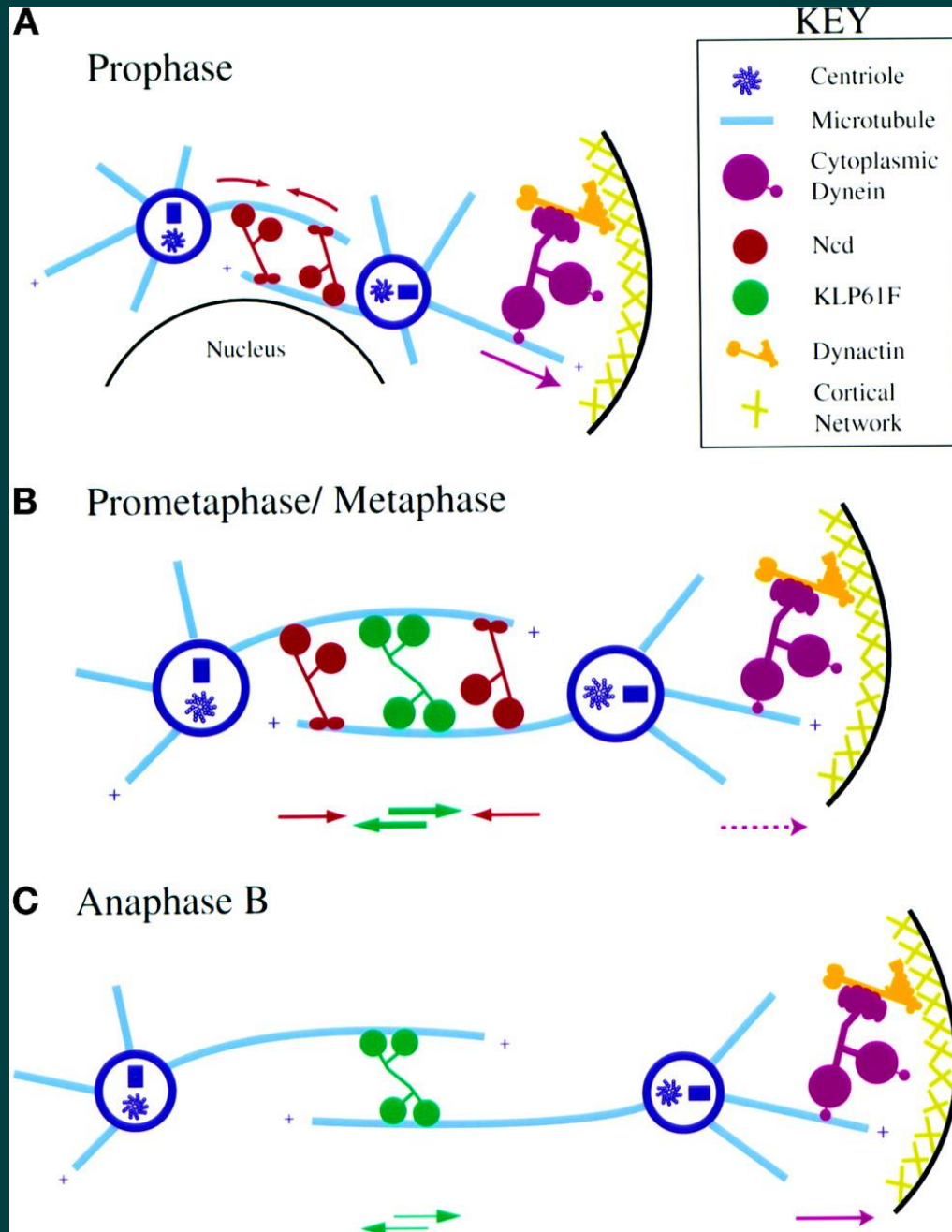


last time: motor proteins in mitosis

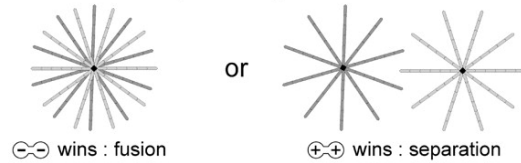


Sharp et al.,
Mol. Biol. Cell, 2000

Conditions for steady-state pole separation

Screen 1: two kinds of *homo-complexes* $\text{U} + \text{V}$

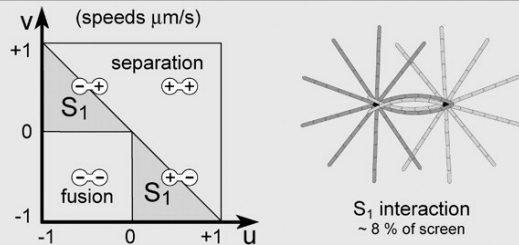
no balance of forces, one complex wins.



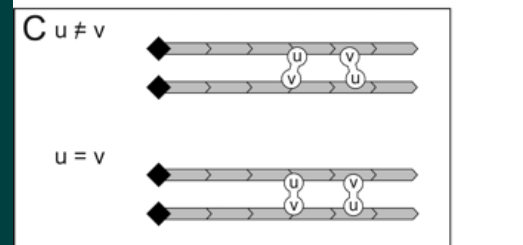
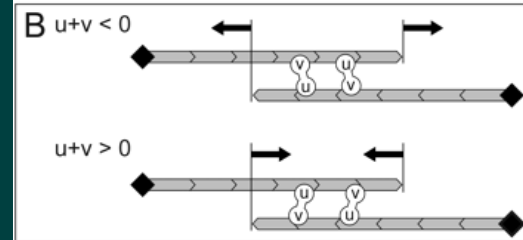
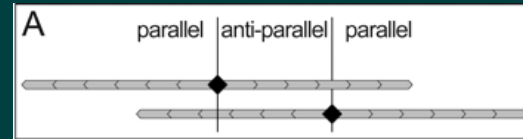
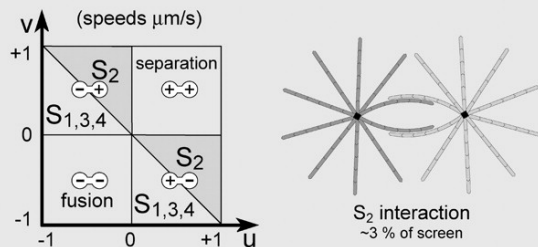
Screen 2: two kinds of *mixed complexes* $\text{U} + \text{V}$

no balance of forces: fusion or full separation

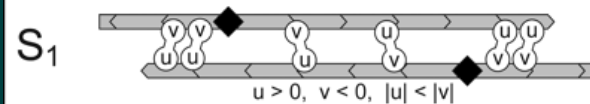
Screen 3a: one kind of *hetero-complex* $\text{U} + \text{V}$



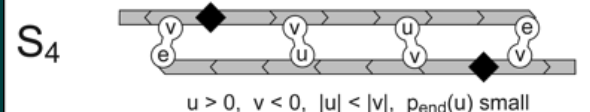
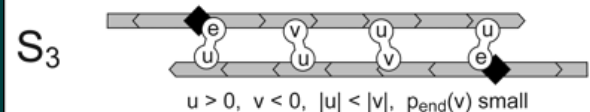
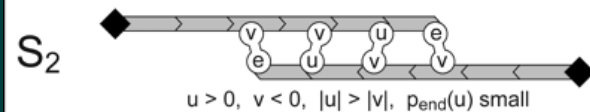
Screen 3b: one kind of *hetero-complex, which can stay at microtubule ends* $\text{U} + \text{V}$



Solutions where motors detach immediately at microtubule ends



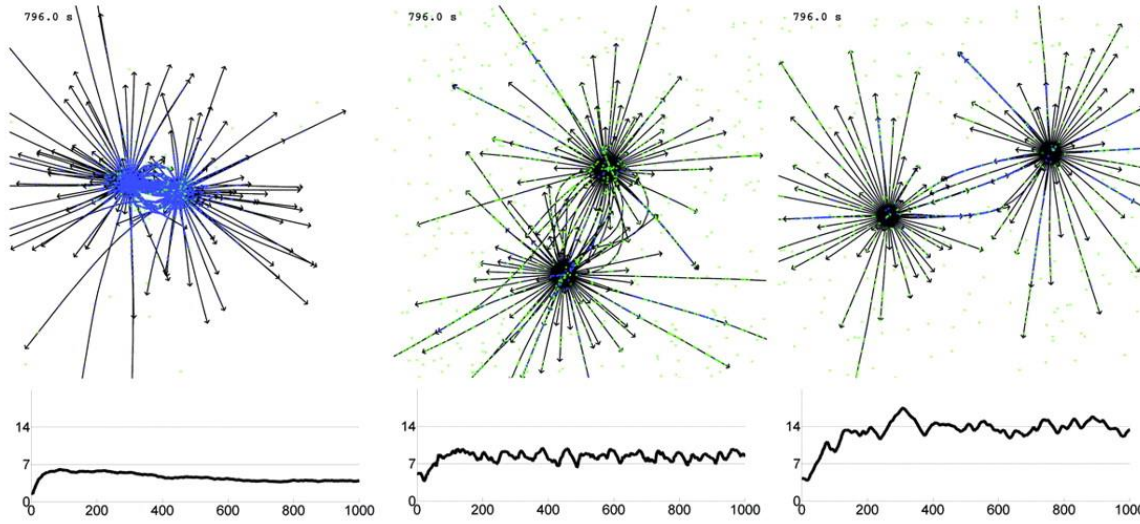
Solutions where motors can halt at microtubule ends



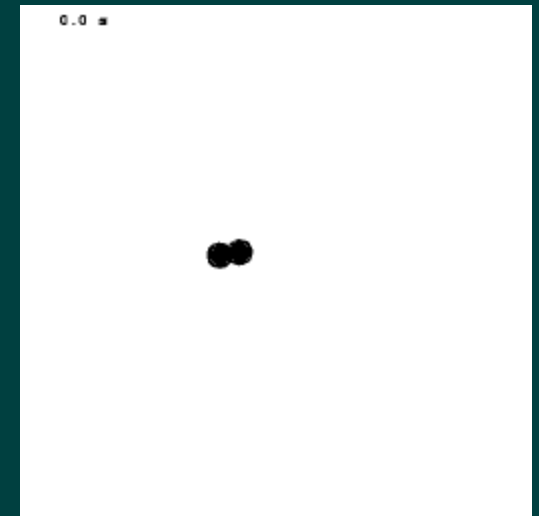
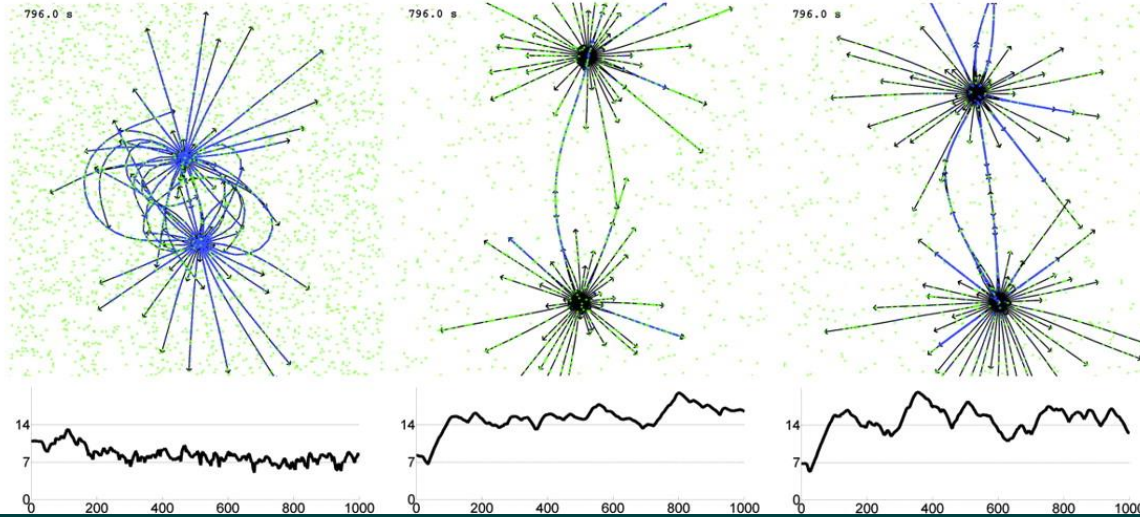
Nedelec, J. Cell Biol., 2002

S₁

10 μ m

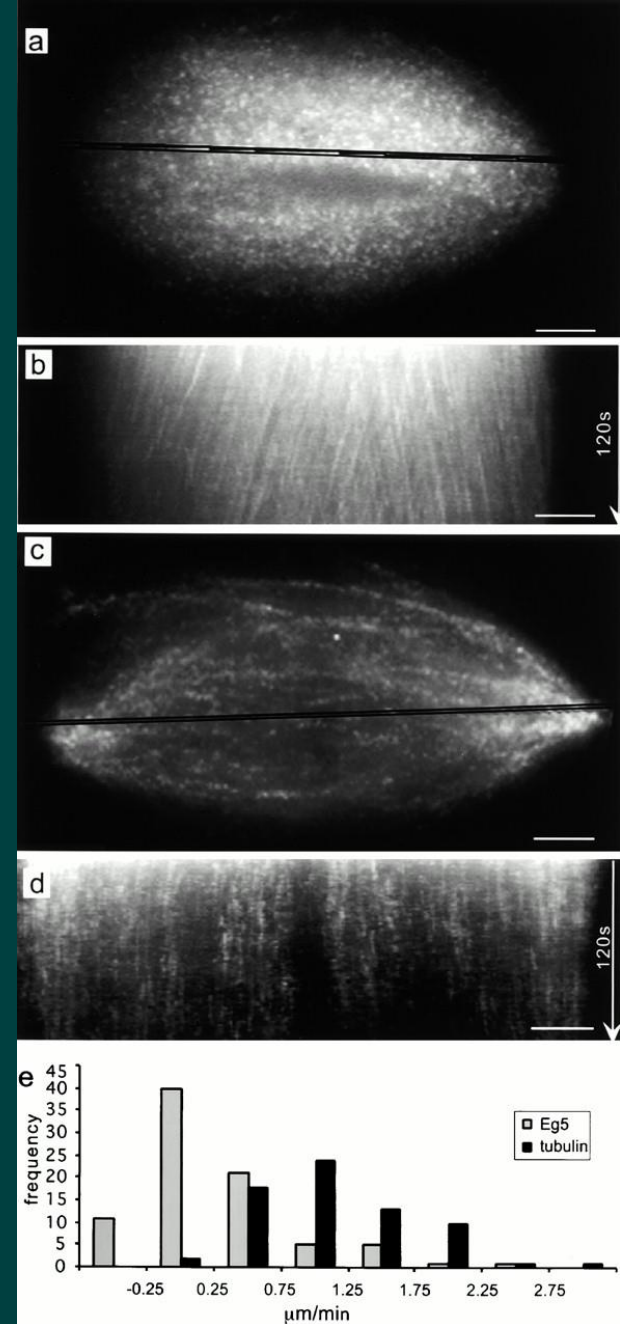
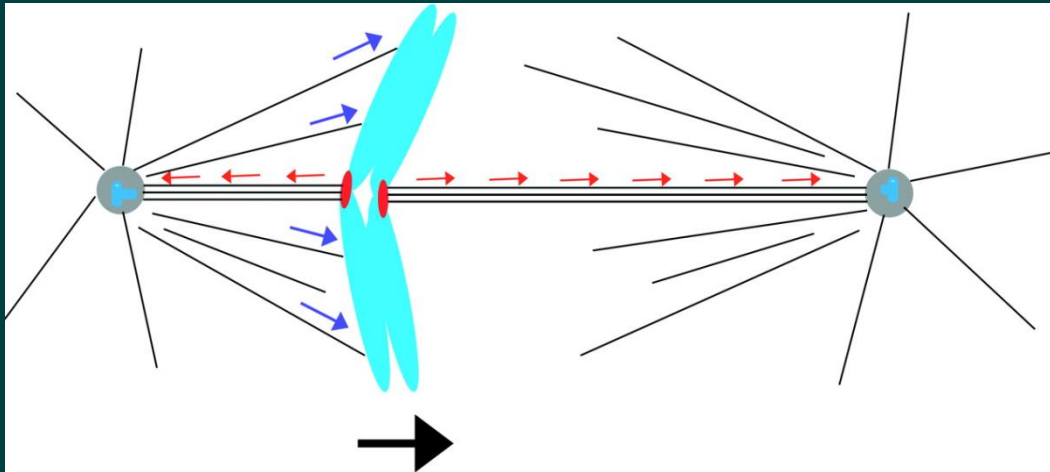


