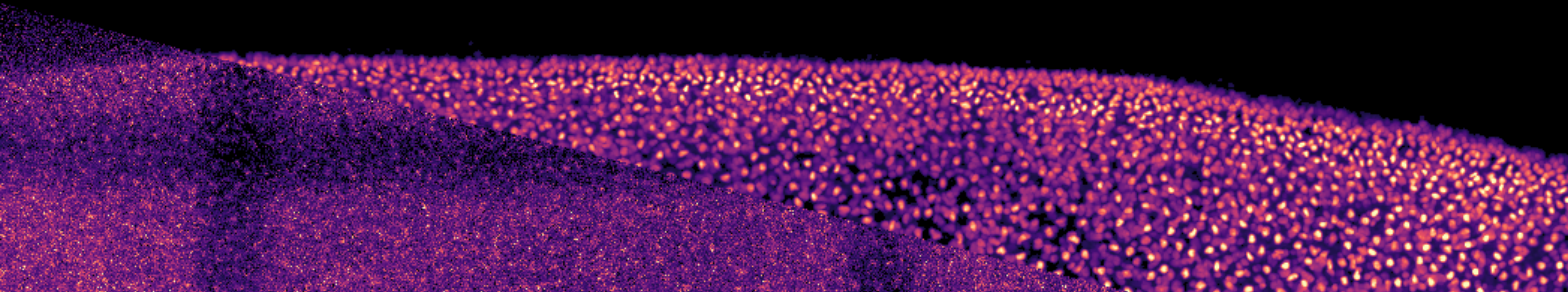


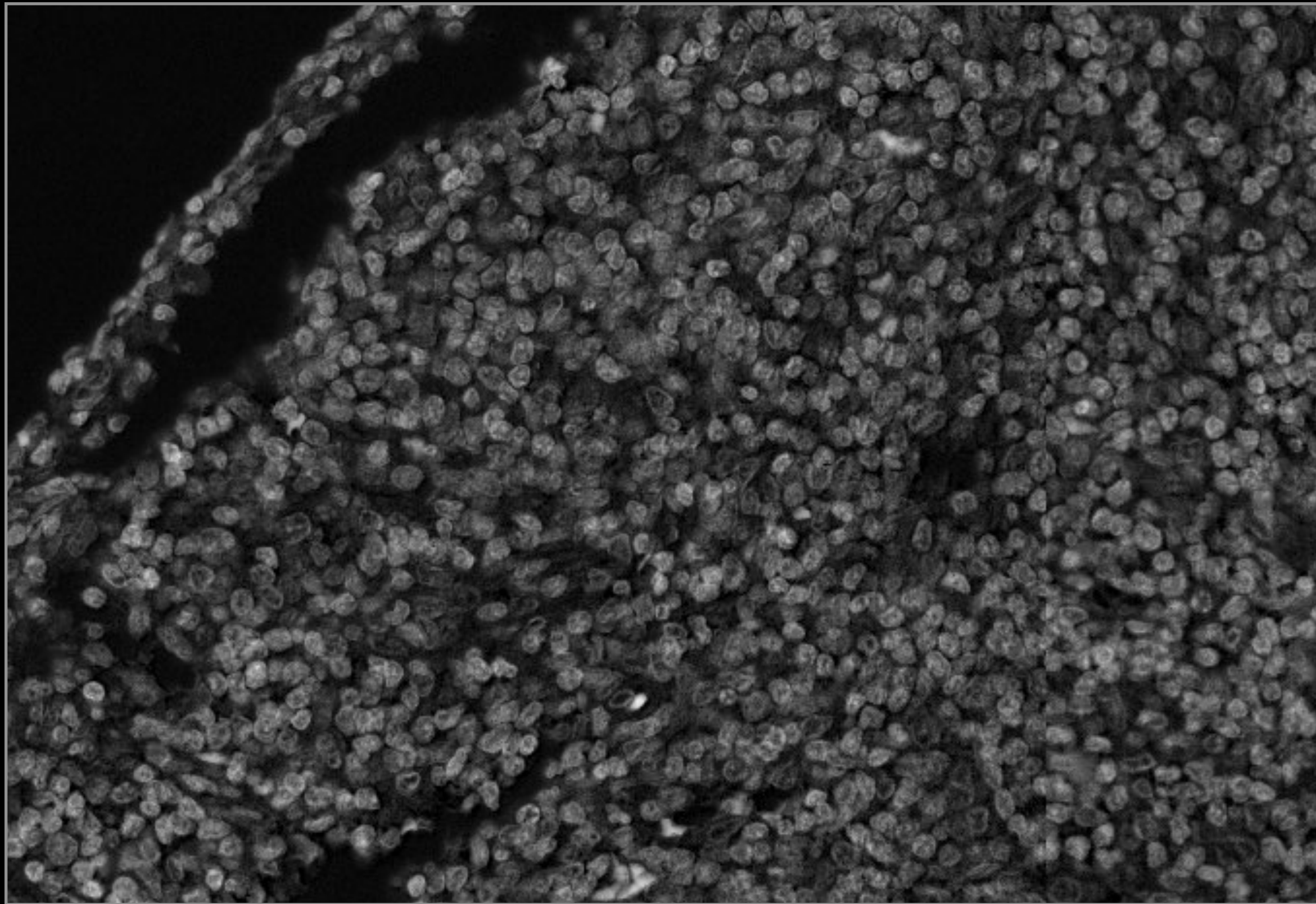
Image analysis for microscopy with machine learning

