

Biochemistry of Membranes

Biomembranes: Introduction.

Phospholipids: Biosynthesis, distribution and dynamics.

Membrane protein biogenesis.

Vesicular transport: From fission to fusion.

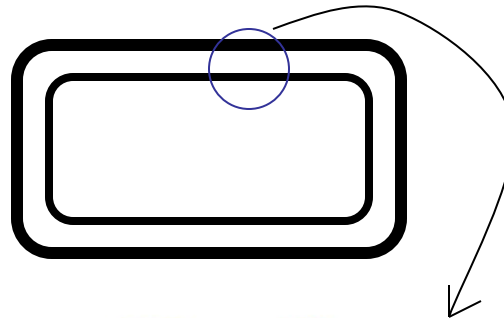
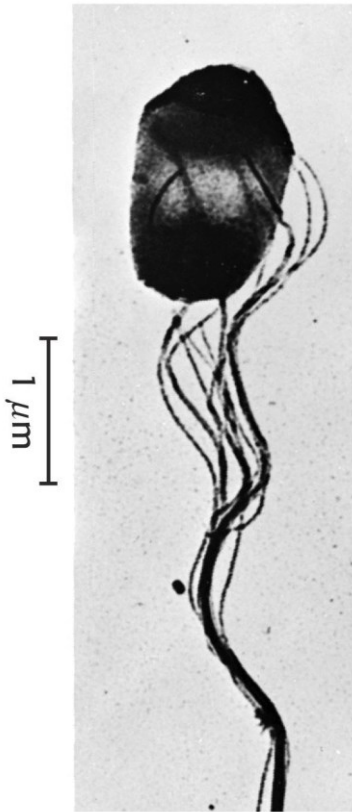
Lateral organization: Dynamic properties of domains.

How does a cell look like?

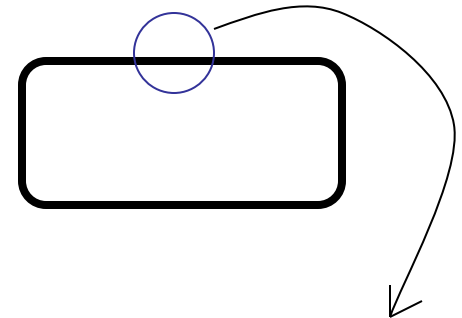
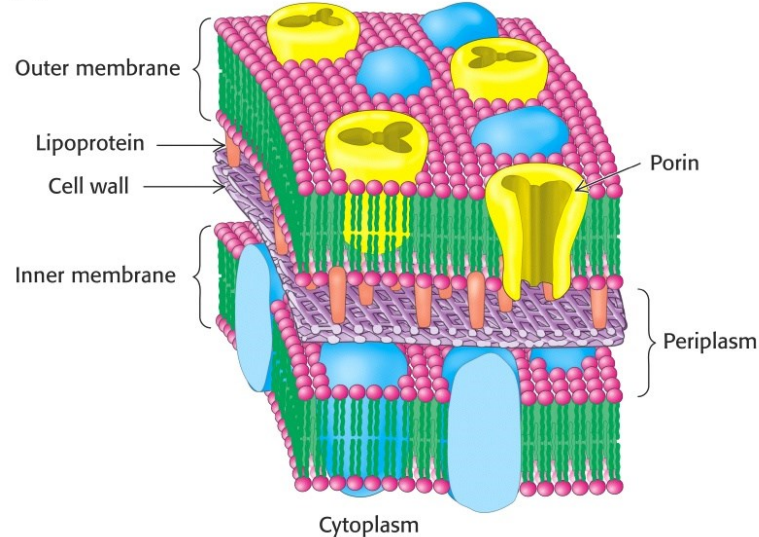
The "simplest" cell: a prokaryote

Gram negative

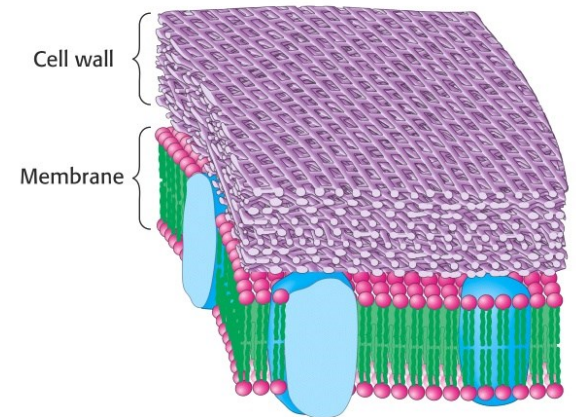
Gram positive



(A)



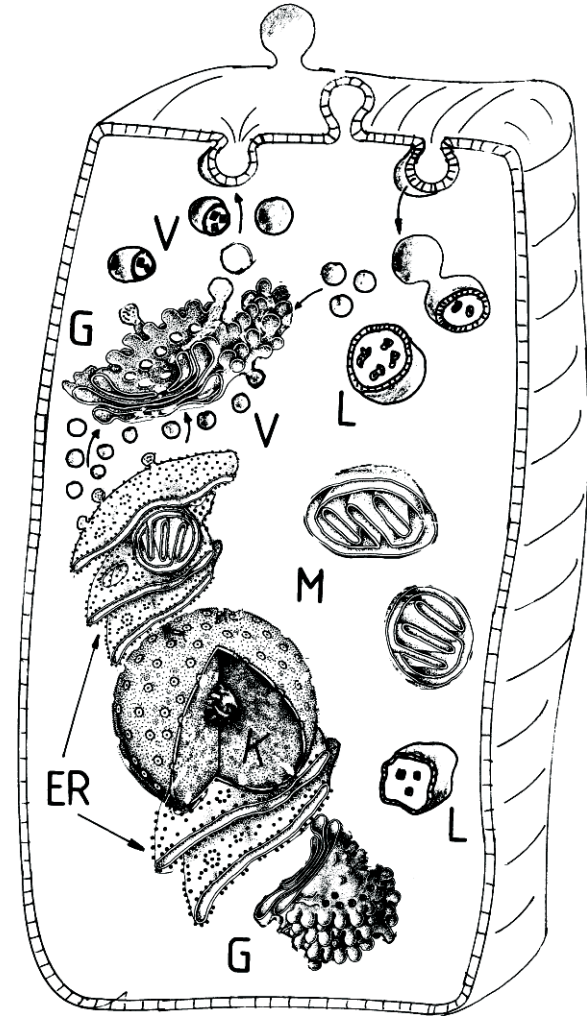
(B)



How does a cell look like?

A more complex cell: a eukaryote

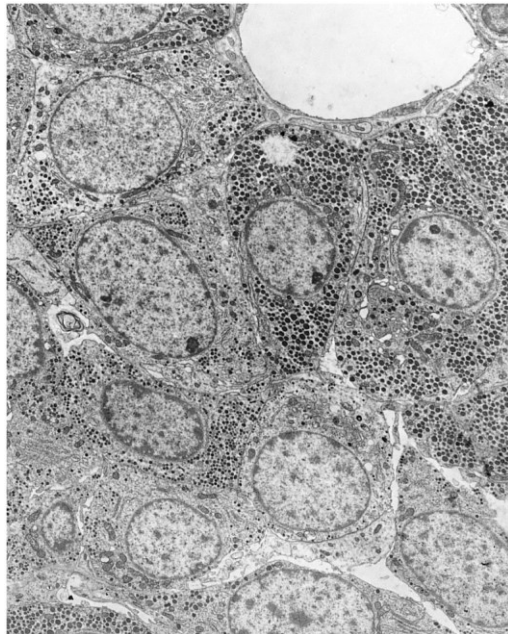
- many internal membranes
- lot of flow
- lot of different morphologies



How does a cell look like?

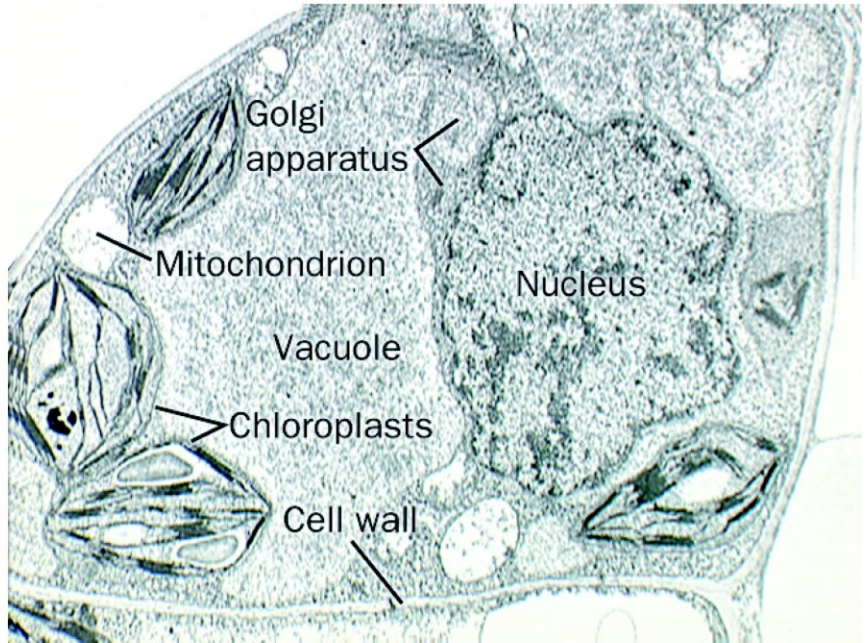
A more complex cell: a eukaryote

- rat pituitary cell



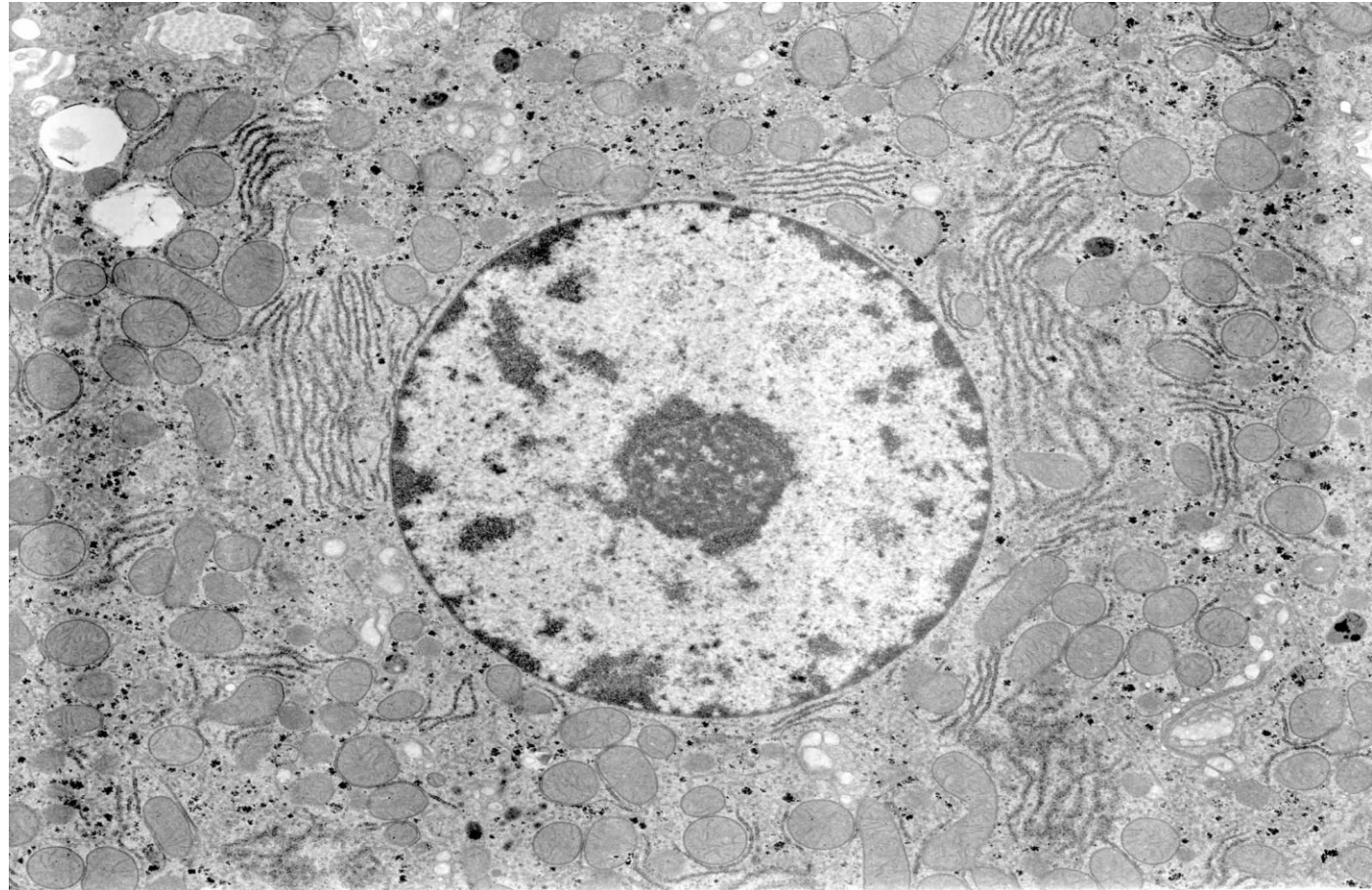
crowded!!

