

Cryo Electron Microscopy (CryoEM)

Bilal M. Qureshi

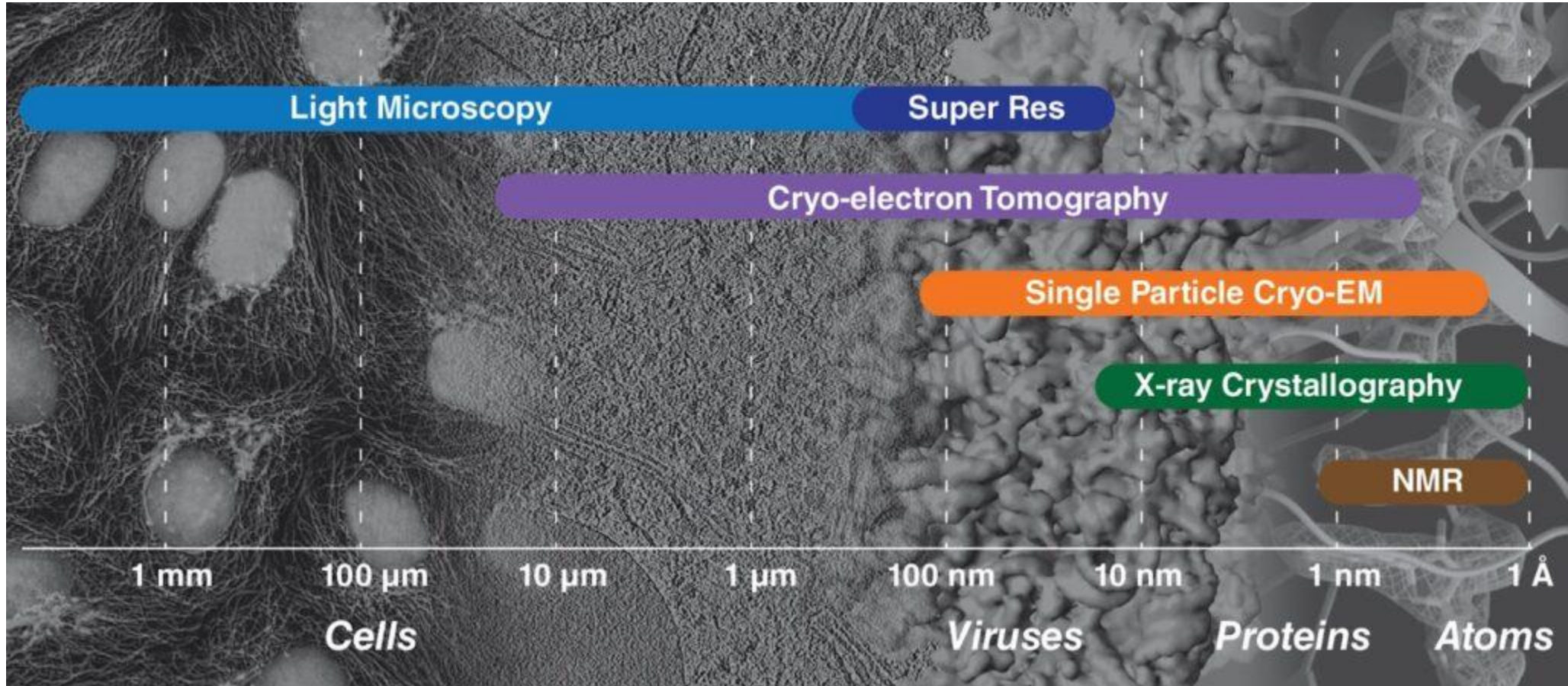
bilal.qureshi@scopem.ethz.ch

CCMX 2022

5th November 2024

- Why cryoEM?
- Radiation damage and low dose EM
- Representative projects:
 - Soft matter
 - Protein complexes
 - hard matter/nanoparticles
- See Demo on Instruments for details about (with Stephan Handschin, Miroslav Peterek):
 - Vitrification
 - Microscopy at cryogenic conditions
 - High throughput and automation

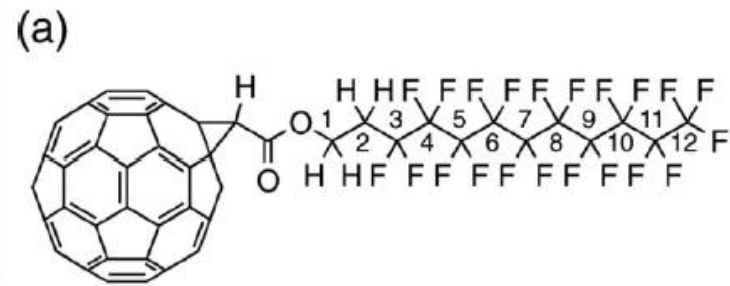
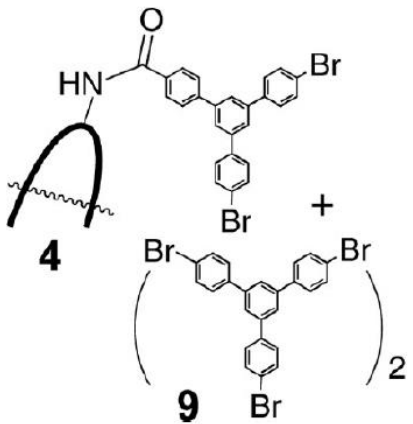
Visualization of biological or soft materials



Cryogenic (transmission) Electron Microscopy

Why EM: The wavelength of electron is very short and thus EM has high resolving power

Why Cryo: Water in biological/soft material samples is retained by rapid freezing (vitrification) making the samples compatible with high vacuum of an electron microscope. Moreover, vitrification preserves near native conformation (structure) of such samples



Scope  Are biological/soft matter samples suitable for electron microscopy?

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Liquids will evaporate in vacuum of TEM



Beam will destroy sample
(temperatures as on the sun)



Scope Are biological/soft matter samples suitable for electron microscopy?

Liquids will evaporate in vacuum of TEM
→ Requires fixing of samples



Beam will destroy sample
(temperatures as on the sun)
→ Requires (i) vitrification, (ii) low-dose
and (iii) rapid data collection (movie
mode) before beam damage

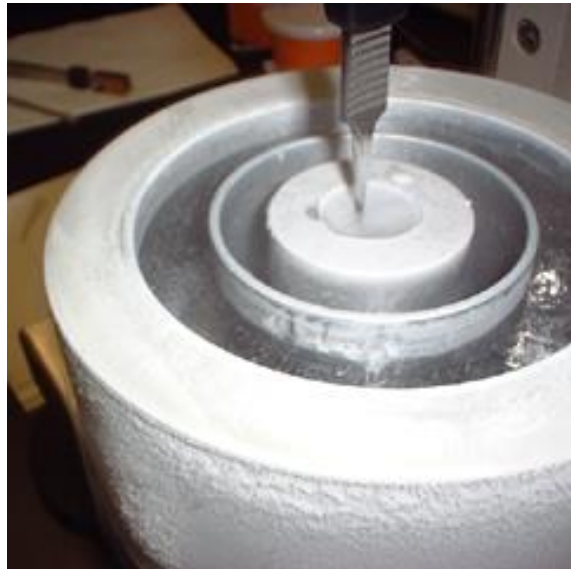


Vitrification and beam damage



Liquid Ethane

Liquid Nitrogen



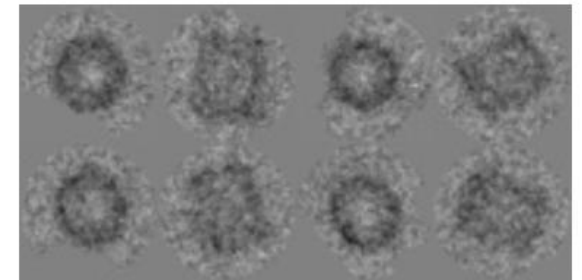
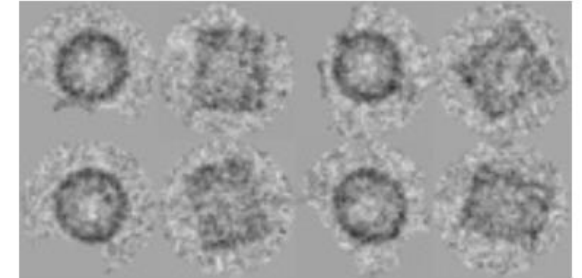
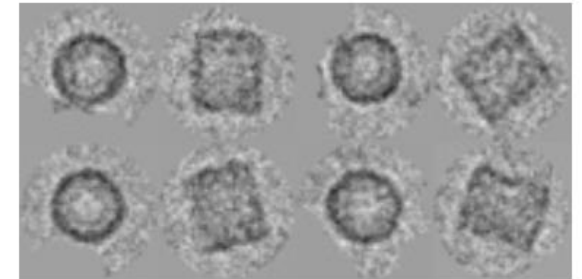
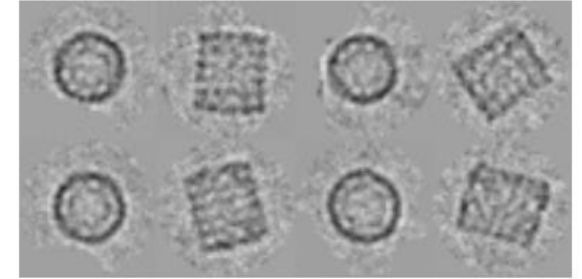
10 or 20 e^-/A^2

120 e^-/A^2

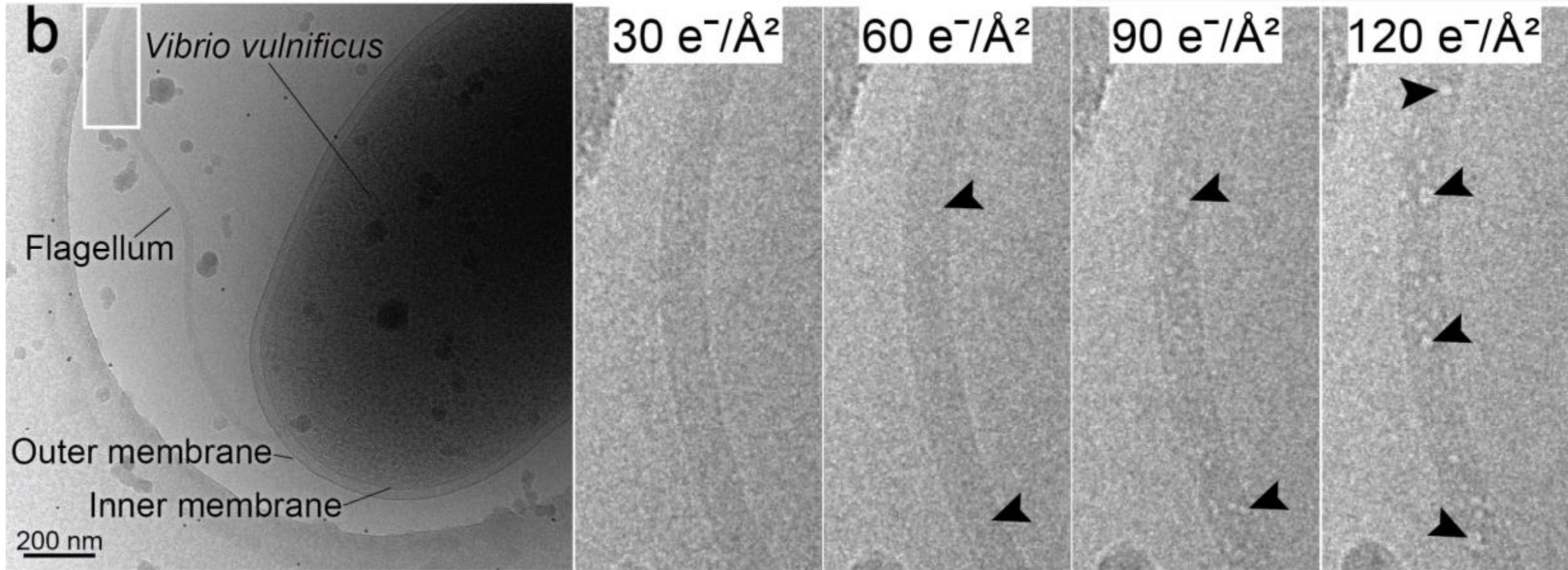
200 e^-/A^2

350 e^-/A^2

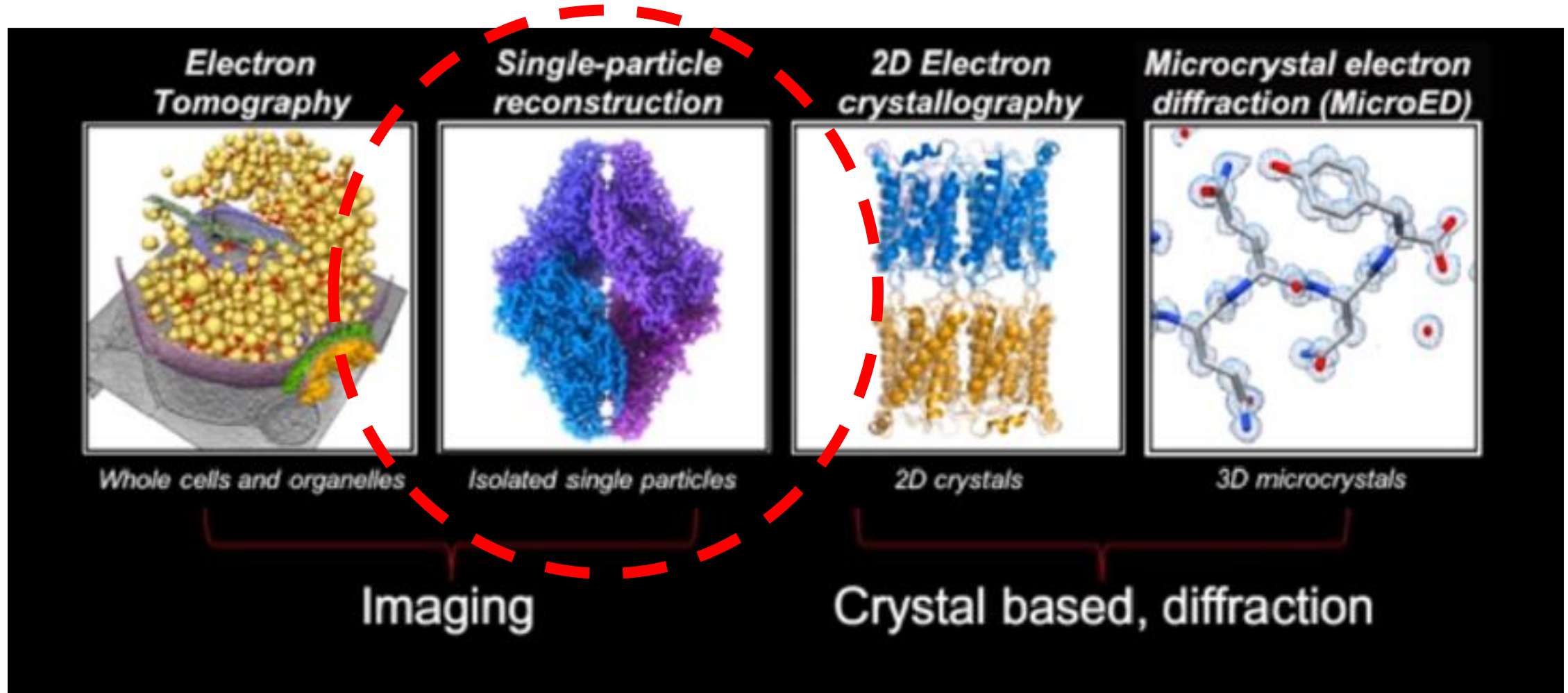
82 K (liquid N₂ cooling)



