Protein domains mediating interactions

- W_03 Protein Protein interactions domains Scaffolds and co-incidence PPI's and networks
- W_04 Protein Membrane
 Lipid-binding domains
 Manipulation of location
 Activation

"the right molecules at the right place at the right time"

Localising proteins to and within membranes

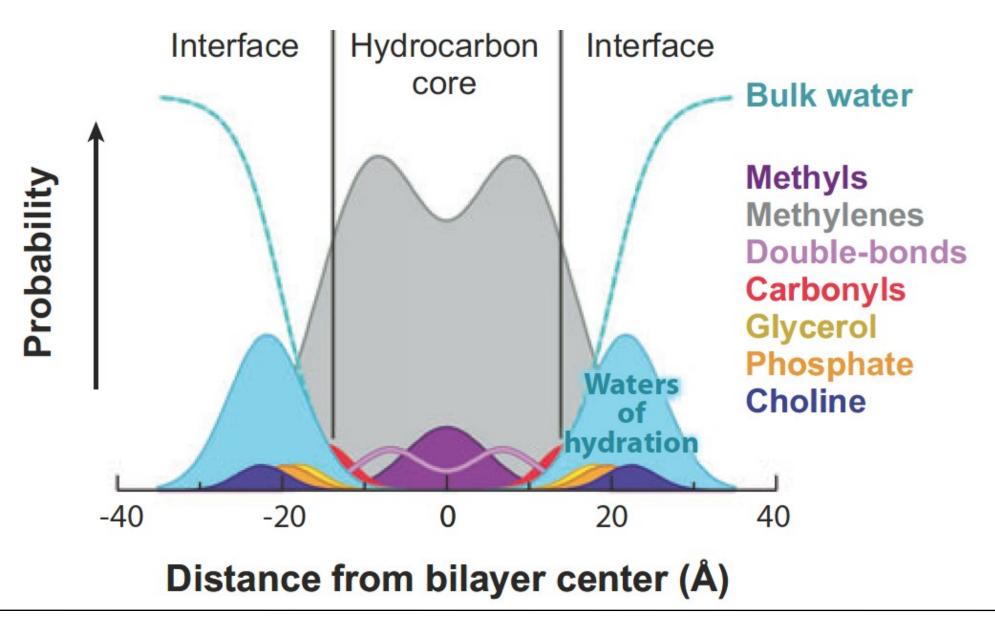
Part 1

Membrane binding domains

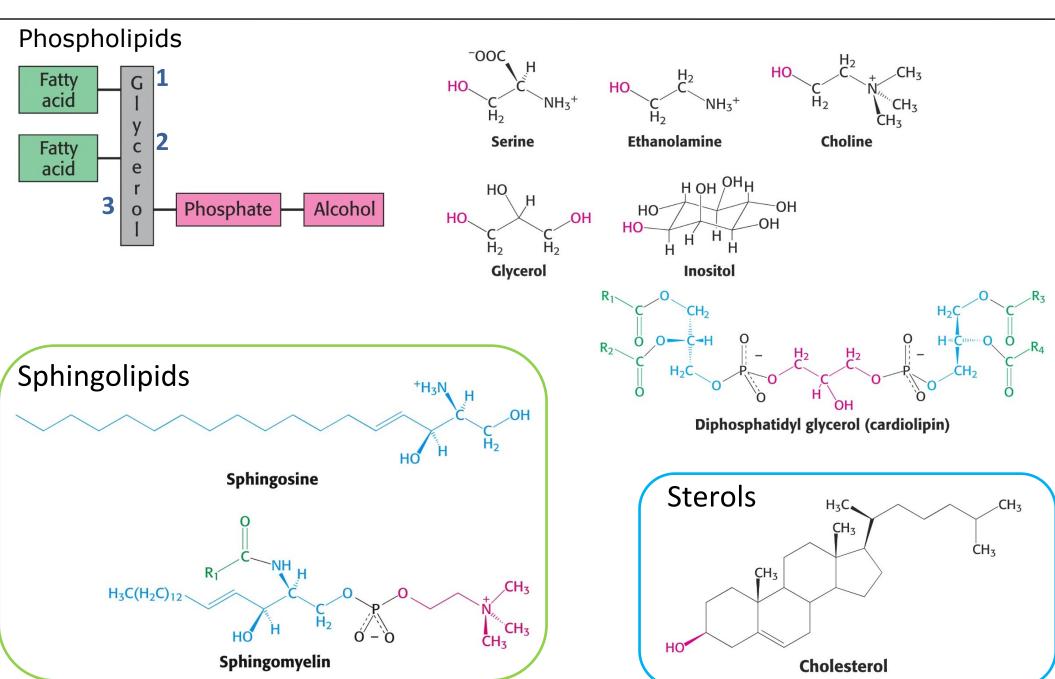
- PIP_x interacting domains
- PIP_x imaging
- Polybasic sequences