- plant cell



Looking at a cell

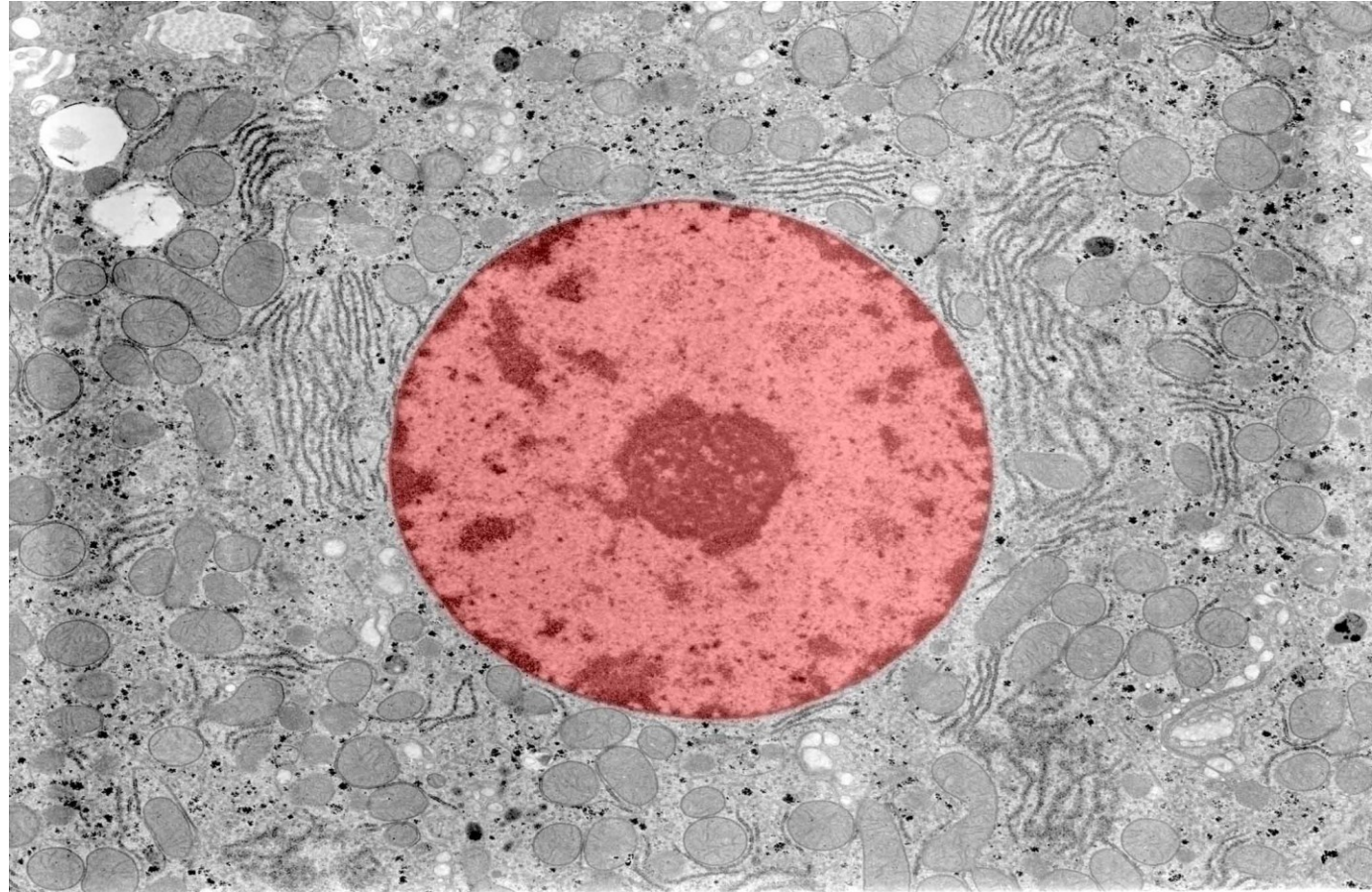
Mammalian cell
viewed by electron
microscopy



Looking at a cell

Nucleus:

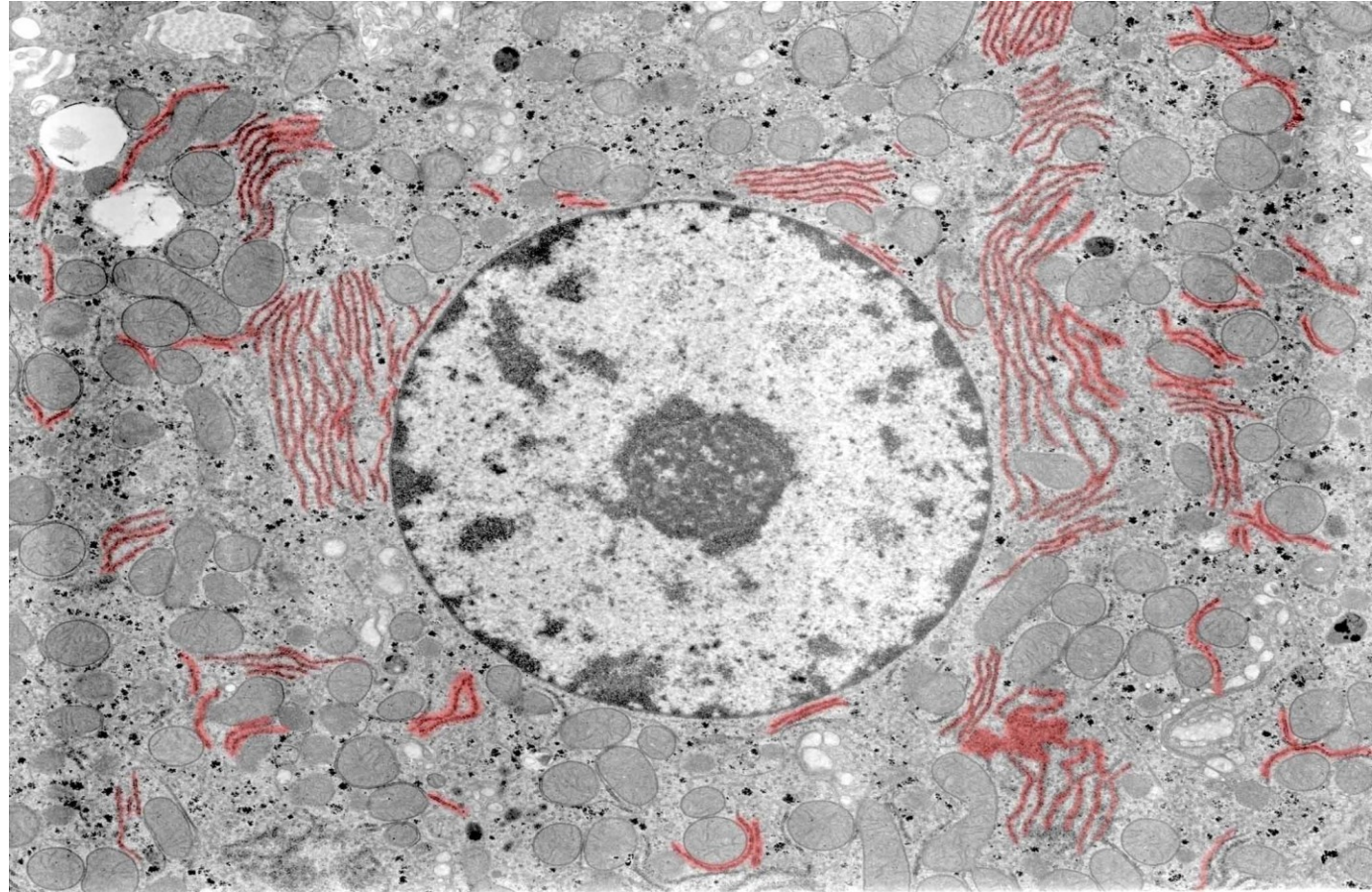
- DNA
- Transcription
- Export of RNA



Looking at a cell

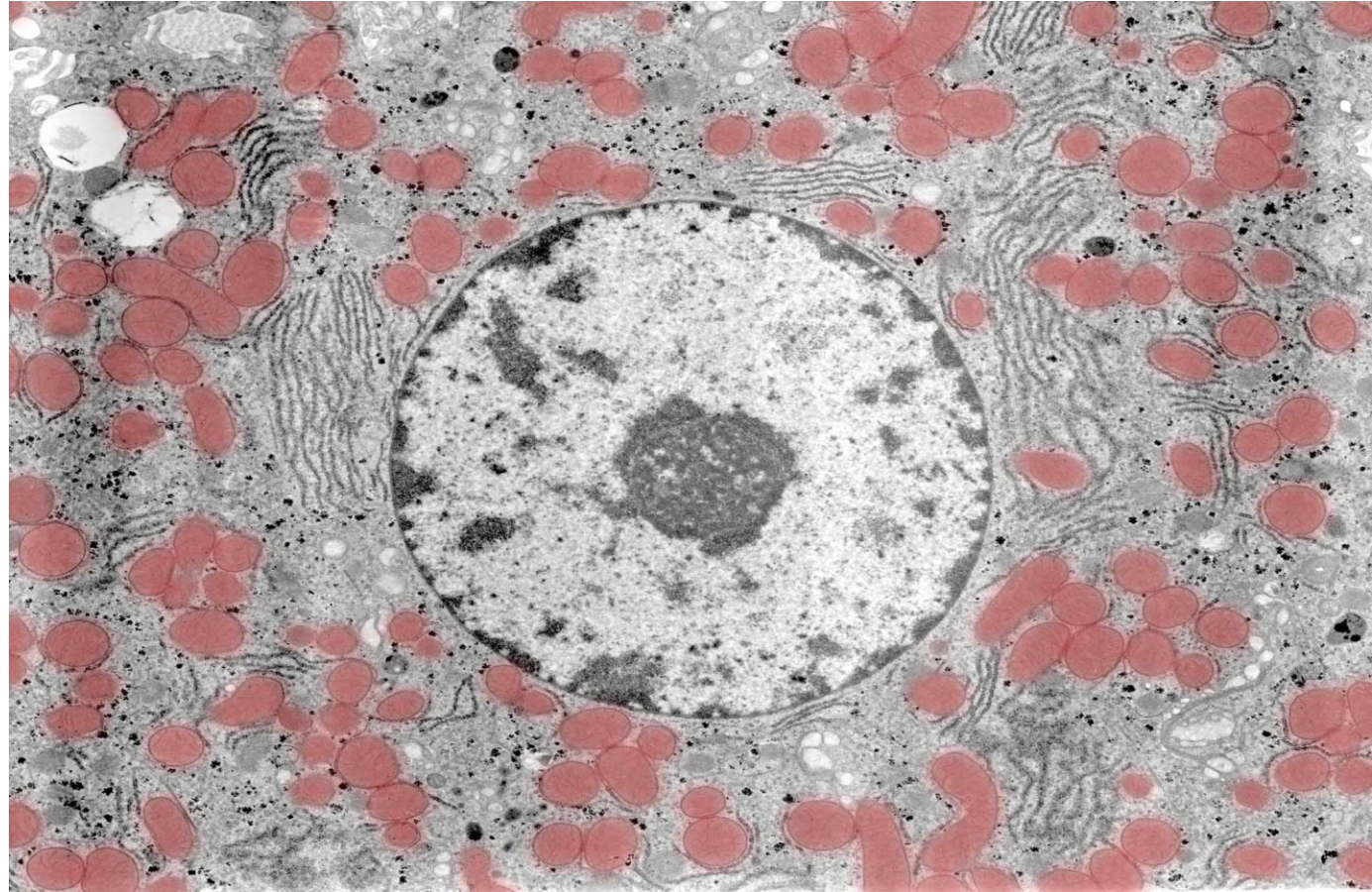
Endoplasmic reticulum (ER):

- Protein synthesis
- Sterol synthesis
- Sorting
- Detoxification



Looking at a cell

Mitochondria:
- Oxidative
phosphorylation



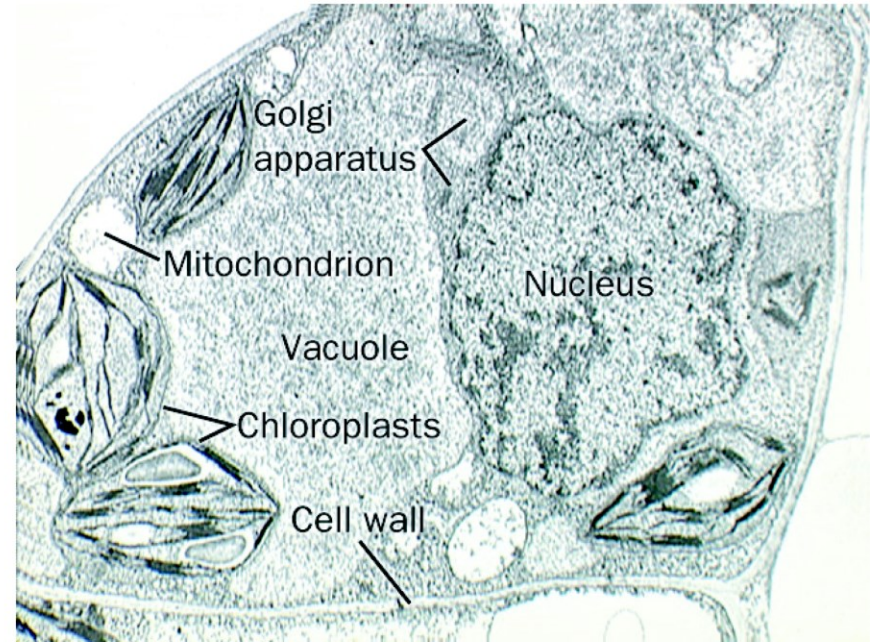
Functions of membranes

- Barrier

“Membranes separate live from dead”

Confinement

- Communication
- Scaffold
- Reservoir
- Transport



Properties of membranes

Self-assembling

Self-healing

Flexible

Shape

Dynamic

Hydrophobic layer

Specificity

QuickTime™ and a Sorenson Video 3 decompressor are needed to see this picture.

Membranes

Wide variety of functions and properties

<=>

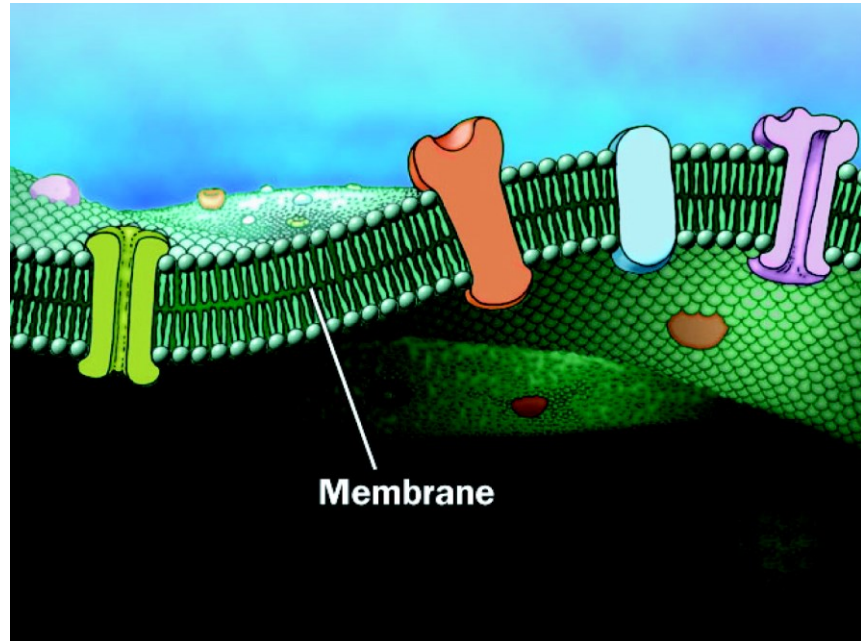
Wide variety of compositions?

