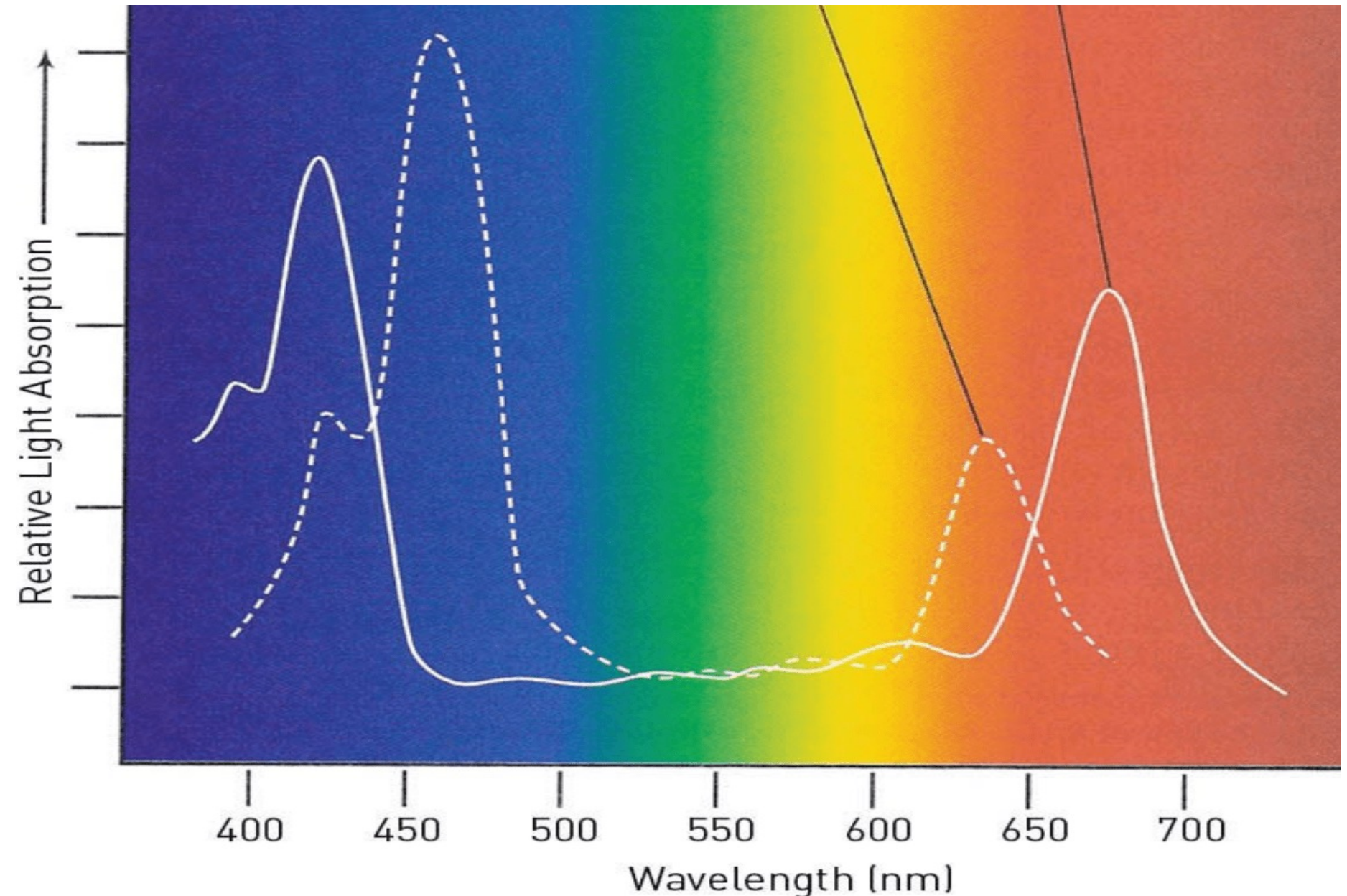


Methods of absorption spectroscopy

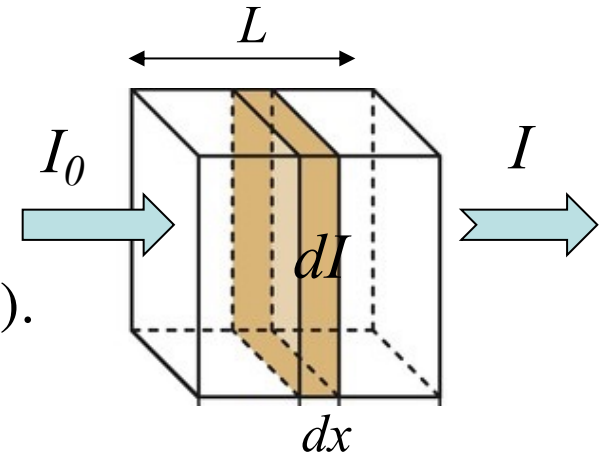


Beer Lambert Law

$$dI(\tilde{\nu}) = -\sigma(\tilde{\nu}) \cdot I(\tilde{\nu}) \cdot n_1 \cdot dx, \quad \text{Loss of intensity}$$

$$\ln I = -\sigma(\tilde{\nu}) \cdot L + \text{const} \Rightarrow$$

$$I(\tilde{\nu}) = I_0(\tilde{\nu}) \exp(-\sigma(\tilde{\nu}) \cdot n_1 \cdot L) = I_0(\tilde{\nu}) \exp(-\alpha(\tilde{\nu}) \cdot L).$$



$\tilde{\nu}$ — frequency in wavenumbers [cm^{-1}] (can be ν , λ as well);

$I(\tilde{\nu})$ — spectral brightness of light; [$\text{Watt}/(\text{cm}^{-1} \cdot \text{m}^2)$];

$\alpha(\tilde{\nu})$ — differential absorption coefficient (sample): sample spectrum [cm^{-1}];

$\sigma(\tilde{\nu})$ — differential absorption cross-section [cm^2]: **molecular spectrum.**

For an isolated transition one must account for its lineshape:

$$\int_0^{\infty} \sigma(\tilde{\nu}) d\tilde{\nu} = \sigma_0 \int_0^{\infty} g(\tilde{\nu}) d\tilde{\nu} = \sigma_0 \quad \text{Integral absorption cross section [cm]}$$

$$\int_0^{\infty} \alpha(\tilde{\nu}) d\tilde{\nu} = \alpha_0 \int_0^{\infty} g(\tilde{\nu}) d\tilde{\nu} = \alpha_0 \quad \text{Absorption coefficient [cm}^{-2}\text{]}$$

What do we measure in experiment?

Beer Lambert Law

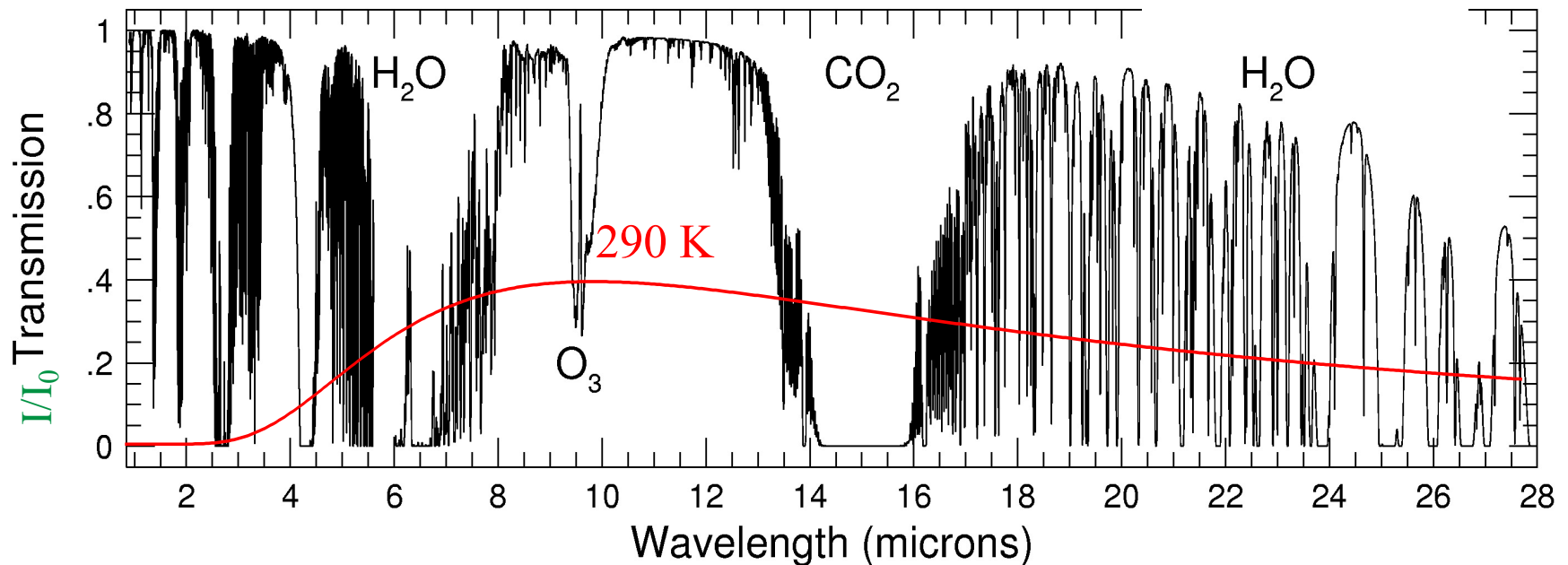
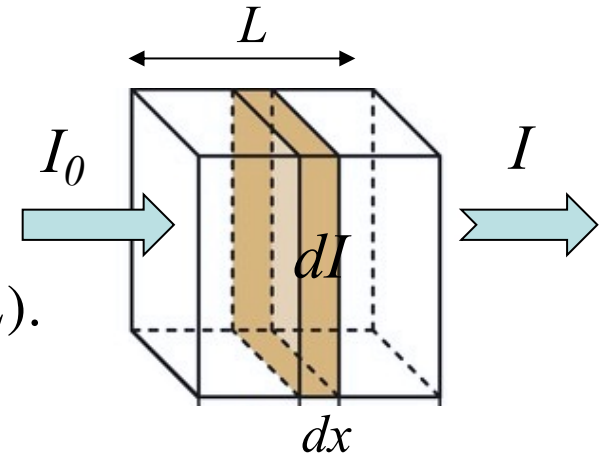
$$dI(\tilde{\nu}) = -\sigma(\tilde{\nu}) \cdot I(\tilde{\nu}) \cdot n_1 \cdot dx, \quad \text{Loss of intensity}$$

$$\ln I = -\sigma(\tilde{\nu}) \cdot L + \text{const} \Rightarrow$$

$$I(\tilde{\nu}) = I_0(\tilde{\nu}) \exp(-\sigma(\tilde{\nu}) \cdot n_1 \cdot L) = I_0(\tilde{\nu}) \exp(-\alpha(\tilde{\nu}) \cdot L).$$

$$\sigma(\nu) = \frac{\ln(I_0 / I(\tilde{\nu}))}{n \cdot L}$$

Earth atmosphere



$$T_{av} = 290K$$

$$T(\lambda_p) = \frac{2900}{\lambda_p (\mu m)}; \quad T(6) = 480K; \quad T(15) = 193K; \quad T(25) = 120K.$$

Beer Lambert Law

Detector measures light intensity integrated over frequency-range

$$P = \int_0^\infty I(\tilde{\nu}) d\tilde{\nu} = \int_0^\infty I_0(\tilde{\nu}) \exp(-\alpha(\tilde{\nu}) \cdot L) \cdot d\tilde{\nu}.$$

For $\alpha(\tilde{\nu}) \cdot L \ll 1$ (optically thin):

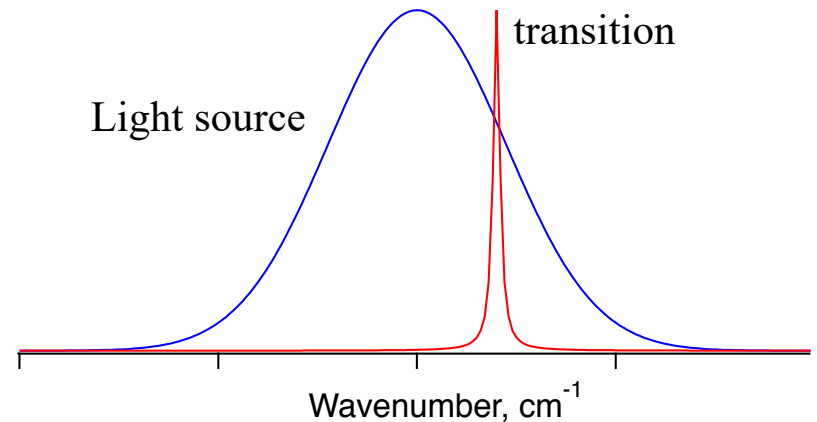
$$I(\tilde{\nu}) = I_0(\tilde{\nu}) \exp(-\alpha(\tilde{\nu}) \cdot L) \simeq I_0(\tilde{\nu}) \cdot (1 - \alpha(\tilde{\nu}) \cdot L).$$

1) $\Delta\nu_I \gg \Delta\nu_g$ (broad light source):

$$P = \int_0^\infty I(\tilde{\nu}) d\tilde{\nu} \simeq \int I_0(\tilde{\nu}) \cdot (1 - \alpha(\tilde{\nu}) \cdot L) \cdot d\tilde{\nu}$$

$$P \simeq P_0 - I_0(\tilde{\nu}_g) \cdot L \cdot \int_0^\infty \alpha(\tilde{\nu}) \cdot d\tilde{\nu}$$

$$\Delta P(\tilde{\nu}_g) = -I_0(\tilde{\nu}_g) \cdot L \cdot \alpha_0;$$



The spectrum will reflect spectral profile of the source, $I_0(\tilde{\nu})$, not of the molecule !

Beer Lambert Law

Detector measures light intensity integrated over frequency-range

$$P = \int_0^\infty I(\tilde{\nu}) d\tilde{\nu} = \int_0^\infty I_0(\tilde{\nu}) \exp(-\alpha(\tilde{\nu}) \cdot L) \cdot d\tilde{\nu}.$$

For $\alpha(\tilde{\nu}) \cdot L \ll 1$ (optically thin):

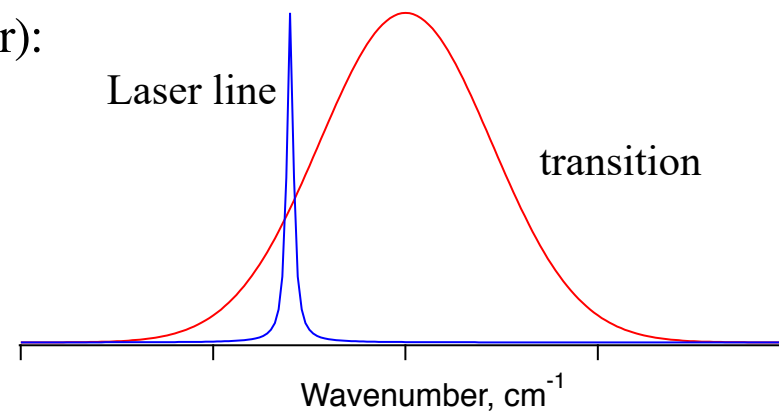
$$I(\nu) = I_0(\tilde{\nu}) \exp(-\alpha(\tilde{\nu}) \cdot L) \simeq I_0(\tilde{\nu}) \cdot (1 - \alpha(\tilde{\nu}) \cdot L).$$

2) $\Delta\nu_I \ll \Delta\nu_g$ (narrow band light source, e.g., laser):

$$P = \int_0^\infty I(\tilde{\nu}) d\nu \simeq \int I_0(\tilde{\nu}) \cdot (1 - \alpha(\tilde{\nu}) \cdot L) \cdot d\tilde{\nu}$$

$$\Delta P(\tilde{\nu}_I) = -P_0 \cdot L \cdot \alpha(\tilde{\nu}_I) \propto \sigma(\tilde{\nu}_I);$$

$$\Delta P(\tilde{\nu}_I) = -P_0 \cdot L \cdot \alpha(\tilde{\nu}_I) \propto \sigma(\tilde{\nu}_I),$$



This is the only truly measurements of molecular spectra

The condition $\Delta\nu_I \ll \Delta\nu_g$ allows for spectral resolution

Einstein coefficient B, cross section and TDM

B and σ are empirical, TDM comes from QM.