Membrane architecture

Depth probability profile of lipid functional groups



Membrane lipids

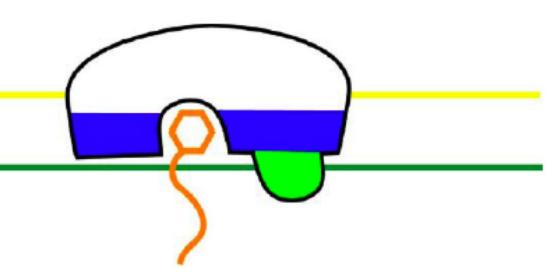


Membrane binding domains

Schematic representation:

Membrane-water interface

Start hydrophobic core



Three interactions are generally observed:

- Polar stereo-selective binding site for a lipid head group
- Hydrophobic loop inserting in membrane core (green)
- Electrostatic interaction with polar head groups (blue)

Hurley, BBA 2007

Membrane binding domains

In human genome many such domains have been identified:

in numan genome many such d	
Domain*	Humans
PH	303 (258)
PKC C2 [‡]	200 (125)
C1	79 (58)
PX	35 (35)
FYVE	27 (26)
Discoidin C2 ^{‡§}	24 (18)
GRAM	18 (15)
F-BAR	14 (14)
Annexin	56 (13)
Gla§	13 (13)
N-BAR	9 (9)
ENTH/ANTH	9 (9)

