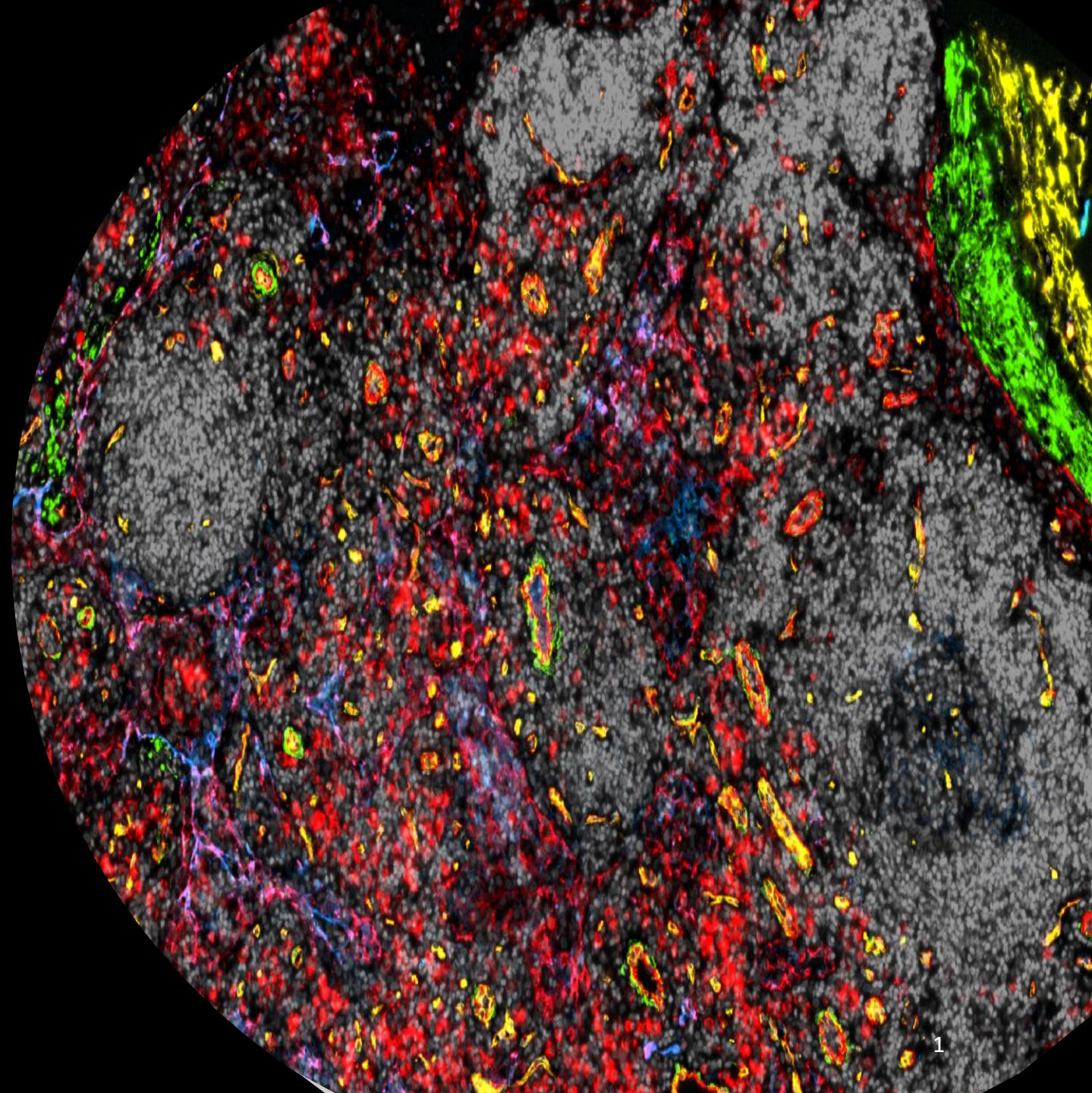


# Cancer Biology I

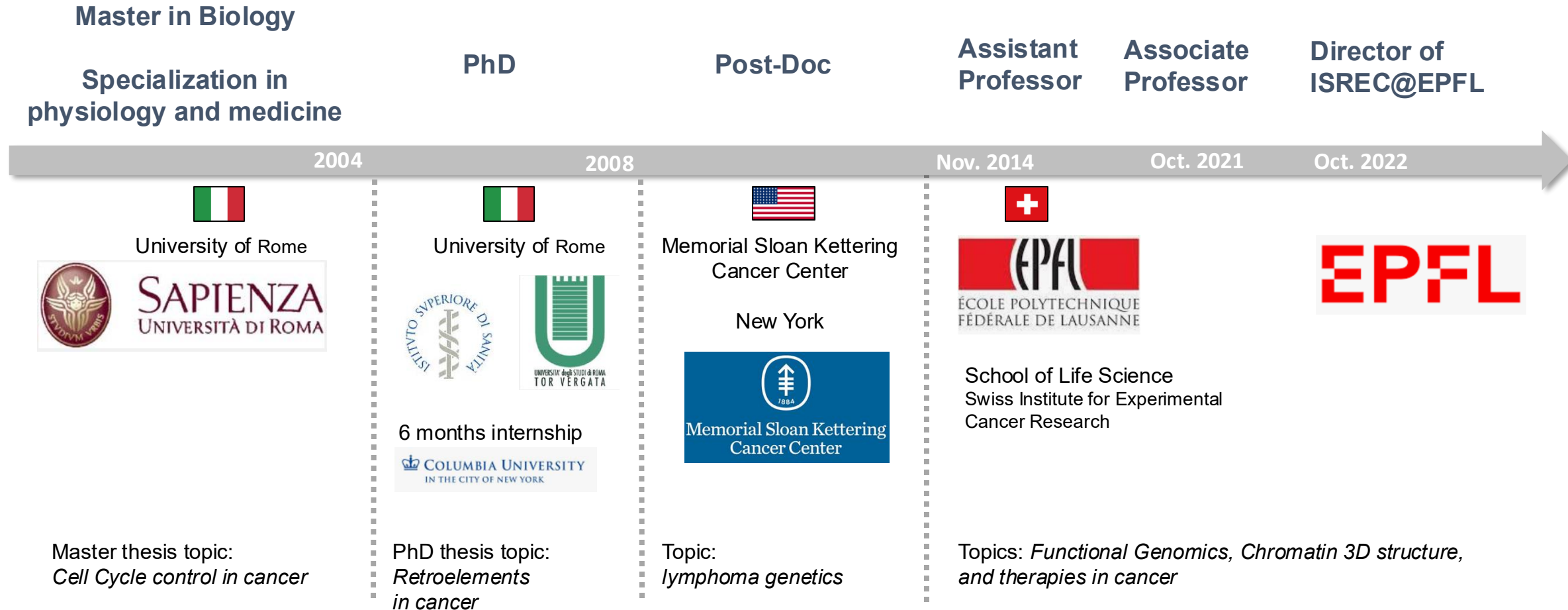
## Part-II

Fall semester 2024

Week 8



# My career path ...

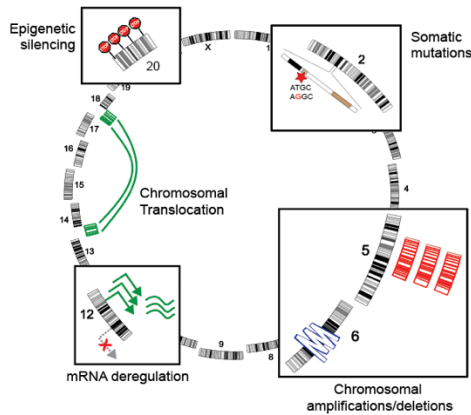


# Main research areas in Oricchio lab

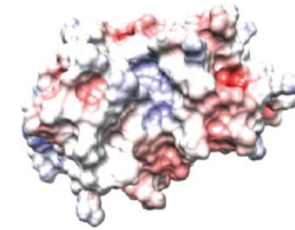
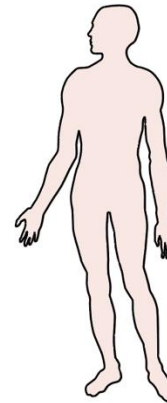
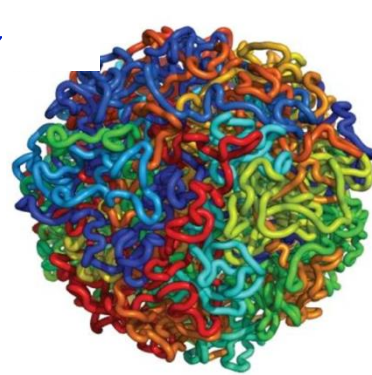
Areas of research



## Genomic alterations in cancer cells



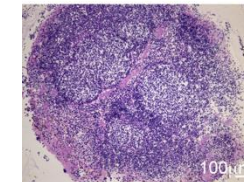
## Chromatin organization



*Target identification*



*therapy development*



*Different models to study response to therapies*

Zufferey, Taveranri et al. *Genome Biology* 2018,  
Donaldson, Sungalee, Zufferey, Tavernari et al. *Nature Genetics* 2019,  
Sungalee et al. *Nature Genetics* 2021,  
Liu et al. *Nature Communications* 2021,  
Lambuta, Nanni et al., *Nature* 2023

Battistello et al. *Blood* 2019,  
Dheilly, Battistello et al. *Cancer Cell* 2020,  
Petruzzella et al. *Nature Chem. Bio.* 2024,  
Santamaria-Martinez et al. *Nature Comm* 2024  
Bruand et al. *in revision*  
Epiney et al. *submitted*

# AGENDA

## Nov 3<sup>rd</sup>: Cancer genomics- mutations

Nov 10<sup>th</sup>: Cancer genomics-copy number alterations, heterogeneity, tumor evolution

Nov 17<sup>th</sup>: Cancer Epigenetics- chromatin 3D structure, cell plasticity

Nov 24<sup>th</sup>: – Major signaling pathways leading to cancer

Dec 1<sup>st</sup>: Cancer Therapies – chemo and targeted therapies

Dec 8<sup>th</sup>: Introduction to immunotherapies –

Dec 15<sup>th</sup>: Exam

(If it conflicts with another exam, it is possible to do the exam on Dec 17<sup>th</sup>)

Dec 17<sup>th</sup>: discussion of unclear points and career development discussion towards a PhD or not...(if the exam will be on Dec 17<sup>th</sup>)

# Exercise

- Two TAs will help in the discussion of the paper

*Alice Pellegrino*

*Dea Muller*

I provide a list of questions that can guide the discussion and help to understand key point of the paper

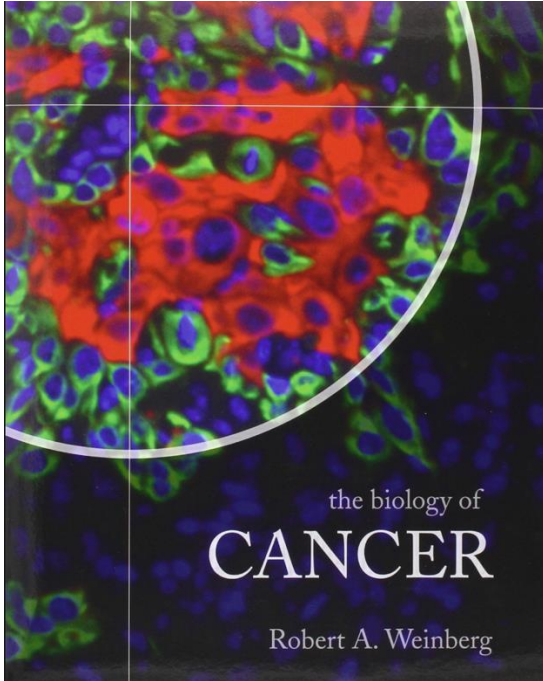
- Wednesday, Nov 19<sup>th</sup> and Wednesday, Dec 10<sup>th</sup>:

I will post two short mock exam sessions with questions from the previous years

# Exam Information

- Exam questions will be in English, but you're allowed to answer in French (*if you do, please use capital letters*)
- *Day to be confirmed:* Dec 15 or 17<sup>th</sup> , 2 - 4 PM (*room will be communicated*)
- ~10 open questions
  - 10 points for each question
  - 2 questions will be on the papers that you will read during the exercises
- The exam will count for 50% of your final grade

# If you want to read on specific topics:



Some information if this book

Send an email:

I can send you recent Reviews written by prominent cancer scientists in the fields

We can discuss interest for your lab immersion and master thesis project

**Why do we develop cancer?**

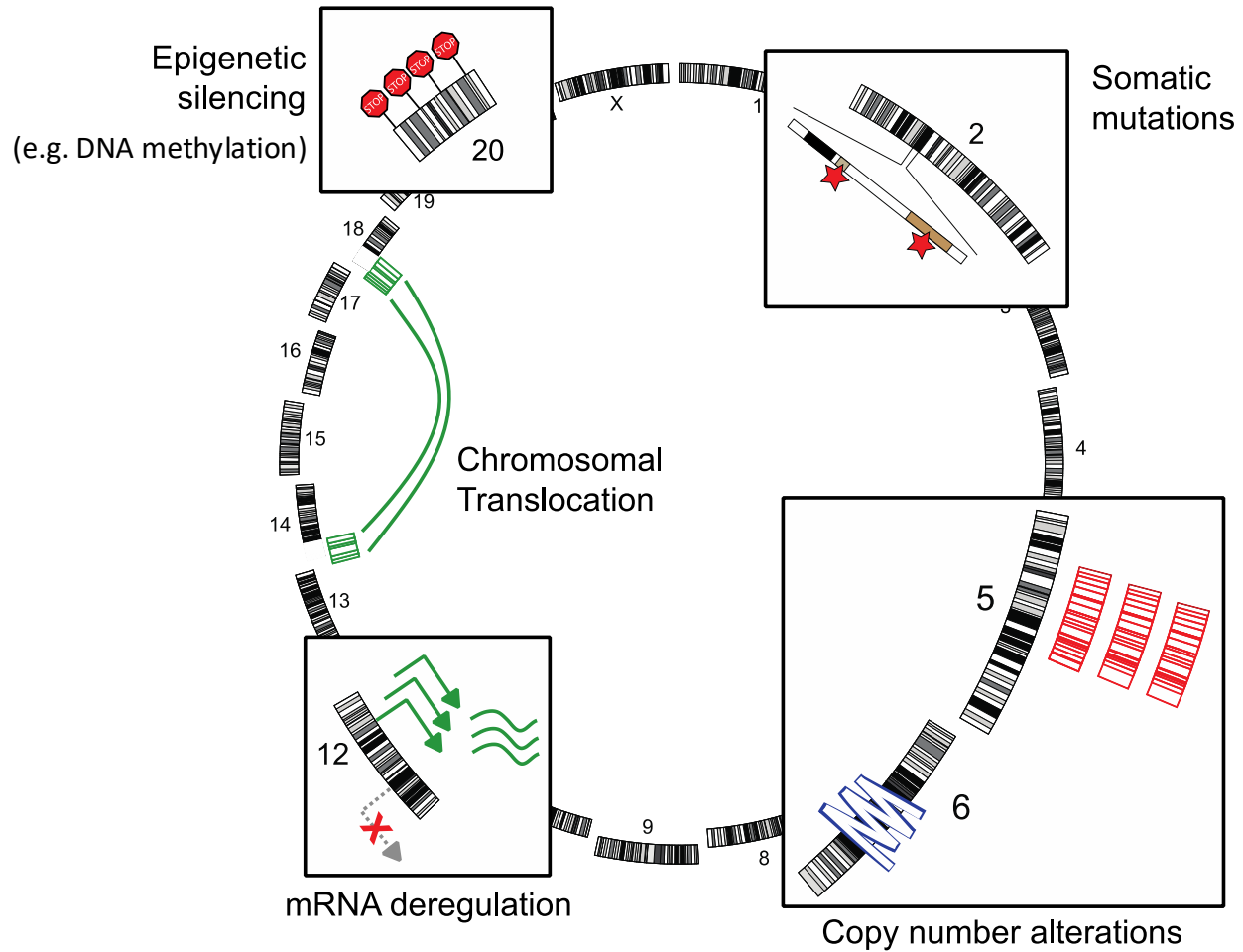
# 1. Cancer is a disease of the genome



Cancer genomics: to study the acquisition of alterations in the genome (i.e. in our DNA) that can cause cancer development

