Seminar - Cochlear Implant

Viva Berlenghi, Florian David, Sophie Meuwly EE-519: Bioelectronics and Biomedical Microelectronics, EPFL December 3, 2024

Abstract—Cochlear implants (CIs) are neuroprosthetic devices providing auditory perception to individuals with hearing loss (HL). This report provides a general overview of cochlear implant technology, including the biological principles underlying hearing restoration, the device's components, surgical insertion, and patient outcomes. Despite their potential, challenges persist. We discuss here the limitations and future directions of such devices.

I. INTRODUCTION

A cochlear implant (CI) is a small, complex neuroprosthetic device that provides a sense of sound to individuals with moderate-to-profound sensorineural hearing loss. In addition to being the pioneering invention to offer a sense of hearing, it holds the distinction of being the first device to restore any of the five human senses [1].

The simplest design consisted of a single stimulating electrode and was implanted in 1957 by French physician André Djourno and otologist Charles Eyriès [2]. The two implanted patients could differentiate between different loudness levels but could not understand speech. The implants sadly failed shortly after implantation. Later, the American otologist William House is widely regarded as the father of cochlear implants. In 1961, he developed a similar device and contributed significantly to clinical trials that demonstrated its feasibility [3].

During the following decade, multiplied efforts yielded promising results, culminating in the Food and Drug Administration (FDA) approving the first cochlear implant in 1984 [4]. Today, over 700,000 registered devices have been implanted worldwide [5], thanks to leading companies such as Med-El Corporation (Austria), Cochlear Corporation (Australia), and Advanced Bionics (Sonova Holding, Switzerland).

II. BIOLOGY BACKGROUND

Cochlear implants address sensorineural hearing loss, which affects the inner ear, unlike conductive hearing loss that impacts the outer or middle ear.

Sound, being airborne, cannot directly reach the inner ear. Instead, it undergoes a transformation in the middle ear, where the tympanic membrane converts low-impedance airborne waves into high-impedance pressure waves within the fluid-filled cochlea (see Figure 1).

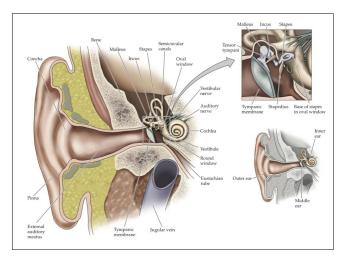


Figure 1: **The human ear.** The outer, middle, and inner ear structures are shown. Sound waves travel through the outer ear to the tympanic membrane, where vibrations are transmitted via the inner ear to the cochlea, where they are converted into neural signals, which are carried by the auditory nerve to the brain. Image taken from [6].

The cochlea performs three key functions: amplifying, converting, and analyzing sound frequencies [7]. It transforms these pressure waves into neural impulses. When uncoiled, the cochlea spans approximately 35mm and features a topographical frequency map described in Figure 2, which cochlear implants leverage.

Inner ear hair cells naturally generate neural impulses in response to shearing forces caused by the movement of the basilar membrane beneath them. Damage to these hair cells results in sensorineural hearing loss, and cochlear implants are considered when the issue is confined to the cochlea and the auditory nerve remains intact.

Candidacy for cochlear implants has evolved over time. In the early 1980s, eligibility was limited to individuals with bilateral total deafness (< 110 dB hearing loss). By the 1990s, it included those with severe hearing loss (< 70 dB hearing loss). Today, candidacy is based on supra-threshold speech criteria, such as <50% open-set sentence recognition with appropriately fitted hearing aids [8].

Outcomes also depend on the brain's hearing-related

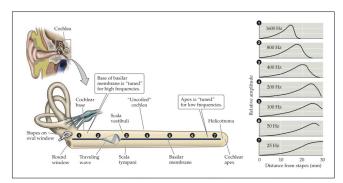


Figure 2: **Traveling waves along the (uncoiled) cochlea.** The graphs on the right profile show the amplitude of the travelling wave along the basilar membrane for different frequencies. Higher frequencies map to the base, and lower frequencies map to the apex. Image taken from [6].

