

Scientific Literature Analysis In Neuroscience

EPFL

Preparation to the analysis of Part 2 “Cell type determination”

BIOENG-451

Academic year: 2025-2026

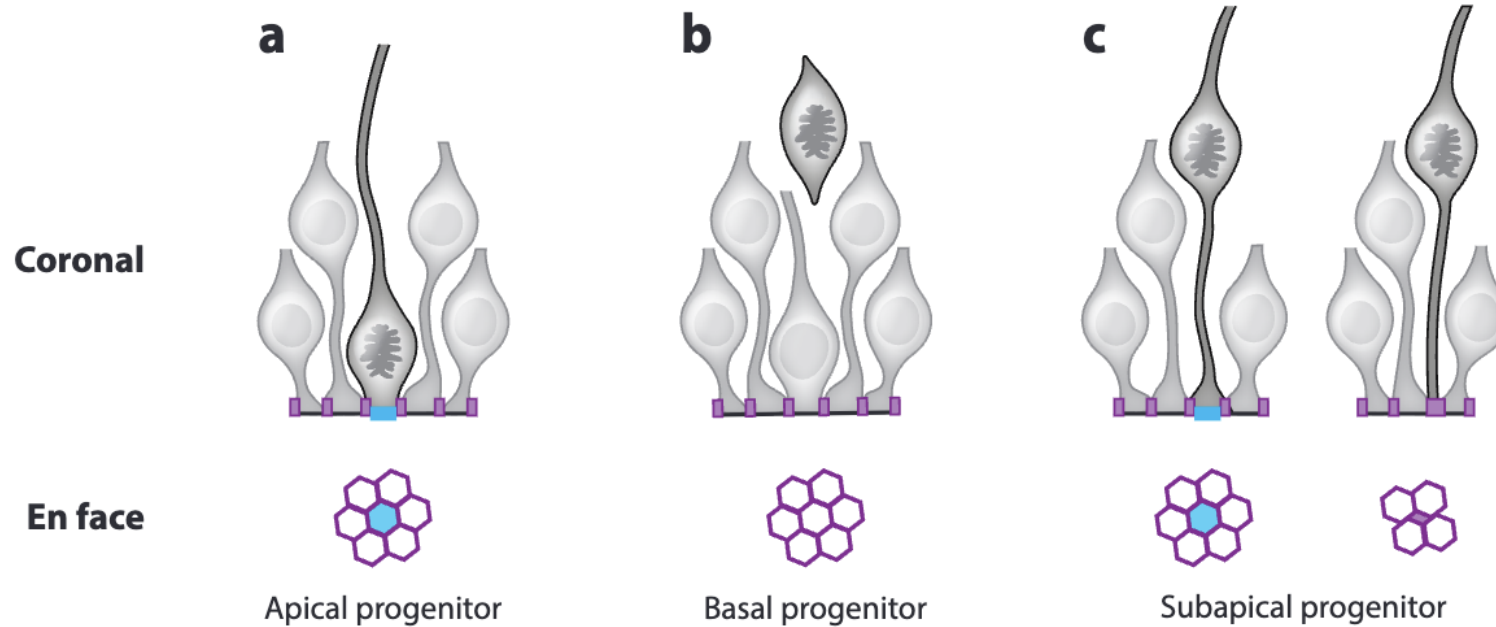
Teacher: Prof. Gioele La Manno

TA: Alessandro Valente

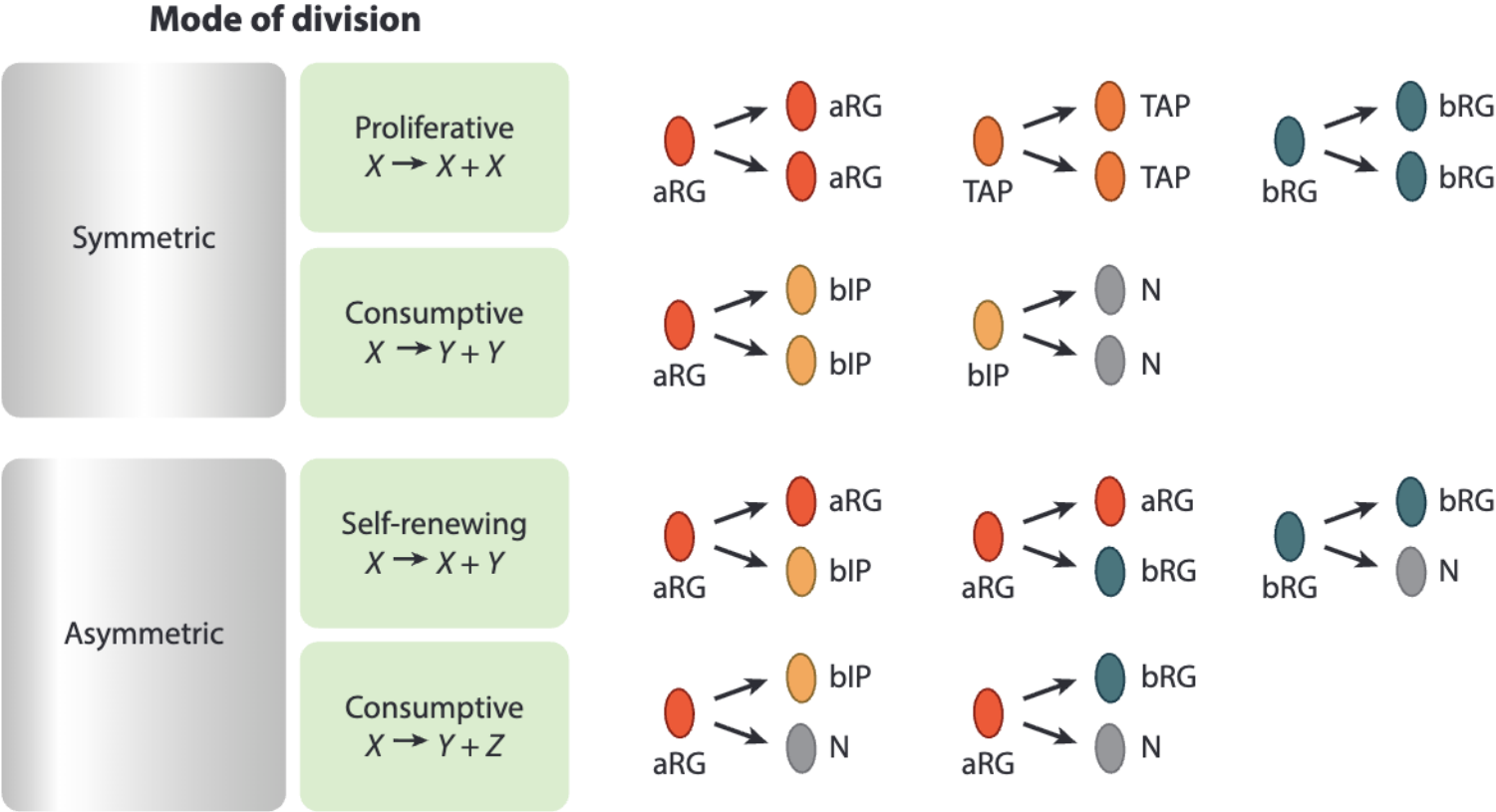
Neural stem and progenitor cell classification

Location of mitosis	Polarity	Proliferation capacity (round of cell division)	Cell type	
Basal	Bipolar	Multiple	Basal radial glia both processes	
	Monopolar		Basal radial glia basal process	
	Nonpolar		Basal radial glia apical process	
	Basal progenitors	Nonpolar	Multiple	Transient basal radial glia
			Single	Transit amplifying progenitor
	Basal progenitors	Nonpolar	Multiple	Basal intermediate progenitor
			Single	Basal intermediate progenitor
	Subapical progenitors	Bipolar or monopolar	Multiple	Subapical progenitor
	Apical progenitors	Bipolar	Single	Apical intermediate progenitor
			Multiple	Apical radial glia
Apical			Neuroepithelial cell	

Apical progenitors



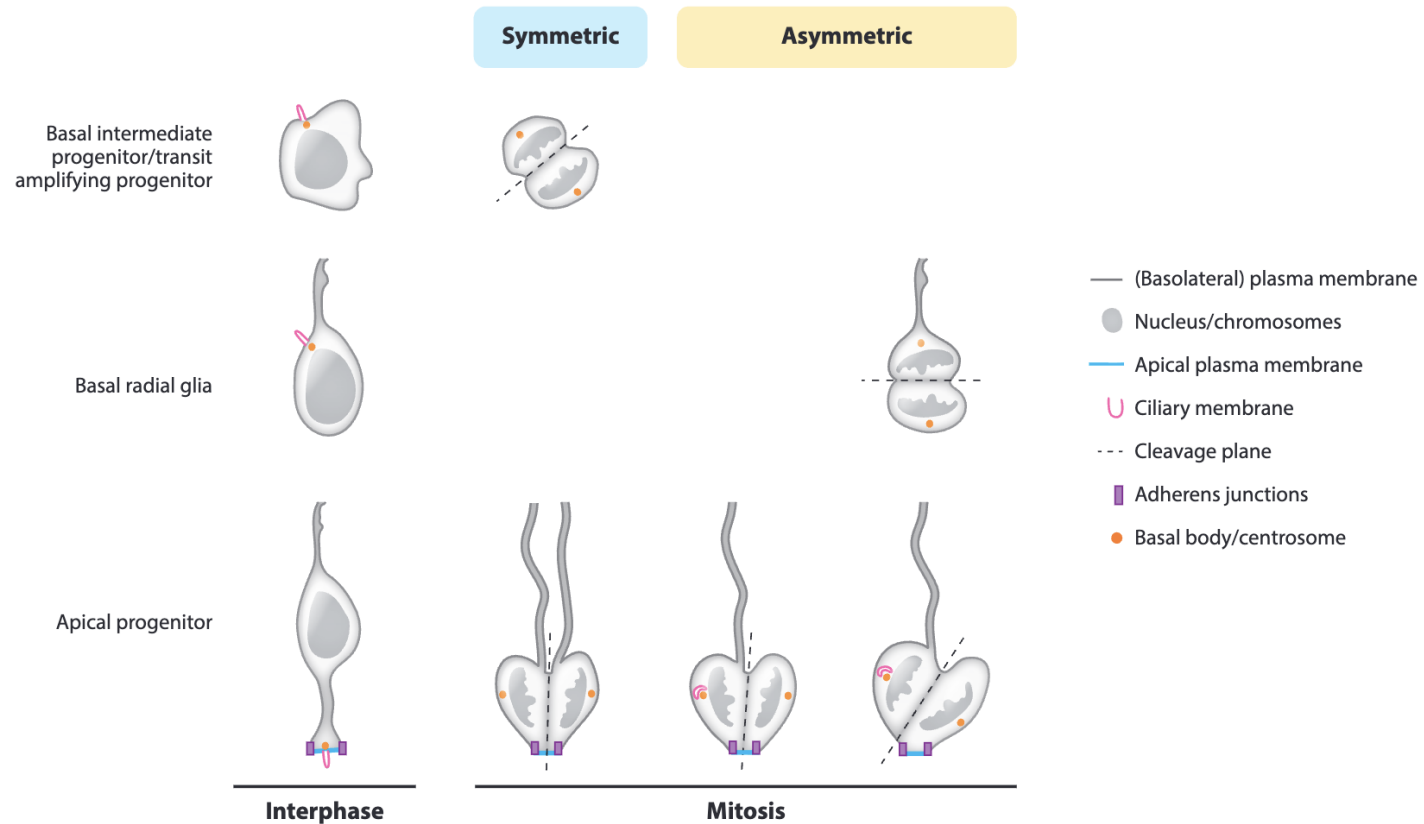
The various modes of cell division of neural stem and progenitor cells