BIO-221
October 31st, 2024

Albert Dominguez Mantes
Weigert Group, EPFL  /TU Dresden 
<https://www.epfl.ch/labs/weigert-lab/>

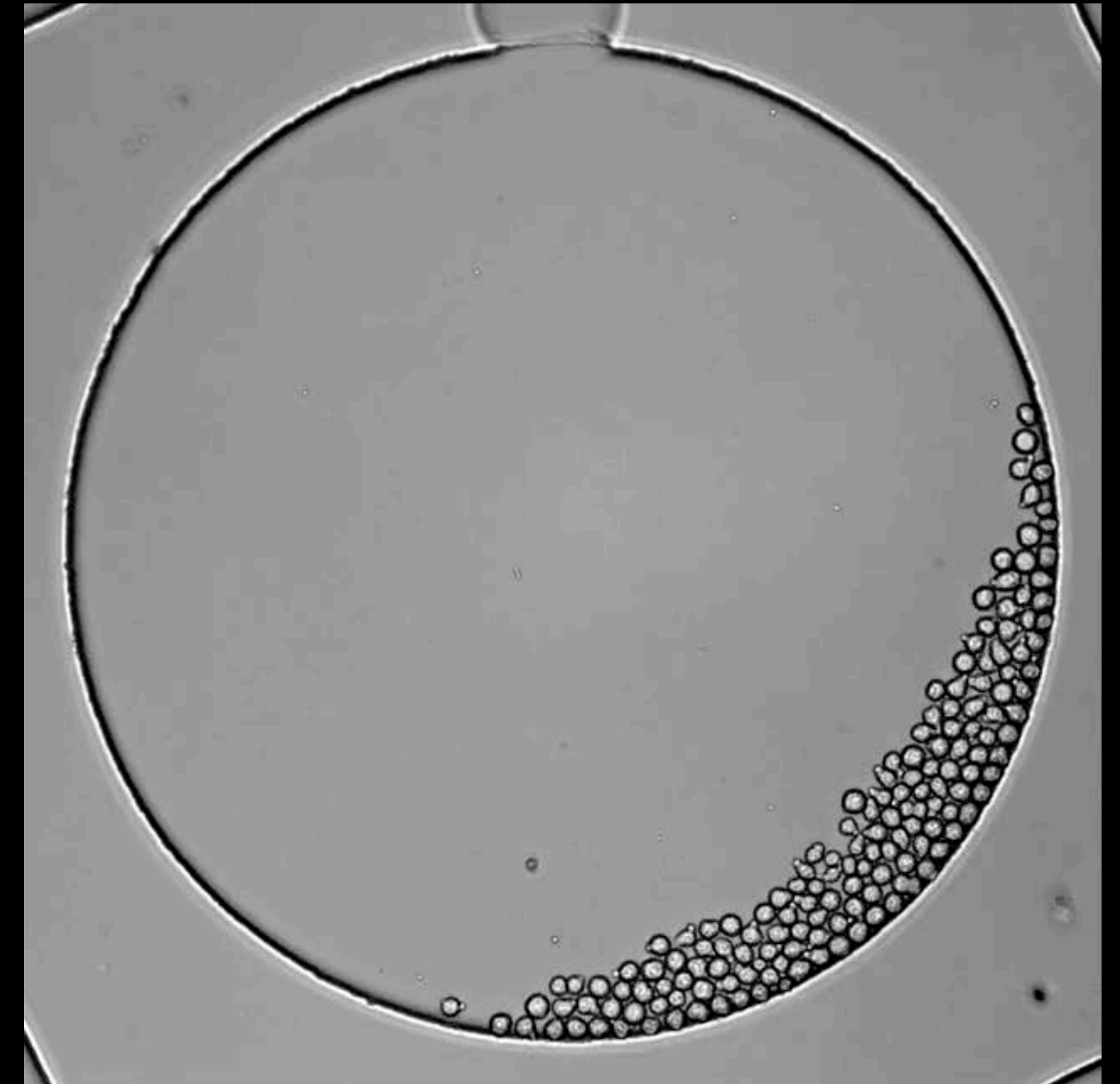


2D (fluorescence)



