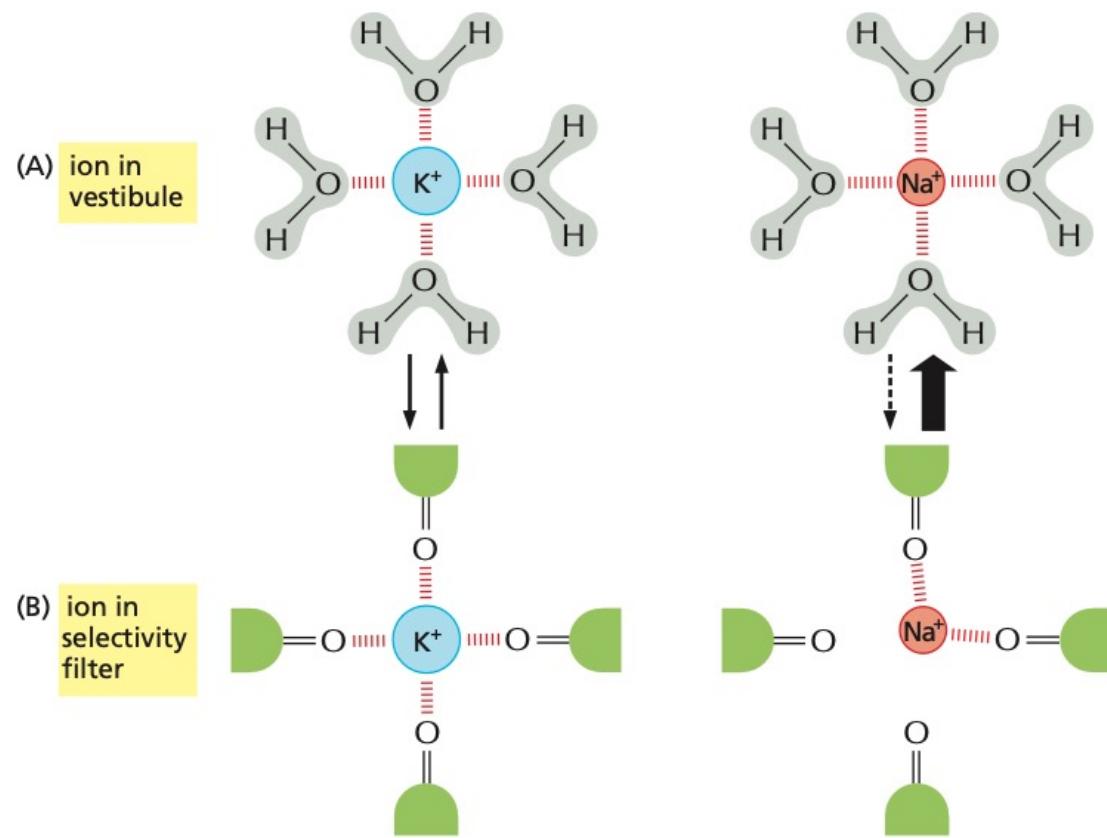
Coordination between carbonyl oxygens and dehydrated K<sup>+</sup>