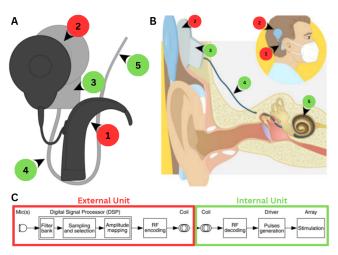


Figure 3: (A-B) Main components of the cochlear implant: (1) external behind-the-ear (BTE) sound processor; (2) inductive radio frequency (RF) link between external and internal units; (3) internal stimulator; (4) wires; (5) electrode array. (C) Scheme of sound coding into the artificial stimulation of the cochlea with cochlear implant systems. Adapted from [9].

regions. Adults with acquired hearing loss are evaluated differently from those born deaf. Early implantation yields better results, as it takes advantage of neural plasticity. Cochlear implants can be provided starting at 9 months of age, allowing for early intervention and improved therapy outcomes.

III. TECHNOLOGY OVERVIEW

An electrical cochlear implant (eCI) converts acoustic signals into electrical stimuli, bypassing damaged hair cells by directly stimulating the spiral ganglion cells (SGNs) from the auditory nerve using an electrode array. The different frequencies are encoded depending on the stimulation lo-

Manufacturer	Advanced Bionics	Cochlear Limited	MED-EL
Internal unit	HiRes Ultra 3D	CI632 Slim Modiolar	Synchrony 2
RF comm. Freq. (MHz)	49	5	12
RF data rate (Mb/sec)	1.0	0.5	0.6
Max stimulation rate (PPS)	83 000	32 000	51 000
Electrodes (#)	16	22	up to 24
Electrode array length (mm)	15.0 to 20.0	14.0	12.1 to 26.4
Pulse amplitude (µA)	0 to 2040	0 to 1750	0 to 1200
Pulse width (µs)	10.78 to 229.0	up to 31.7	2.1 to 425.0
Implant dimensions (mm ³)	56.2 x 28.6 x 4.5	55.0 x 31.0 x 3.9	47.3 x 24.0 x 4.5
Implant weight (g)	11.0	9.2	7.6
External unit	Sky CI M90	Nucleus 8 CP1110	Sonnet 2
Input dynamic range (dB)	<80	<45	<75
Rechargeable battery life (h)	9 to 18	17 to 34	7 to 10

Figure 4: Comparison of 3 devices already available on the market. Based on [11], [10], [12], [13]

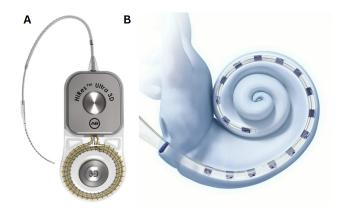


Figure 5: (A) HiRes Ultra 3D Cochlear Implant (internal unit) with (B) HiFocus SlimJ electrodes. From [10].

cation along the tonotopic map. eCIs are composed of two main units []: an external unit to capture and process sounds and an internal unit to generate electrical pulse patterns to stimulate the cochlea (see Fig. 3).

While many devices exist, they all have different characteristics. We provide a short comparison of three different devices in Figure 4. An internal unit from the manufacturer Advanced Bionics [10] can be found in Figure 5.

A. External unit

The external unit of the cochlear implant is responsible for converting acoustic sounds into signals that can be sent to the internal unit for stimulating the cochlea.

Sound waves are captured by a microphone and converted into electrical signals, which are then processed in the speech processor. While the design of cochlear implant systems may vary, they all follow the same core steps in sound processing []. First, the acoustic signal is digitized using a digital signal processor (DSP). Next, advanced signal processing methods, often enhanced by machine learning nowadays, extract key sound features such as amplitude and frequency to characterize the auditory input. Finally, the speech processor translates these features into stimulation

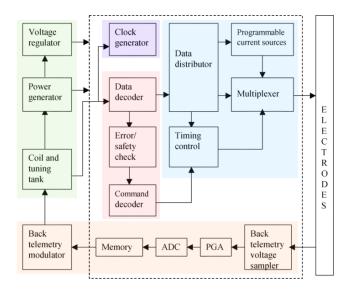


Figure 6: Block diagram of the cochlear implant internal unit. Most devices commercially available today include the following units: a signal receptor and a power generator (green), a stimulation parameter decoder (red), a current generator (blue), a feedback modulator (orange), and a control unit (purple). Adapted from [14].

parameters that the internal unit can interpret, enabling the recreation of sound for individuals with hearing loss.