Vitrification and beam damage

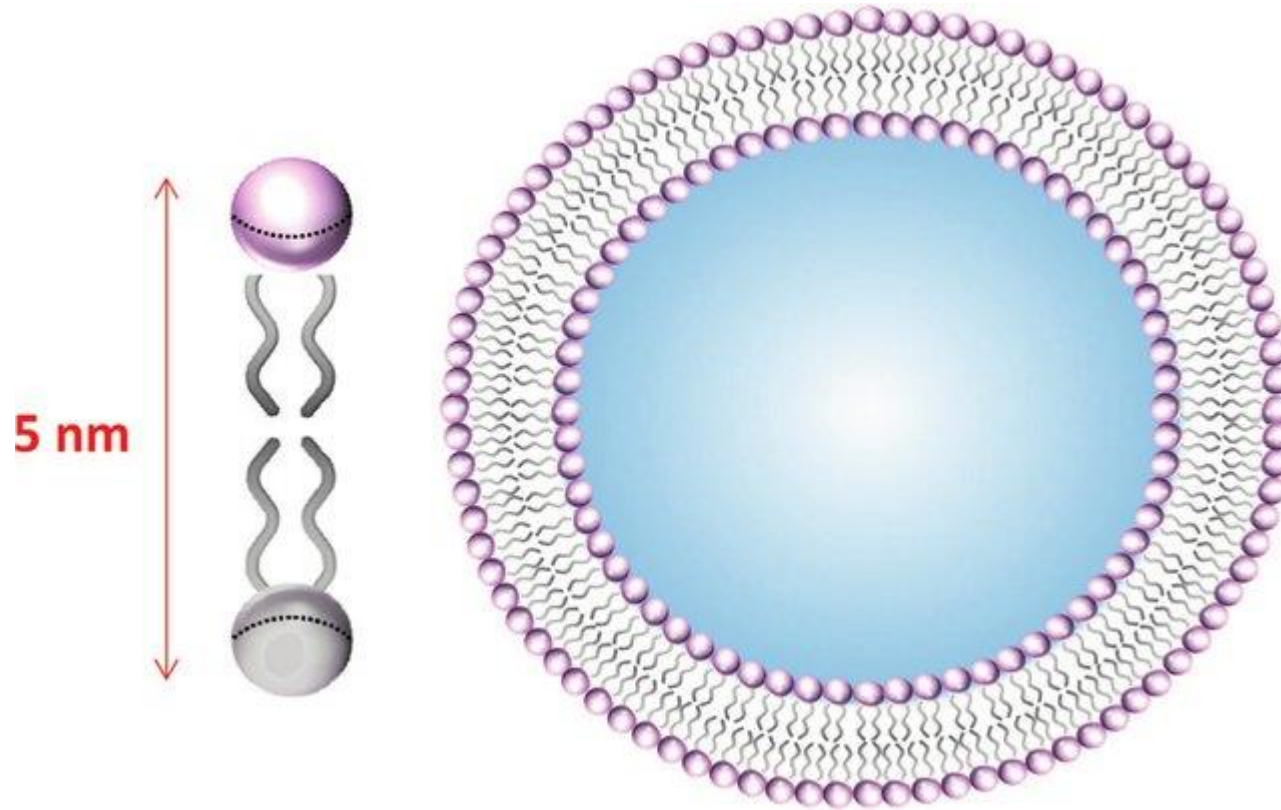


CryoEM: four basic “3D techniques” plus some variations!



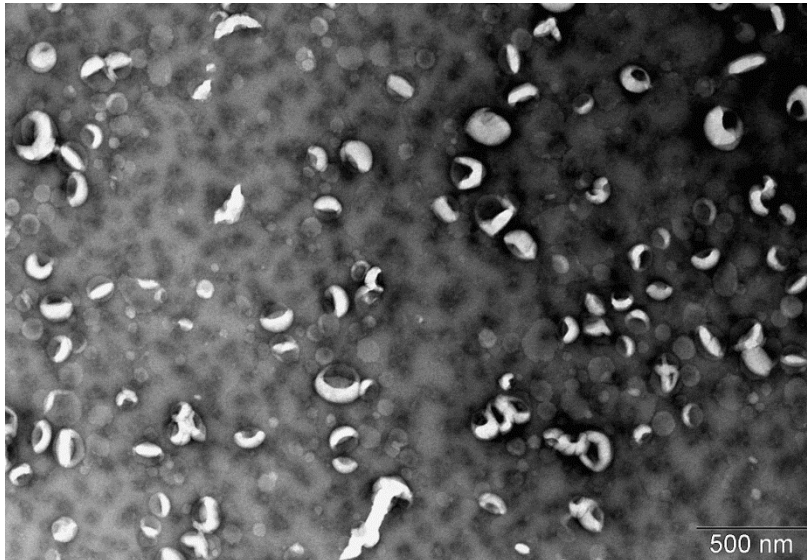
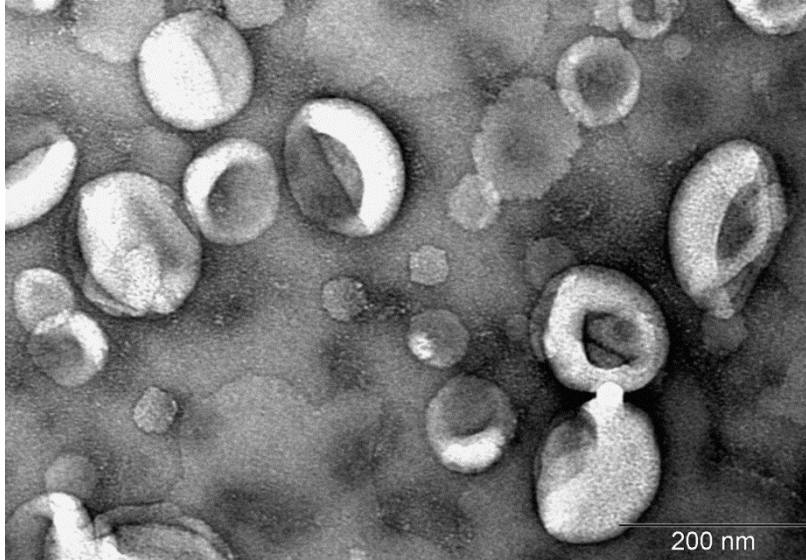
Representative projects...

Project 1: Vesicles



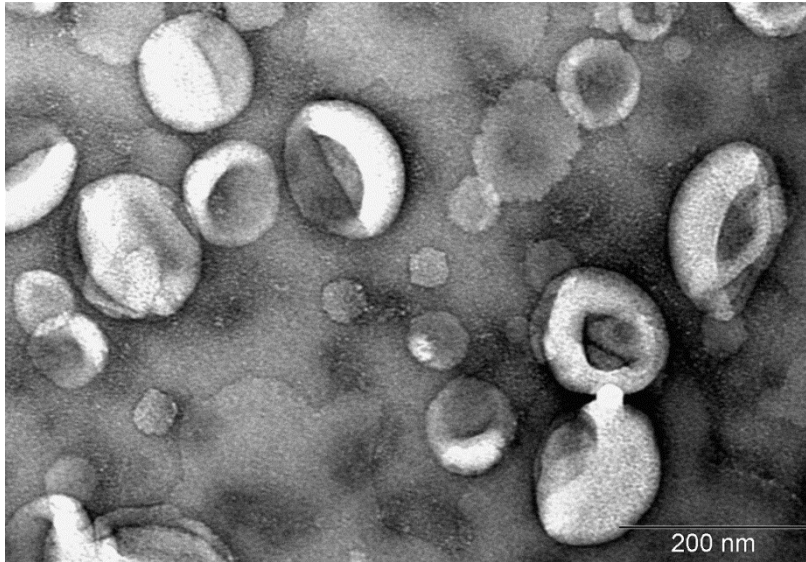
Project 1: Vesicle structural analysis

Room temperature/stained

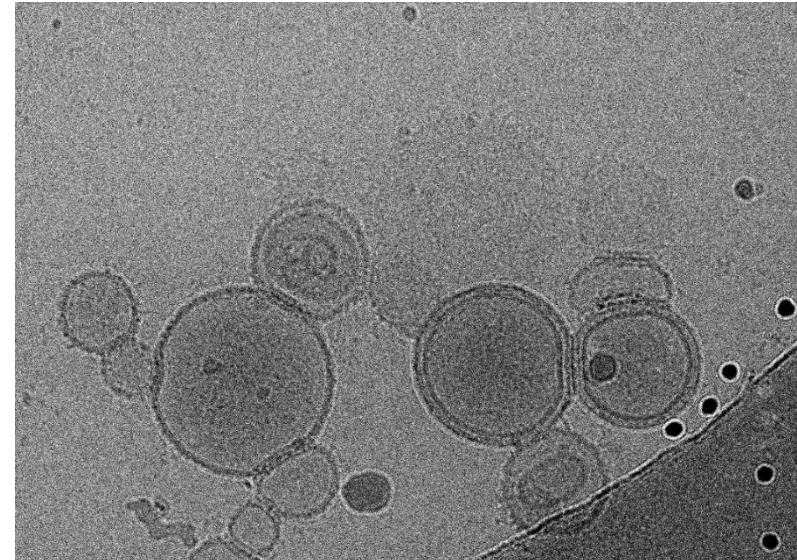
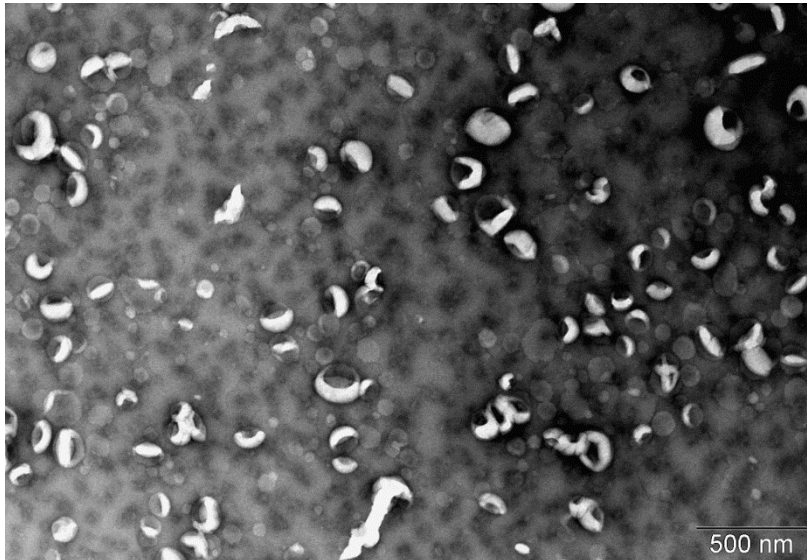
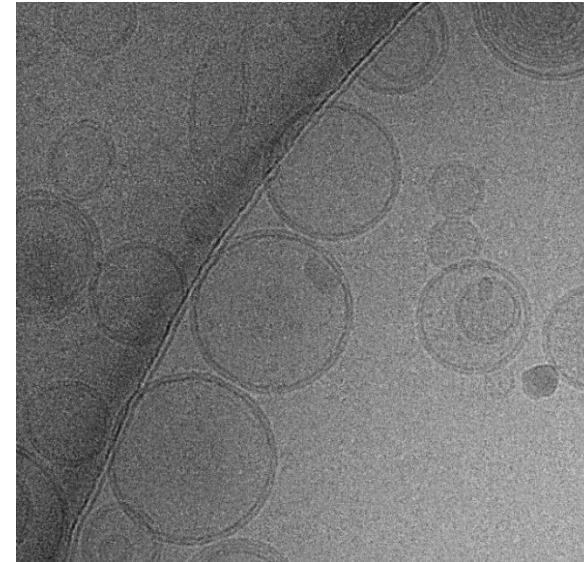