In the vestibule, there can still be Na<sup>+</sup> and K<sup>+</sup>

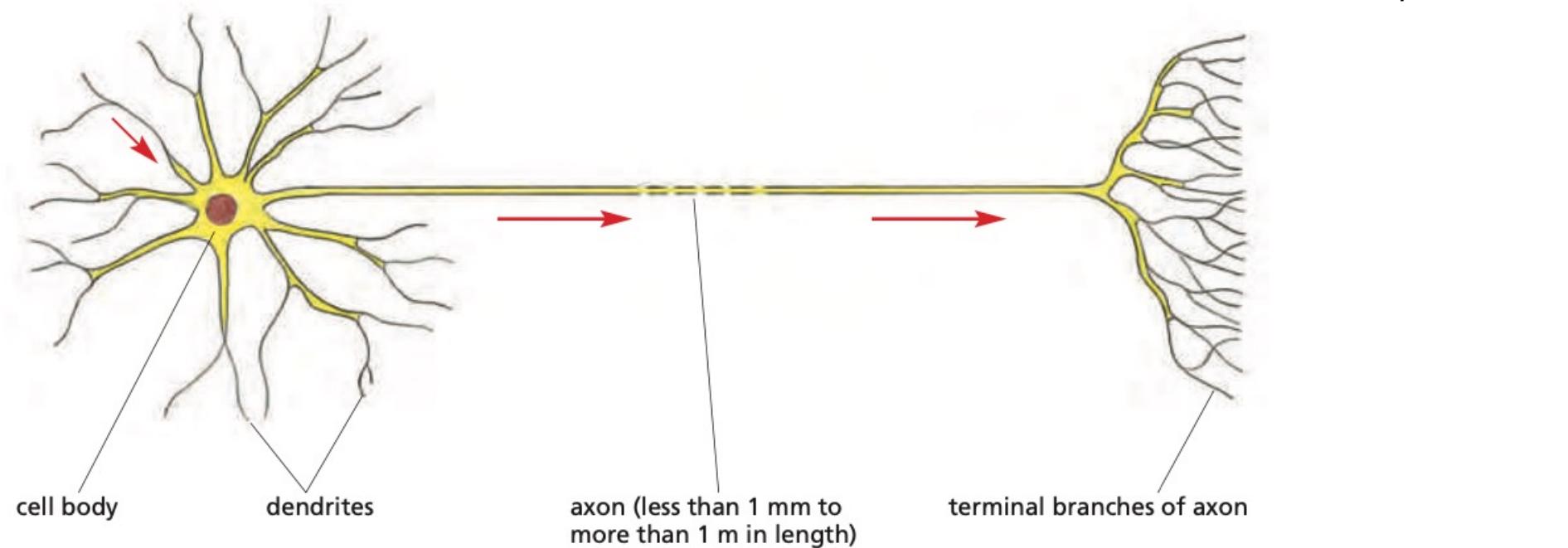
# $K^+$ specificity of the selectivity filter in a $K^+$ channel.



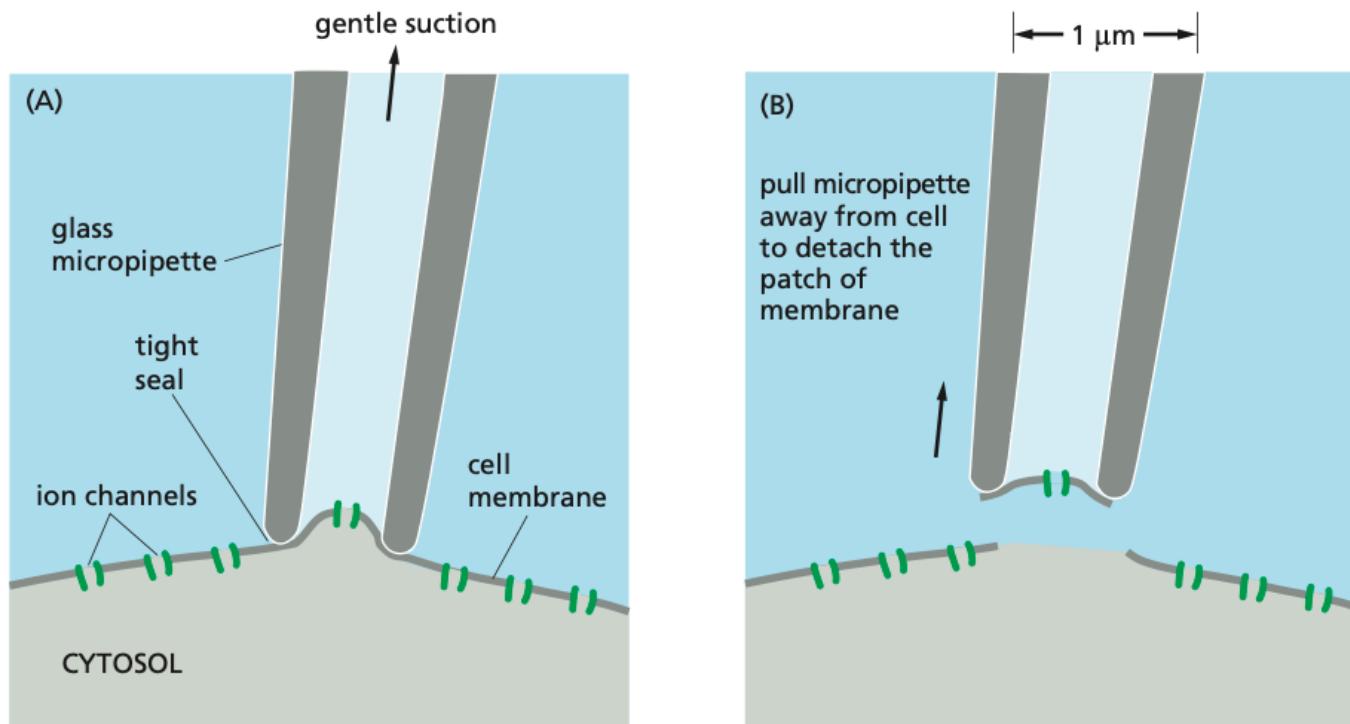
$1s^2 2s^2 2p^6 3s^1$        $1s^2 2s^2 2p^6 3s^2 3p^6 4s^1$