A battery is also located behind the ear to power both the external and internal units. Modern designs incorporate rechargeable or disposable batteries for convenience and extended use. Transmission of information between the two units is wireless via a radio frequency (RF) transmitter. Located within a headpiece and aligned to the internal receiver by a magnet, it sends both the stimulation patterns and power through the skin of the patient.

B. Internal unit

The internal unit of a cochlear implant is responsible for processing external signals and delivering electrical stimulation to the auditory nerve. While many devices exist today, they all share similar building blocks (see Fig. 6) described in [14], each serving a specific function. Overall, they ensure safe, effective, and reliable stimulation while offering feedback for continuous monitoring and adjustment.

First, a coil receives RF signals from the external processor. RF signals contain the preprocessed data required to stimulate the cochlea but also the power needed to operate the internal unit [15]. Specifically, a power generator is necessary to convert RF energy into electrical power, which is then stabilized by a voltage regulator.

Digital commands are then extracted from the RF signal and converted into detailed stimulation parameters using decoders. Such parameters include electrode selection, stimulation amplitude, and pulse duration, which all need to be controlled through a safety module to ensure that stimulation is safe and doesn't induce damages to the tissues or the electrodes [16].

Once stimulation parameters have been decoded, programmable current sources generate precise electrical pulses [17]. Then, a multiplexer ensures the generated current is delivered to the correct electrode channel (described in detail in Section III-C), enabling targeted stimulation of the auditory nerve.

All cochlear implants available today include a back telemetry system to monitor the electrode-tissue interface and neural response [18]. Voltage is measured at the electrode, amplified (PGA), converted into digital signals (ADC), and finally transmitted to the external unit for diagnostic purposes and real-time feedback.

Finally, synchronization of operations across all components is necessary to coordinate precise delivery of stimulation patterns.

C. Electrodes

A cochlear implant's electrodes typically consist of multiple stimulation contacts or channels, each corresponding to different frequency ranges along the tonotopic organization of the patient's cochlea [19]. Modern electrodes aim to balance effective stimulation with minimal trauma to the delicate structures of the inner ear.

Electrodes are made of biocompatible materials to prevent tissue rejection and metal corrosion. For example, platinum and iridium can be used in electrode contacts for their excellent electrical conductivity and durability. Silicone and parylene are also often used to encapsulate the electrodes due to their insulating and hermetic properties [20]. Also, it is important that the electrode array remains flexible to allow smooth insertion into the cochlea (see Section IV).

The number of channels varies by manufacturer and model, typically ranging from 12 to 24. The placement and spacing of these channels must be optimized to align with the cochlea's tonotopic organization [21]. While increasing the number of channels could, in theory, increase sound reconstruction quality, it is not feasible for two main reasons [22]. First, the cochlea is a fluid-filled structure, which limits the focusing of electric current and increases crosstalk between channels. Second, more channels require higher power supply and wiring, which is difficult to accomplish in this fragile area.

IV. SURGICAL PROCEDURE

Performed under general anesthesia, the surgery typically lasts between two and three hours per ear [23]. The surgeon makes a small incision behind the ear to access the mastoid bone and the middle ear. A mastoidectomy is performed to create a pathway to the cochlea, allowing for the insertion of the electrode array [24]. The internal receiver-stimulator is then secured under the skin and fixed on the skull, and the

electrode array is carefully inserted into the cochlea through the round window to stimulate the auditory nerve. Postoperative assessments, including impedance testing and neural response telemetry, are conducted to ensure the device's functionality. Most patients can return home the same day, with follow-up appointments scheduled to monitor recovery and initiate the device's programming [25].