Polarity cues are partitioned during mitosis

In stem and progenitor cells, polarity cues are represented by the apical plasma membrane (APs) the junctional complexes (APs)the primary cilium (APs and BPs), the centrosomes (APs and BPs), and the basal process (Aps and basal radial glial cells).

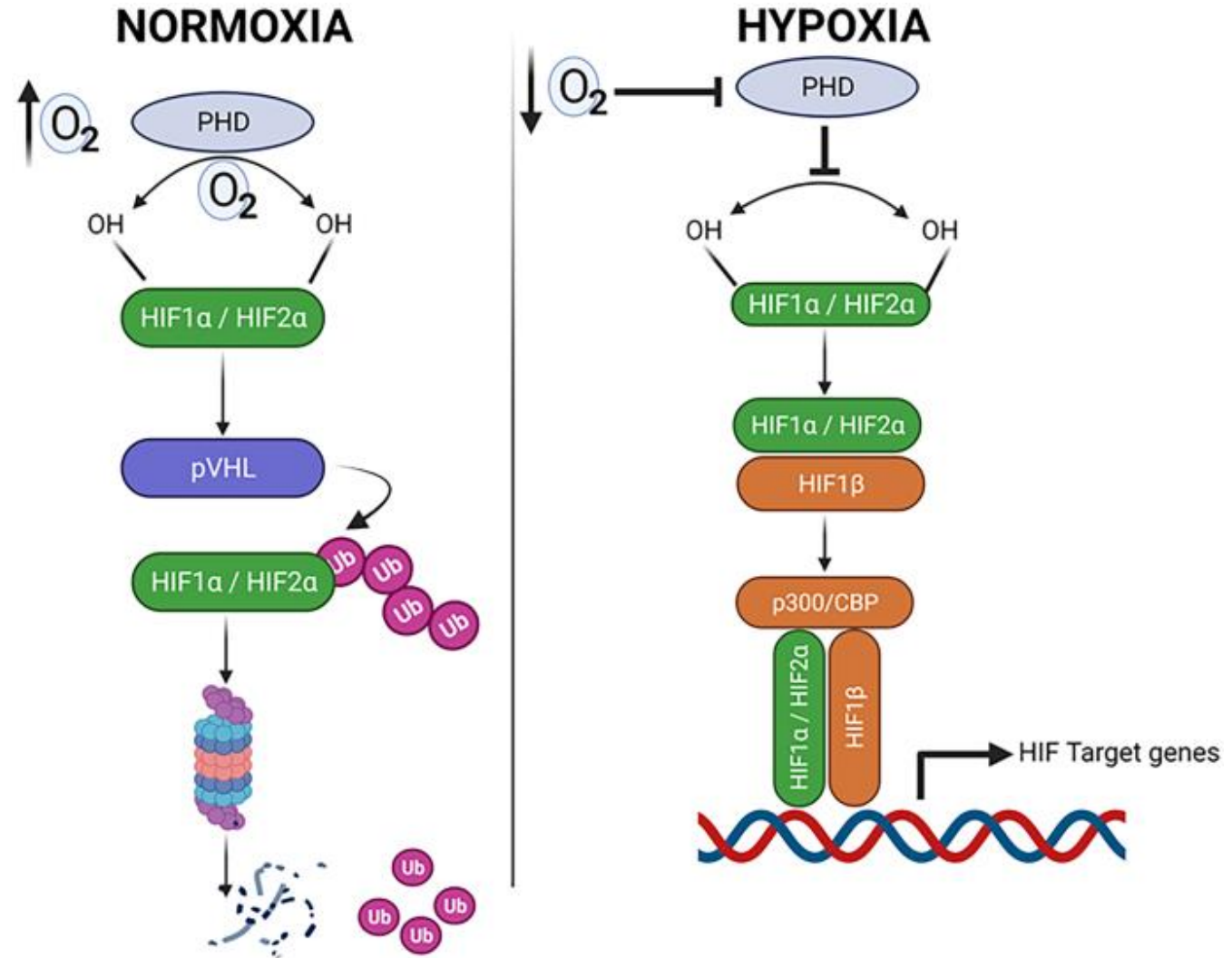
These cues are also present in interphase and can be asymmetrically or symmetrically partitioned during mitosis..



HIF 1alpha pathway

HIF-1 α is a transcription factor that mediates the cellular response to oxygen availability.

It allows cells to adapt to hypoxia by reprogramming metabolism, angiogenesis, and differentiation.



HIF 1alpha pathway

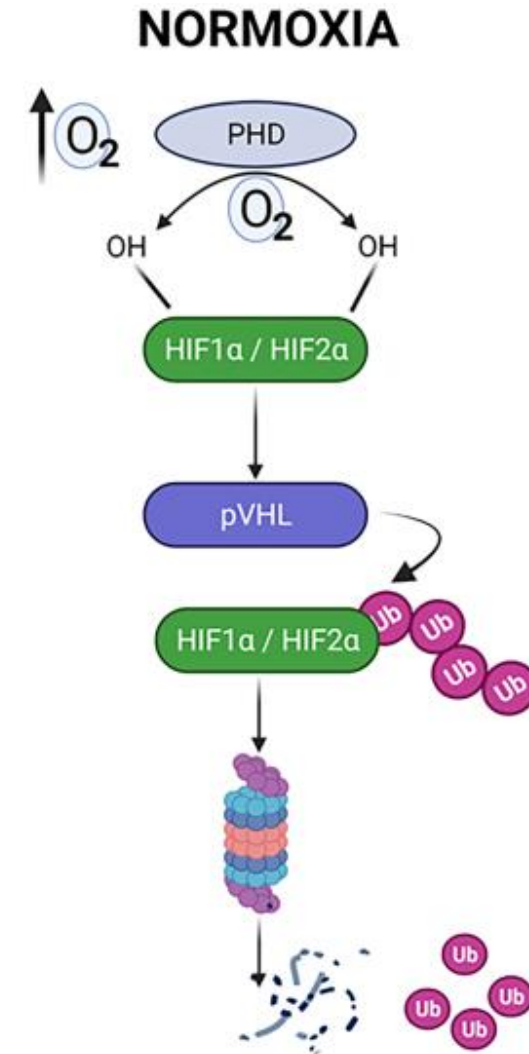
Normoxia (normal O₂):

HIF-1 α protein is continuously synthesized but **rapidly degraded**.

Prolyl hydroxylase domain enzymes (**PHD1-3**) hydroxylate specific proline residues on HIF-1 α using O₂ and α -ketoglutarate.

Hydroxylated HIF-1 α is recognized by **VHL (von Hippel-Lindau)** E3 ubiquitin ligase complex \rightarrow **ubiquitination** \rightarrow **proteasomal degradation**.

Thus, under normoxia, HIF-1 α is absent from the nucleus.



HIF 1alpha pathway

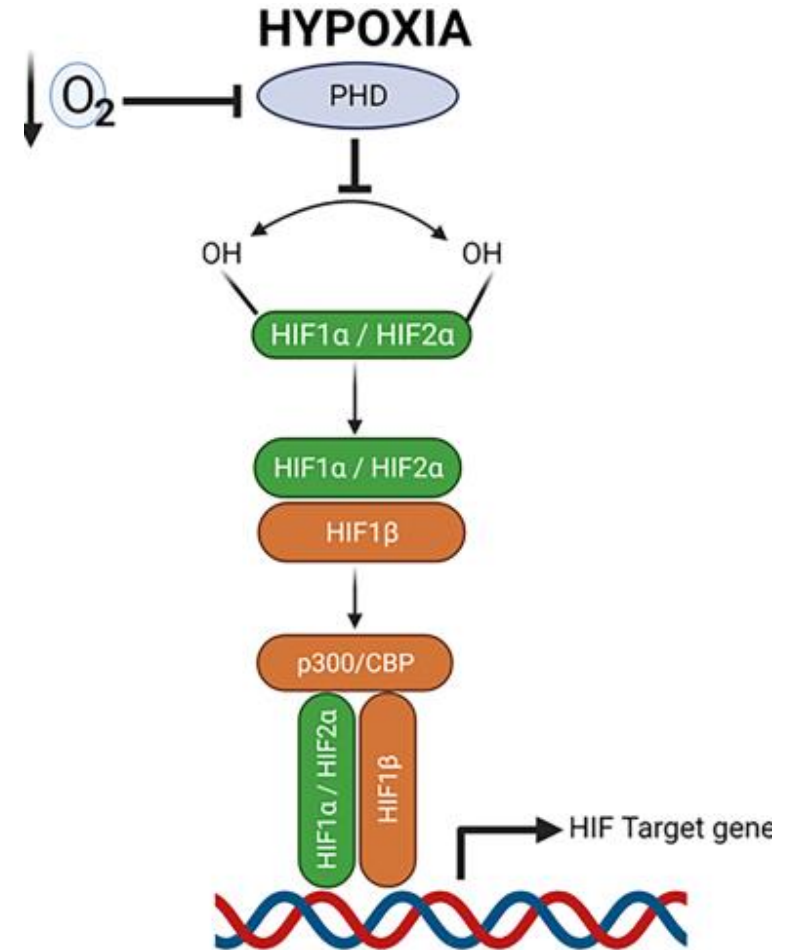
Hypoxia (low O₂):

PHD enzymes are inactive (need O₂).