# Importance of ion channels in neurons



# The technique of patch-clamp recording



**Figure 11–34** The technique of patch-clamp recording. Because of the extremely tight seal between the micropipette and the membrane, current can enter or leave the micropipette only by passing through the ion channels in the *patch* of membrane covering its tip. The term *clamp* is used because an electronic device is employed to maintain, or “clamp,” the membrane potential at a set value while recording the ionic current through individual channels. The current through these channels can be recorded with the patch still attached to the rest of the cell, as in (A), or detached, as in (B). The advantage of the detached patch is that it is easy to alter the composition of the solution on either side of the membrane to test the effect of various solutes on channel behavior. A detached patch can also be produced with the opposite orientation, so that the cytoplasmic surface of the membrane faces the inside of the pipette.

# The technique of patch-clamp recording

## The Nobel Prize in Physiology or Medicine 1991



Erwin Neher



Bert Sakmann

The Nobel Prize in Physiology or Medicine 1991 was awarded jointly to Erwin Neher and Bert Sakmann *"for their discoveries concerning the function of single ion channels in cells"*

# The technique of patch-clamp recording

To record ionic current flow through individual channels, while membrane potential is clamped

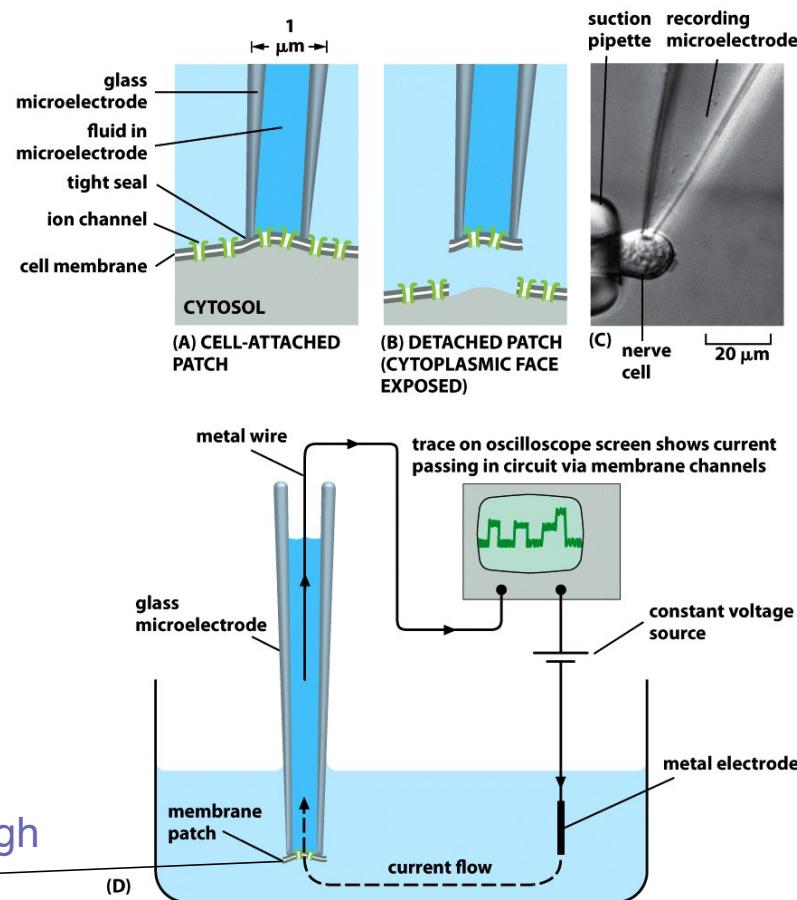


Figure 12-23 *Essential Cell Biology* (© Garland Science 2010)