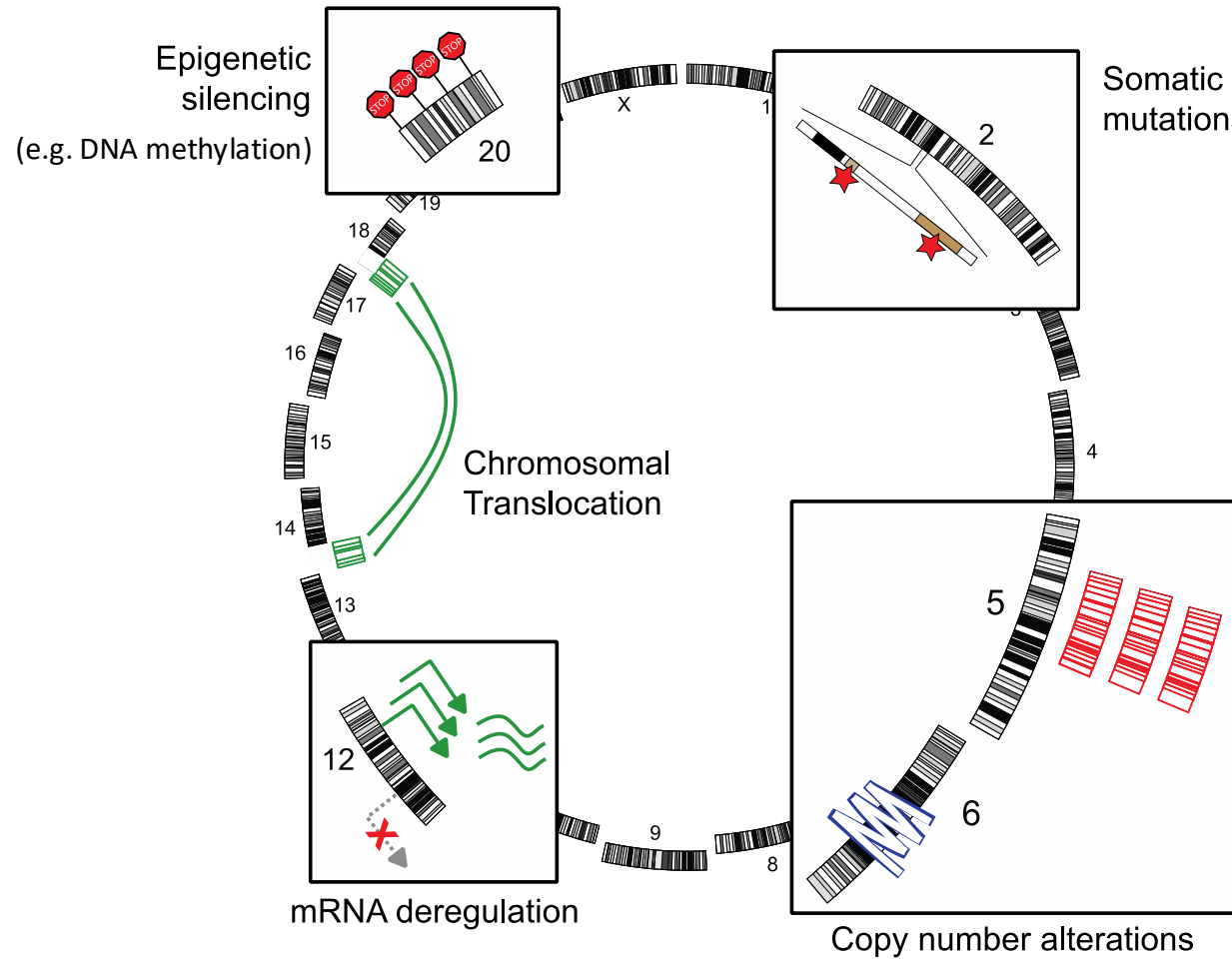
What are they?

# Cancer Genomic Alterations



What are they?

# Cancer Genomic Alterations



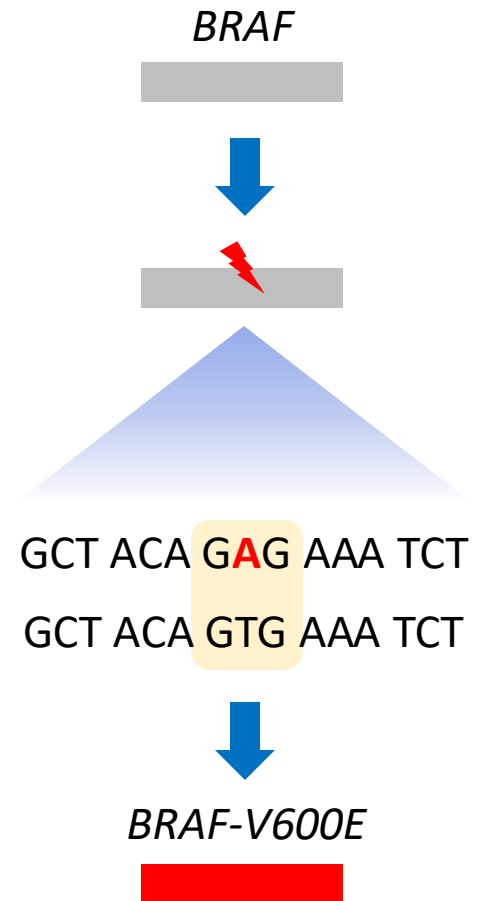
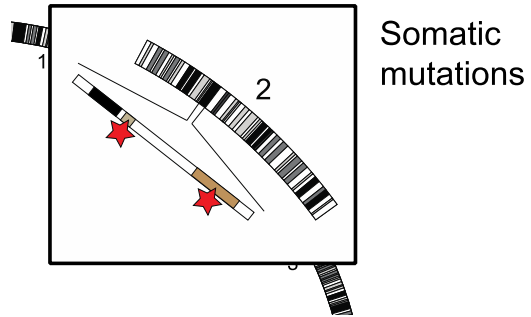
You like my dogs

You like my **dots**



# Mutations

- Single nucleotide changes



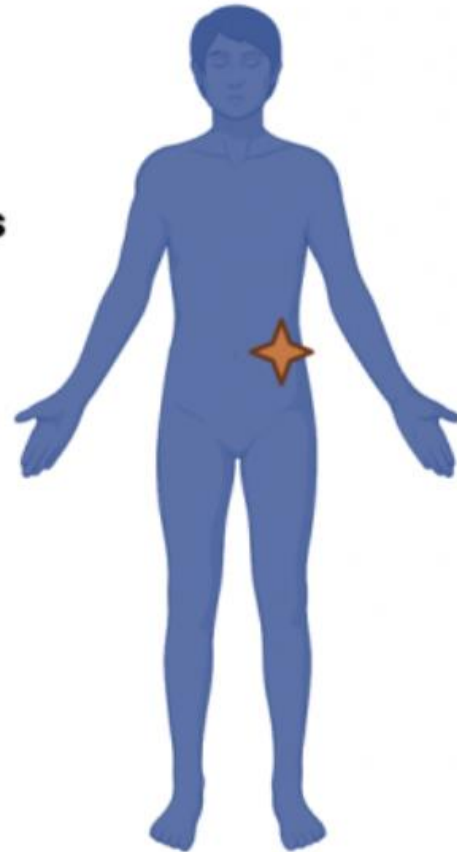
# Mutations

## Somatic DNA changes

Acquired over a person's lifetime in single cells

Can lead to cancer

Can NOT be inherited



## Germline DNA changes

Present in every cell of the body including egg and sperm

Can increase cancer susceptibility



Can be inherited



**How can we find mutations?**

**How can we find **somatic** mutations?**

# Somatic Mutations have the same properties of germline mutations

- Single nucleotide changes

- Missense Mutations

- GAG = Valine
- GTG = Glutamate

- Silent Mutations

- TCT = Serine
- TCC = Serine

- Nonsense Mutations

- TAC = Tyrosine
- TAG = Stop Codon!

	T			C			A			G		
T	TTT	Phe	F	TCT	Ser	S	TAT	Tyr	Y	TGT	Cys	C
	TTC	Phe	F	TCC	Ser	S	TAC	Tyr	Y	TGC	Cys	C
	TTA	Leu	L	TCA	Ser	S	TAA	stop	*	TGA	stop	*
	TTG	Leu	L	TCG	Ser	S	TAG	stop	*	TGG	Trp	W
C	CTT	Leu	L	CCT	Pro	P	CAT	His	H	CGT	Arg	R
	CTC	Leu	L	CCC	Pro	P	CAC	His	H	CGC	Arg	R
	CTA	Leu	L	CCA	Pro	P	CAA	Gln	Q	CGA	Arg	R
	CTG	Leu	L	CCG	Pro	P	CAG	Gln	Q	CGG	Arg	R
A	ATT	Ile	I	ACT	Thr	T	AAT	Asn	N	AGT	Ser	S
	ATC	Ile	I	ACC	Thr	T	AAC	Asn	N	AGC	Ser	S
	ATA	Ile	I	ACA	Thr	T	AAA	Lys	K	AGA	Arg	R
	ATG	Met	M	ACG	Thr	T	AAG	Lys	K	AGG	Arg	R
G	GTT	Val	V	GCT	Ala	A	GAT	Asp	D	GGT	Gly	G
	GTC	Val	V	GCC	Ala	A	GAC	Asp	D	GGC	Gly	G
	GTA	Val	V	GCA	Ala	A	GAA	Glu	E	GGA	Gly	G
	GTG	Val	V	GCG	Ala	A	GAG	Glu	E	GGG	Gly	G

# Somatic Mutations

- **Frame-shift mutations:** insertion or deletion that change the reading frame
- **Deletion:** deletion of 1 or more nucleotides

ACC AGC TGC ACT  
Thr Ser Cys Thr

ACC AGC TGA CT  
Thr Ser **stop**

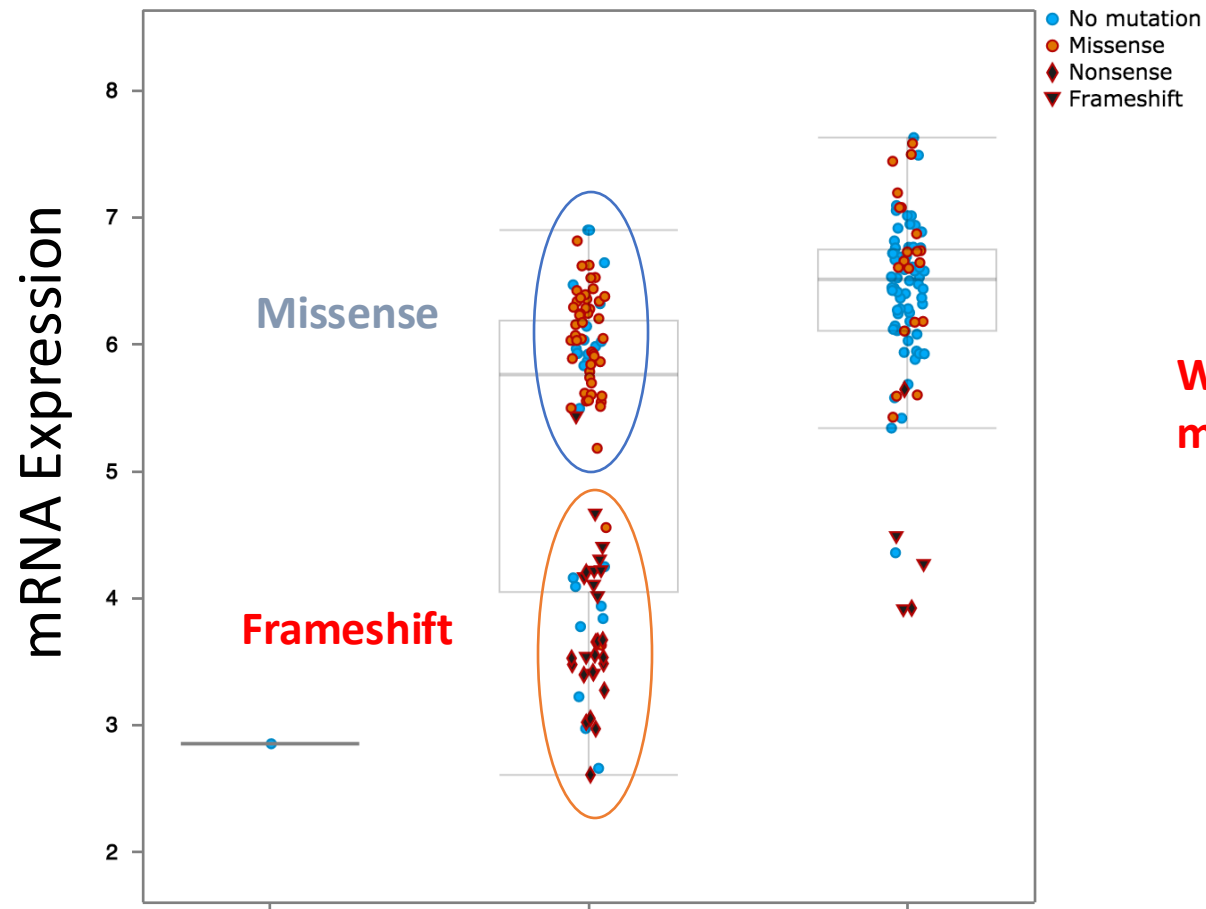
- **Insertion:** Addition 1 or more extra-nucleotides

ACC AGC TGC ACT  
Thr Ser Cys Thr

ACC AGC TGC CAC CT  
Thr Ser Cys His

# Truncating Mutations (inactivating a **tumor suppressor**)

- TP53 mutations in Colorectal cancer patient



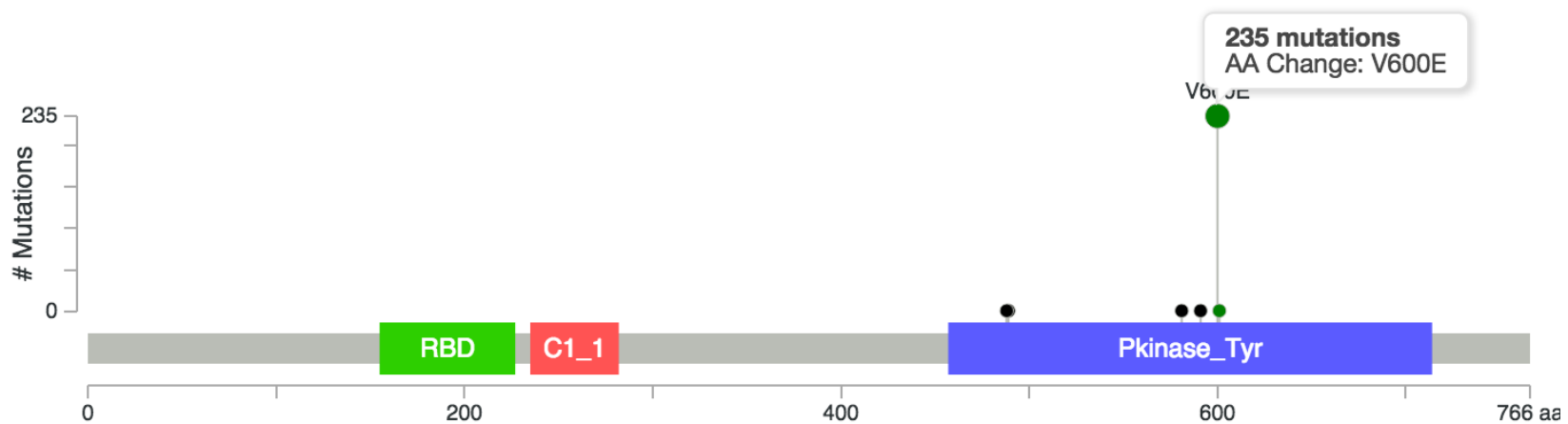
**Why this difference between missense and frameshifts?**

