



BIOC | Bioinformatics Competence Center

Bioinformatics Analysis of RNA-sequencing

BIO-693

Unil **EPFL**

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This morning

- Connect to the cluster
- Collect and organize all the raw fastq data
- Prepare reference data
- Map fastq to reference
- Check mapping statistics
- Obtain counts

Getting started on helvetios

- Connect to helvetios and request an interactive session
 - `ssh <username>@helvetios.epfl.ch`
 - `module add tmux`
 - `tmux, or tmux attach`
 - `cd /scratch/<username>`
 - `Sinteract -a bio-693 -c 20 -t 08:00:00 -m 40g`
- On the cluster node
 - `cp /scratch/iseli/course/SraRunTable.txt .`
 - Do not forget/omit the dot (.)
 - `singularity shell -s /bin/bash -B /scratch/$USER -B /scratch/iseli /scratch/iseli/MicMap-2.4-f38.sif`

Example SRA – retrieve

- Retrieve all the samples

```
for n in `cut -d , -f 1 SraRunTable.txt | grep ^SRR`; do
    echo "Processing $n";
    fasterq-dump --split-3 --progress $n;
    pigz --fast ${n}*.fastq;
    sra-stat --quick $n >${n}_stat.txt;
done
```

- Check retrieved data

```
for f in SRR*_stat.txt; do
    n=`basename $f _stat.txt`;
    cntFQ=`perl -ne '($c)=$_~/^[^:]+\|(\d+):/;$t+=$c;END{print "$t\n"}' $f`;
    linesFQ=`gzip -dc ${n}_1.fastq.gz | wc -l`;
    if [[ $cntFQ -ne $((linesFQ / 4)) ]]; then
        echo "We have a problem for $n - $cntFQ vs $((linesFQ / 4))";
    else
        echo "count is correct for $n - $cntFQ";
    fi;
done
```

Retrieving SRA data

```
helvetios_iseli: ls -alth
total 76G
drwxr-xr-x 2 iseli BICC-unit 4.0K Jun 17 07:42 fasterq.tmp.helvetios.4169460
drwxr-xr-x 3 iseli BICC-unit 4.0K Jun 17 07:36 .
-rw-r--r-- 1 iseli BICC-unit 39K Jun 17 07:36 SRR16219945_stat.txt
-rw-r--r-- 1 iseli BICC-unit 2.2G Jun 17 07:31 SRR16219945_1.fastq.gz
-rw-r--r-- 1 iseli BICC-unit 2.4G Jun 17 07:31 SRR16219945_2.fastq.gz
-rw-r--r-- 1 iseli BICC-unit 39K Jun 17 05:45 SRR16219944_stat.txt
-rw-r--r-- 1 iseli BICC-unit 2.3G Jun 17 05:39 SRR16219944_1.fastq.gz
-rw-r--r-- 1 iseli BICC-unit 2.5G Jun 17 05:39 SRR16219944_2.fastq.gz
-rw-r--r-- 1 iseli BICC-unit 0 Jun 17 03:44 SRR16219943_stat.txt
-rw-r--r-- 1 iseli BICC-unit 2.8G Jun 17 03:38 SRR16219943_2.fastq.gz
-rw-r--r-- 1 iseli BICC-unit 2.7G Jun 17 03:38 SRR16219943_1.fastq.gz
-rw-r--r-- 1 iseli BICC-unit 40K Jun 17 01:34 SRR16219940_stat.txt
-rw-r--r-- 1 iseli BICC-unit 2.5G Jun 17 01:28 SRR16219940_1.fastq.gz
-rw-r--r-- 1 iseli BICC-unit 2.8G Jun 17 01:28 SRR16219940_2.fastq.gz
-rw-r--r-- 1 iseli BICC-unit 40K Jun 16 23:25 SRR16219938_stat.txt
-rw-r--r-- 1 iseli BICC-unit 2.8G Jun 16 23:19 SRR16219938_2.fastq.gz
-rw-r--r-- 1 iseli BICC-unit 2.6G Jun 16 23:19 SRR16219938_1.fastq.gz
```