# The technique of patch-clamp recording

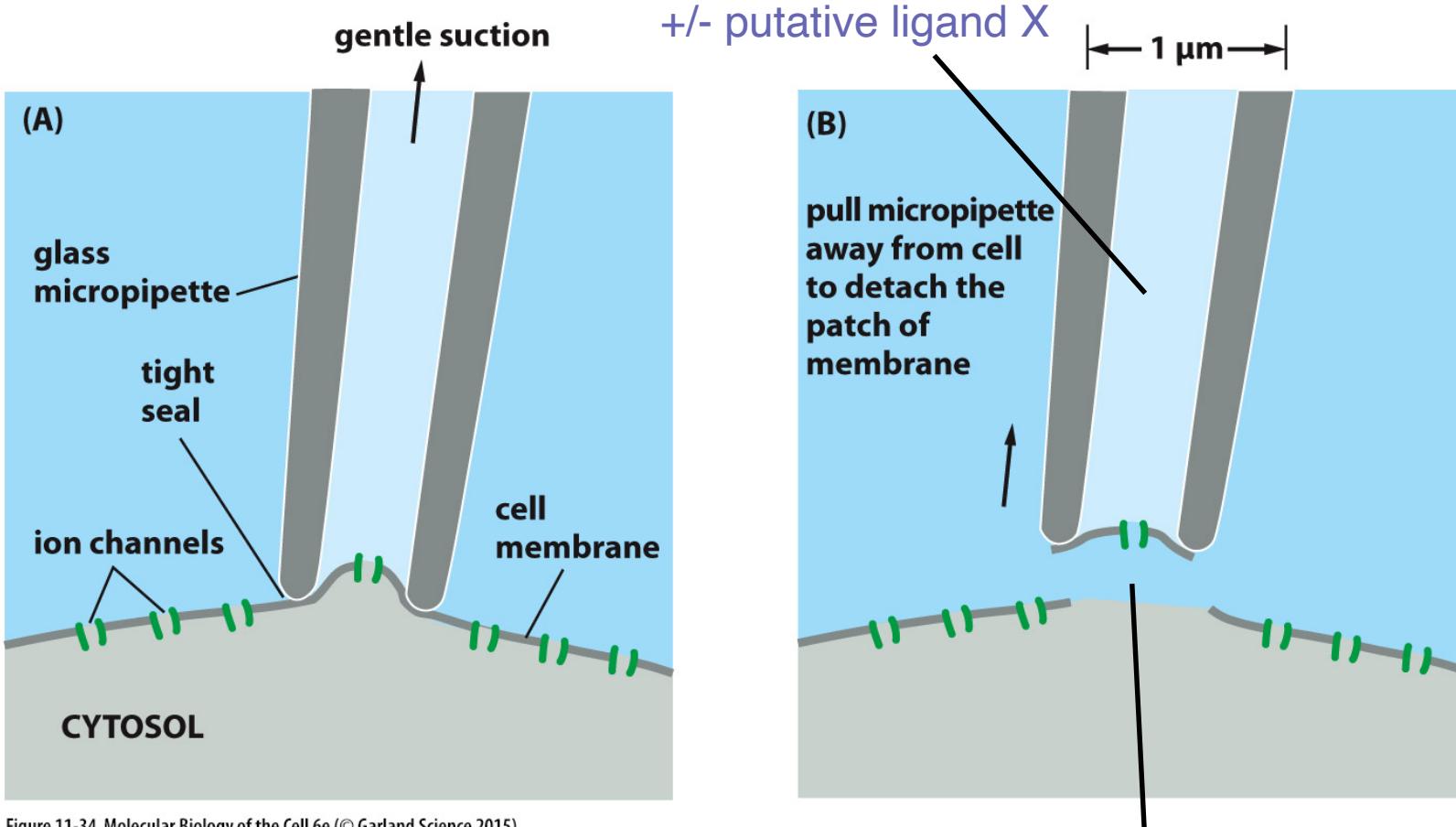
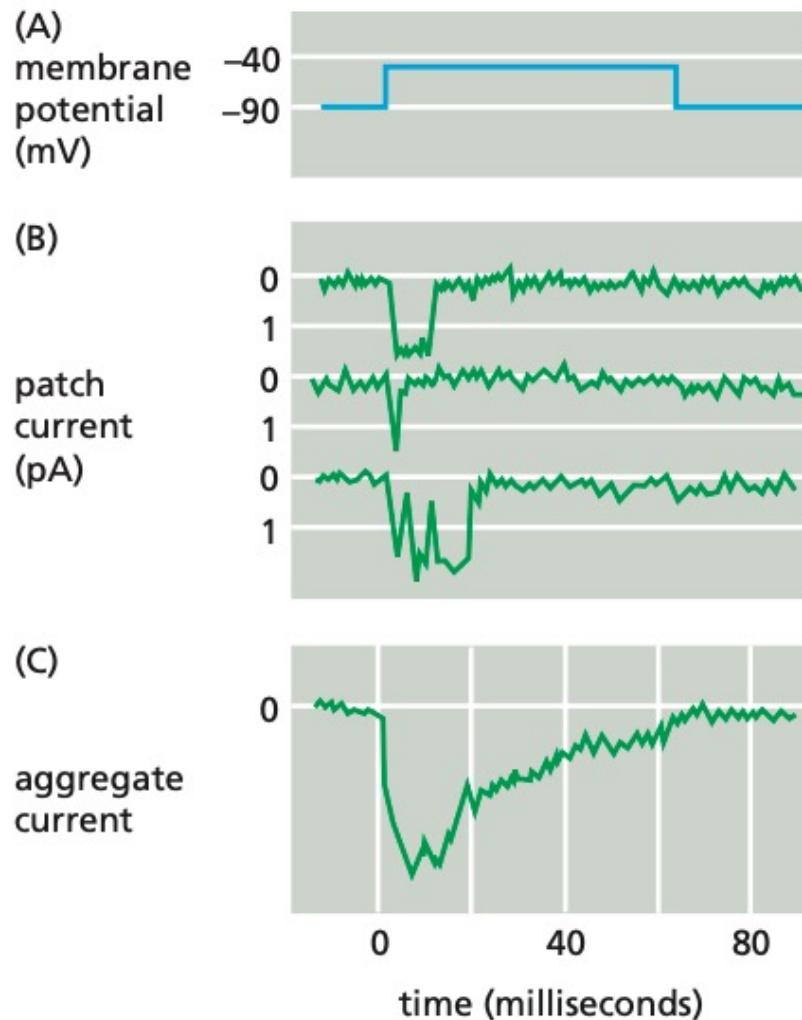