# HOTSPOT mutations (activating an **oncogene**)

## BRAF V600E mutations in Thyroid Carcinoma (399 patients)

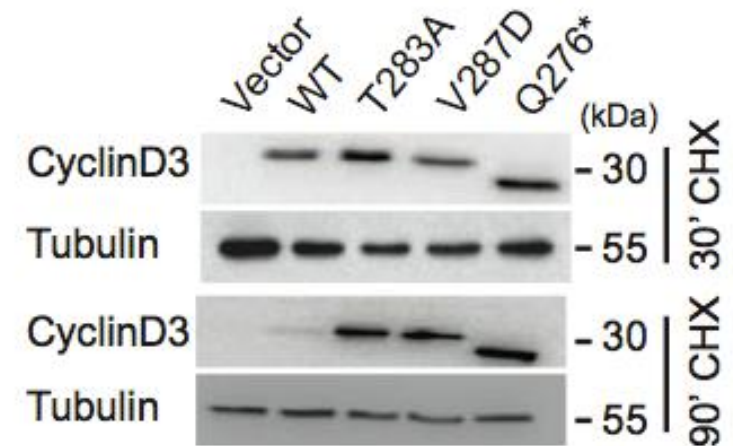
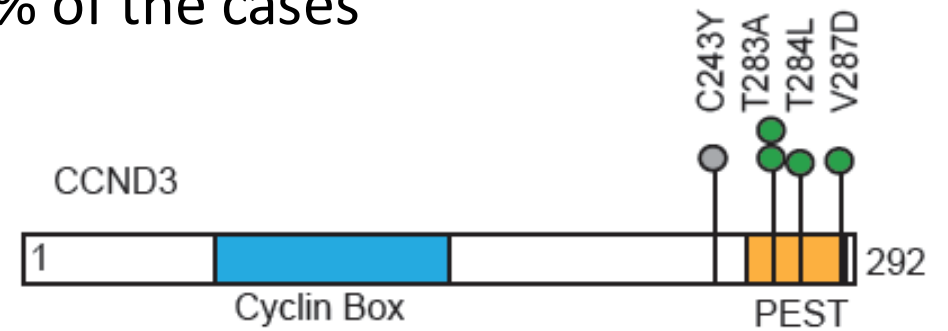
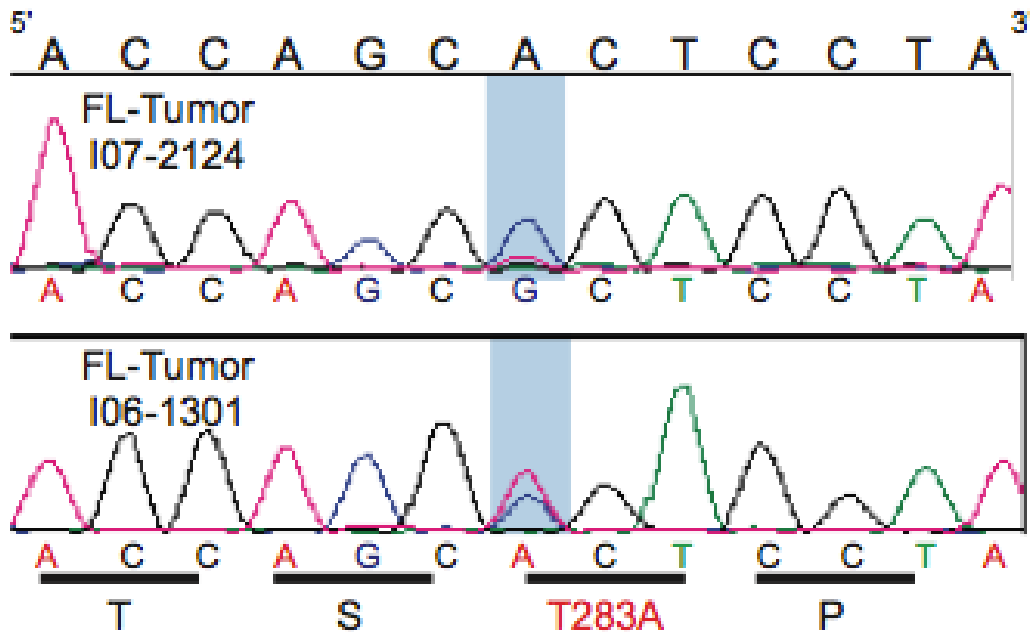
**GTG** = Valine (V)

**GAG** = Glutamate (E)



# Truncating Mutations (activating an **oncogene**)

In Lymphoma mutation in CyclinD3 occurs in ~10% of the cases

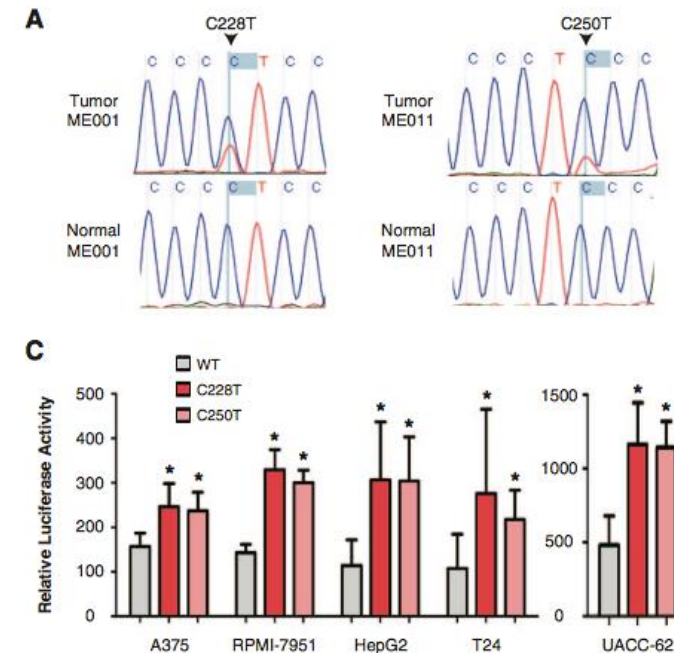
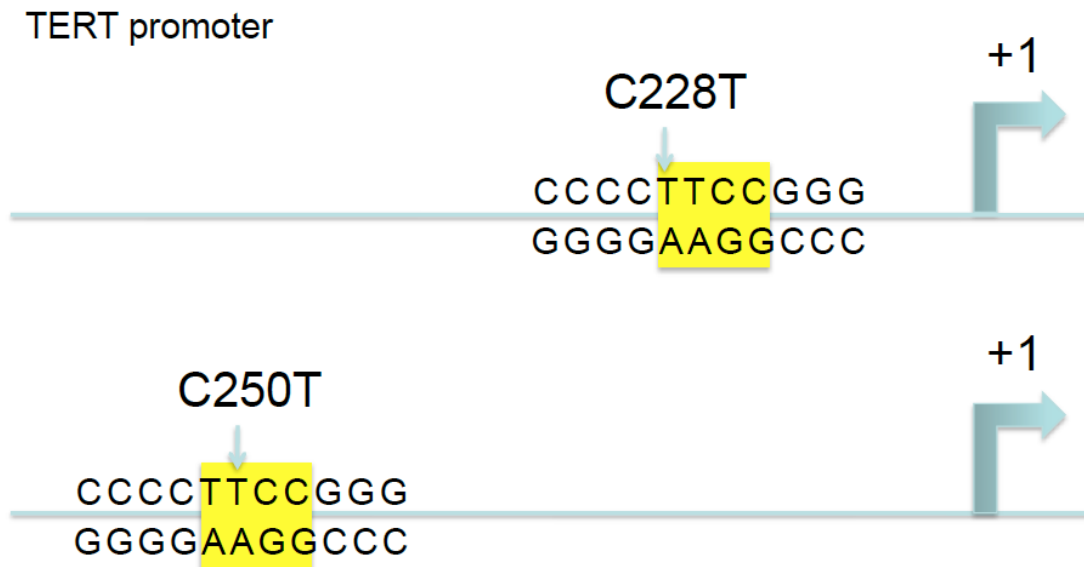


# Non-coding Mutations

## Highly Recurrent *TERT* Promoter Mutations in Human Melanoma

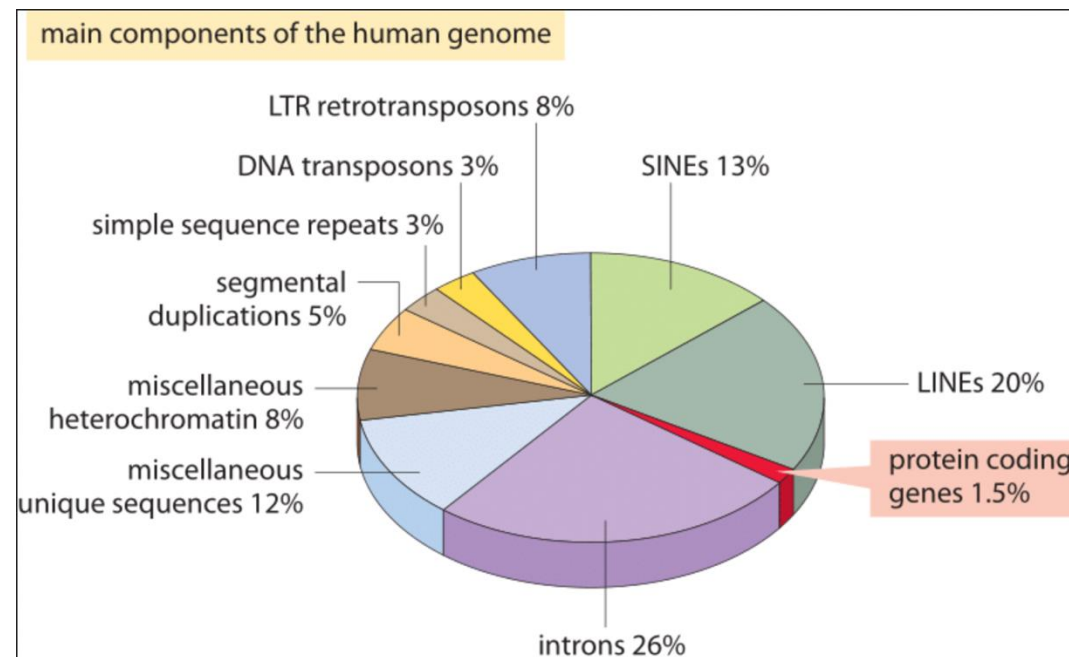
Franklin W. Huang,<sup>1,2,3\*</sup> Eran Hodis,<sup>1,3,4\*</sup> Mary Jue Xu,<sup>1,3,4</sup> Gregory V. Kryukov,<sup>1</sup>  
Lynda Chin,<sup>5,6</sup> Levi A. Garraway<sup>1,2,3†</sup>

(Science, 2013)



# What should we sequence?

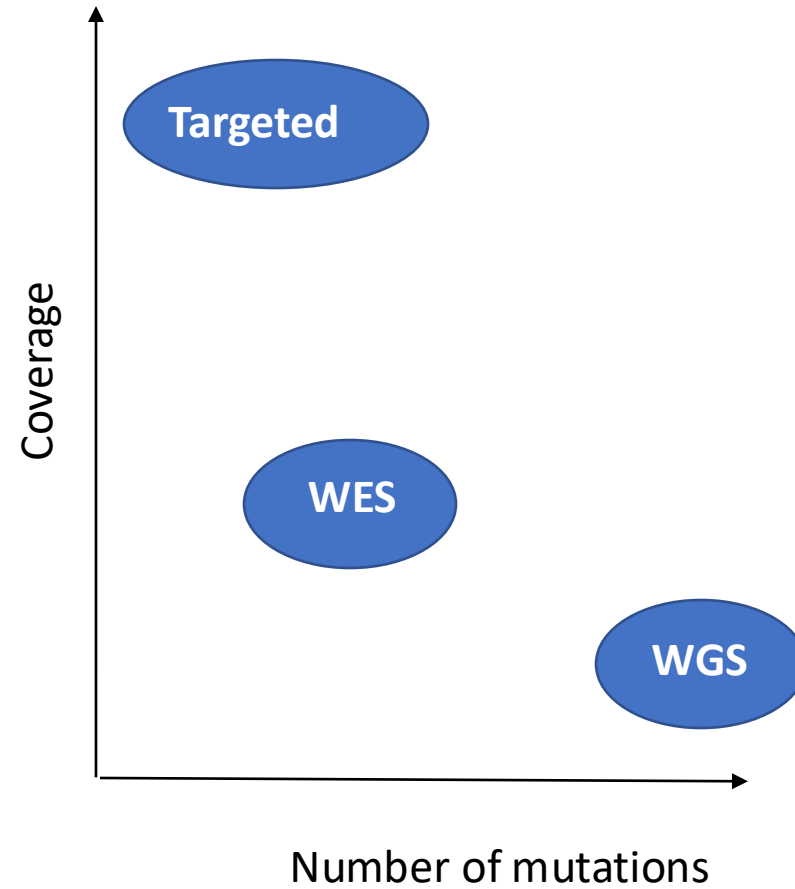
## How much should we sequence?



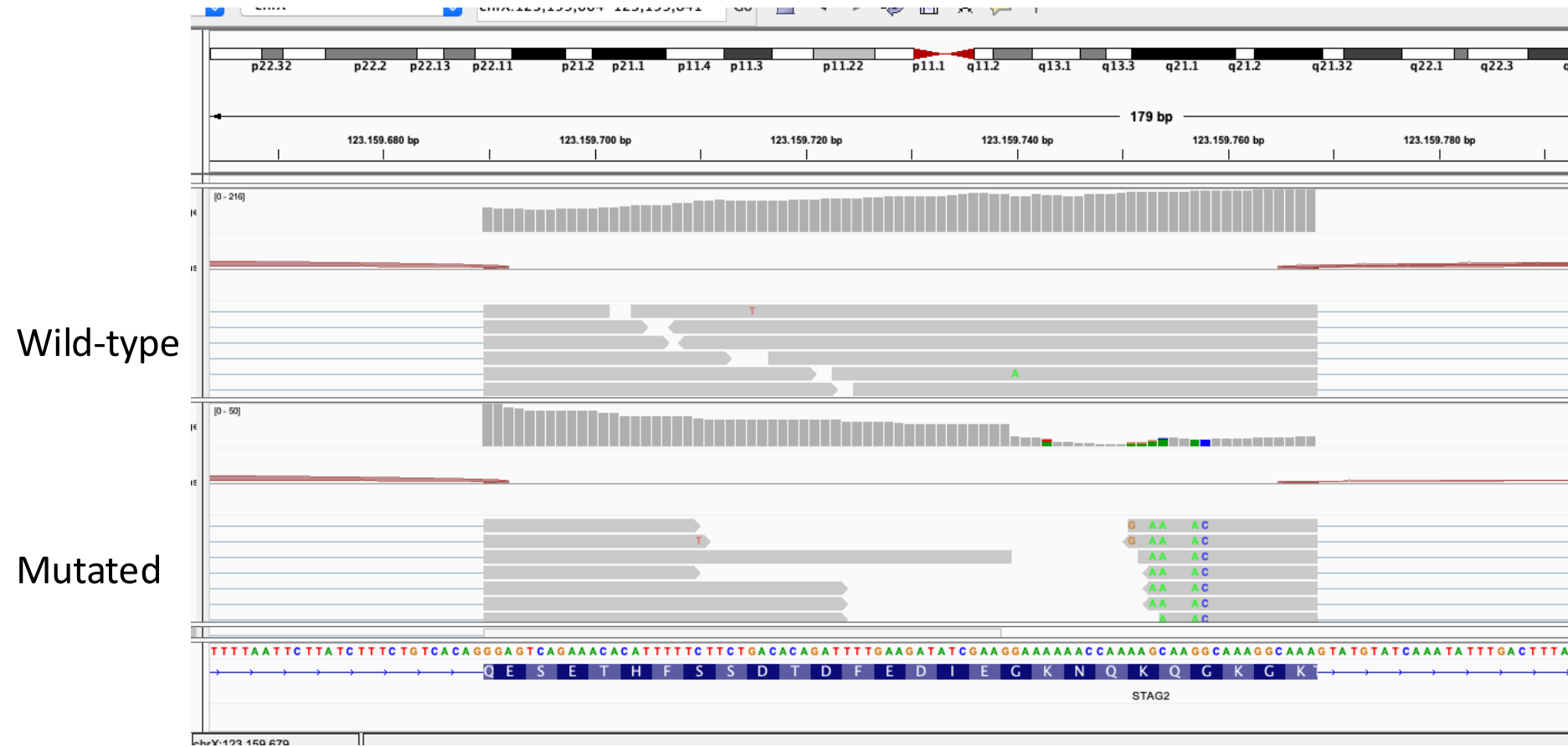
# Cancer Genomics

- Sequencing the blood and tumor biopsy
- **Targeted sequencing** – select a limited number of genes that will be sequenced (e.g. the most frequent mutated genes)
- **Whole-exome sequencing** – sequence the coding genome
- **Whole-genome sequencing**- sequence coding and non-coding regions