S₂

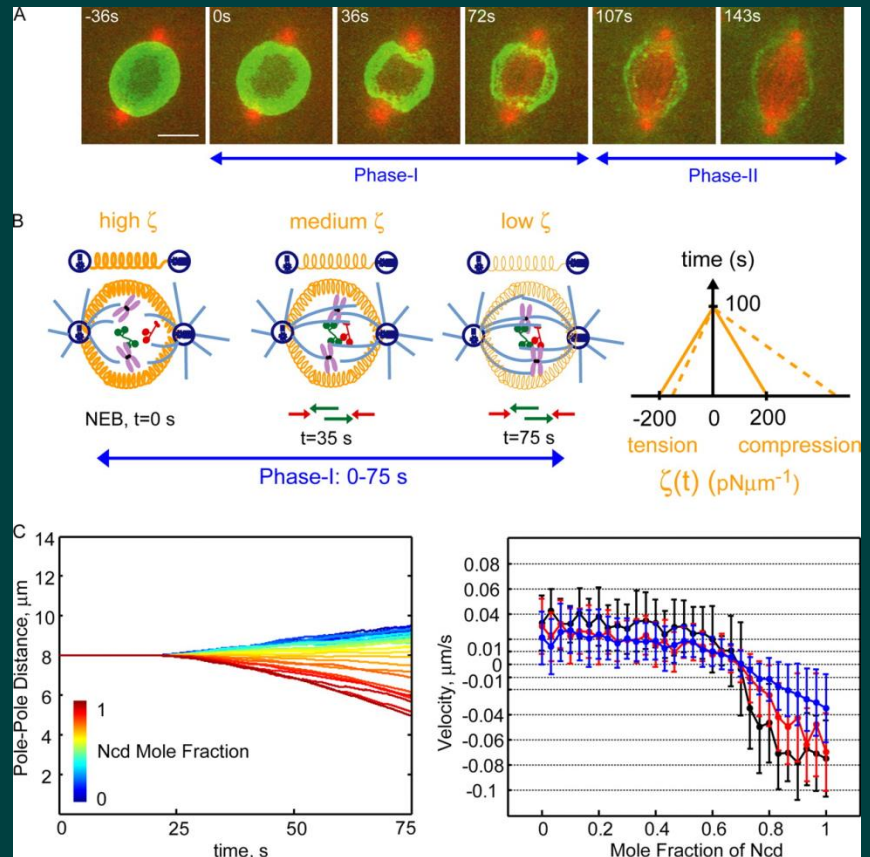
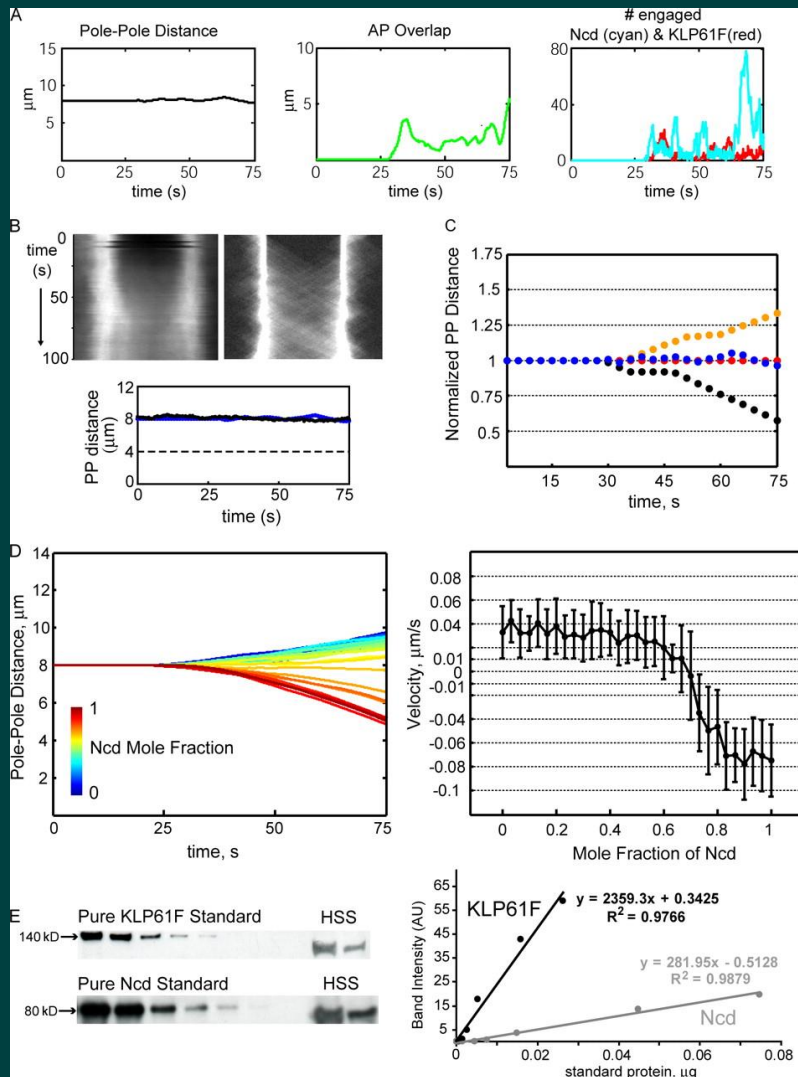


Nedelec, J. Cell Biol., 2002

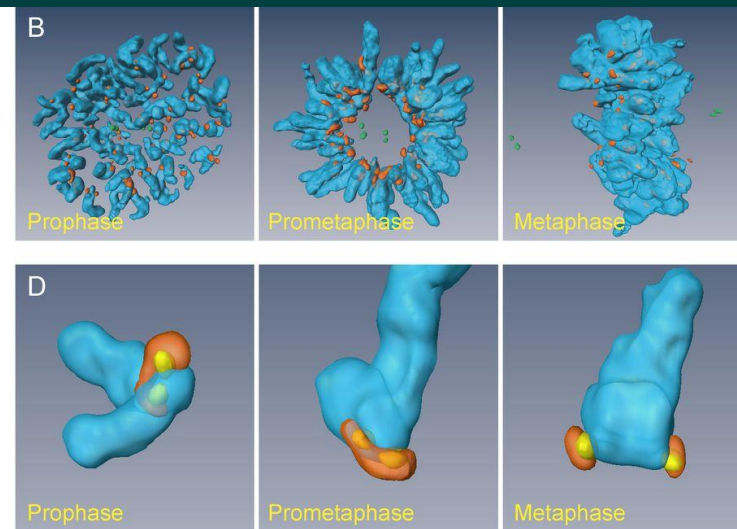
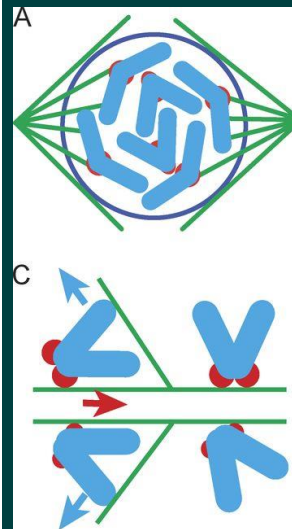
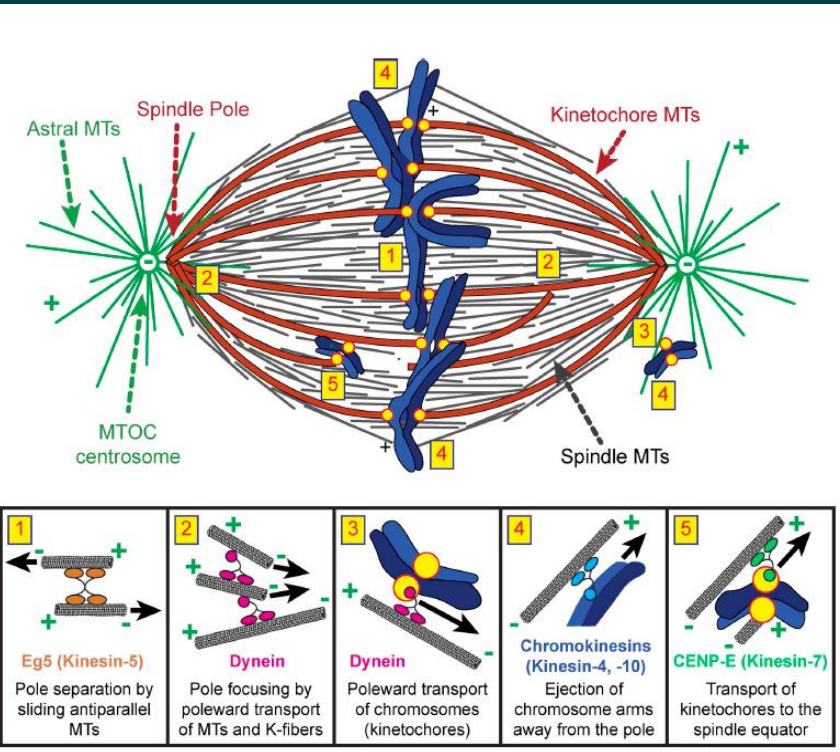
Early ideas about forces in mitotic spindle:
traction fiber, polar wind and spindle matrix



more recent force-balance model: force-dependent detachment of motors Nuclear lamina can stabilize the separation

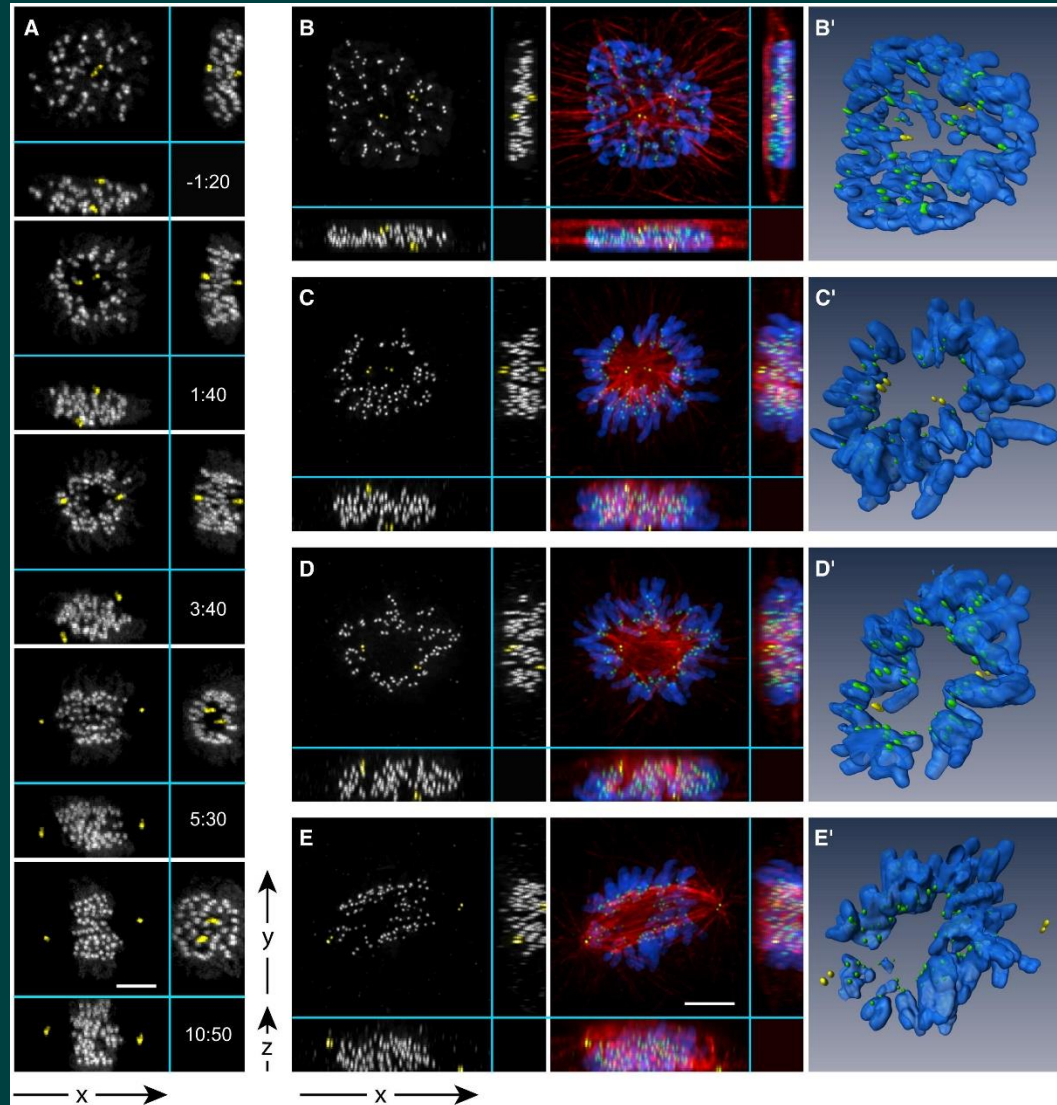
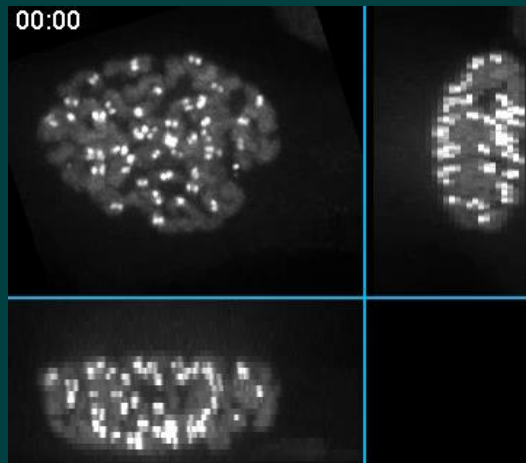
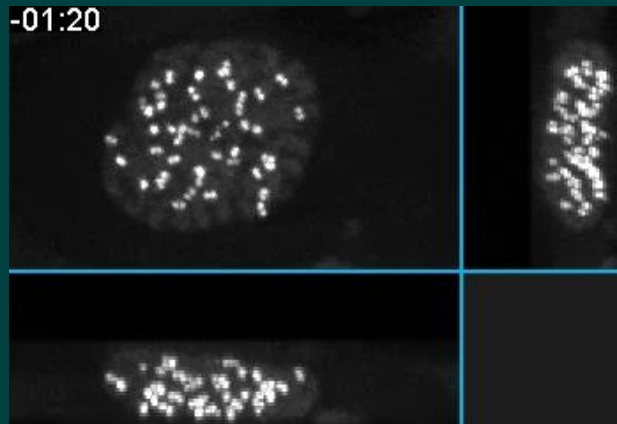


How do microtubules (kinetochore fibers) find chromosomes (kinetochores): search and capture hypothesis and what makes it work