All three reflect probability for a transition

$$\sigma_0 = \frac{h\tilde{\nu}B_{12}}{c^2} \quad [B_{12}] = (\text{cm} \cdot \text{cm}^2 \cdot \text{s}^{-2}) / \text{J} = \text{cm}^3 / (\text{J} \cdot \text{s}^2)$$

$$A_{21} = \frac{8\pi h}{c^3} \cdot \nu_{12}^3 B_{21} = 8\pi h \cdot \tilde{\nu}_{12}^3 B_{21} \Rightarrow B_{21} = \frac{A_{12}}{8\pi h \cdot \tilde{\nu}_{12}^3};$$

$$\sigma_0 = \frac{h\tilde{\nu}B_{21}}{c^2} = \frac{h\tilde{\nu}}{c^2} \frac{A_{12}}{8\pi h \cdot \tilde{\nu}_{12}^3} = \frac{1}{8\pi c^2 \cdot \tilde{\nu}_{12}^2} A_{12} \Rightarrow \tau_{sp} \propto \frac{1}{\sigma_0};$$

- The stronger is a transition the shorter is the lifetime of excited state

$$\sigma_0 = M_{21}^2 \frac{\pi \cdot \tilde{\nu}}{3\epsilon_0 \cdot h \cdot c} \Rightarrow A_{21} \propto \tilde{\nu}^3 \cdot M_{21}^2 \Rightarrow \tau_{12} \propto \frac{1}{\tilde{\nu}^3};$$

- Lifetime of excited state reduces quickly with its energy

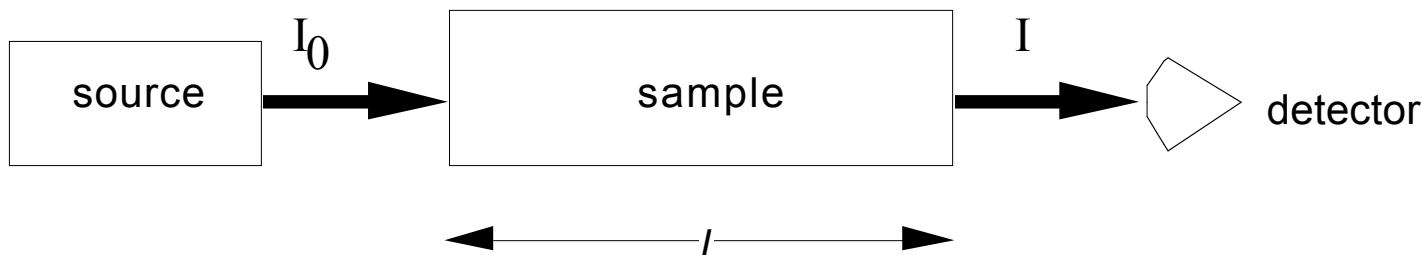
Spectroscopic techniques

1. Direct absorption (frequency and abs. intensity):
 - ✓ Intracavity spectroscopy (IC),
2. Indirect absorption (frequency only):
 - ✓ Photoacoustic spectroscopy (PA),
 - ✓ Laser induced fluorescence (LIF),
 - ✓ Photodissociation spectroscopy (UV/IR-PD).
3. Fourier-transform IR (FTIR)
4. (Stimulated) Raman scattering (RS or SRS)

What one can do?

- Measure frequencies and intensities of transitions to calculate structure and energy levels;
- Detect presence of different chemical species by detecting specific transitions;
- Measure concentration of chemical species, including short-living intermediates;
- Study dissociation/chemical kinetics;
- Study intramolecular ultra-fast processes:
 - a) by time-resolved spectroscopy with ps/fs lasers,
 - b) from frequency resolved spectra.

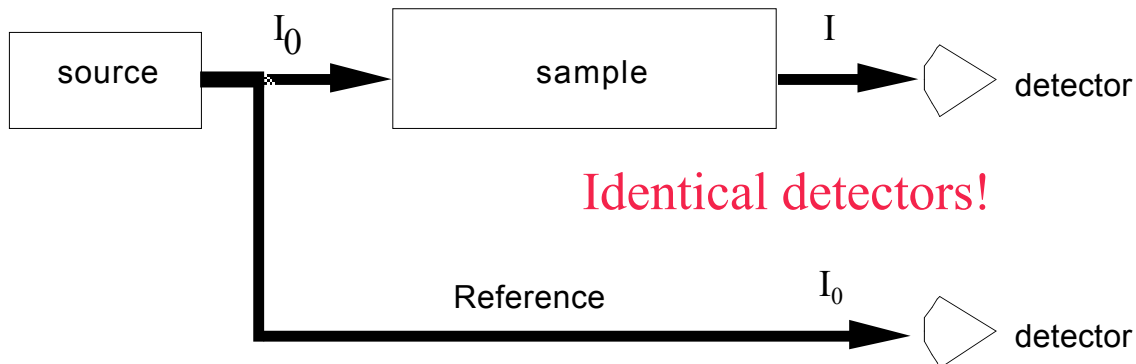
Absorption Spectroscopy



$$I(\nu) = I_0 e^{-nl\sigma(\nu)} \quad \Delta I(\nu) \simeq I_0 nl\sigma(\nu)$$

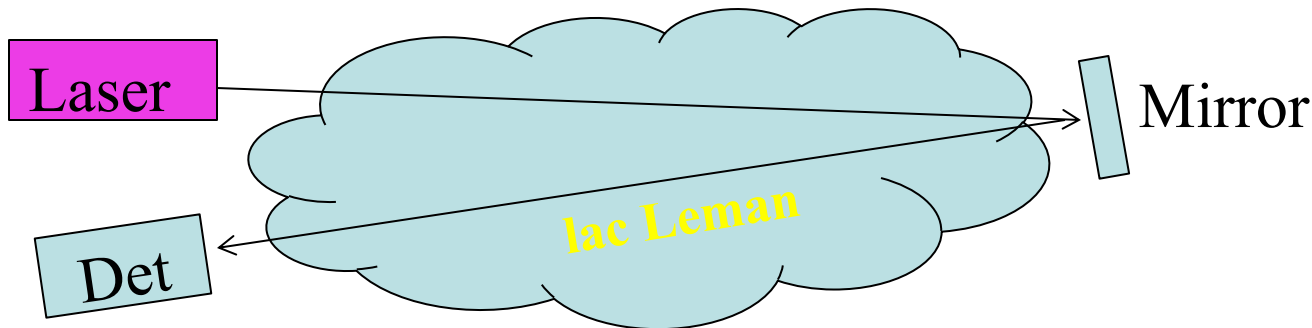
CF_3H ($\nu_{\text{CH}}=3$): $p=1\text{mBar}$ ($n=3 \cdot 10^{16} \text{ cm}^{-3}$), $l=10\text{cm}$, $\sigma=10^{-21} \text{ cm}^2$

$$\Delta I(\nu)/I_0 = 3 \cdot 10^{-4} = 0.03\%$$



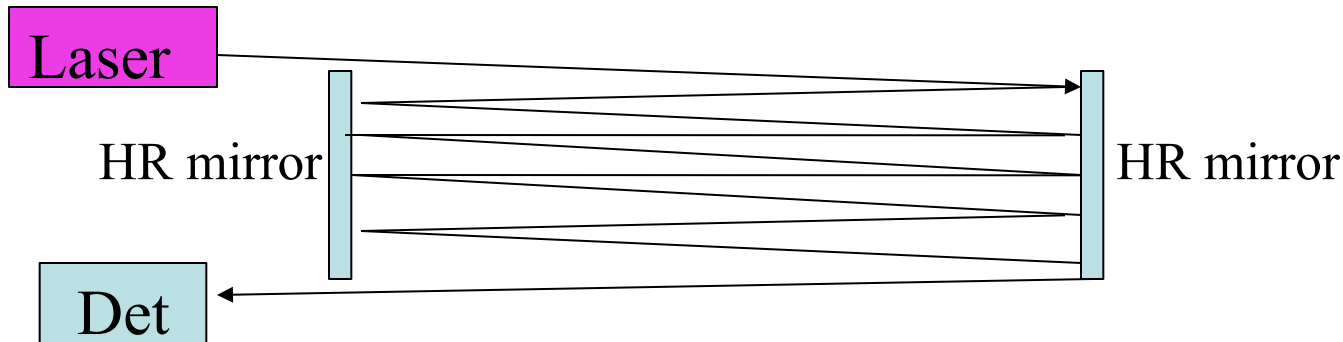
Direct laser absorption methods

1. Single long pathway: ($(\text{H}_2\text{O})_2$ above lake, $L=8$ km)
2. Multipath cells;
3. Intra-cavity spectroscopy;



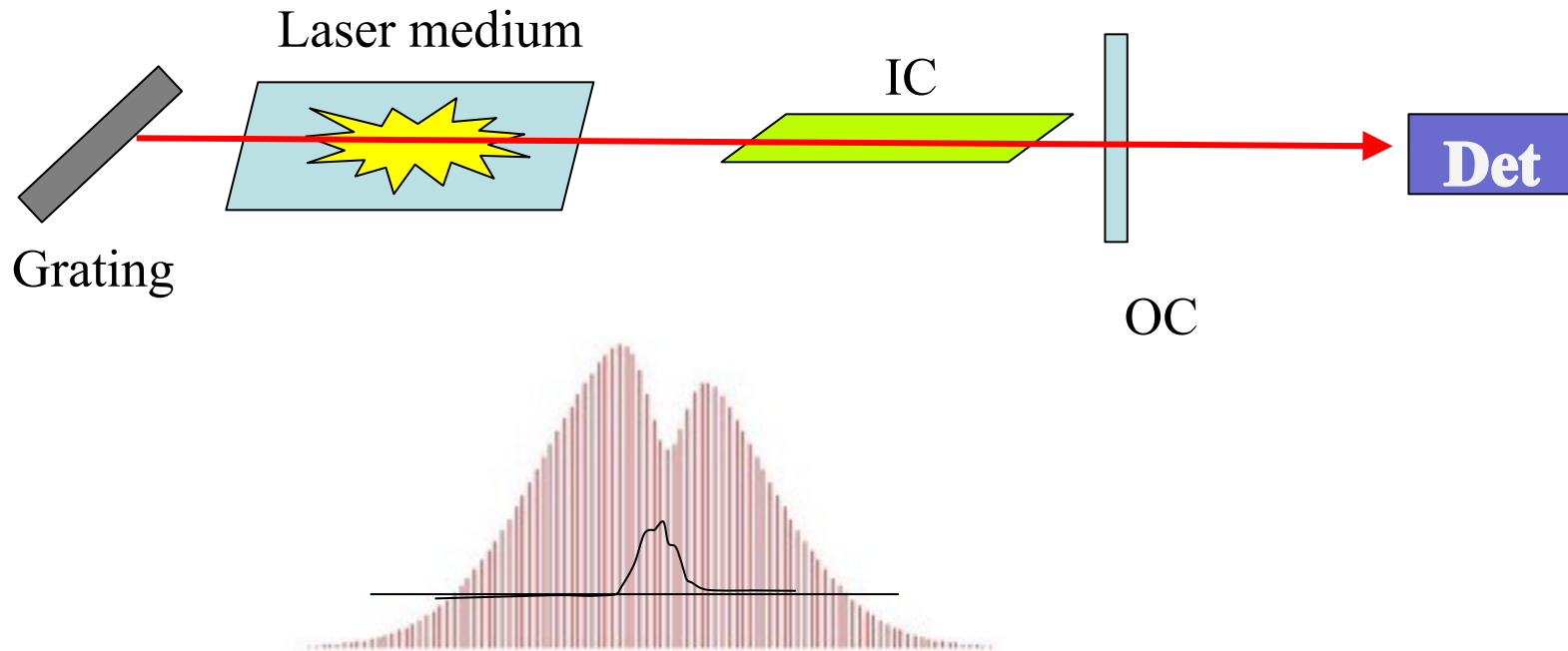
Direct laser absorption methods

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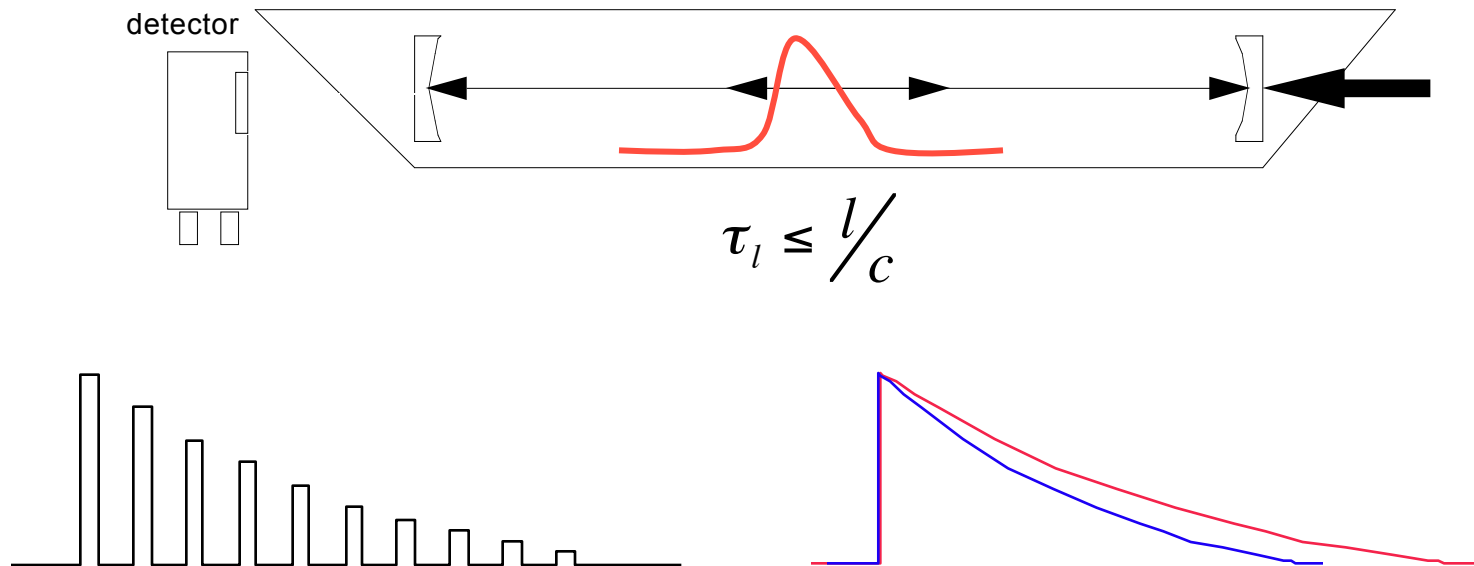
Direct laser absorption methods

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Direct absorption methods

1. Single long pathway: $((\text{H}_2\text{O})_2$ above lake, $L=8$ km)
2. Multipath cells;
3. Intra-cavity spectroscopy;
4. Cavity ring-down spectroscopy:



$$I = I_0 \exp(-\alpha(\lambda) \cdot t)$$

Application of absorption spectroscopy

- Concentration of chemical species;
- Frequencies and intensities of transitions.