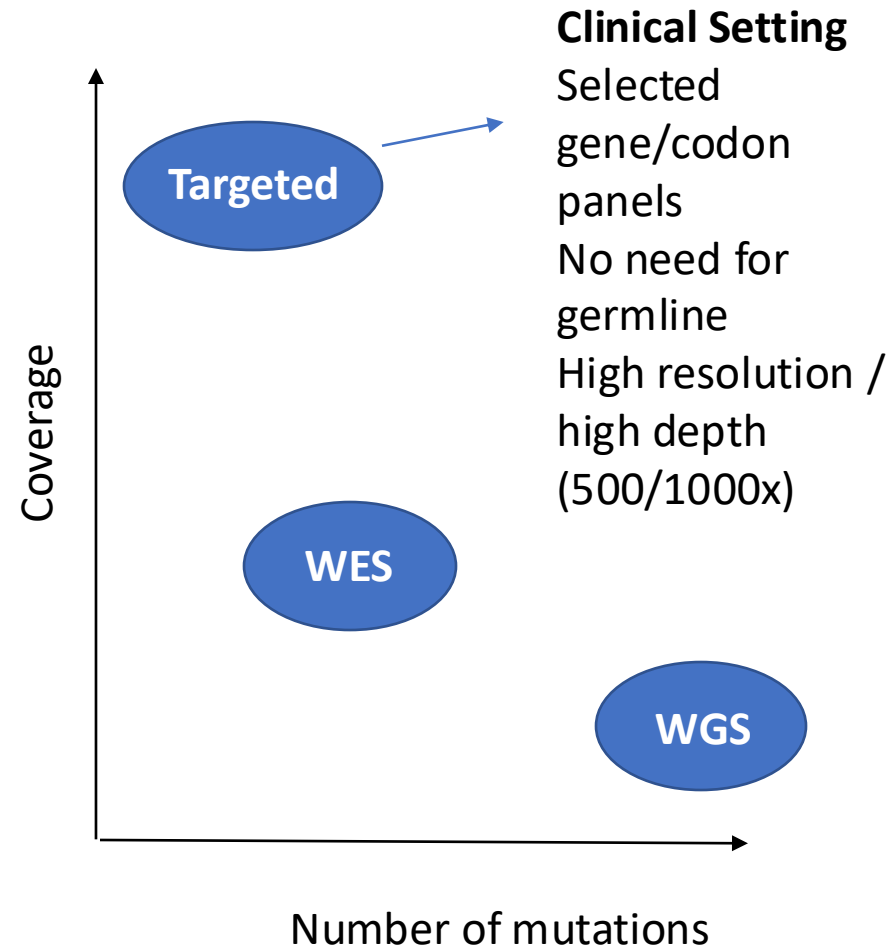
# Sequencing a tumor



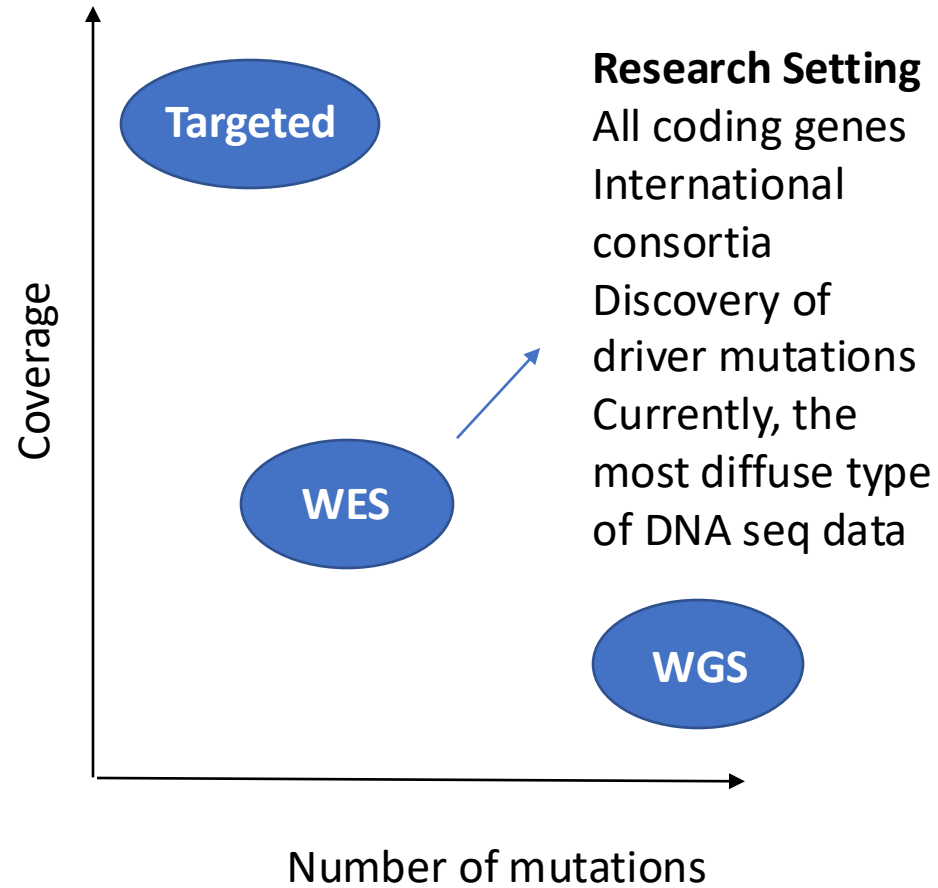
# Coverage versus number of mutations



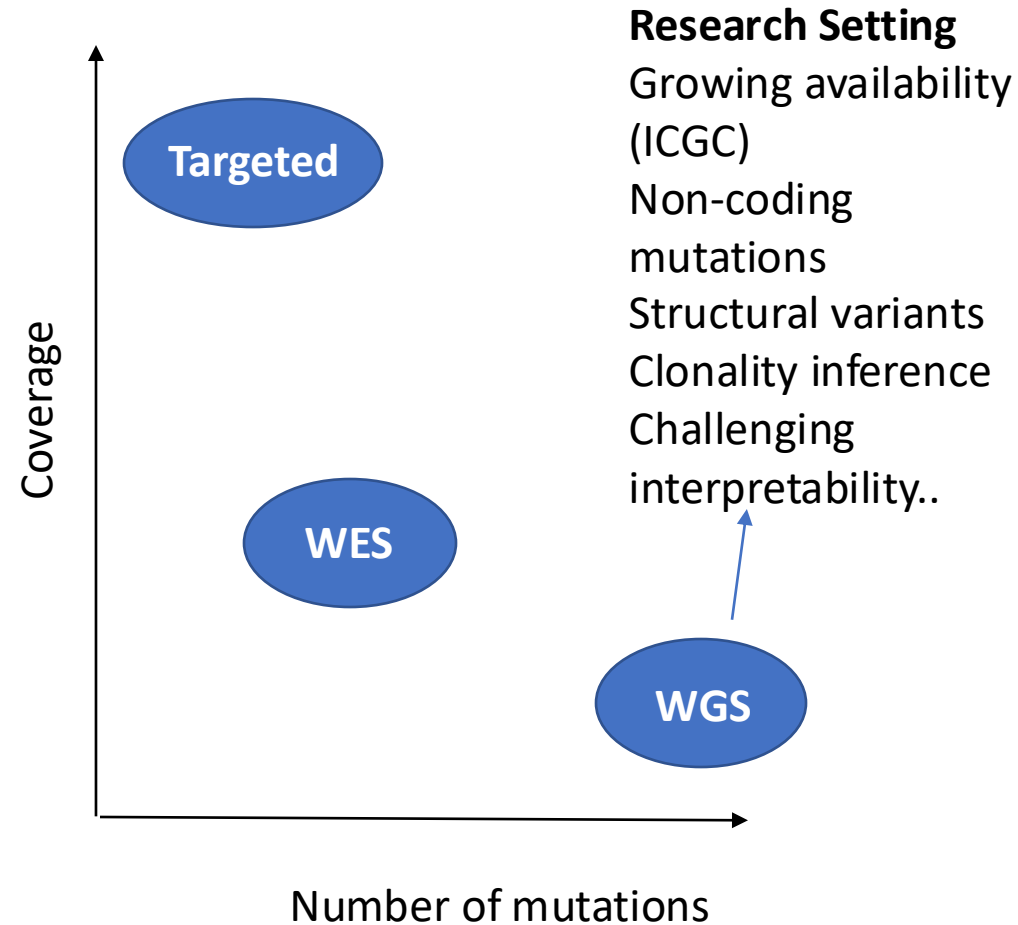
# Targeted Sequencing



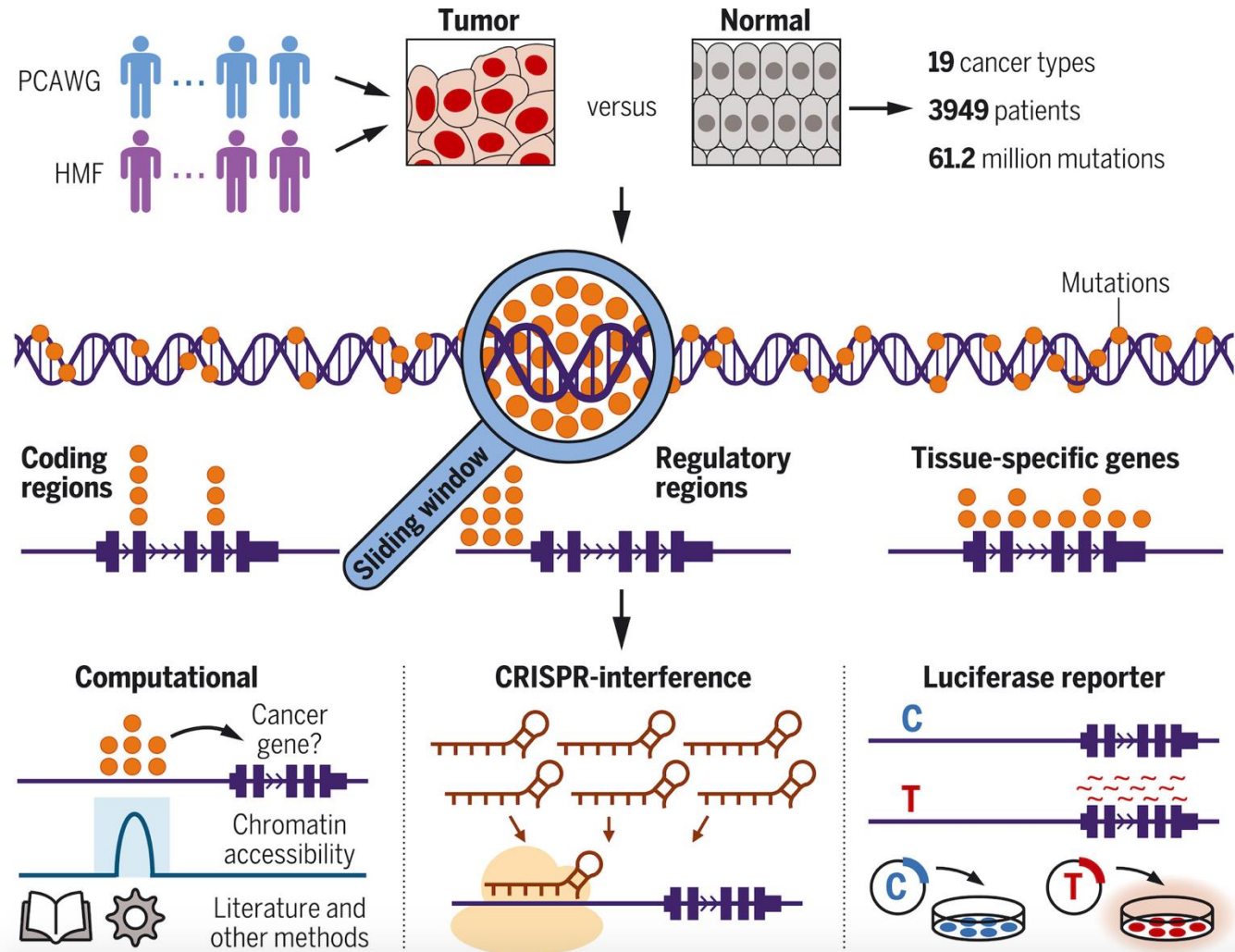
# Whole Exome Sequencing



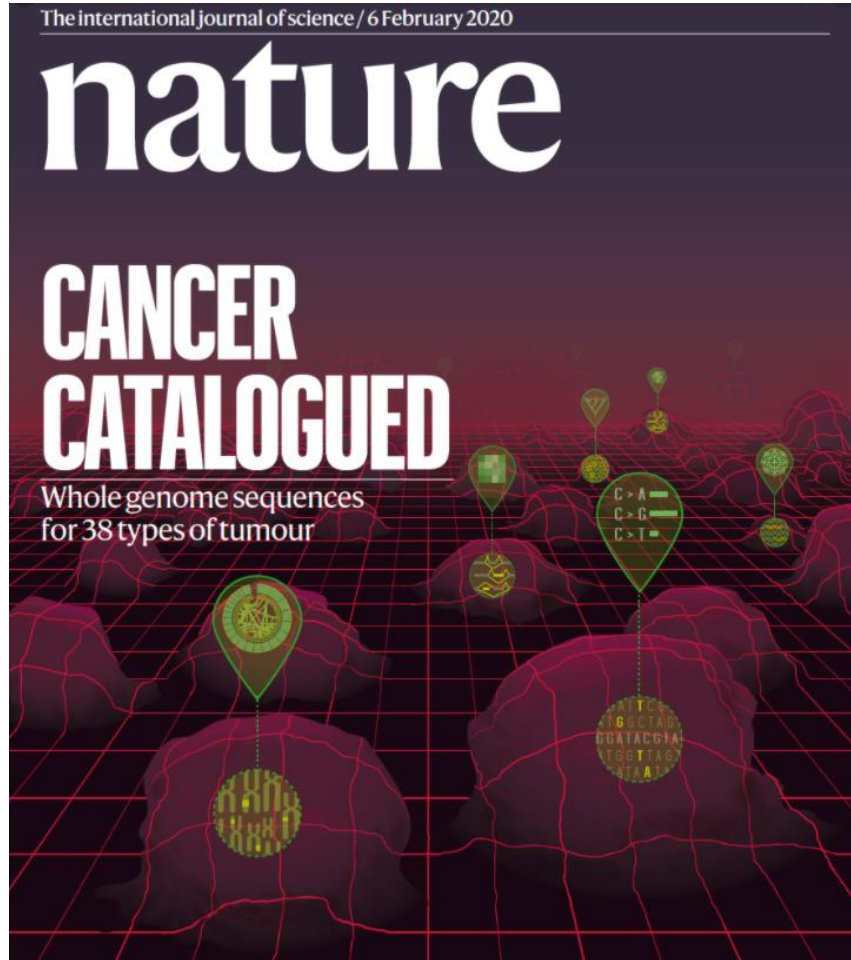
# Whole genome sequencing



# Non-coding Mutations



# Pan-Cancer Analyses of Whole Genomes



Article | [Open Access](#) | Published: 05 February 2020

## Analyses of non-coding somatic drivers in 2,658 cancer whole genomes

Esther Rheinbay, Morten Muhligh Nielsen, [...] PCAWG Consortium

*Nature* 578, 102–111(2020) | [Cite this article](#)

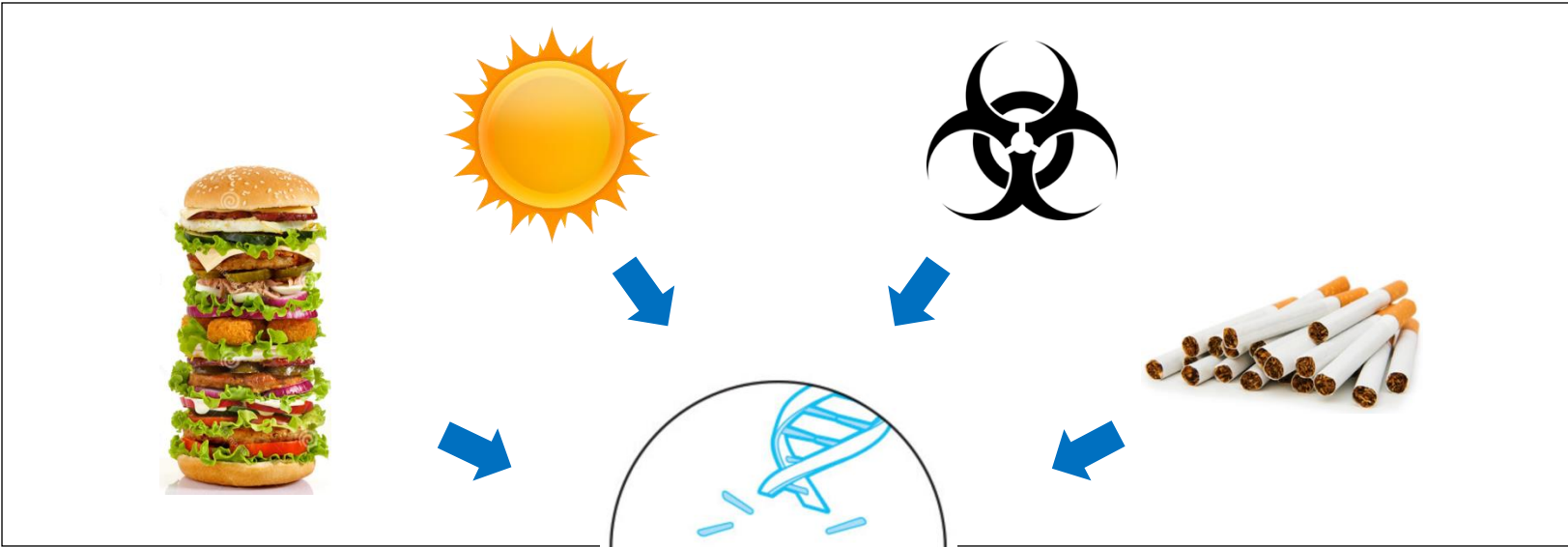
**Paucity of non-coding drivers in cancer**

Why?