Checking SRA data

```
Singularity> for f in SRR*_stat.txt; do n=`basename $f _stat.txt`; cntFQ=`perl -ne '($c)=$_~/^[^:]+\|(\d+):/;$t+=$c;END{print  
"$t\n"}' $f`; linesFQ=`gzip -dc ${n}_1.fastq.gz | wc -l`; if [[ $cntFQ -ne $((linesFQ / 4)) ]]; then echo "We have a problem  
for $n - $cntFQ vs $((linesFQ / 4))"; else echo "count is correct for $n - $cntFQ"; fi; done  
  
count is correct for SRR16219921 - 41125131  
count is correct for SRR16219922 - 31498370  
count is correct for SRR16219923 - 37099819  
count is correct for SRR16219924 - 35957634  
count is correct for SRR16219925 - 38804161  
count is correct for SRR16219926 - 33688333  
count is correct for SRR16219928 - 39271560  
count is correct for SRR16219929 - 42674023  
count is correct for SRR16219930 - 30197829  
count is correct for SRR16219931 - 34847378  
count is correct for SRR16219932 - 34691862  
count is correct for SRR16219933 - 33625186  
count is correct for SRR16219935 - 40275240  
count is correct for SRR16219936 - 34729325  
count is correct for SRR16219937 - 37121145  
count is correct for SRR16219938 - 38216703  
count is correct for SRR16219940 - 38089443  
count is correct for SRR16219941 - 37228589  
count is correct for SRR16219942 - 36950994  
count is correct for SRR16219943 - 40066968  
count is correct for SRR16219944 - 34343035  
count is correct for SRR16219945 - 32508162  
count is correct for SRR16219946 - 34186012  
count is correct for SRR16219947 - 36344402  
count is correct for SRR16219948 - 35999453  
count is correct for SRR16219949 - 33542412  
count is correct for SRR16219950 - 39697257  
count is correct for SRR16219951 - 38813449  
count is correct for SRR16219952 - 33811791  
count is correct for SRR16219954 - 33188661  
count is correct for SRR16219955 - 33387404  
count is correct for SRR16219956 - 37428628  
Singularity>
```

Organize data

- Create a working directory in `/scratch/$USER`
- The container is available here :
 - `/scratch/iseli/MicMap-2.4-f38.sif`
- Data is available here :
 - `/scratch/iseli/course`
- Try to use symlinks when possible

Getting alignment (BAM) files

- STAR aligner – <https://github.com/alexdobin/STAR>
- Source of reference data
 - Gencode – <https://www.gencodegenes.org/>
 - Ensembl – <https://www.ensembl.org/index.html>
- Ideally use genome (FASTA) + gene annotation (gtf, gff, gff3)
 - Use primary assembly for the genome
 - Use comprehensive gene annotation (gencode) for the genes
 - Use `wget` to retrieve the data on the cluster

Preparing STAR index

- Use **genomeGenerate** command
 - `STAR --runMode genomeGenerate \`
 `--genomeDir STAR_mm39vM32/ \`
 `--genomeFastaFiles \`
 `GRCm39.primary_assembly.genome.fa \`
 `--sjdbGTFfile gencode.vM32.annotation.gtf`
- Beware the **genomeSAindexNbases** parameter for small genomes

Preparing STAR index files

```
Sinteract -a bio-693 -c 4 -t 08:00:00 -m 30g
```

```
Singularity> STAR --runMode genomeGenerate --genomeDir STAR_mm39vM32/ --genomeFastaFiles GRCm39.primary_assembly.genome.fa --sjdbGTFfile gencode.vM32.annotation.gtf
STAR --runMode genomeGenerate --genomeDir STAR_mm39vM32/ --genomeFastaFiles GRCm39.primary_assembly.genome.fa --sjdbGTFfile gencode.vM32.annotation.gtf
STAR version: 2.7.10b compiled: 2022-11-08T13:46:18+01:00 pccig4007.unil.ch:/root/src/github/STAR/source
Jun 19 10:32:47 ..... started STAR run
Jun 19 10:32:47 ... starting to generate Genome files
Jun 19 10:33:24 ..... processing annotations GTF
Jun 19 10:33:42 ... starting to sort Suffix Array. This may take a long time...
Jun 19 10:33:56 ... sorting Suffix Array chunks and saving them to disk...
Killed
Singularity>
```