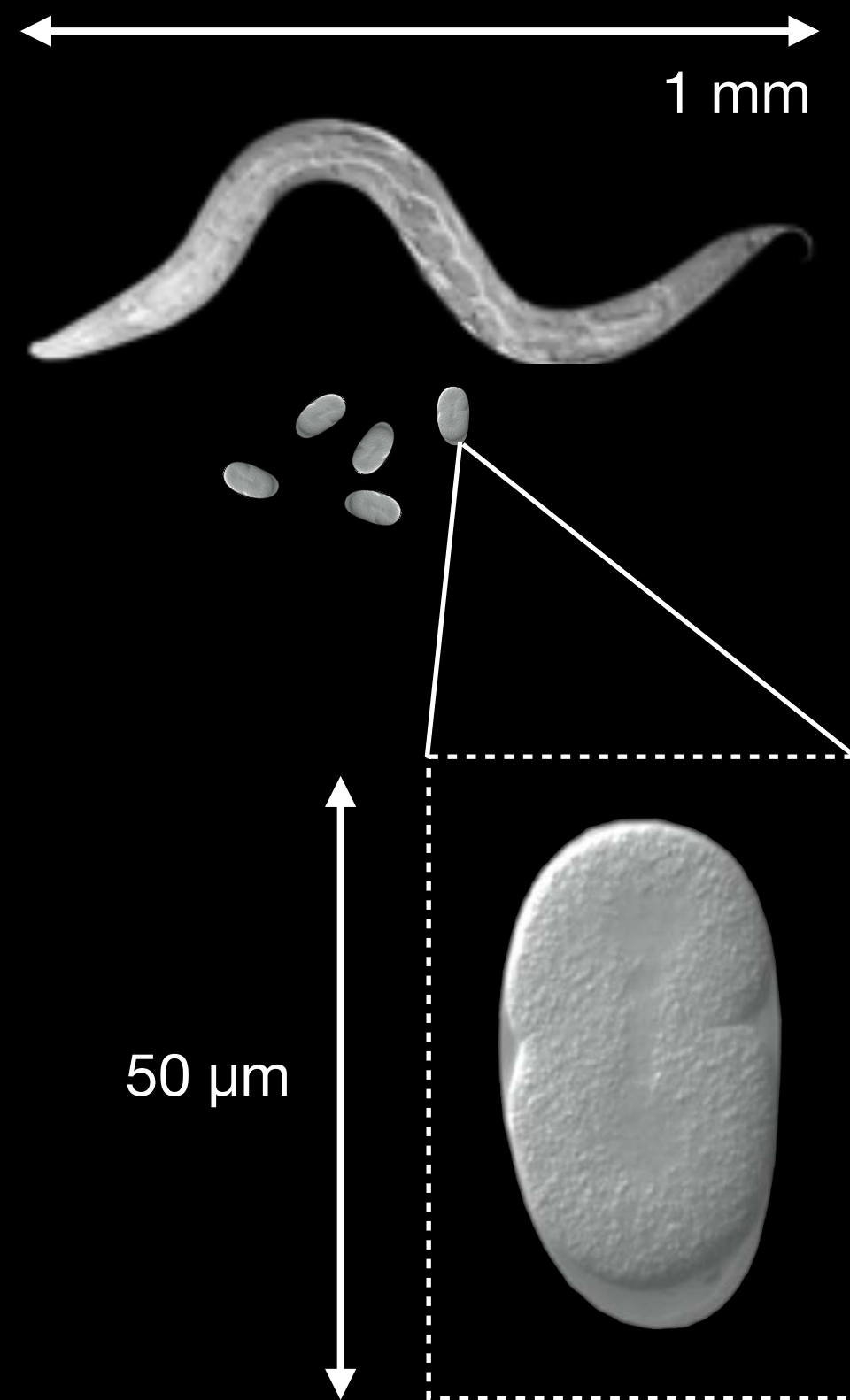
Data from Anna Maria Tsakiroglou (Manchester)

2D + time (brightfield)