Membrane components

Initial ideas:

Lipids => Barrier

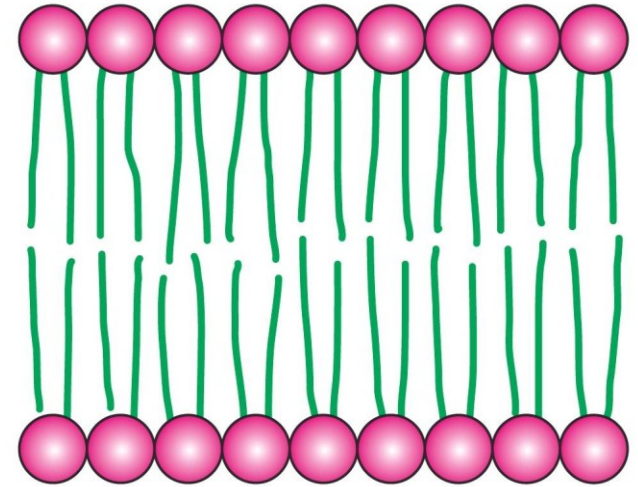
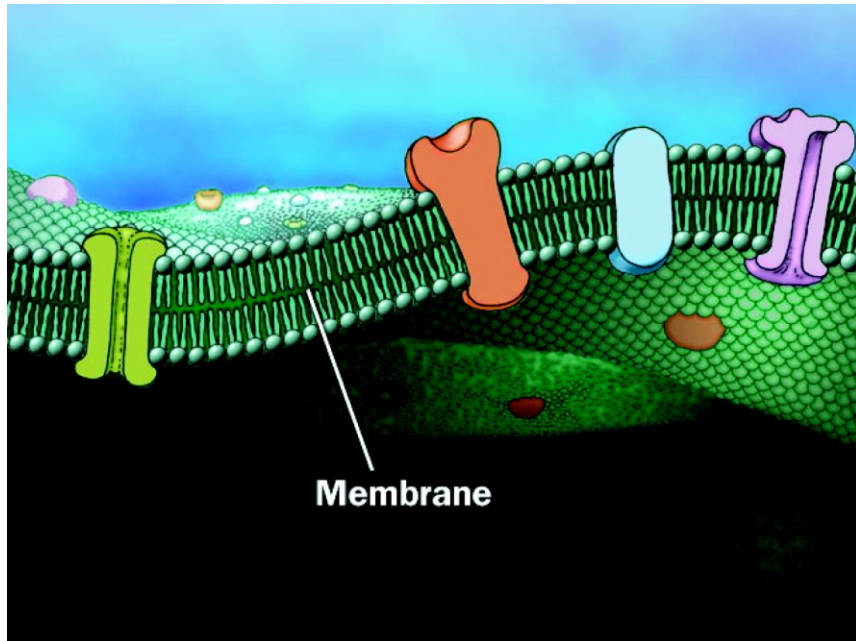
Proteins => Function



Architecture of membranes

1925

Lipidic bilayer (Gorter & Grendel)



1972

Fluid mosaic model

(Singer & Nicolson):

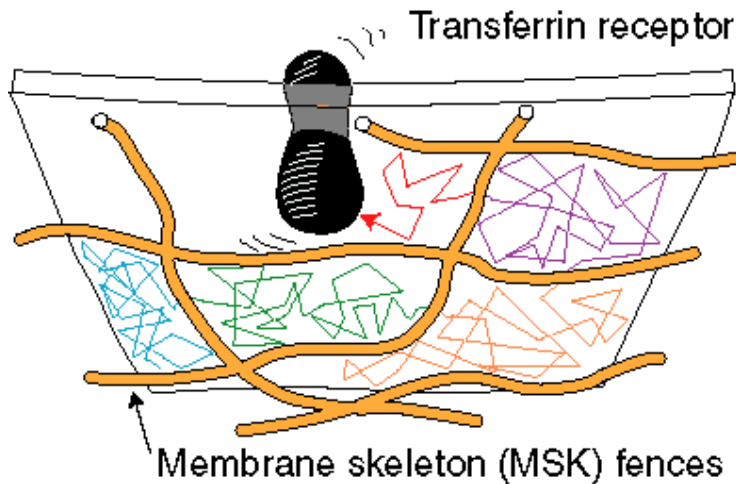
Free diffusion of all components within the membrane.

Architecture of membranes

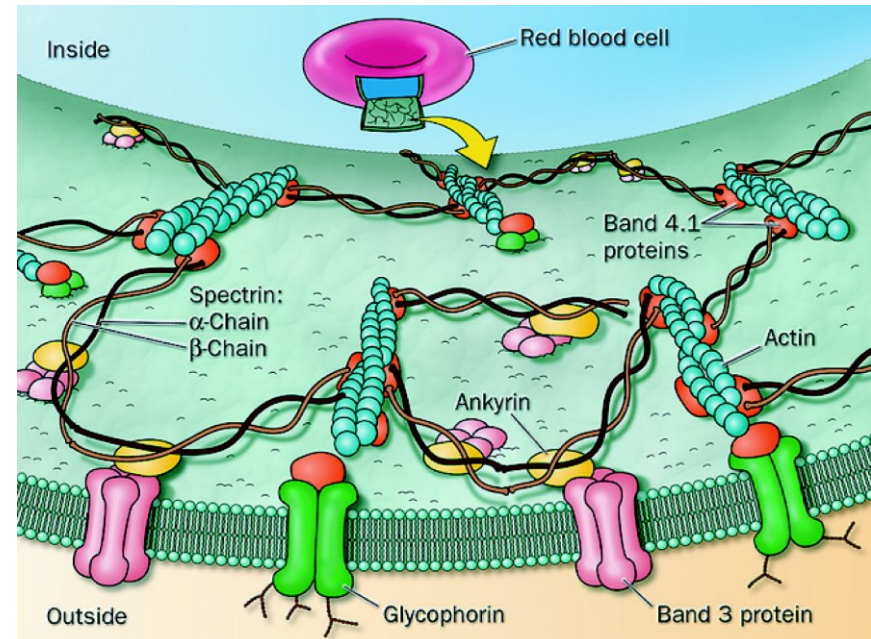
A skeleton of membrane-attached proteins:

- mechanical stability
- scaffold for molecules
- diffusion barrier

Membrane Skeleton Fence



Kusumi (Nagoya, Japan)



Architecture of membranes

All is moving!

QuickTime™ and a YUV420 codec decompressor are needed to see this picture.

Membranes

Some numbers:

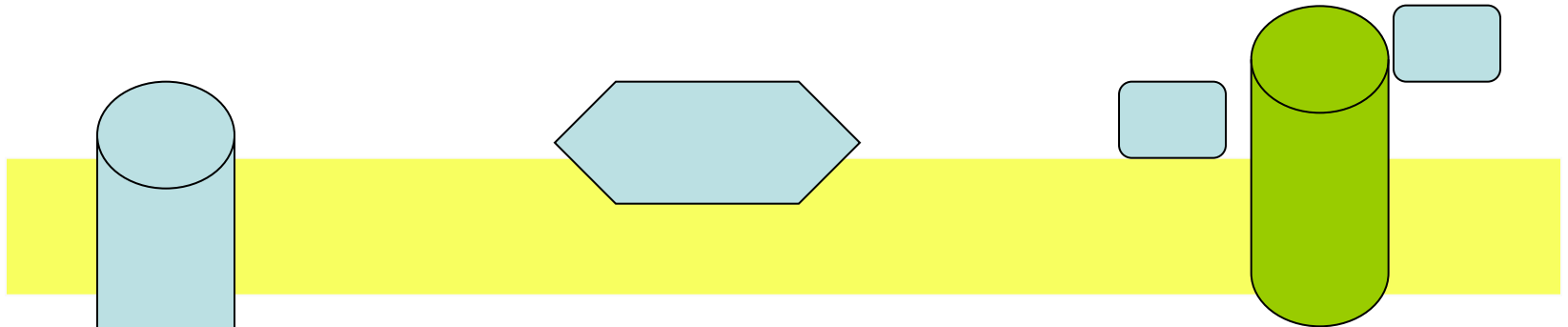
- ◇ Dimensions
 - cell 1 to 10's μm
 - membrane $\pm 4 \text{ nm thick}$
 - lipid $\pm 50 \text{ \AA}^2$
- ◇ Diffusion
 - lipid in membrane $10^{-10} \text{ to } 10^{-8} \text{ cm}^2/\text{s}$
- ◇ Potential
 - $-80 \text{ mV} \Rightarrow 20 \text{ MV/m}$

Membrane proteins

Integral

Peripheral

Extrinsic



Detergents

Weak detergents
Chaotropic agents

High Salt
Chelators

Extraction:

Hydrophobic

Hydrophobic &
Electrostatic

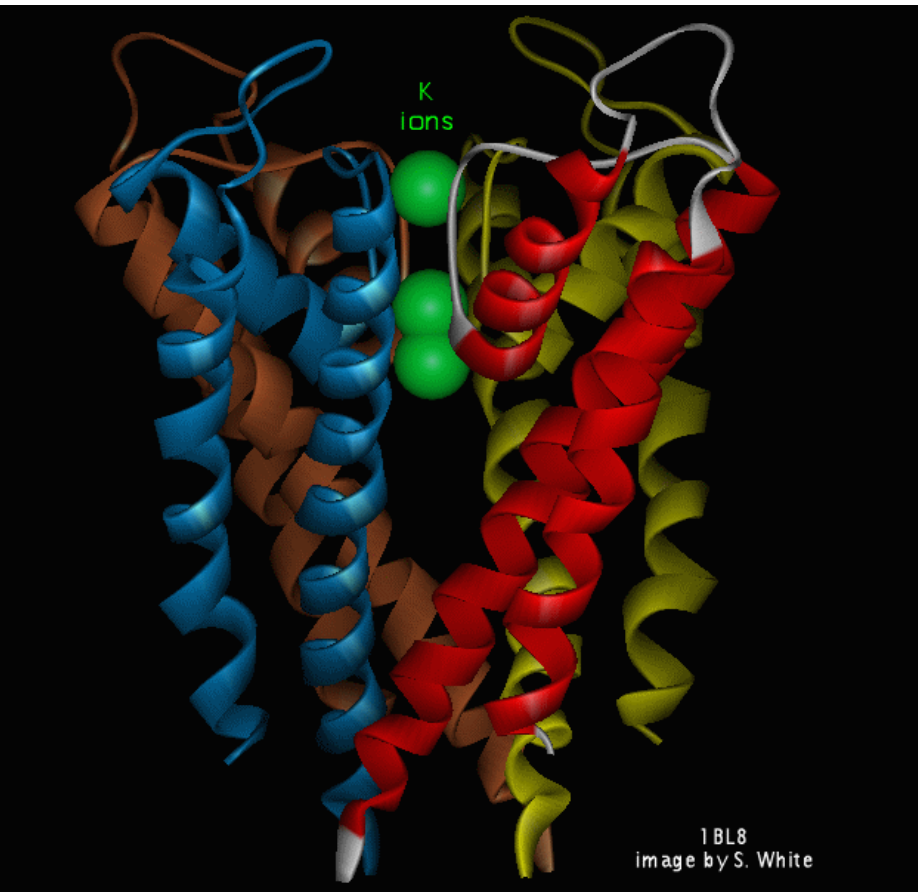
Electrostatic

Interaction:

Integral membrane proteins

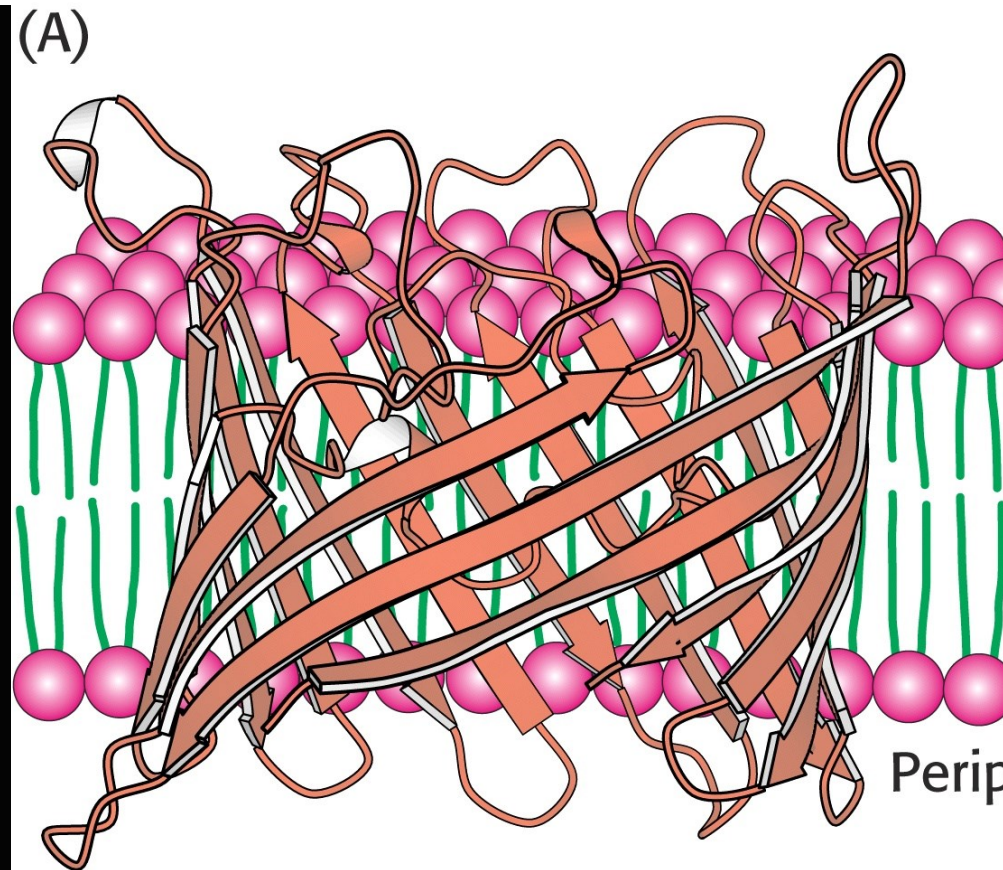
α -Helical bundle

Voltage-gated K⁺-channel



β -Barrel

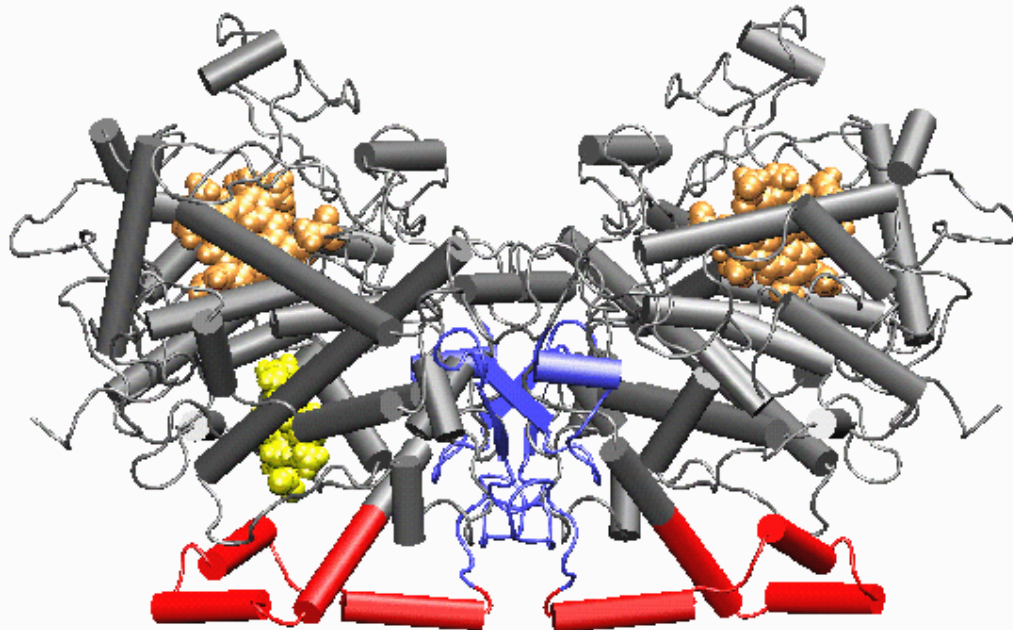
Porin OmpF



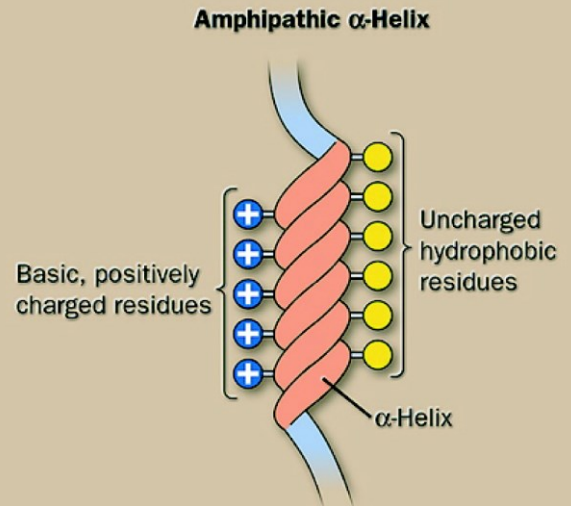
Peripheral membrane proteins

Prostaglandin H2 synthase

Amphipathic α -helix



Theoretical Biophysics Group, University of Illinois at Urbana-Champaign



Membrane proteins

◇ Highly ordered structural motifs:

- α -helix

hydrophobic stretch of 15-25 amino acids.