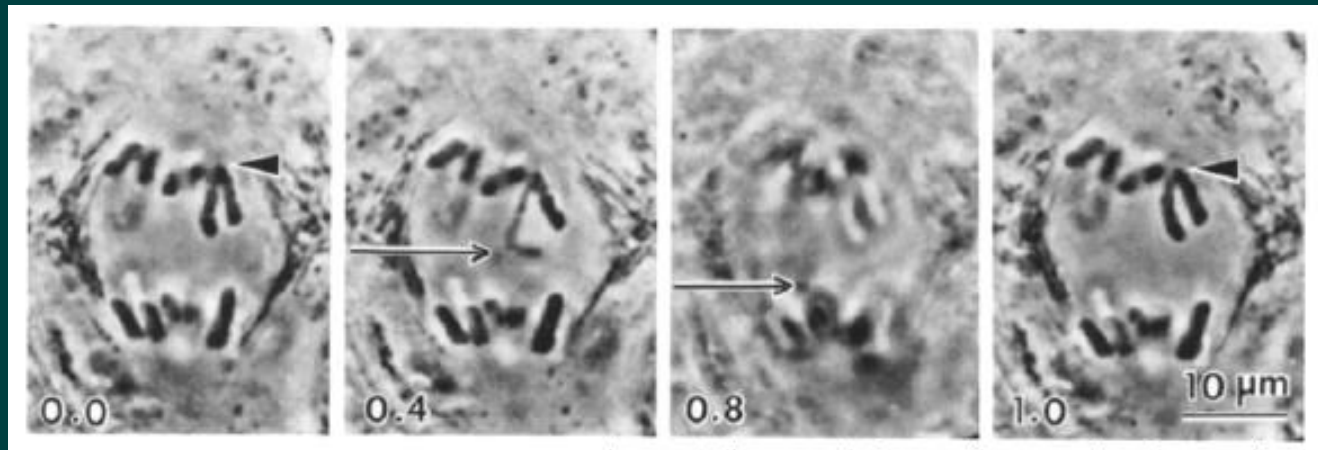
Heald and Khodjakov, 2015

Chromosome ring at the spindle periphery helps search and capture



Magidson et al., 2011

Nanonewton forces in mitotic spindle

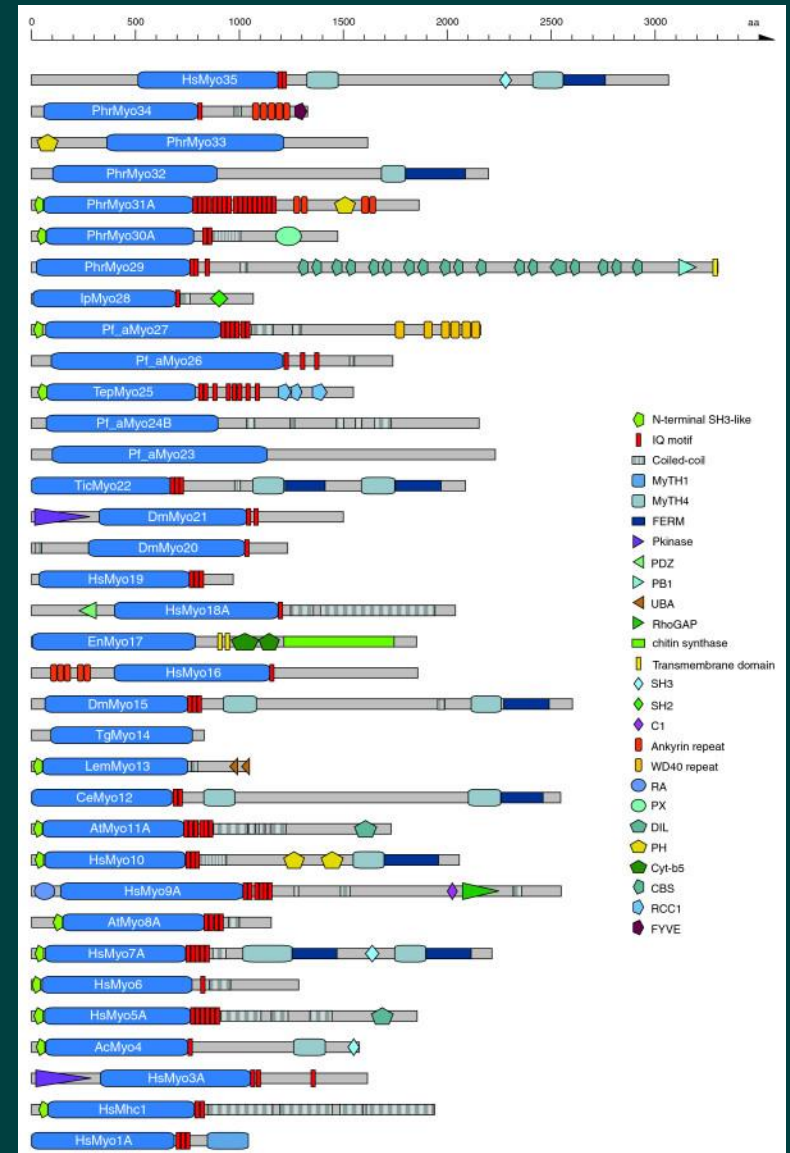
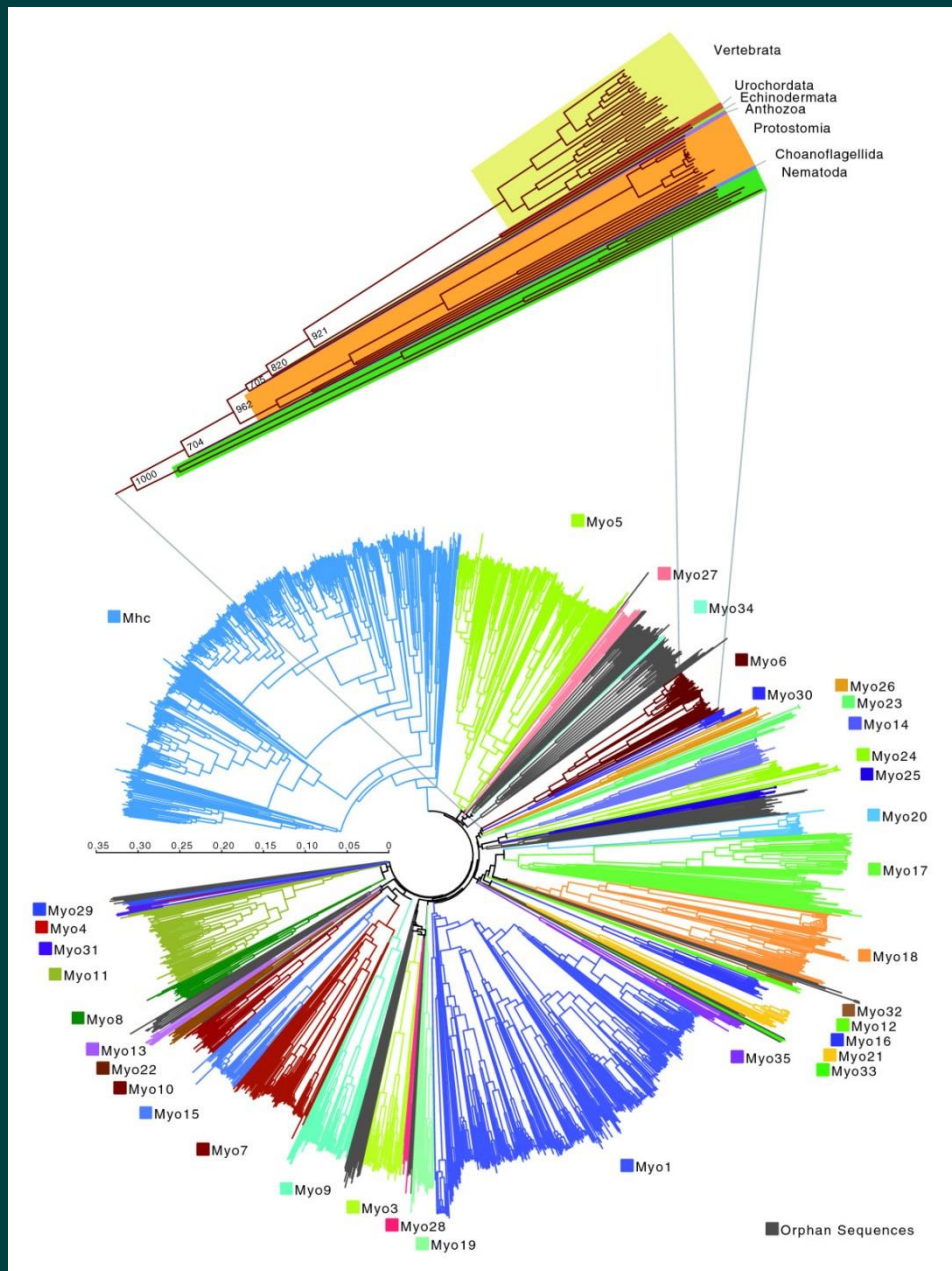


microneedle in grasshopper spermatocytes
Nicklas, J. Cell Biol., 1983

100 nN, but 1 nN is enough to overcome viscous resistance

[illegible]

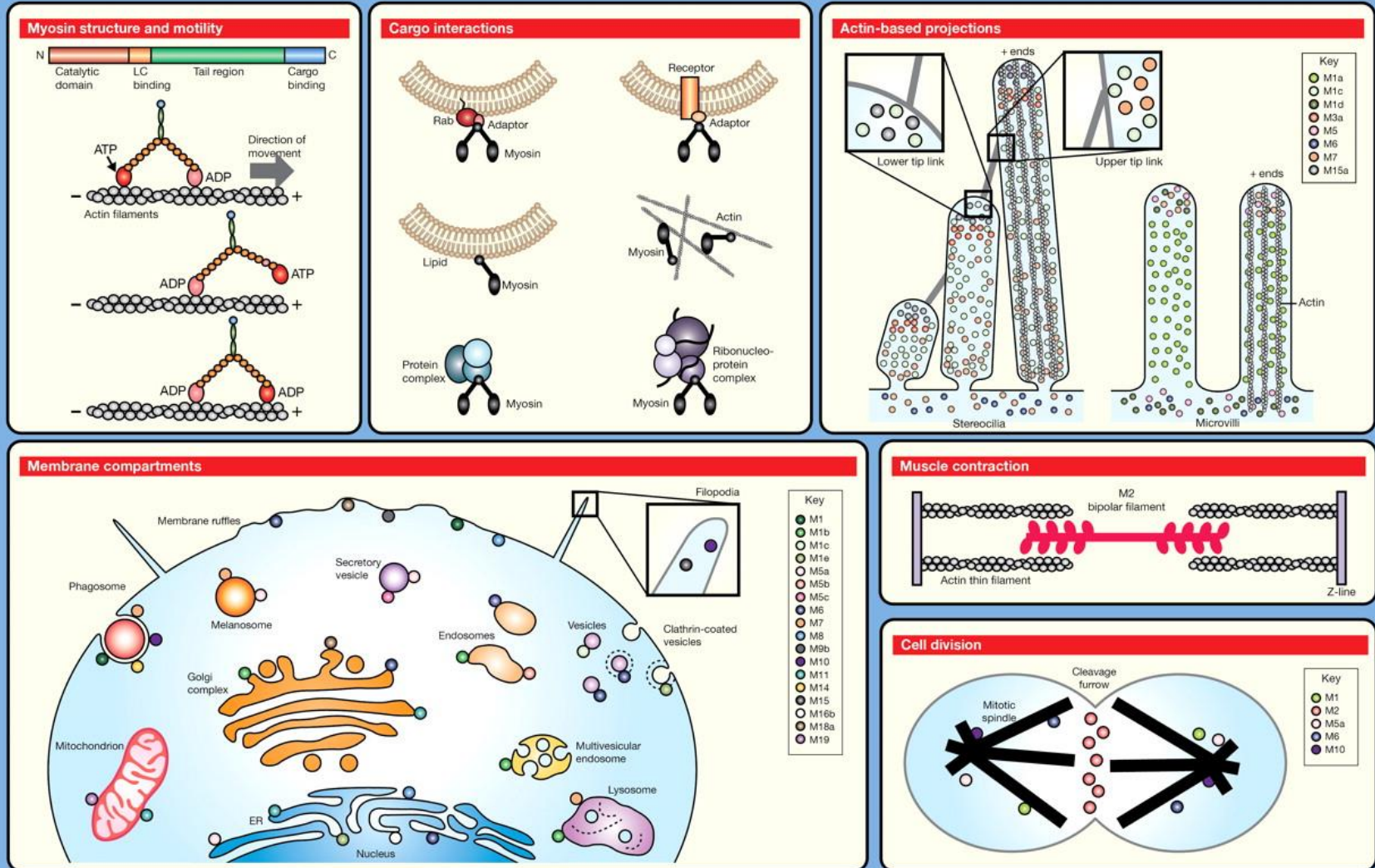
Filogenetic tree of myosin superfamily



Summary of myosin functions

The Myosin Superfamily at a Glance

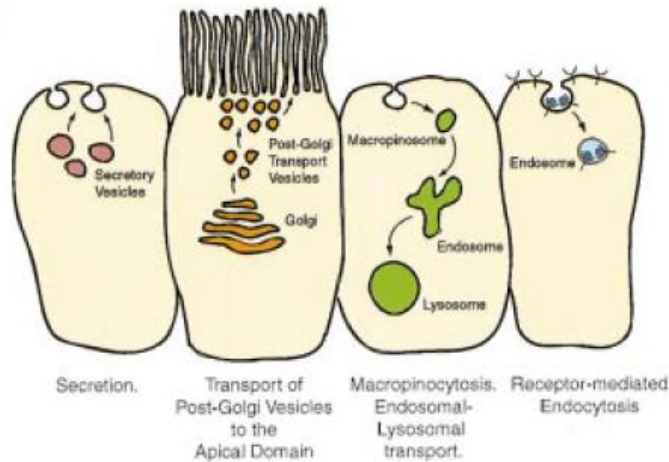
M. Amanda Hartman and James A. Spudis



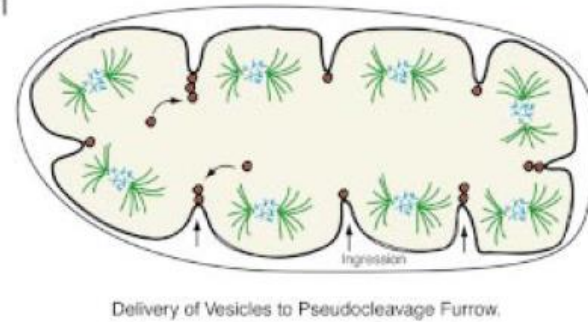
Abbreviations: LC, light chain; M, myosin; + ends, actin plus ends.

Unconventional myosins in membrane traffic

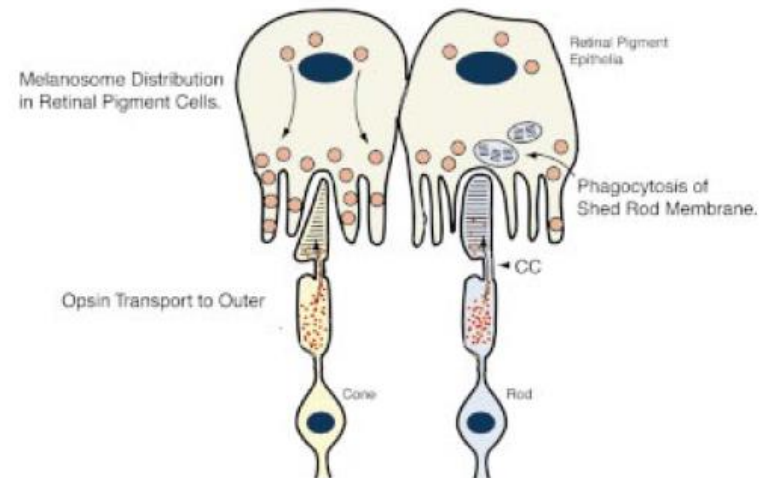
Myosin I



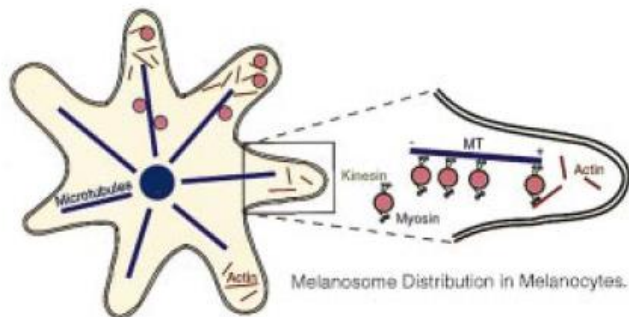
Myosin VI



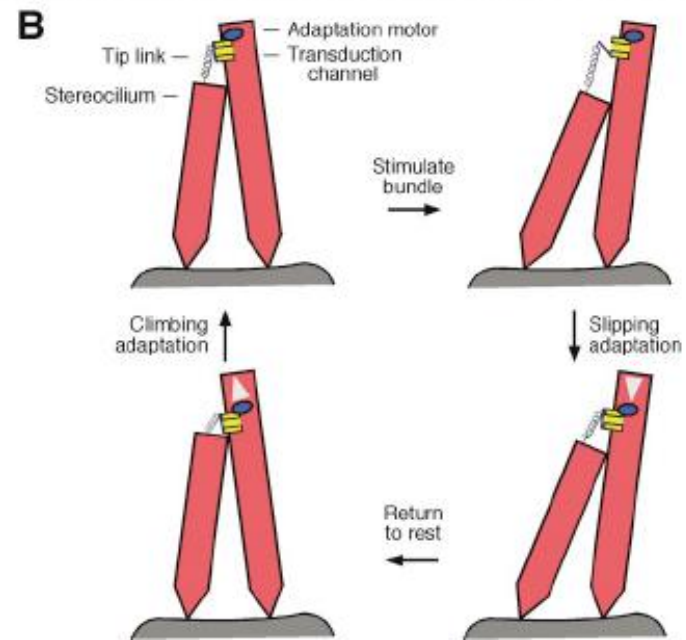
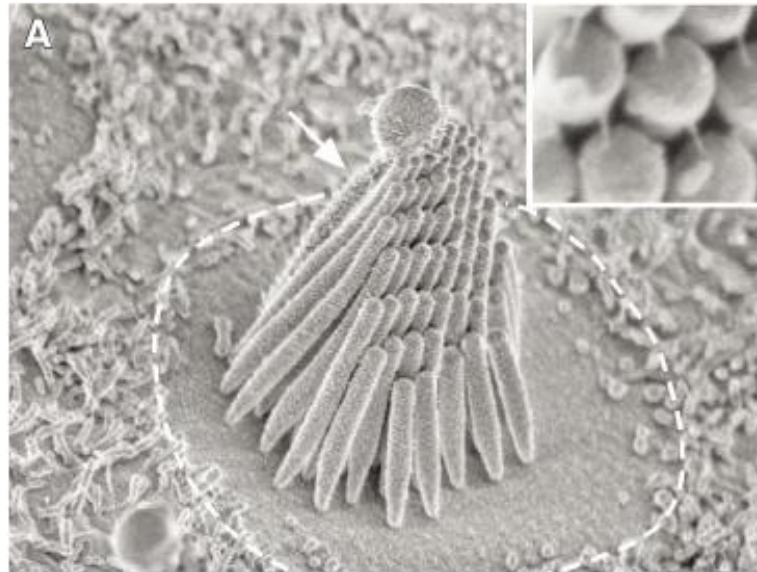
Myosin VII



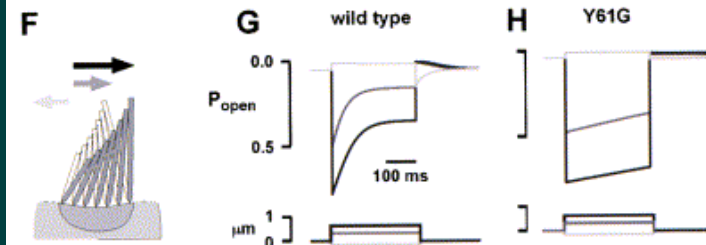
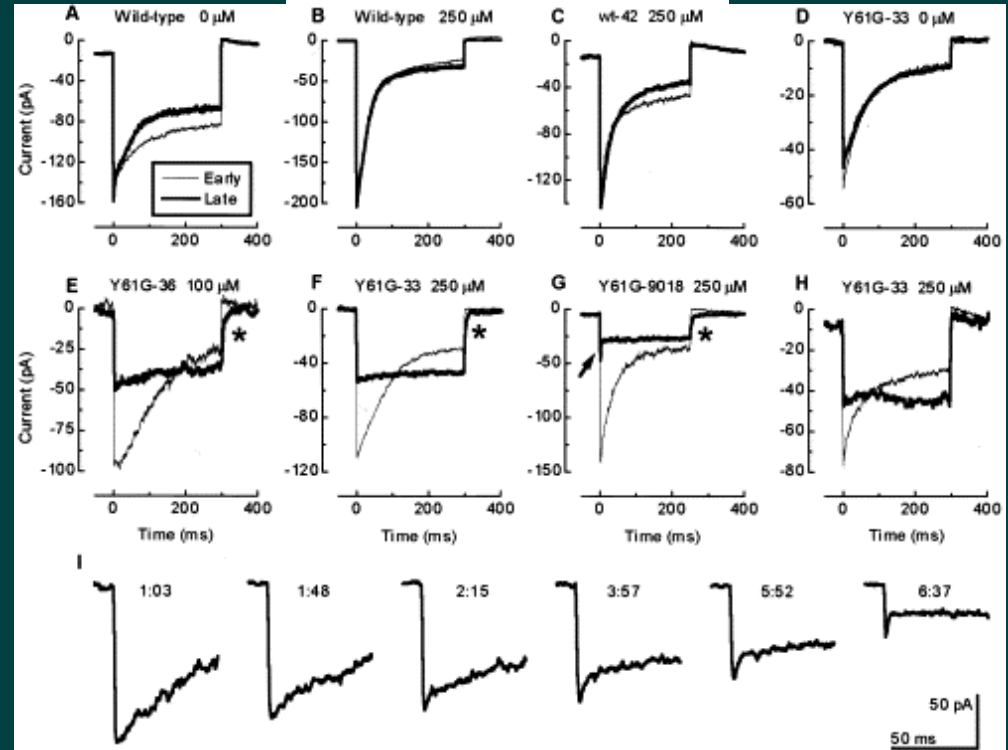
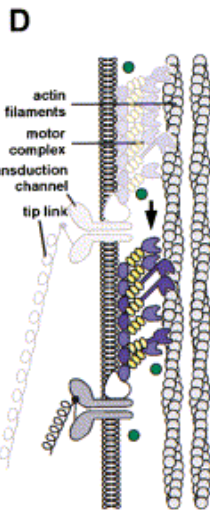
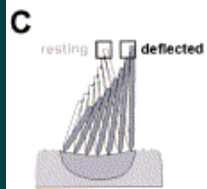
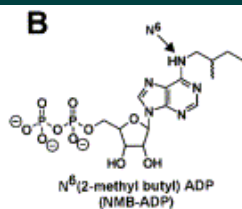
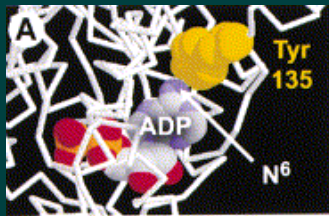
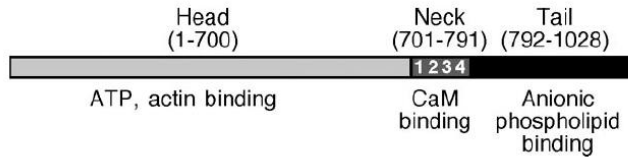
Myosin V