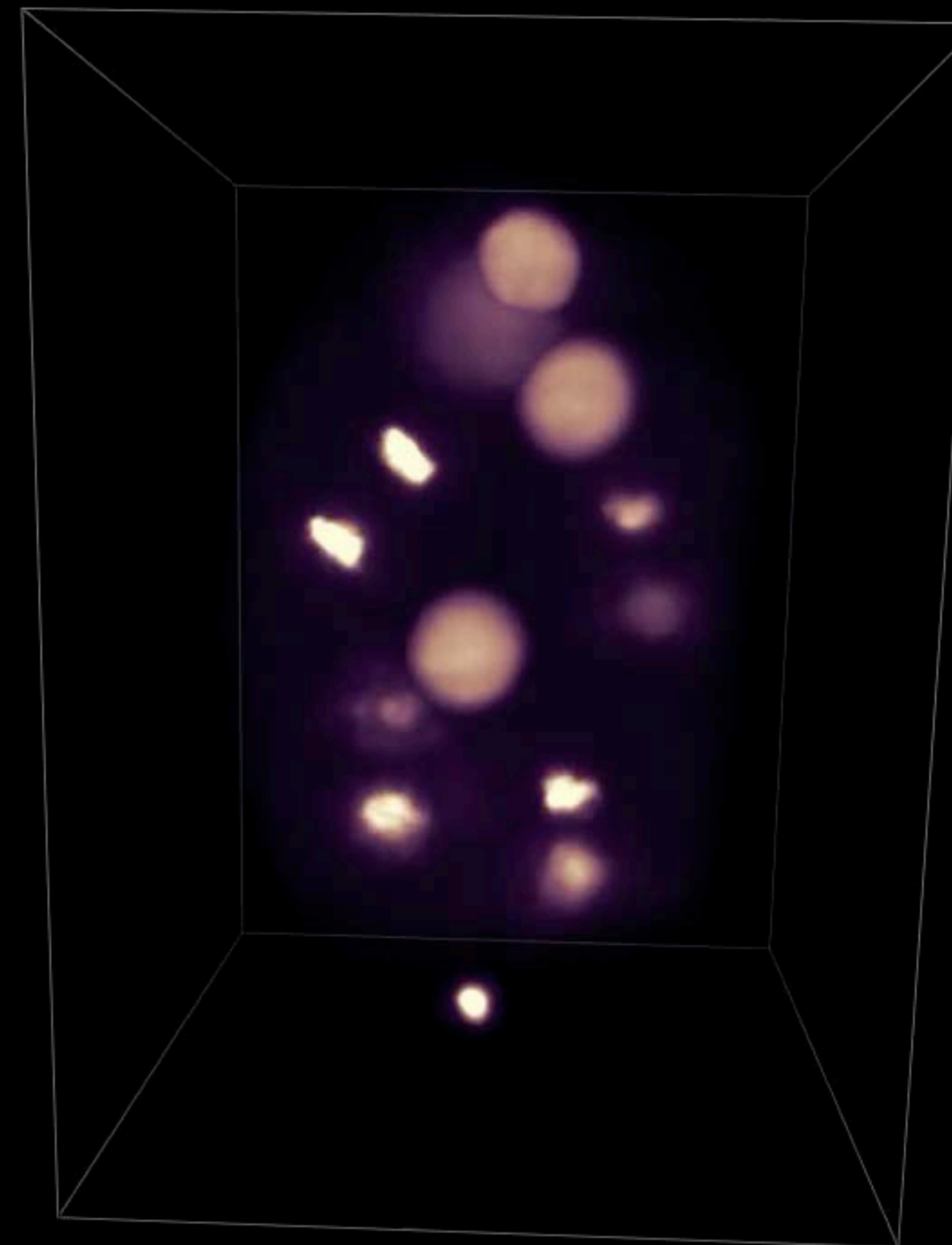


Mouse stem cells, data from cell tracking challenge

3D + time - fluorescence microscopy