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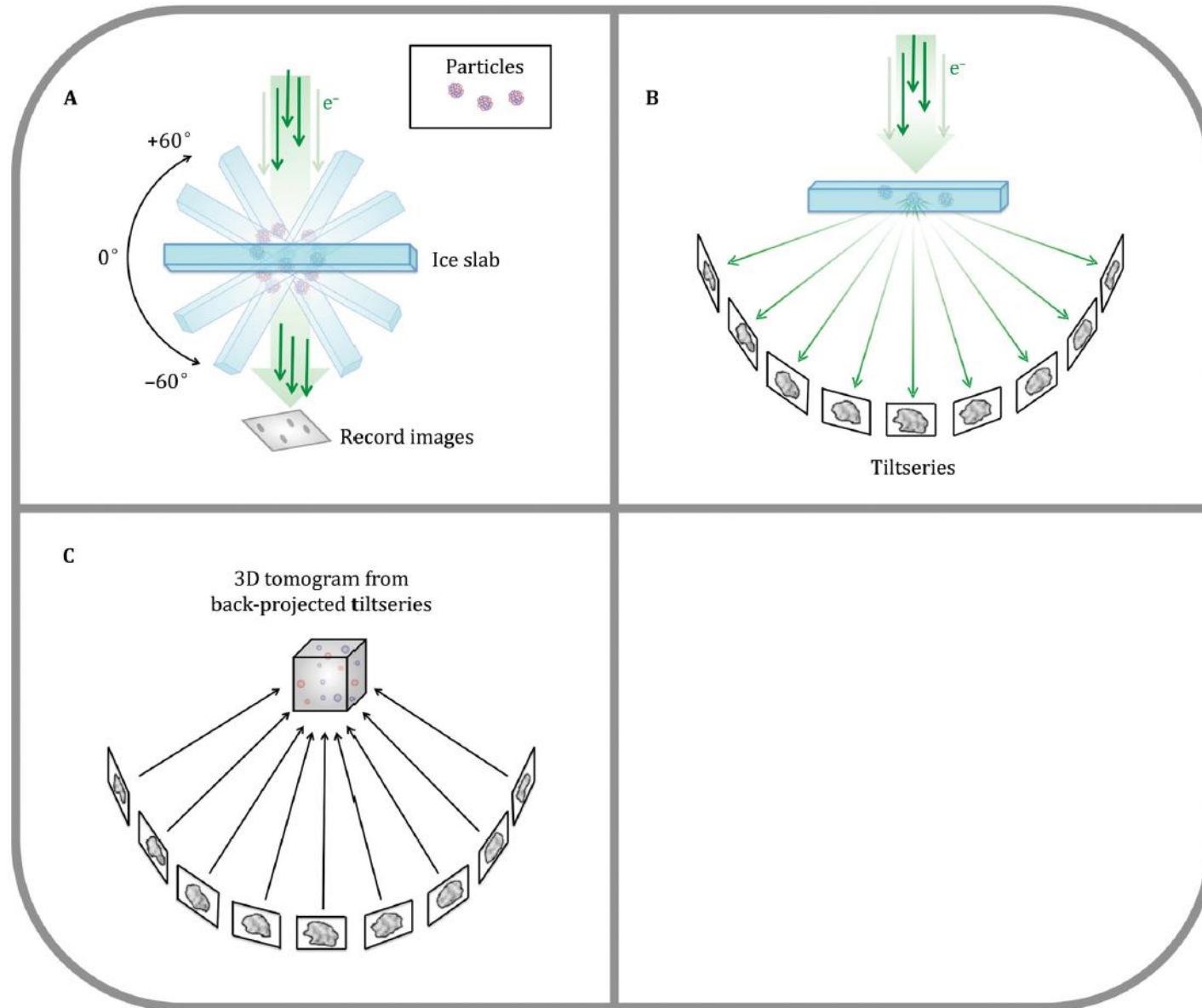
Room temperature/stained