HIF-1 α is **stabilized**, accumulates, and translocates to the **nucleus**.

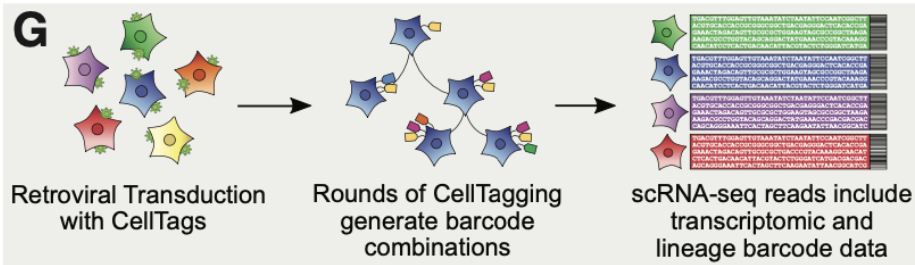
Dimerizes with **HIF-1 β (ARNT)** \rightarrow forms the active HIF-1 complex.

Binds **hypoxia response elements (HREs)** in target gene promoters \rightarrow transcriptional activation.



Single-cell Biology

2010s-



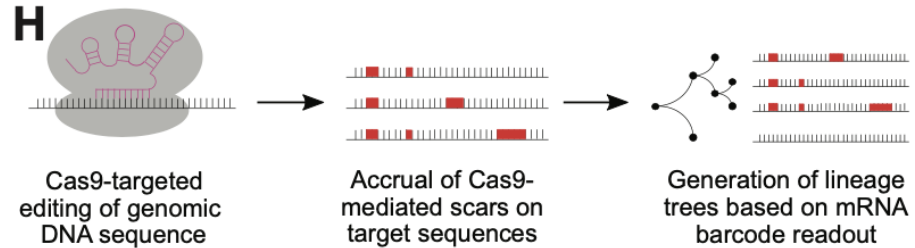
Single-cell

1000s -
10,000s of
cells

Resolved to clonal
and sub-clonal
populations

**Retroviral mRNA
Barcode Accrual**
Yao et al, 2017
Biddy et al, 2018
Weinreb et al, 2020

2010s-



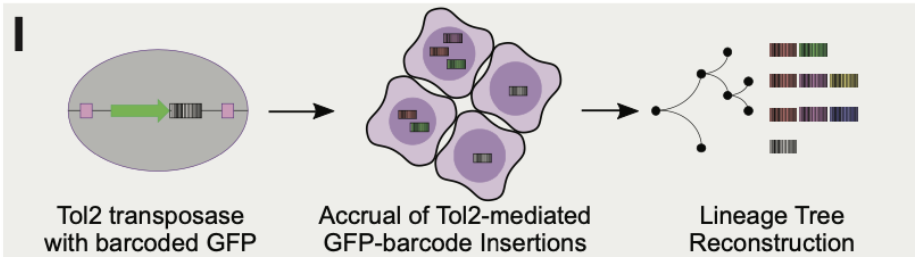
Single-cell

1000s -
10,000s of
cells

Information dropout
due to Cas9
induced deletion
of previous scars

Cas9 mRNA Scars
Spanjaard et al, 2018
Raj et al, 2018
Chan et al, 2019
Bowling et al, 2020

2010s-



Single-cell

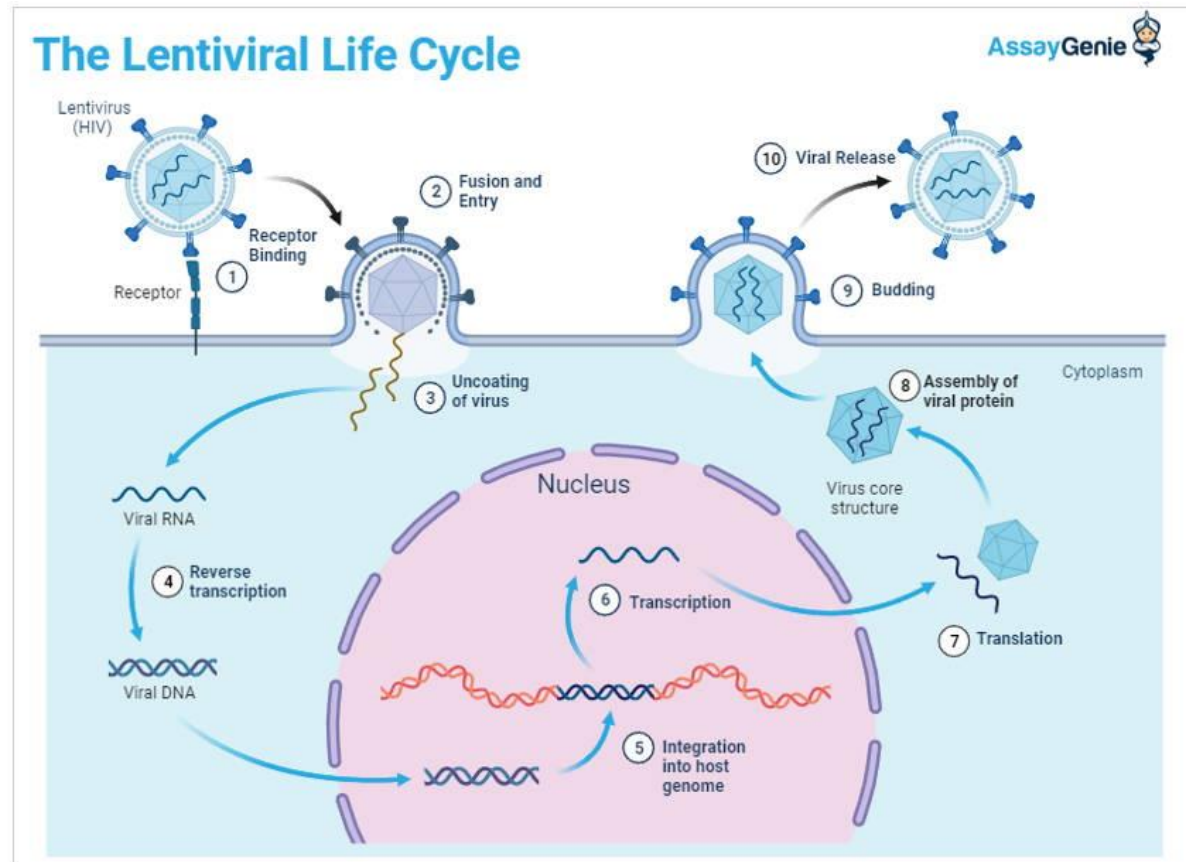
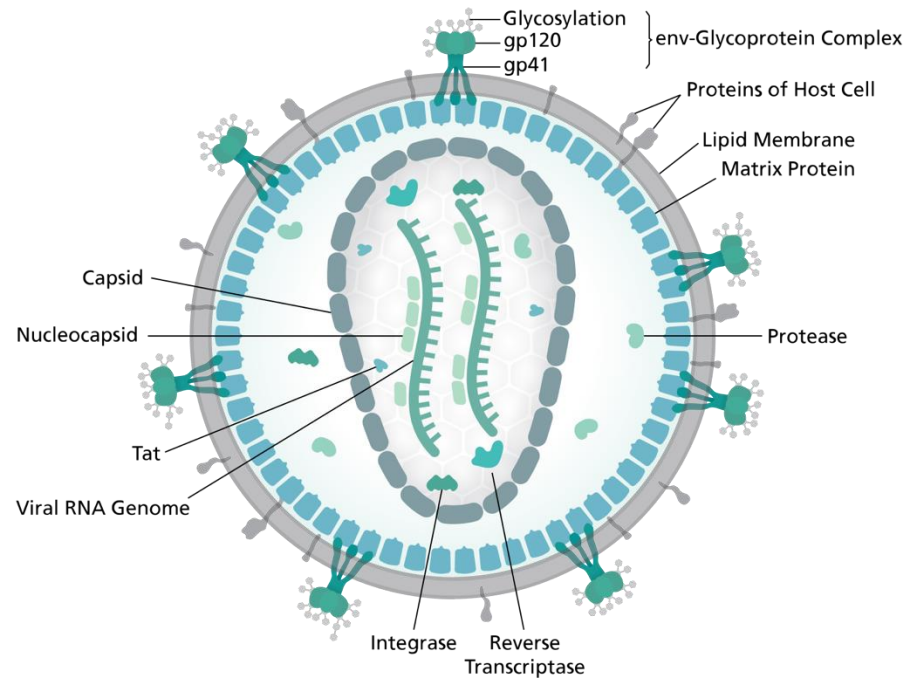
1000s -
10,000s of
cells

