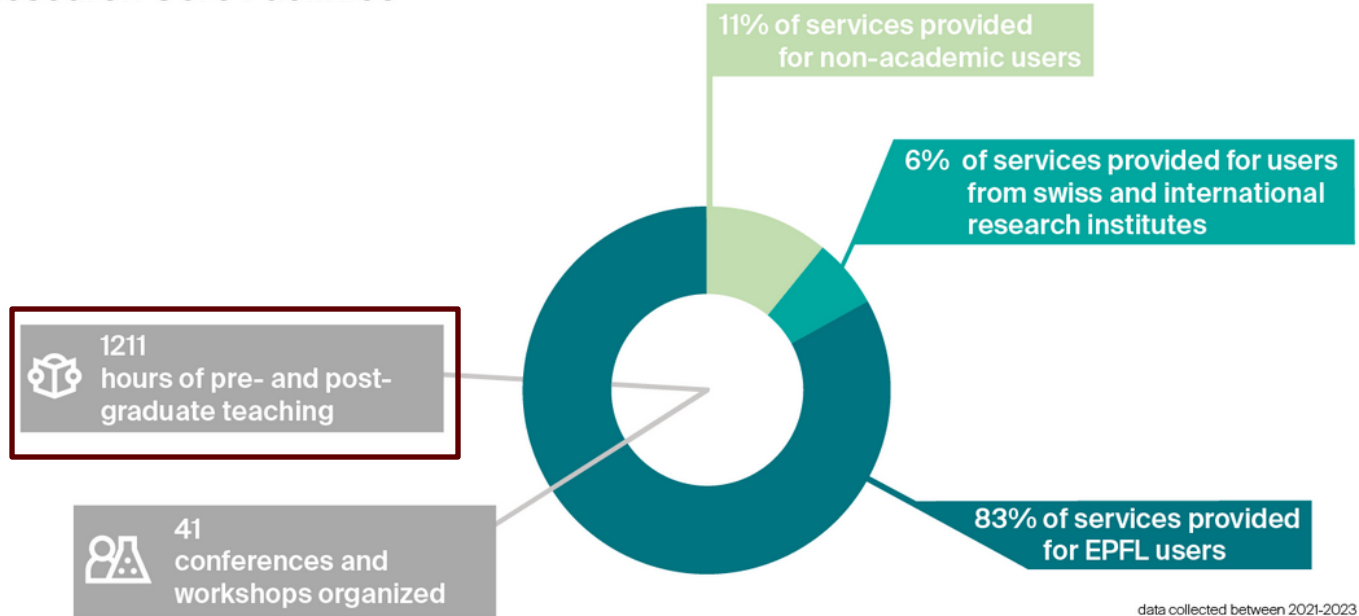


OMICS in Biomedical Research

BIOENG-519

11

Research Core Facilities



SV Research Core Facilities (RCF)



Head of RCFs
Roman Chrast



Gene Expression Core Facility

Head Bastien Mangeat



Proteomics Core Facility

Head Maria Pavlou



Flow Cytometry Core Facility

Head Miguel Garcia



Bioinformatics Competence Center

Head Christian Iseli



Protein Production and Structure Core Facility

Head Florence Pojer



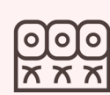
Bioimaging and Optics Facility

Head Arne Seitz



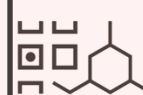
Biological Electron Microscopy Facility

Head Graham Knott



Histology Core Facility

Head Jessica Sordet



Biomolecular Screening Facility

Head Gerardo Turcatti



Bertarelli Foundation Gene Therapy Facility

Head Bernard Schneider



Bio-engineering and Organoids Facility


Head Gaspard Pardon






*Head of
RCFs*
**Roman
Chrast**




**Gene
Expression
Core Facility**


Head
**Bastien
Mangeat**




**Proteomics
Core Facility**


Head
**Maria
Pavlou**




**Flow
Cytometry
Core
Facility**


Head
**Miguel
Garcia**




**Bioinformatics
Competence
Center**

Head
**Christian
Iseli**




**Protein
Production
and
Structure
Core Facility**

Head
**Florence
Pojer**




**Bioimaging
and
Optics
Facility**

Head
**Arne
Seitz**




**Biological
Electron
Microscopy
Facility**

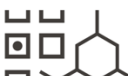
Head
**Graham
Knott**




**Histology
Core Facility**

Head
**Jessica
Sordet**




**Biomolecular
Screening
Facility**


Head
**Gerardo
Turcatti**




**Bertarelli
Foundation
Gene Therapy
Facility**

Head
**Bernard
Schneider**




**Bio-
engineering
and Organoids
Facility**

Head
**Gaspard
Pardon**

Course organization

- ✓ Course will be held in person (no Zoom - no recordings)
- ✓ Practicals are mandatory
- ✓ Course material will be available via Moodle
- ✓ Exam