CryoEM

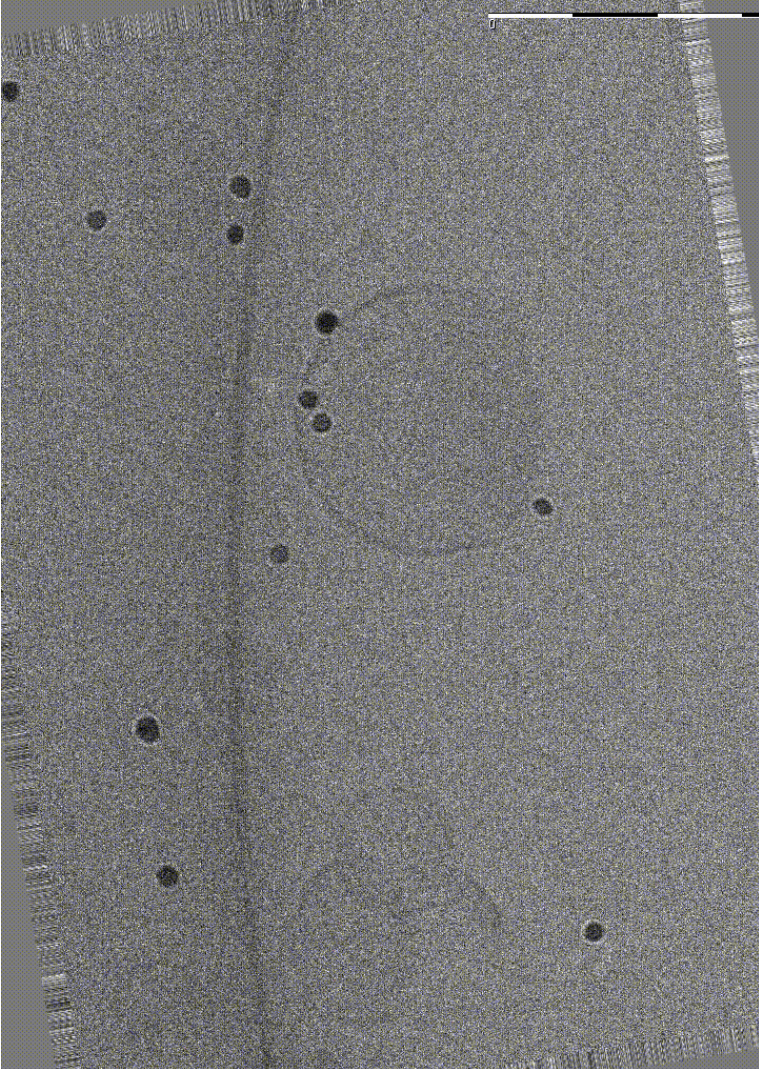


Cryo tilt series & electron tomography

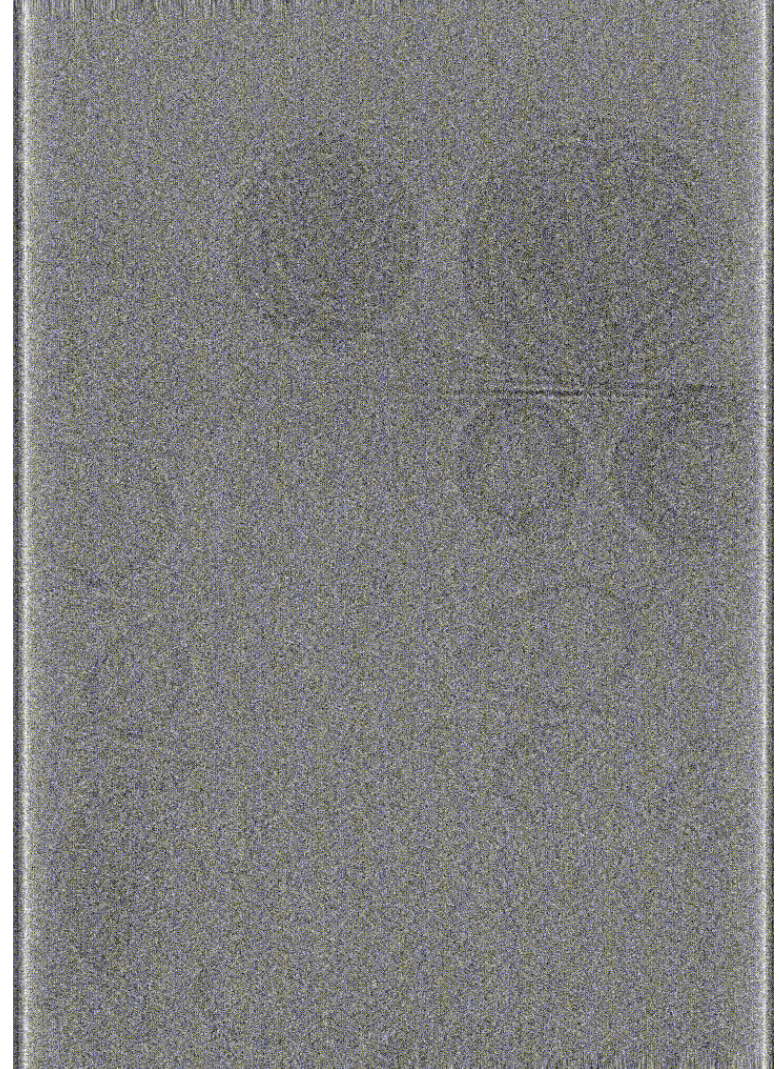


Project 1: Vesicle structural analysis

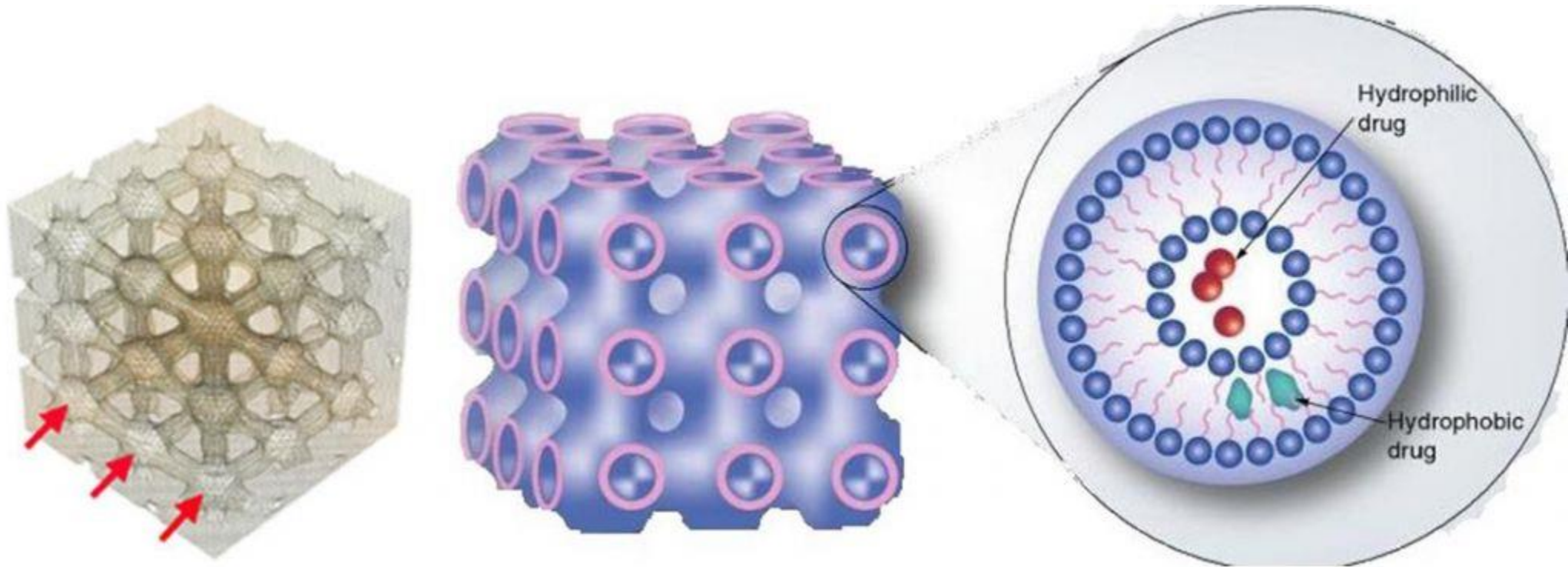
Movie1



Movie2



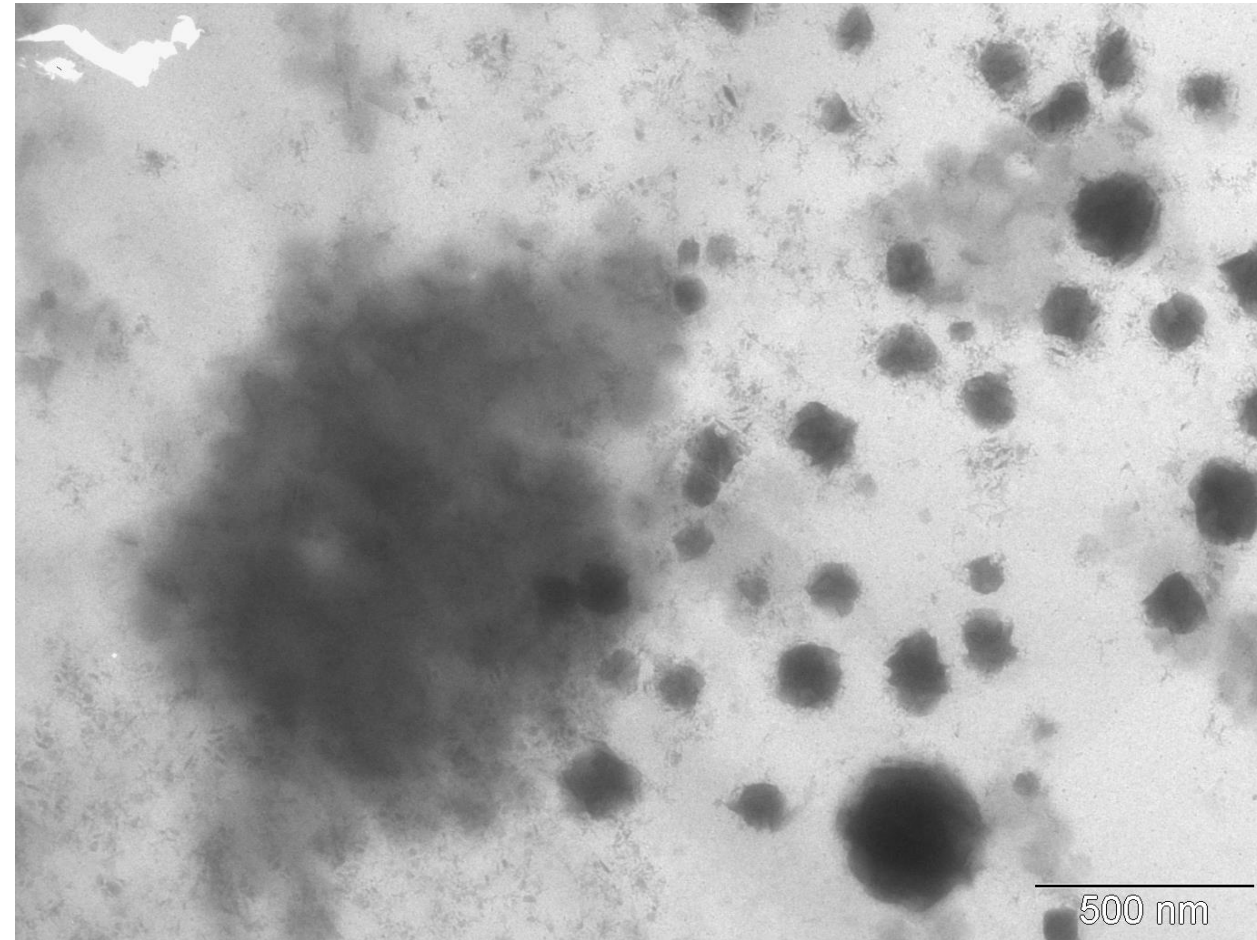
Project 2: Cubosomes



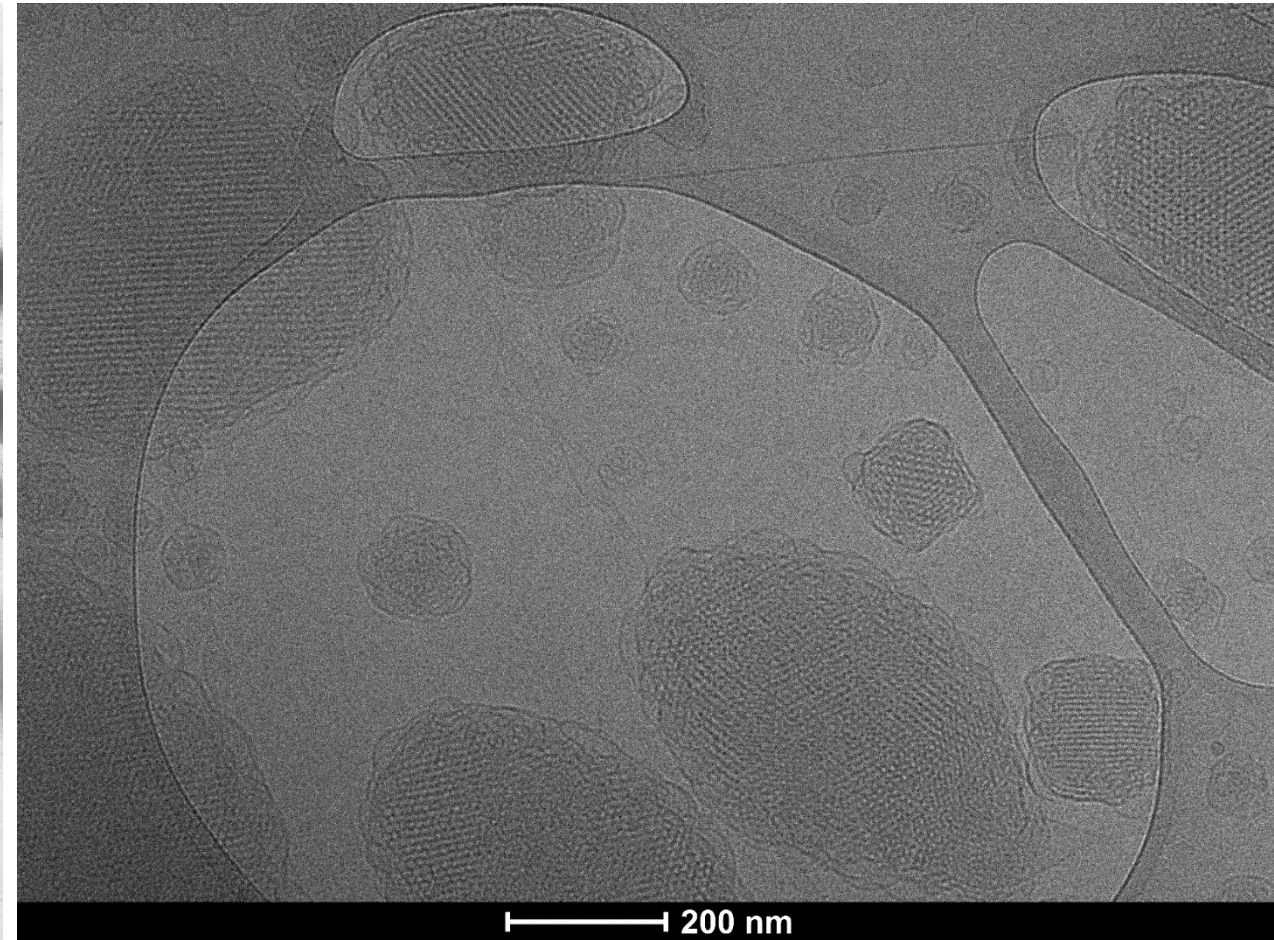
Cubosomes are discrete, sub-micron, nanostructured particles of the bicontinuous cubic liquid crystalline phase. The term "bicontinuous" refers to two distinct hydrophilic regions separated by the bilayer (Wikipedia)

Project 2: Cubosome structural analysis

Room temperature/stained



CryoEM



Project 3: Proteins



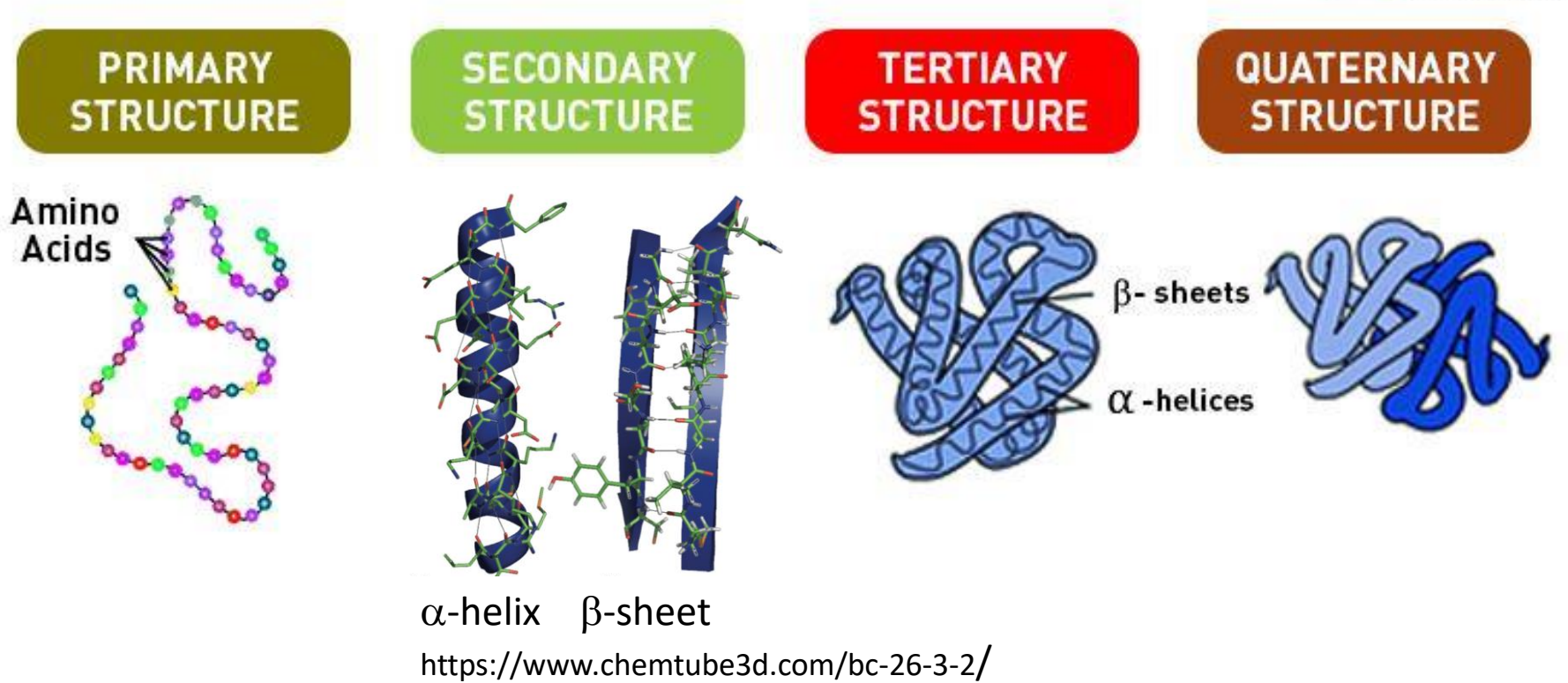
Protein shake (source: Wikipedia)

Project 3: Proteins

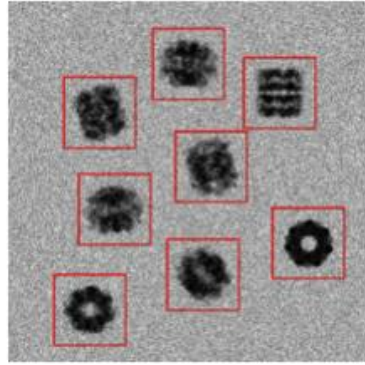
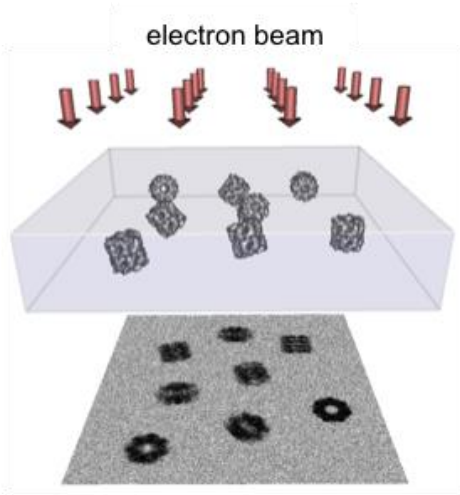
<https://byjus.com/biology/proteins-structure-and-functions/>



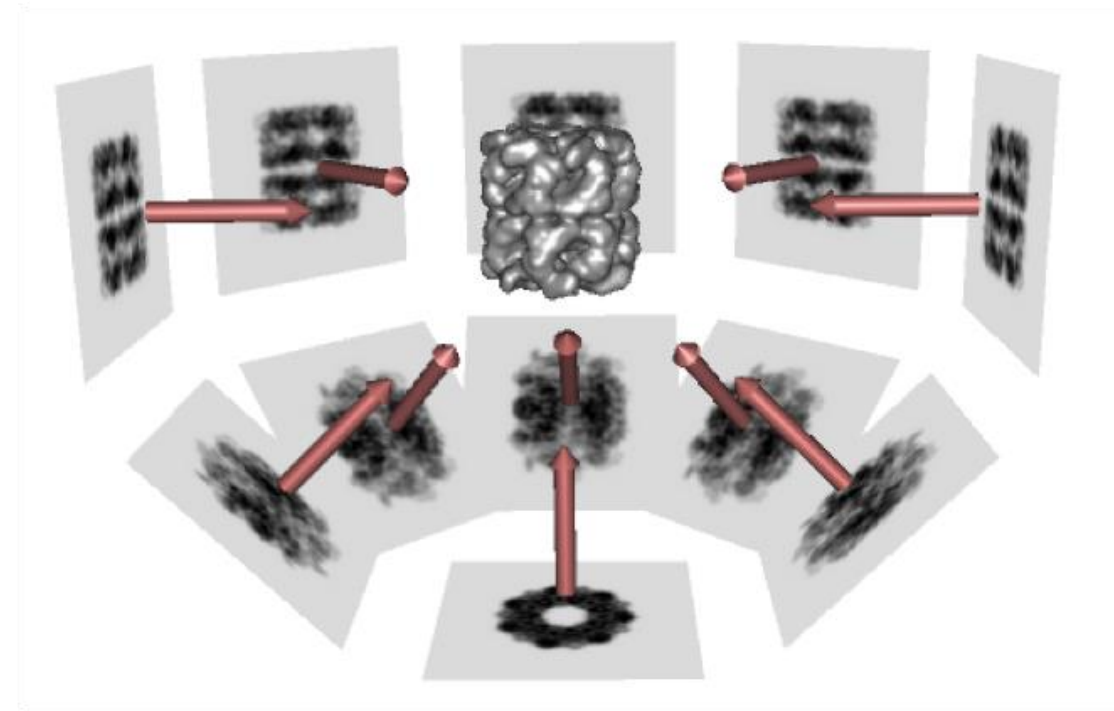
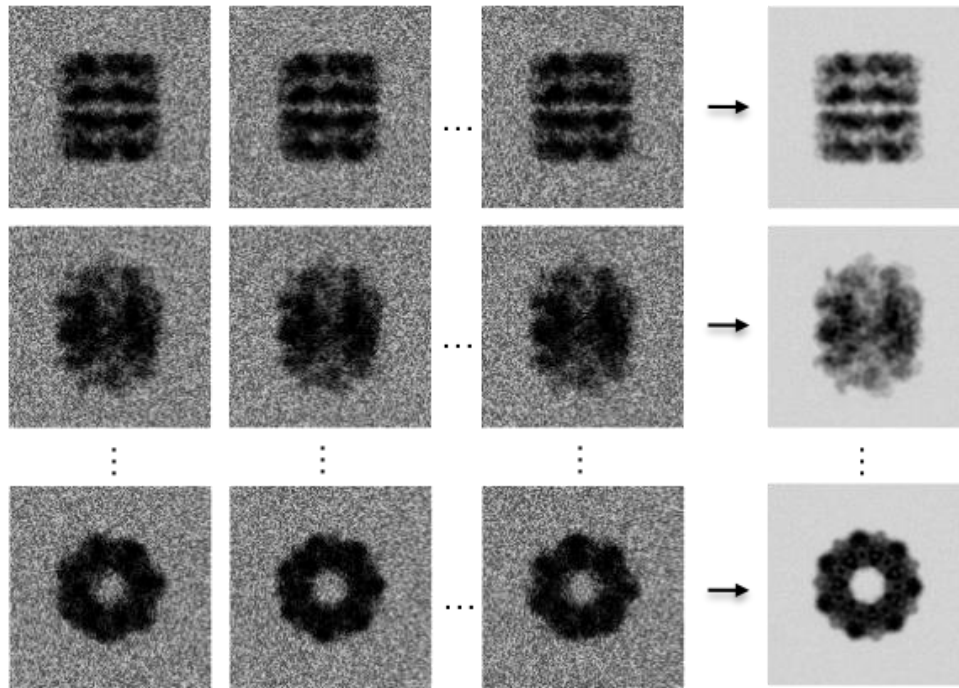
Protein shake (source: Wikipedia)