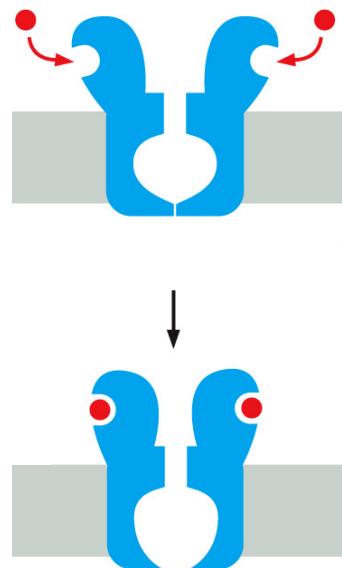
Figure 11-34 Molecular Biology of the Cell 6e (© Garland Science 2015)

The patch-clamp enables to determine which molecule(s) activate(s) the channel, and on which side

# Patch-clamp measurements for a single voltage-gated $\text{Na}^+$ channel



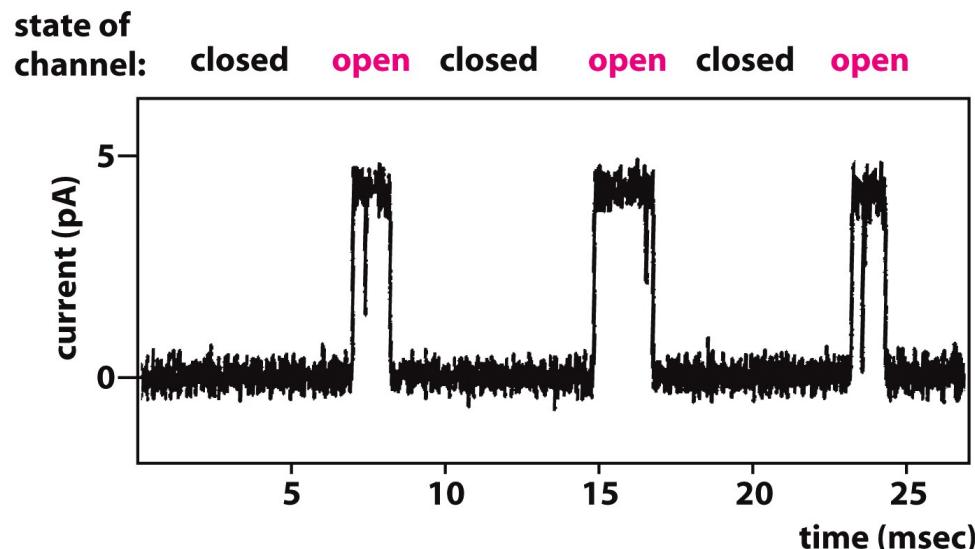
Ex: acetylcholine bound to an ion channel in muscle cells



ligand-gated  
(extracellular  
ligand)

and Science 2015)

Although acetylcholine is always present, the channel does not stay open



When open, the channel shows the same conductance

Figure 12-24 *Essential Cell Biology* (© Garland Science 2010)

# Done for the day!

Proteins involved in membrane transport

Active vs Passive transport

ATP-driven pumps and  $\text{Na}^+/\text{K}^+$  ATPase - ABC transporters

Channels

- Aquaporins

Introduction to ion channels

- Ion selectivity

- Ion current measurements

Examples of the importance of these transporters