None beyond
usual scRNA-seq
transgene dropout

**Transposon
mRNA Barcode
Accrual**
Wagner et al, 2019

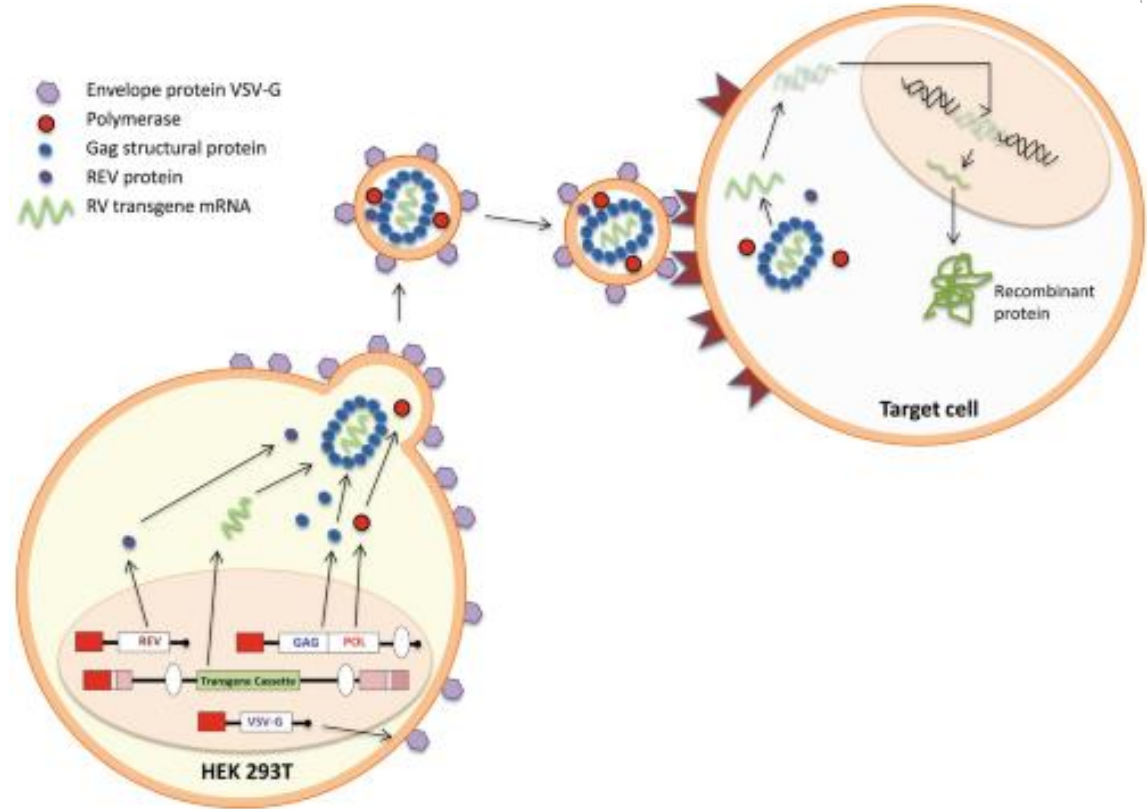
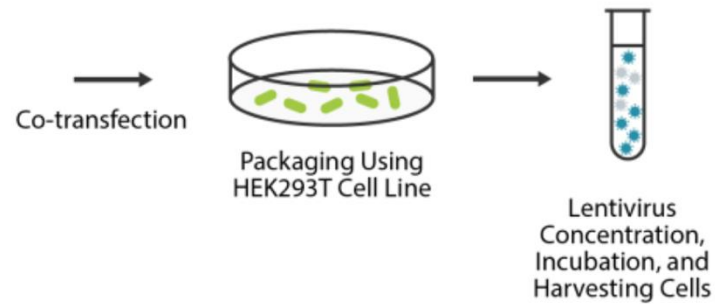
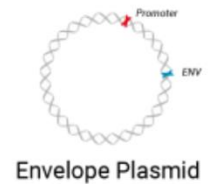
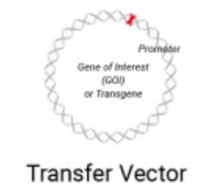
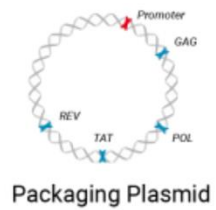
Lentiviral Libraries: Principle and Purpose

- Lentiviruses are modified retroviruses integrating into host genomes.
- Used for stable delivery of genes, reporters, or barcodes.
- Applications: molecular tagging, perturbation screens, lineage marking.



Lentiviral Libraries: How They Are Made

- Steps:
- plasmid backbone
- insert library
- bacterial amplification
- viral packaging in HEK293T
- titering.



Lentiviral Libraries: Experimental Logic

- Library = pool of constructs with randomized sequence (barcode or guide RNA).
- Infect target tissue at low multiplicity of infection (MOI).
- Each cell integrates a unique barcode → stable expression.
- Expression under chosen promoter (ubiquitous or specific).

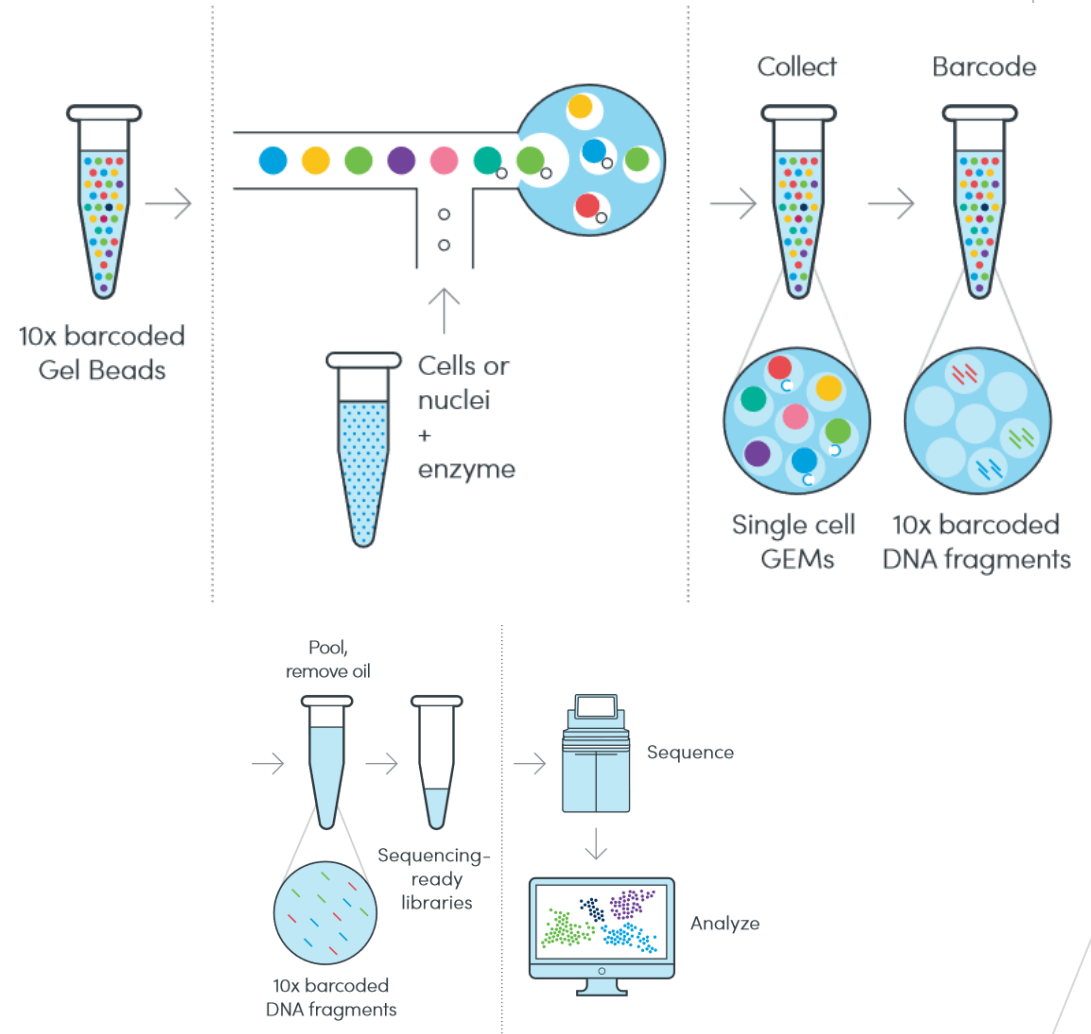
Critical points:

- MOI control: too high → multiple integrations, too low → unlabeled cells
- Integration site variability can affect gene expression
- Titering accuracy essential for reproducible transduction
- Potential for insertional mutagenesis in host genome

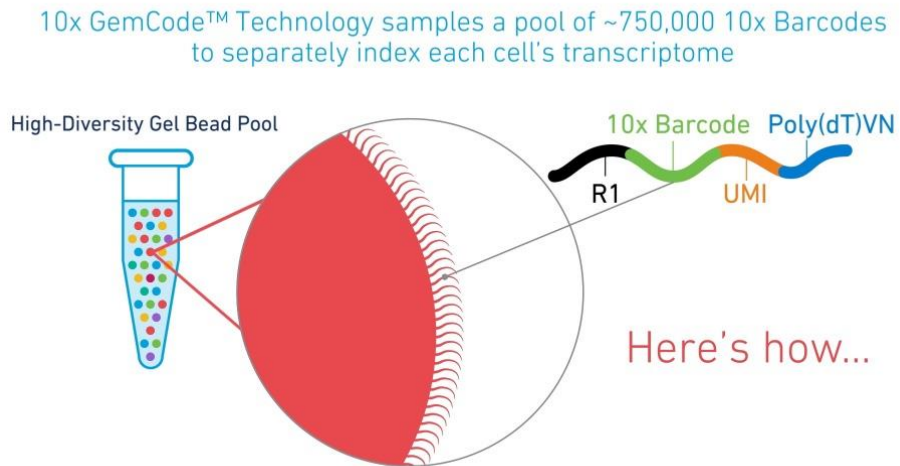
Single-cell RNA Sequencing (scRNA-seq)