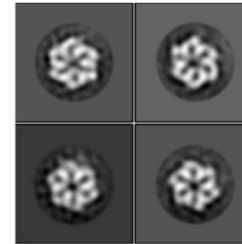
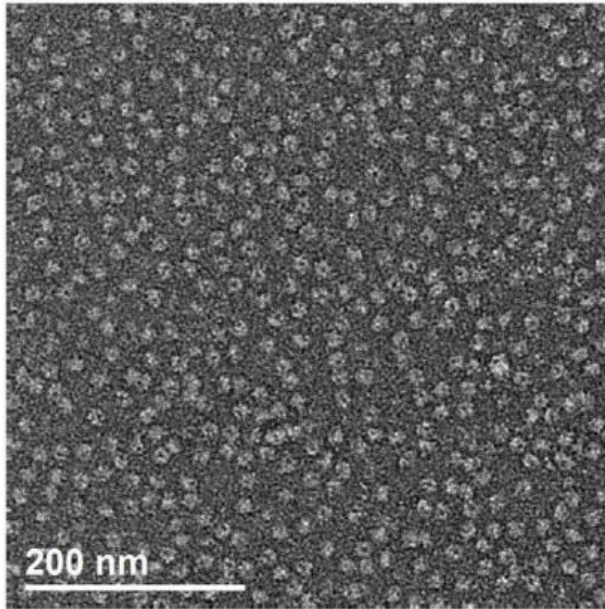
Project 3: Single Particle Analysis (SPA)



→ Reconstruct 3D map from “back projected” 2D projections

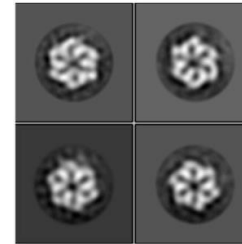
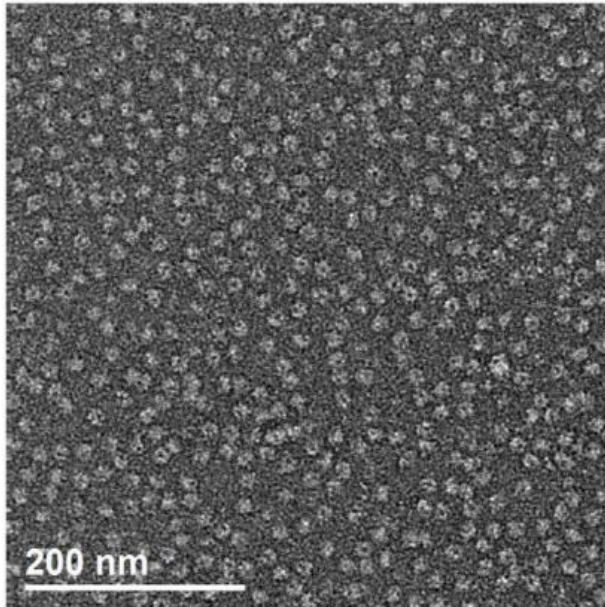


Project 3: Single Particle Analysis (SPA)

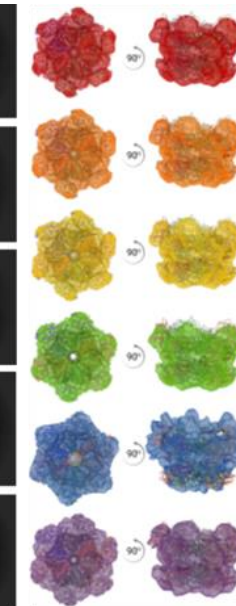
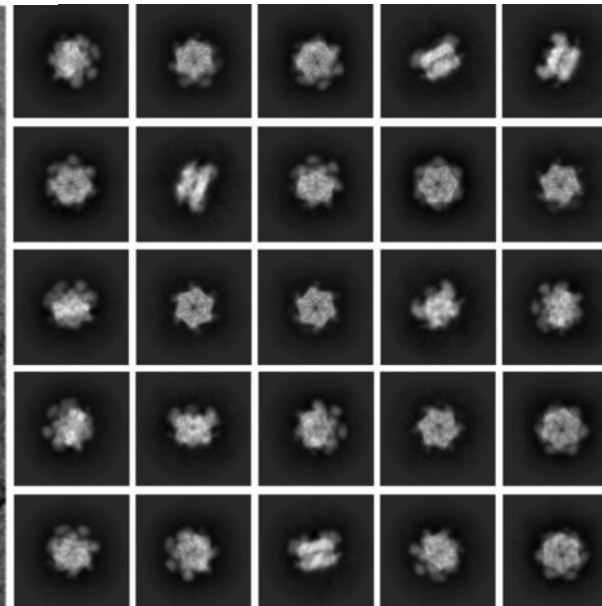
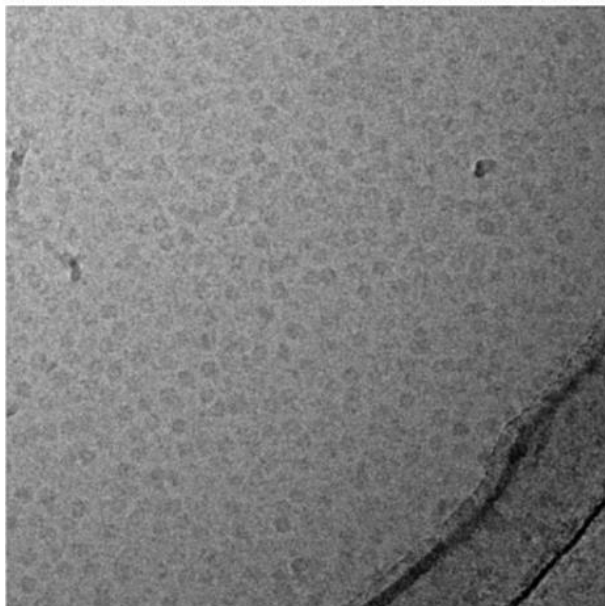


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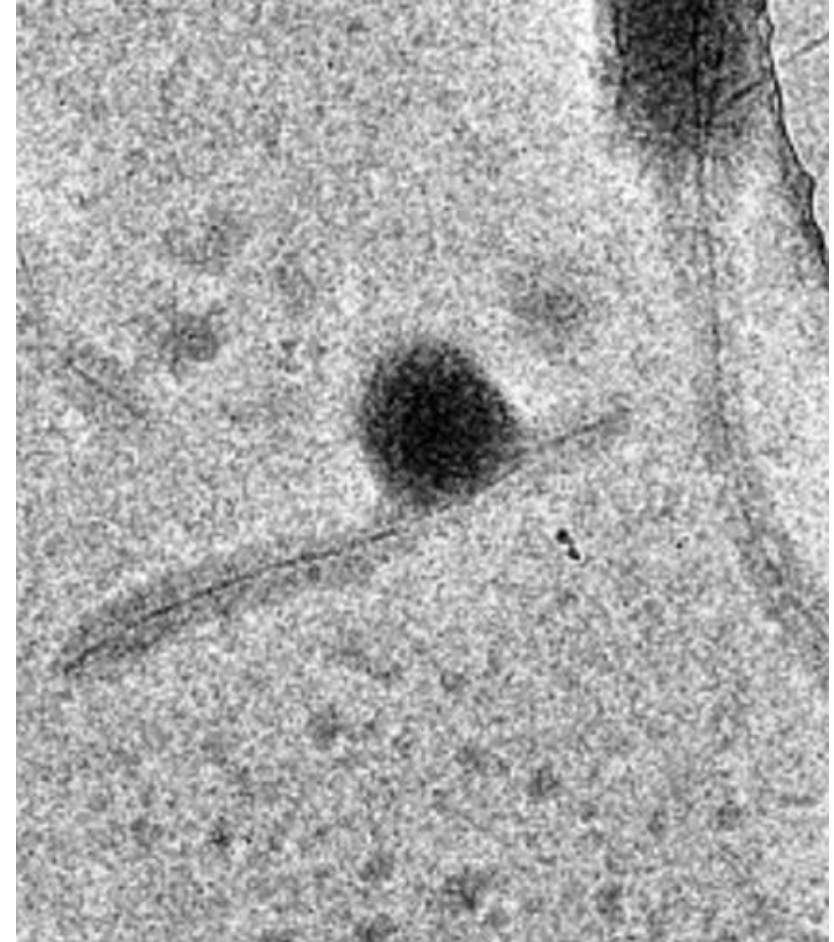
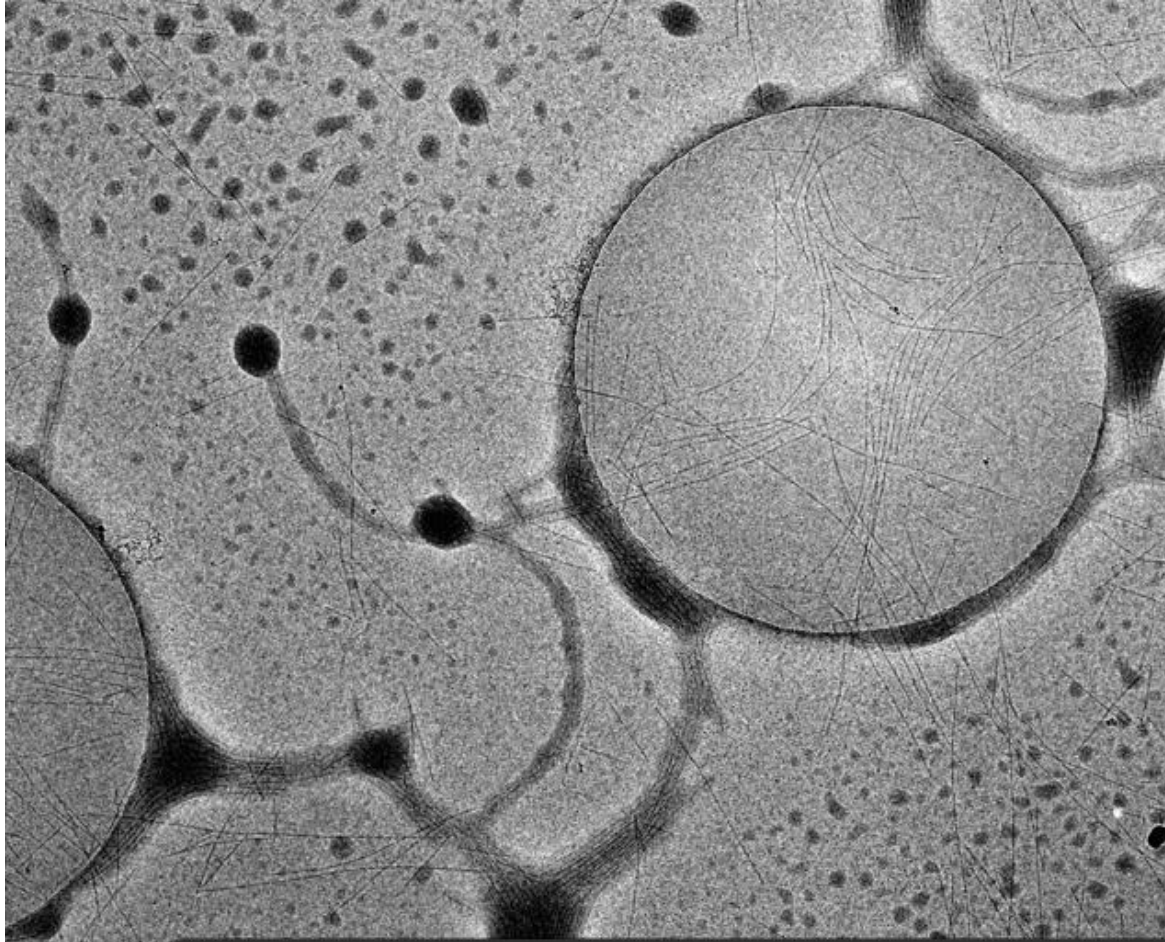
Room temperature/stained



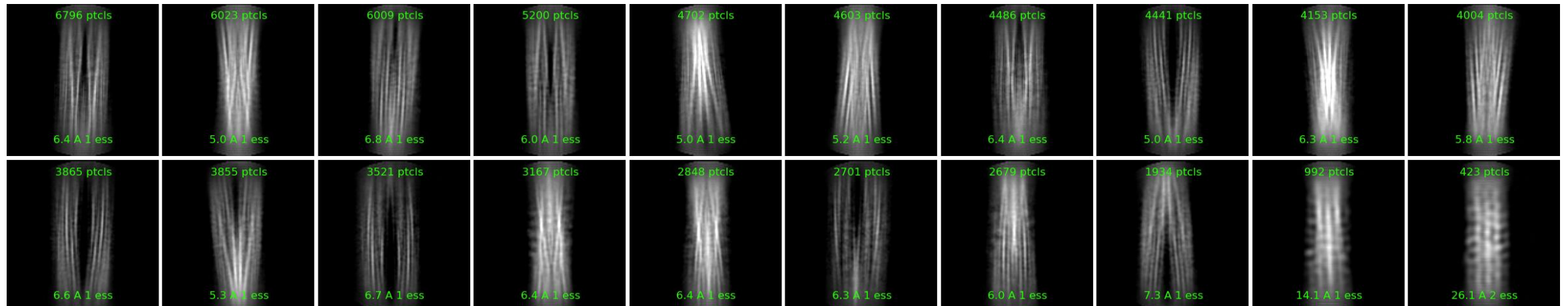
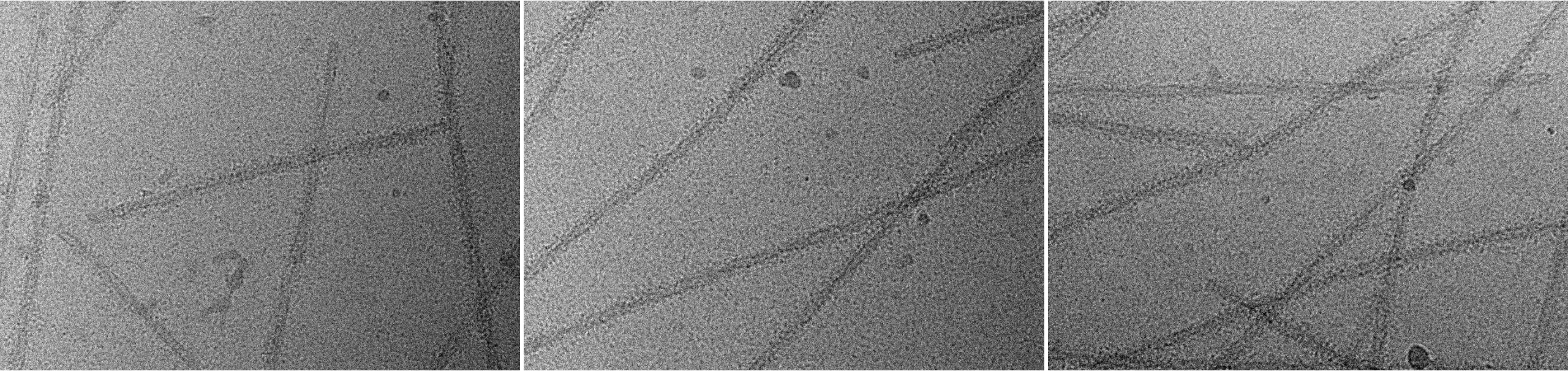
Cryo