of developing organisms



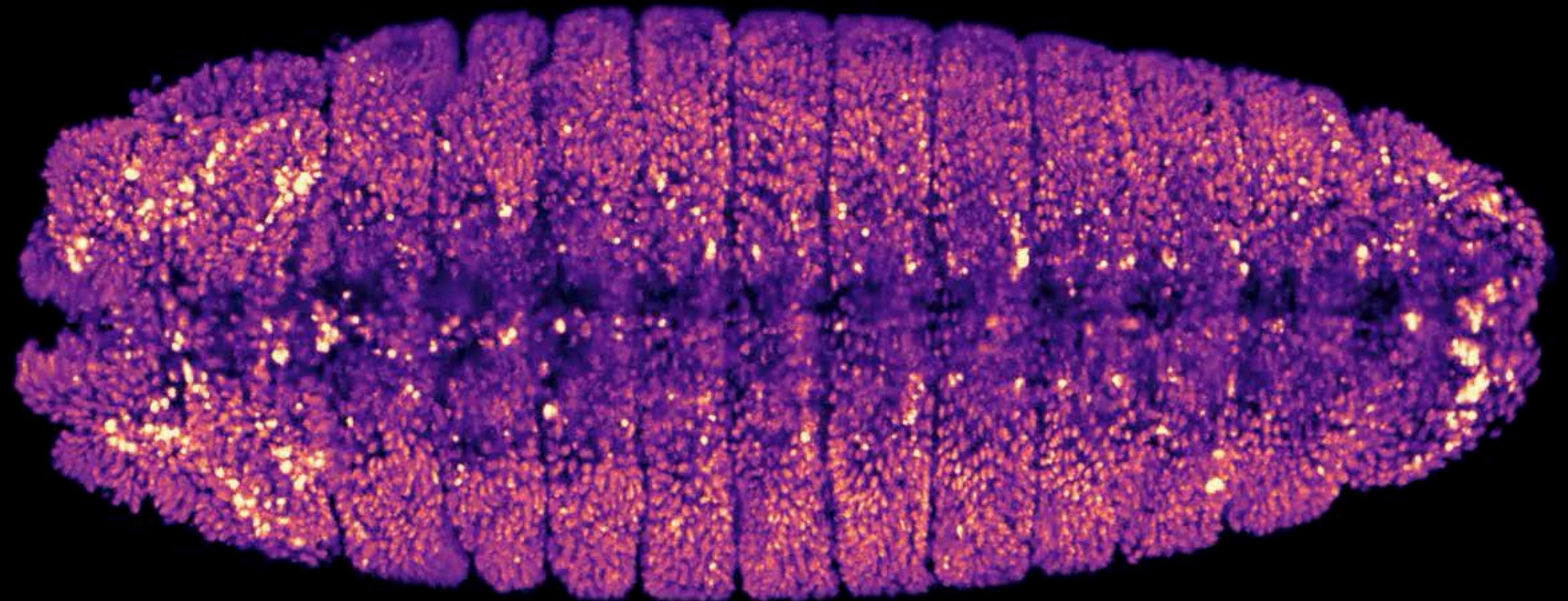
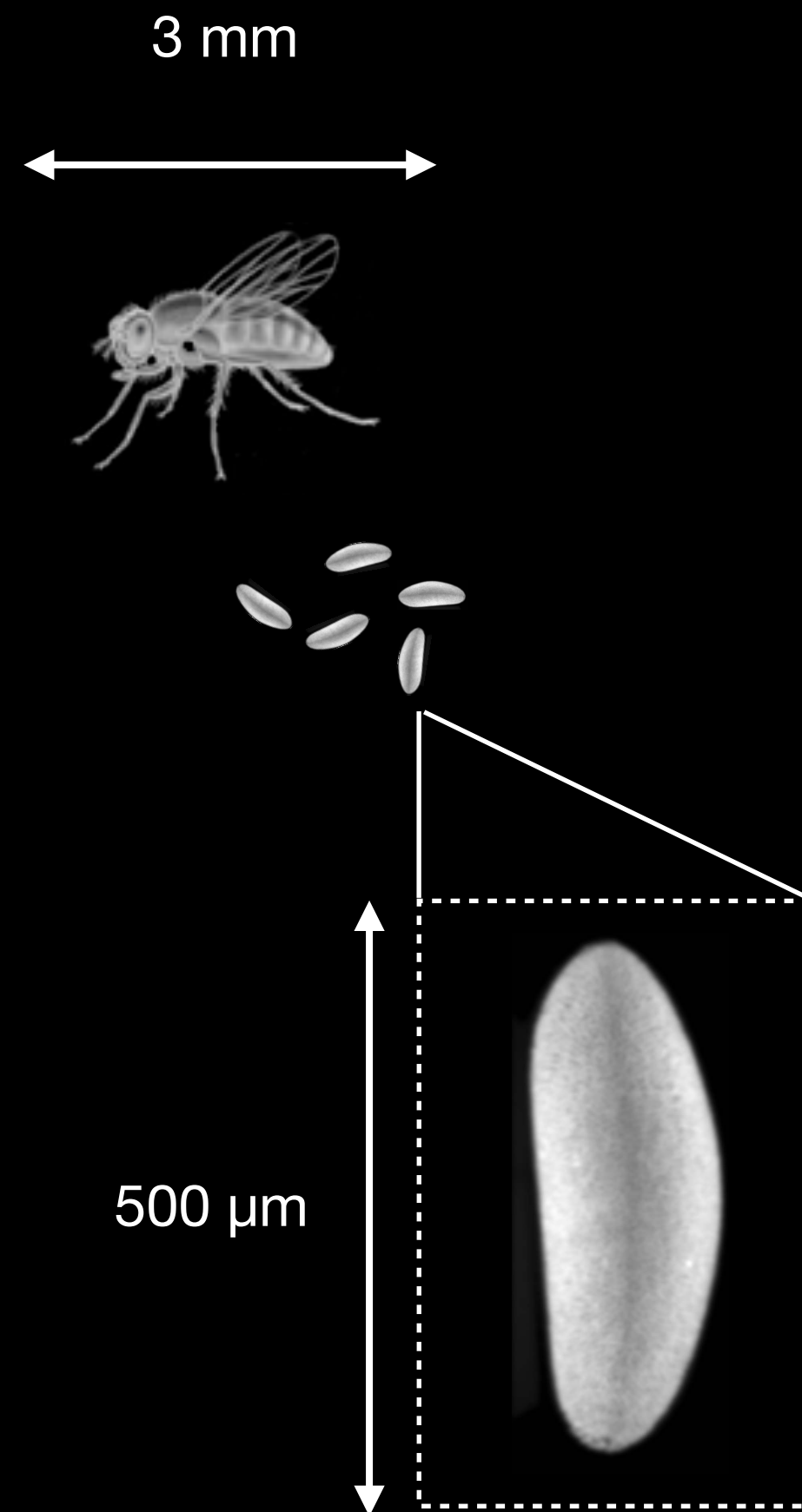
C. elegans