- β -sheet

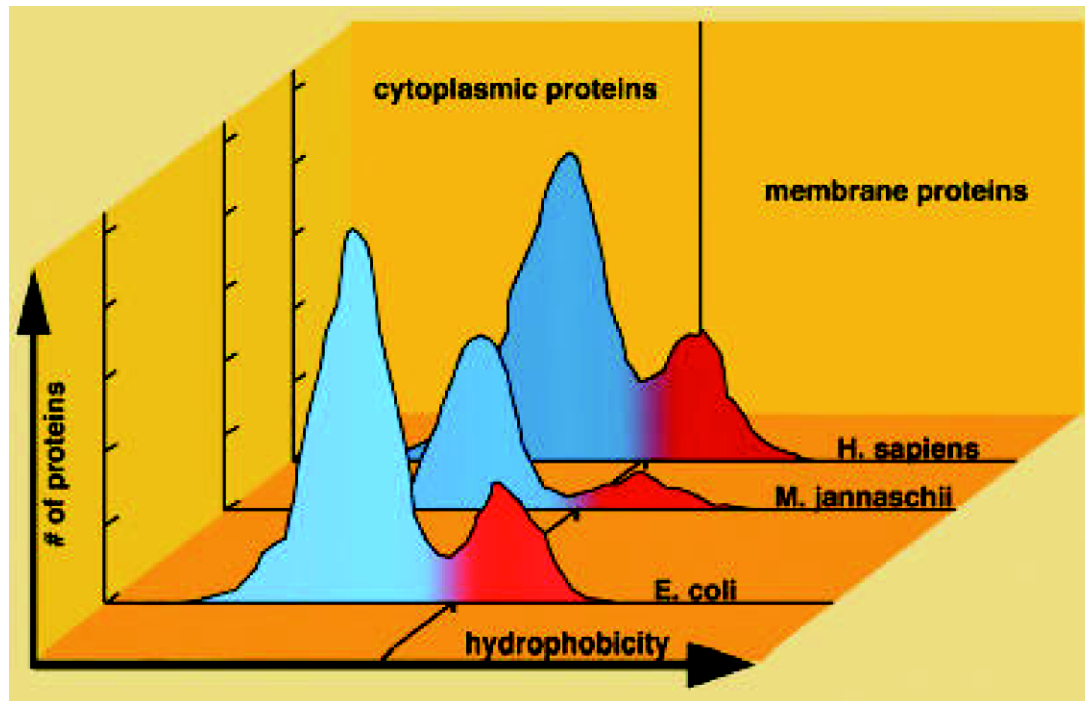
alternating hydrophilic and hydrophobic amino acids.

◇ Few charged amino acids.

◇ Proteins move to function.

Membrane proteins

Genome-wide analysis of mean hydrophobicity of proteins:



=> 30 % of all proteins are membrane proteins, i.e. about 10'000.

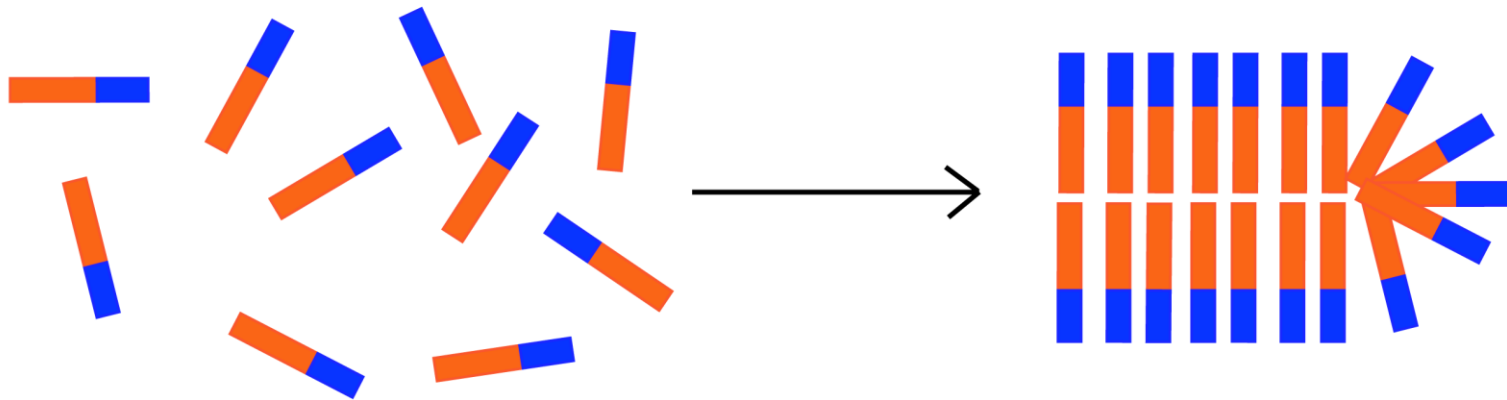
Membrane lipids

General structure of membrane components



In water the amphipatic membrane components segregate **spontaneously** driven by **entropy**.

This process is called **self assembly**



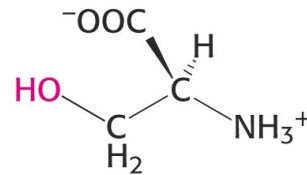
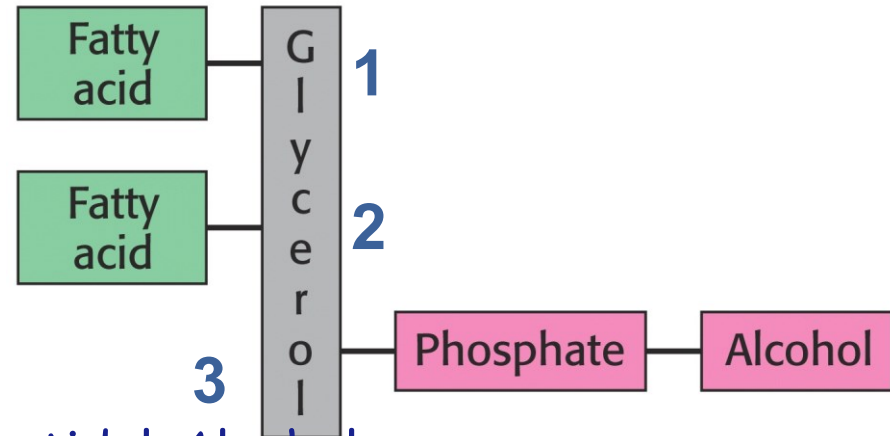
=> flexible

=> dynamic

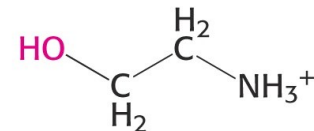
=> self healing

Glycero-phospholipids

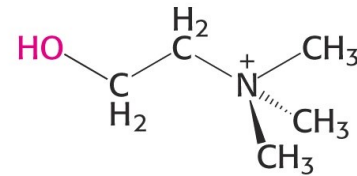
- Glycerol backbone
- Fatty acids
 - sn1: saturated
 - sn2: unsaturated
- Alcohol- P_i -ester on sn-3
- Name "1-FA₁-2-FA₂-sn-phosphatidyl-Alcohol"



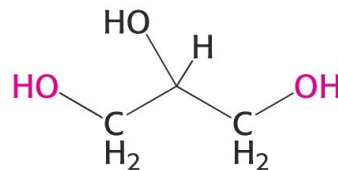
Serine



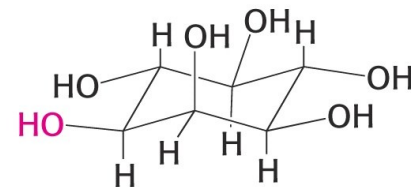
Ethanolamine



Choline



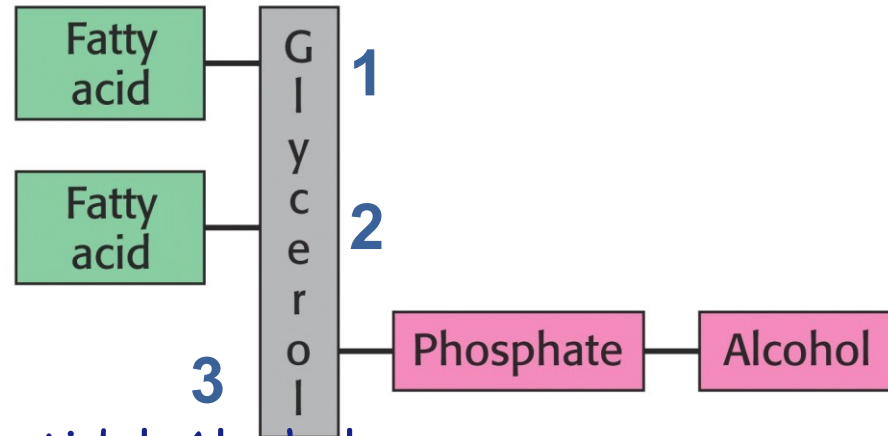
Glycerol



Inositol

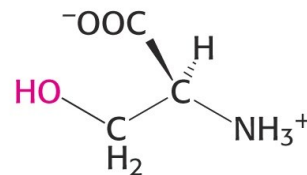
Glycero-phospholipids

- Glycerol backbone
- Fatty acids
 - sn1: saturated
 - sn2: unsaturated
- Alcohol- P_i -ester on sn-3
- Name "1-FA₁-2-FA₂-sn-phosphatidyl-Alcohol"

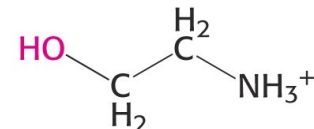


• Charge:

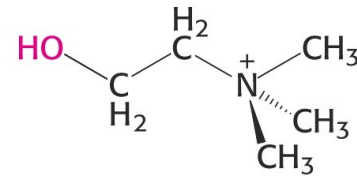
±	PC, PE
-	PI, PG
2-	CL
-/2-	PA



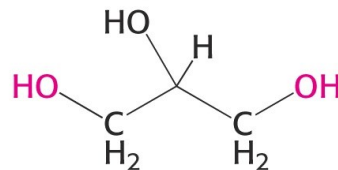
Serine



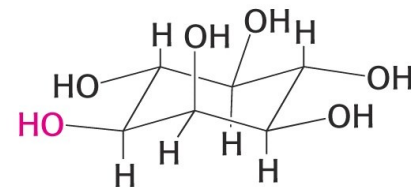
Ethanolamine



Choline



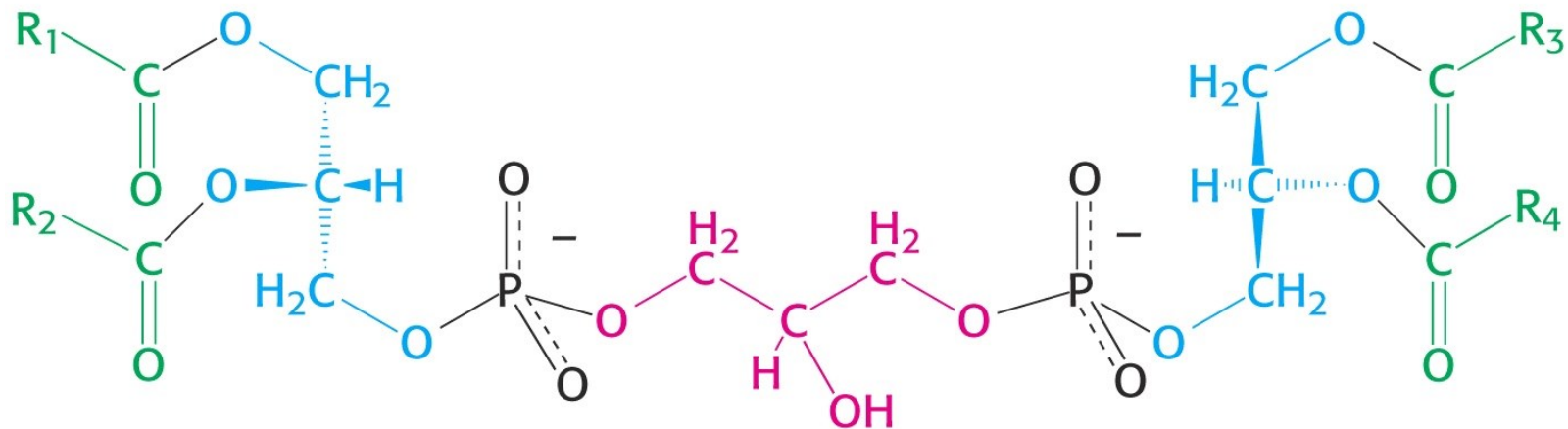
Glycerol



Inositol

Glycero-phospholipids

- Cardiolipin: a double lipid.
In eukaryotes, only present in mitochondria.