Advantages:

- Absolute measurements of concentration/absorption cross section;
- Simplicity and universality.

Drawbacks:

- Low sensitivity, limited by laser power stability;
- Low selectivity.
- Significant thermal broadening ($T > 300$ K).

Indirect Absorption Methods

Photoacoustic spectroscopy:



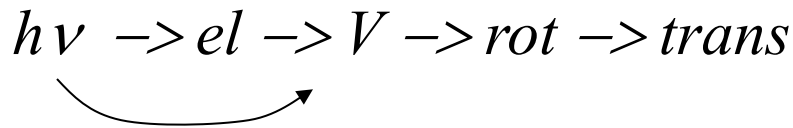
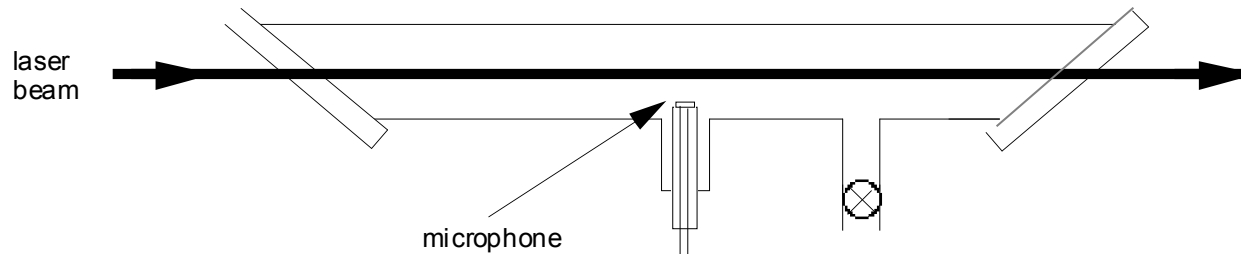
Thunderstorm:
Lightning followed
by **T**hunder

Why **T**hunder?

1. Heating of air inside the discharge channel,
2. Acoustic shockwave

Indirect Absorption Methods

Photoacoustic spectroscopy:



Direct absorption:

$$S_a = I_0 \cdot (1 - n \cdot l \cdot \sigma), \quad n \cdot l \cdot \sigma \ll 1;$$

Baseline : I_0 .

Photo acoustic signal: $S_{pa} = \alpha \cdot I_0 \cdot n \cdot l \cdot \sigma,$

Zero baseline!

Photoacoustic spectroscopy:

$$d \gg \frac{1}{n \cdot \sigma}$$

Advantages:

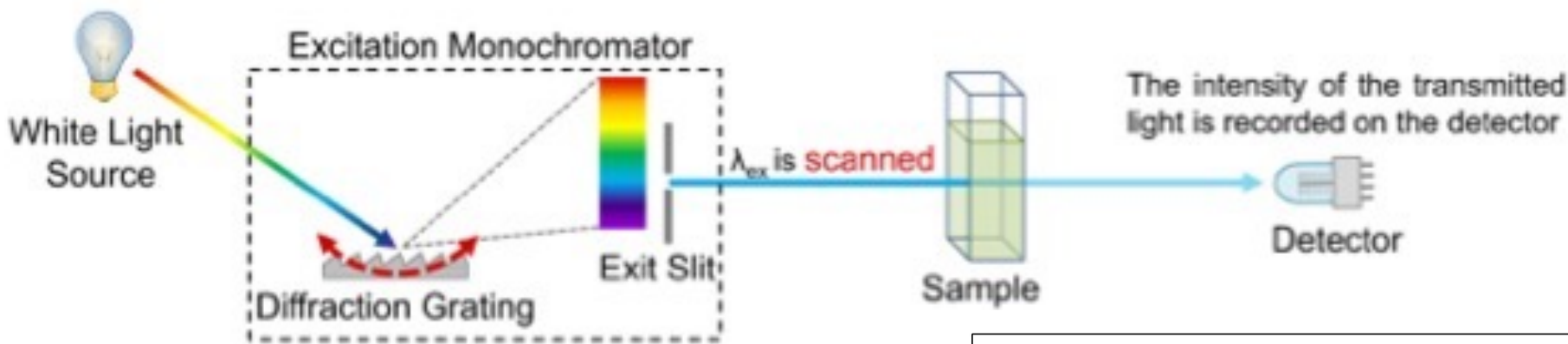
- 1) Extremely high sensitivity;
- 2) Simple and inexpensive;
- 3) Universal.

Disadvantage:

- 1) Collisional =>
 - Pressure broadening,
- 2) No absolute intensity
- 3) Requires pulsed tunable laser
- 4) Requires a vacuum pump

Used for laboratory studies/detections

Direct absorption spectrometers

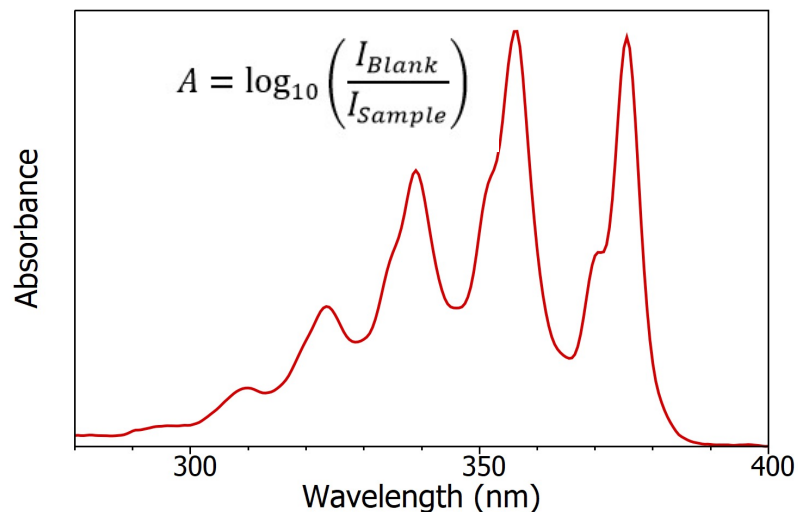


UV-Vis or IR absorption:

“Blanc” spectrum with no sample, I_0

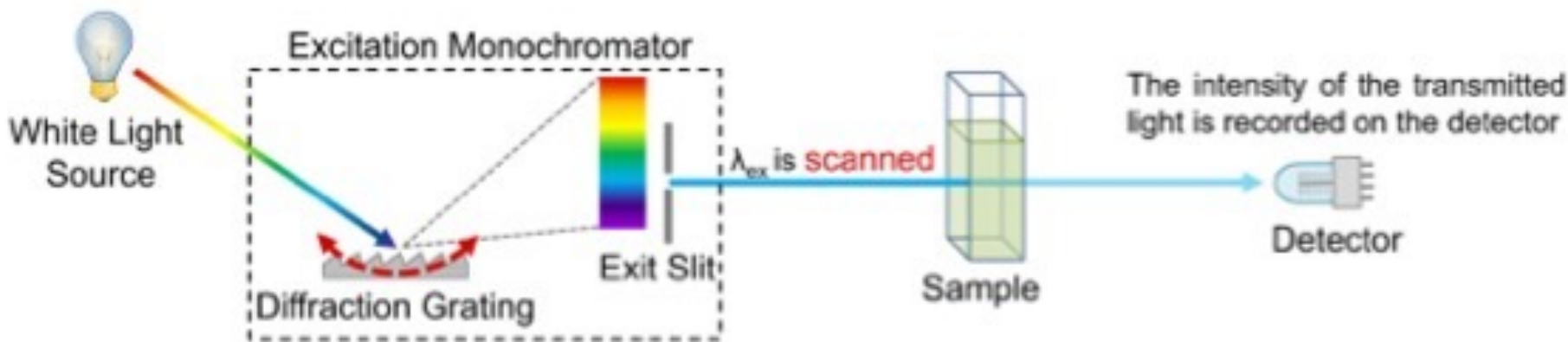
”Sample” spectrum with sample, I .

$$A = \log_{10} \left(\frac{I_0}{I} \right) = \epsilon c l$$



Typically: the spectrum is fit by a linear combination of the library spectra to get concentrations of the analytes in solution

Direct absorption spectrometers



- + Low cost, reliable
- + Universal, suitable for multi compound samples
- + Direct determination of concentration.
- Low sensitivity (solution; high concentration of sample)
- Non-selective
- Small dynamic range (=min to max concentration)
- Sensitive to sample T

Emission spectroscopy

$$I = \alpha \cdot \left\langle \varphi_{el}^i \left| \overrightarrow{\mu}_{el} \right| \varphi_{el}^f \right\rangle \left\langle \varphi_{vib}^i(r) \left| \varphi_{vib}^f(r) \right\rangle \right\};$$

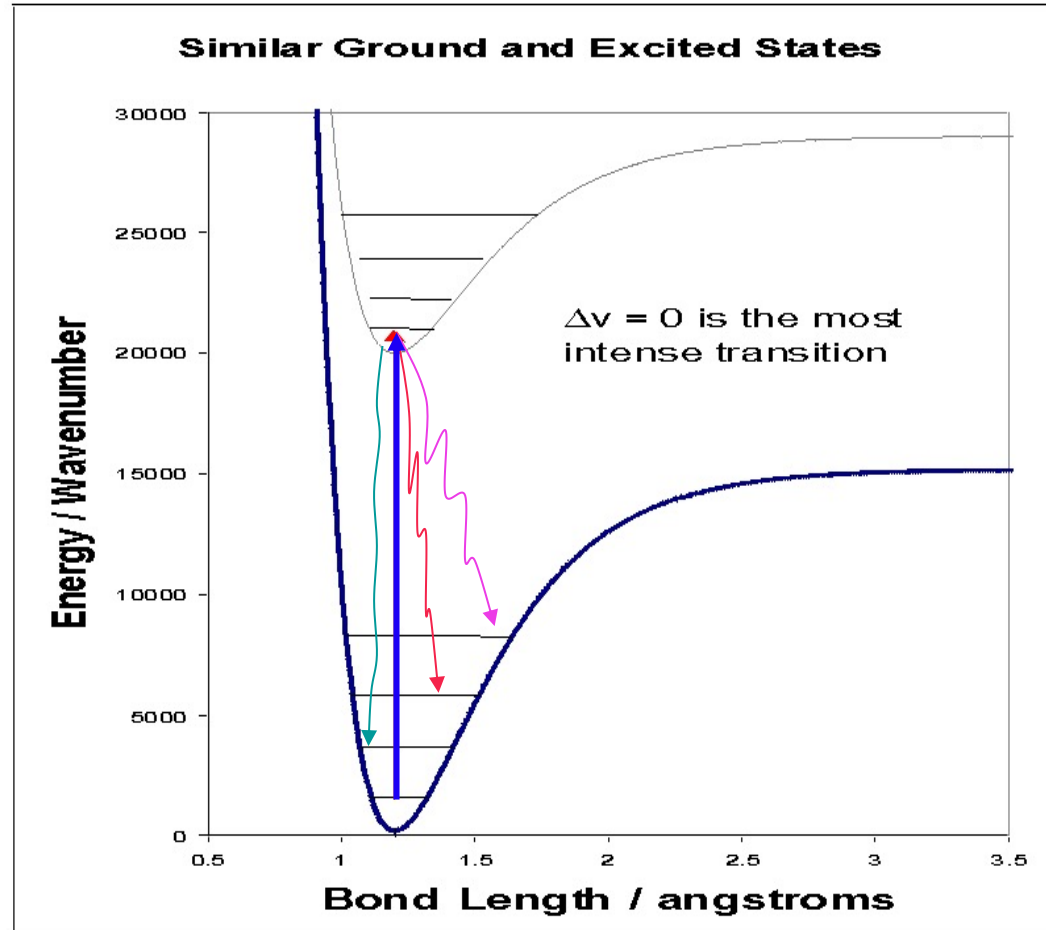
Excitation + emission =>
Compared with absorption:
higher selectivity.

$$\lambda_{em} > \lambda_{exc}$$

- Lines positions
<=> Energy levels;
- Lines intensities
<=> Franck-Condon factor

Background free:

- high sensitivity
- high dynamic range



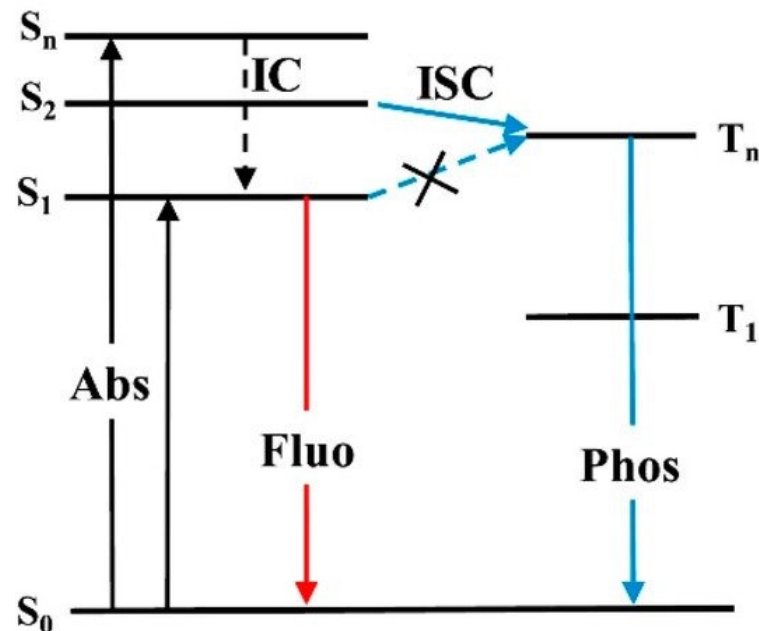
Induced Fluorescence/Phosphorescence

Kasha's/Vavilov rule:

In many polyatomics in solution an emission spectrum often is nearly independent of the excitation wavelength.