1:22:04

Data: Gene Myers Lab (Dresden)

3D + time - fluorescence microscopy of developing organisms



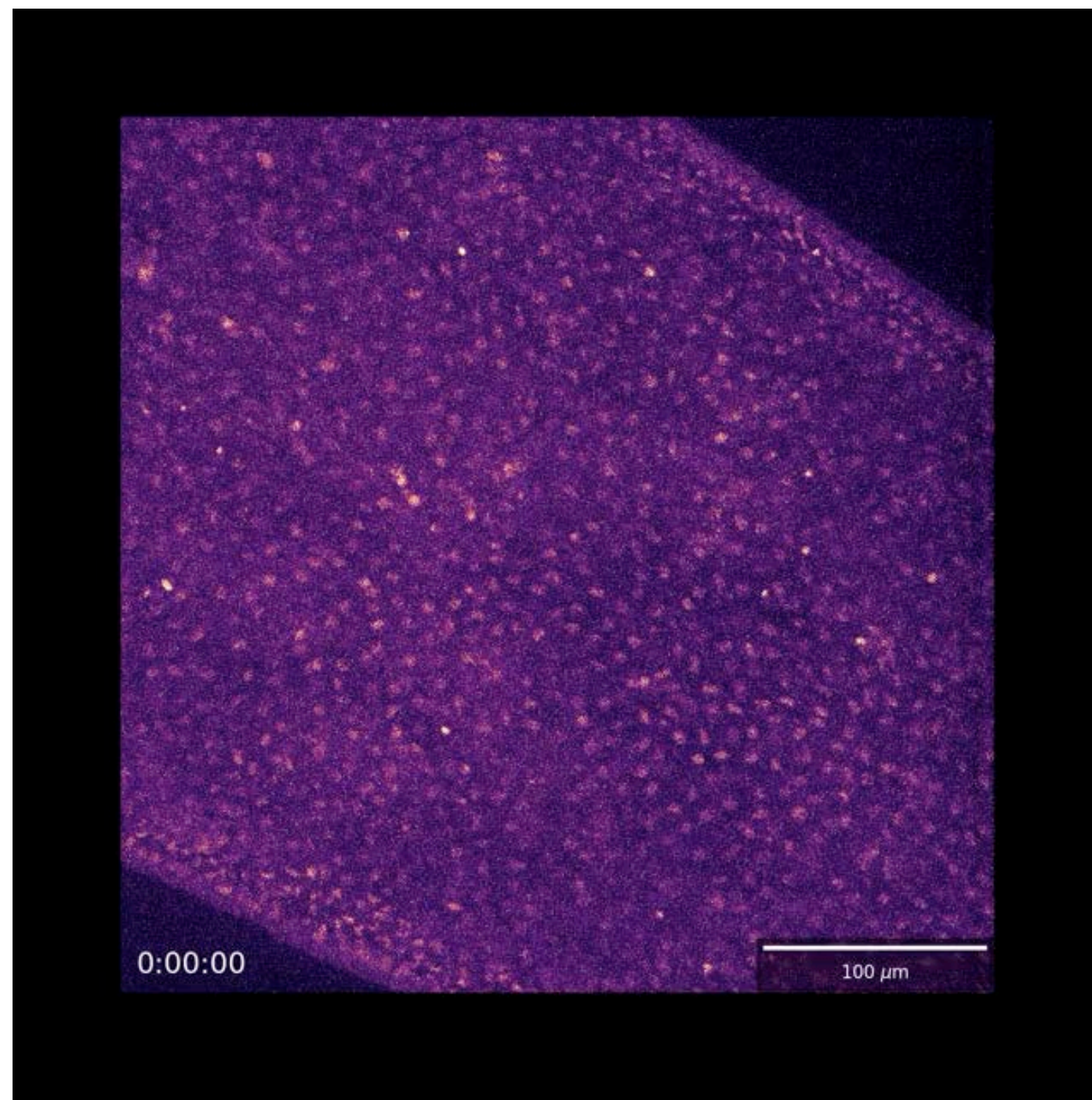
Drosophila melanogaster

Royer et al. (2016)

Extraction of quantitative biological information from digitised camera images

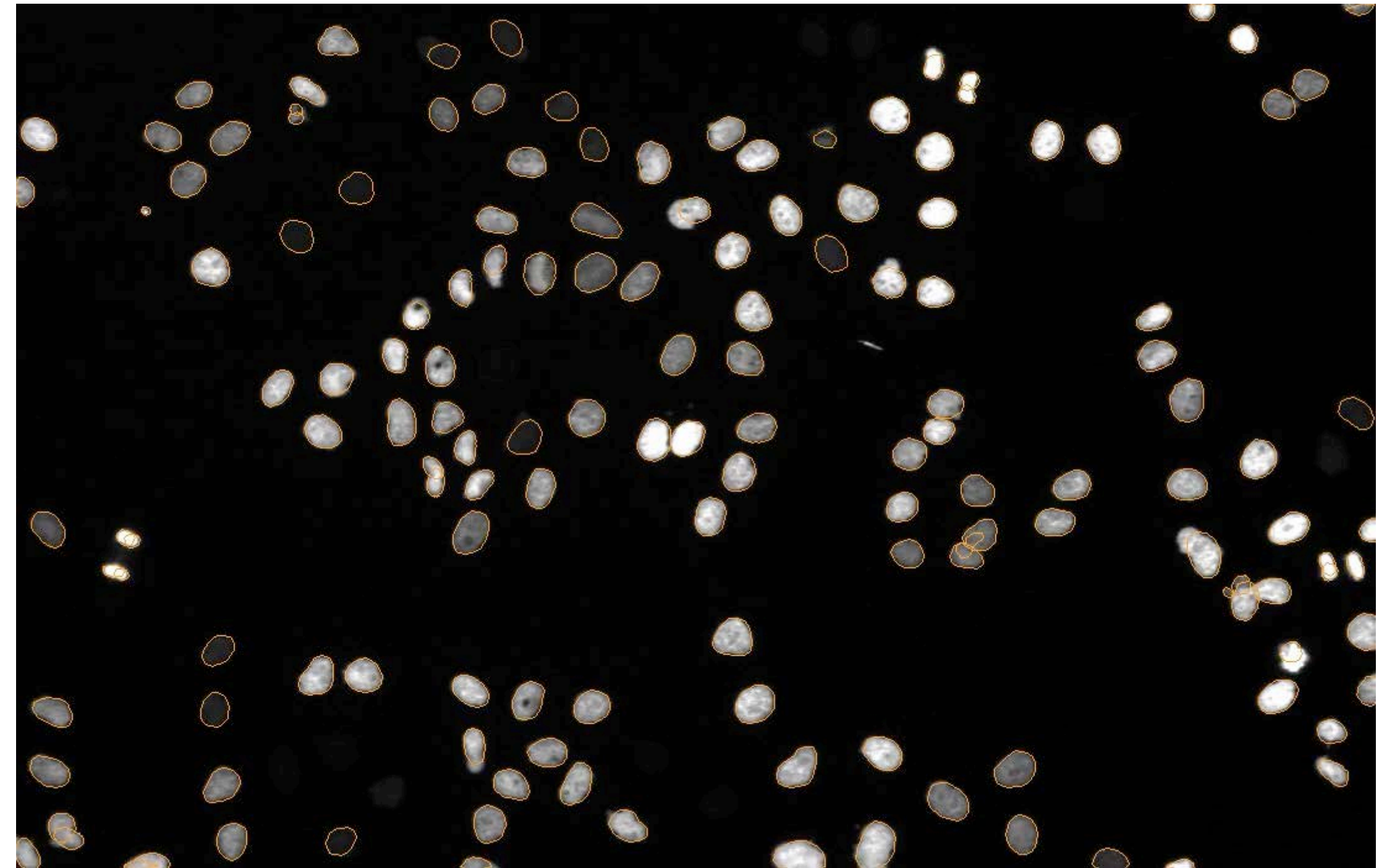
Image denoising/restoration

Remove camera noise, remove blur...