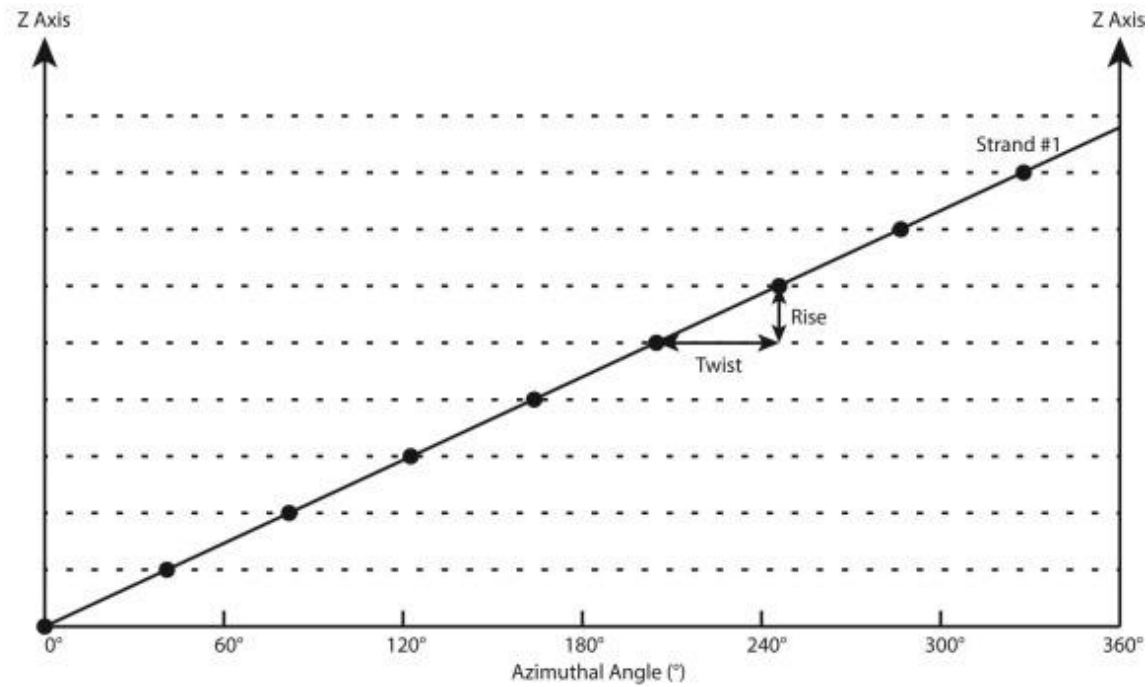
Project 4: Helical Reconstruction



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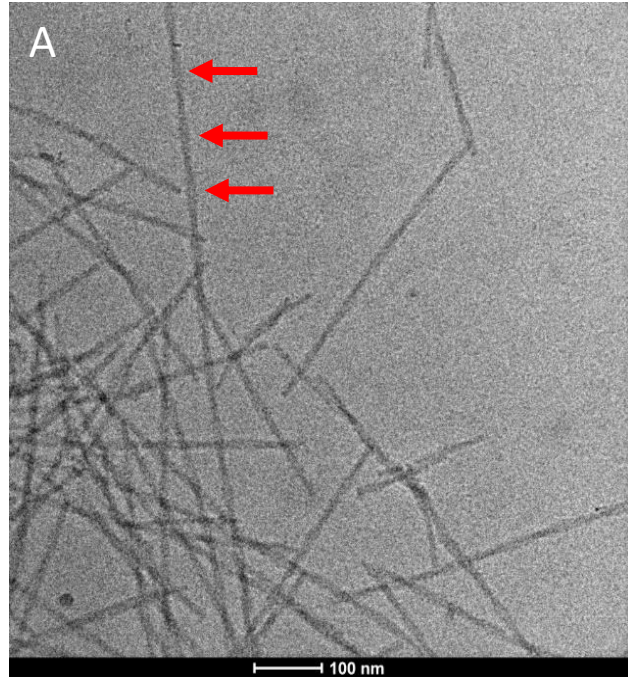


Twist and Rise parameters

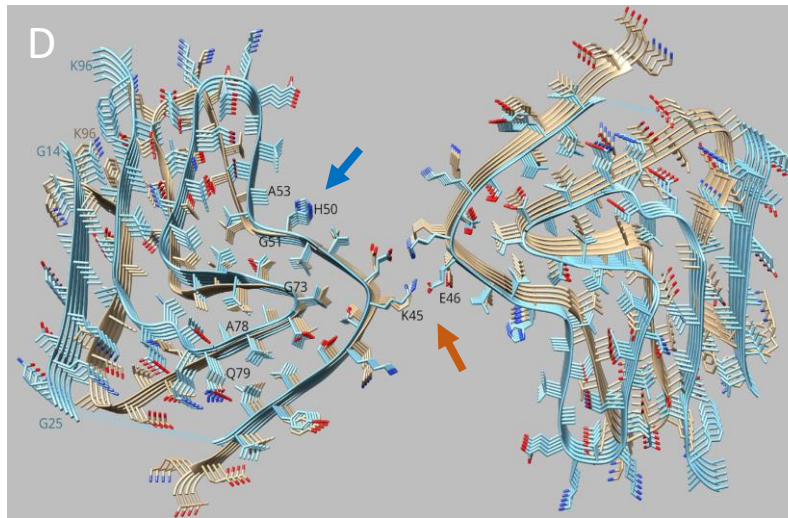
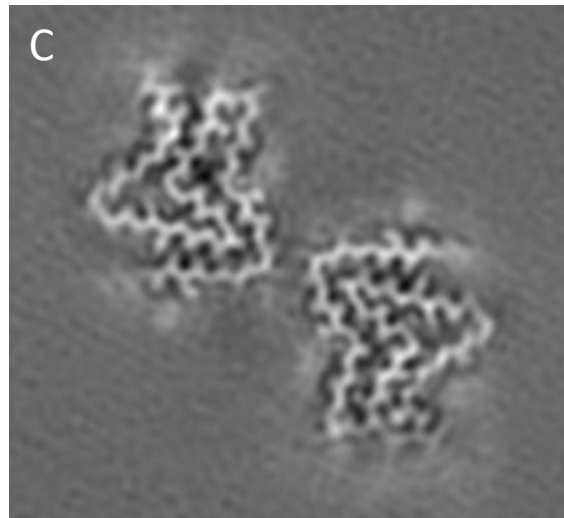
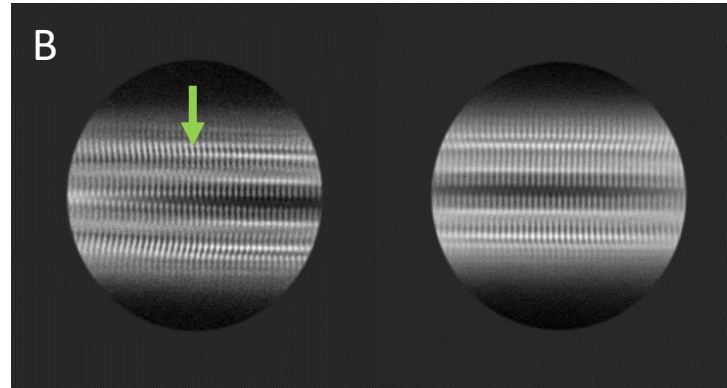


→ Symmetry operations of translation and rotation in a screw like fashion reconstruct the helix

Collaboration with Riek Group, ETH Zurich



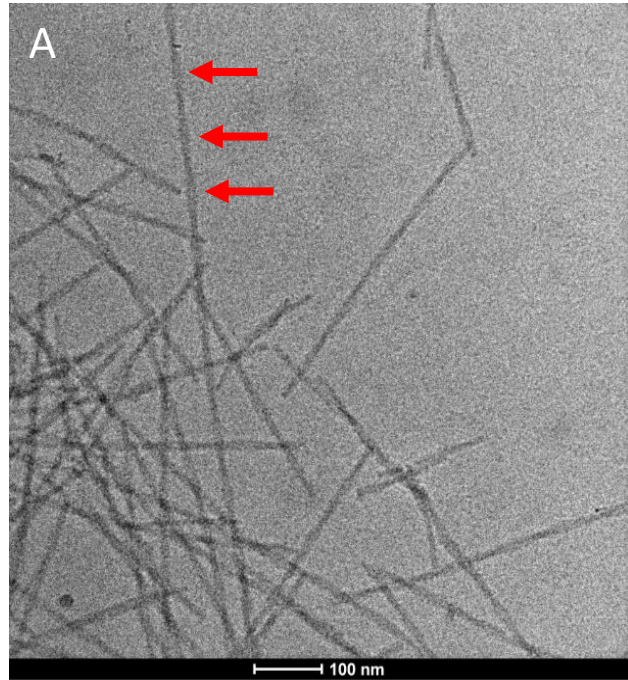
A Representative cryo-EM image of the α -synuclein fibrils showing twist of the two strands (red arrows), B Representative 2D class-average images showing striations visible due to helical arrangement of the monomers (green arrow pointing to a monomer) in the two protofibrils, C Cross-section of cryo-EM map showing the relative orientation of the monomers D Model structures (gold) compared with published structure (aquamarine)



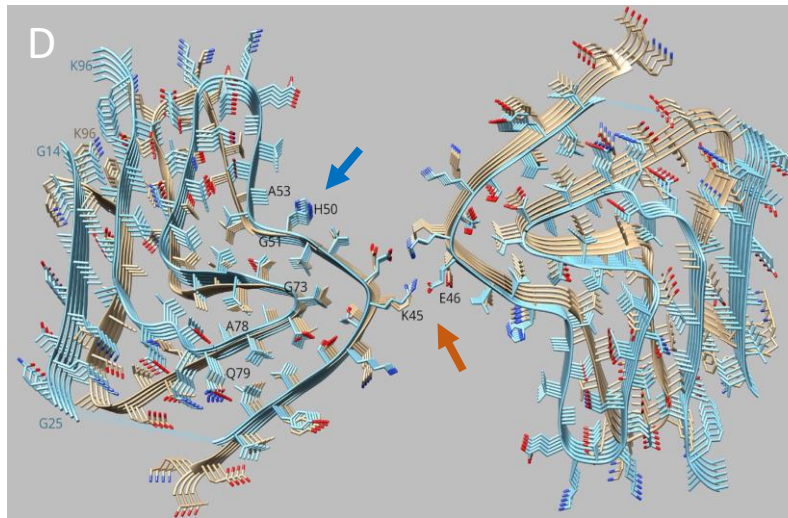
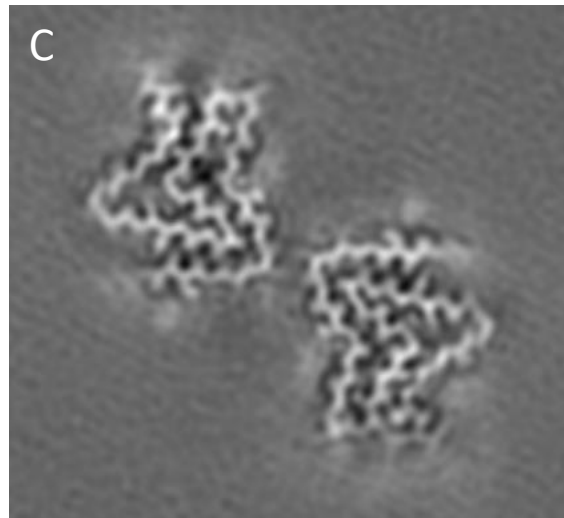
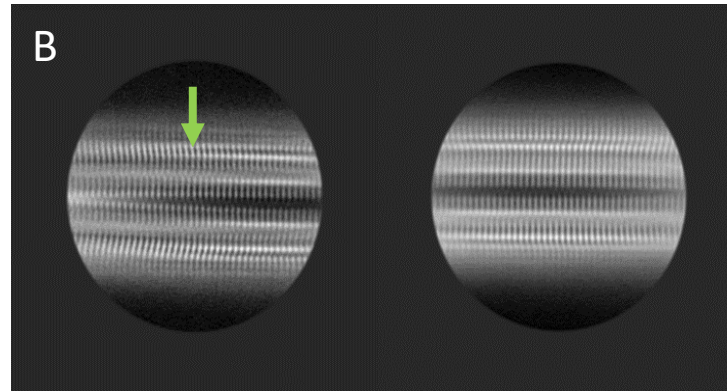
Movies



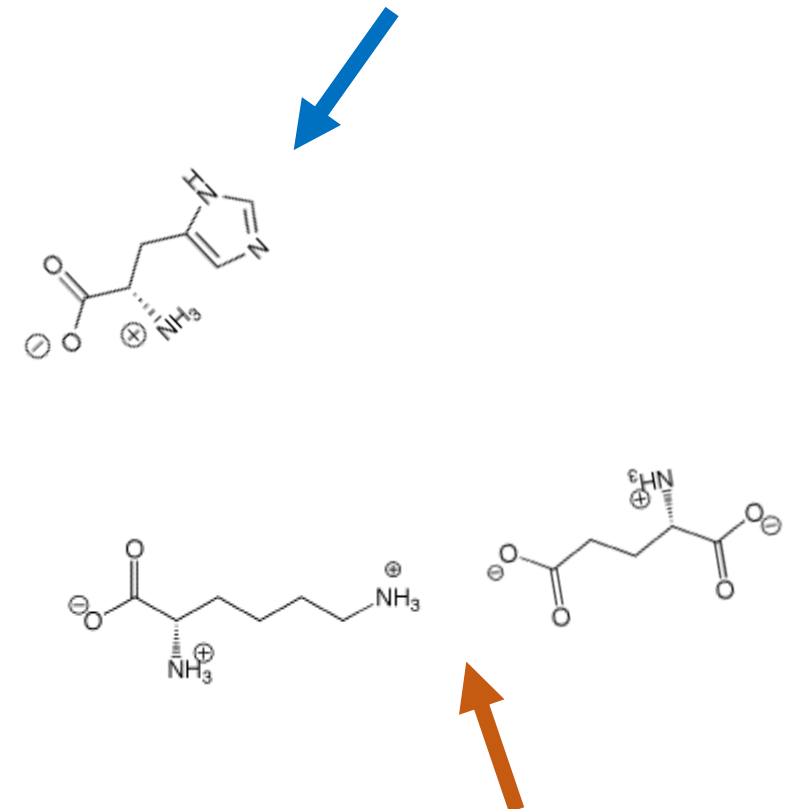
Collaboration with Rolad Riek Group, ETH Zurich