Insufficient RAM

```
Sinteract -a bio-693 -c 4 -t 08:00:00 -m 80g
```

```
Singularity> STAR --runMode genomeGenerate --genomeDir STAR_mm39vM32/ --genomeFastaFiles GRCm39.primary_assembly.genome.fa --sjdbGTFfile gencode.vM32.annotation.gtf
STAR --runMode genomeGenerate --genomeDir STAR_mm39vM32/ --genomeFastaFiles GRCm39.primary_assembly.genome.fa --sjdbGTFfile gencode.vM32.annotation.gtf
STAR version: 2.7.10b compiled: 2022-11-08T13:46:18+01:00 pccig4007.unil.ch:/root/src/github/STAR/source
Jun 19 11:00:12 ..... started STAR run
Jun 19 11:00:12 ... starting to generate Genome files
Jun 19 11:00:47 ..... processing annotations GTF
Jun 19 11:01:04 ... starting to sort Suffix Array. This may take a long time...
Jun 19 11:01:17 ... sorting Suffix Array chunks and saving them to disk...
Jun 19 12:04:55 ... loading chunks from disk, packing SA...
Jun 19 12:05:44 ... finished generating suffix array
Jun 19 12:05:44 ... generating Suffix Array index
Jun 19 12:08:42 ... completed Suffix Array index
Jun 19 12:08:42 ..... inserting junctions into the genome indices
Jun 19 12:16:08 ... writing Genome to disk ...
Jun 19 12:16:08 ... writing Suffix Array to disk ...
Jun 19 12:16:14 ... writing SAindex to disk
Jun 19 12:16:15 ..... finished successfully
Singularity>
```

Successful

Run the aligner

n=SRR16219925

```
STAR --runMode alignReads \  
  --runThreadN 20 \  
  --genomeDir STAR_mm39vM32 \  
  --twopassMode Basic \  
  --outFileNamePrefix ${n}_mm39vM32/ \  
  --outSAMtype BAM SortedByCoordinate \  
  --outSAMattributes All \  
  --outSAMattrRGline ID:${n} LB:${n} SM:${n} PL:ILLUMINA \  
  --readFilesCommand "gzip -dc" \  
  --quantMode GeneCounts \  
  --limitBAMsortRAM 160000000000 \  
  --readFilesIn ${n}_?.fastq.gz
```

Example run

```
Singularity> STAR --runMode alignReads --runThreadN 20 --genomeDir STAR_mm39vM32 --twopassMode Basic --outFileNamePrefix ${n}_mm39vM32/ --outSAMtype BAM SortedByCoordinate --outSAMattributes All --outSAMattrRGline ID:$n LB:$n SM:$n PL:ILLUMINA --readFilesCommand "gzip -dc" --quantMode GeneCounts --limitBAMsortRAM 16000000000 --readFilesIn ${n}?.fastq.gz

STAR --runMode alignReads --runThreadN 20 --genomeDir STAR_mm39vM32 --twopassMode Basic --outFileNamePrefix SRR16219925_mm39vM32/ --outSAMtype BAM SortedByCoordinate --outSAMattributes All --outSAMattrRGline ID:SRR16219925 LB:SRR16219925 SM:SRR16219925 PL:ILLUMINA --readFilesCommand "gzip -dc" --quantMode GeneCounts --limitBAMsortRAM 16000000000 --readFilesIn SRR16219925_1.fastq.gz SRR16219925_2.fastq.gz
STAR version: 2.7.10b   compiled: 2022-11-08T13:46:18+01:00 pccig4007.unil.ch:/root/src/github/STAR/source
Jun 19 14:17:37 ..... started STAR run
Jun 19 14:17:37 ..... loading genome
Jun 19 14:17:58 ..... started 1st pass mapping
Jun 19 14:19:36 ..... finished 1st pass mapping
Jun 19 14:19:37 ..... inserting junctions into the genome indices
Jun 19 14:20:52 ..... started mapping
Jun 19 14:22:46 ..... finished mapping
Jun 19 14:22:47 ..... started sorting BAM
Jun 19 14:23:39 ..... finished successfully
Singularity>
```

samtools

- Need to get familiar with this tool to work with BAM files
- <https://github.com/samtools/samtools>