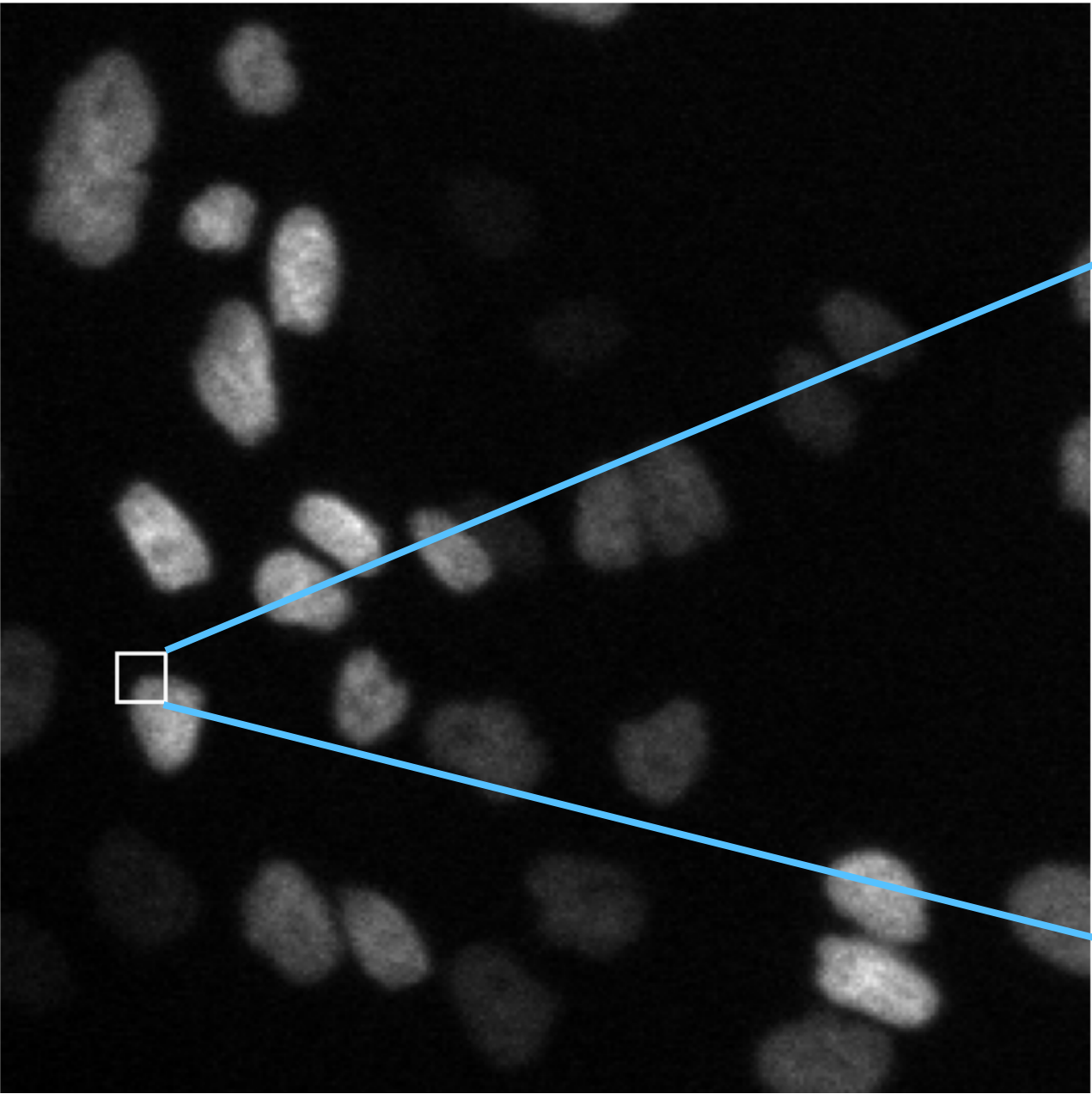


Object Detection and Segmentation

Find contours of cells, detect spots, track cells over time, ...