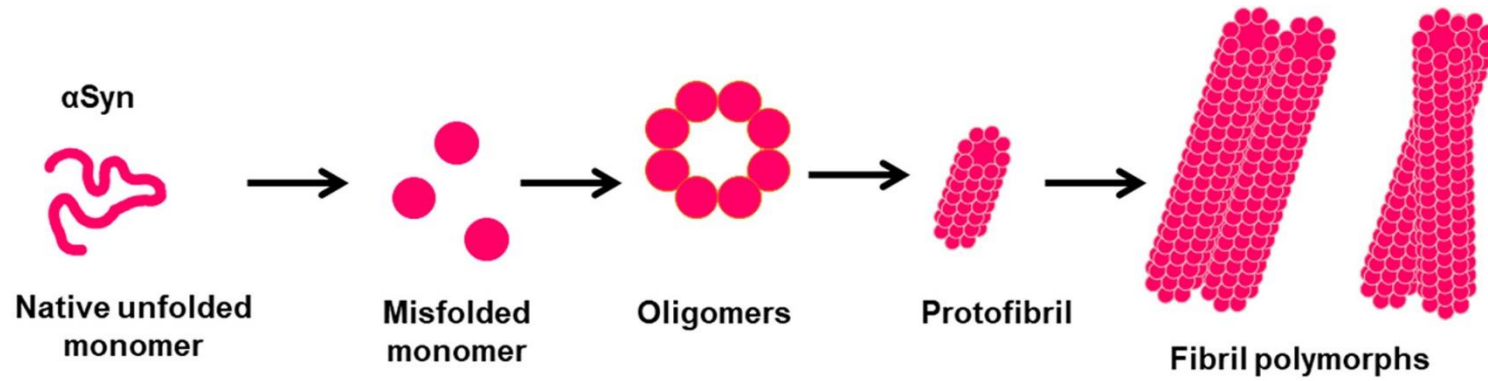
A Representative cryo-EM image of the α -synuclein fibrils showing twist of the two strands (red arrows), B Representative 2D class-average images showing striations visible due to helical arrangement of the monomers (green arrow pointing to a monomer) in the two protofibrils, C Cross-section of cryo-EM map showing the relative orientation of the monomers D Model structures (gold) compared with published structure (aquamarine)



Resolving power at atomic level:
histidine, lysine, glutamate residues depicted



α -synuclein has a role health and disease (synucleinopathies)

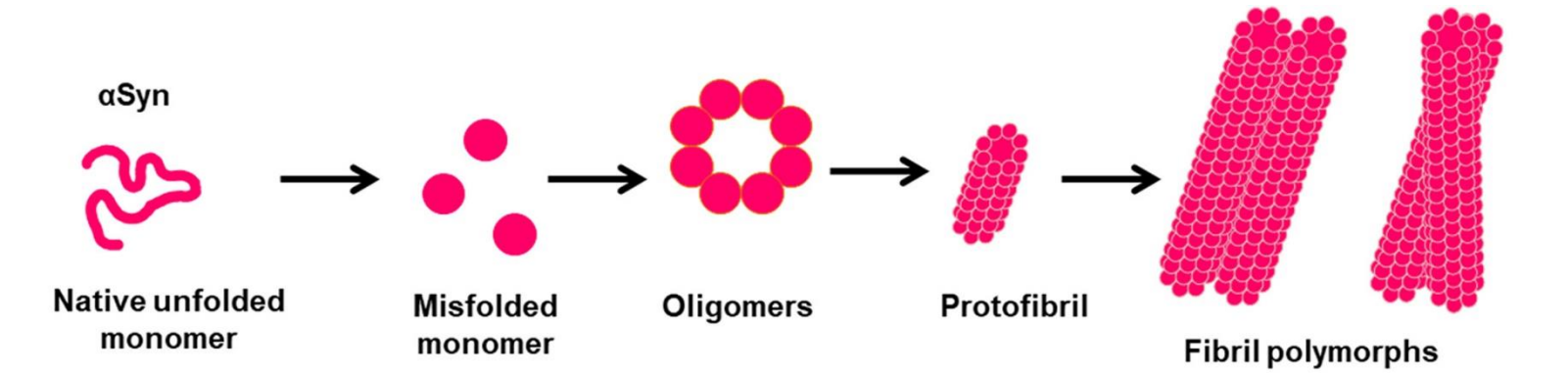


α -synuclein has a role health and disease (synucleinopathies)

Health:

Vesicle trafficking

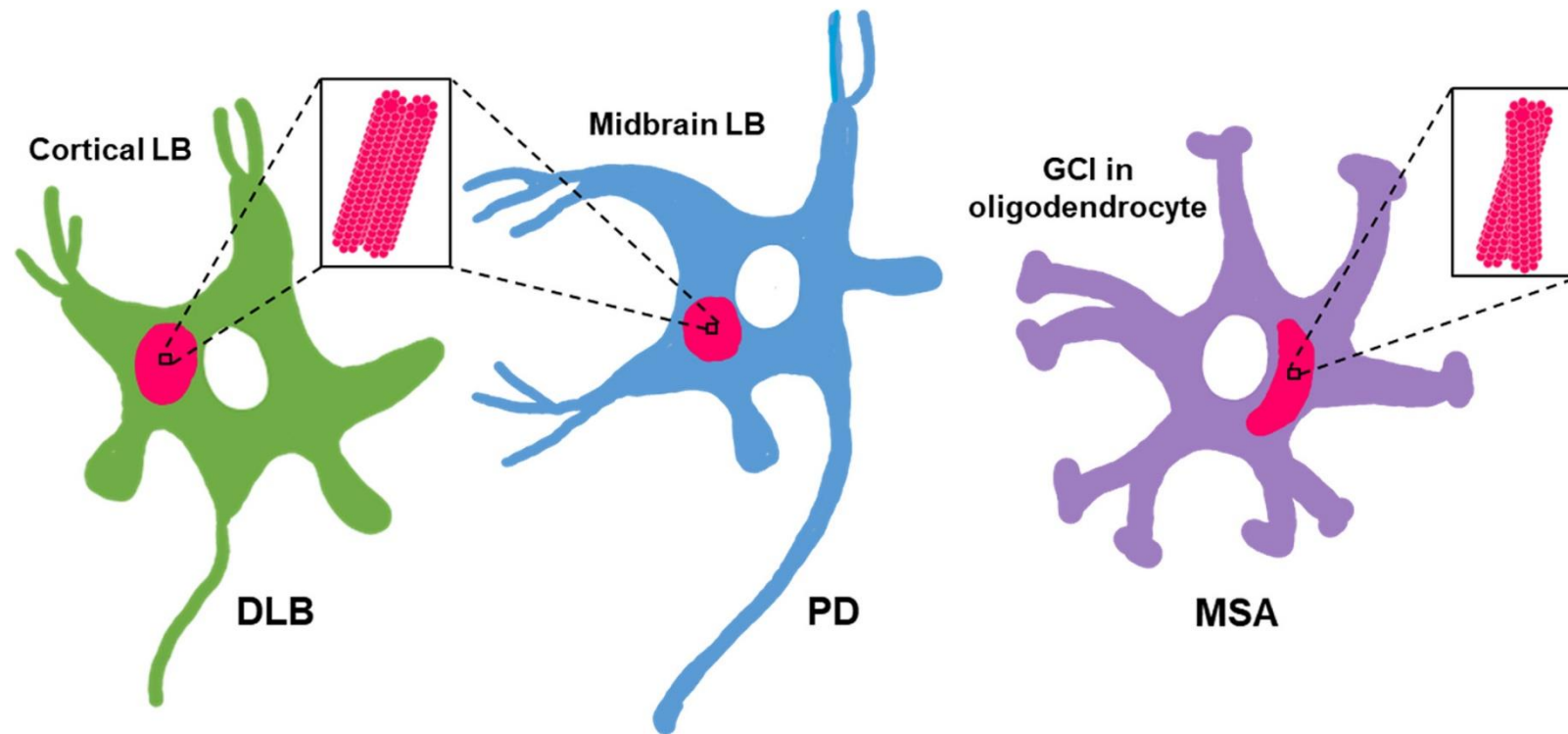
...



Disease:

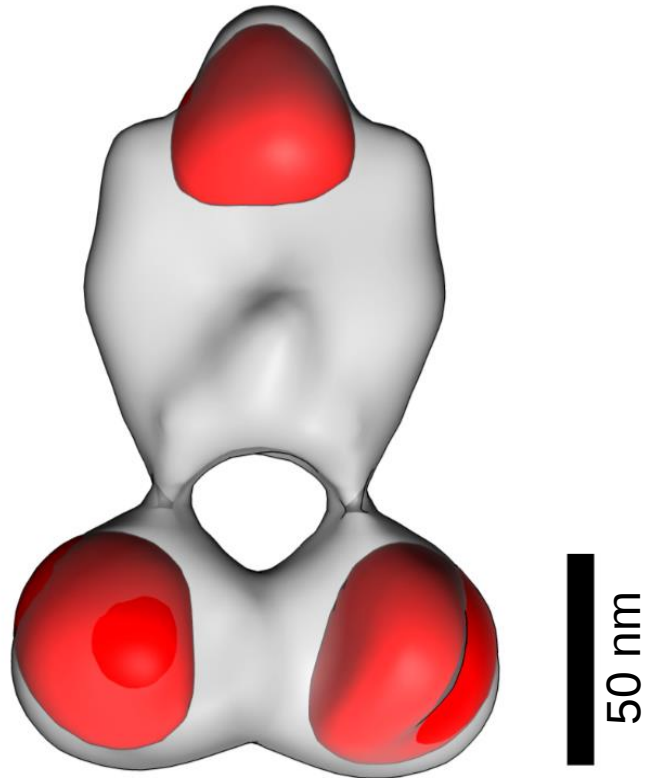
Parkinson's disease

...

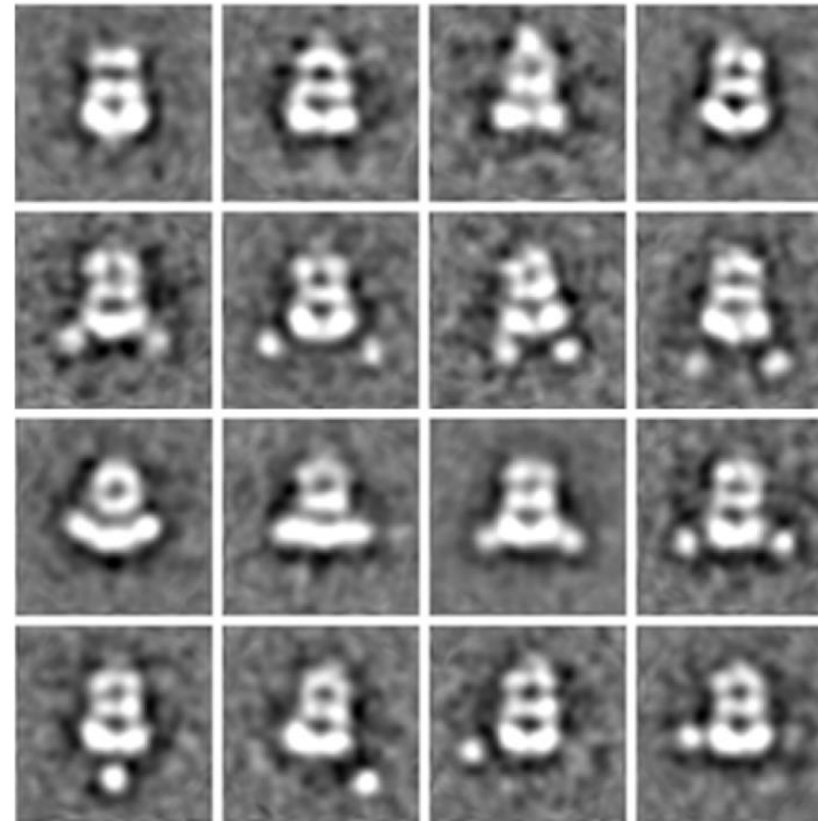


Project 6: Domain mobility or co-factor occupation

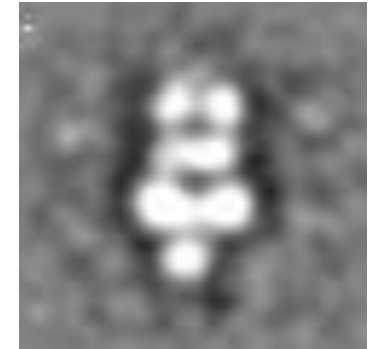
cryo-EM structure (tPDE):
3D variability