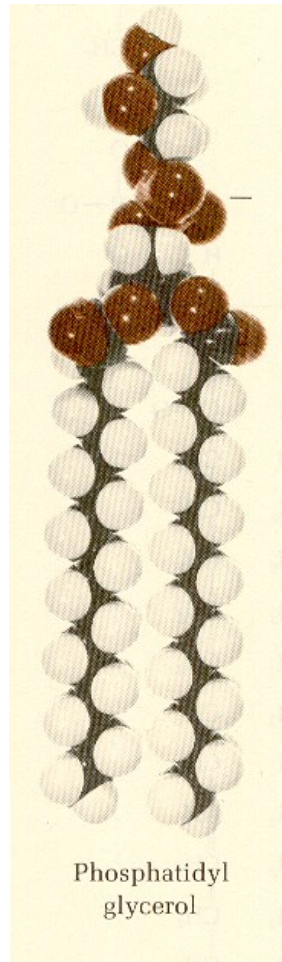


Diphosphatidyl glycerol (cardiolipin)

Model of phospholipids

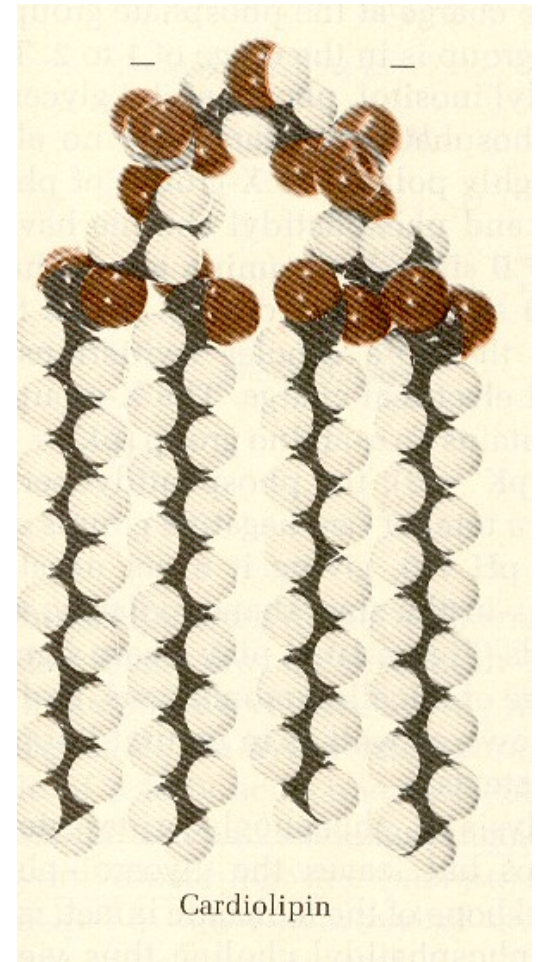


PE



Phosphatidyl
glycerol

PG



Cardiolipin

Cardiolipin

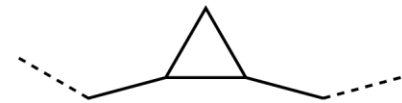
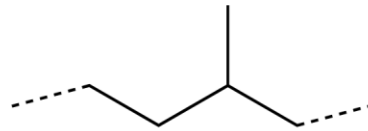
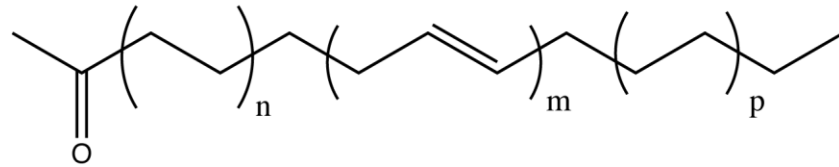
Fatty acids

- Fatty acids

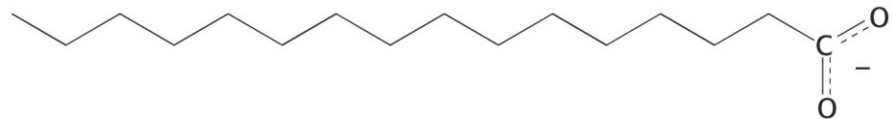
even - chain length 12 - 36

cys - saturation $0 \leq m \leq 6$

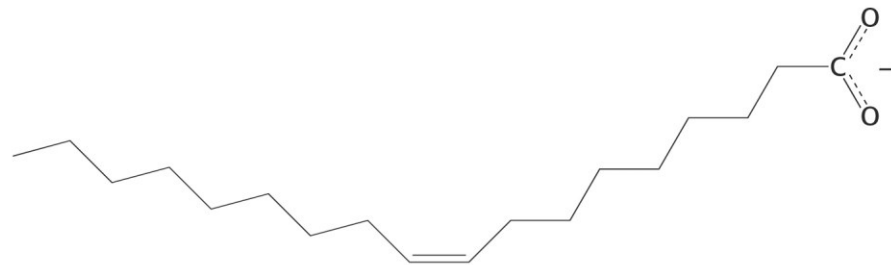
- branching



- Highly variable
- Phase transition
- Accommodate proteins
- Phase separation
- Signalling
-
-



Palmitate

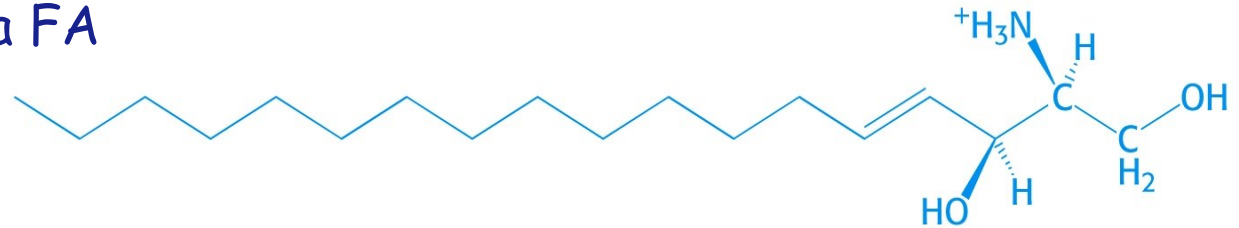


Oleate

(ionized form of oleic acid)

Phospho-sphingolipids

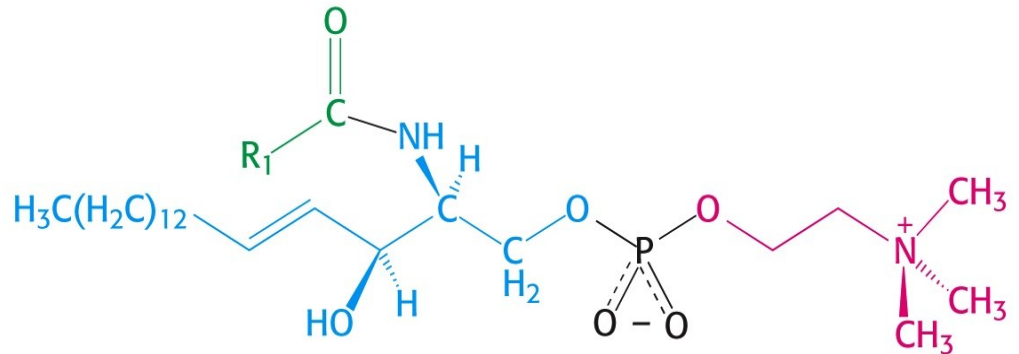
- Sphingosine backbone
 - already includes a FA



Sphingosine

- + Fatty acid linked via peptide bond
 - => ceramide

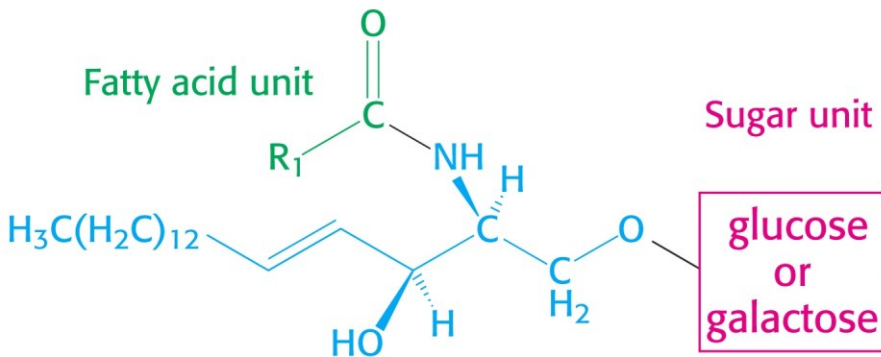
- Alcohol-Pi-ester
 - choline => Sphingomyelin
 - ethanolamine
 - glycerol



Sphingomyelin

Glyco-sphingolipids

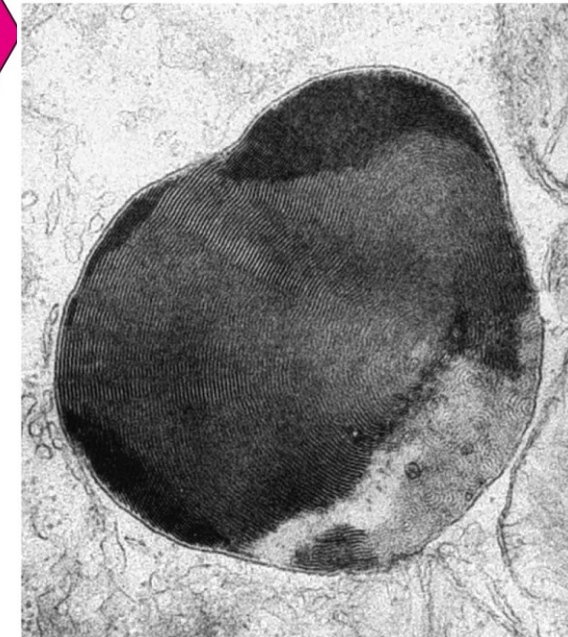
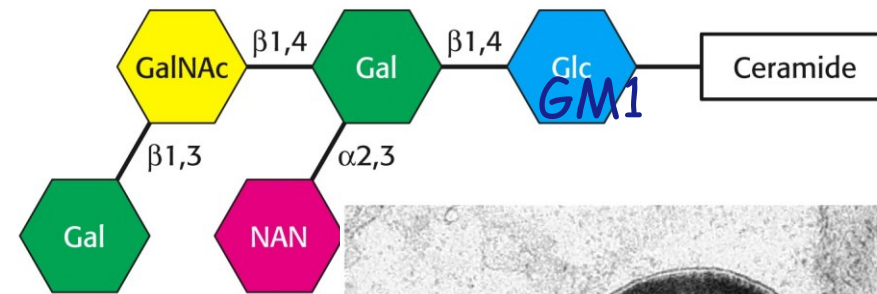
- Ceramide having sugars as polar head group
 - simple, e.g. 1 sugar => cerebroside
 - complex, e.g. 5 sugars => ganglioside GM1



Cerebroside

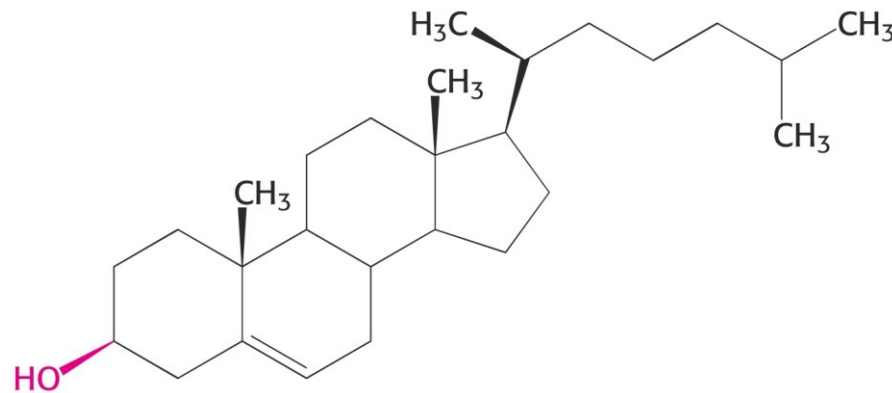
(glycolipid)

- Outer surface of plasma membrane; myelin
- Blood group antigens on erythrocyte.



Sterols

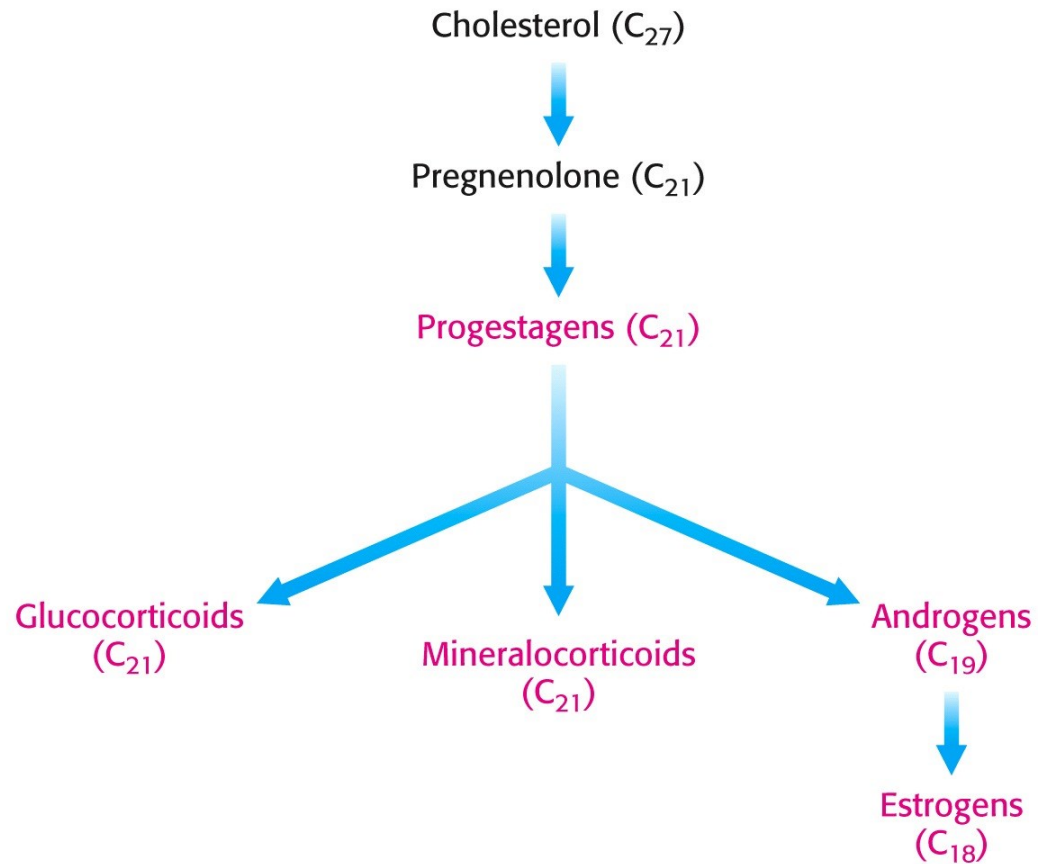
- Cholesterol is main sterol in mammals
 - mainly present in plasma membrane, but also in endo- and exo-cytotic pathway.
 - profound effect on phase behavior of membranes.
 - domain formation.



Cholesterol

Sterols

- Cholesterol a precursor of steroid hormones.



General properties of membranes

- | | | |
|----------------------|----|------------------------------|
| • Dynamic nature | => | protein function & diffusion |
| • Phase transitions | => | domain formation |
| • Polymorphism | => | fission and fusion |
| • Surface pressure | => | protein insertion |
| • Permeability | => | barrier |
| • Electrical effects | => | concentration gradients |

Dynamic nature of membranes

Simulation:

All is moving!

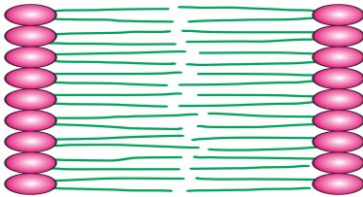
QuickTime™ and a YUV420 codec decompressor are needed to see this picture.