40% - 4 platform rotations

Genes to proteins (PTPSP) - Evaluation: Participation during platform rotation, student presentations

Proteomics (PCF) - Evaluation: Participation during platform rotation, quiz

Flow cytometry (FCCF) - Evaluation: Participation during platform rotation, quiz

Gene expression (PTEG) - Evaluation: Participation during platform rotation, report

60% - written exam during official exam session (for the written exam, each of the covered topics will count 1/5)

- ✓ Contact for general course organization related questions: roman.chrast@epfl.ch

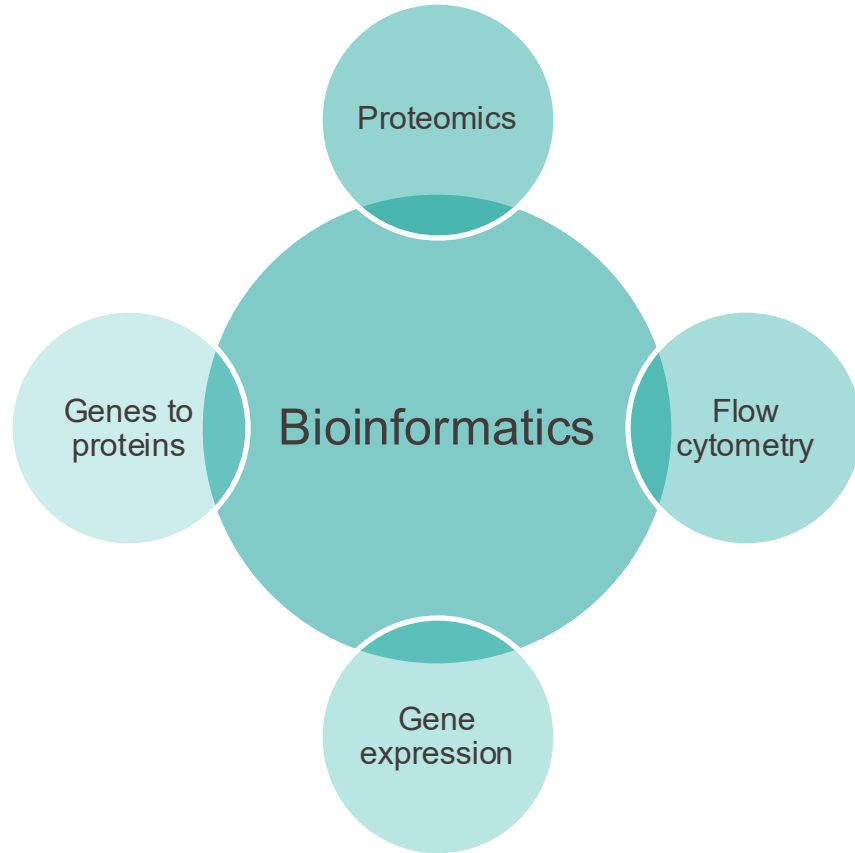
Course time-table

Date/Time	Sept 12	Sept 19	Sept 26	Oct 3	Oct 10	Oct 17	Oct 24	Oct 31	Nov 7	Nov 14	Nov 21	Nov 28	Dec 5	Dec 12	Dec 19	
Week	W1	W2	W3	W4	W5	W6		W7	W8	W9	W10	W11	W12	W13	W14	
8:15 - 9:00	Intro	Genes to	Flow	Hands-on at platforms	Hands-on at platforms	FREE	Hands-on at platforms	Hands-on at platforms	Hands-on at platforms	Hands-on at platforms	Hands-on at platforms	Hands-on at platforms	Hands-on at platforms	Hands-on at platforms	Hands-on at platforms	Bioinformatics
9:15 - 10:00		proteins	cytometry													
10:15 - 11:00		Proteomics	Gene													
11:15 - 12:00			expression													

CM1106

CM1106

EPFL Course description





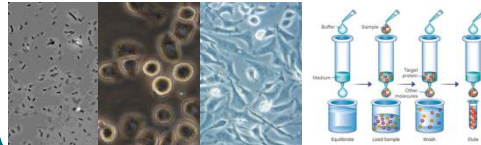
Genes
to
proteins
(PTPSP)

Lecture

Why are recombinant proteins important in science?