negative stain:
holo-PDE6 in complex with PrBP/ δ



Movie



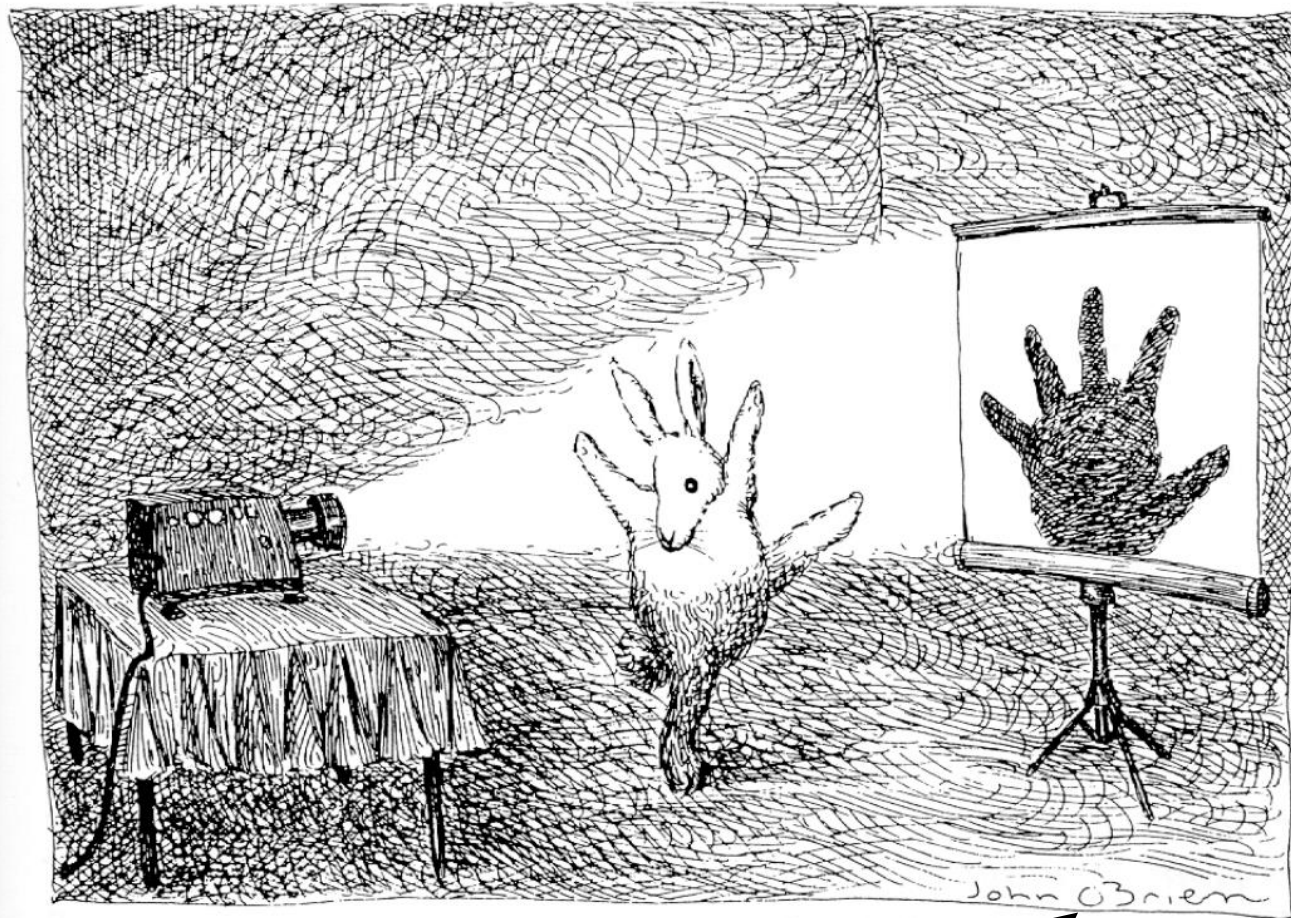
Qureshi, Schmidt et al (2018) *Nat Comms* 9 (1), 90

Thank you for you attention!

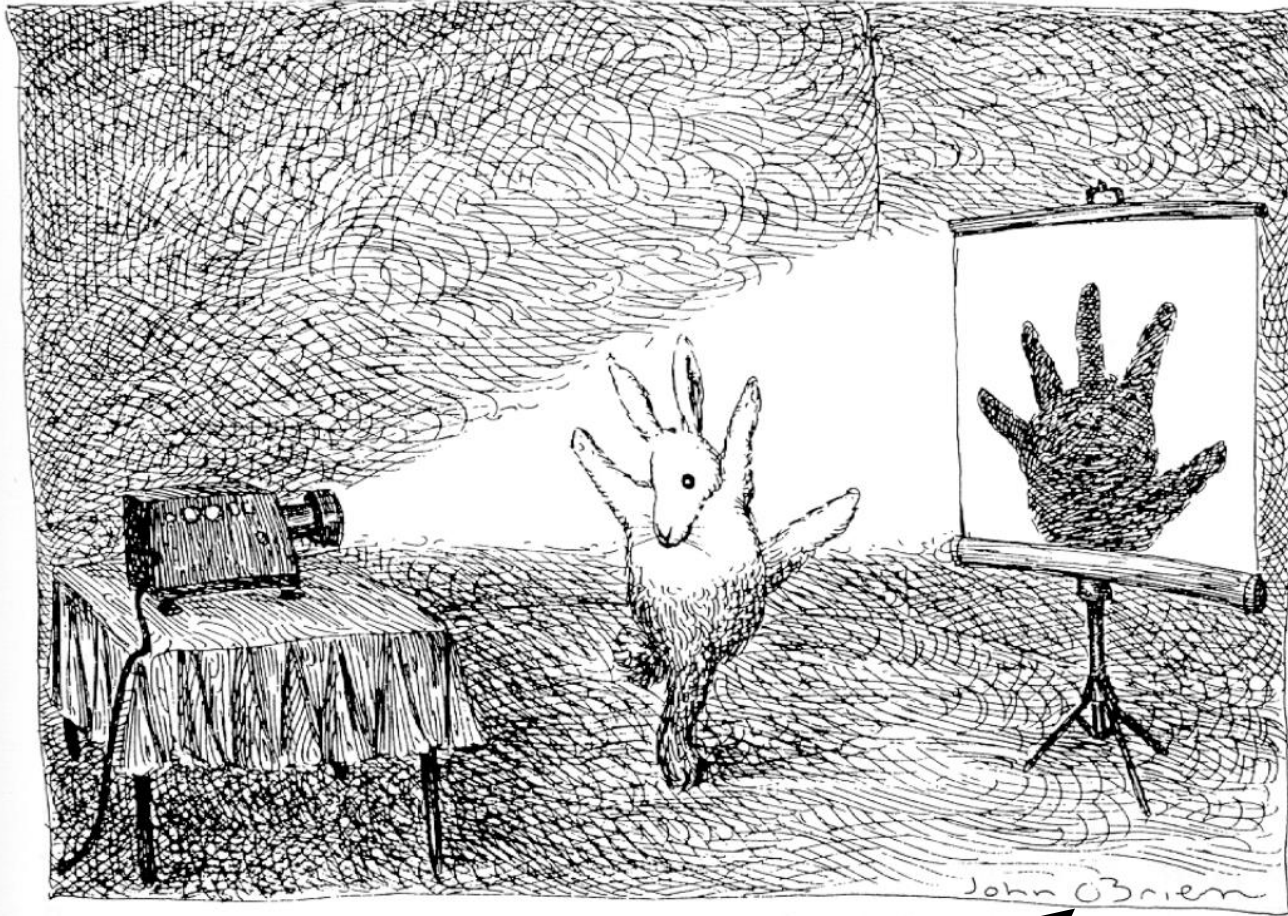
Questions?

Additional slides for potential questions?

2D images (shadows/photos vs. projects) of 3D objects

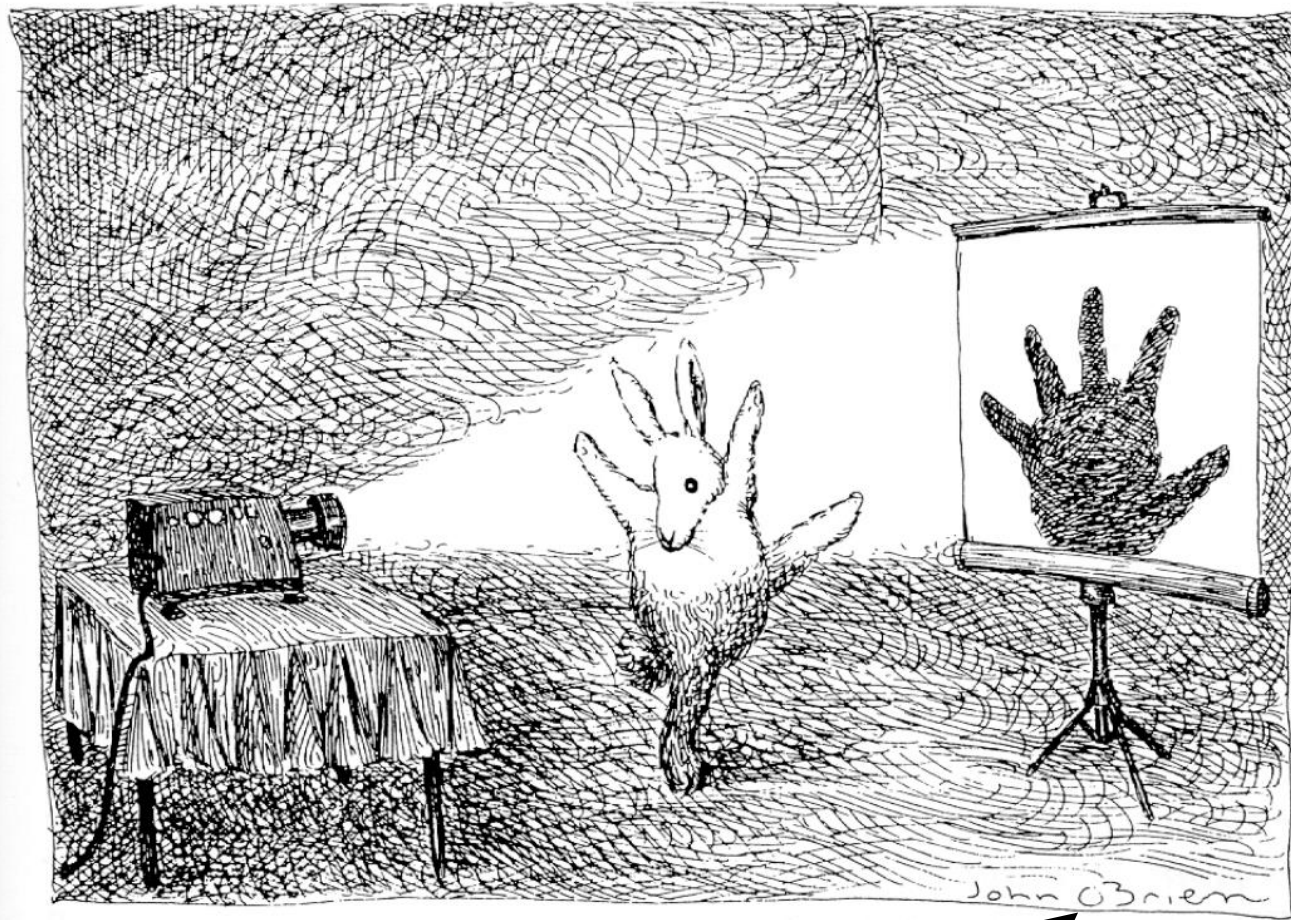


2D images (shadows/photos vs. projects) of 3D objects



Courtesy: Wikipedia

2D images (shadows/photos vs. projects) of 3D objects



Courtesy: Wikipedia

→ No transmission (shadow, photo) vs. transmission in projection image i.e., internal structure

Electron interactions with matter in TEM

Two different types of interactions appear:

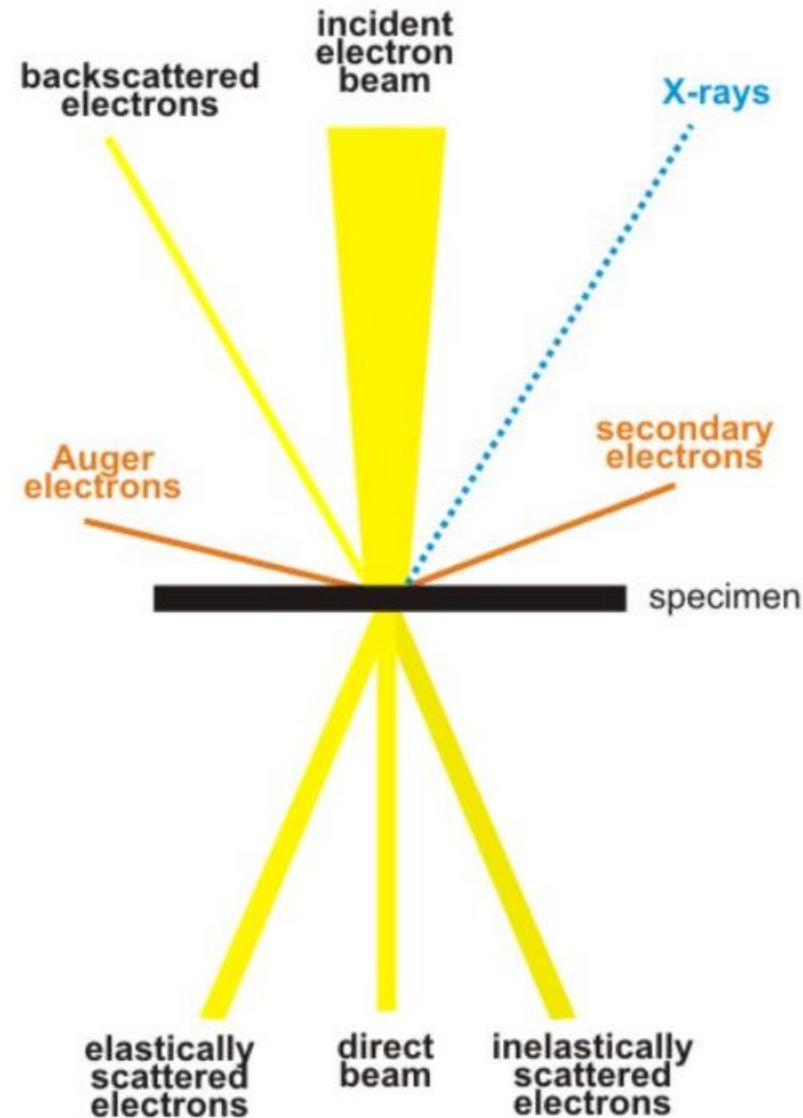
Elastic Interactions

No energy is transferred from the electron to the sample. The electron either passes without any interaction (direct beam) or is scattered by the electrostatic attraction of the positive potential inside the electron cloud. The arising signals are mainly exploited in [TEM](#) and [electron diffraction](#).

Inelastic Interactions

Energy is transferred from the incident electrons to the sample producing secondary electrons, phonons, UV quanta or cathodo-luminescence. Ionization of atoms by removing inner shell electrons leads to the emission of X-rays or Auger electrons. These signals are used in [analytical electron microscopy](#).

A script giving an introduction can be downloaded here: [interactions](#).



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