=> PIPx

=> Ca²⁺-dependent to phospholipids

=> diacylglycerol

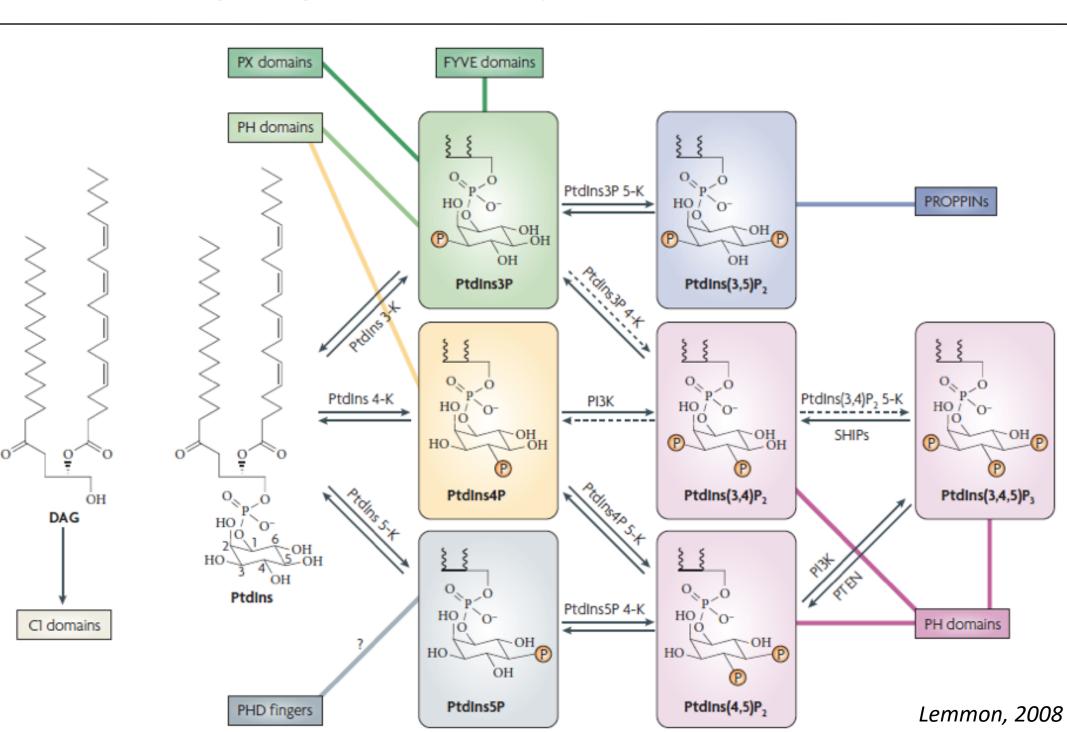
=> PI(3)P

=> PI(3)P

(#): number of proteins

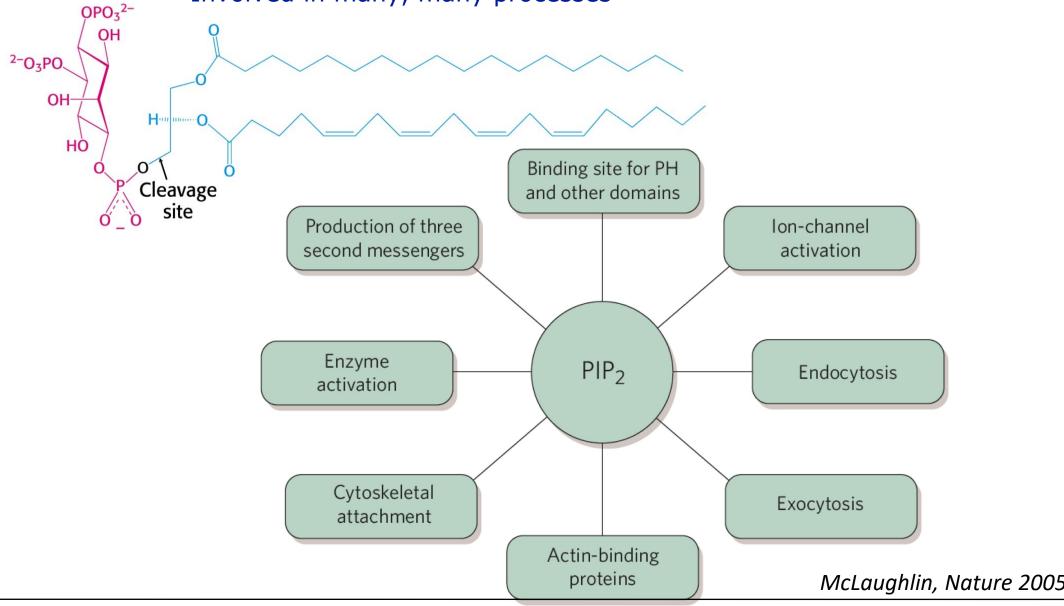
<u>Lemmon, 2008</u>

Domains recognising different PIP_x species



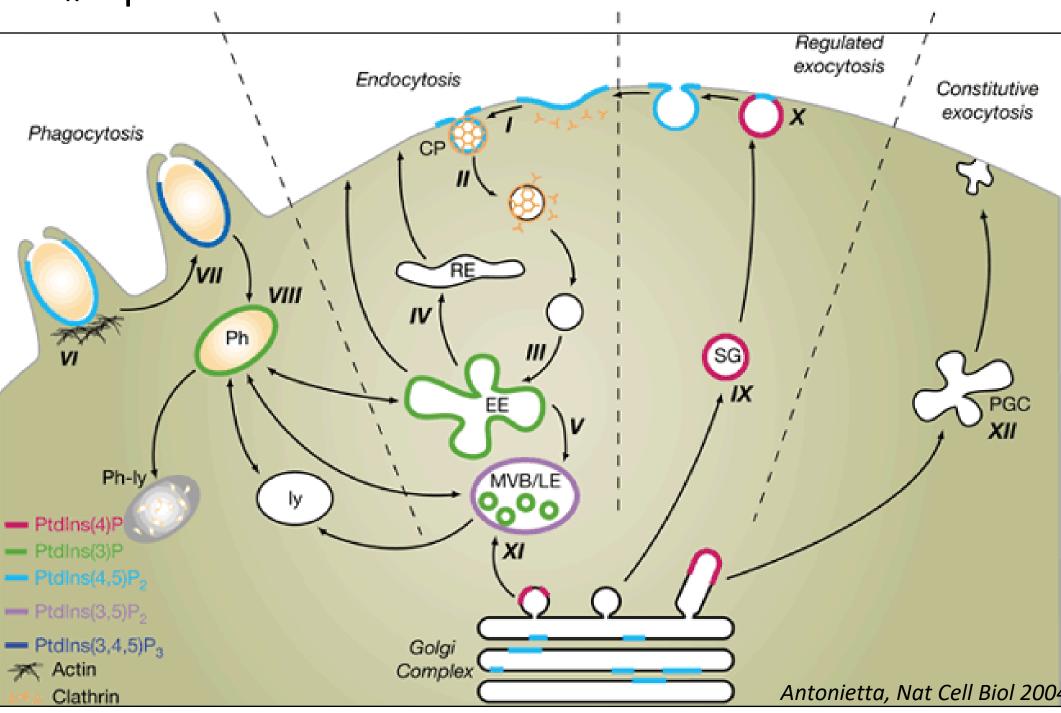
Phosphatidylinositol-(4,5)-bisphosphate (PIP₂)

- About 1% of PM lipids (about 1-10 μM); the most abundant of the PIPx
- Involved in many, many processes