Aim: Profile gene expression at individual cell resolution

- Isolates individual cells
- Captures and amplifies mRNA from each cell
- Sequences and quantifies transcripts
- Computational analysis for cell clustering and gene expression patterns



Single-cell RNA Sequencing (scRNA-seq)



Controls:

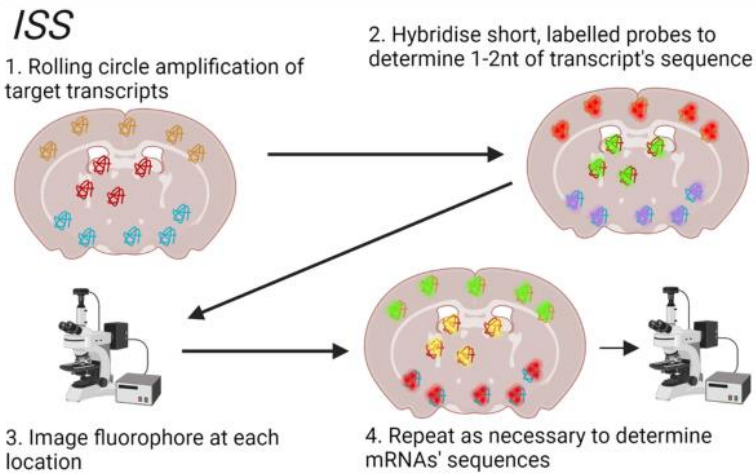
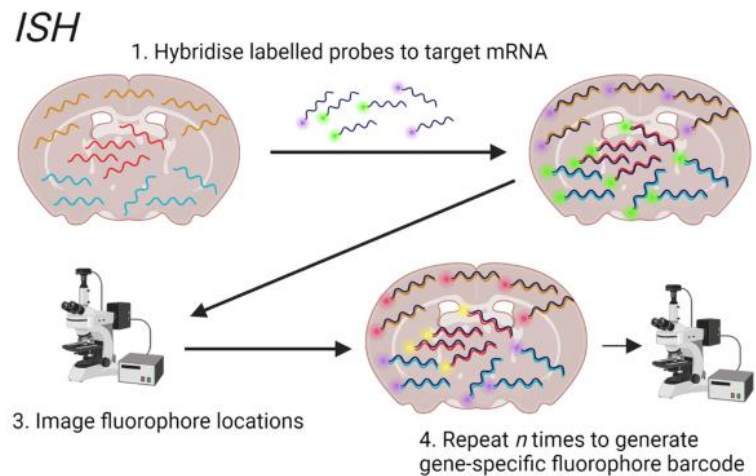
- Spike-in controls for technical variability
- Cell type-specific markers for validation

Critical points:

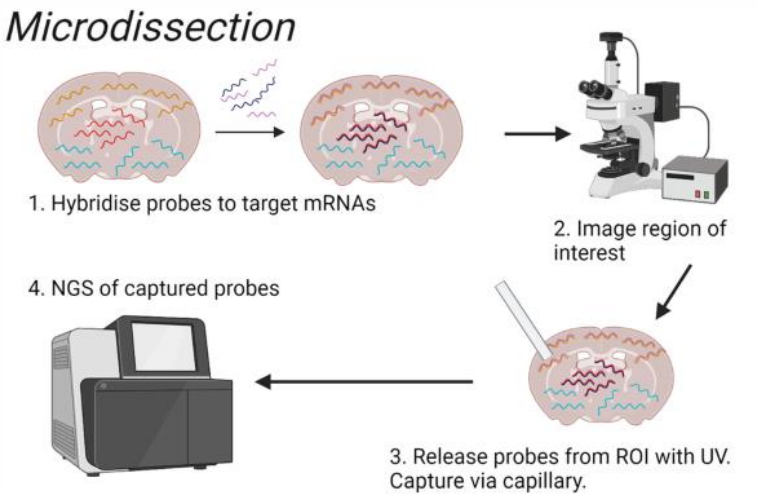
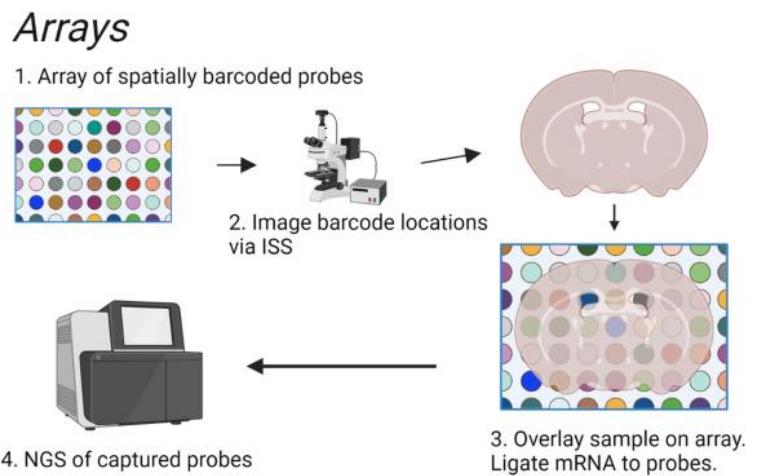
- Cell dissociation and viability
- Dropout events and technical noise
- Batch effects and data normalization