Dynamic nature of membranes

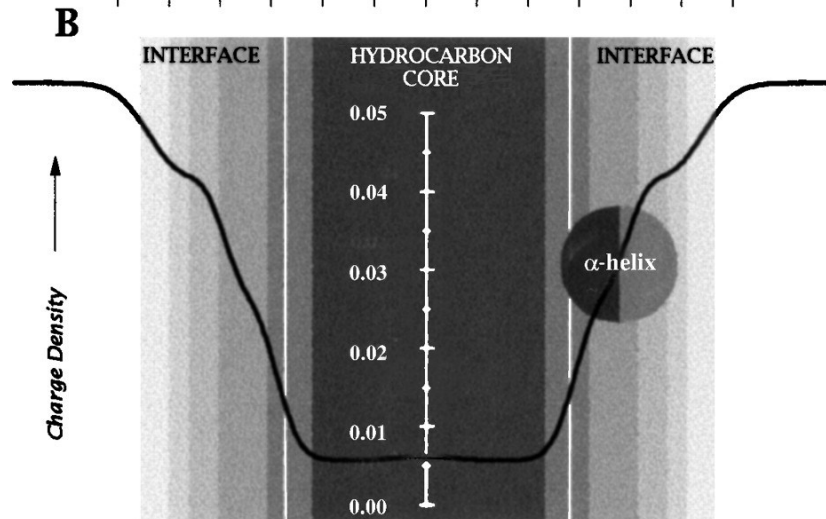
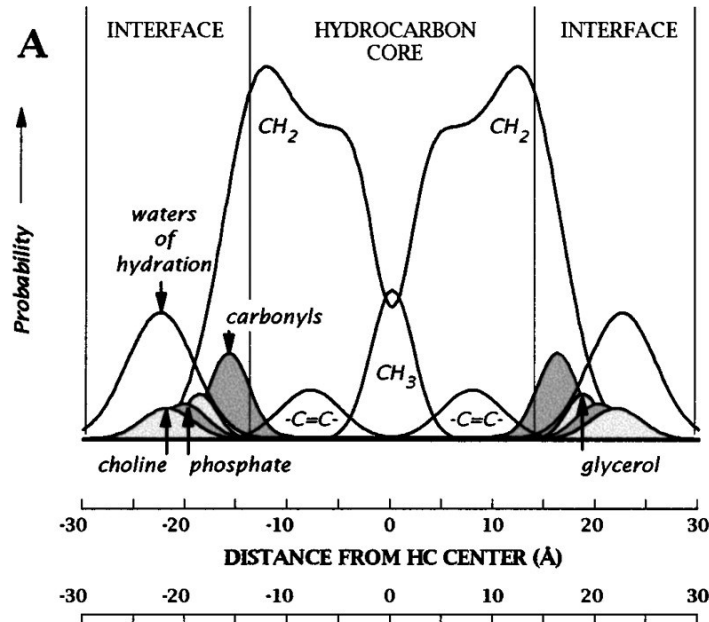
Diffraction studies:

Distributions are wide:

- Reflecting simulations
- Rejecting



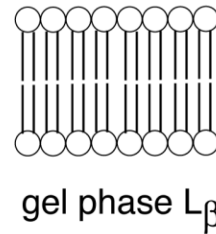
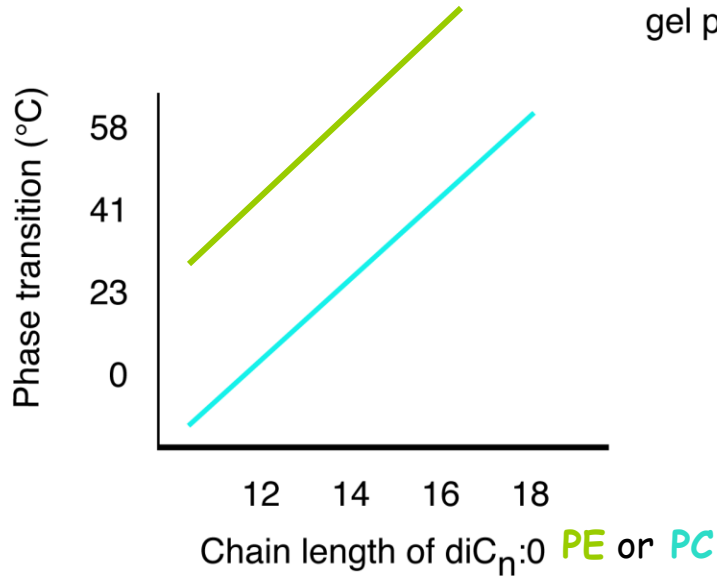
- Hydrophobic core of $\pm 25 \text{ \AA}$
- Interfacial region of $\pm 15 \text{ \AA}$ with a large range of properties.



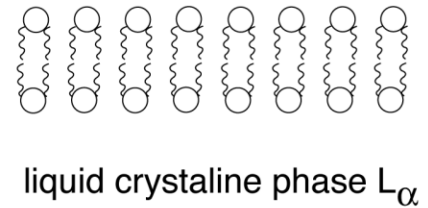
Phase transitions of lipids

- Temperature

- melting of the chains
- => membrane thickness
- => molecular packing



Temperature
increase



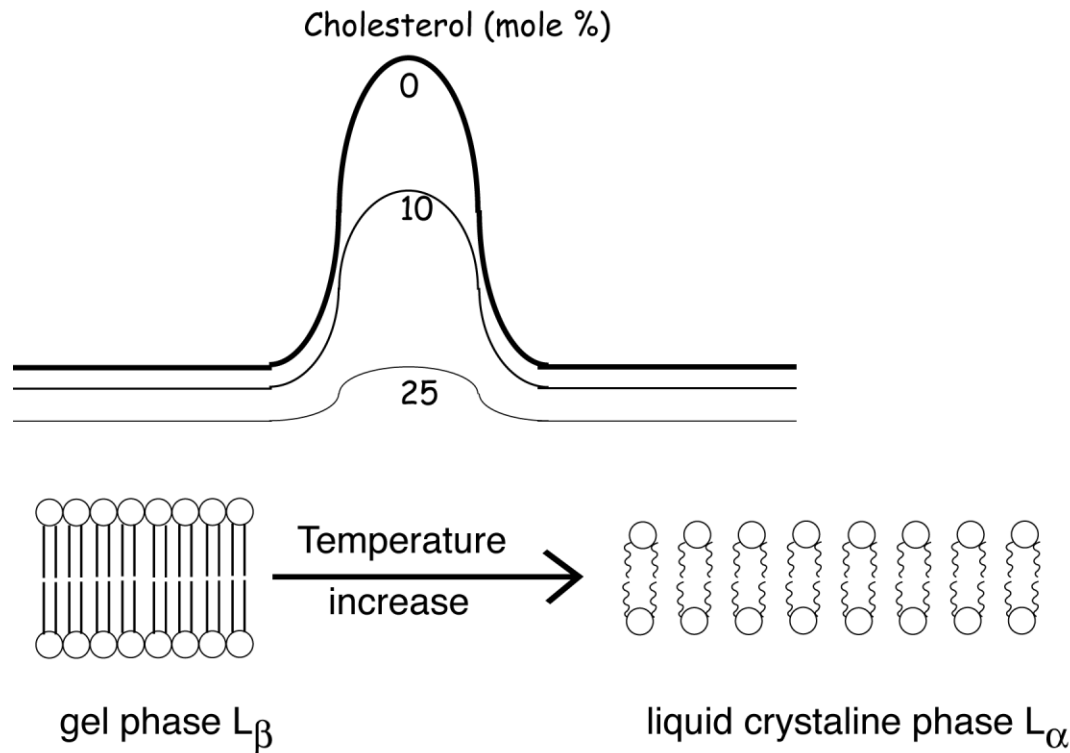
- => $\pm 20^\circ\text{C}$ per two C2-units
- => $\pm 30^\circ\text{C}$ per cis-double bond
- => choline $\pm 30^\circ\text{C}$ lower than ethanolamine
- =>

- ◇ Temperature affects fluidity => diffusion increases 100-fold from gel to liquid phase
- ◇ FA-composition affects phase transition
- ◇ Cell membranes want to be liquid crystalline.

Phase transitions of lipids

- Cholesterol

- "liquifies" gel phase & "solidifies" liquid crystalline phase
 - membrane permeability decreases
- => phase transition disappears



Phase transitions of lipids

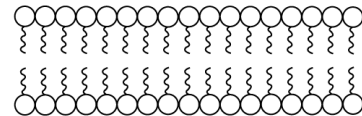
- Buffer composition

- pH

e.g. PA

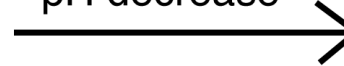
- Divalent cations

e.g. CL

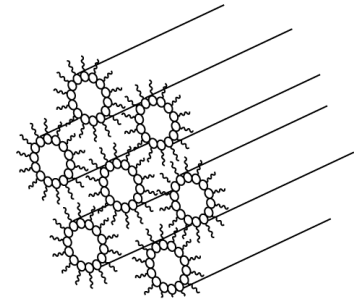
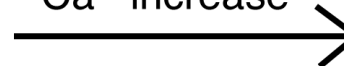


L-phase

pH decrease



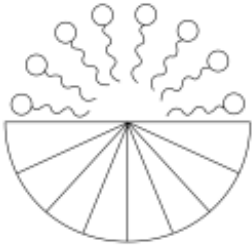
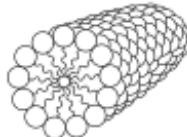
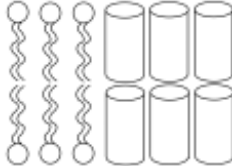
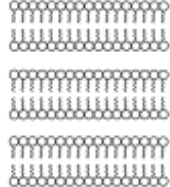
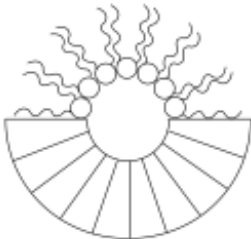
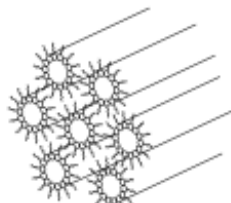
Ca[#] increase



H_{II}-phase

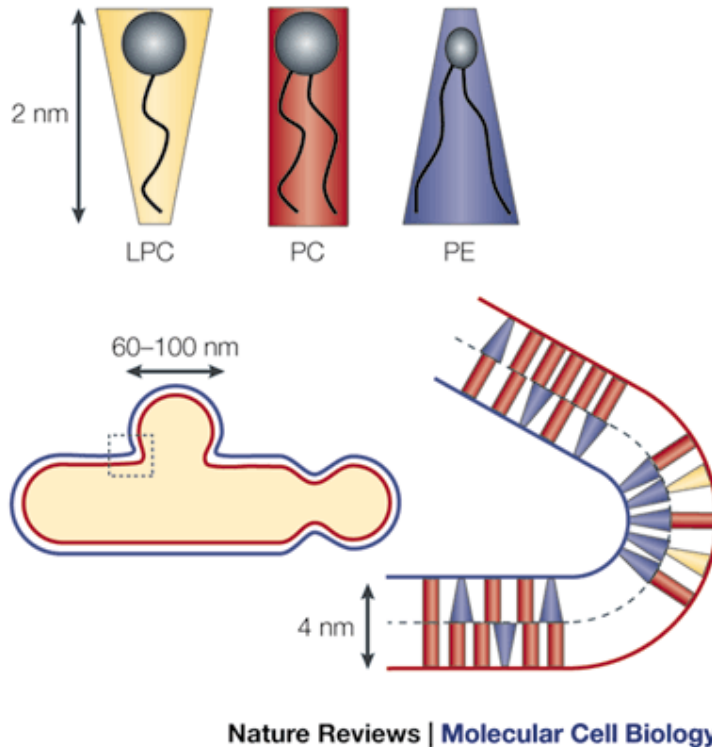
Lipid polymorphism

- Relative size of polar head group and hydrophobic part of lipid

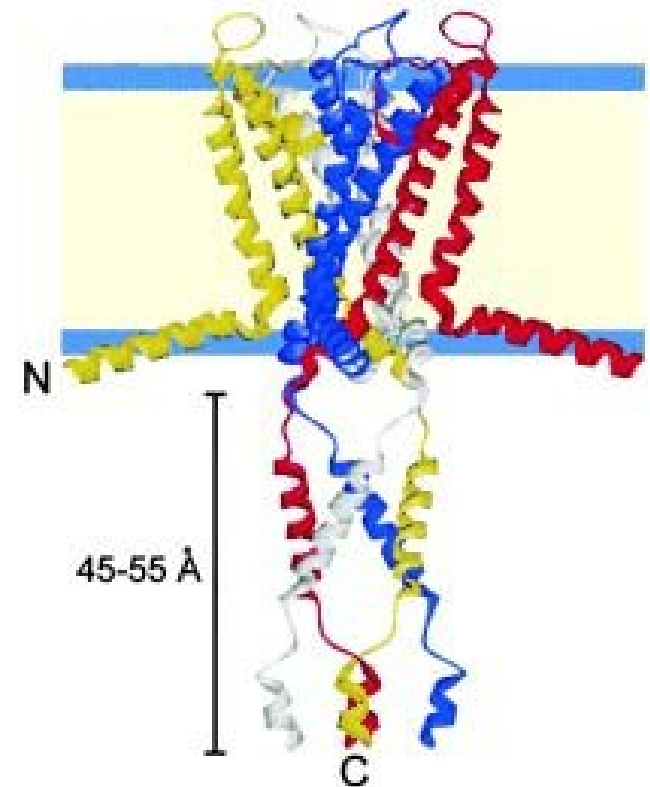
Species	Shape	Organization	Phase	
Soaps □ Detergents Lysophospholipids	 Inverted cone	 Micelles	Isotropic Hexagonal 1	
Phosphatidylcholine Phosphatidylserine Phosphatidylinositol Sphingomyelin Dicetylphosphate DODAC	 Cylinder	 Bilayer	Lamellar (Cubic)	
Phosphatidylethanolamine Phosphatidic acid Cholesterol Cardiolipin Lipid A	 Cone	 Reverse micelles	Hexagonal 2	