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PIP_x: Specific locations



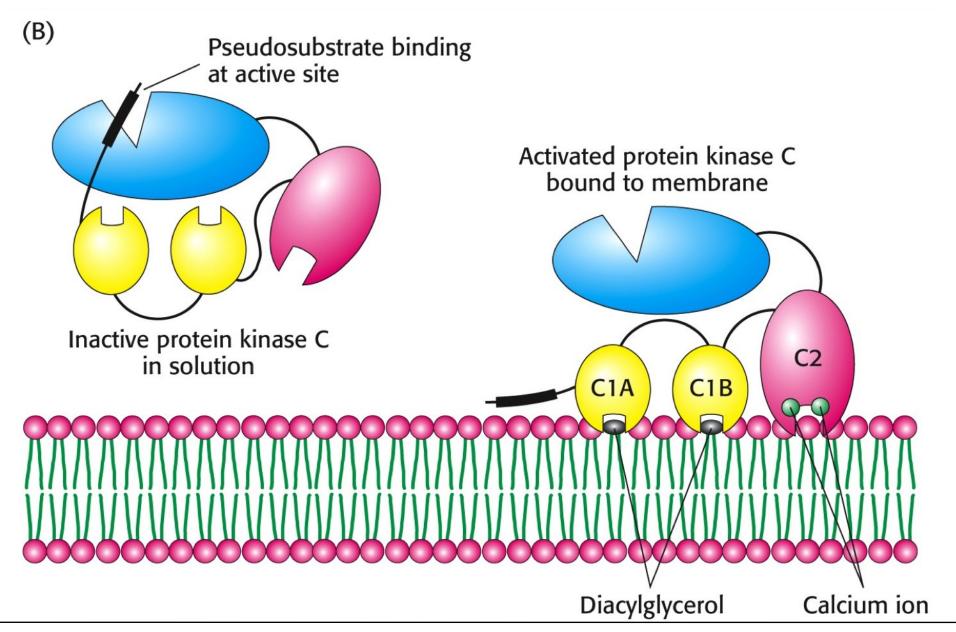
4 - Membrane binding domains

• PIPx

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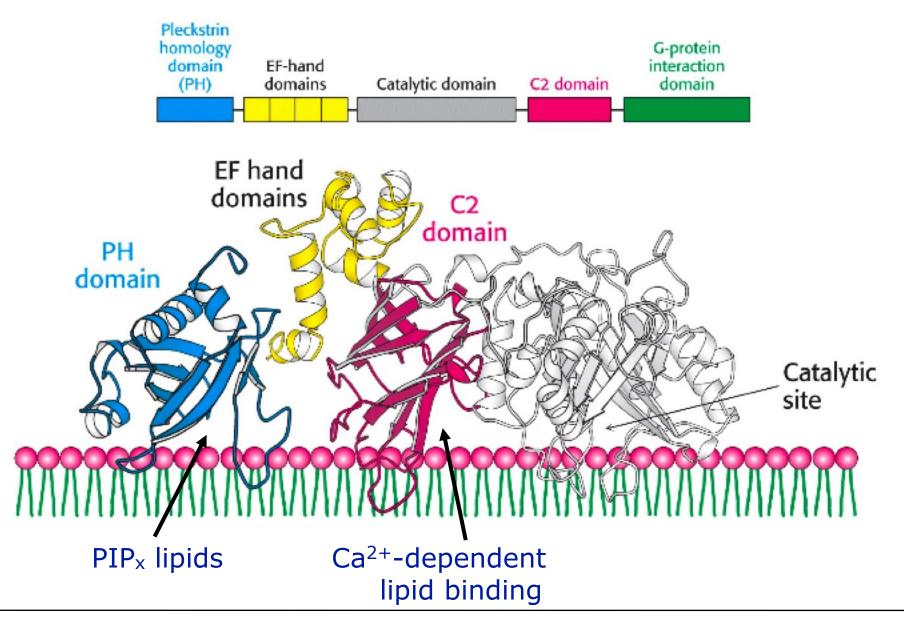
Reversible membrane location and activation

Protein kinase C combines the 2 mechanisms, both for <u>localisation</u> & <u>activation</u>:



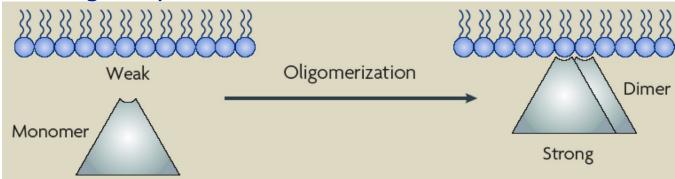
Reversible membrane location and activation

Phospholipase Cβ combines the 2 mechanisms, both for <u>localisation</u> & <u>activation</u>: but also for it's **in-activation**!!

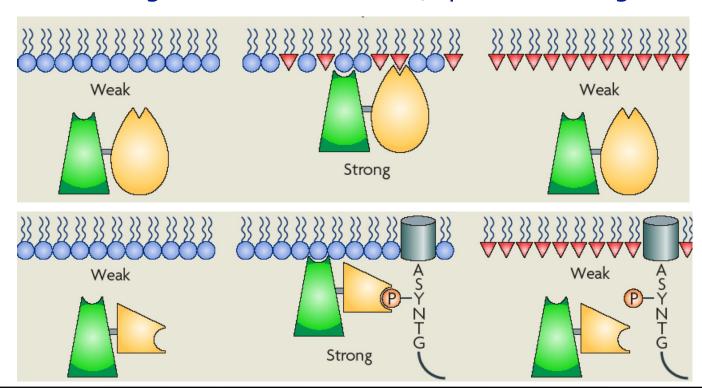


Membrane binding: Combination of domains

=> Stronger by more of the same



=> => Stronger & more selective/specific through **co-incidence**



Lemmon, 2008

4 - Protein domains

PIP_x undergo signal-dependent turnover; e.g. PI(4,5)P₂

GPCR signalling via $G_{\alpha q}$:

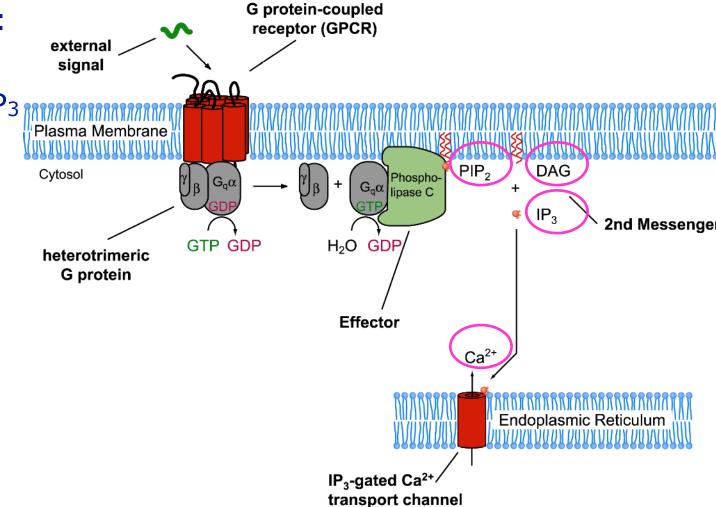
=> PLC activation

$$PI(4,5)P_2 => DAG + IP_3$$

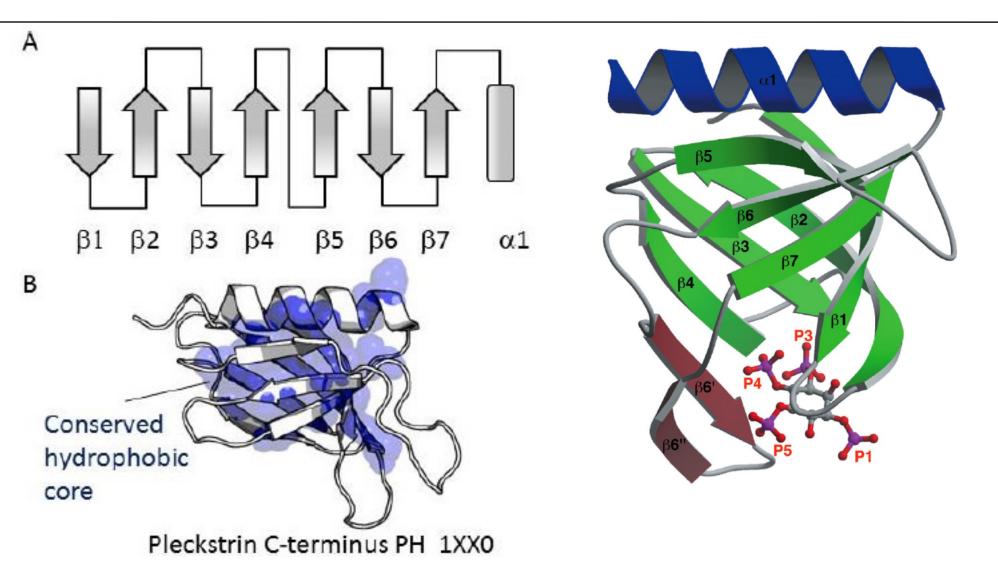
 $IP_3 = Inositol(1,4,5)$ triphosphate

DAG = Diacylglycerol

PLC = Phospholipase C



Pleckstrin Homology (PH) domains: Common Structure



Lenoir et al, Membranes 2015

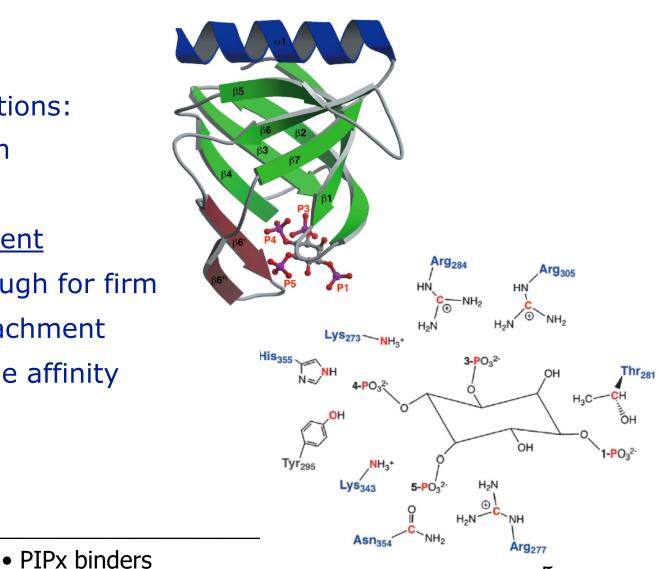
PH domains recognising PI(3,4,5)P₃

- PI(3,4,5)P₃ has a very low abundance in resting cells
 - upon activation 40-fold increase due to phosphorylation of PI(4,5)P₂ by PI-3-kinase

=> *see* : *1fgy.pdb*

Important regions for interactions:

- β 1-β2 loop is a basic region
- 17-19 H-bonds
- Large hydrophobic loop <u>absent</u>
- K_d = 27 nM => strong enough for firm membrane attachment
- I(1,3,4,5)P₄ binds with same affinity

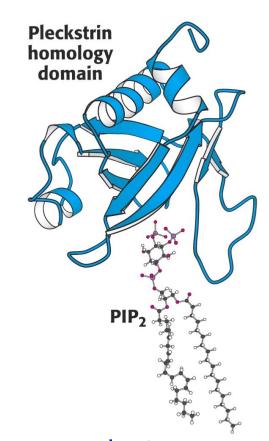


Domains recognising PI(4,5)P₂

- PI(4,5)P₂
 - most abundant PIP_x
 about 1% of plasma membrane PL's
 to 100-fold more abundant than other PIP_x's
 - most domains bind with low affinity
 - => see : 1mai.pdb