Spatial Transcriptomics - many methods, not one

Imaging methods

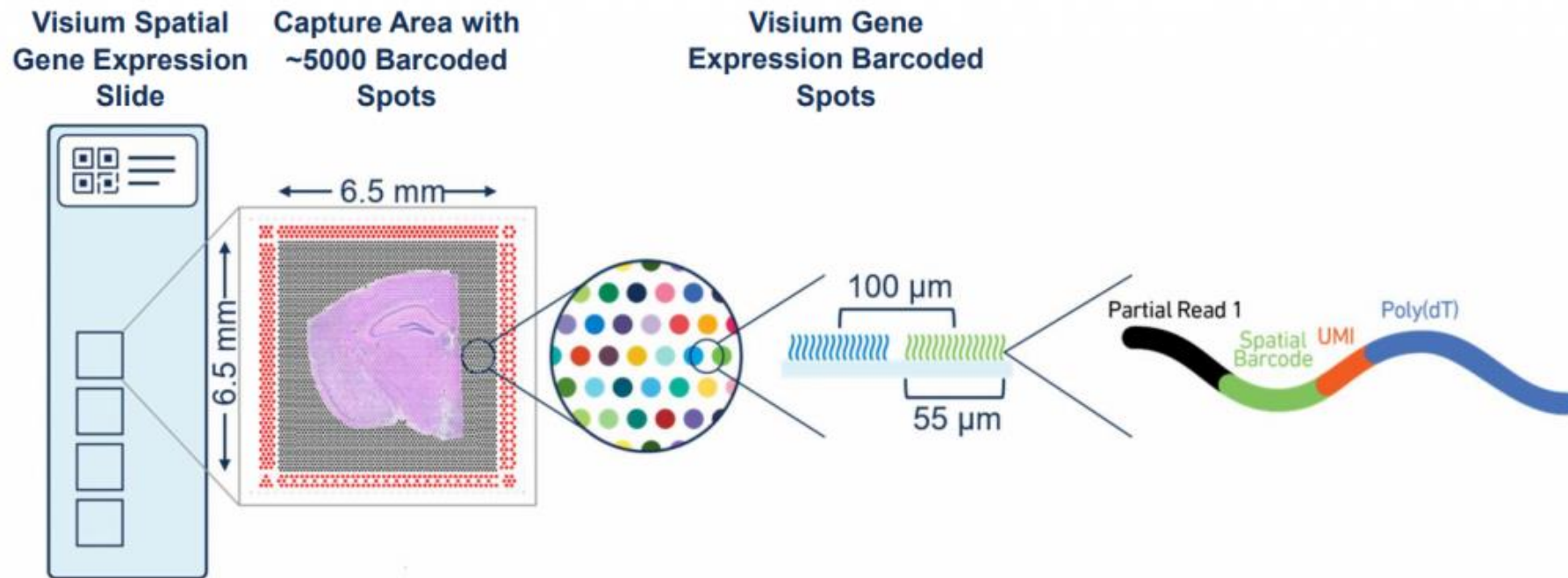


Sequencing methods



Spatial Transcriptomics: Principle

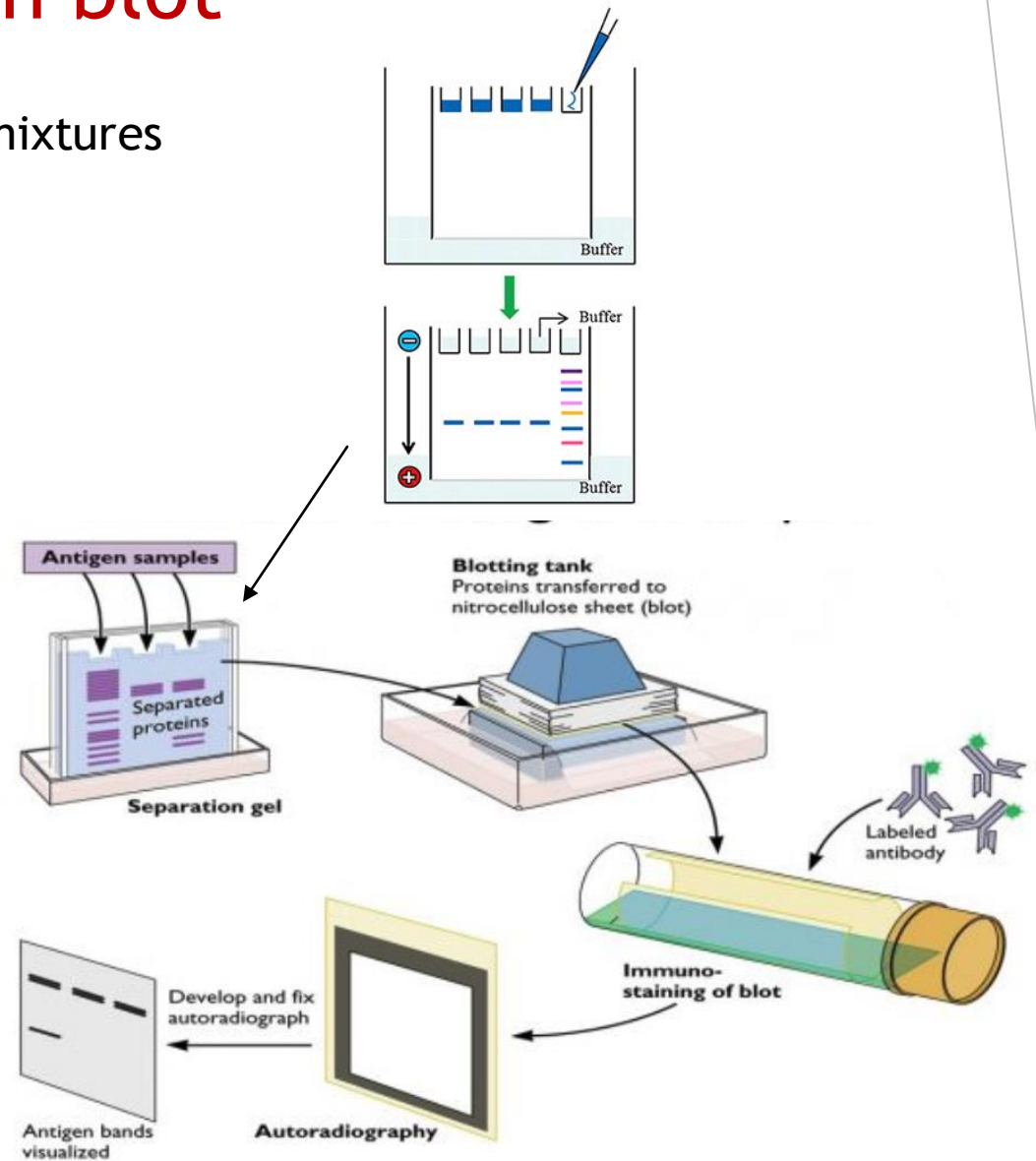
- Fresh-frozen tissue section placed on array of spatially barcoded oligos.
- mRNA hybridizes → reverse transcription → spatial barcodes captured in cDNA.
- Sequencing yields transcriptomes with coordinates.



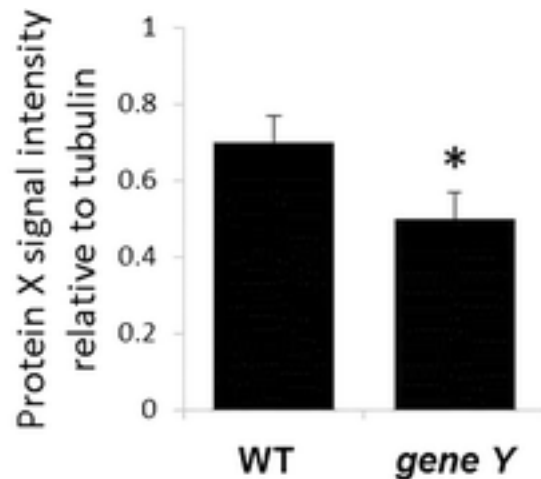
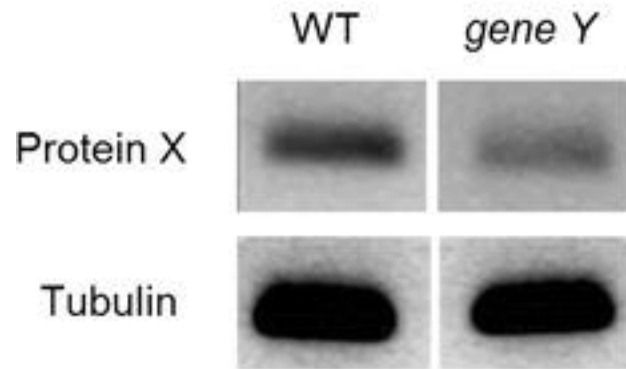
Western blot

Aim: Detect and quantify specific proteins in complex mixtures

- Separates proteins by size using gel electrophoresis.
- Transfers proteins to a membrane.
- Detects target proteins using specific antibodies.
- Visualizes using chemiluminescence or fluorescence.



Western blot



Controls:

- Loading control (e.g., housekeeping proteins)
- Positive and negative controls for antibody specificity

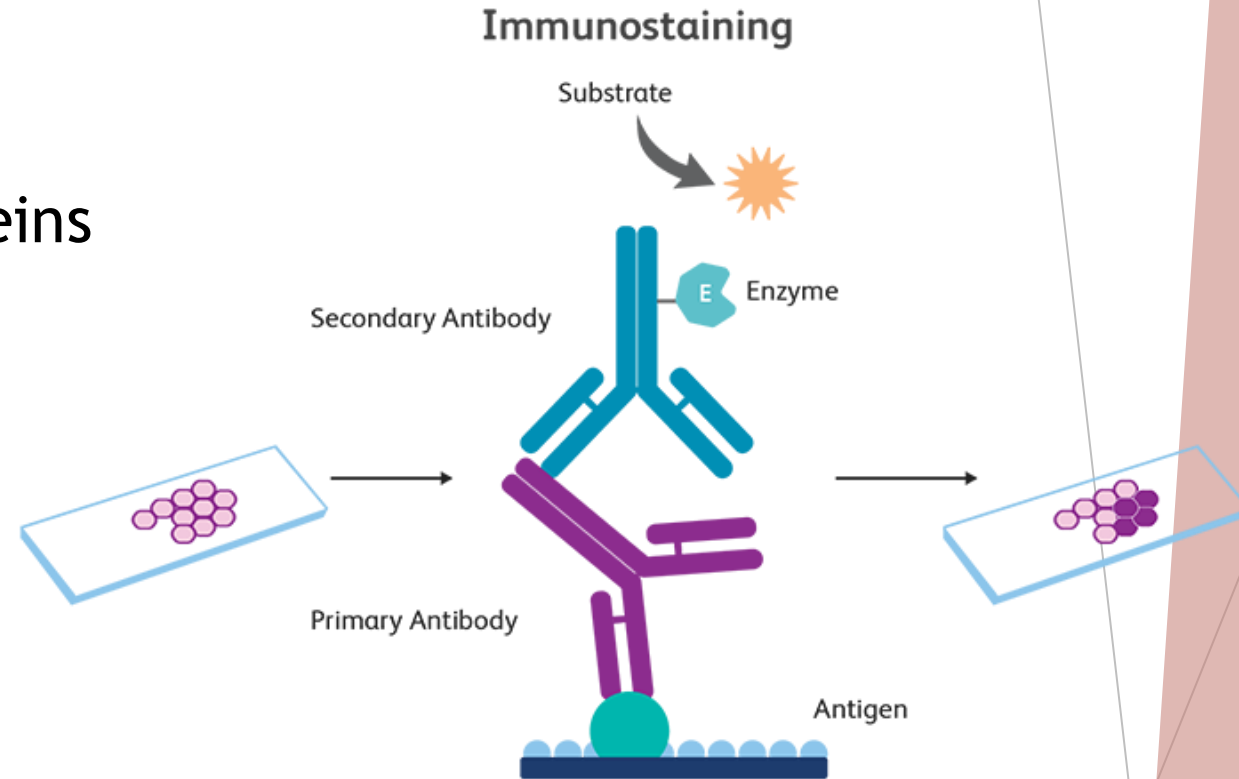
Critical points:

- Antibody specificity and validation
- Proper normalization for quantification
- Linear range of detection

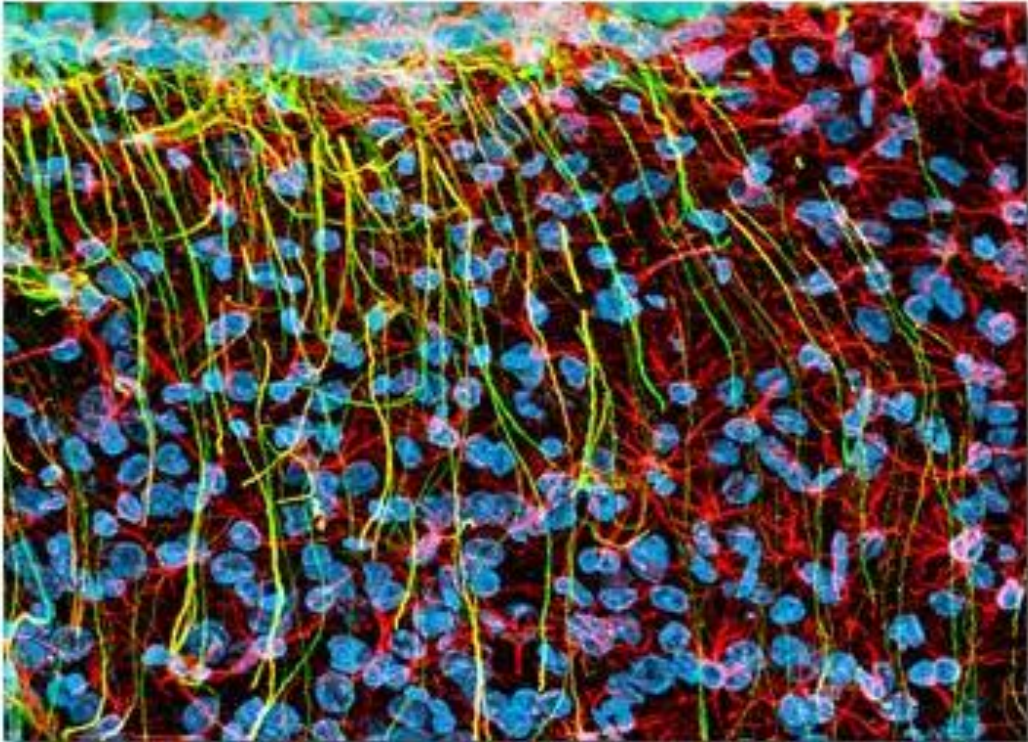
Immunostaining

Aim: Visualize and localize specific proteins within cells or tissues

- Uses antibodies to detect target proteins
- Primary antibody binds to protein of interest
- Secondary antibody (labeled) binds to primary antibody
- Fluorescent (or chromogenic) detection