When K-V rule is obeyed:

vibrational levels of S_0 ;
if, in addition S_1 and S_0 are similar:
IF/IP spectrum $\sim$ Absorption

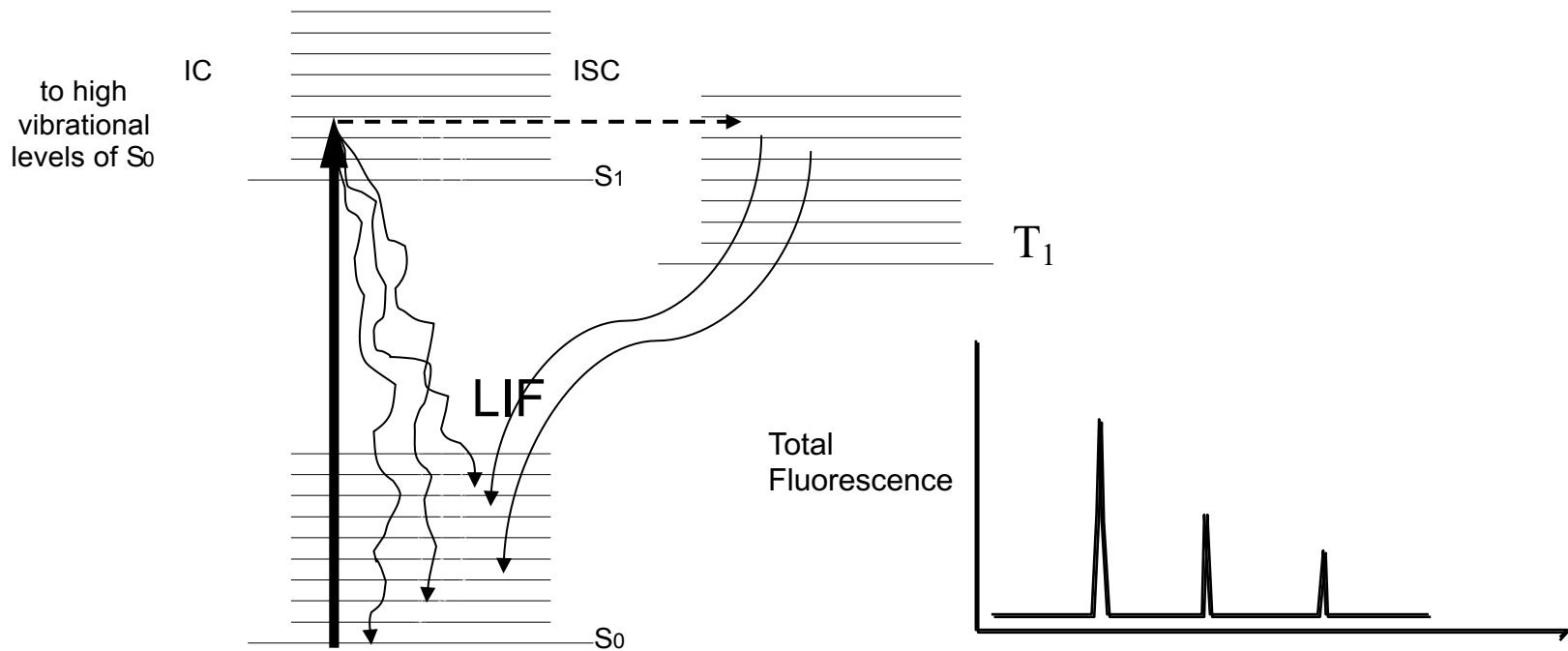


Excitation to $S_2, S_3, \dots$ often is followed by fast internal conversion to S_1 and, subsequently, by fast vibrational relaxation to $v=0$ in S_1

This rule has many exceptions.

There is no vibrational relaxation in the gas phase.

Induced Fluorescence/Phosphorescence



$S_0 \leftarrow S_1$: Fluorescence (10-100 ns in UV/Vis, 10-100 μ s in IR)

$$\frac{1}{\tau_{21}} = A_{21} = \frac{8\pi h \nu_{12}^3 B_{21}}{c^3}$$

$S_0 \leftarrow T_1$: Phosphorescence (formally forbidden; ms to s)

Emission spectroscopy

Case (1):

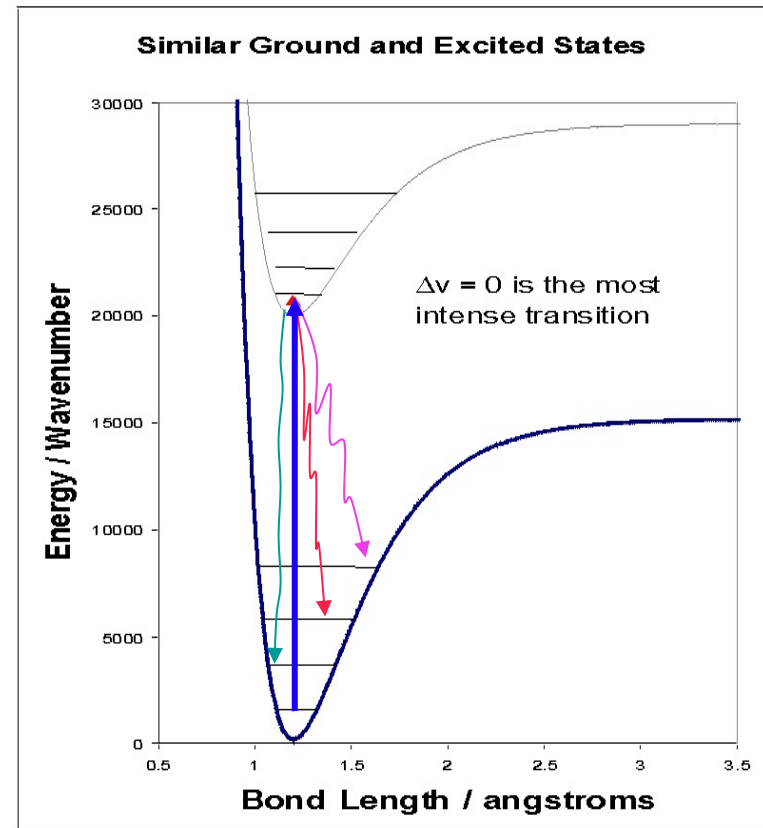
Excitation wavelength is scanned, while total fluorescence is collected (LIF):
(rovibronic structure of S_1 and FC factor)

Case (2):

Excitation wavelength is scanned, emission at a specific transition only is detected (rovibronic structure of S_1 and additional selectivity)

Case (3):

Excitation wavelength is fixed, emission is detected as a function of wavelength (rovibronic structure of S_0 , FC factor)



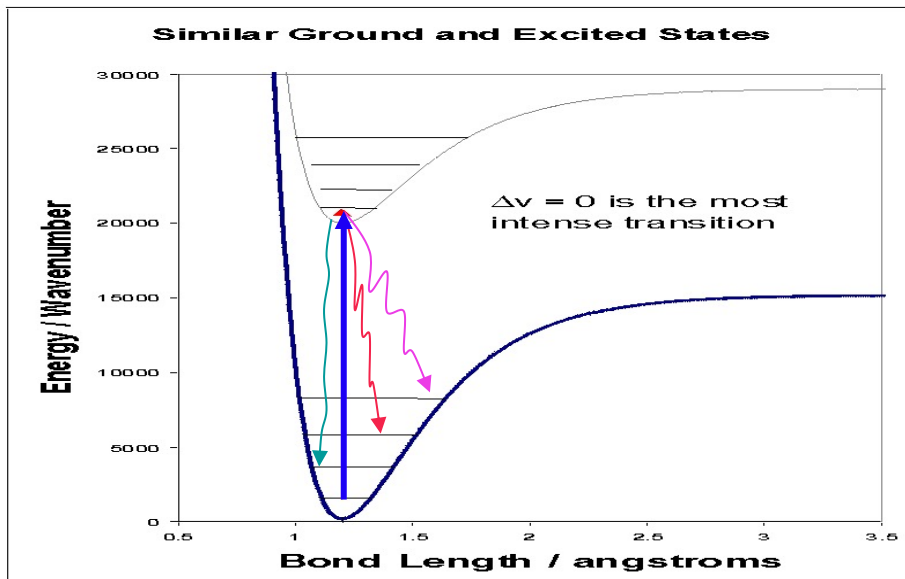
Emission spectroscopy

Advantages:

- 1) Extremely sensitive (1);
limited by quantum yield and
design of light collection system
- 2) Cases (2) and (3) are less sensitive
but highly selective.

Disadvantages:

- 1) Require high fluorescence
quantum yield;
- 2) Scattered light discrimination;
- 3) Difficult for triplet states and for
vibrational spectroscopy



$$\frac{1}{\tau_{21}} = A_{21} = \frac{8\pi h \nu_{12}^3 B_{21}}{c^3}$$

Laser-induced fluorescence spectroscopy

(Case 1. Research only)

High spectral resolution =

Selectivity

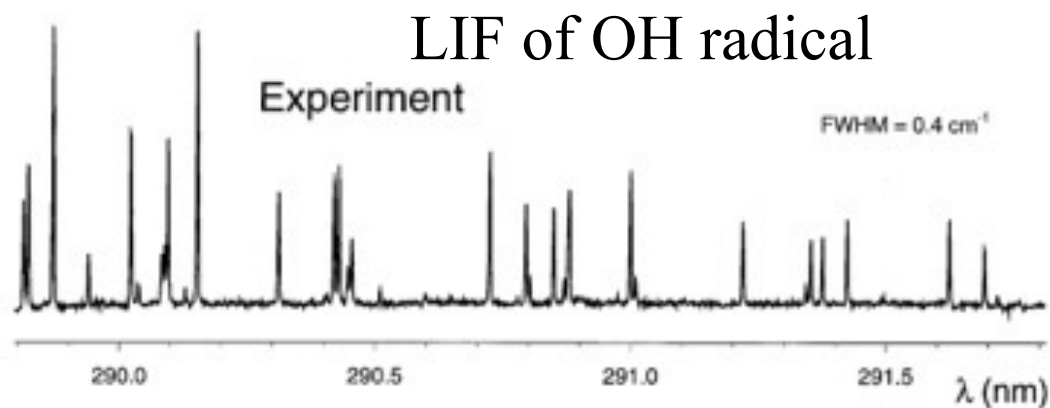
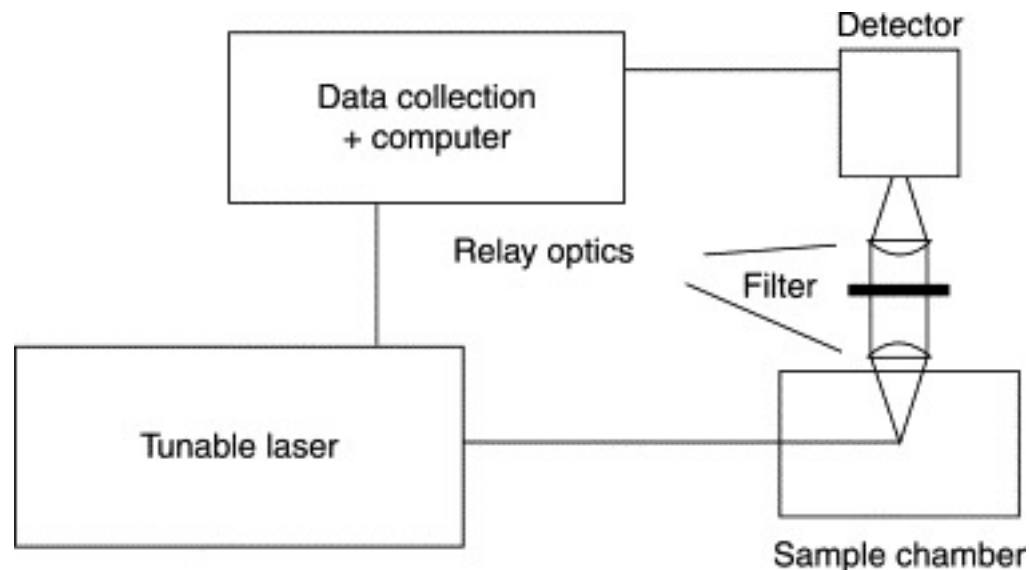
High spectral brightness
and directionality =>

Sensitivity

- Optimized geometry of optics to reduce scattered light (baffles, cut-off glass filters).

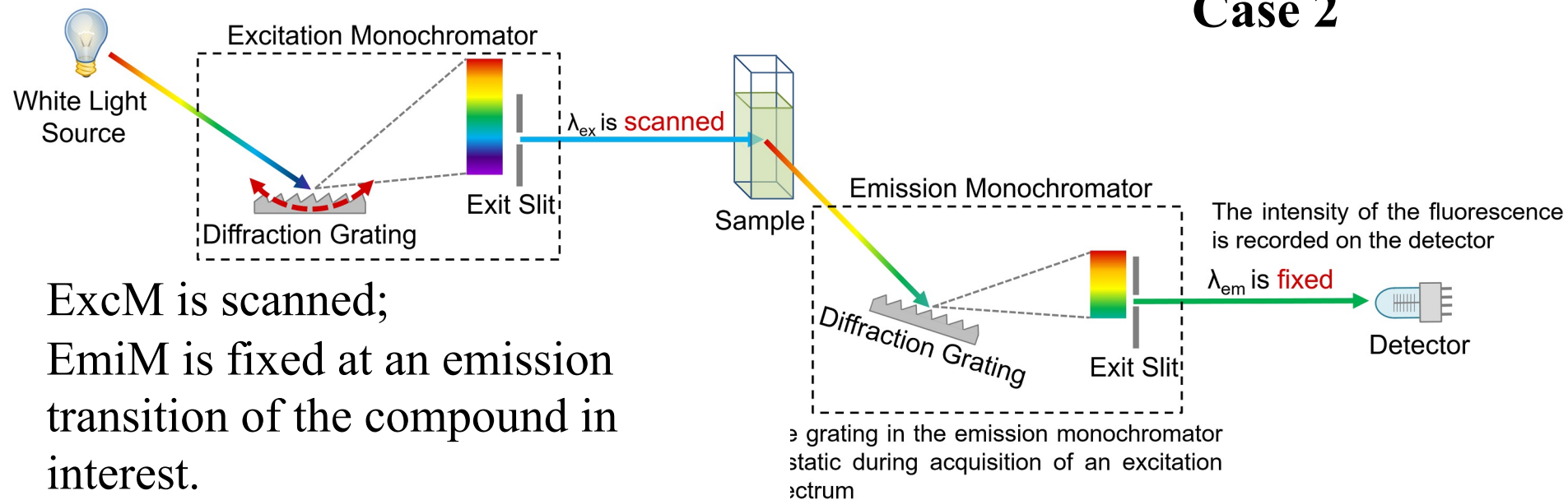
$$\lambda_{LIF} > \lambda_{Laser}$$

- Requires a tunable laser



Excitation spectra with spectrofluorometer

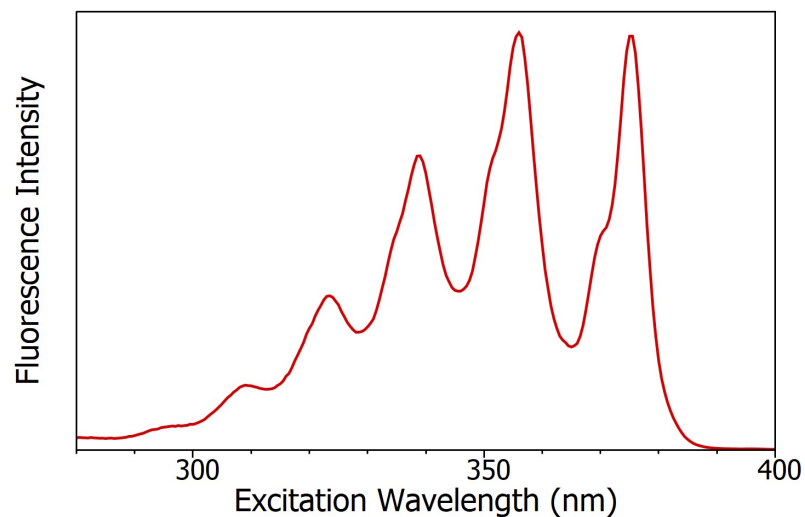
Case 2



ExcM is scanned;
EmiM is fixed at an emission transition of the compound in interest.