Adaptation motor



Myosin 1c in adaptation by hair cells



Holt et al., Cell, 2002

myosin II in the muscle

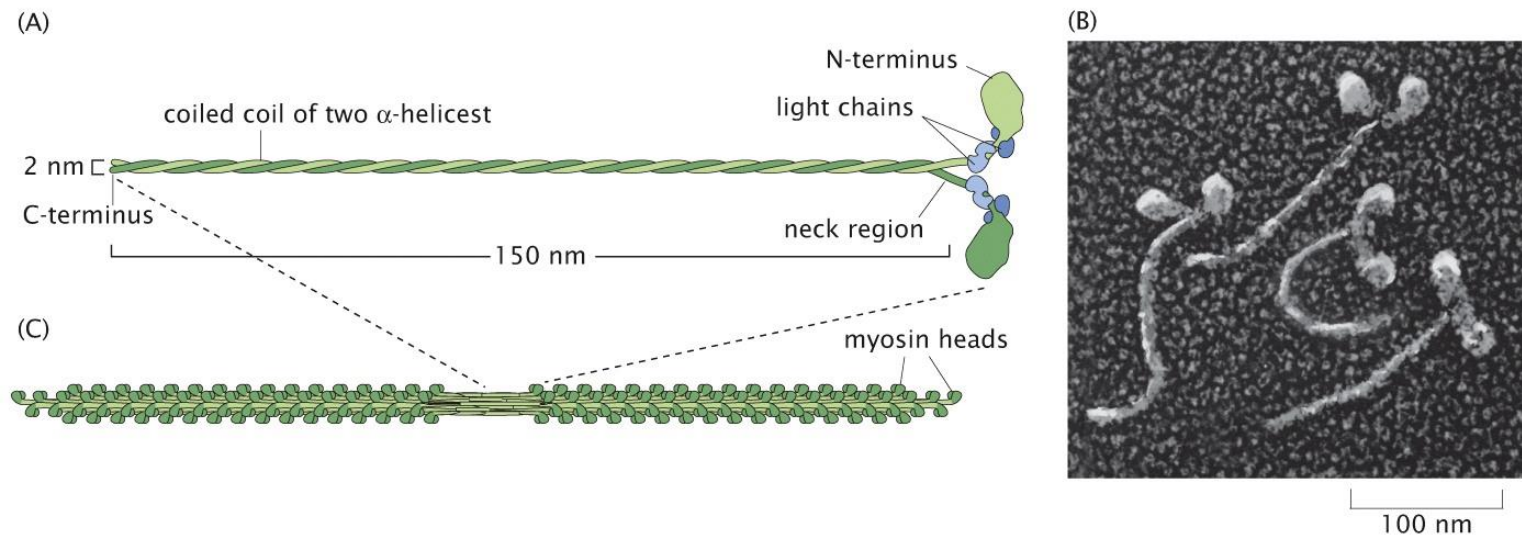


Figure 16.6 Physical Biology of the Cell, 2ed. (© Garland Science 2013)

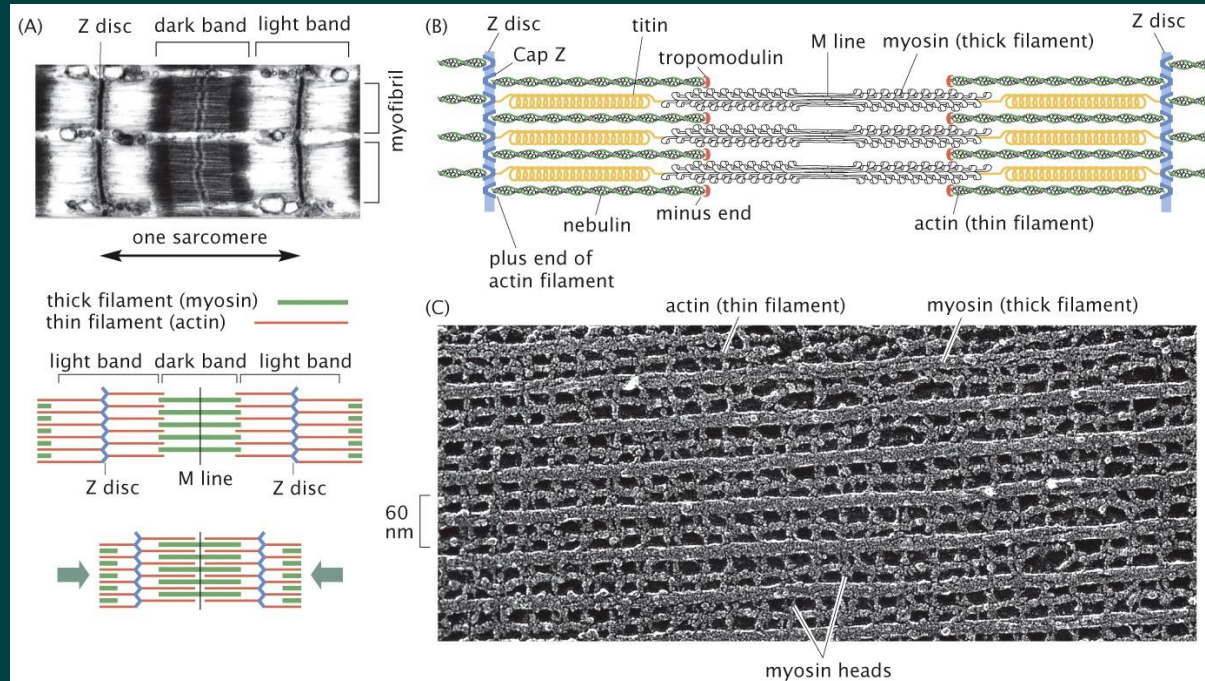
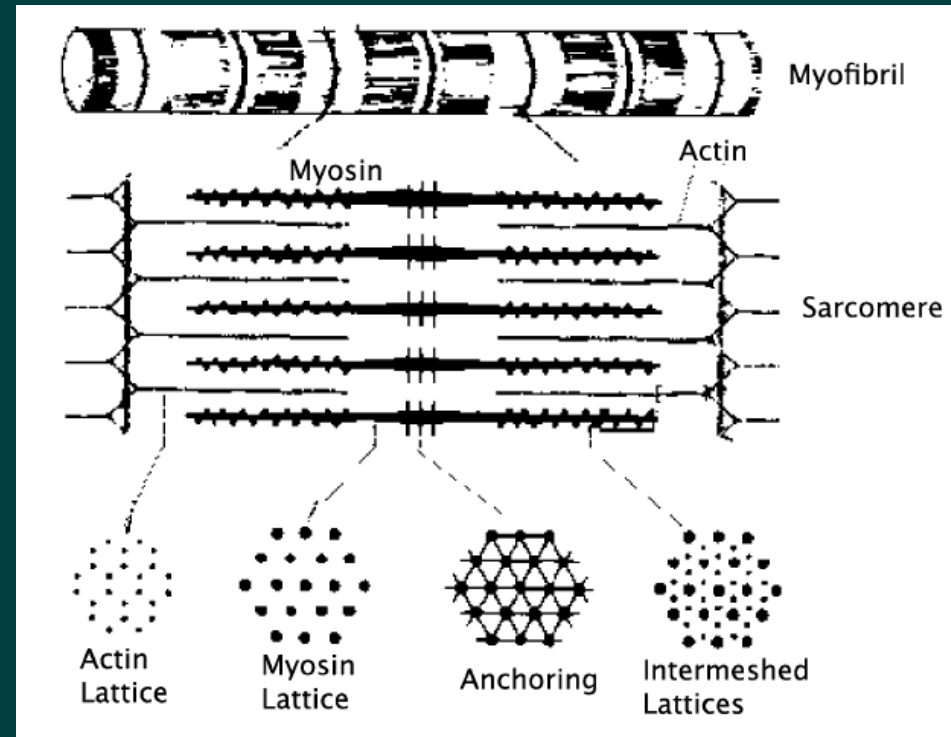
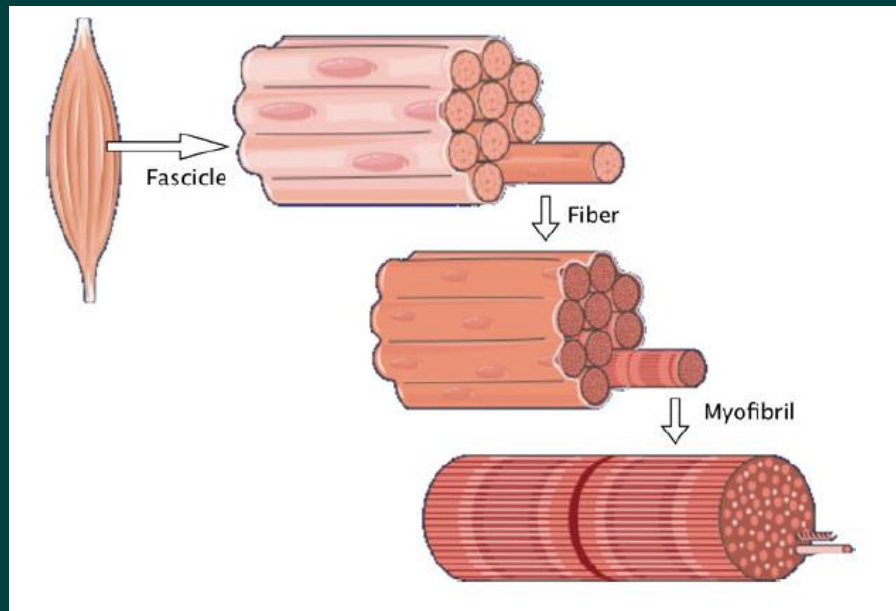


Figure 16.7 Physical Biology of the Cell, 2ed. (© Garland Science 2013)

Regular organization of contractile units in skeletal muscle



Williams, 2011

Muscle contraction is activated through release of Ca^{2+} from sarcoplasmic reticulum leading to a conformational change in troponin-tropomyosin complex making actin filaments accessible to myosin

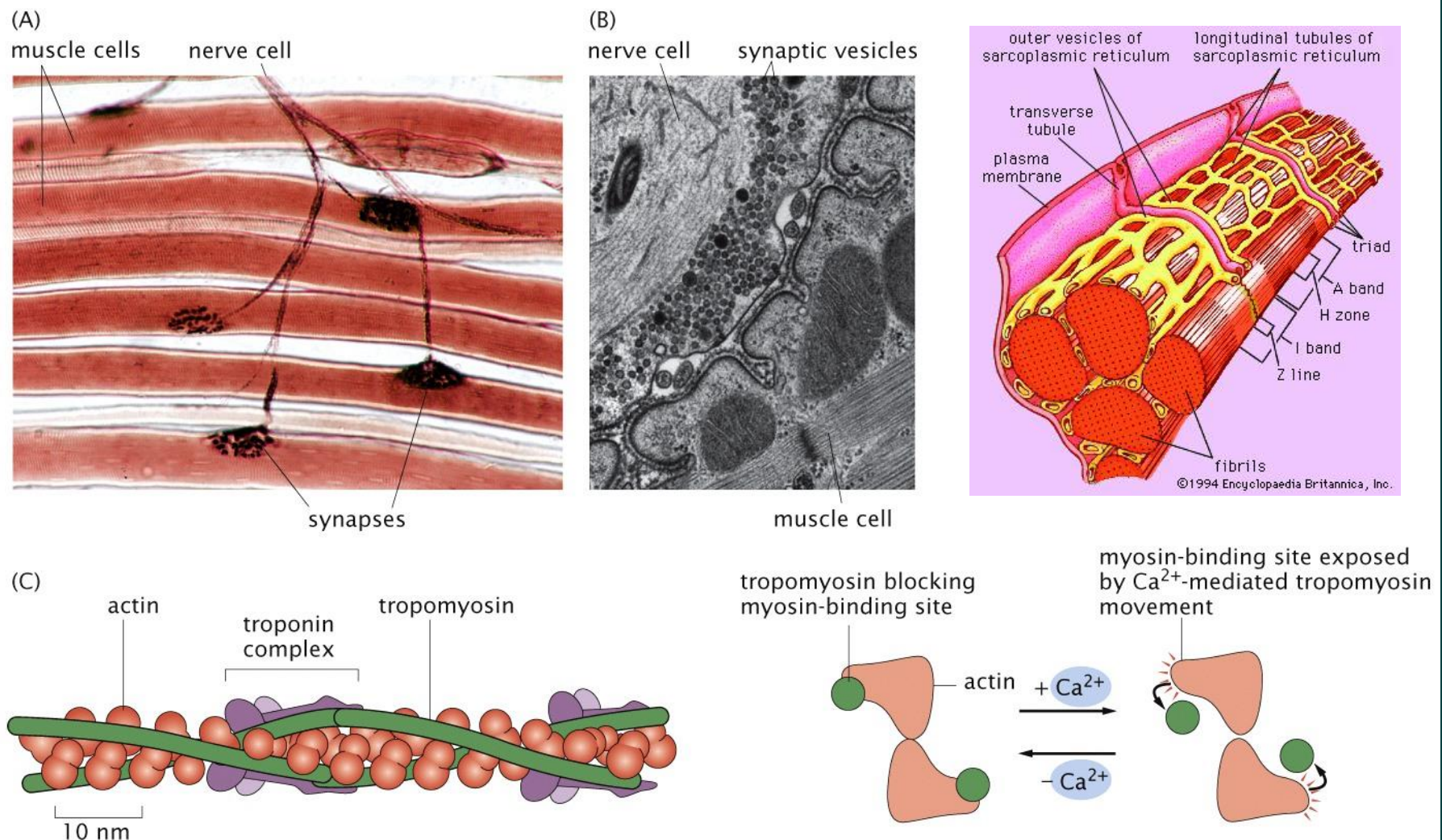
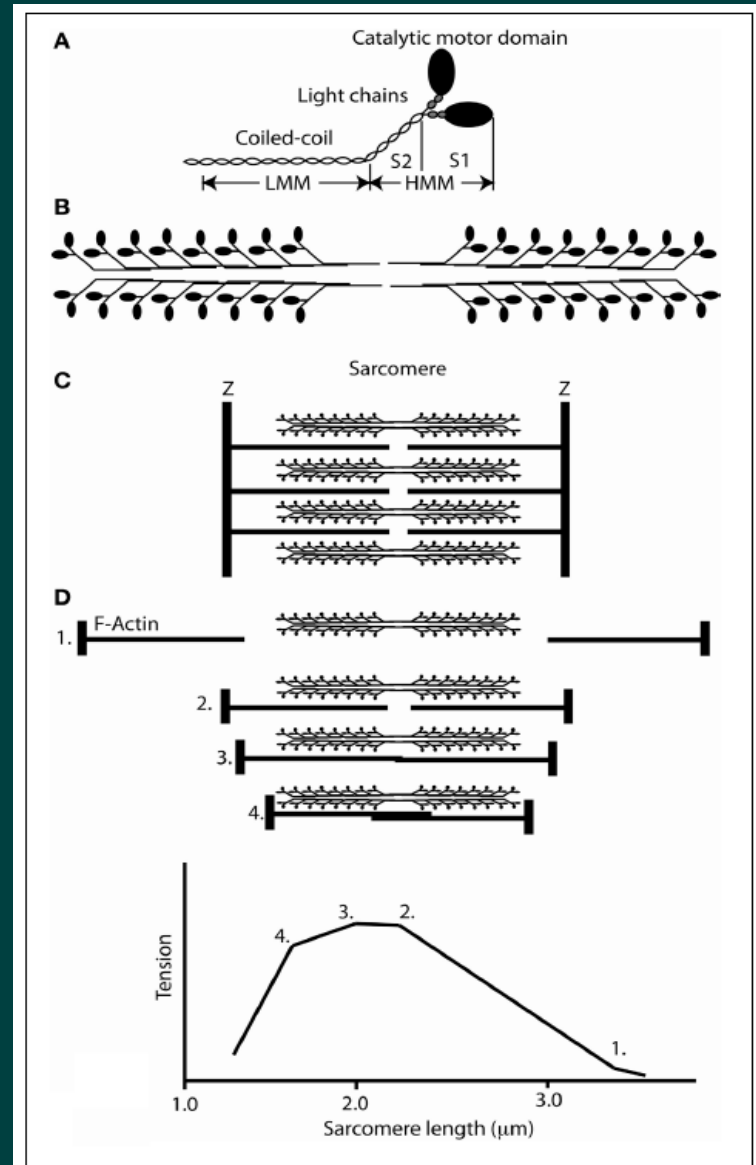
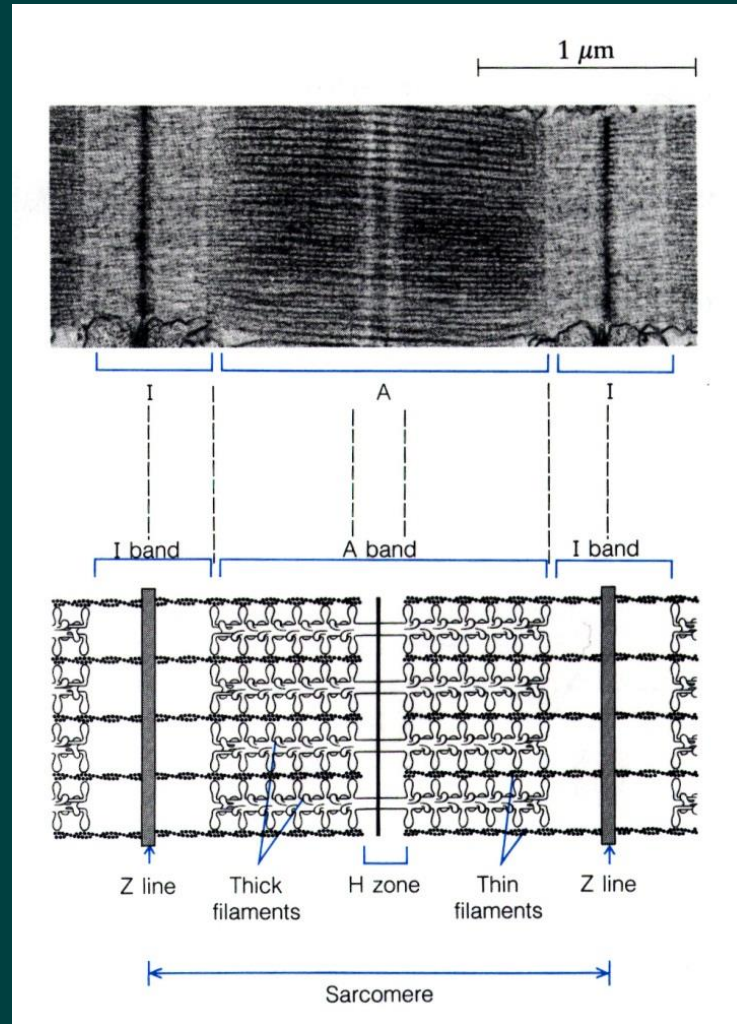