An exception:

 PH domain of phospholipase-Cδ1 binds with high affinity, however: affinity for I(1,4,5)P₃ is 10-fold higher

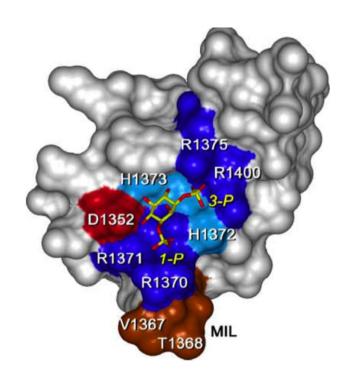


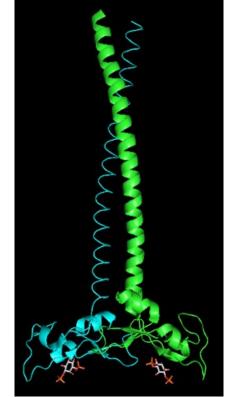
=> The enzyme is removed from the membrane by it's own product.

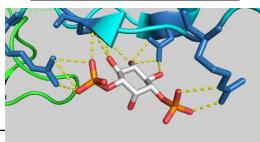
FYVE domains recognising PI(3)P₁

- PI(3)P₁ present in endosomes, 0.25 % of cellular lipid, or 200 μM
 - important in regulating trafficking in endosomal pathway and Golgi-lysosome sorting
- ♦ FYVE domains: Important for interaction with PI(3)P:
- basic (R/K)(R/K)HHCR in 1st β -strand $\}$ => selectivity
- 12 H-bonds with P_i and OH groups
- Monomer too weak for membrane attachment

=> dimers do !!







4 - Membrane binding domains

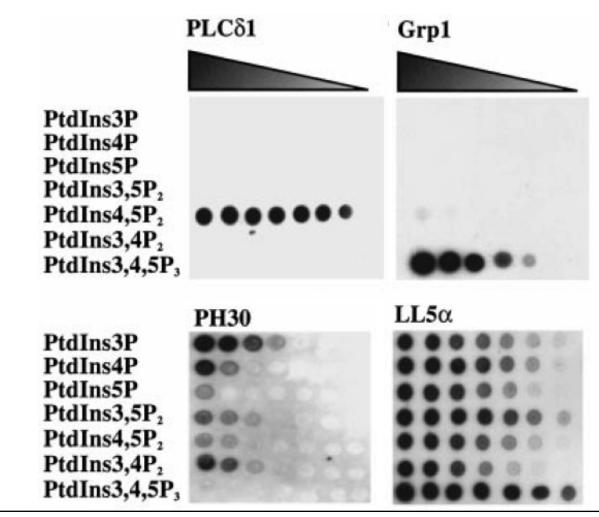
PIPx binders

Testing specificity and affinity of PIP_x domains

"Dot blot assay"

Dowler, Biochem J 2000

- Spot the different lipids on nitrocellulose
- Overlay with PH-domain-GST fusion proteins as dilution series
- Detect bound fusion with an anti-GST antibody



GST = Glutathion-S-Transferase an affinity tag for purification

4 - Membrane binding domains

PIPx - Omics

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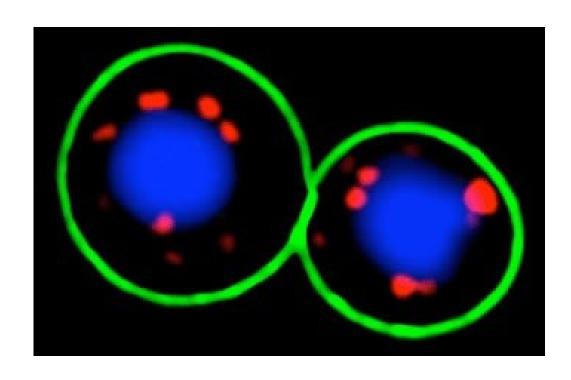
Imaging PIP_x in vivo using domain-GFP chimeras

In yeast cells:

Green: PH-domain - GFP fusion $=> PI(4,5)P_2$ in plasma membrane

Red: FYVE-domain - DsRed fusion => PI(3)P in endosomes

Blue: Vacuoles



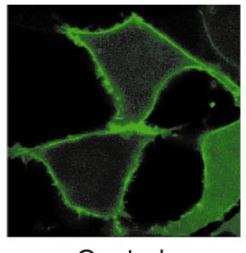
Imaging PIP_x metabolism in vivo using PH-GFP chimeras

In mammalian cells:

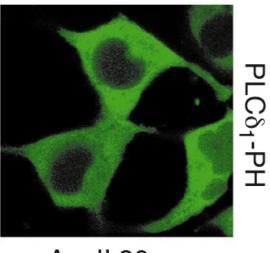
- If target lipid is present => PH-GFP probe on membrane

• If not:

=> probe in cytosol



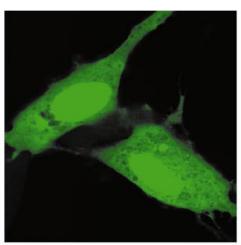
Control



Angll 30 s

A: PI(4,5)P₂-specific probe Control: PI(4,5)P₂ in plasma membrane

+ Angiotensin II (AngII) promotes breakdown via PLase-C



Control

PDGF 4 min

B: PI(3,4,5)P₃-specific probe Control: little PI(3,4,5)P₃ in plasma membrane

+ PDGF stimulate synthesis by activation of PI-3-Kinase

Balla, TiPS 2000

4 - Membrane binding domains

PIPx - Omics

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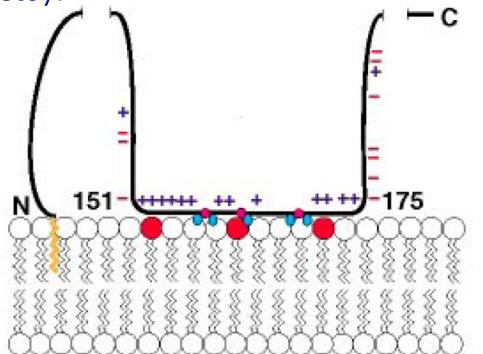
Poly-basic sequence: Membranes, PIP₂ and Ca²⁺²

Poly-basic sequence: A short stretch of amino acids with many K and/or R

- Bind strongly to negatively charged membranes
- Sequester PIP₂

E.g. MARCKS

- " Myristoylated Alanine-Rich C-Kinase Substrate"
- Present in the cell at µM concentrations
- Polybasic domain ...KKKKKRFSFKKSFKLSGFSFKKNKK
- N-terminal myristoyl



red dot = PIP₂

McLaughlin, Nature 2005

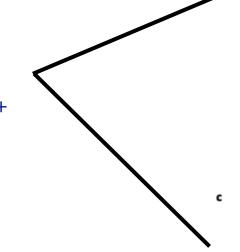
Poly-basic sequence: Release from membrane

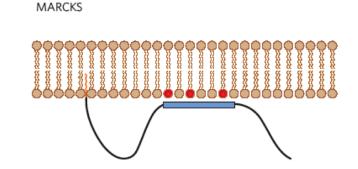
Reduction of MARCKS binding to membranes:

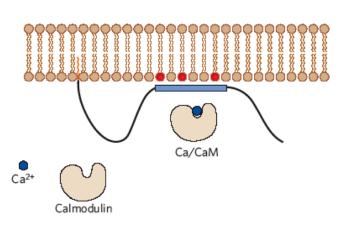
- i) Hydrolysis of PIP₂
- ii) Phosphorylation of serine **S** residues by protein kinase C (PK-C)

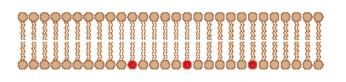
KKKKKRFSFKKSFKLSGFSFKKNKK

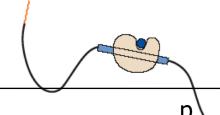
iii) Binding of calmodulin-Ca²⁺











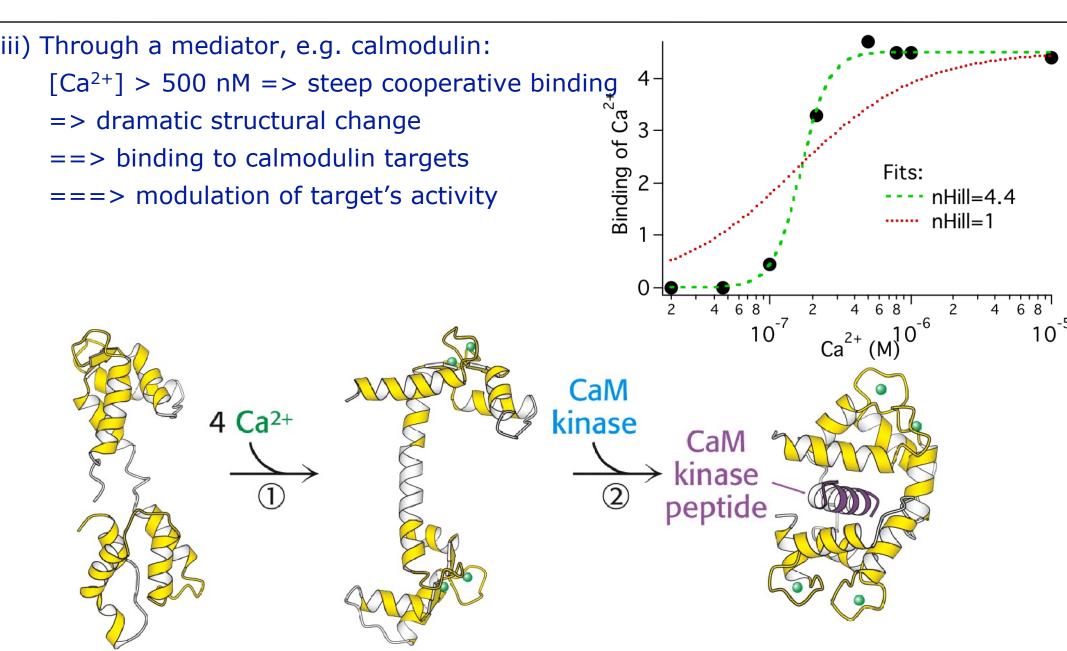
McLaughlin, Nature 2005.