Postoperative complications are rare or easily treatable. Complications are classified as minor and major, and they respectively have a rate of 14.9% and 5% [26]. The most common minor complications are infectious complications like acute otitis media (AOM) or wound infections. Those are treated with antibiotics and do not have a long-term impact. Another common minor complication is cochleovestibular, which includes tinnitus and vestibular complications. For both, no causal or specific cause was identified. Vestibular complications consist of vertigo and loss of balance and can rapidly be treated with medications. The last type of minor complications are neurological, like facial nerve palsy or taste disorder. They are more rare than the others and resolved spontaneously without any medication.

Major complications include infection-related issues, surgical revision issues, and device failure. A few cases of meningitis and mastoids were observed, but all patients were successfully treated with no long-term impact. There are around 3.7% [26] cases that needed a surgical revision. Device failures rarely happen and are mostly linked to well-known trauma (for instance, if the patient hits their head) [27].

V. LIMITATIONS

A. Technical considerations

When performing an MRI or a CT scan on a patient with a CI on the ipsilateral side of interest, we need to be careful not to damage the implant and ensure that the area of interest is clearly visible. Due to the magnet and other metallic parts of the CI, MRI images might be affected and parasitized with artifacts [28]. If the area of interest is hidden by the artifacts, then the CI might need to be removed, although some specific imaging techniques are less sensitive to metallic artifacts.

Another issue with the electric cochlear implant is that the precision of stimulation is limited. Broad areas of the tonotopic map are simultaneously activated. Although selectivity can be improved by using tripolar electrodes, this also requires more cables and more battery consumption, which reduces the device's practicality for everyday use [29]. This is why more electrodes doesn't imply higher precision. They haven't found a significant performance difference between implants with 10 or 20 electrodes [30].

B. Additional considerations

Only about 6% of adults eligible for cochlear implantation (CI) undergo the surgery [31]. The decision is complex,

factoring in costs, surgical risks, and lifelong follow-up. Implantation permanently eliminates any residual natural hearing and requires therapy to relearn how to hear, with improvements in quality of life taking time and effort.

Cultural factors also play a role, as the strong identity within the deaf community often creates a divide. Some reject implants, emphasizing the preservation of deaf culture, while others feel torn between the hearing and non-hearing worlds. For children, the decision lies with guardians, while adults decide for themselves [32].

VI. FUTURES DIRECTIONS

Hearing loss affects hundreds of millions of people world-wide today, and the numbers keep growing. The sound quality provided by current devices is still far from natural hearing. Ongoing research efforts aim to improve the performance of the user experience of cochlear implants. For example, some researchers focus on signal processing improvement by using advanced speech processing strategies [33] and novel machine learning-based noise reduction methods [34], [35]. Others develop new techniques to reduce the current spread during stimulation [36] or combine adjacent electrodes to improve spectral resolution [37].

One particularly promising emerging technology is the use of optogenetics to increase spatial selectivity of the stimulation using light delivered by an optical cochlear implant (oCI) [9]. First, the SGNs need to express an opsin, a light-activated ion channel. Then, instead of introducing an electrode in the spiraled organ, a light emitter shines light on the genetically modified neurons, inducing a neural response [38]. The other building blocks of the device remain highly similar to those of modern eCIs. Two main approaches have already been explored in animal models and human postmortem models to deliver light to the neurons of interest. In active oCIs or LED-based devices, the light source is in direct contact with the neurons inside the cochlea through the insertion of low-voltage organic LEDs. Conversely, in passive oCIs, or waveguide-based devices, the light source is located in the external part of the device, and light is brought inside the cochlea through inserted optical fibers.

VII. CONCLUSION

This report delves into key aspects of cochlear implants, highlighting their reliance on the cochlea's tonotopic organization to restore hearing. By stimulating specific regions of the cochlea, the device effectively generates distinct sound frequencies. Composed of an external unit and an internal unit, a cochlear implant converts acoustic signals into electrical impulses delivered via electrodes. While the implantation procedure is routine, it carries potential risks. Despite some limitations, such as imprecise stimulation and imaging artifacts, multiple future directions exist, such as optical cochlear implants.