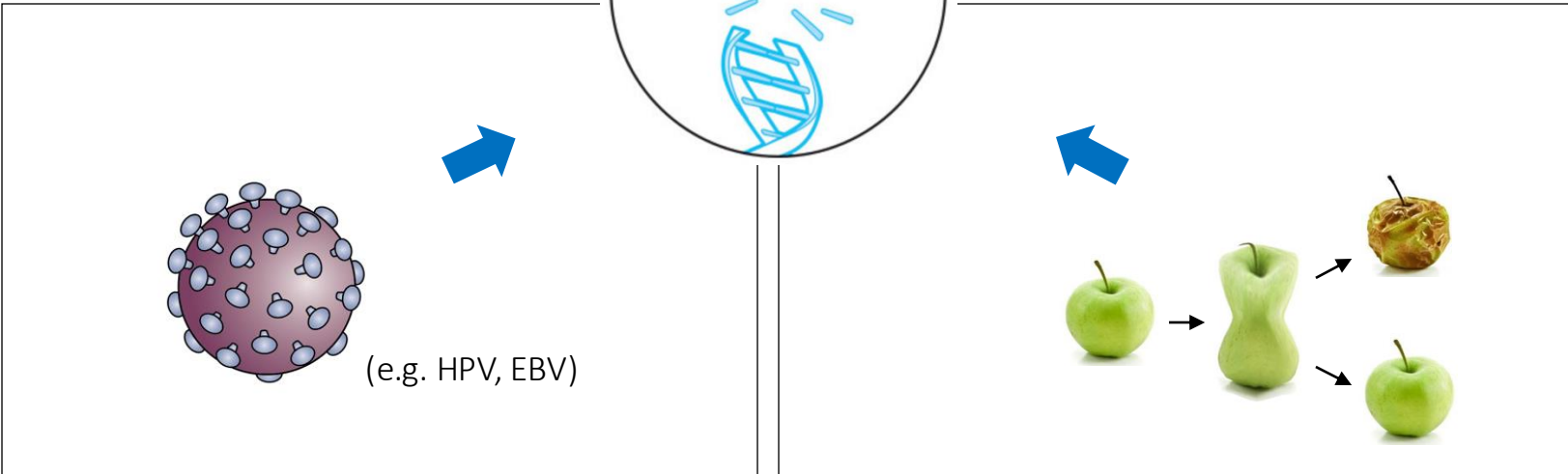
How do mutations emerge?

# How do *mutations* emerge?

Exogenous mutagens

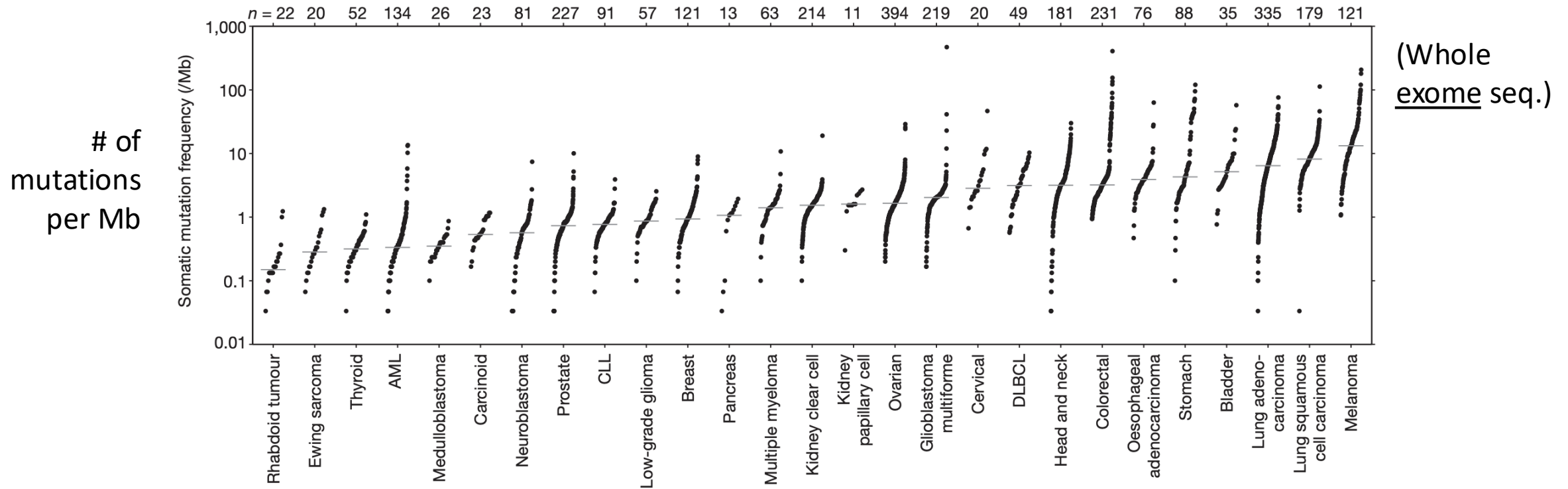


Viral infection

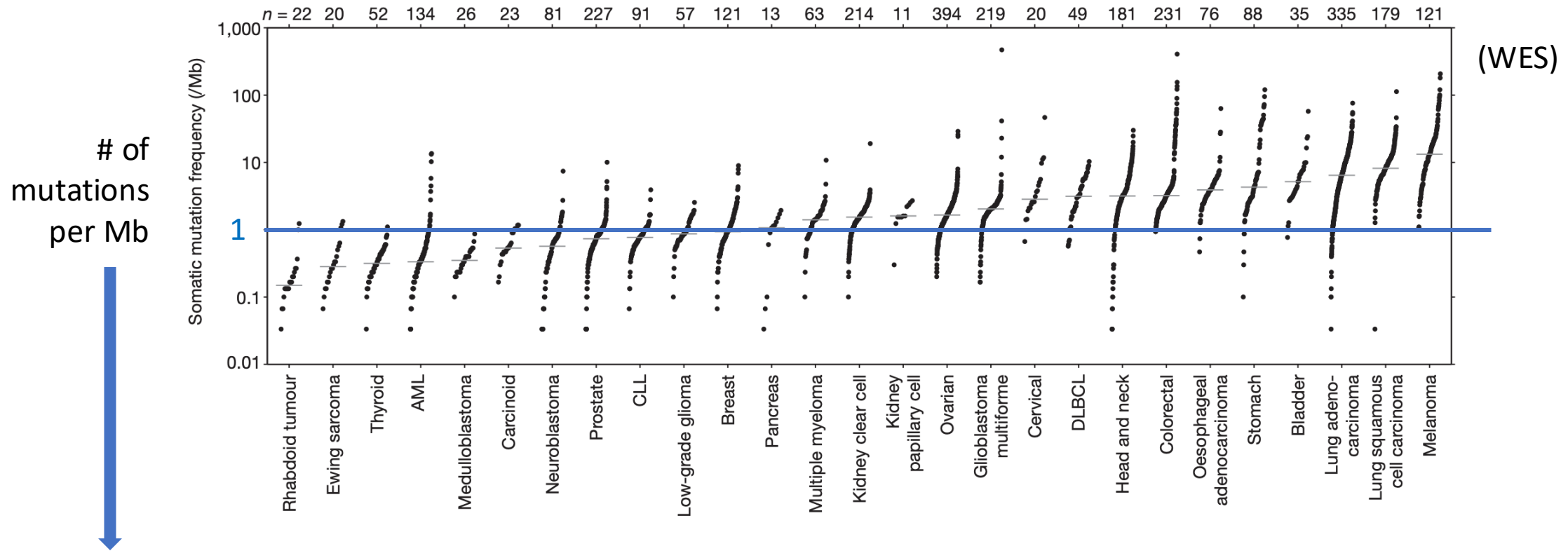


Unrepaired Replication Errors

# What do you get after sequencing 1000 tumors?



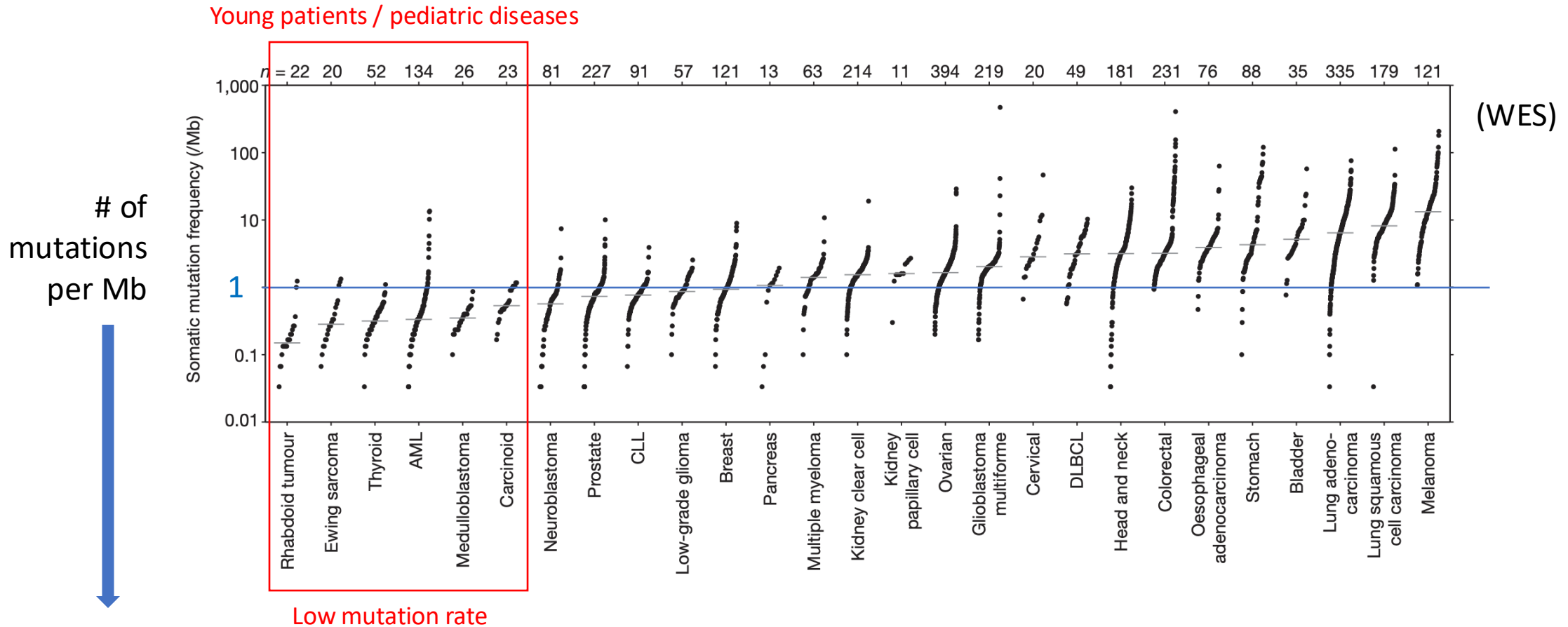
# What do you get after sequencing 1000 tumors?



1 → 1/Mb \* 3000Mb ~ 3000 mutations

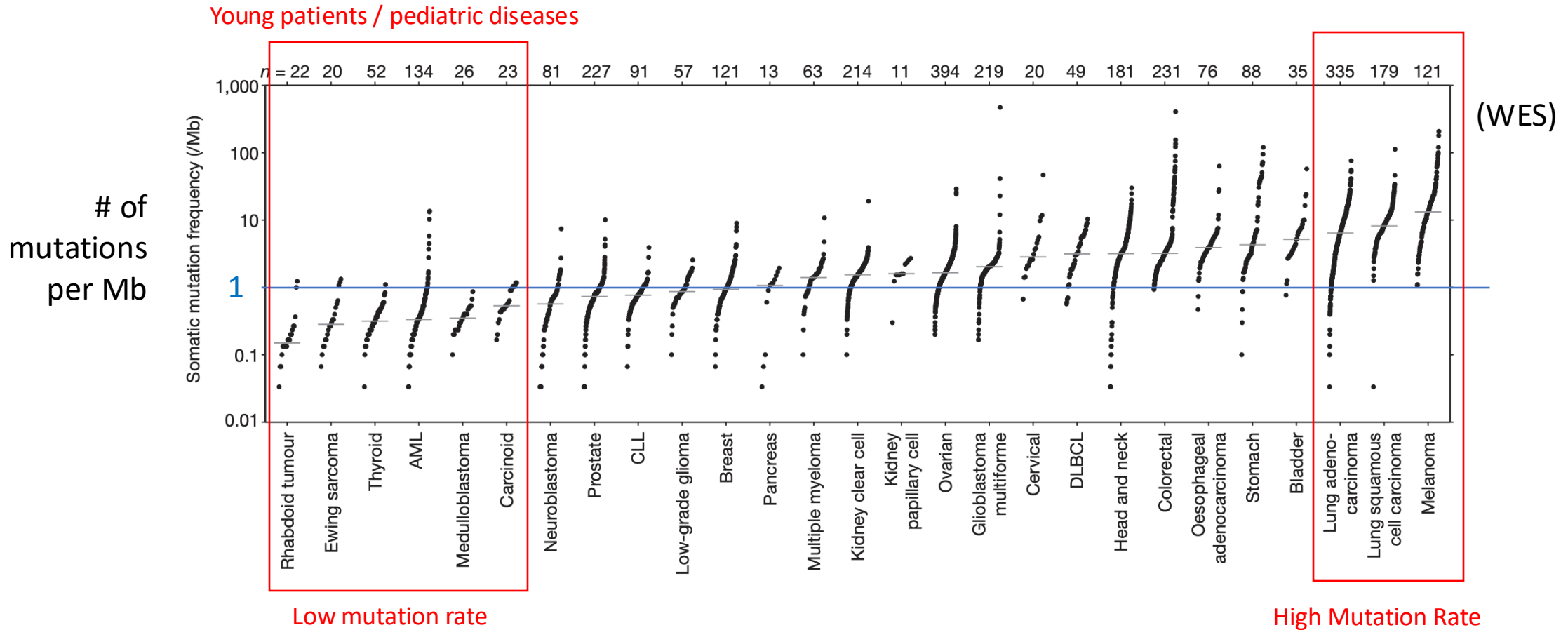
2% are coding sequences → ~60 mutations in gene sequences