After STAR alignment completes

- Examine the directory `${n}_mm39vM32/`
- Examine the logs, counts and BAM files
 - `cat Log.final.out`
 - `less ReadsPerGene.out.tab`
 - `sort -k 4,4nr ${n}_mm39vM32/ReadsPerGene.out.tab | less`
 - `grep ENSMUSG00000064351.1 gencode.vM32.annotation.gtf | less`
 - `samtools view -h Aligned.sortedByCoord.out.bam | less`
 - `samtools index Aligned.sortedByCoord.out.bam`
 - `samtools tview \`
 - `-p chr1:5153351 \`
 - `${n}_mm39vM32/Aligned.sortedByCoord.out.bam \`
 - `GRCm39.primary_assembly.genome.fa`
 - Chr1:5171281 – probable alt splicing
 - Chr1:5194491 – variants

Get a feel for duplicates

- `java -jar /usr/local/share/java/picard.jar MarkDuplicates \
-I ${n}_mm39vM32/Aligned.sortedByCoord.out.bam \
-O ${n}_mm39vM32/Aligned.sortedByCoord.MD.bam \
-M ${n}_mm39vM32/marked_dup_metrics.txt`
- `grep -A 1 ^LIBRARY \
${n}_mm39vM32/marked_dup_metrics.txt | column -t`

SAM / BAM file format

- SAM is a text file with
 - A header with lines starting by @
 - The rest with at least 11 columns, tab-delimited
- BAM is a compressed version of SAM

1	2	3	4	5	6	7	8	9	10	11
Name	Flag	Chr	Pos	MpQ	Cigar	PChr	Ppos	Len	Seq	Qual
String	Int	String	Int	Int	String	String	Int	Int	String	String

- To view:
 - `samtools view <file>.bam | less -S`

SAM/BAM flags

Decimal	Hex	Description
1	0x1	template having multiple segments in sequencing
2	0x2	each segment properly aligned according to the aligner
4	0x4	segment unmapped
8	0x8	next segment in the template unmapped
16	0x10	SEQ being reverse complemented
32	0x20	SEQ of the next segment in the template being reverse complemented
64	0x40	the first segment in the template
128	0x80	the last segment in the template
256	0x100	secondary alignment
512	0x200	not passing filters, such as platform/vendor quality controls
1024	0x400	PCR or optical duplicate
2048	0x800	supplementary alignment

Secondary and duplicates

- `samtools view \`
 `${n}_mm39vM32/Aligned.sortedByCoord.out.bam \`
 `chr1:5153351-5153600 | less`
- `samtools view \`
 `${n}_mm39vM32/Aligned.sortedByCoord.MD.bam \`
 `chr1:5153351-5153600 | less`
- Can play with `-f 256` and `-f 1024` to filter reads

Get raw counts

- Several possibilities – **htseq-count**, **featureCounts**
 - Htseq-count – <https://doi.org/10.1093%2Fbioinformatics%2Fbtu638>
 - featureCounts – <https://doi.org/10.1093/bioinformatics/btt656>

Get raw counts using featureCounts

- `featureCounts -T 20 \`
 `--primary -s 2 -p -B -Q 10 \`
 `-a gencode.vM32.annotation.gtf \`
 `-o featureCounts.txt \`
 `SRR162199*_mm39vM32/Aligned.sortedByCoord.out.bam \`
 `2>featureCounts_log.txt`
- `sed 's/_mm39vM32\//Aligned.sortedByCoord.out.bam//g' \`
 `featureCounts.txt.summary | column -t`
- `cut -f1,7- featureCounts.txt | sed 1d | \`
 `sed 's/_mm39vM32\//Aligned.sortedByCoord.out.bam//g' \`
 `> final_counts.txt`