Figure 16.39 Physical Biology of the Cell, 2ed. (© Garland Science 2013)

Muscle contraction: sliding filament hypothesis

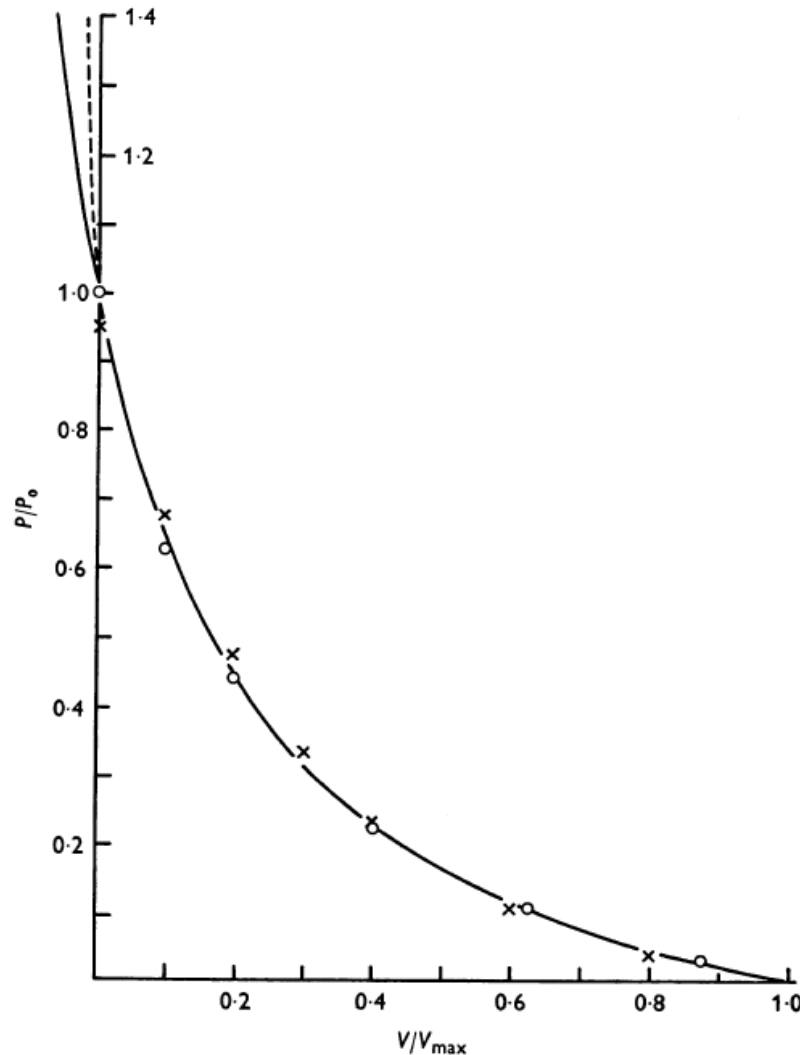
Huxley and Hanson, Huxley and Niedergerke, 1954



Linear force-velocity relation (an abstraction often used in modeling)

$$F = F_0 (1 - V/V_{\max})$$

Force-velocity relationship for skeletal muscle is hyperbolic (Hill, 1938)



Text-fig. 1. Force-velocity relation. The degree of curvature of the relation varies between different preparations, and with temperature. Continuous line: Hill's (1938) relation,

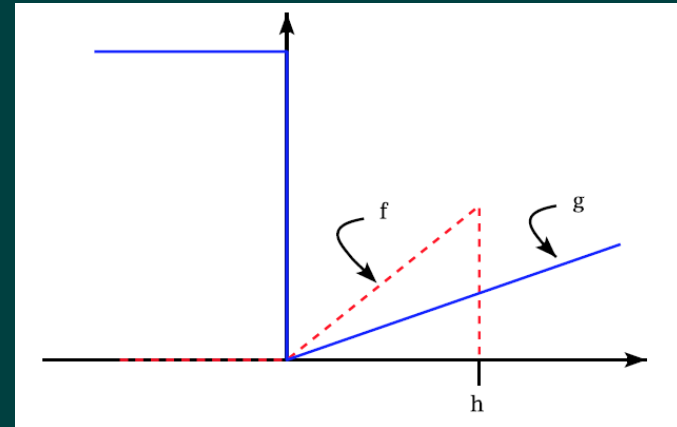
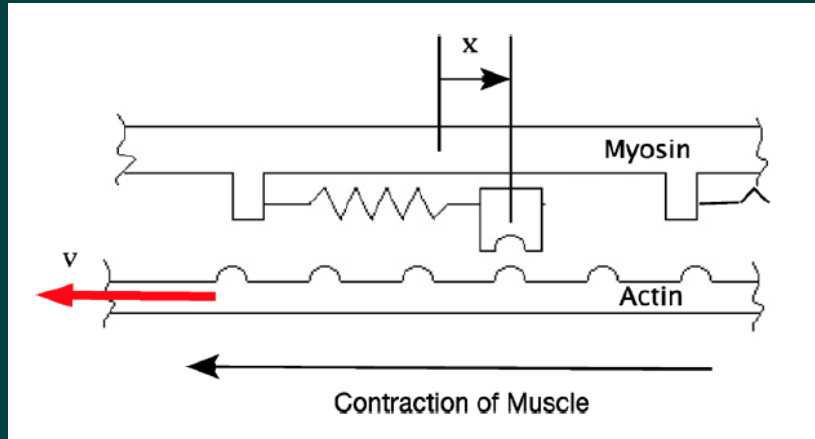
$$P/P_0 = (1 - V/V_{max}) / (1 + (P_0/a)V/V_{max}),$$

with $P_0/a = 4$, appropriate for frog muscle at 0°C .

$$(F+a)(V+b)=b(F_0+a)$$

A.F. Huxley, 1974

A.F. Huxley's model of muscle contraction: force and the probability to attach and detach depend on the cross-bridge deformation



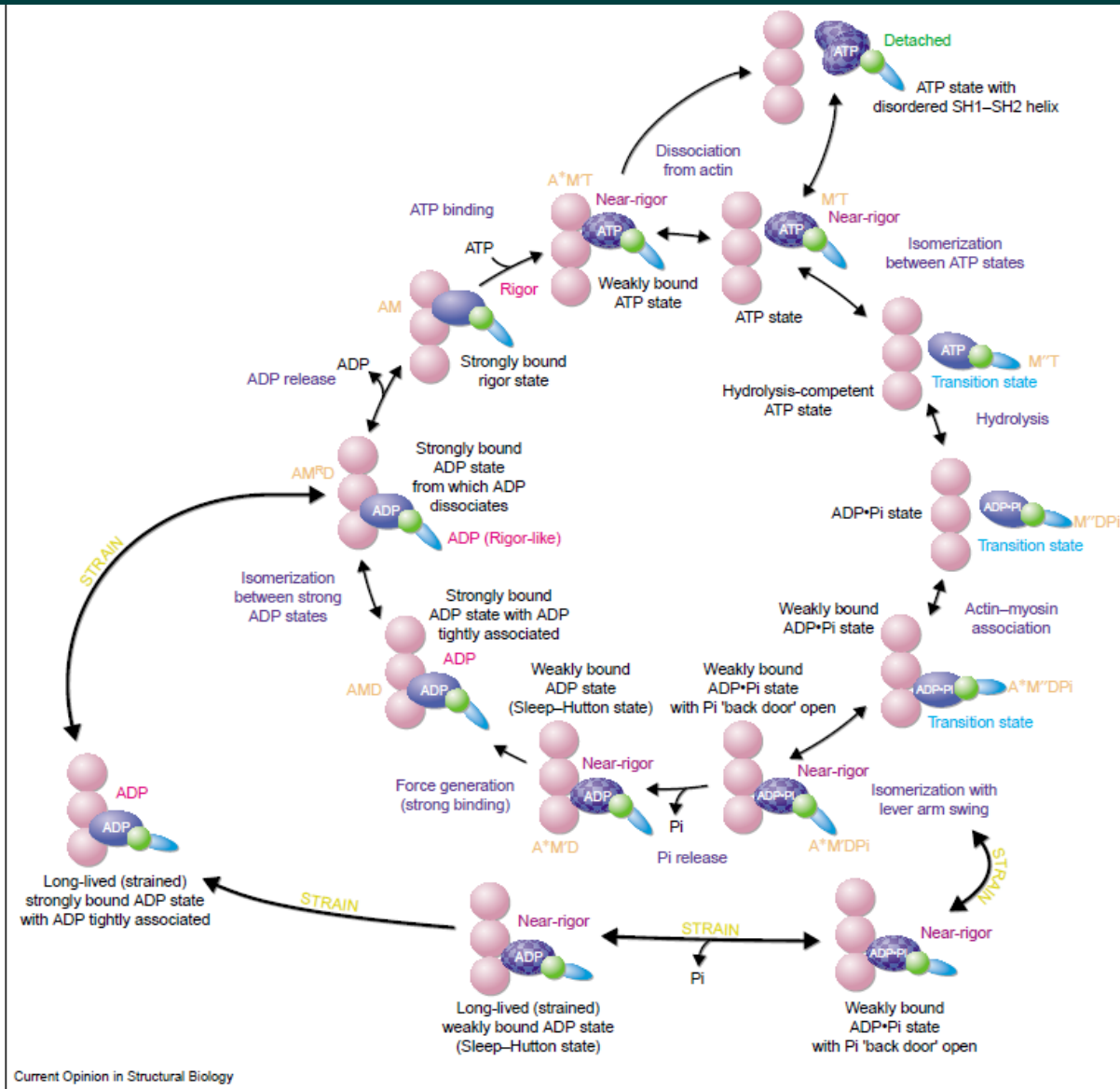
$$\begin{aligned} \frac{d}{dt}N((a, b), t) &= \int_a^b \frac{\partial n}{\partial t}(x, t) dx \\ &= \int_a^b (F(x, t) - G(x, t)) dx + v(t)[n(b, t) - n(a, t)]. \end{aligned}$$

$$\frac{\partial n(x, t)}{\partial t} = F(x, t) - G(x, t) + v(t) \left(\frac{\partial n(x, t)}{\partial x} \right)$$

$$P(t) = \int_{-X}^X \mathcal{E}(x) n(x, t) dx$$

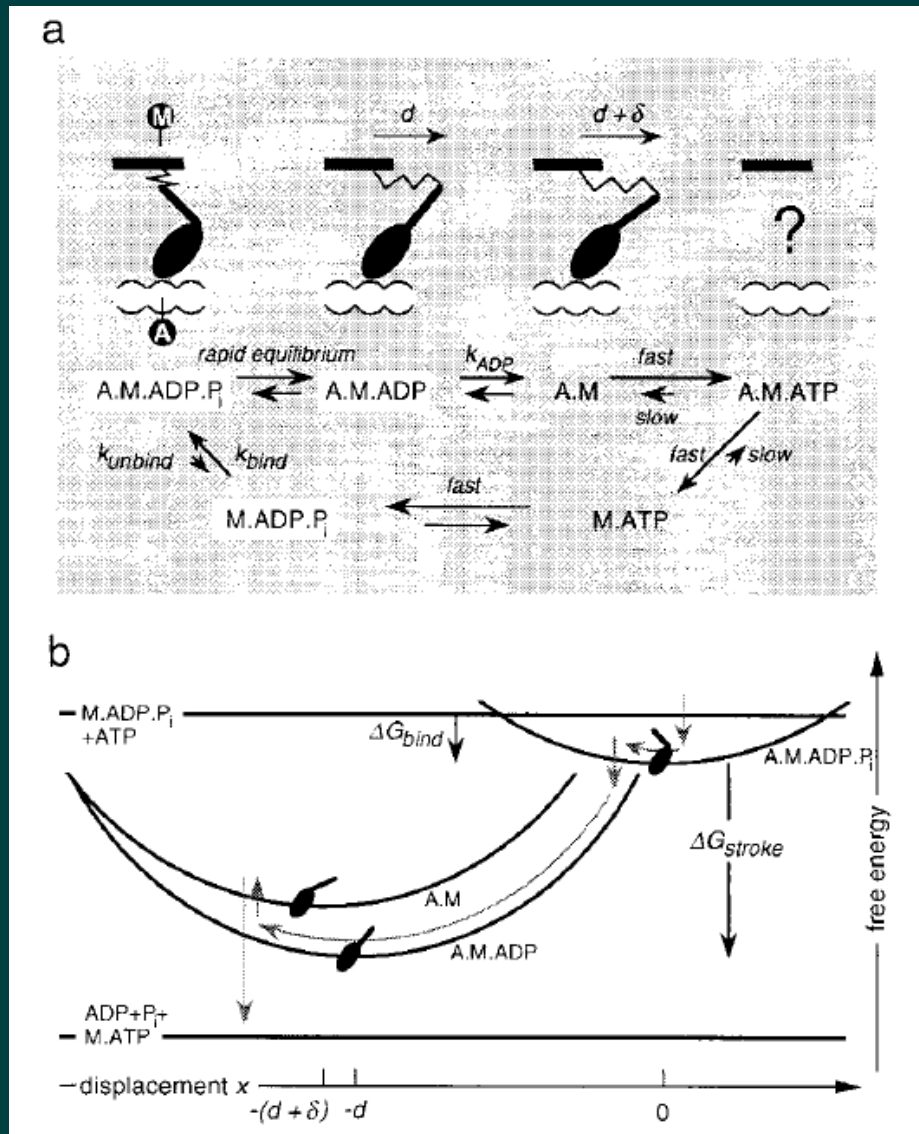
Williams, 2011

Huxley's model does not consider the complexity of myosin cycle of ATP-hydrolysis and power stroke

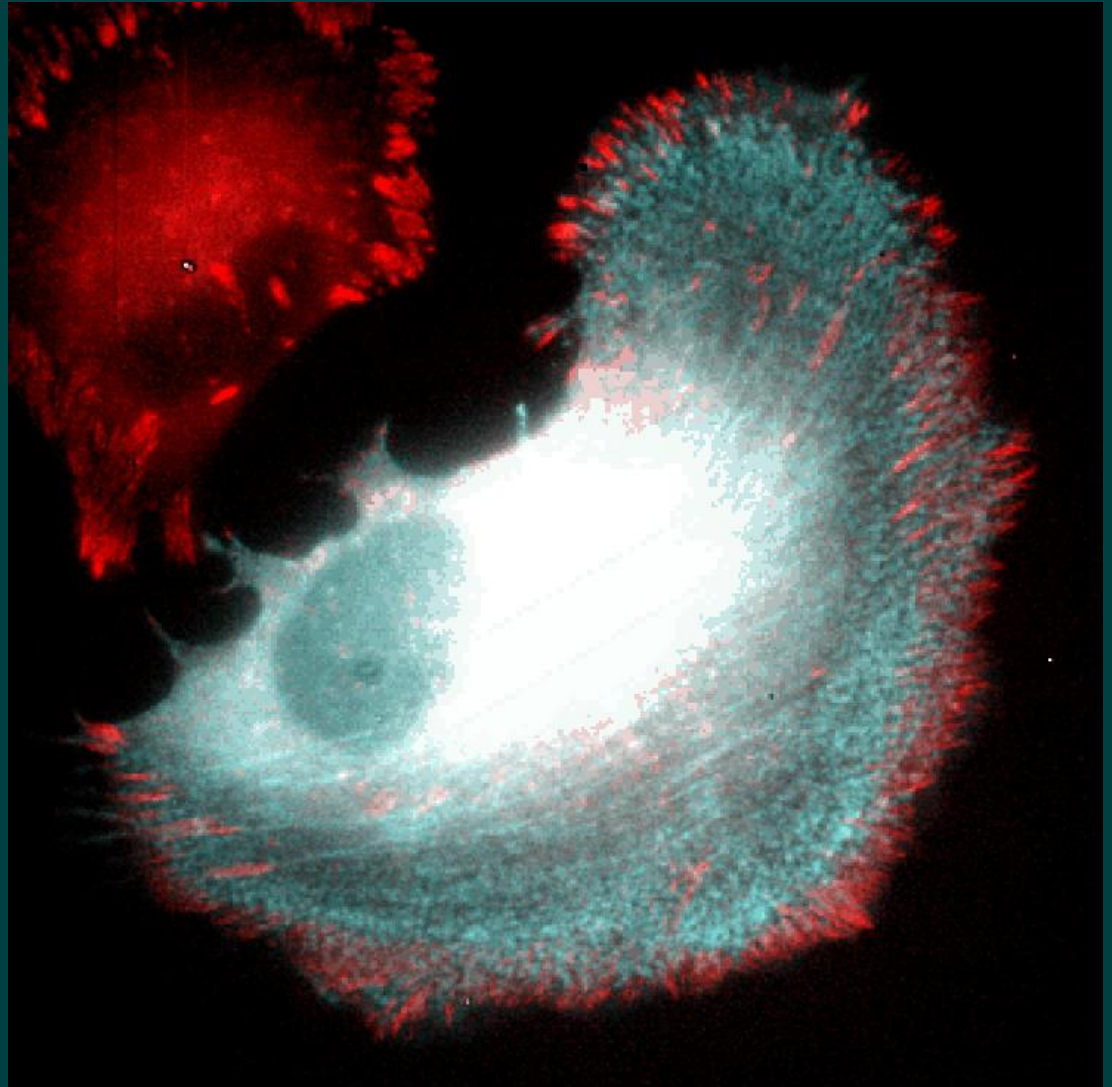
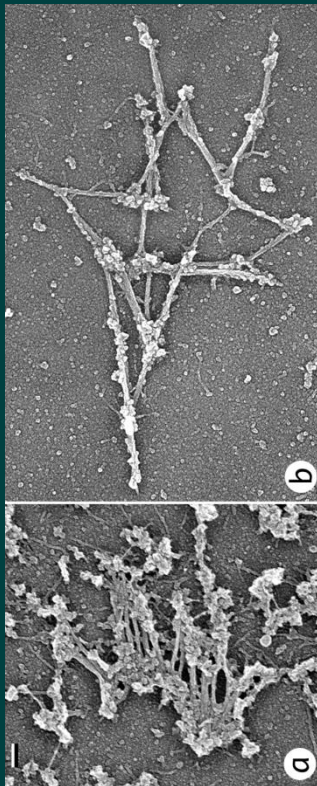
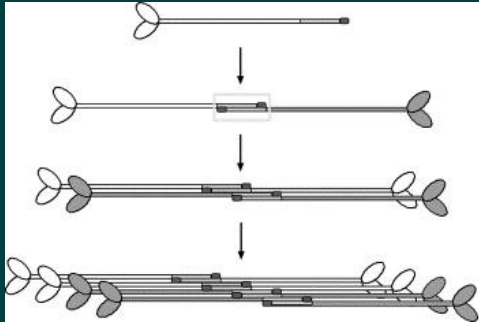
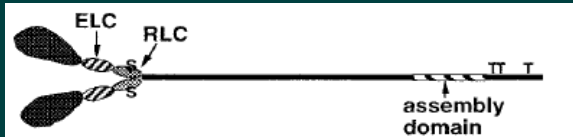