# What do you get after all this sequencing?



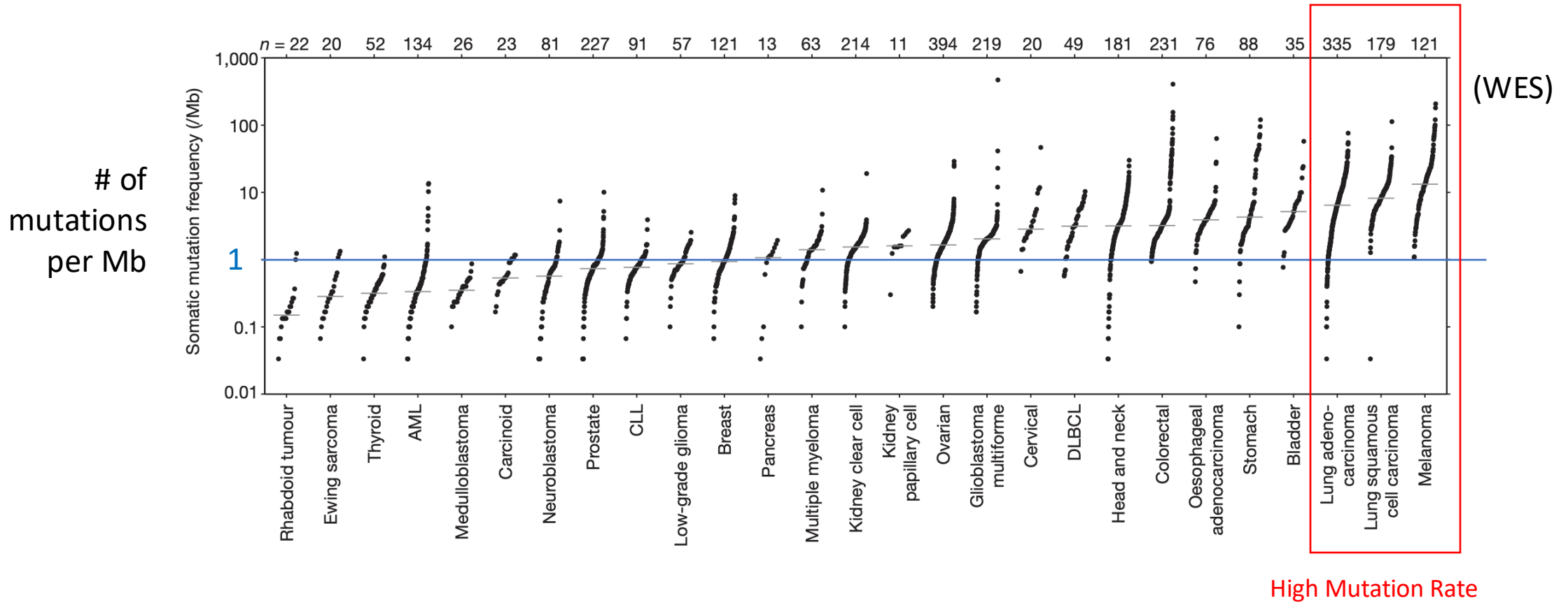
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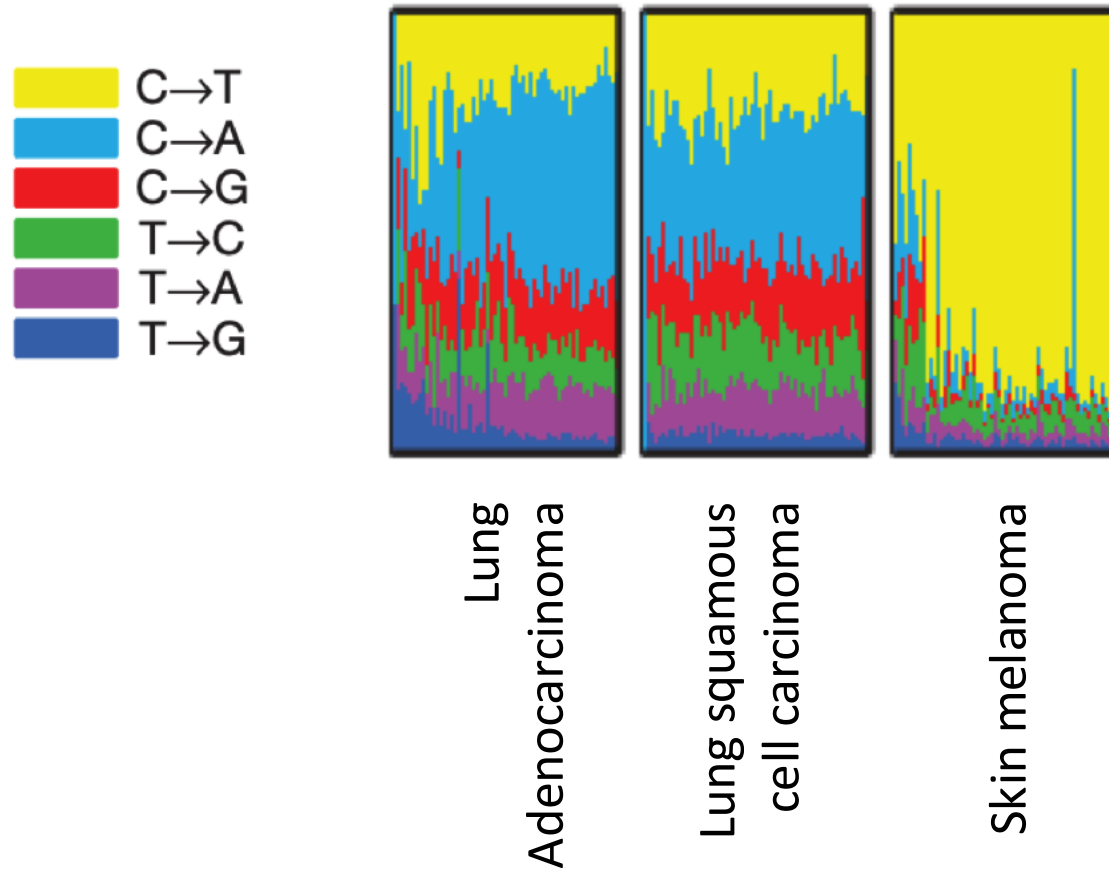


**Why a difference between tumor that develops in children vs the one that develop in adults?**

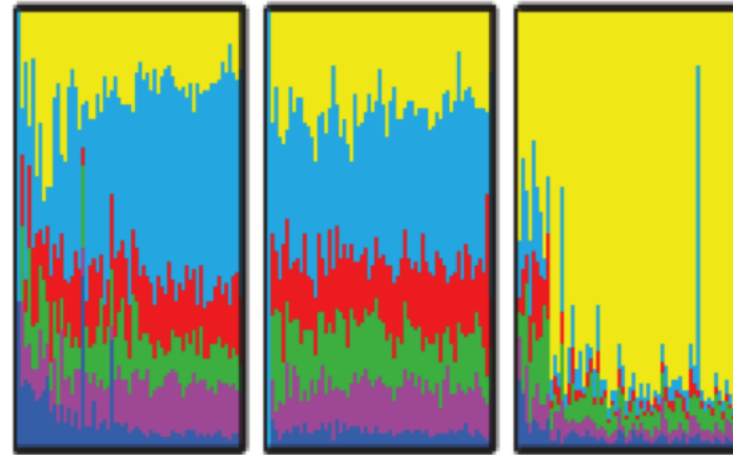
# What do you get after all this sequencing?



# Mutational Signatures



# Mutational Signatures



Lung  
Adenocarcinoma

Lung squamous  
cell carcinoma

Skin melanoma



C → A



C → T



# Mutational Signatures

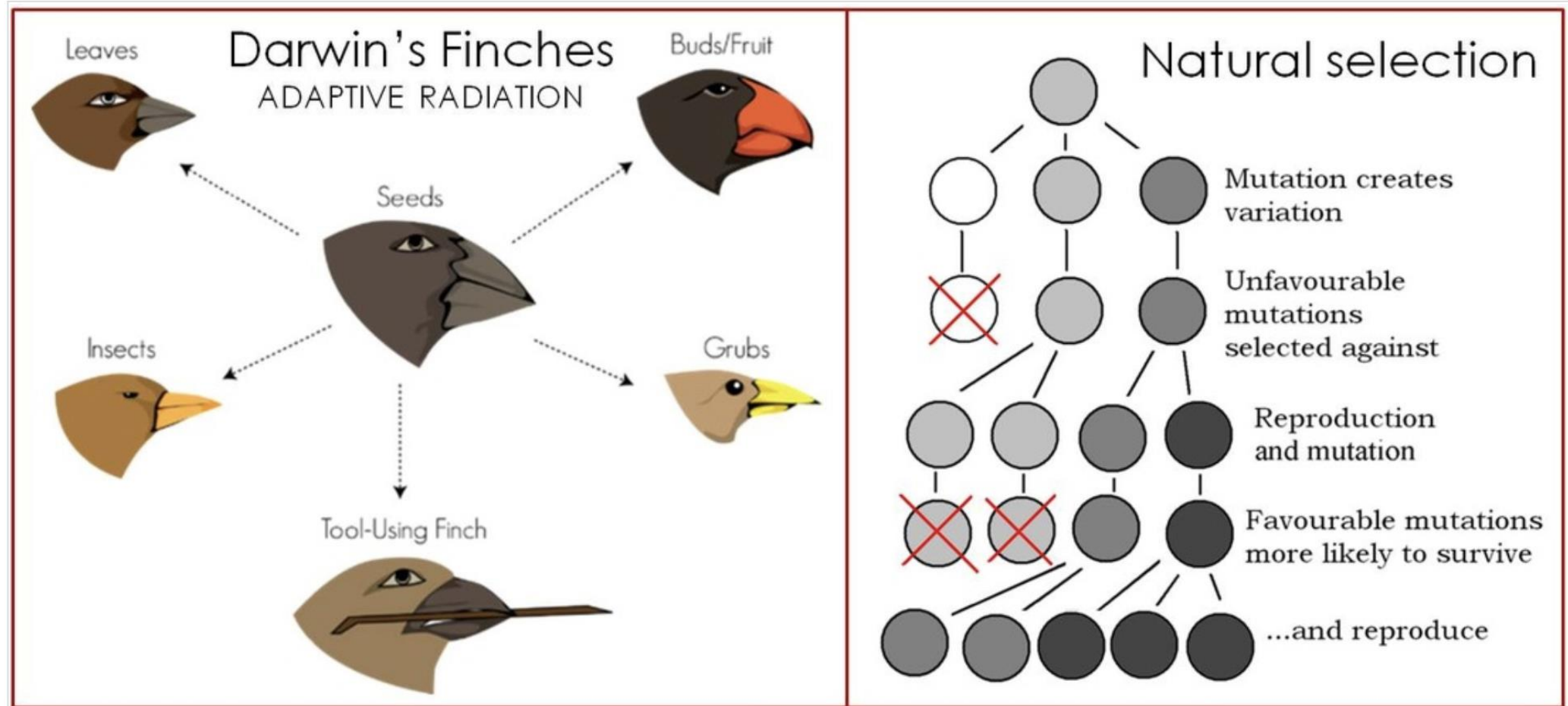
Spontaneous  
De-amination  
(defective base  
excision repair)



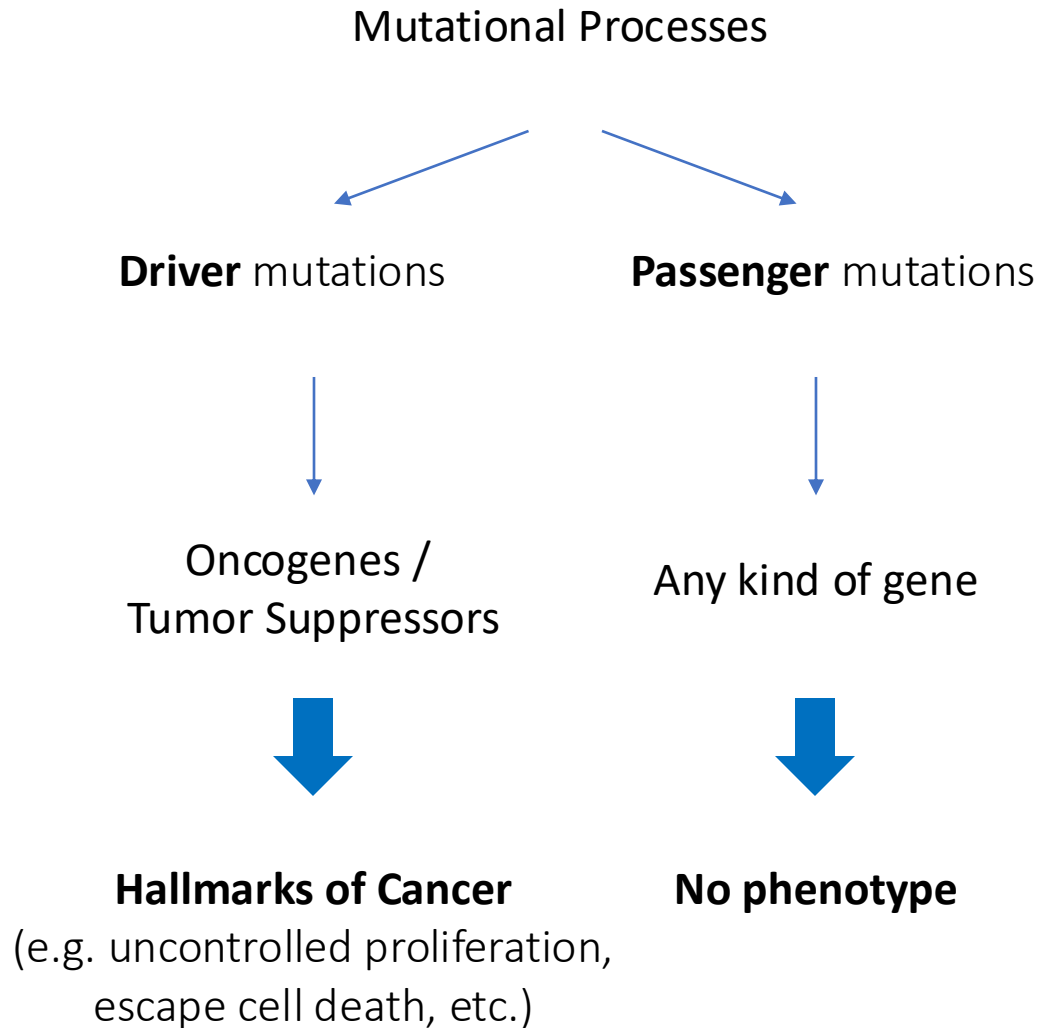
Tobacco  
smoking

Are all mutations bad, and do they all  
cause cancer?

# Mutation in species evolution

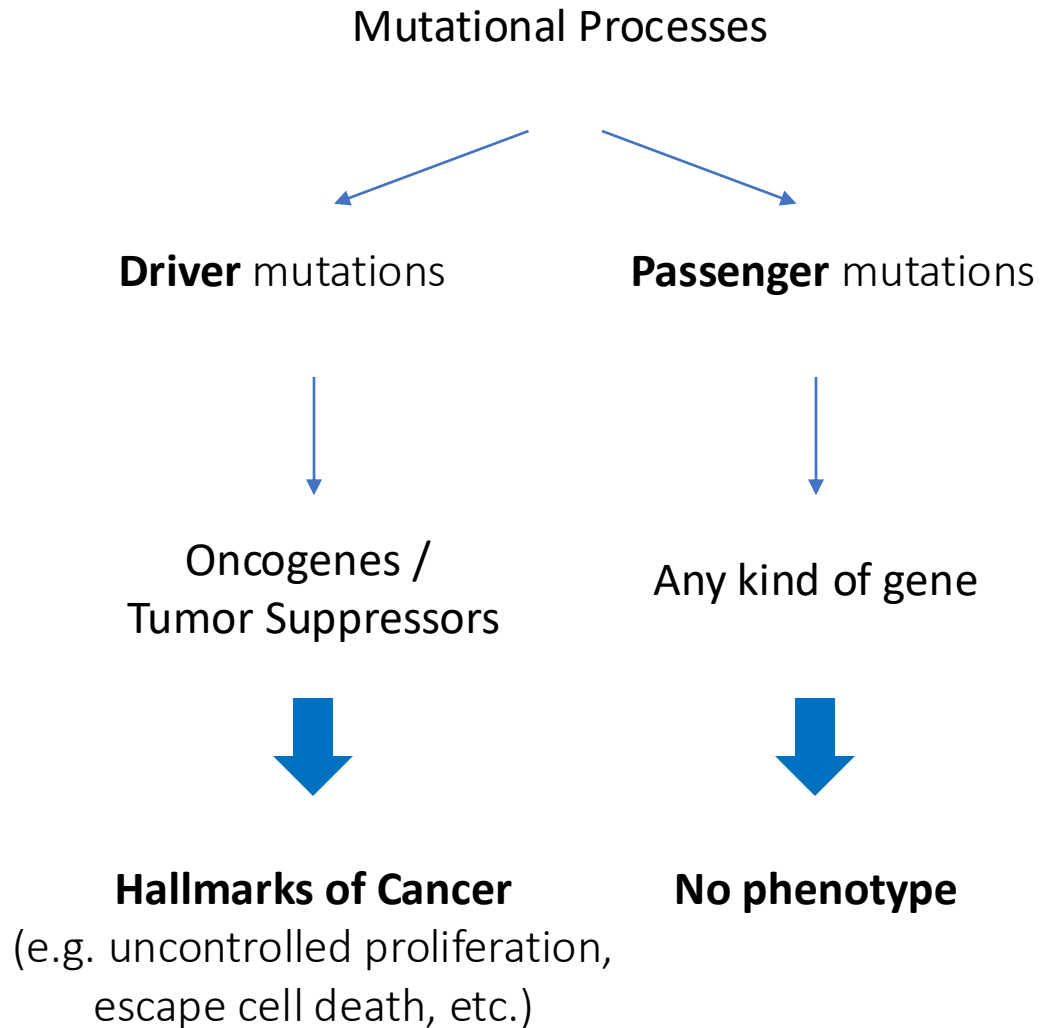


# Driver vs Passenger mutations



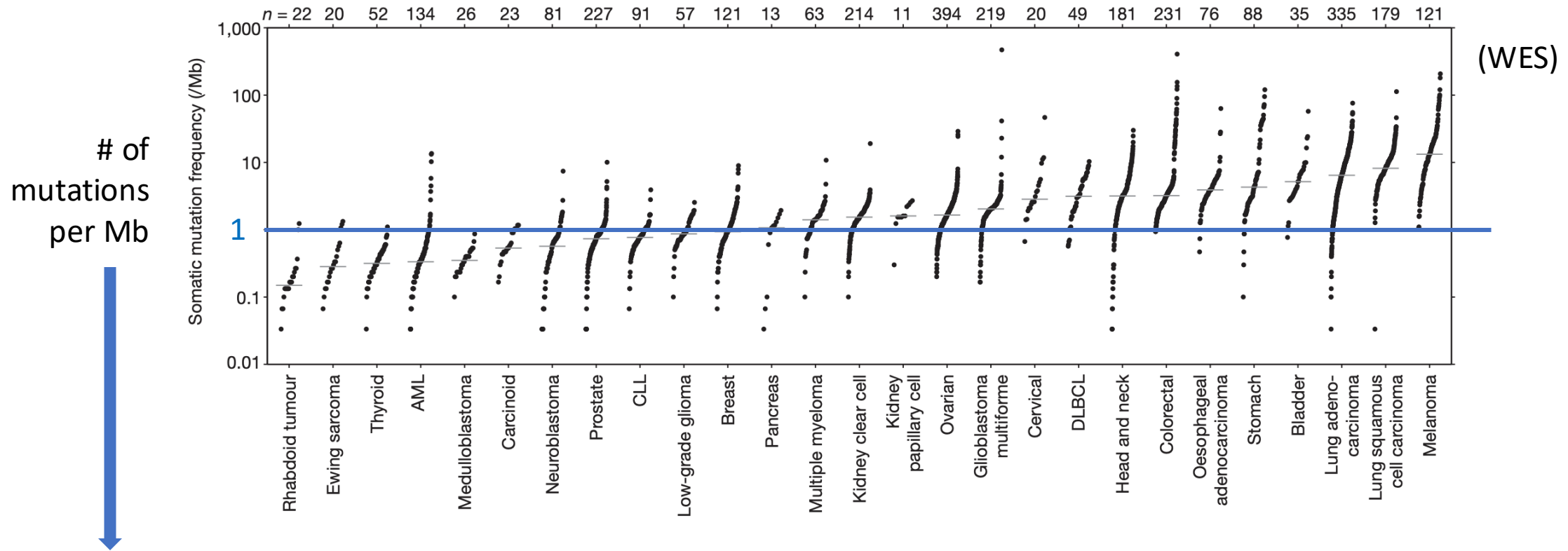
If you have a driver, usually  
you also have a passenger

# Driver vs Passenger mutations



Often you have many more passengers than drivers!