5 - Membrane binding domains

Polybasic

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Calcium, a ubiquitous messenger



Calmodulin (apo)

4 - Membrane binding domains

Polybasic

Poly-basic sequence: Release from membrane

Regulation of MARCKS binding to membranes:

- Hydrolysis of PIP₂
- Phosphorylation of Ser
- Binding of calmodulin-Ca²⁺

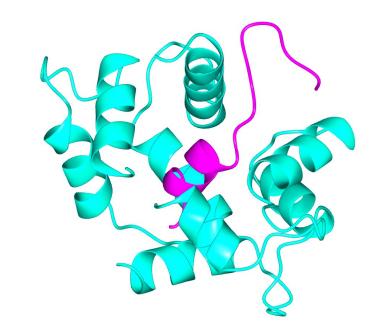
Crystal structure of complex o calmodulin-MARCKS cartoon:

cyan: calmodulin

- purple: MARCKS

Iso-potential surface:

- red -25 mV
- blue + 25 mV
- => see 1iwq.pdb



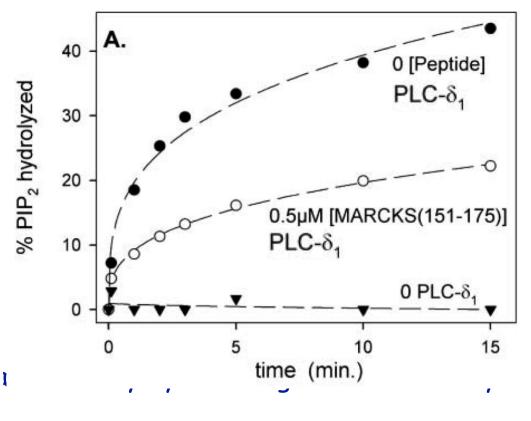
McLaughlin, Nature 2005.

19.9 Å

Poly-basic sequence - Effect on other PIP₂ binding proteins

 Inhibition of PIP₂-hydrolysis by MARCKS

Lipid vesicles containing 0.15 % [3 H]-PIP₂ Hydrolysis by phospholipase $C_{\delta 1}$

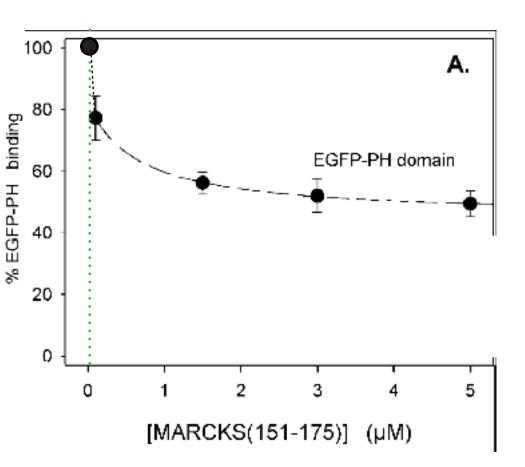


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Poly-basic sequence - Effect on other PIP₂ binding proteins

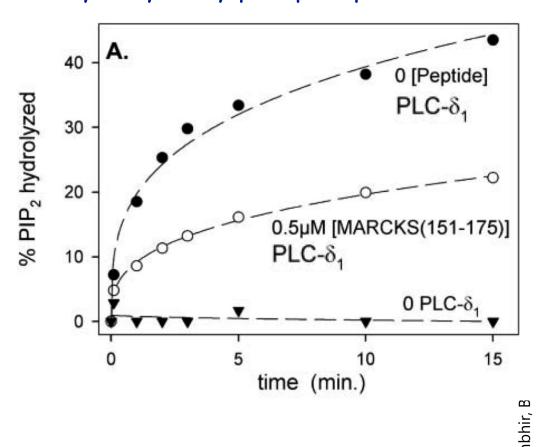
 Inhibition of PH-domain binding to PIP₂ by MARCKS

Lipid vesicles containing 1% PIP₂ 10 nM PIP₂ binders



 Inhibition of PIP₂-hydrolysis by MARCKS

Lipid vesicles containing 0.15 % [${}^{3}H$]-PIP₂ Hydrolysis by phospholipase $C_{\delta 1}$



=> MARCKS decreasing PLC binding and activity by reducing PIP2-availability

Localising proteins to and within membranes

Cytosolic proteins can bind in a spatio-temporally regulated manner to the membrane surface through

- lipid binding domains
- poly-basic sequences

Reversible membrane attachment depends

- on **type** and **number** of interactions
- cofactors like Ca²⁺
- composition and concentration of lipids recognized
- co-incidence of different conditions and
- co-operation between domains
- ÷ This is regulated by signalling processes

Further reading:

Balla: "How accurately can we image inositol lipids in living cells?"

TiPS 2000

McLaughlin: "Plasma membrane phosphoinositide organization by protein electrostatics" Nature 2005

Lemmon: "Membrane recognition by phospholipid-binding domains"

Nature Reviews Molecular Cell Biology, 2008

Next week:

Part 2: Lipidation & transmembrane domains