Houdusse and Sweeney, 2001

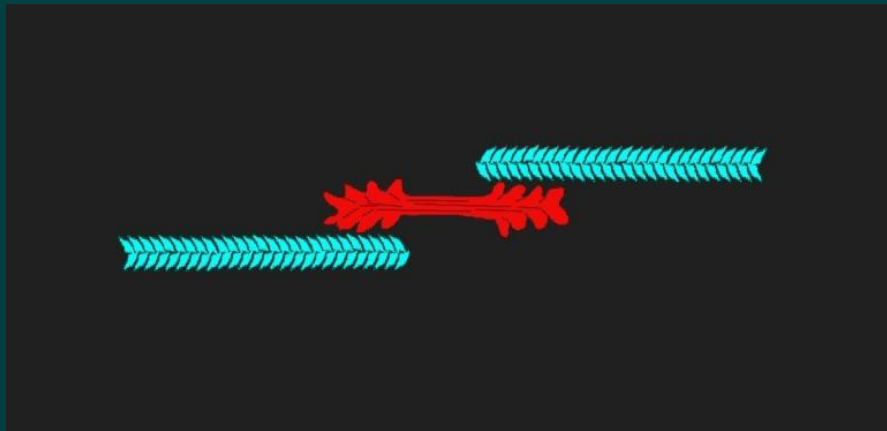
Molecular model taking into account myosin cycle



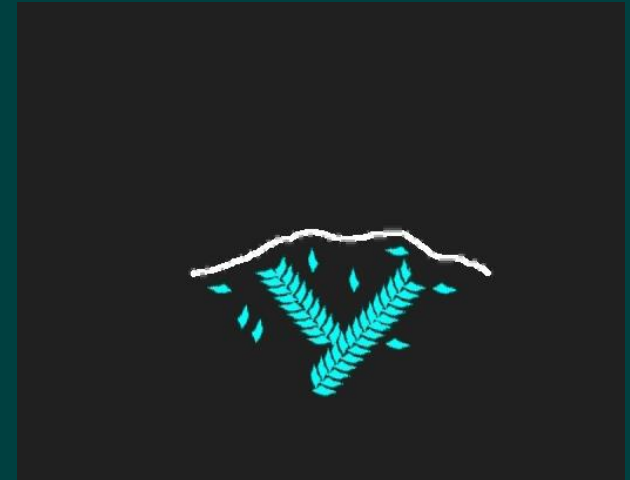
Myosin II in non-muscle cells



proposed role for myosin II in crawling cell motility

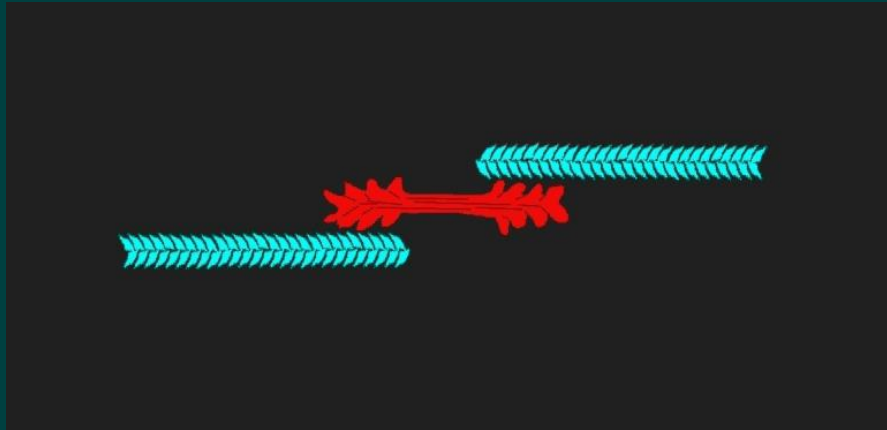


actin/myosin contraction
(analogous to muscle)

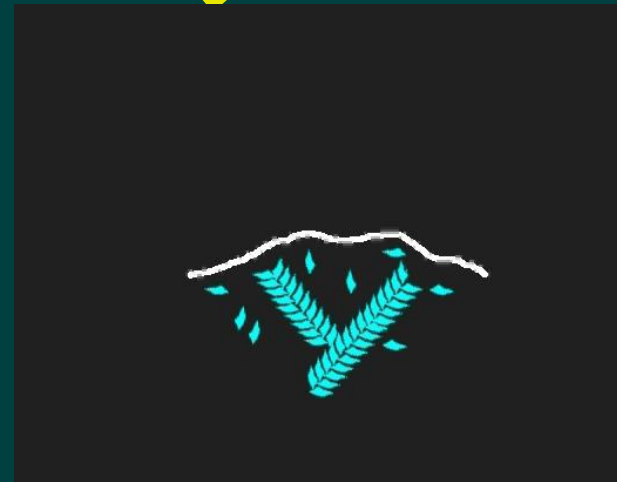


actin assembly

proposed role for myosin II in crawling motility



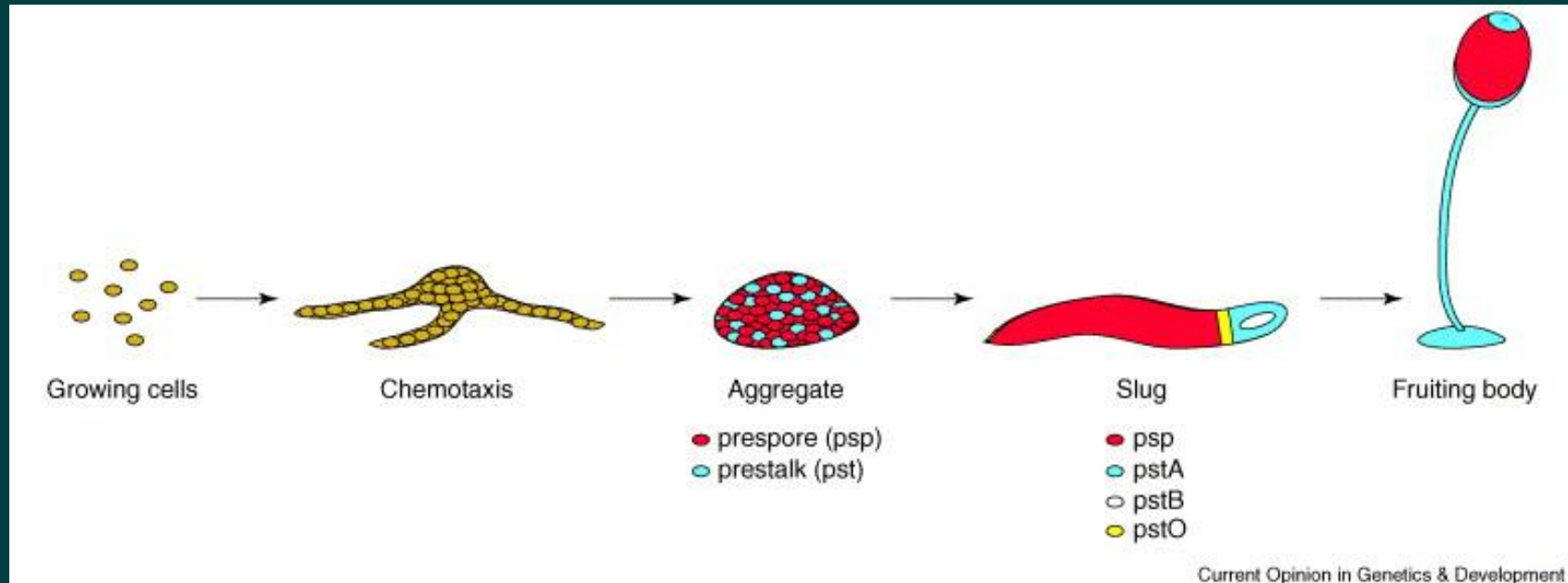
actin/myosin contraction



actin assembly

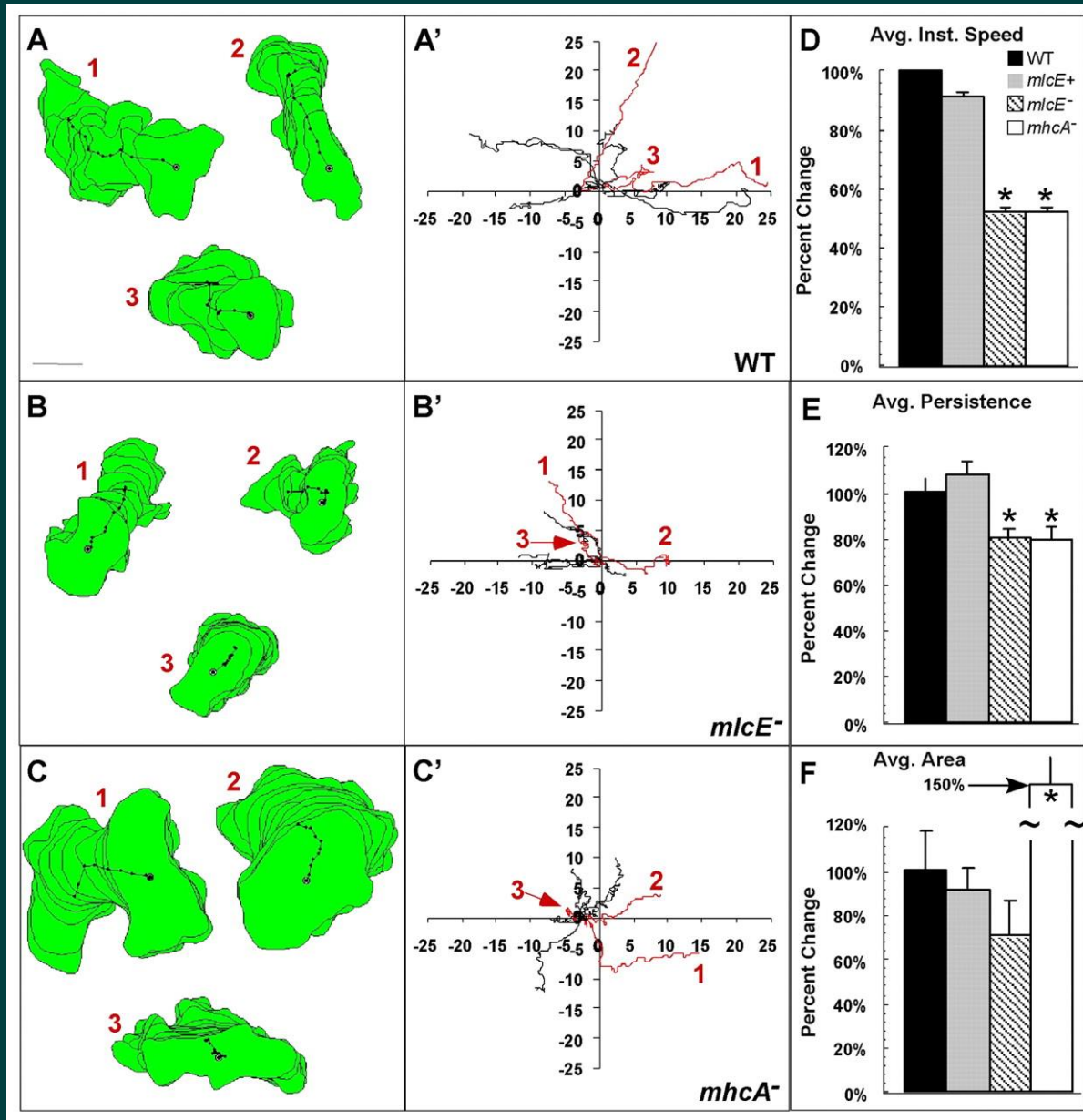
Is myosin II essential for retraction?
What is the contraction mechanism?

Dictyostelium : a model for motility and chemotaxis (accessible to genetic manipulation)



Myosin II mutants exhibit motility and chemotaxis, but do not form fruiting bodies

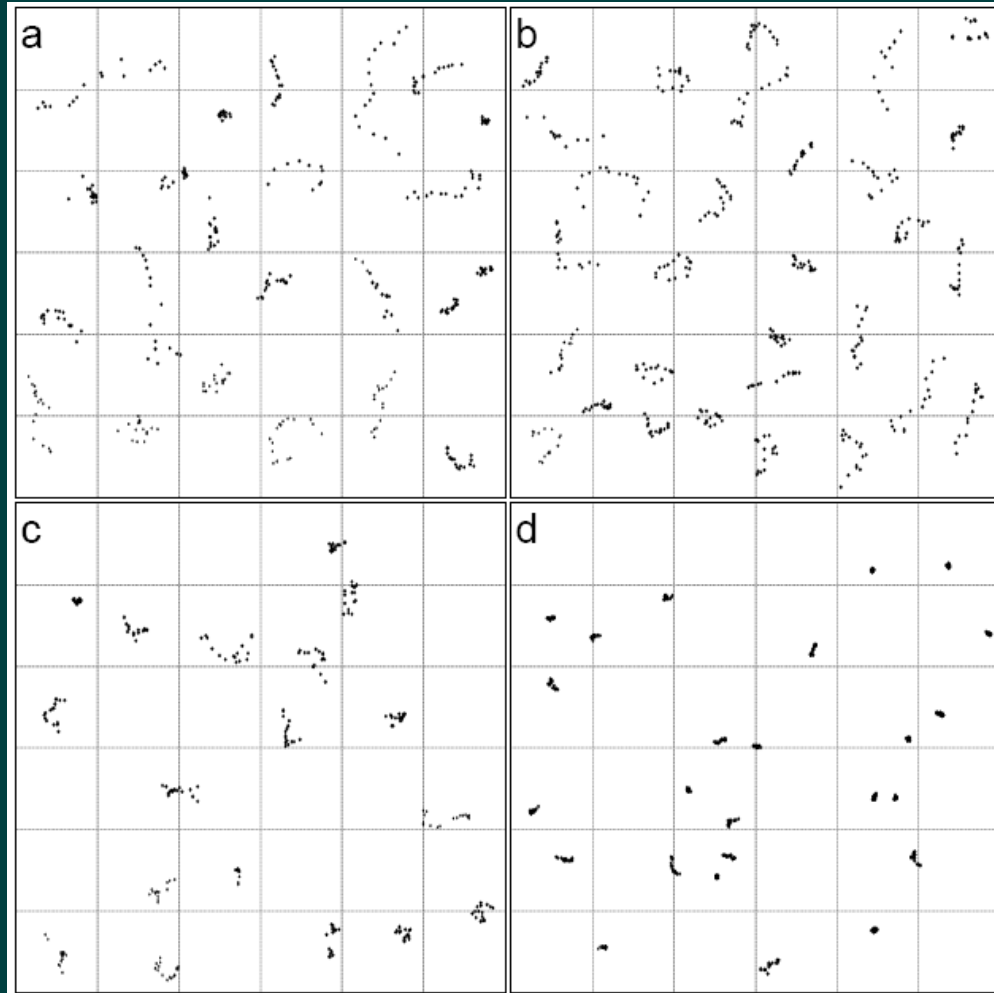
Motility in Dictyostelium myosin II mutants