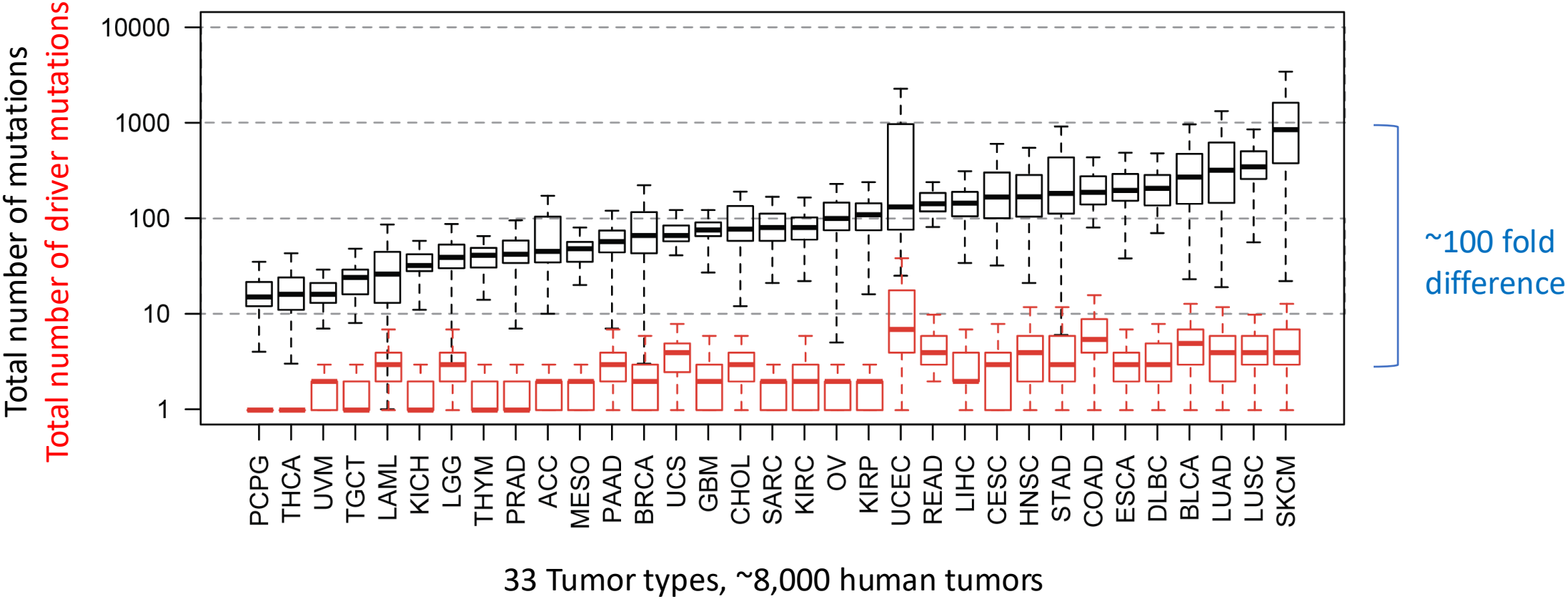
# What do you get after sequencing 1000 tumors?



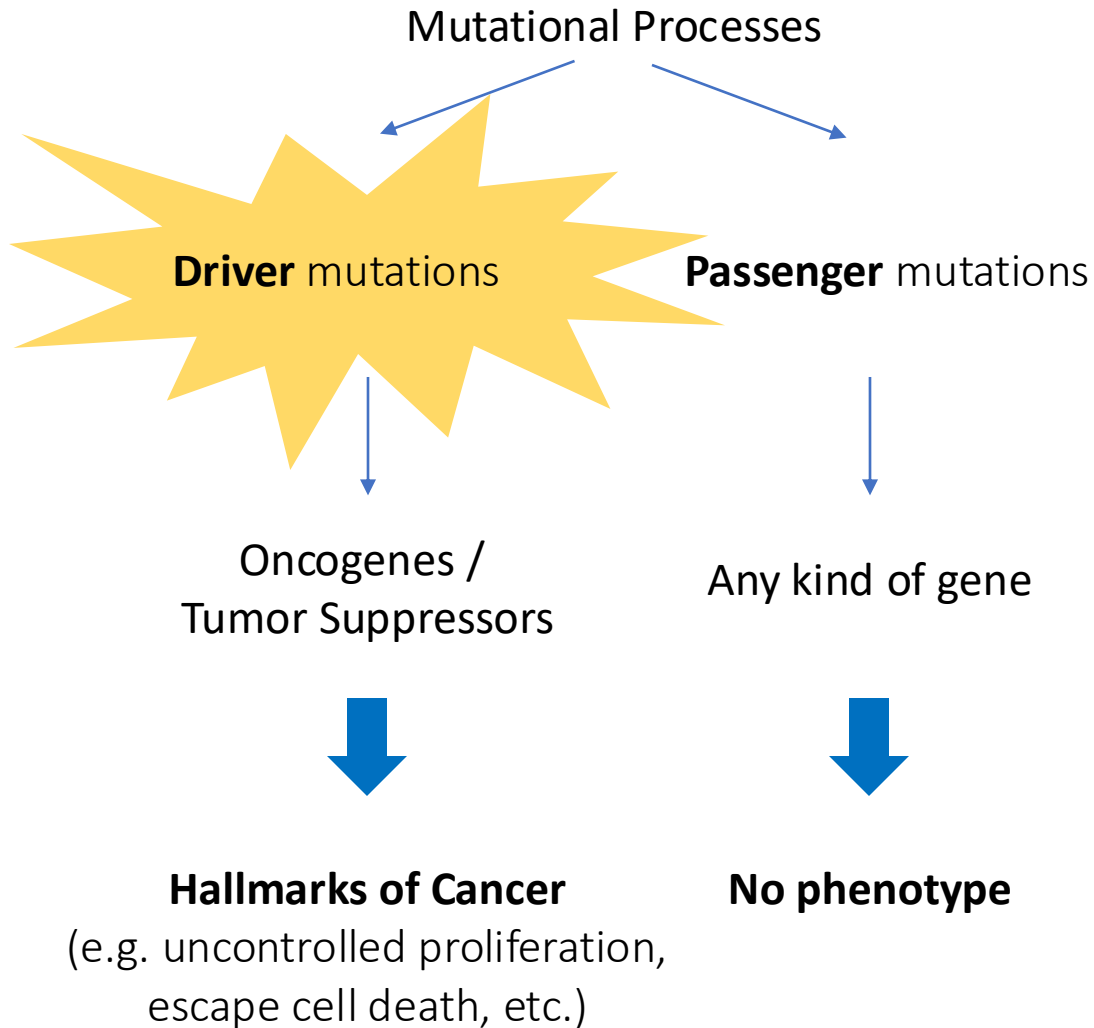
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2% are coding sequences → ~60 mutations in gene sequences

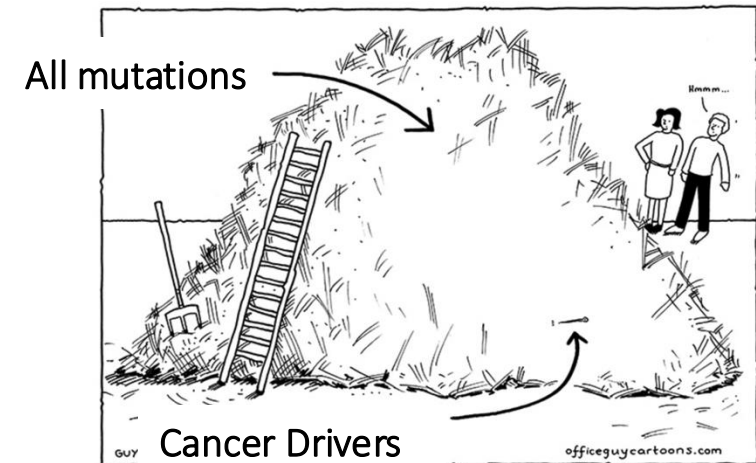
# Driver vs. passenger alterations



# Driver vs Passenger mutations



## How do we identify cancer drivers?



# How do we identify driver mutations?

- “driver alterations” vs. “passenger alterations”
- Design an experiment to prove a given alteration can induce cancer
  - Cell lines / Mouse models / tumor organoids / etc.

# How do we identify driver mutations?

- “driver alterations” vs. “passenger alterations”
- Design an experiment to prove a given alteration can induce cancer
  - Cell lines / Mouse models / tumor organoids / etc.
- *However, it is unfeasible to test >10,000 mutations!*

In the genomics era, we need strategies to prioritize

# How do we identify driver mutations?

- “driver alterations” vs. “passenger alterations”
- **Main idea:** *A cancer driver gene is a gene that is mutated more frequently than expected in a large tumor cohort*

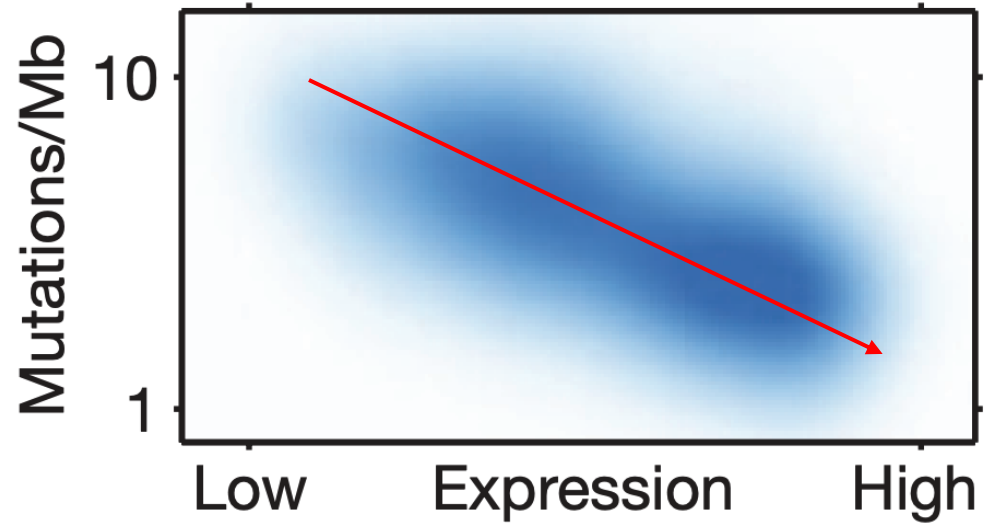
# How do we identify driver mutations?

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- **Main idea:** *A cancer driver gene is a gene that is mutated more frequently than expected in a large tumor cohort*

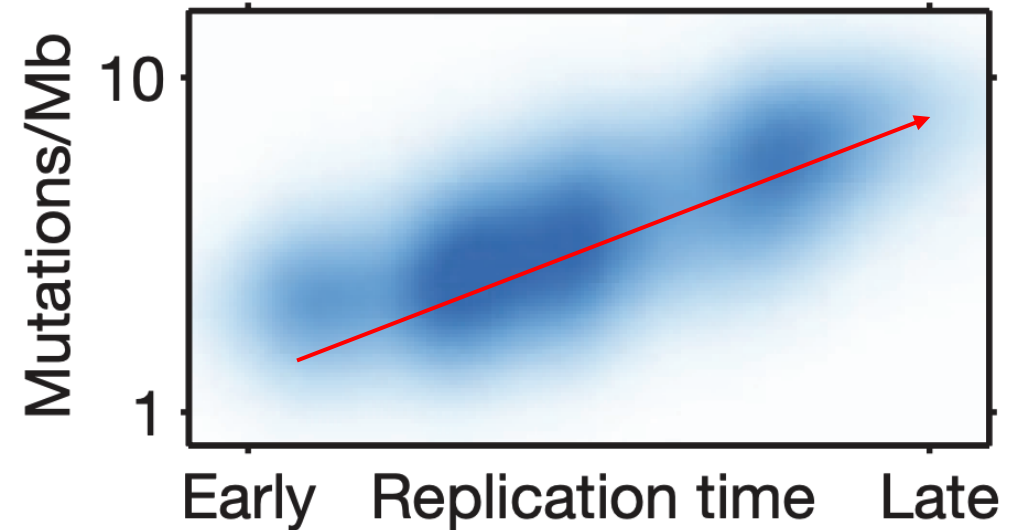
- Count the number of mutations observed in each gene in the cohort
- Determine the expected background mutation rate (BMR)
- Estimate significance of observed vs. expected number of mutations

# What does influence the BMR?

Gene expression



Replication Time





# What does influence the BMR?

- **Sample specific features**
  - Tissue-type
  - Impact of specific alterations
- **Underlying mutational processes**
  - (e.g. UV-light or tobacco consumption)
- **Regional genome properties:**
  - Gene expression
  - Replication time
  - Heterochromatin vs. euchromatin

# Estimating Coding Drivers in 2020

Review Article | Published: 10 August 2020

## **A compendium of mutational cancer driver genes**

Francisco Martínez-Jiménez, Ferran Muiños, Inés Sentís, Jordi Deu-Pons, Iker Reyes-Salazar, Claudia Arnedo-Pac, Loris Mularoni, Oriol Pich, Jose Bonet, Hanna Kranas, Abel Gonzalez-Perez  & Nuria Lopez-Bigas 

*Nature Reviews Cancer* **20**, 555–572(2020) | [Cite this article](#)

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What is the evidence of selection?