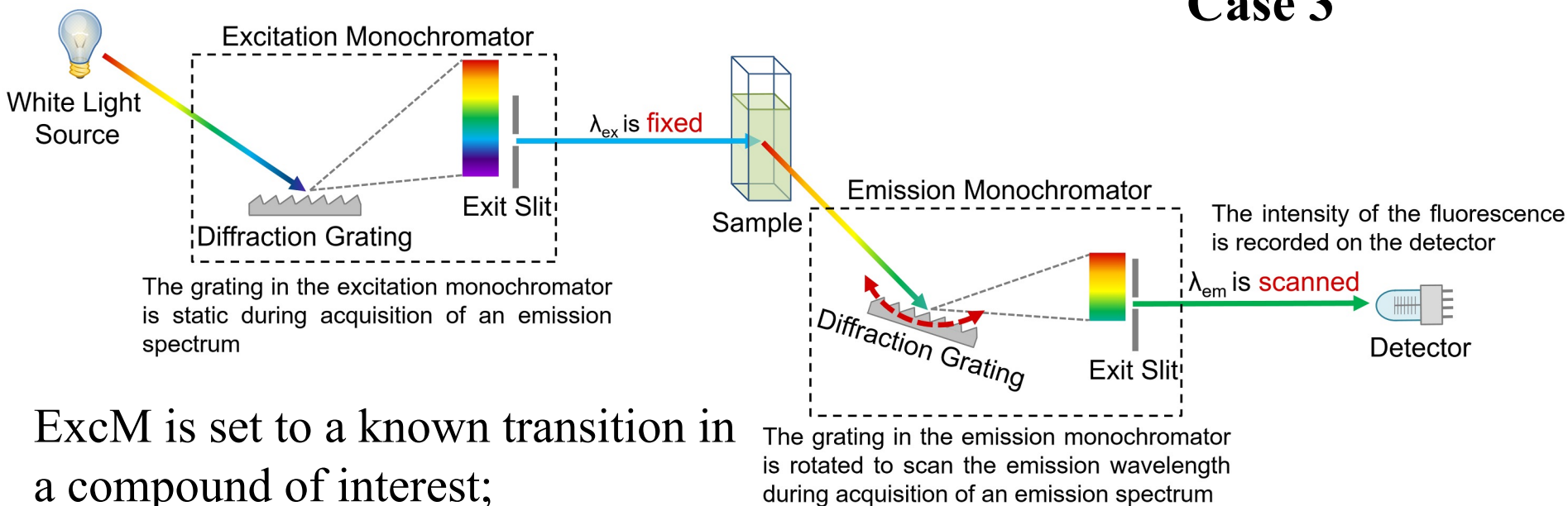
Often (K-V rule) yields nearly absorption spectrum

- + High selectivity
- + High sensitivity
- + High dynamic range
- One molecule at time
- Technical complexity



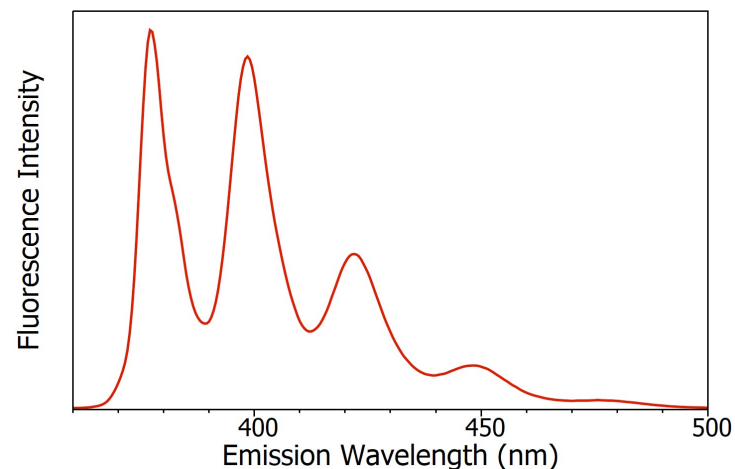
Emission spectra with spectrofluorometer

Case 3



ExcM is set to a known transition in a compound of interest;
EmiM is scanned across the region of emission from the excited state of the compound in interest.

- + High selectivity
- High sensitivity
- One molecule at time
- Technical complexity



Fluorescence emission spectrum of anthracene in cyclohexane

Photodissociation action spectroscopy



The more light is absorbed the more fragments appear

A or B fragment is detected; the number of fragments $\propto \sigma_{MA}$

Background-free technique => *highly sensitive*

Photofragmentation techniques:

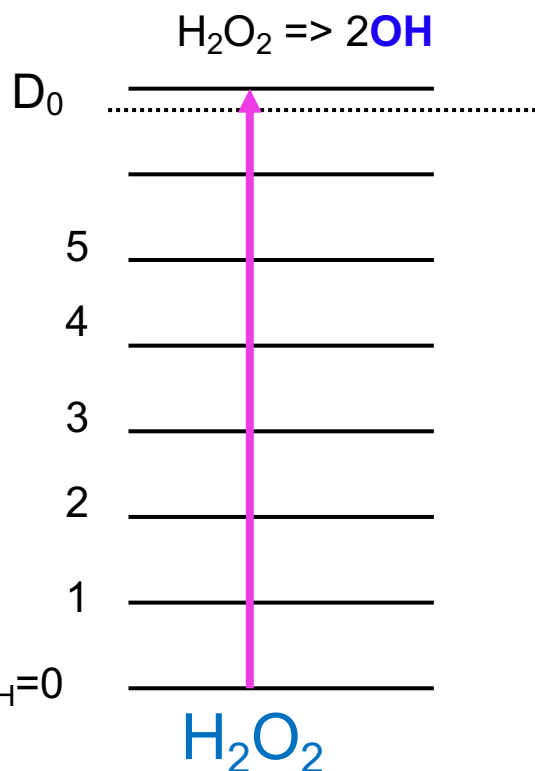
- Overtone excitation
- UVPD direct
- Vibrationally mediated UVPD
- IRMPD

Detection techniques:

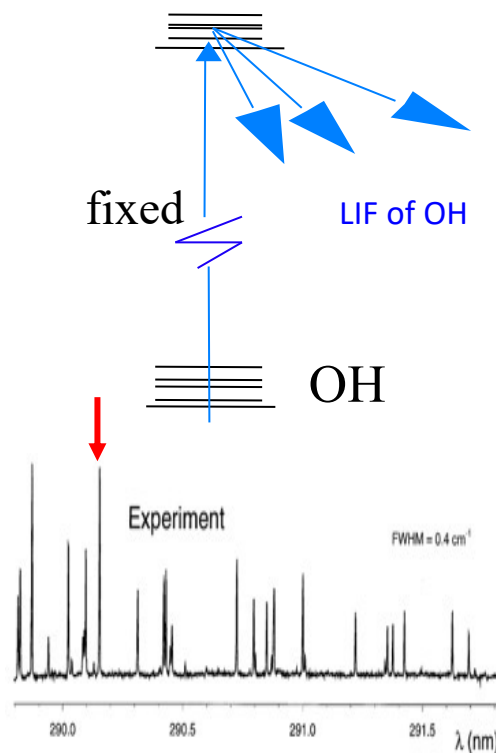
- LIF
- Mass spectrometry

Photodissociation action spectroscopy

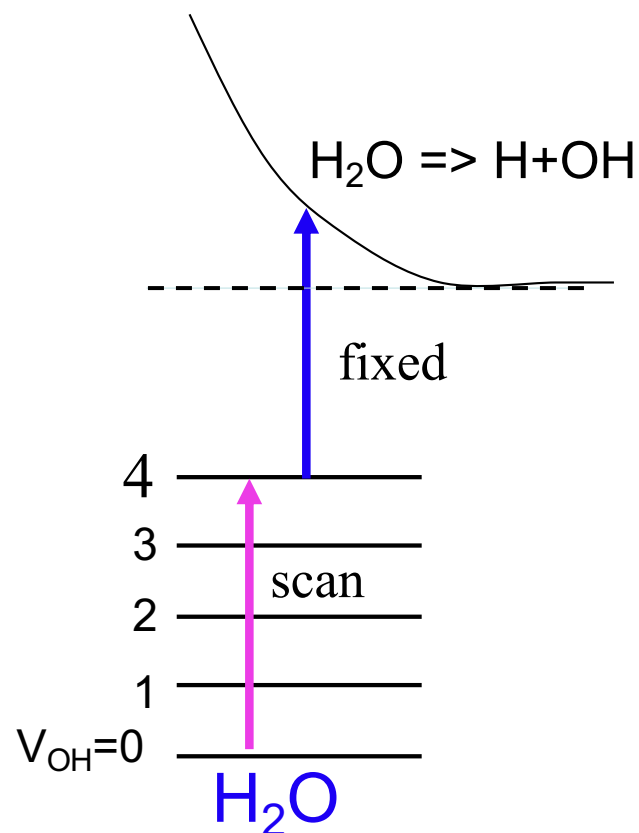
Vibrational overtone photodissociation



LIF detection

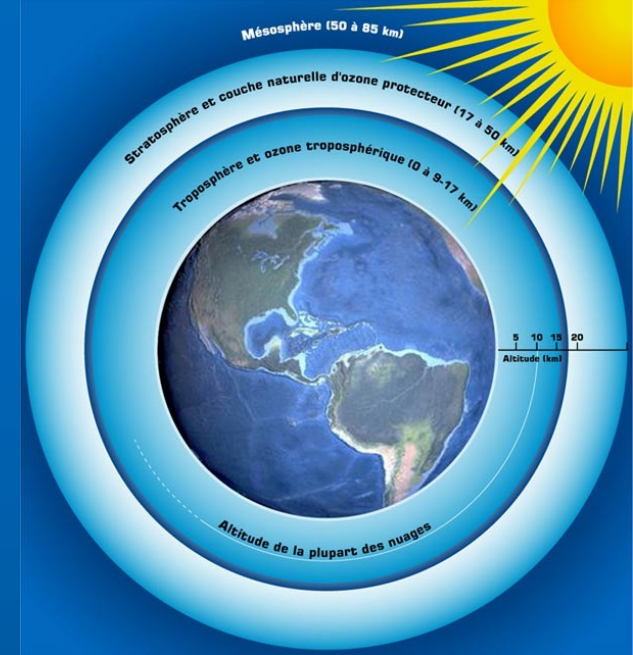


Vibrationally mediated UV dissociation

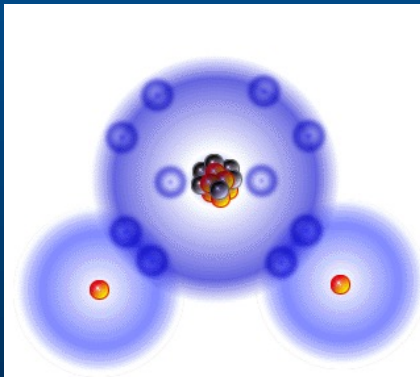


Atmospheric water vapor :

- Main absorber (70 %) of the incoming sunlight;
- Strong green house gas (IR absorption);



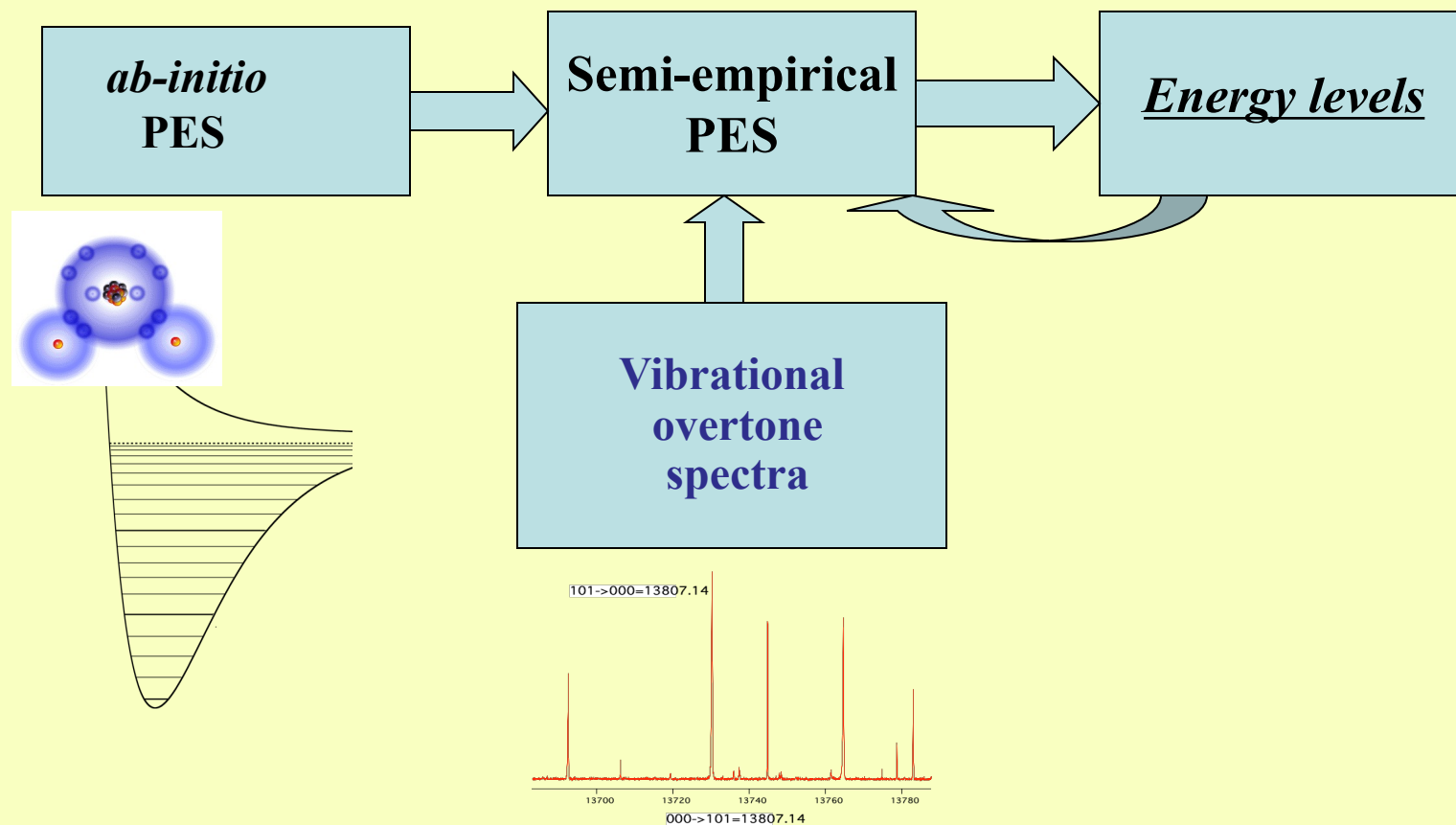
Water vapor absorption must be well modeled!



Fundamental to H₂O molecule:

- Ro-vibrational energy levels,
 - Intensity of transitions.