Lombardi et al.,
J. Cell Sci., 2007

Myosin II-null cells cannot move on strongly adhesive substrates

wild type



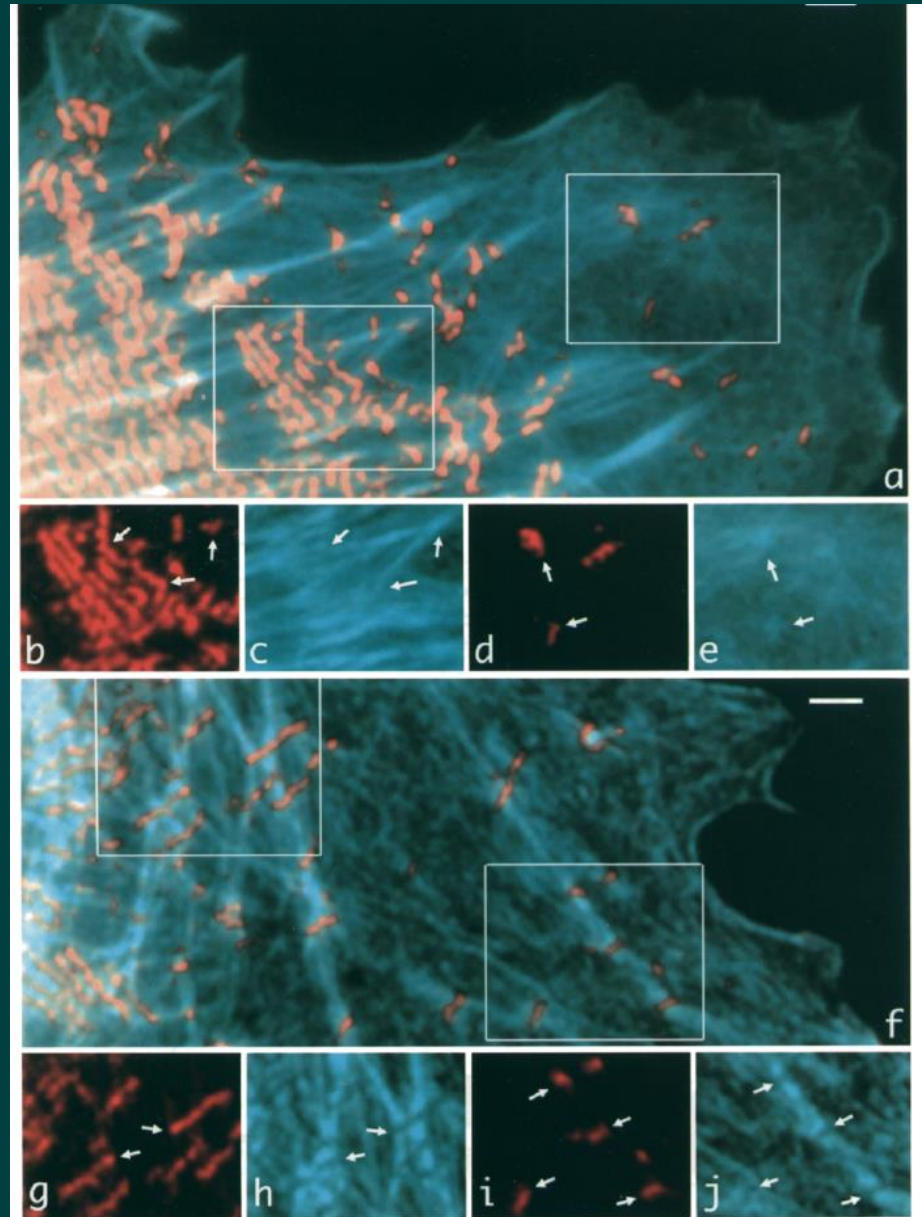
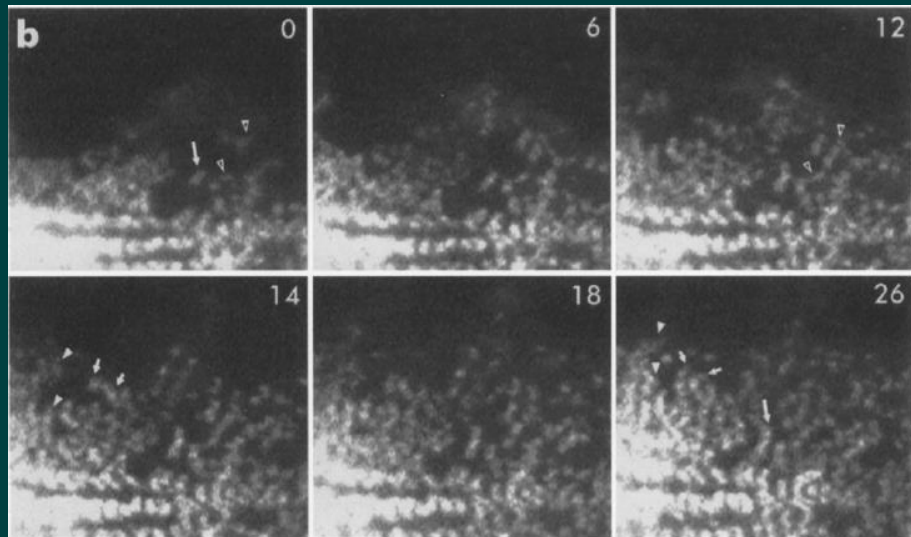
mhcA⁻

uncoated

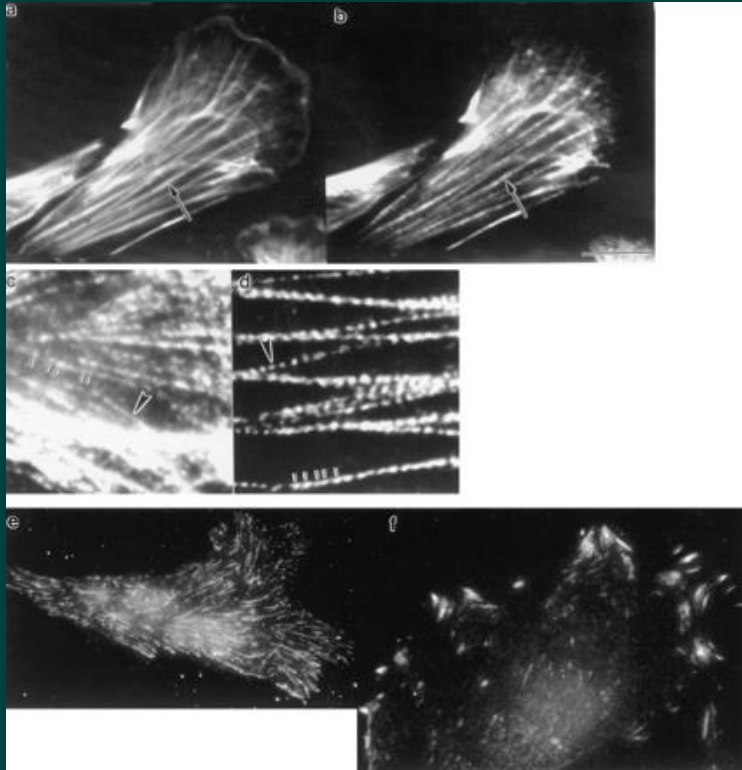
polylysine-coated

Jay et al.,
J.Cell Sci., 1995

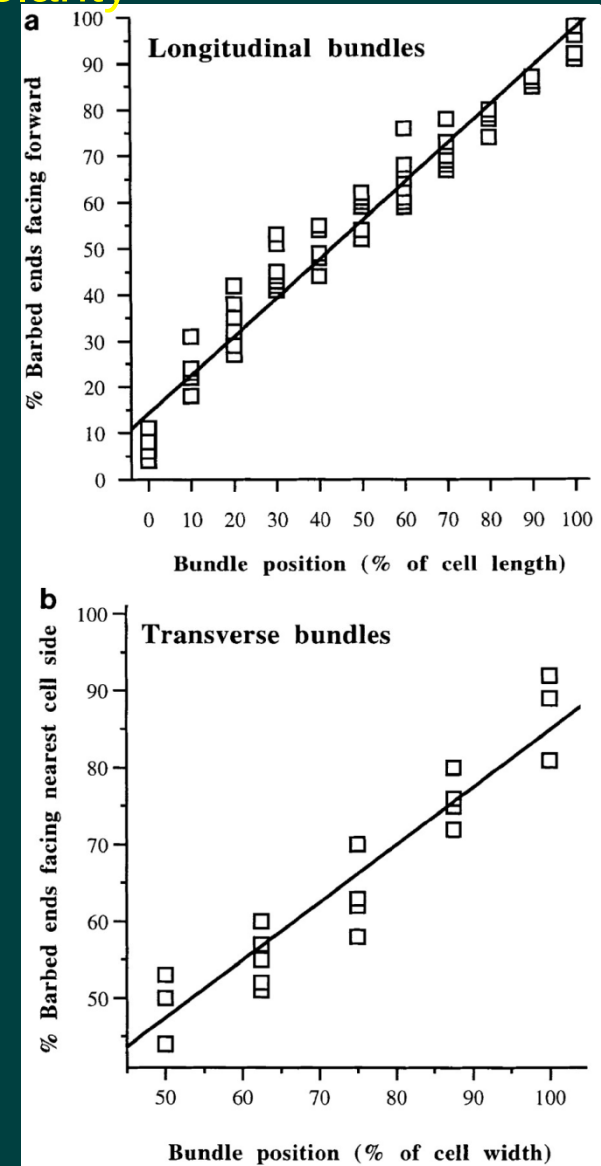
Myosin II organization in non-muscle cells (fibroblasts)



In migrating fibroblasts, actin filament bundles
have irregular myosin II organization
and graded actin filament polarity



Cramer et al., JCB, 1997



How could actomyosin bundles contract in non-muscle cells if there is no regularly alternating actin polarity?

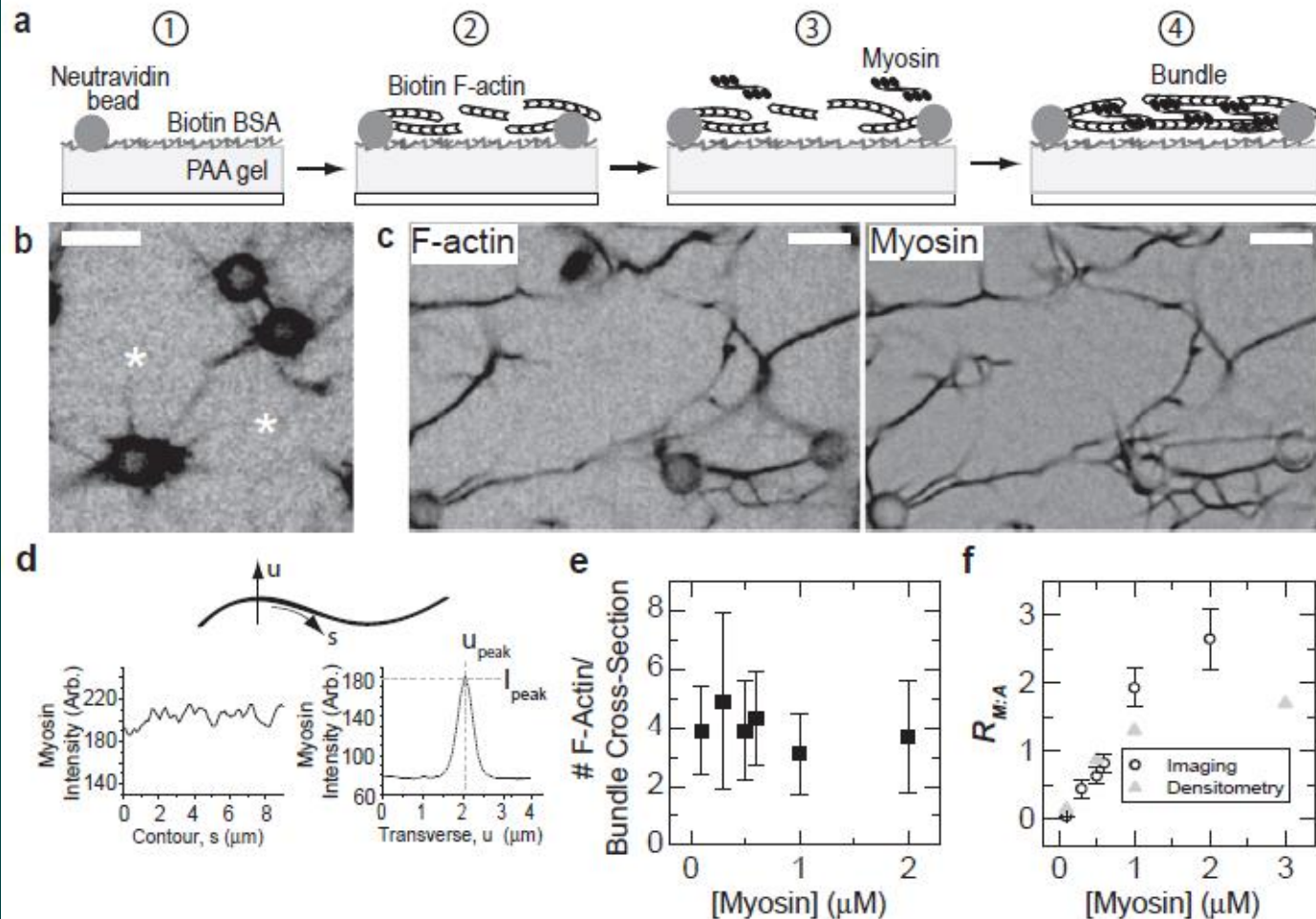
2698

Biophysical Journal Volume 100 June 2011 2698–2705

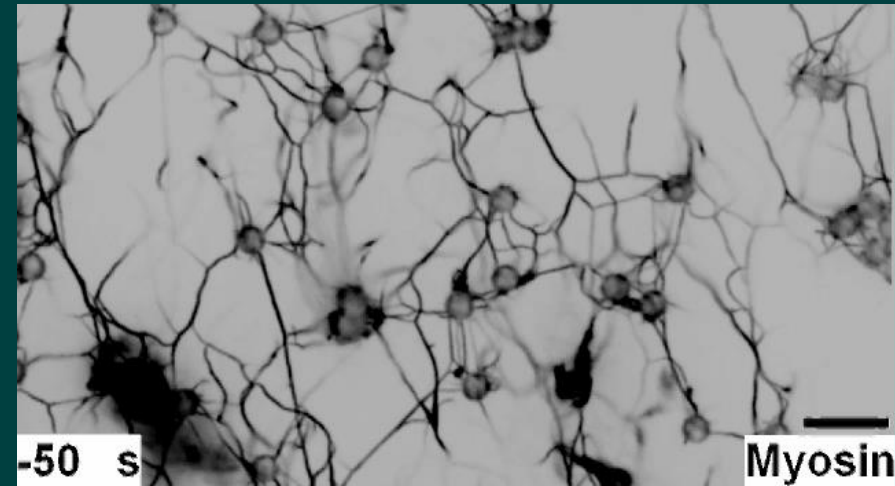
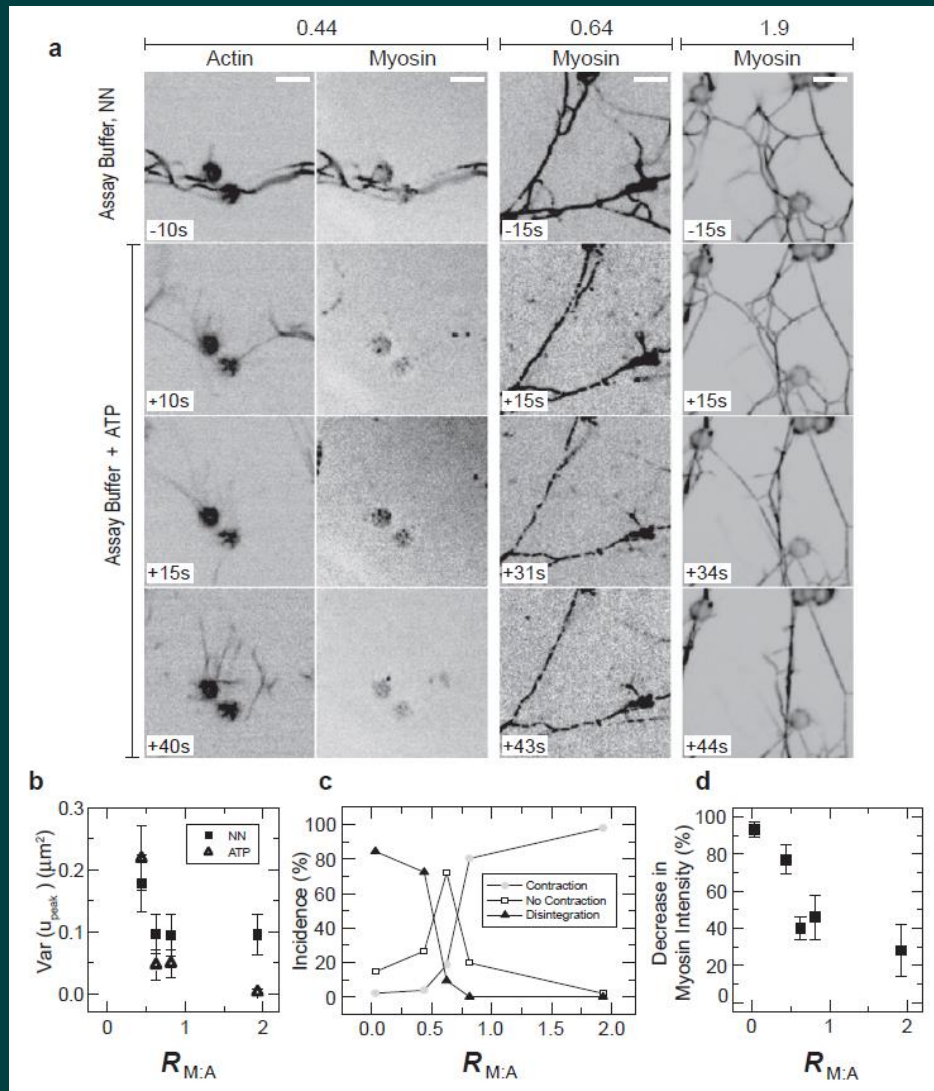
Reconstitution of Contractile Actomyosin Bundles