REFERENCES

- A. Mudry and M. Mills, "The early history of the cochlear implant: a retrospective," *JAMA Otolaryngol Head Neck Surg.*, 2013.
- [2] P. Seitz, "French origins of the cochlear implant," Cochlear Implants Int, 2002.
- [3] L. Eisenberg, "The contributions of william f. house to the field of implantable auditory devices," *Hear Res*, 2015.
- [4] M. Envoy, "The history of the cochlear implant," 2024. Accessed: 2024-11-30.
- [5] NID, "What are cochlear implants for hearing?," 2024. Accessed: 2024-11-30.
- [6] D. Purves, G. Augustine, D. Fitzpatrick, W. C. Hall, A. LaMantia, and R. Mooney, *Neuroscience*. 6th Edition. Sinauer Associates, New York, 2018.
- [7] S. J. Elliott and C. A. Shera, "The cochlea as a smart structure," *Smart Materials and Structures*, vol. 21, no. 6, p. 064001, 2012.
- [8] World Health Organization (WHO), "World report on hearing executive summary." https://www.who.int/publications/i/item/9789240021570. Accessed: 2024-12-03.
- [9] L. Khurana, T. Harczos, T. Moser, and L. Jablonski, "En route to sound coding strategies for optical cochlear implants," iScience, vol. 26, no. 10, p. 107725, 2023.
- [10] Advanced Bionics, "Internal components of a cochlear implant." https://www.advancedbionics.com/gb/en/portals/ professional-portal/products/ci-internal-components.html. Accessed: 2024-12-01.
- [11] Cochlear Implant HELP, "Comparison chart." https://cochlearimplanthelp.com/cochlear-implant-comparison-chart/, 2023. Accessed: 2024-12-01.
- [12] Cochlear, "Slim modiolar electrode (ci632)." https://www.cochlear.com/us/en/professionals/products-and-candidacy/nucleus/implant/electrodes/slim-modiolar. Accessed: 2024-12-01.
- [13] MED-EL, "Synchrony 2 cochlear implant." https://www.medel.com/hearing-solutions/cochlear-implants/synchrony2. Accessed: 2024-12-01.
- [14] F.-G. Zeng, S. Rebscher, W. Harrison, X. Sun, and H. Feng, "Cochlear implants: System design, integration, and evaluation," *IEEE Reviews in Biomedical Engineering*, vol. 1, pp. 115–142, 2008.
- [15] I. J. Hochmair and E. S. Hochmair, "Transcutaneous power and signal transmission system and methods for increased signal transmission efficiency," tech. rep., U.S. Patent 5 070 535, 1991.
- [16] R. V. Shannon, "A model of safe levels for electrical stimulation," *IEEE Transactions on Biomedical Engineering*, vol. 39, no. 4, pp. 424–426, 1992.