Lipid polymorphism in membranes

- Extreme curvature of membrane

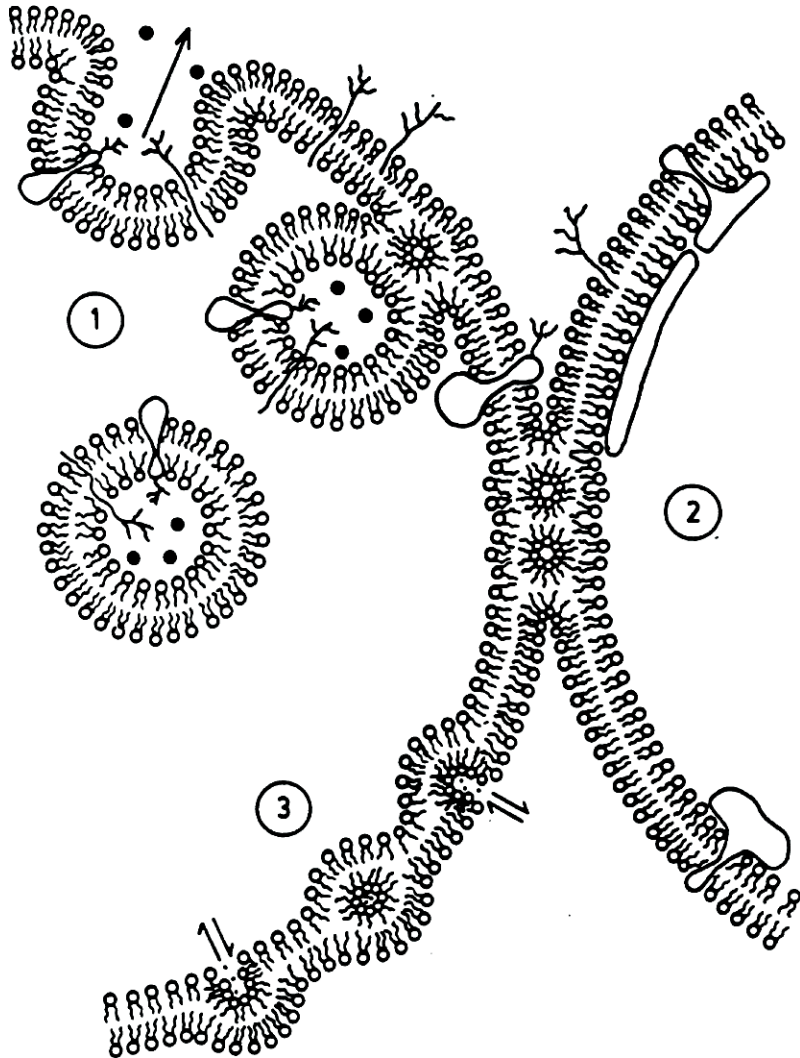


- Accommodate proteins

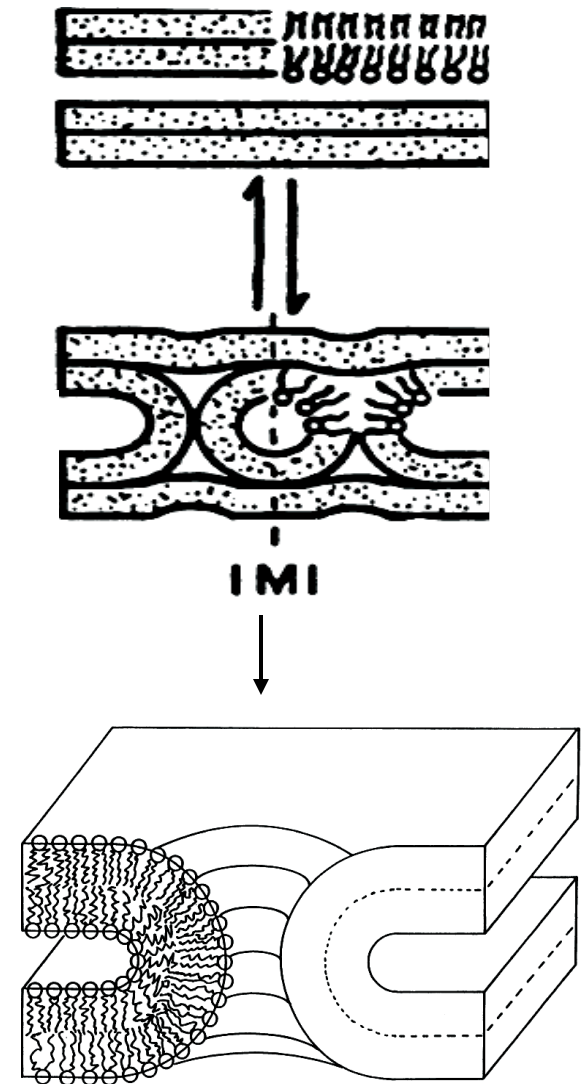


Lipid polymorphism in membranes

- Membrane fusion & vesicle formation



Biochem



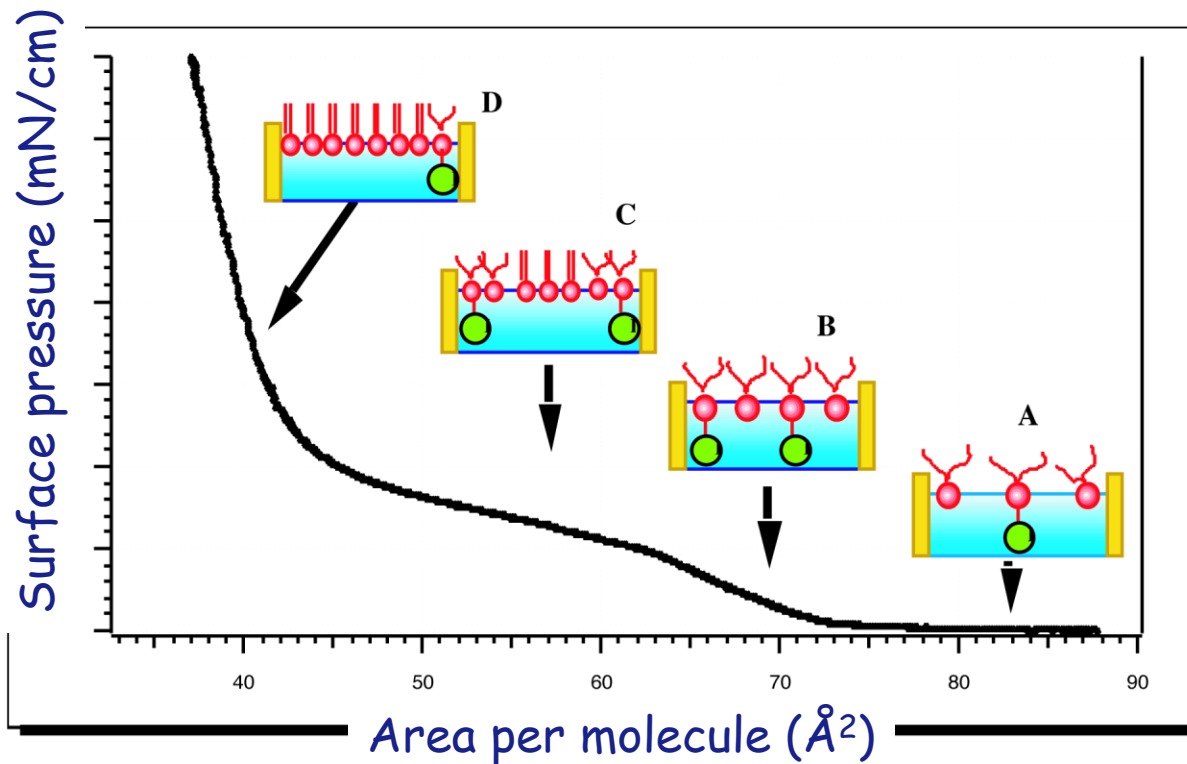
ILA

Phase transitions of lipids

- Surface pressure Π

A lipid monolayer at the air-water interface of a trough with variable surface

Measure surface tension $\gamma \Rightarrow \Pi = \gamma_o - \gamma.$



Phases:

A "gass"

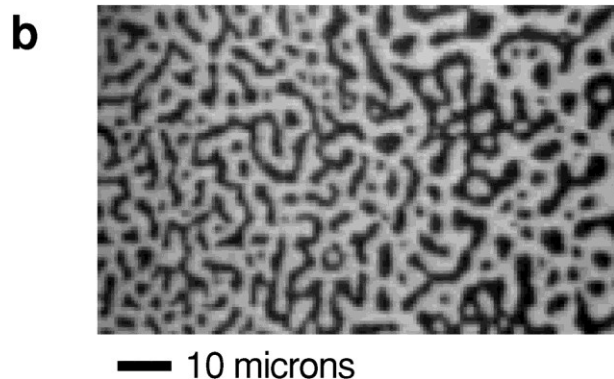
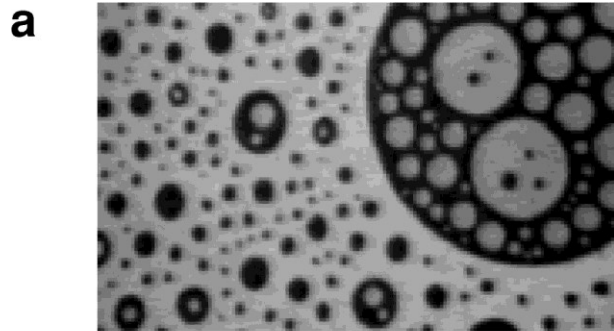
B "liquid"

C co-existence

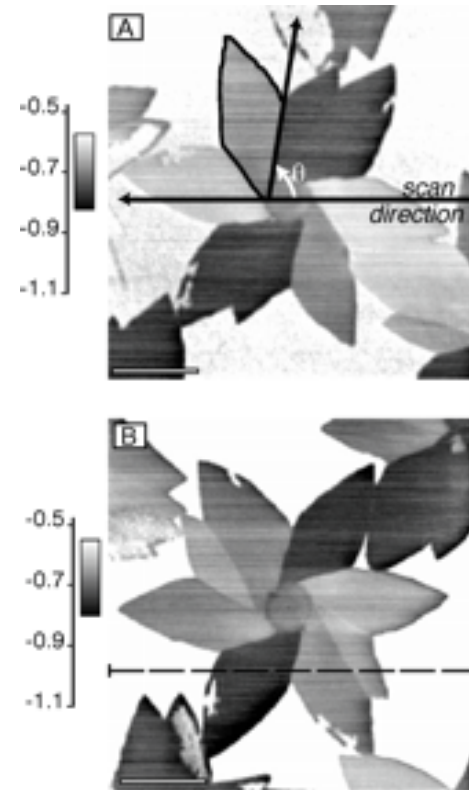
D "solid"

Phase separations of lipids

- Co-existence of condensed and disordered phase
micrometer-sized “solid” domains



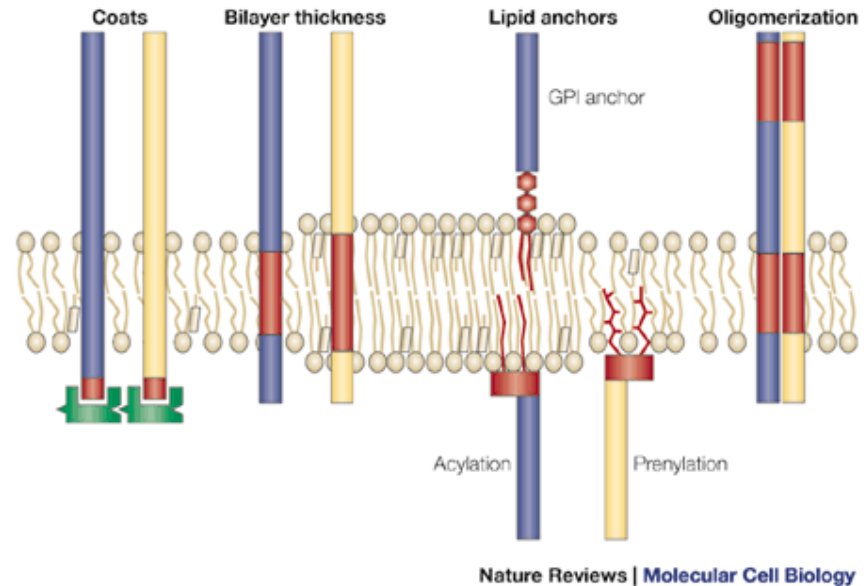
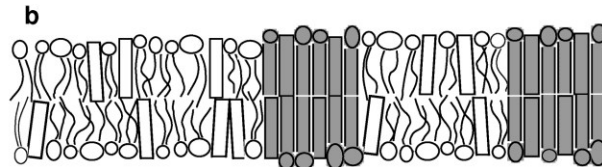
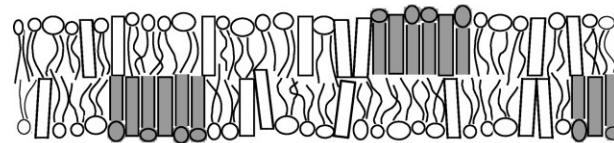
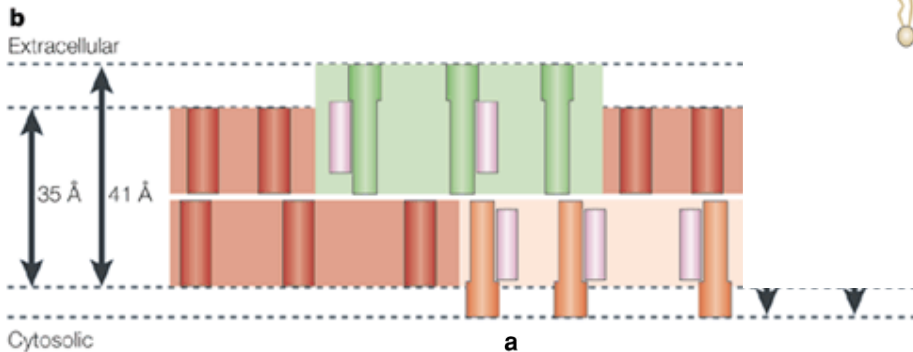
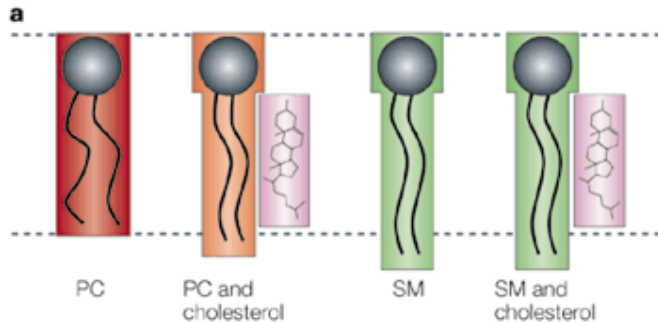
- Fluorescence microscopy



- Atomic force microscopy

Phase separations within membranes

- Cholesterol: Preference for sphingomyelin and not so "fluid" lipids



- independent monolayers

- coupled monolayers

Phase separations within artificial membranes



- Co-existence of ordered and disordered phase in artificial vesicles made of:
 - cholesterol
 - sphingomyelin
 - di-oleoyl-phosphatidyl-choline

Blue: ordered phase

Red: disordered phase

W. Webb 23.10.2003

Phase separations within artificial membranes

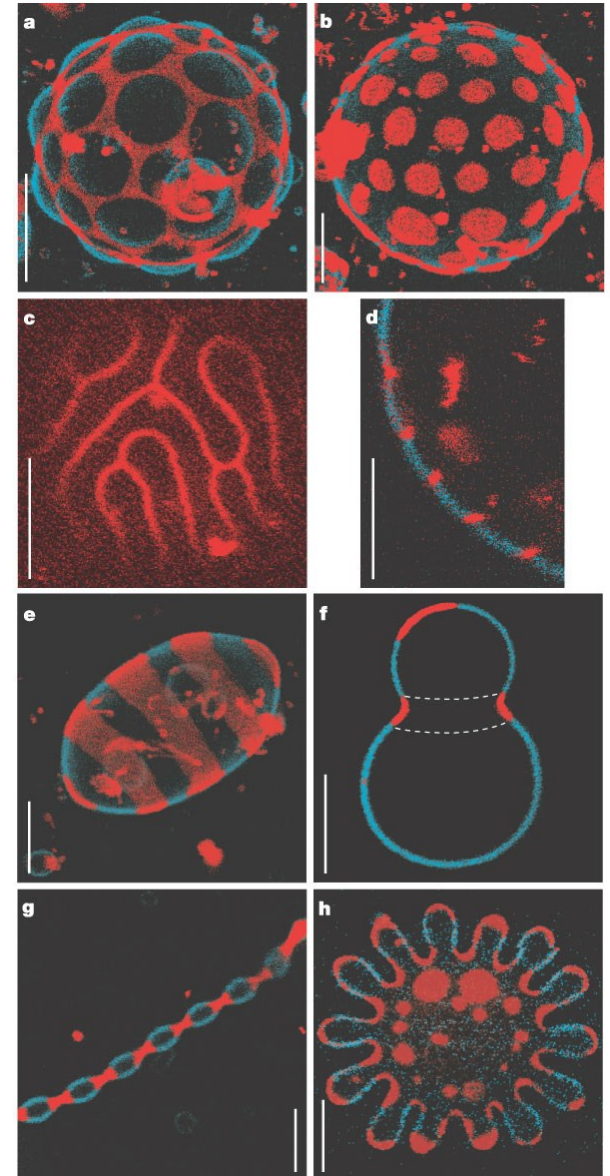
- Co-existence of ordered and disordered phase in artificial vesicles made of:
 - cholesterol
 - sphingomyelin
 - di-oleoyl-phosphatidyl-choline

Variables:

Composition & Temperature

Blue: ordered phase

Red: disordered phase

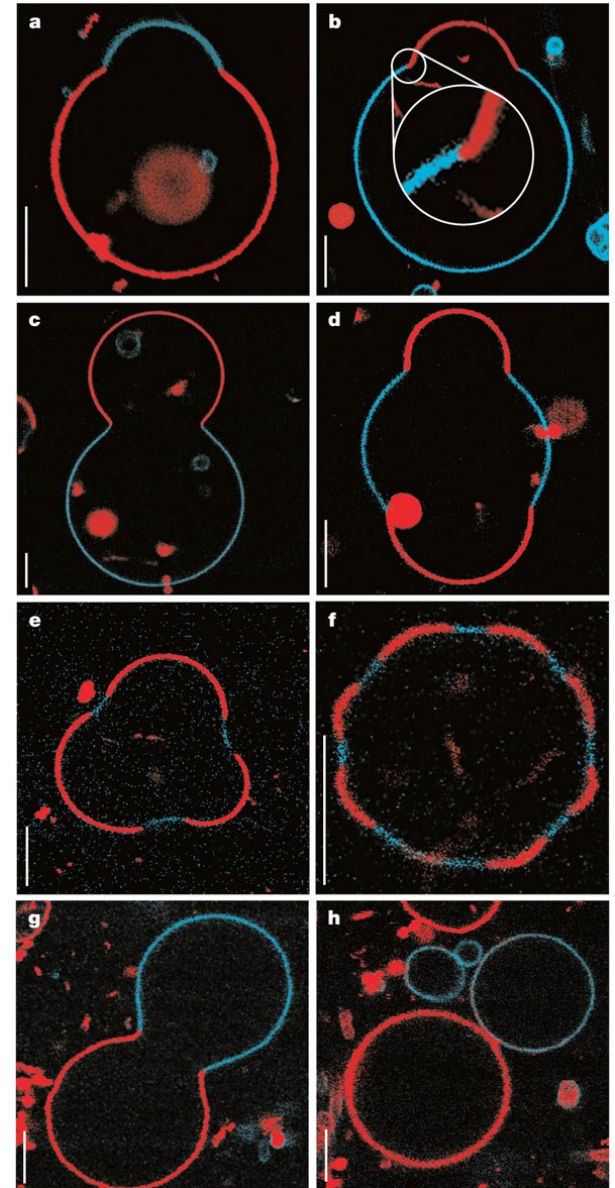


Phase separations within artificial membranes

- Co-existence of condensed and disordered phase
micrometer-sized “solid” domains

Blue: ordered phase

Red: disordered phase



Phase separations within artificial membranes

- Co-existence of condensed and disordered phase
micrometer-sized “solid” domains

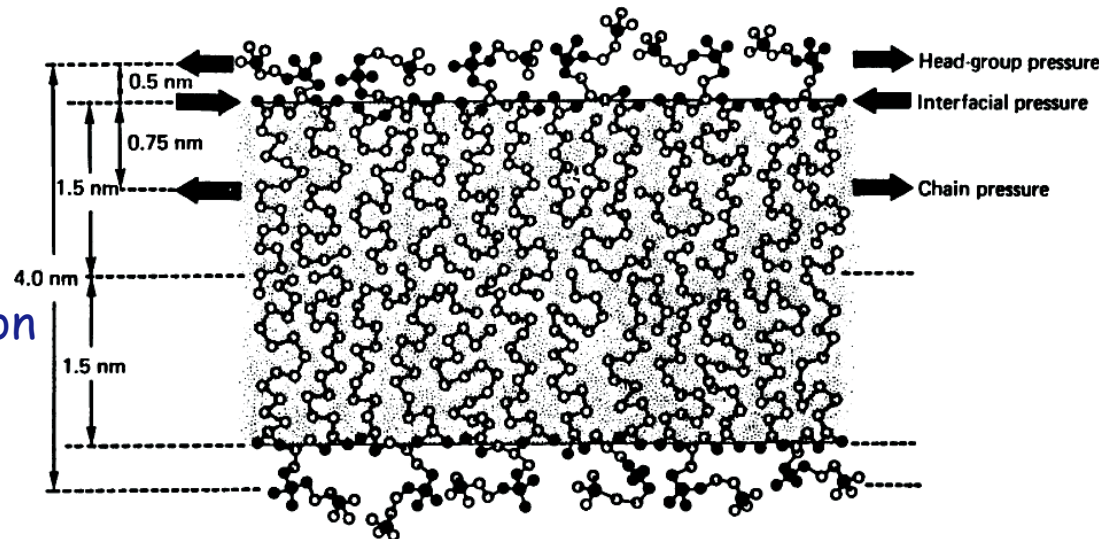
Black: ordered phase

White: disordered phase

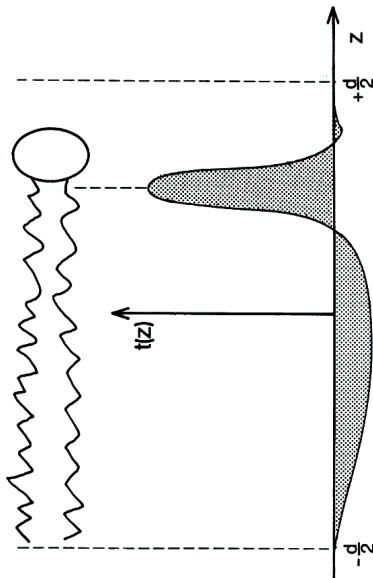
Surface pressure of membranes

◇ "Pressures" within a membrane

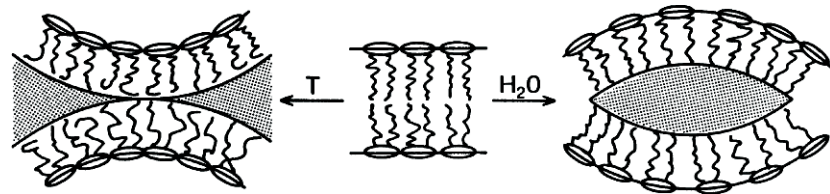
- Attraction ÷ H-bonds
÷ hydrophobic effect
- Repulsion ÷ head group hydration
÷ thermal fluctuation



◇ Stress profile



◇ Effect of "stress"



Surface pressure of membranes

Relative hydrolytic activity of phospholipases on erythrocytes and monolayers

Erythrocytes

Monolayer

max Π (mN/m)

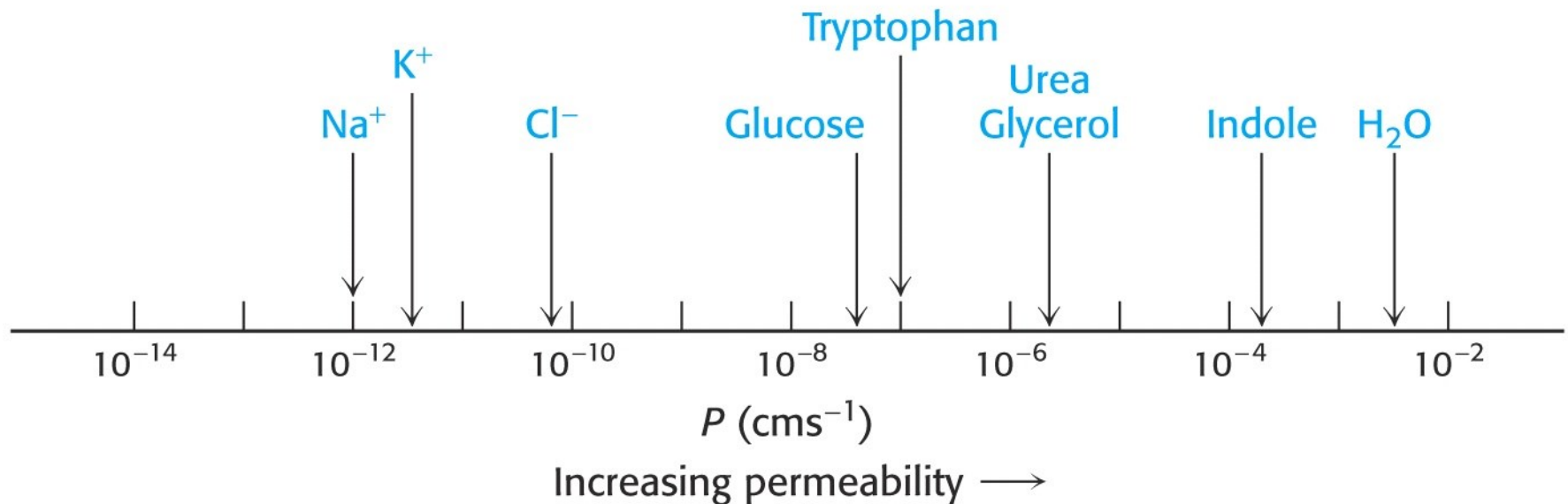
PL-A ₂	Pig pancreas	-	16.5
PL-D	Cabbage	-	20.5
PL-A ₂	C. Adamanteus	-	23
PL-C	B. Cereus	-	31
PL-A ₂	n. Naja	+	35
PL-A ₂	Bee venom	+	35.5
PL-C	Cl. Welchii	+	> 40

=> For erythrocyte membrane $31 < \Pi < 35$ mN/m

Permeability of Membranes

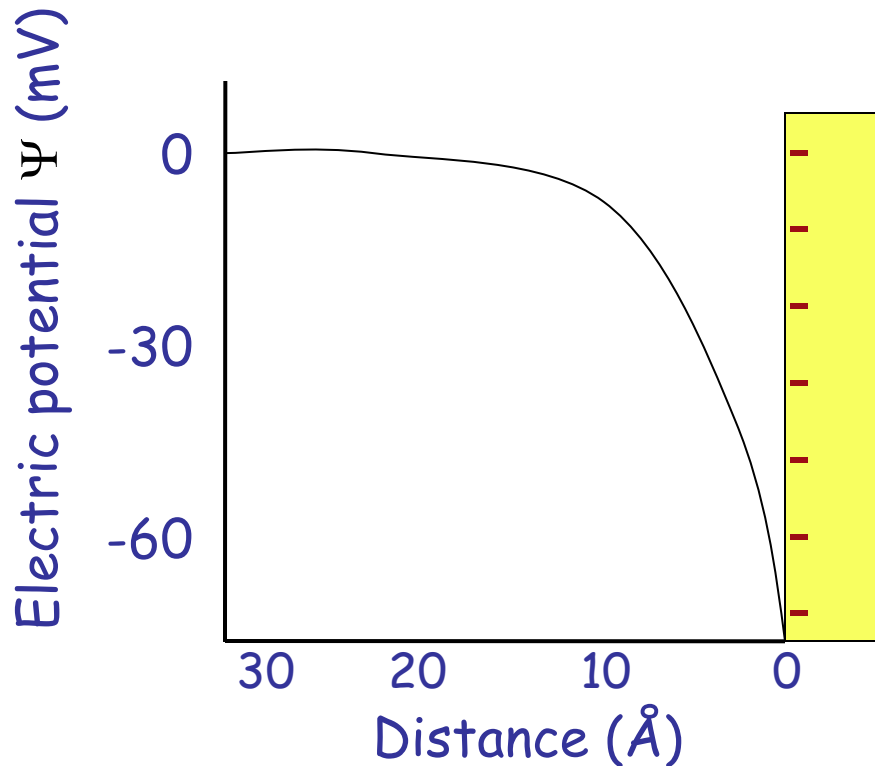
- The hydrophobic core of the membrane is an ideal barrier for hydrophilic (charged and polar) solutes.

=> Ion gradients can be created that are highly energetic.



Electric potential of membranes

- Membrane surfaces are in general negatively charged

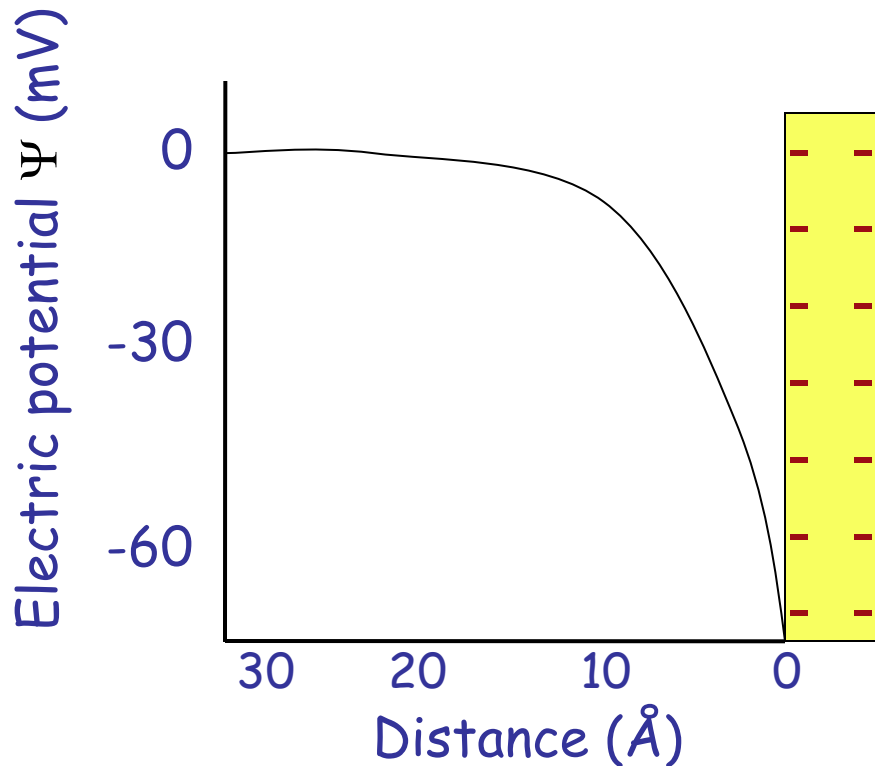


Gouy-Chapman

<= Molecular dimensions!

Electric potential of membranes

=> Concentration of cation & depletion of anion



Surface concentration C_o
with $\Psi = -60$ mV

-2	$0.01 \times C_\infty$
-1	0.1
+1	10
+2	100

Integrative questions

- ◇ Formation of a membrane: Entropy?
- ◇ Place of double bond on phase transition temperature.
- ◇ What would be the problem of crossing a phase transition?
- ◇ Why are proteins within membranes highly structured?
- ◇ Peptide insertion in a membrane.
- ◇ Phase transitions, phase separation, diffusion & lipid polymorphism.

Further reading

Text books:

Biochemistry - Stryer	Chpt 12,26
The Cell - Alberts	Chpt 10
Biomembranes - Gennis	Chpt 1, 2 & 7

Review articles:

Understanding the diversity of membrane lipid diversity, Nature reviews Molecular Cell Biology; from Riezman (Uni Geneva) 2018

- W. Dowhan "Molecular basis for membrane phospholipid diversity" Annu Rev Biochem 66 (1997) 199-232.
- A.R. Cossins & M. Sinensky "Adaption of membranes to temperature, pressure and exogenous lipids" in "Physiology of membrane fluidity" (1984) 1-20.

Membrane protein structures: see the PDB : Protein Data Bank

<https://www.rcsb.org>

Next time: Phospholipid biosynthesis

Review articles:

- W. Dowhan "Molecular basis for membrane phospholipid diversity" *Annu Rev Biochem* 66 (1997) 199-232.
- A.R. Cossins & M. Sinensky "Adaption of membranes to temperature, pressure and exogenous lipids" in "Physiology of membrane fluidity" (1984) 1-20.