Immunostaining



Controls:

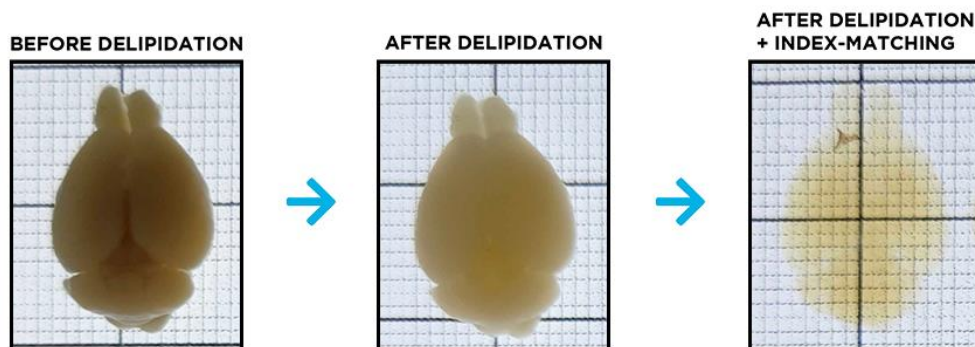
- Negative control (no primary antibody)
- Positive control (known expression pattern)

Critical points:

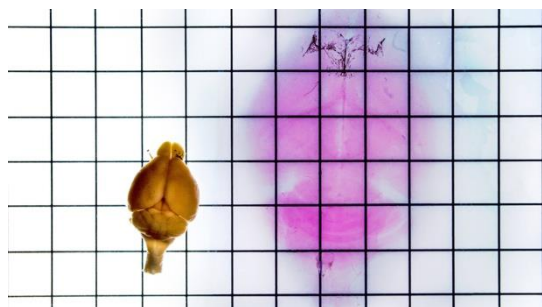
- Antibody specificity and validation
- Fixation and permeabilization methods
- Autofluorescence and background signal

Tissue clearing methods and light sheet imaging

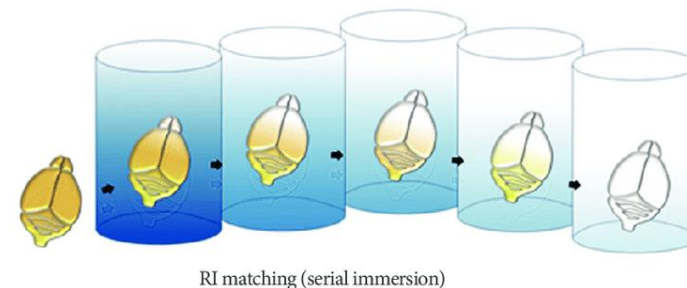
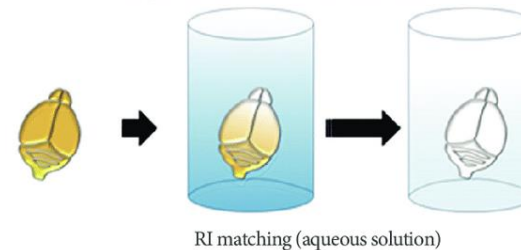
Aim: Visualize intact organs and large tissue volumes in three dimensions at cellular or subcellular resolution



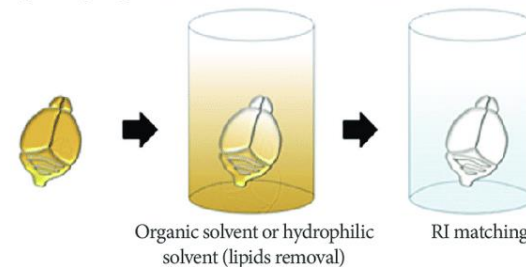
- Tissue clearing renders samples optically transparent by removing lipids and matching refractive index



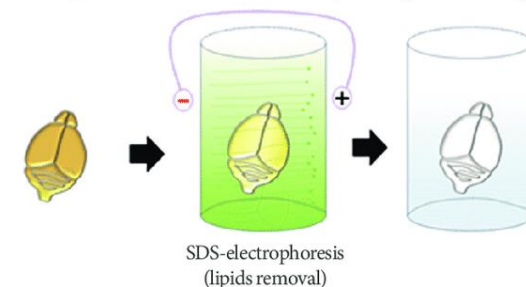
Simple immersion methods: *scale*, ClearT and SeeDB



Organic/hydrophilic solvent based methods: BABB, 3DISCO, iDISCO, and CUBIC

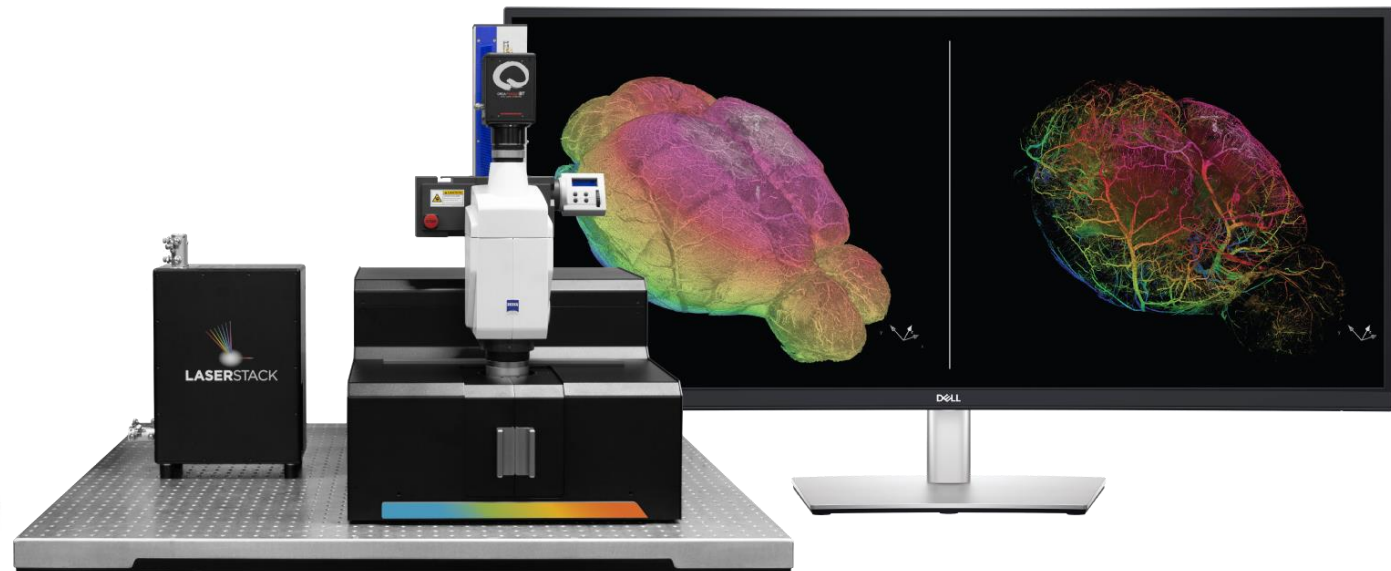
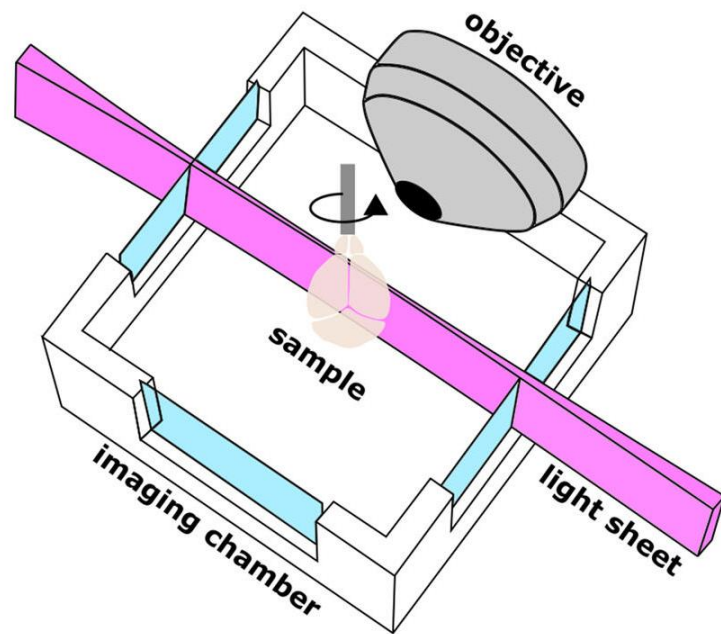


Polymer-based methods: CLARITY, PACT (passive clearing), and ACT



Tissue clearing methods and light sheet imaging

- Light sheet microscopy illuminates only the focal plane with a thin sheet of light perpendicular to the detection axis
- Enables rapid volumetric imaging of millimeter to centimeter-scale specimens



Tissue clearing methods and light sheet imaging - critical aspects

Clearing method compatibility: Does the method preserve the fluorophores or antibody epitopes used? Solvent-based methods quench many fluorescent proteins, while aqueous methods better preserve endogenous fluorescence.

Tissue distortion and shrinkage: Some protocols cause significant tissue expansion or shrinkage that affects spatial measurements and anatomical interpretation.

Imaging depth and resolution: Resolution degrades with depth due to light scattering and aberrations. Validate that quantitative analyses are performed within the reliable imaging depth range.

Controls and validation: Proper controls include uncleared tissue comparisons, antibody specificity checks, and validation of cell counts or morphology against established methods.