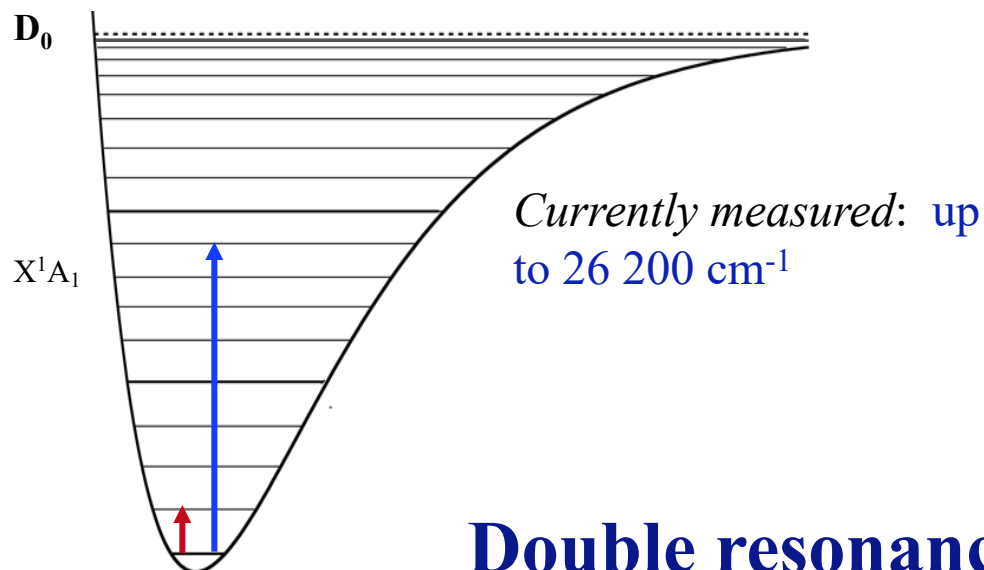
Obtaining overtone transitions



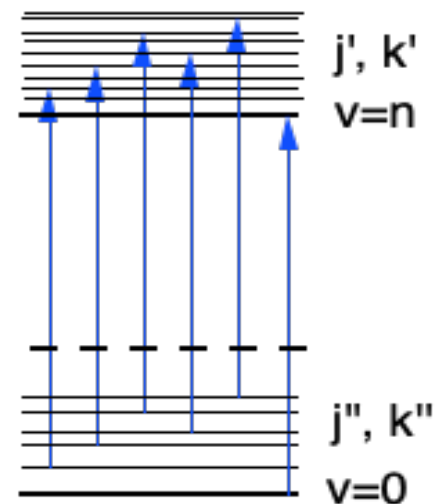
Problems and solutions

- Very low intensity of high vibrational overtone transitions:

$$\frac{\sigma(n \leftarrow 0)}{\sigma(1 \leftarrow 0)} \approx 10^{n+1}$$

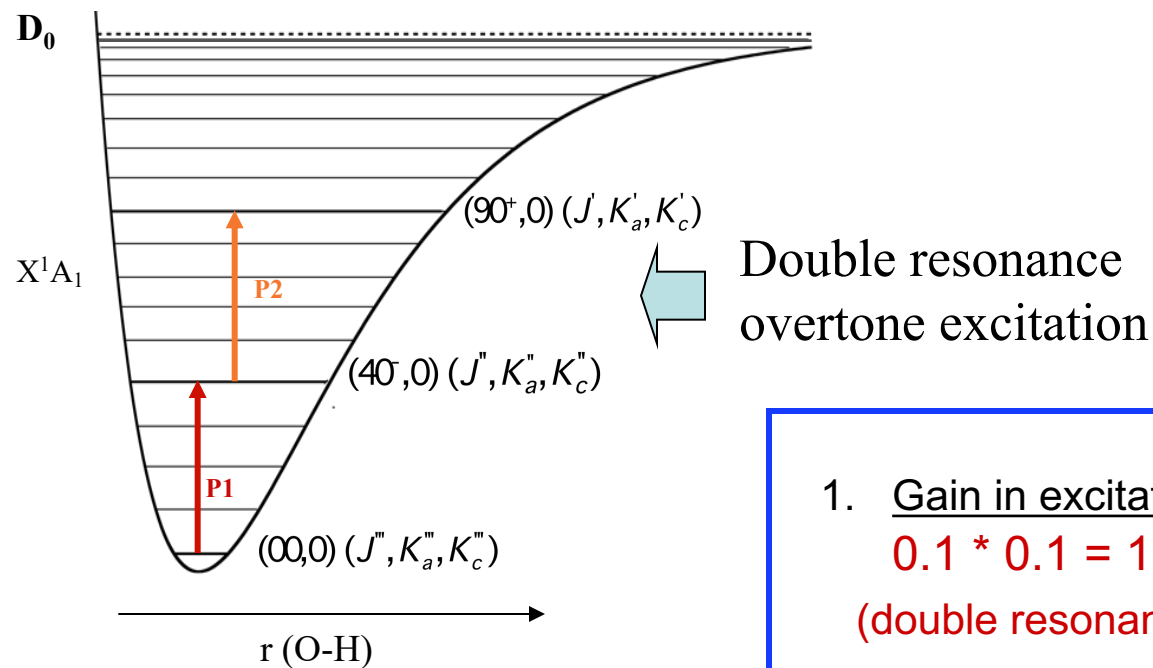


- Rotational congestion due to thermal distribution



Double resonance overtone excitation
combined with photofragment detection

Energy Level Diagram

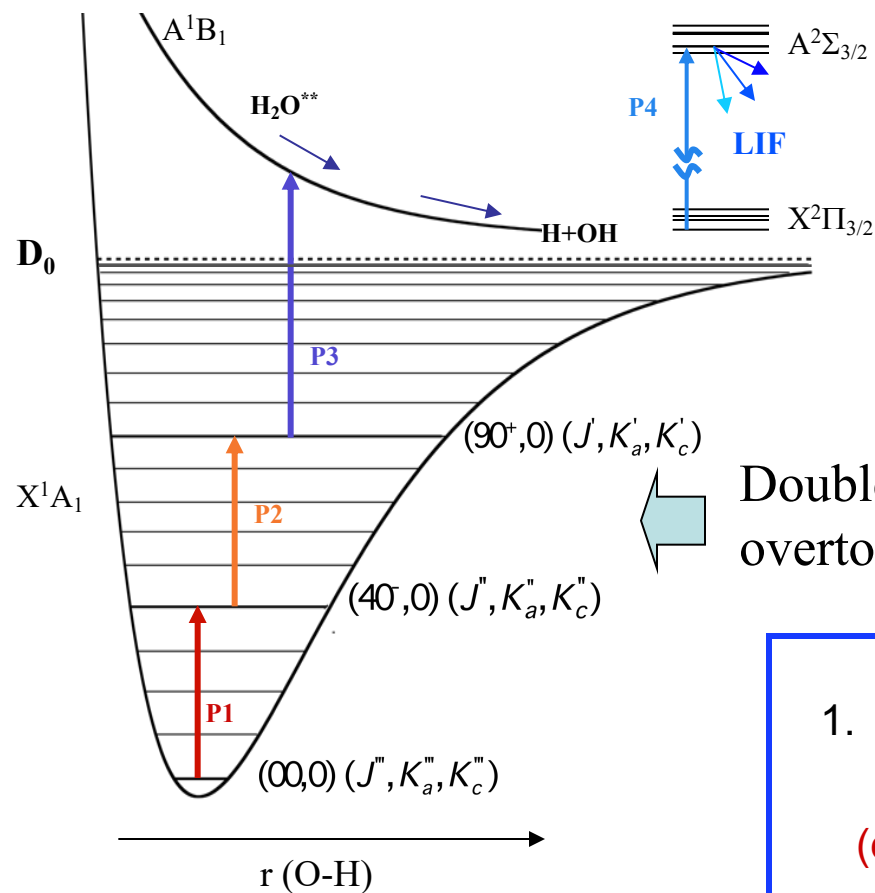


1. Gain in excitation efficiency:

$0.1 * 0.1 = 10^{-2}$ **vs** $10^{-5}-10^{-6}$
 (double resonance) (direct excitation)

2. Rotational pre-selection => simpler assignment

Energy Level Diagram



Photofragmentation
detection

High selectivity and sensitivity

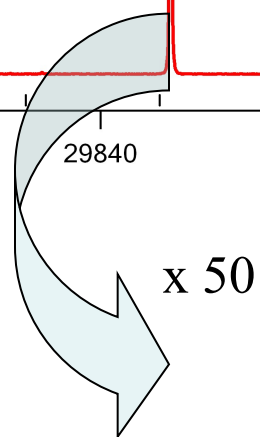
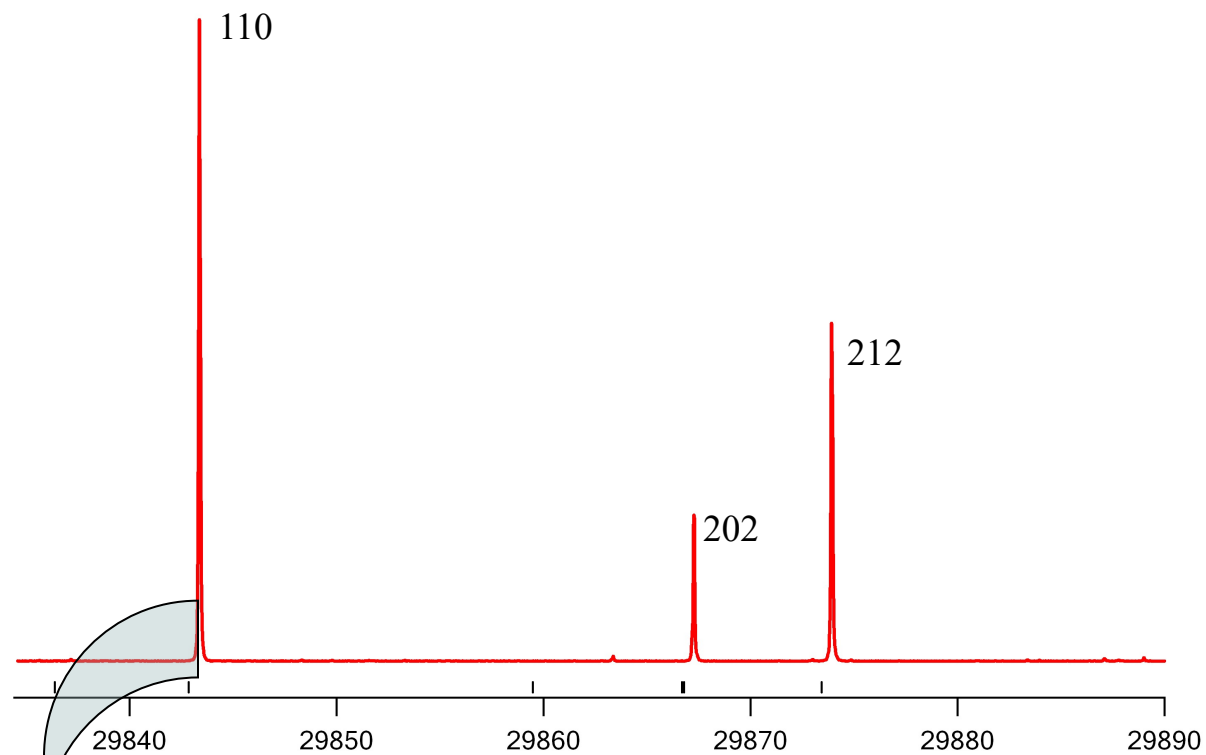
Double resonance
overtone excitation

1. Gain in excitation efficiency:

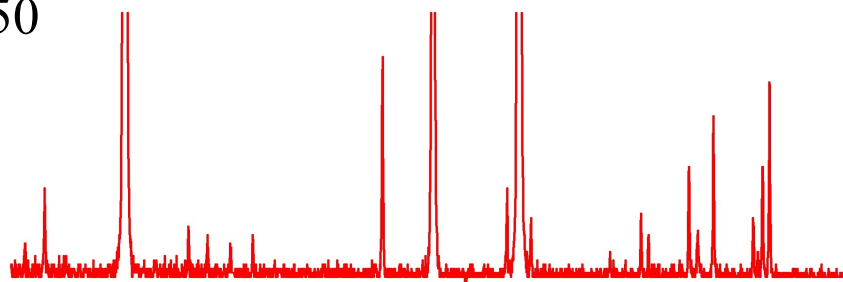
$0.1 * 0.1 = 10^{-2}$ **vs** $10^{-5}-10^{-6}$
(double resonance) (direct excitation)