Strategies for production of recombinant proteins in different hosts

- Purification of proteins by affinity tags and other chromatography techniques
- State-of-the-art biophysical methods to ensure quality of purified proteins



Week 1: Purification of recombinant proteins by affinity

Week 2: Protein Quality control (QC) using biophysical techniques such as 2MP and DLS

Covered topics:

- Hands-on experience and expertise in the most advanced techniques used in the production and purification of recombinant proteins.
- Strategies to optimize protein production and ensure high-quality outputs.

Practicals



Proteomics
(PCF)

Lecture

Introduction in Proteomics

Mass Spectrometry-based
Proteomics Workflows

Week 1: In-gel digestion and basic
mass spectrometry principles

Week 2: Database search and
protein annotation principles

Covered topics:

- Bottom-up mass spectrometry-based proteomics (from sample preparation to data analysis)
- Mass spectrometer hands-on experience

Practicals



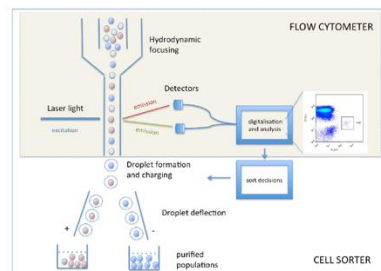
Flow
cytometry
(FCCF)

Lecture

Introduction to Flow Cytometry & Cell Sorting

How it works

Examples & applications



Schematic representation of a flow cytometer and a cell sorter

Week 1: Acquisition of stained cells on instruments

Week 2: Sorting demo & analysis +
Wrap up and discussion/conclusion

Covered topics:

- Instrument set up & QC
- Acquisition settings
- Data analysis and QC
- Standardization
- Cell Sorting

Practicals



Gene
expression
(PTEG)

Lecture

The NGS toolbox

Gene expression analysis (single cell and bulk)

Week 1: quantification and quality control of nucleic acids

Week 2: single cell transcriptomics

Covered topics:

- Methods for quantification of RNA/DNA
- Methods for QC of RNA/DNA
- 10X Genomics single cell transcriptomics experiment and downstream data QC

Practicals

Bioinformatics - Christian Iseli (EPFL) and Nicolas Guex (UNIL)



BICC

Lecture & Practicals

Week 1: Analysis of flow cytometry data

Week 2: Analysis of RNAseq data

Week 3: Analysis of proteomics data

Covered topics:

- Mission of a bioinformatic facility
- Experimental design and sample nomenclature
- Protein, transcript and genome reference databases, nomenclature
- Biological function knowledge, protein and gene annotations
- Data QC, mapping, analysis and visualization
- Data clustering and its applications: theory and practice
- Data formats and handling
- Variant calling, peak calling, quantification, GSEA
- Evaluation of genetic mutation impact

What about you?

Session ID:
bio519polling



SCAN ME

Group exercise

- ✓ Your group is working on a new blood disease that affects one type of white blood cell: the eosinophils. All other white blood cell types (neutrophils, basophils, monocytes, T cells, and B cells) are unaffected. A mouse model that mimics the human pathology is available, and your initial characterization of this model showed that eosinophils in adult mutant mice (2 months old) exhibit an increased accumulation of eosinophil granules.
- ✓ To develop a therapy for the disease, your goal is to characterize and correct the observed phenotype (increased accumulation of eosinophil granules).
- ✓ Think about (45–60 min):
 - Your strategy (since your company has limited resources, consider factors like project cost, duration and risk mitigation)
 - The model / samples you will need
 - The methods you will use to analyse the collected samples (preferably omics approaches)
 - How you will analyse the generated data (data integration, bioinformatics etc)
- ✓ Prepare a 10-15 minute presentation to pitch your project to a panel of investors who will decide whether to fund it.

Ground rules:

- No internet research allowed
- When working on your strategy, you can ask questions to anyone in the room
- You can use PowerPoint and/or white-board to present your approach
- You can decide who will be presenting your project (it can be 1 person or a group of persons)

Cellular structure, receptors and mediators of wild-type eosinophils