- [17] J. C. Gord, "Programmable current output stimulus stage for implantable device," tech. rep., U.S. Patent 6 181 969, 2001.
- [18] L. H. M. Mens, "Advances in cochlear implant telemetry: Evoked neural responses, electrical field imaging, and technical integrity," *Trends in Amplification*, vol. 11, no. 3, pp. 143– 159, 2007.
- [19] A. Dhanasingh and C. Jolly, "An overview of cochlear implant electrode array designs," *Hearing Research*, vol. 356, pp. 93– 103, 2017.
- [20] S.-H. Ahn, J. Jeong, and S. J. Kim, "Emerging encapsulation technologies for long-term reliability of microfabricated implantable devices," *Micromachines*, vol. 10, no. 8, p. 508, 2019.
- [21] C. C. Finley, T. A. Holden, L. K. Holden, B. R. Whiting, R. A. Chole, J. G. Neely, T. E. Hullar, and M. W. Skinner, "Role of electrode placement as a contributor to variability in cochlear implant outcomes," *Otology & Neurotology*, vol. 29, no. 7, pp. 920–928, 2008.
- [22] M. F. Dorman, P. C. Loizou, J. Fitzke, and Z. Tu, "The recognition of sentences in noise by normal-hearing listeners using simulations of cochlear-implant signal processors with 6–20 channels," *The Journal of the Acoustical Society of America*, vol. 104, no. 6, pp. 3583–3585, 1998.
- [23] S. F. University of California, "Cochlear implant surgery," 2024. Accessed: 2024-11-24.
- [24] N. L. Deep, E. M. Dowling, D. Jethanamest, and M. L. Carlson, "Cochlear implantation: an overview," *Journal of Neurological Surgery Part B: Skull Base*, vol. 80, no. 02, pp. 169–177, 2019.
- [25] J. H. Medicine, "Cochlear implant surgery," 2024. Accessed: 2024-11-24.
- [26] A. Farinetti, D. B. Gharbia, J. Mancini, S. Roman, R. Nicollas, and J.-M. Triglia, "Cochlear implant complications in 403 patients: comparative study of adults and children and review of the literature," *European annals of otorhinolaryngology, head and neck diseases*, vol. 131, no. 3, pp. 177–182, 2014.
- [27] A. Ciorba, R. Bovo, P. Trevisi, M. Rosignoli, C. Aimoni, A. Castiglione, and A. Martini, "Postoperative complications in cochlear implants: a retrospective analysis of 438 consecutive cases," *European Archives of Oto-rhino-laryngology*, vol. 269, pp. 1599–1603, 2012.
- [28] R. Srinivasan, C. So, N. Amin, D. Jaikaransingh, F. D'arco, and R. Nash, "A review of the safety of mri in cochlear implant patients with retained magnets," *Clinical Radiology*, vol. 74, no. 12, pp. 972–e9, 2019.
- [29] Z. Zhu, Q. Tang, F.-G. Zeng, T. Guan, and D. Ye, "Cochlear-implant spatial selectivity with monopolar, bipolar and tripolar stimulation," *Hearing research*, vol. 283, no. 1-2, pp. 45–58, 2012.
- [30] A. Kral, R. Hartmann, D. Mortazavi, and R. Klinke, "Spatial resolution of cochlear implants: the electrical field and excitation of auditory afferents," *Hearing research*, vol. 121, no. 1-2, pp. 11–28, 1998.

- [31] L. Zhang, A. S. Ding, D. X. Xie, and F. X. Creighton, "Understanding public perceptions regarding cochlear implant surgery in adults," *Otology & Neurotology*, vol. 43, no. 3, pp. e331–e336, 2022.
- [32] I. D. Miziara, C. S. M. G. Miziara, R. K. Tsuji, and R. F. Bento, "Bioethics and medical/legal considerations on cochlear implants in children," *Brazilian Journal of otorhino-laryngology*, vol. 78, pp. 70–79, 2012.
- [33] D. Wang and J. H. L. Hansen, "Speech enhancement for cochlear implant recipients," *The Journal of the Acoustical Society of America*, vol. 143, no. 4, pp. 2244–2254, 2018.
- [34] L. P.-H. Li, J.-Y. Han, W.-Z. Zheng, R.-J. Huang, and Y.-H. Lai, "Improved environment-aware–based noise reduction system for cochlear implant users based on a knowledge transfer approach: Development and usability study," *Journal of Medical Internet Research*, vol. 23, no. 10, p. e25460, 2021.
- [35] L. Bramsløw, G. Naithani, A. Hafez, T. Barker, N. H. Pontoppidan, and T. Virtanen, "Improving competing voices segregation for hearing impaired listeners using a low-latency deep neural network algorithm," *The Journal of the Acoustical Society of America*, vol. 144, no. 1, pp. 172–185, 2018.
- [36] C. van den Honert and D. C. Kelsall, "Focused intracochlear electric stimulation with phased array channels," *The Jour*nal of the Acoustical Society of America, vol. 121, no. 6, pp. 3703–3716, 2007.
- [37] X. Luo, C.-C. Wu, and K. Pulling, "Combining current focusing and steering in a cochlear implant processing strategy," *International Journal of Audiology*, vol. 60, no. 3, pp. 232– 237, 2020.
- [38] A. Dieter, D. Keppeler, and T. Moser, "Towards the optical cochlear implant: optogenetic approaches for hearing restoration," EMBO Molecular Medicine, vol. 12, no. 4, 2020.