2. Rotational pre-selection => simpler assignment

9th OH-stretch overtone

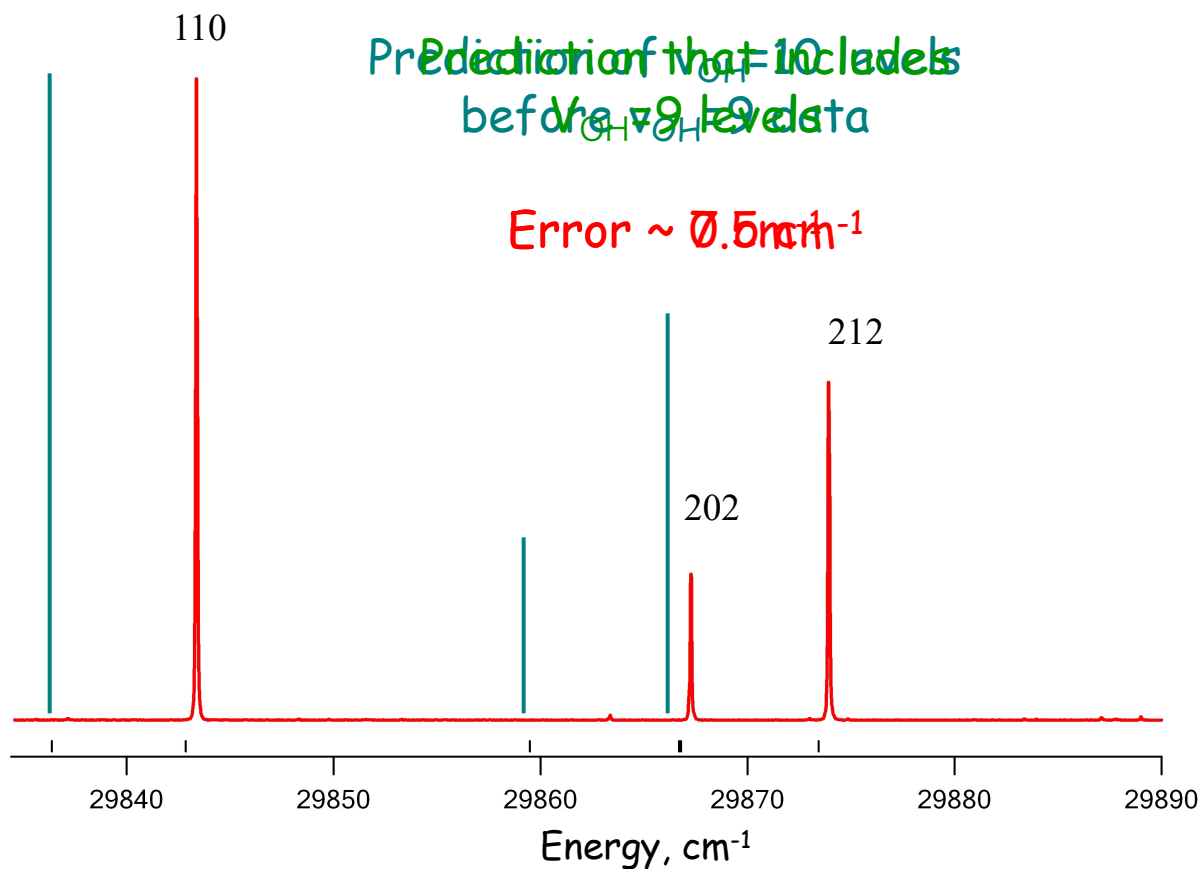


x 50

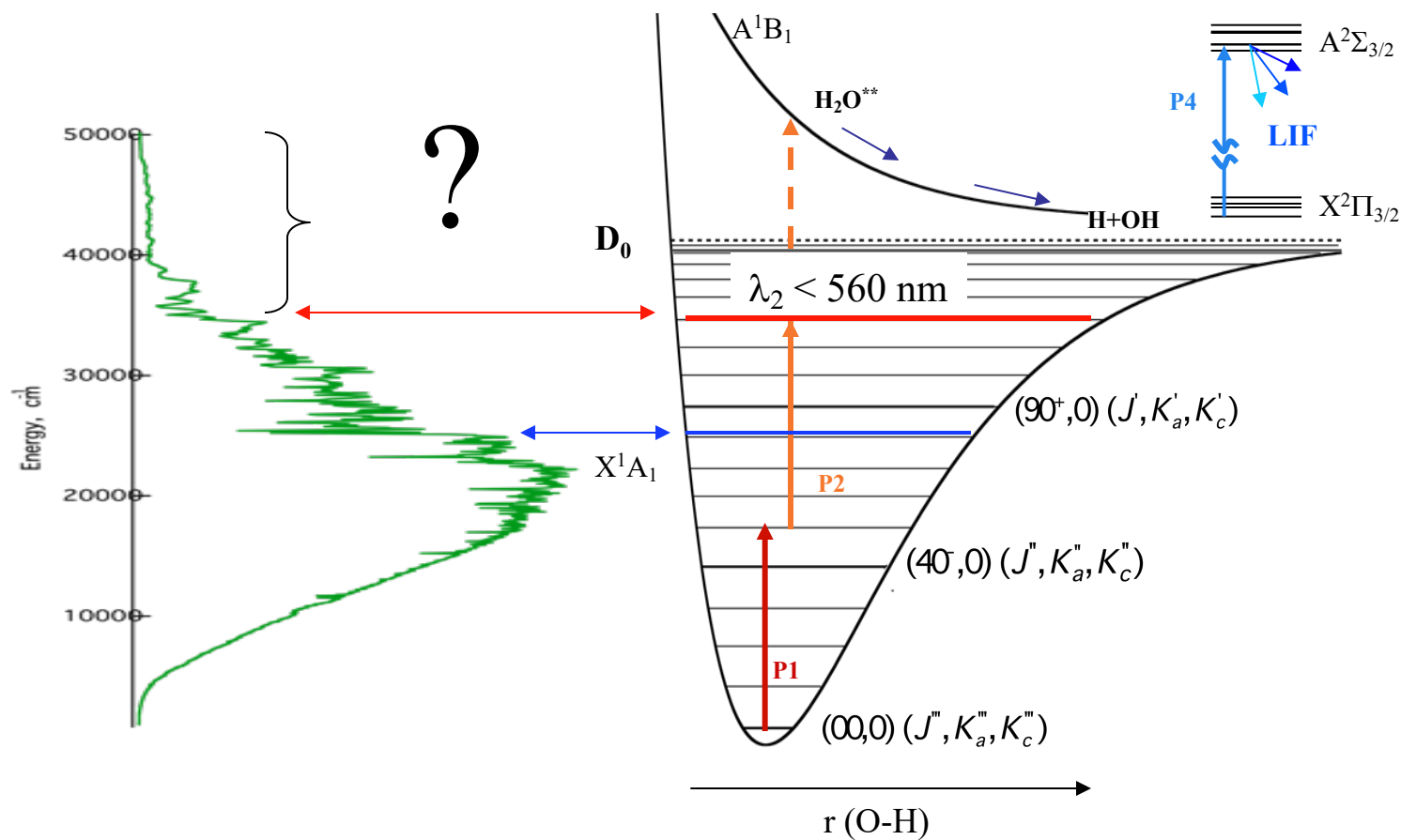


S/N > 3000

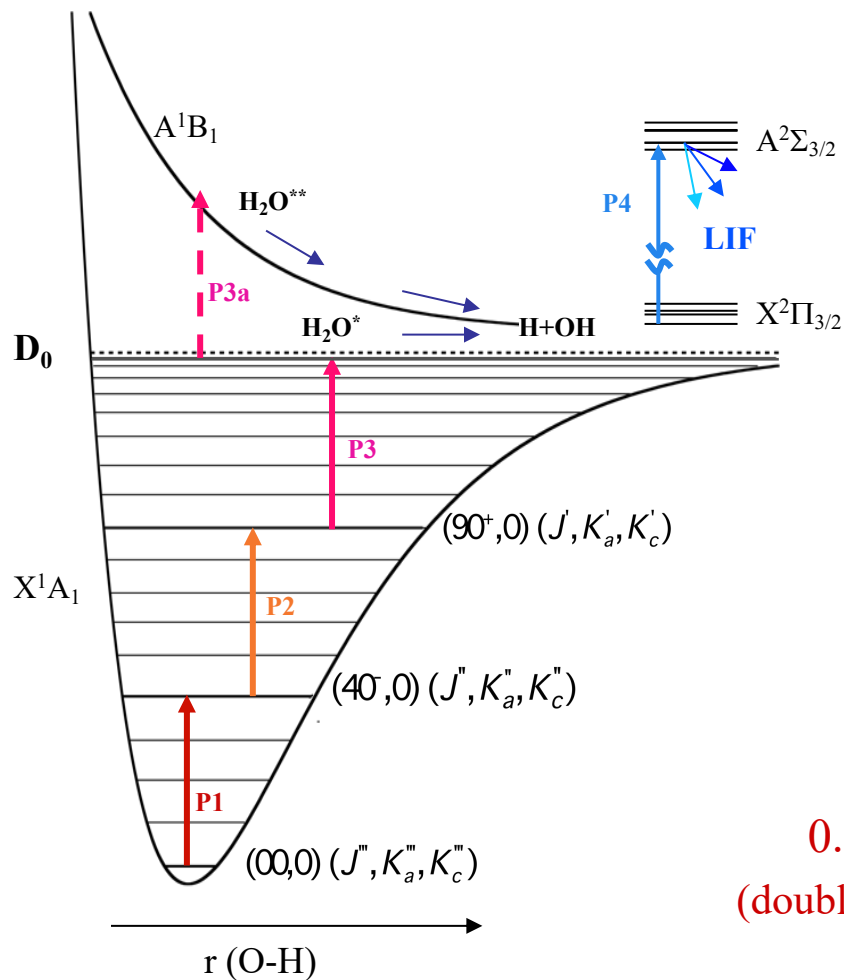
9th OH-stretch overtone



Solar irradiance spectrum



Triple-Resonance Excitation

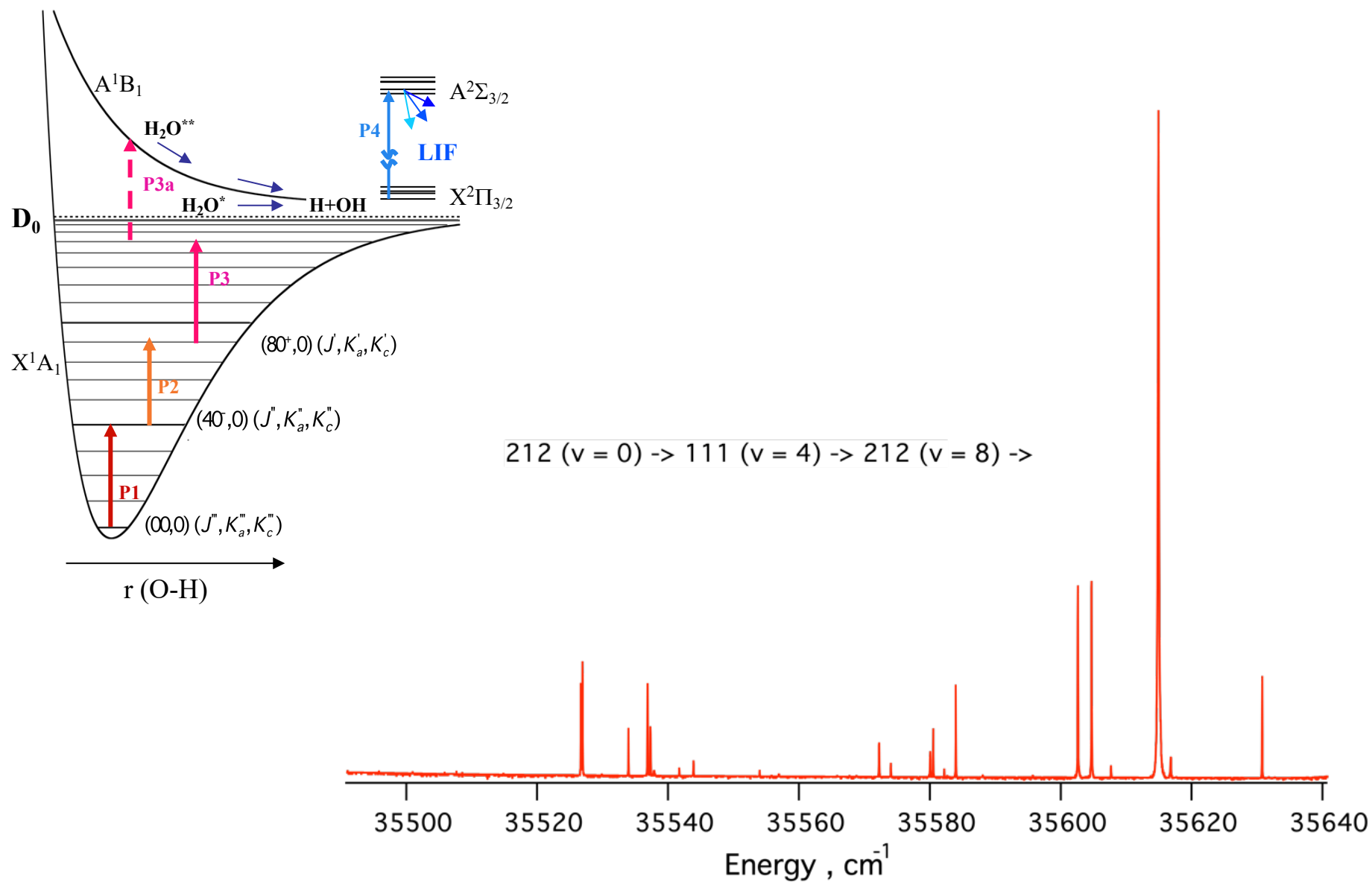


Gain in excitation efficiency:

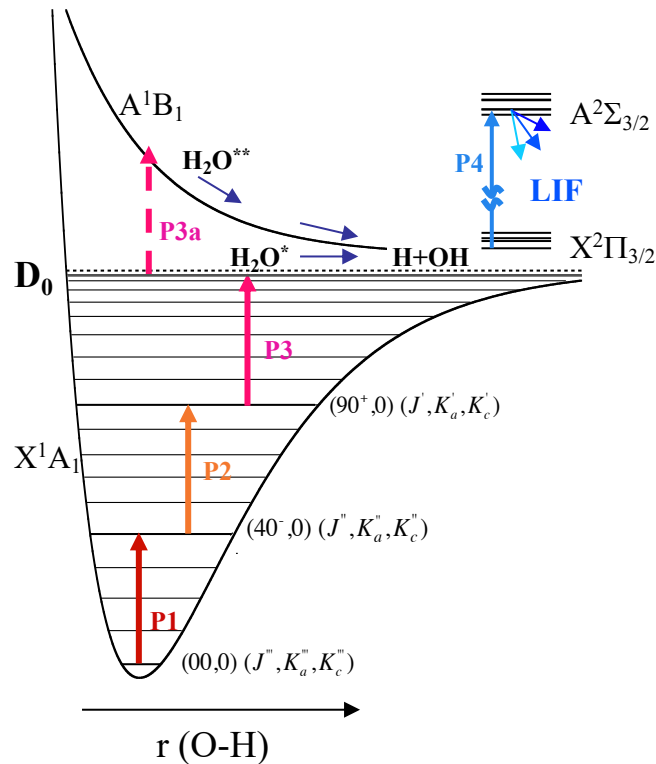
$$0.1 * 0.1 * 0.1 = 10^{-3} \quad \text{vs} \quad 10^{-15} - 10^{-16}$$

(double resonance) (direct excitation)

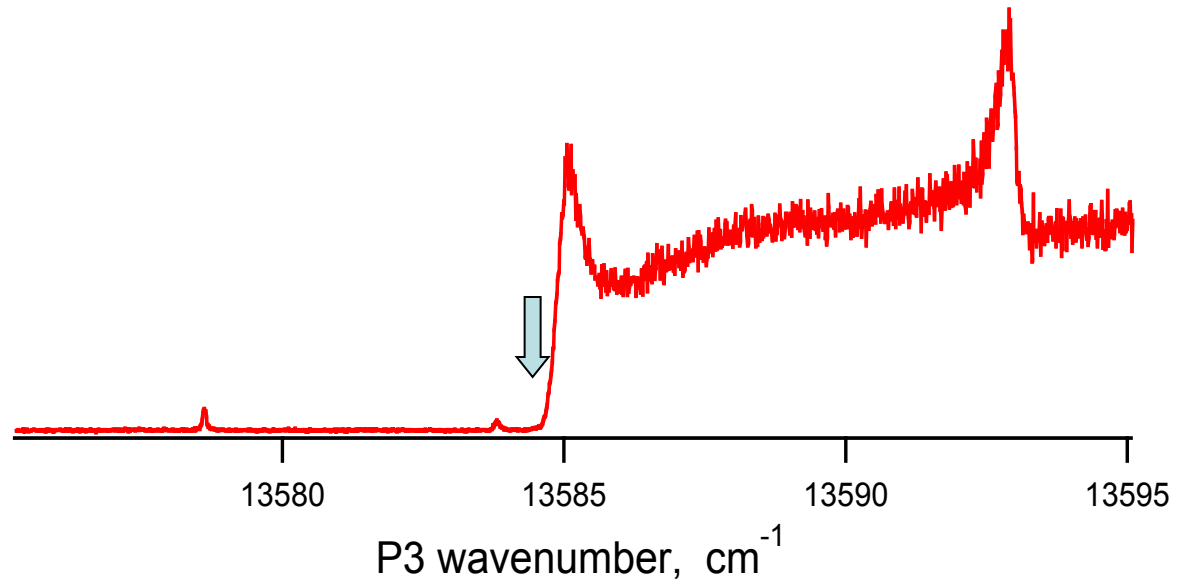
129th OH-stretch overtone



Precise determination of D_0 in water



- Start from rotationless state of H_2O
- Prepare H_2O with no rotational energy
- Detect OH with no rotational energy



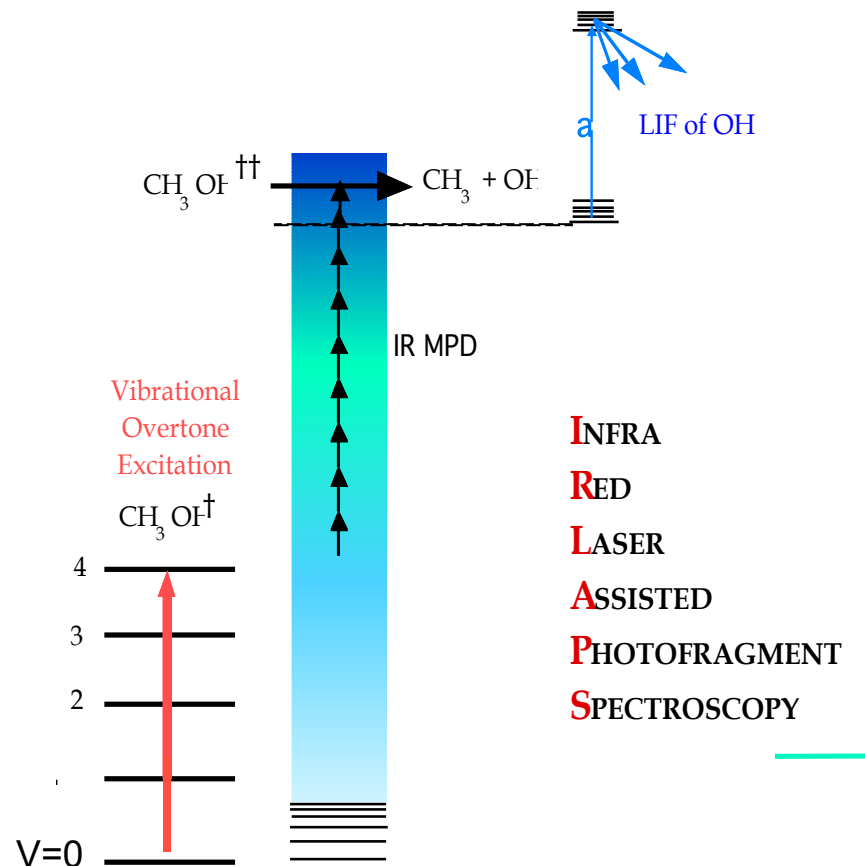
Triple resonance
overtone excitation

$$D_0 = 41\,145.94 \pm 0.15 \text{ cm}^{-1}$$

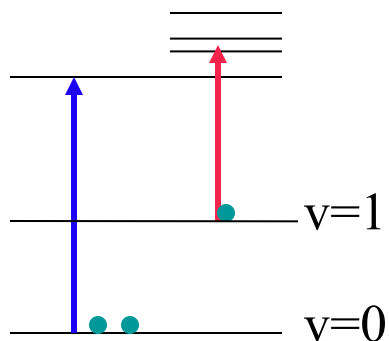
Made in EPFL

Photofragmentation

Energy diagram of double
resonance overtone IRLAPS:



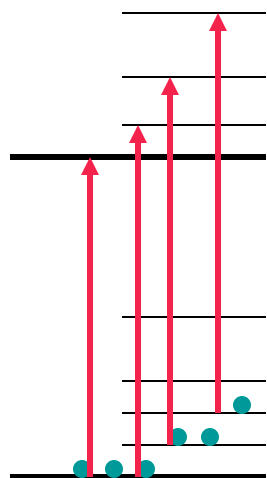
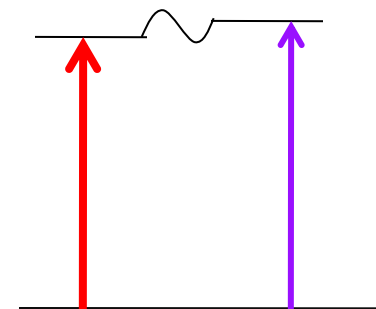
State-resolved laser Spectroscopy



Spectral congestion:

1. Hot bands,
2. Rotational structure,
3. Anharmonic couplings,...

$$E_{vib}(5OH) \approx E_{vib}(4OH + CH)$$

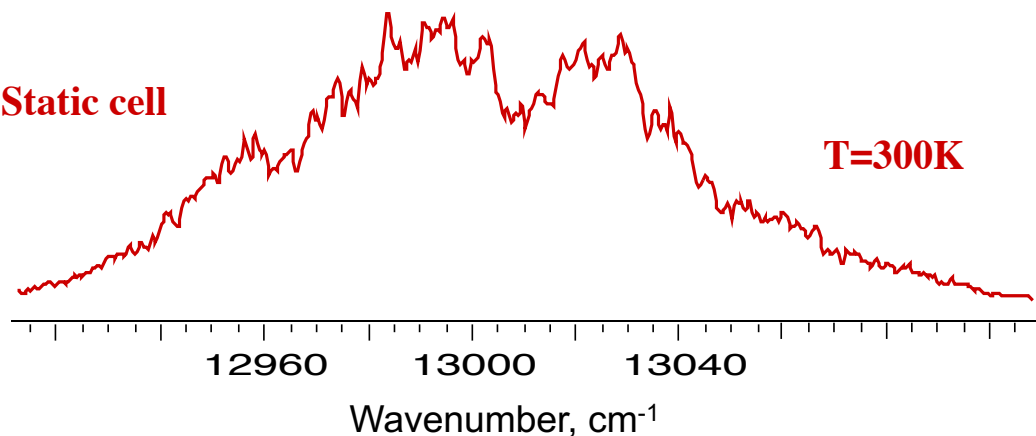


CH₃OH

5ν₁

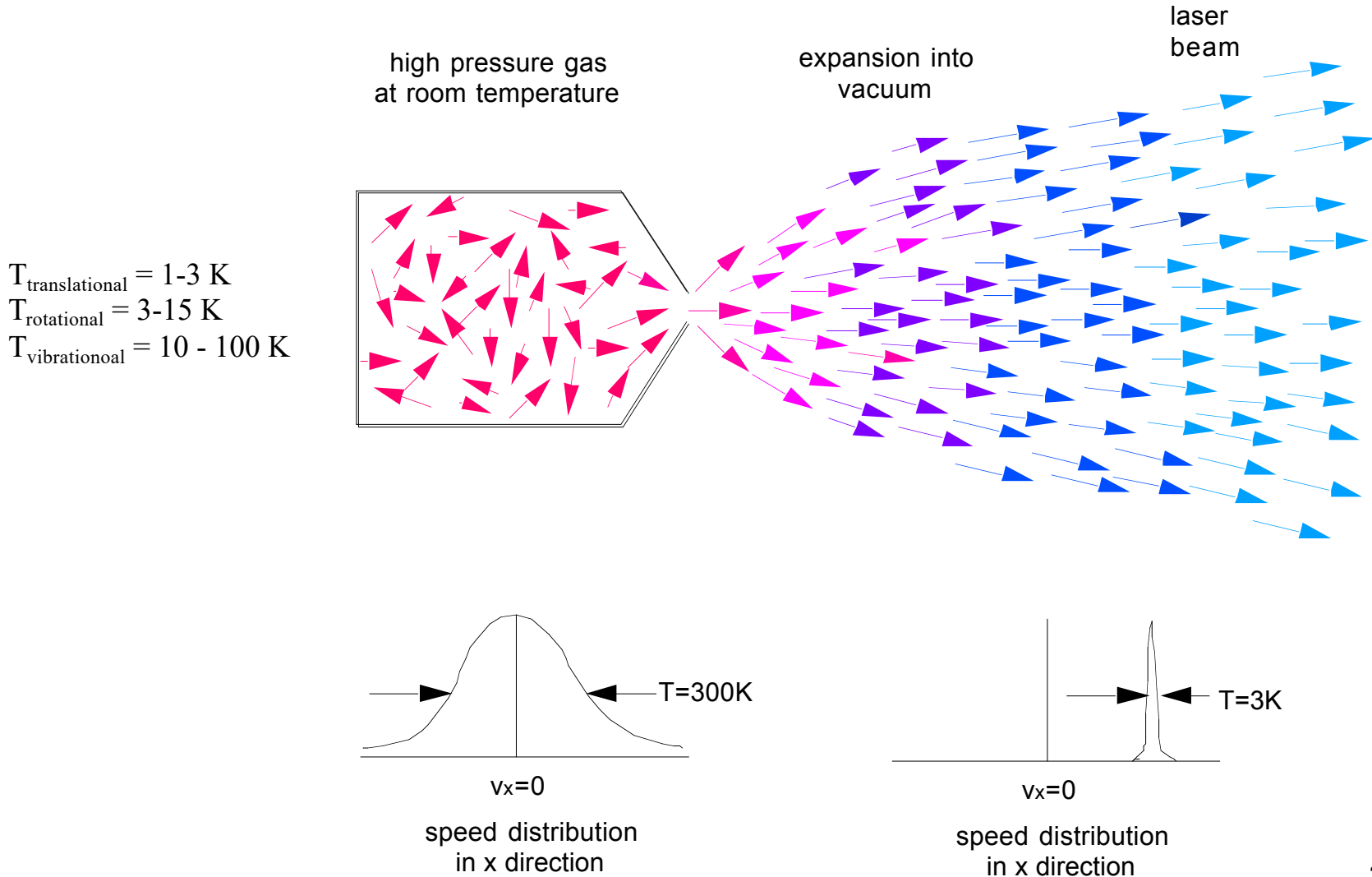
Static cell

T=300K



Spectral Simplification Techniques

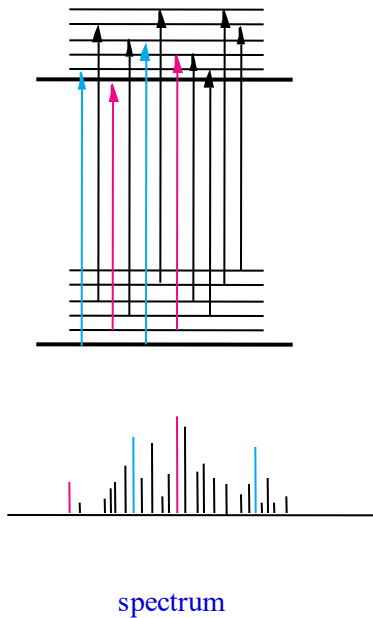
Supersonic cooling



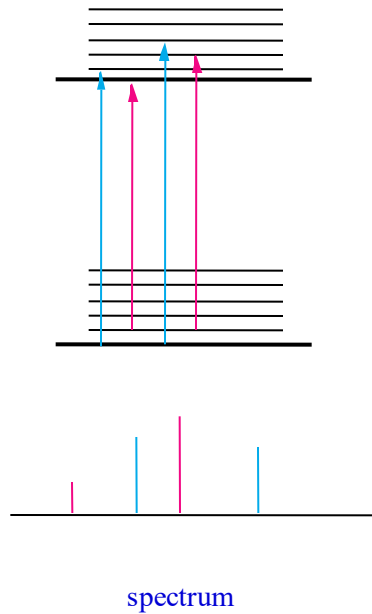
Supersonic cooling

Spectral Simplification via Supersonic Expansion

Room temperature experiment



In supersonic expansion



CH3OH

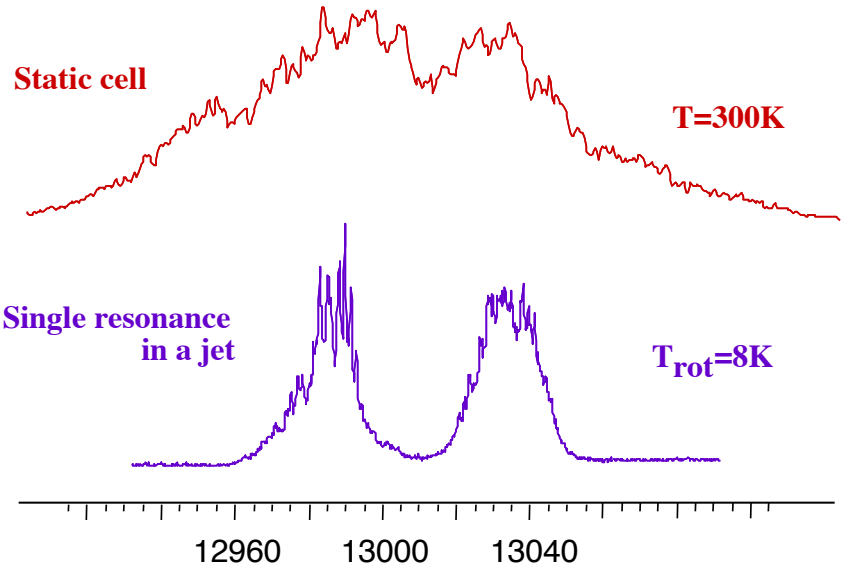
$5\nu_1$

Static cell

$T=300\text{K}$

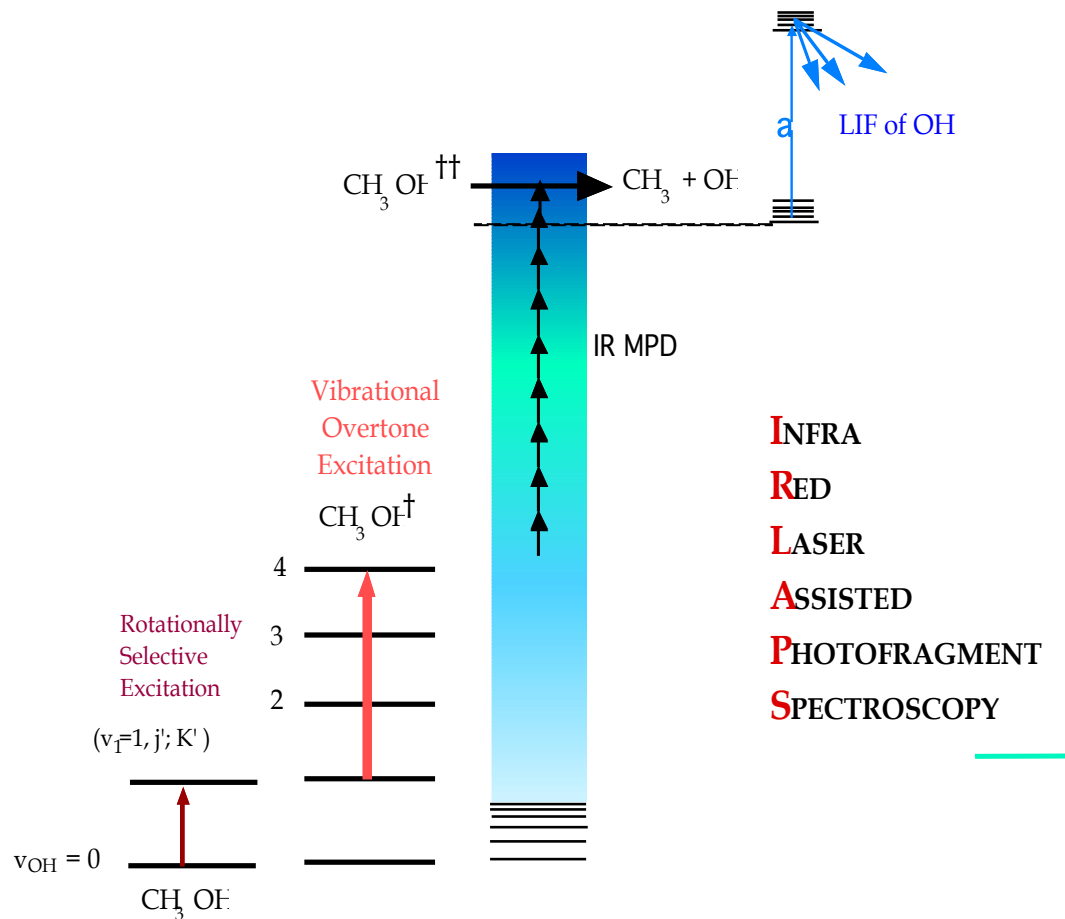
Single resonance
in a jet

$T_{\text{rot}}=8\text{K}$



Photofragmentation

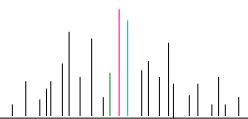
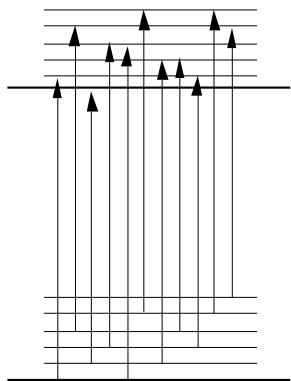
Energy diagram of double
resonance overtone IRLAPS:



Double resonance

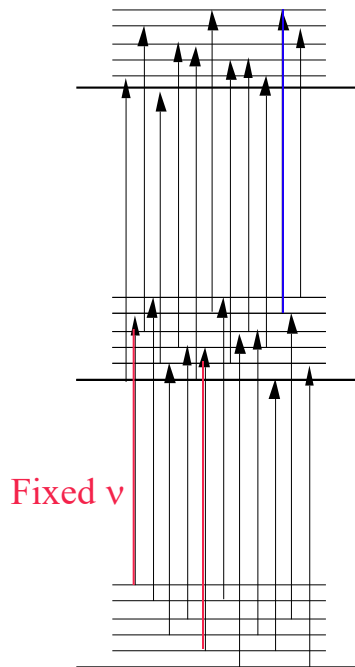
Spectral Simplification via Double Resonance Techniques

Single laser experiment

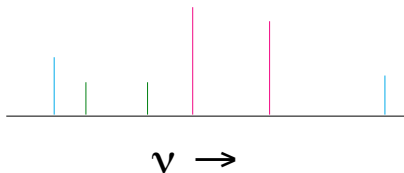


$\nu \rightarrow$
spectrum

Double resonance experiment

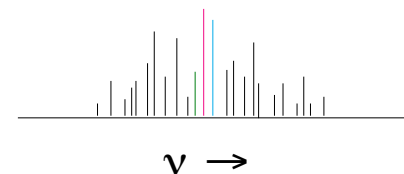


spectrum of
second step



$\nu \rightarrow$

spectrum of
first step



$\nu \rightarrow$