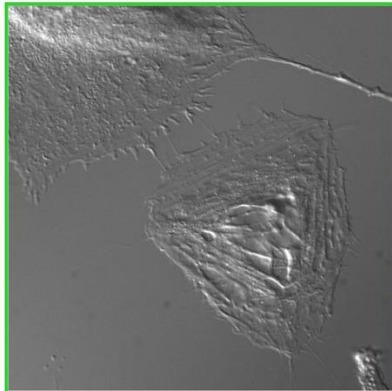
Computational processing: Image stitching, deconvolution, and cell segmentation algorithms can introduce artifacts. Check whether methods are described and validated, especially for automated quantification.

Live Imaging: Principle and Setup

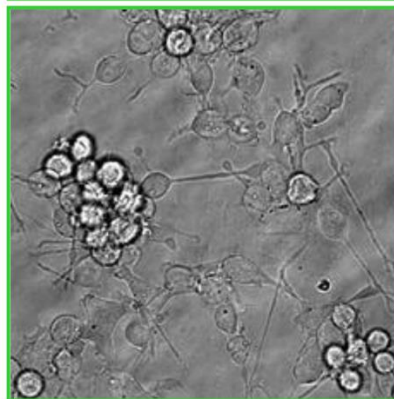
- Observes dynamic processes in real time.

Non-epifluorescence
techniques

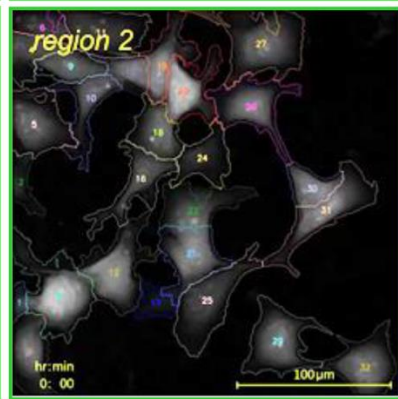
DIC



Phase Contrast

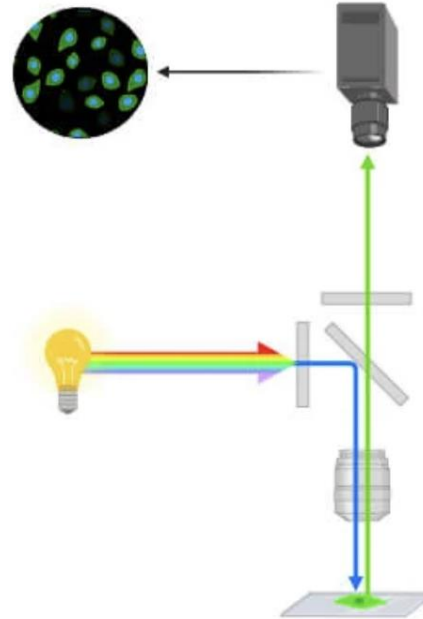


Brightfield

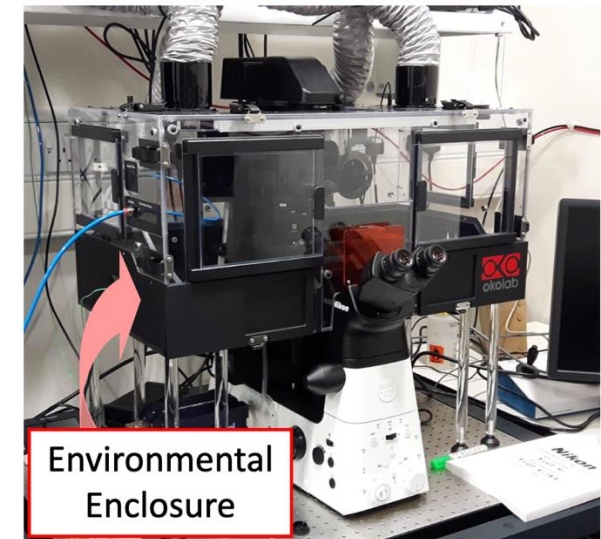


Phase Imaging

Epifluorescence

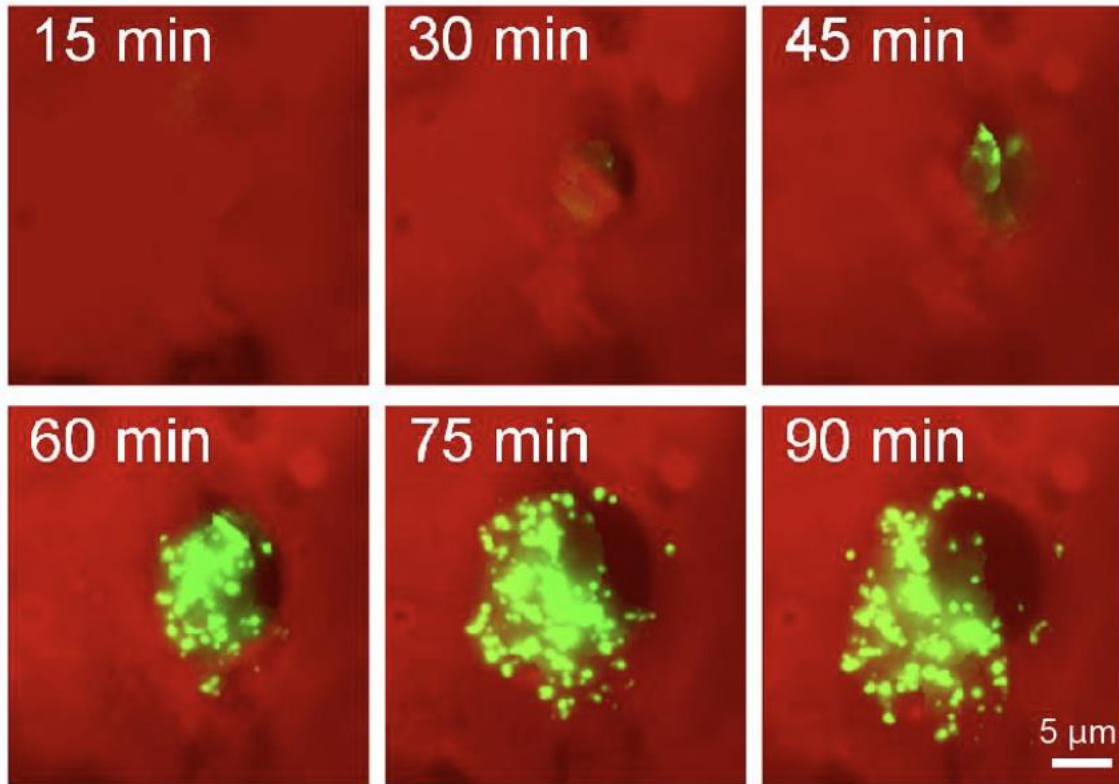


Enclosure

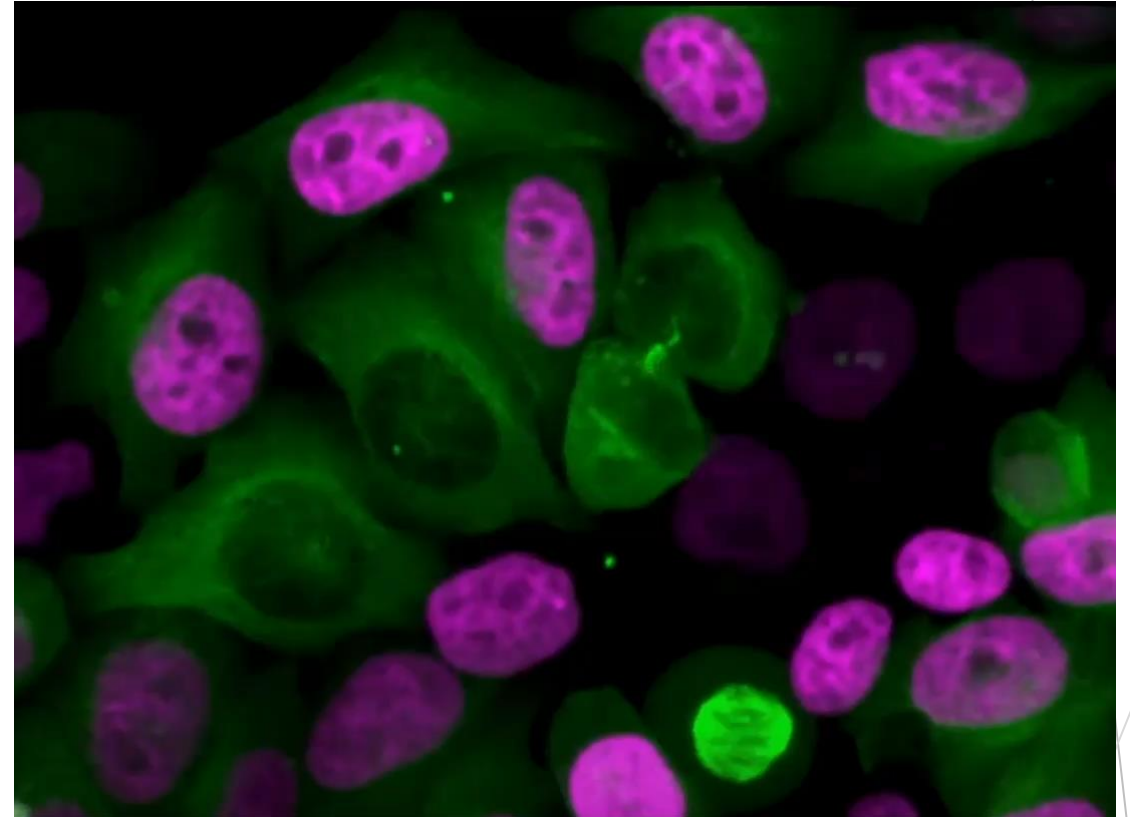


Live Imaging

Fluorescent reporters



Example



Live Imaging: Interpretation and Critical aspects

- Phototoxicity: excessive light exposure damages cells and alters behavior
- Photobleaching: fluorophore degradation limits observation time
- Environmental control: maintain temperature, CO₂, pH, and humidity
- Z-drift during long acquisitions affects tracking accuracy
- Out-of-plane movement can cause cell loss during tracking
- Reporter expression level can affect cell physiology