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### **Evidence of selection:**

- Recurrence ( it requires to estimate BMR )

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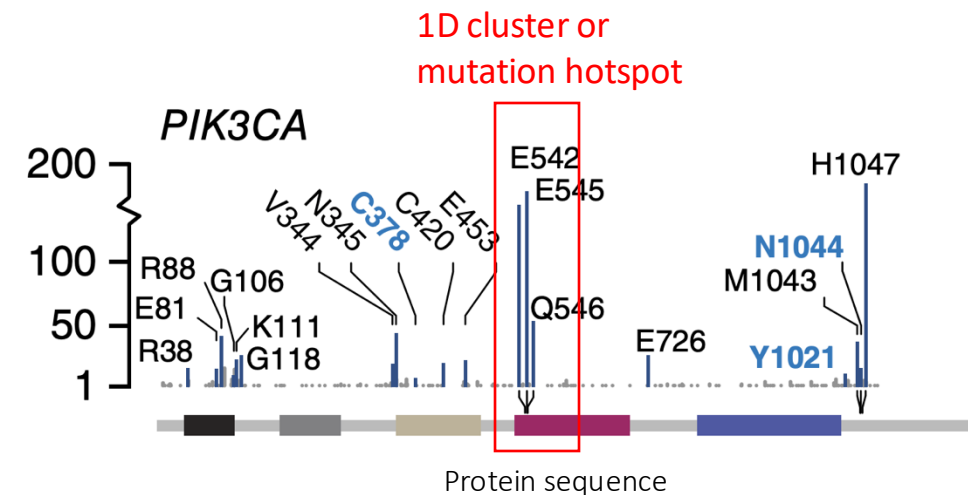
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### Evidence of selection:

- Recurrence ( it requires to estimate BMR )
- Distribution of mutations (1D and 3D clusters)



(Chang et al. Nat. Biotech 2016)

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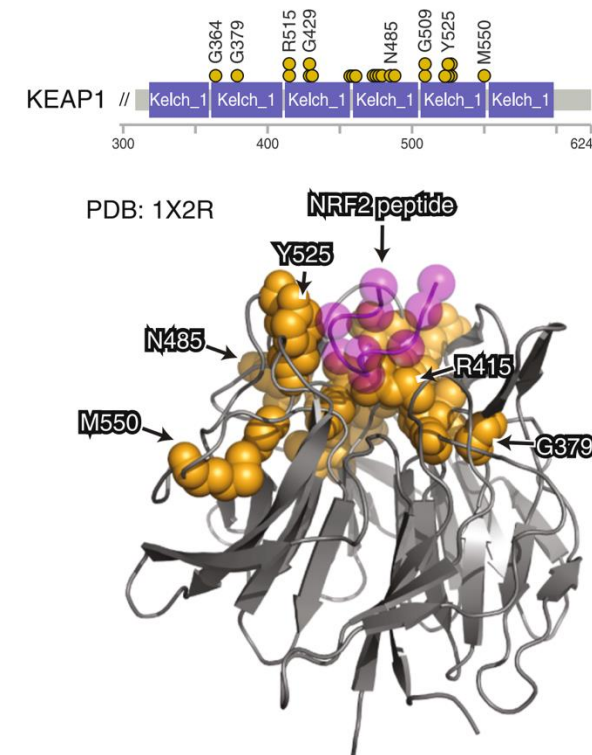
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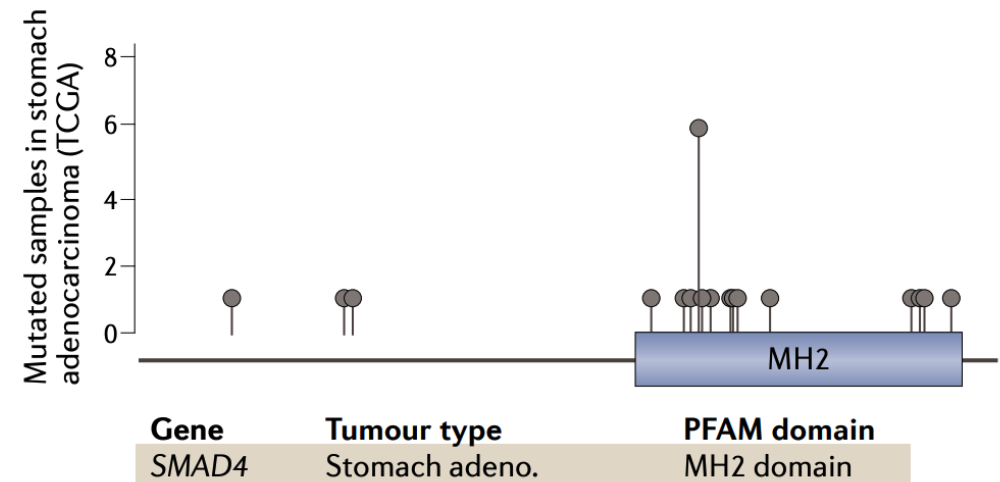
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### Evidence of selection:

- Recurrence ( it requires to estimate BMR )
- Distribution of mutations (1D and 3D clusters)
- Mutations in functional domains



# Estimating Coding Drivers in 2020

Review Article | Published: 10 August 2020

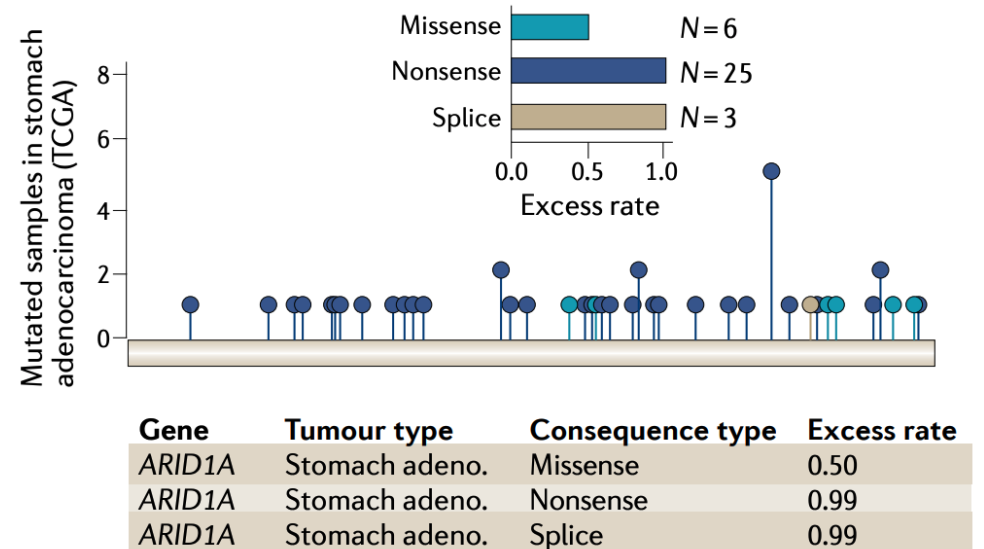
## A compendium of mutational cancer driver genes

Francisco Martínez-Jiménez, Ferran Muiños, Inés Sentís, Jordi Deu-Pons, Iker Reyes-Salazar, Claudia Arnedo-Pac, Loris Mularoni, Oriol Pich, Jose Bonet, Hanna Kranas, Abel Gonzalez-Perez & Nuria Lopez-Bigas

*Nature Reviews Cancer* 20, 555–572(2020) | [Cite this article](#)

### Evidence of selection:

- Recurrence ( it requires to estimate BMR )
- Distribution of mutations (1D and 3D clusters)
- Mutations in functional domains
- *Functional impact bias*
  - (also evolutionary conservation)



What did we learn about cancer from all these mutation analyses?

# *Exercise*

Exercise: 1.15h-2h30 Prepare the paper presentation  
2.30h-3h30 presentation of the paper

<https://www.nature.com/articles/s41586-020-1965-x>

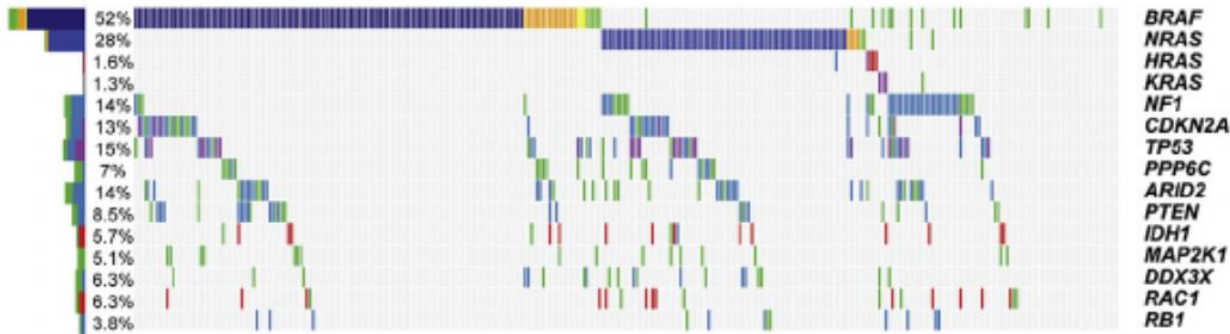
The pdf is in Moodle with a list of questions

Each group presents 1 figure, as it was organized in the previous exercise session.

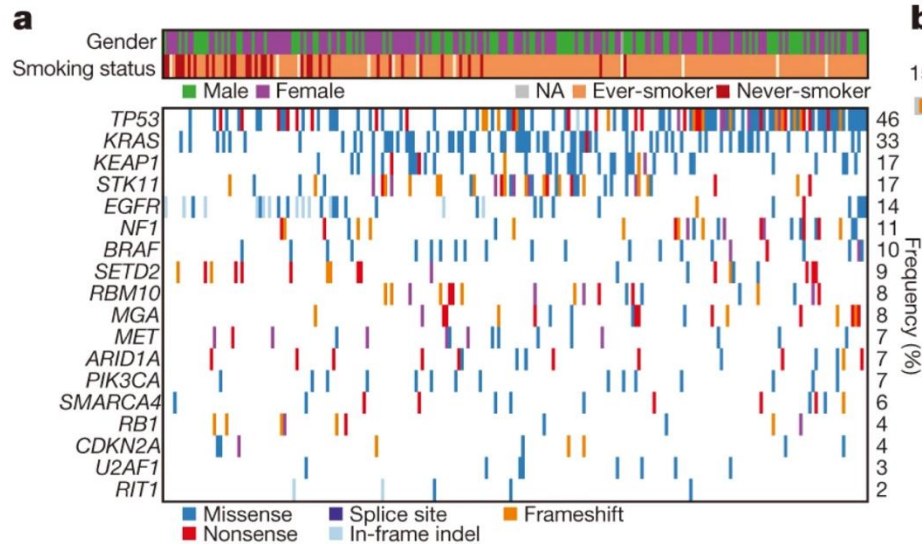
Address some of the questions

# Cancer heterogeneity between different tumor types

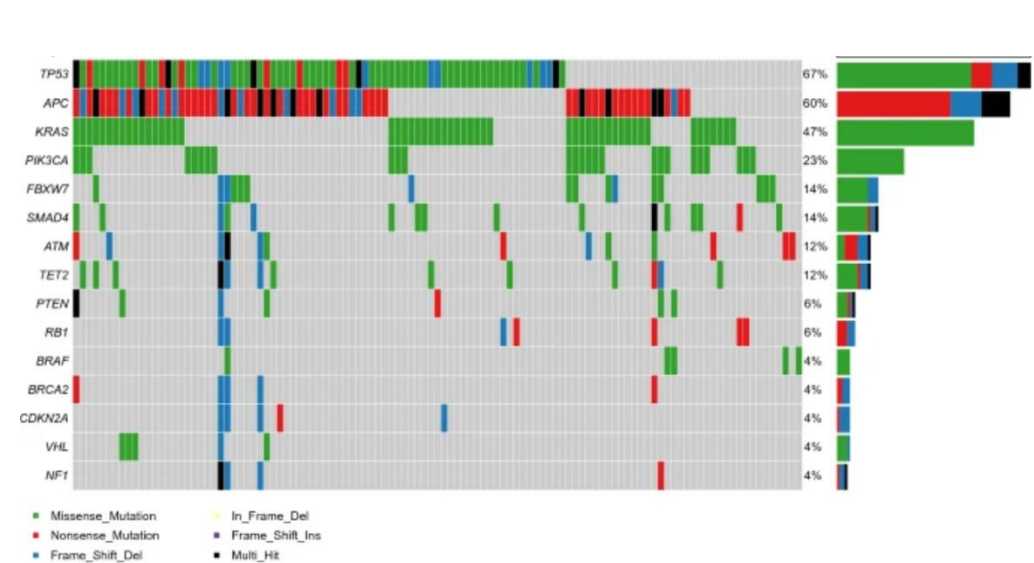
What are the most frequent mutated genes in each tissue?



Melanoma



Lung cancer

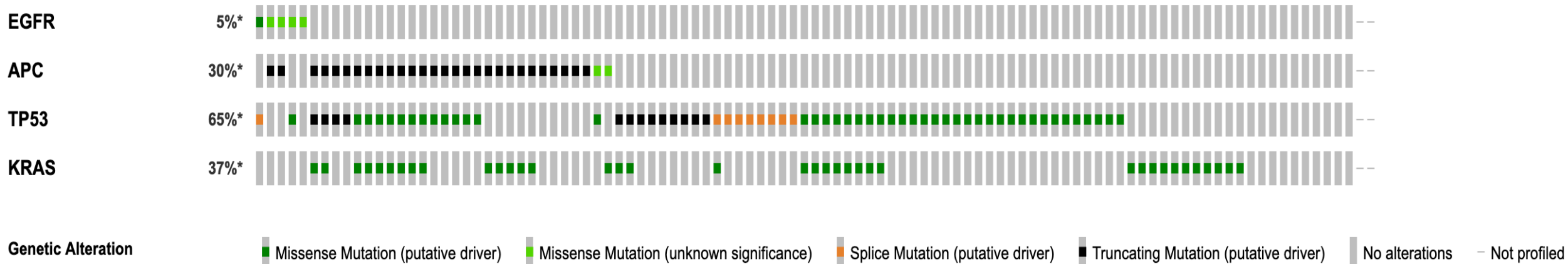


Colon cancer

# Cancer inter-patient heterogeneity

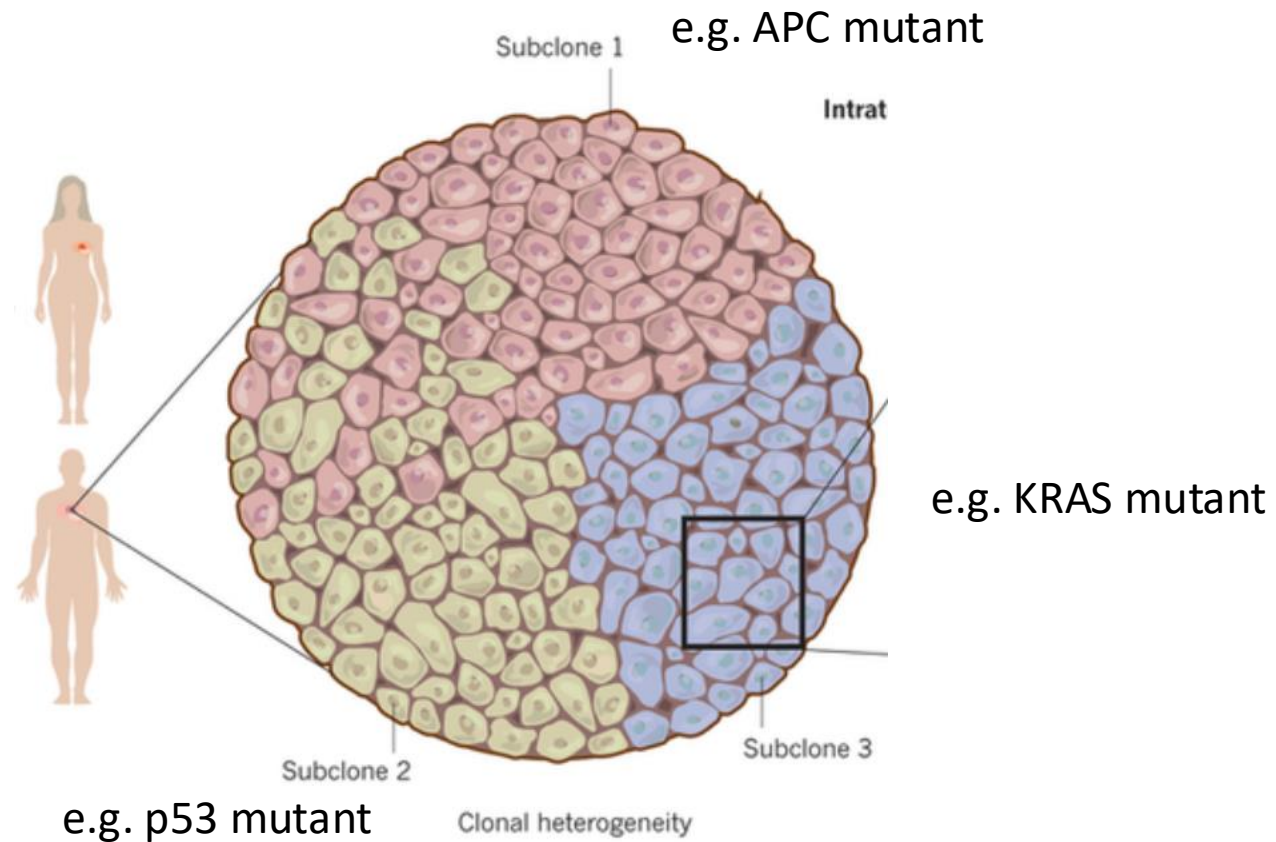
Same tumor: different mutation profile in each patient

103 patients colon cancer patient: each bar in this plot represents a patient



# Cancer intra tumor heterogeneity

The same patient in different areas of the tumor have different mutations



# *Exercise*

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