Todd Thoresen,[†] Martin Lenz,^{†‡} and Margaret L. Gardel^{†‡*}

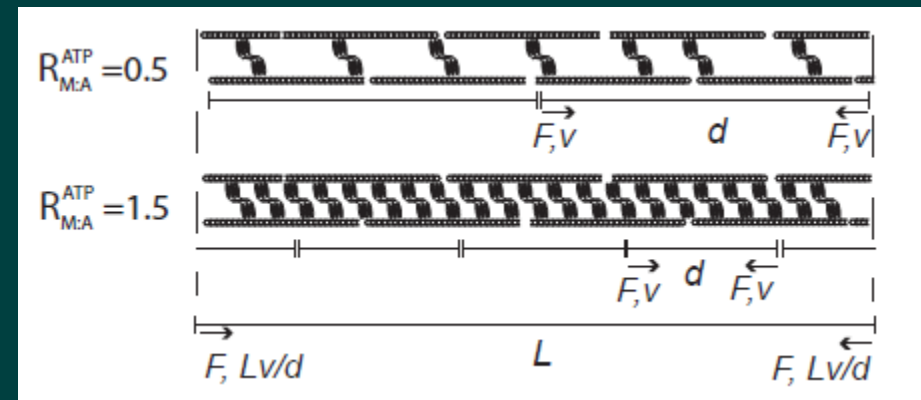
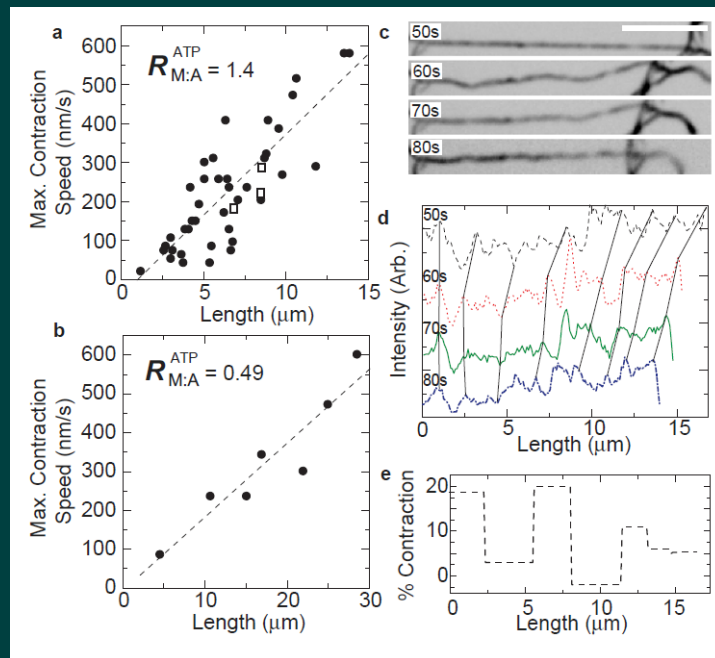
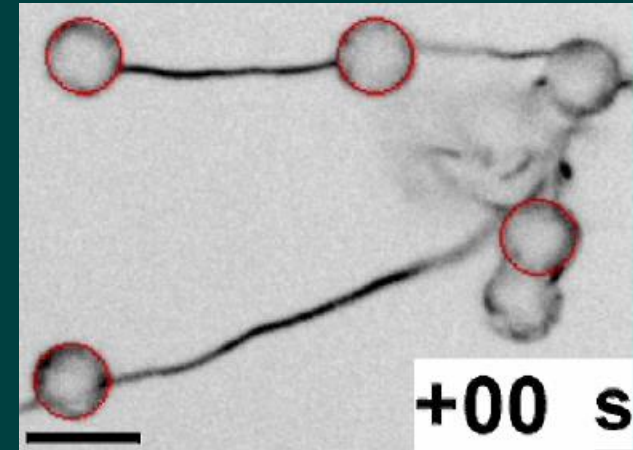
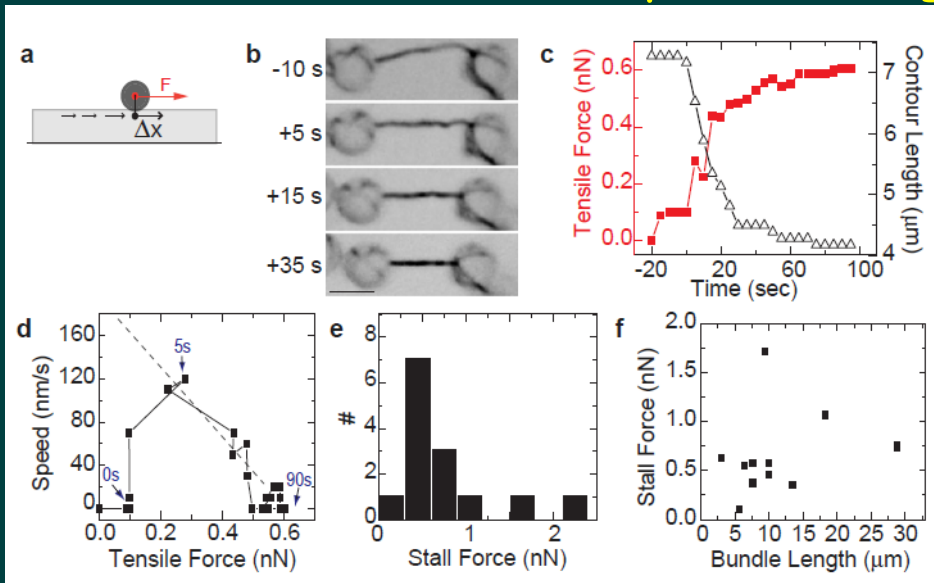
[†]Institute for Biophysical Dynamics and [‡]James Franck Institute and Department of Physics, University of Chicago, Chicago, Illinois



Reconstituted bundles contract



Force does not depend on length, velocity proportional to length

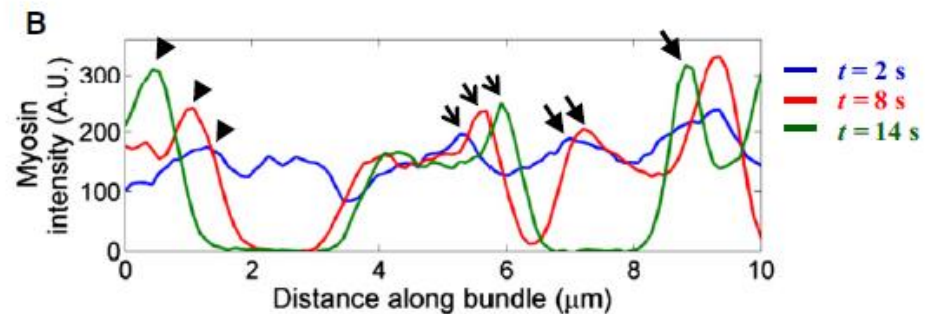
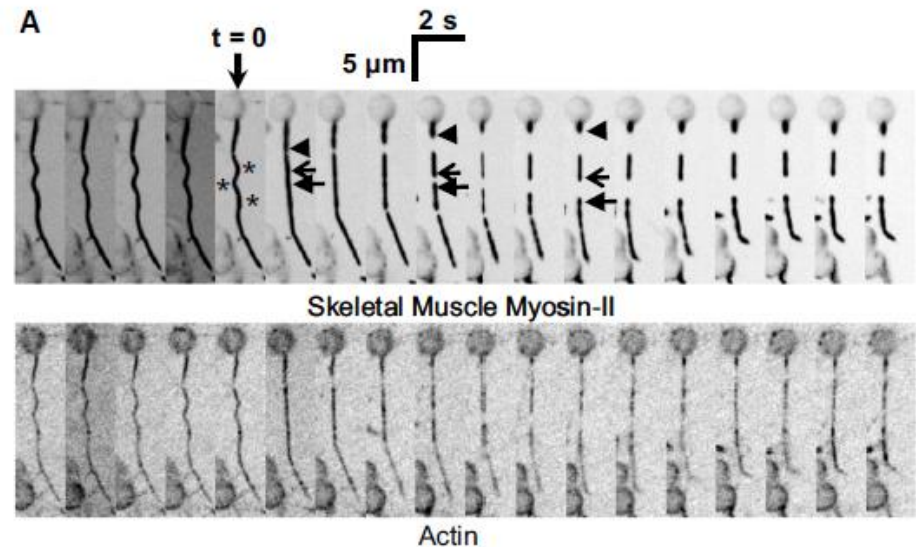
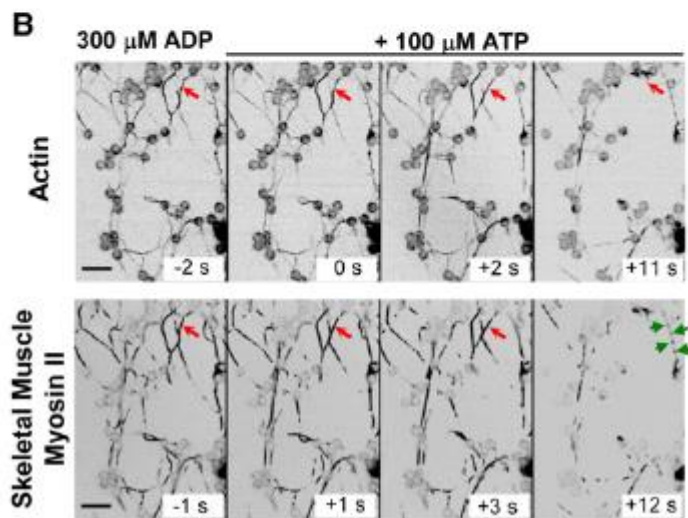
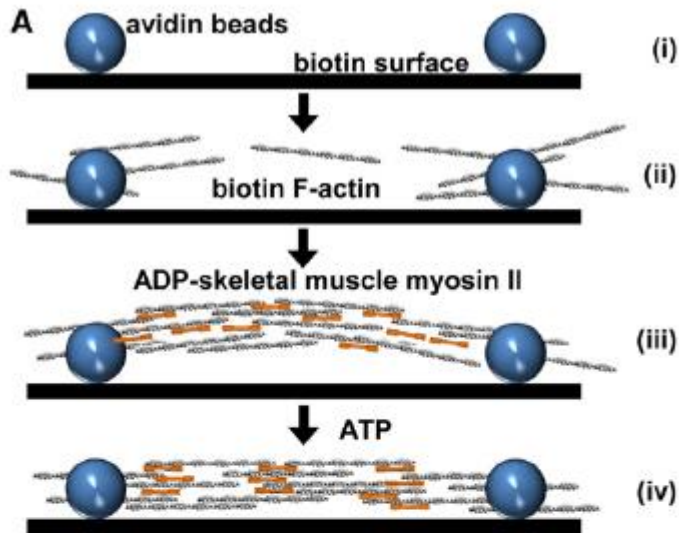


Self-Organization of Myosin II in Reconstituted Actomyosin Bundles

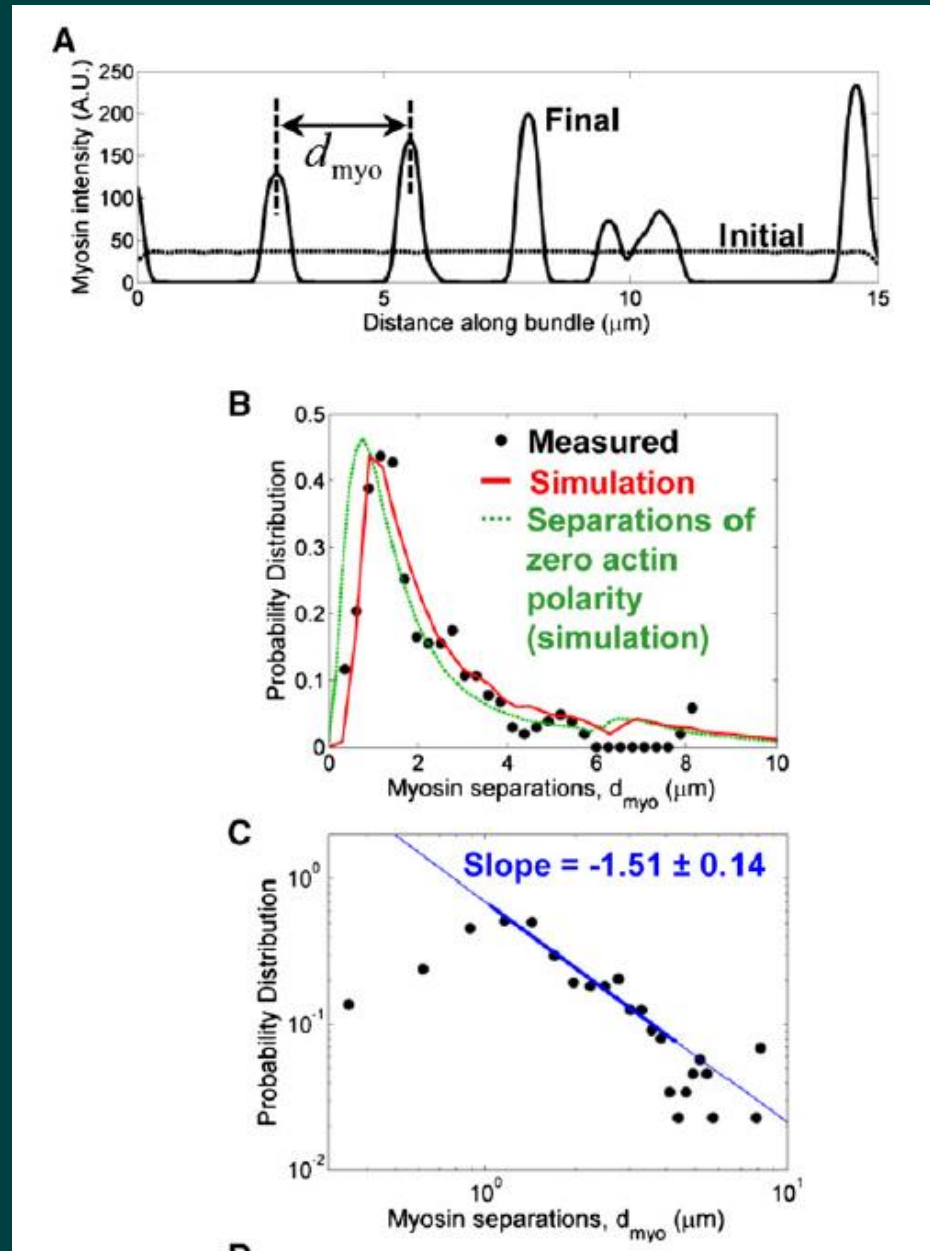
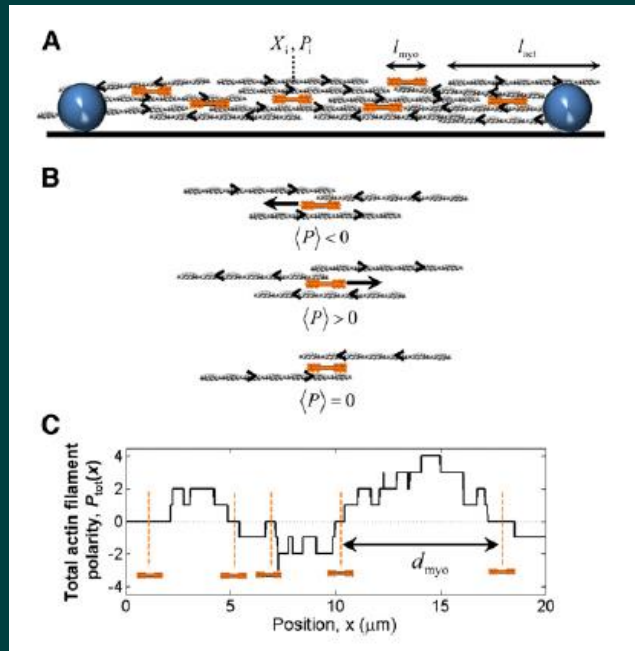
Matthew R. Stachowiak,[†] Patrick M. McCall,^{‡§} Todd Thoresen,^{‡¶} Hayri E. Balcioglu,[†] Lisa Kasiewicz,[†] Margaret L. Gardel,^{‡§¶} and Ben O'Shaughnessy^{†*}

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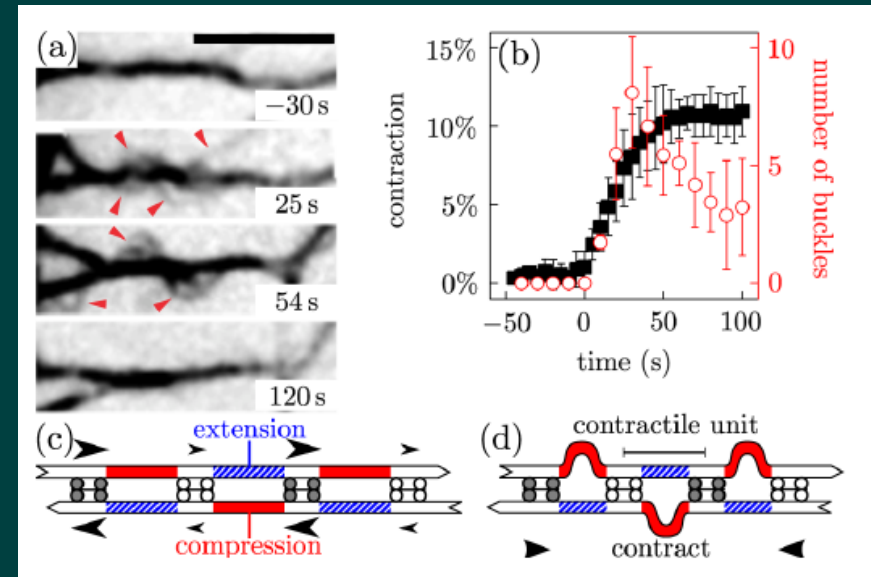
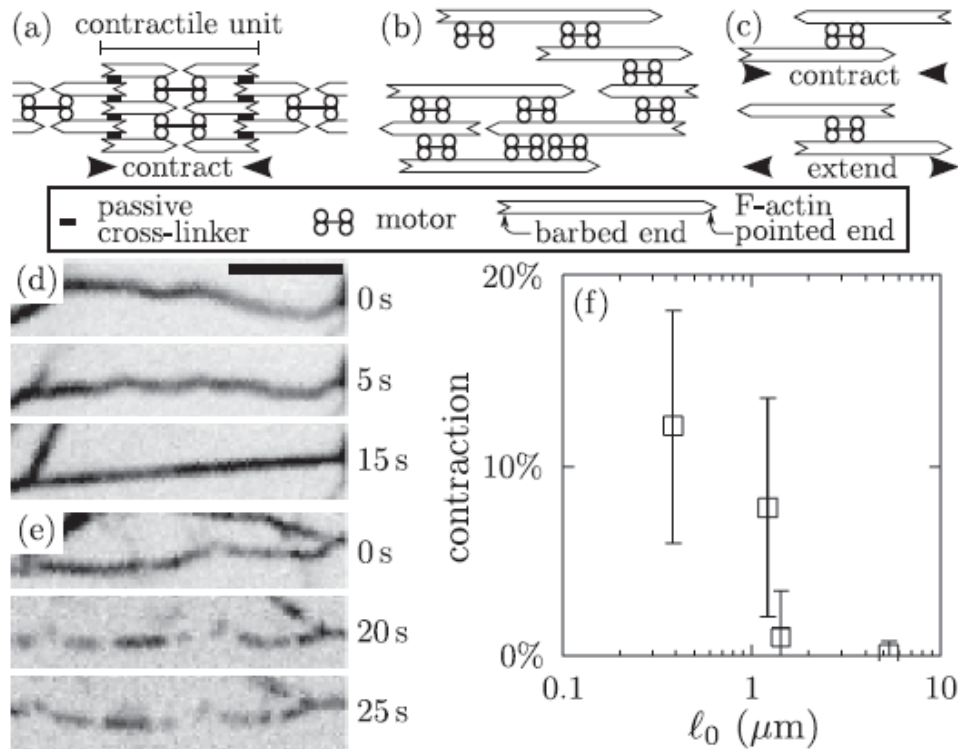


myosin concentrates at zero actin polarity



Contractile Units in Disordered Actomyosin Bundles Arise from F-Actin Buckling

Martin Lenz,¹ Todd Thoresen,² Margaret L. Gardel,^{1,2,3} and Aaron R. Dinner^{1,2,4,*}



next time: how does it work in the cells?