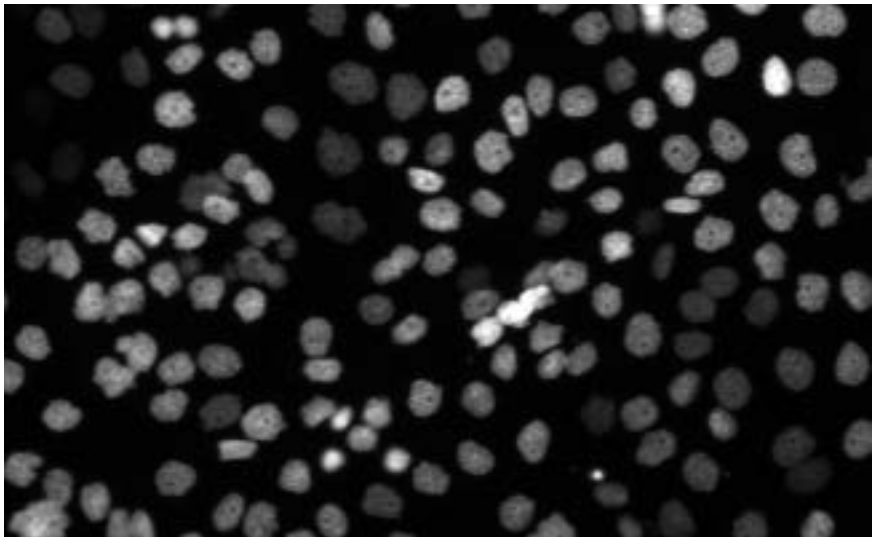
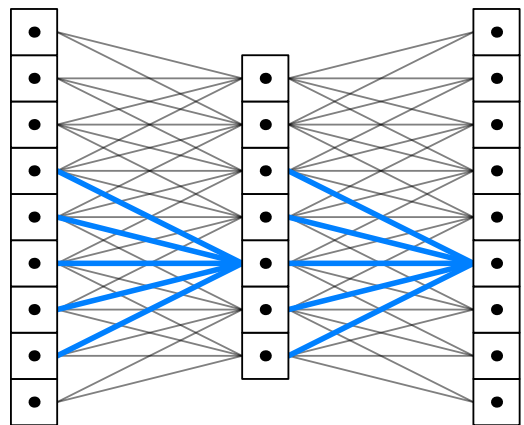
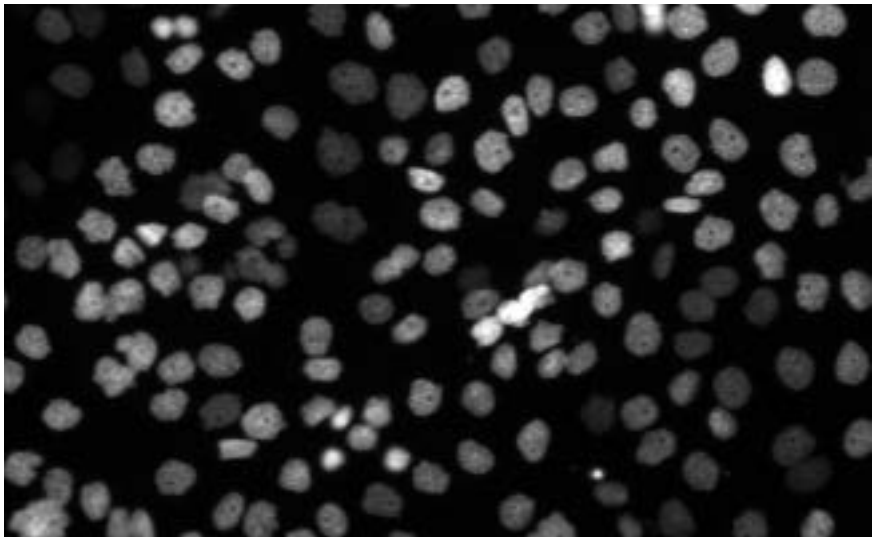
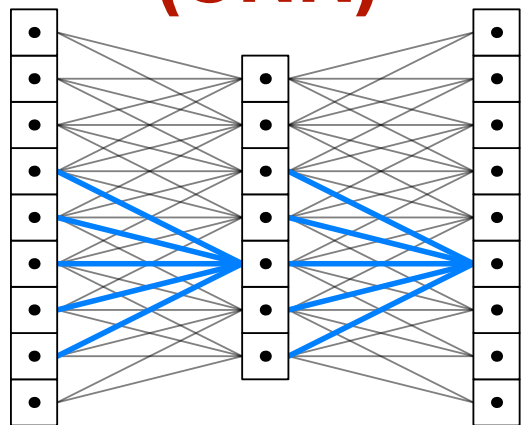
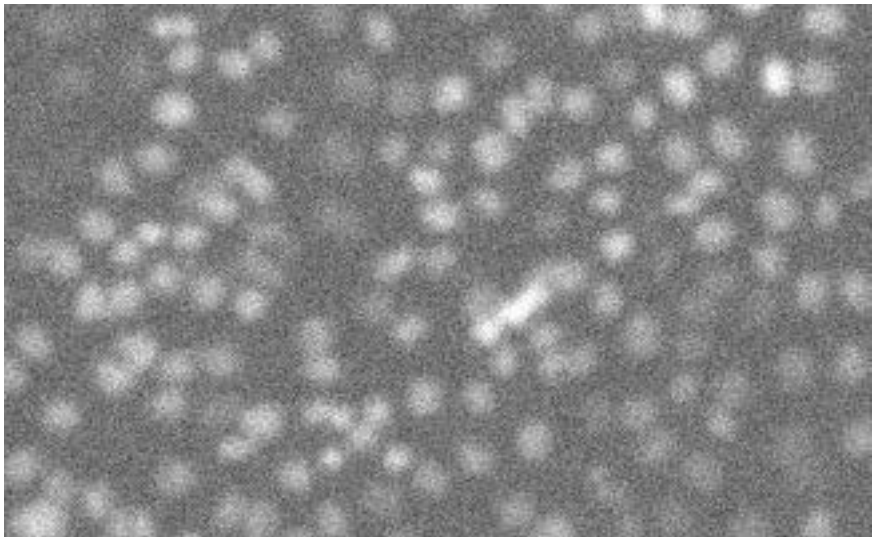
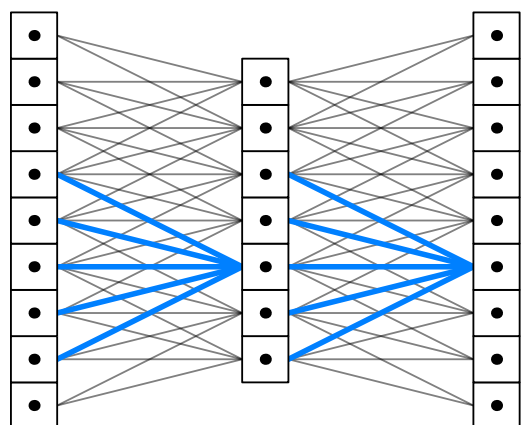
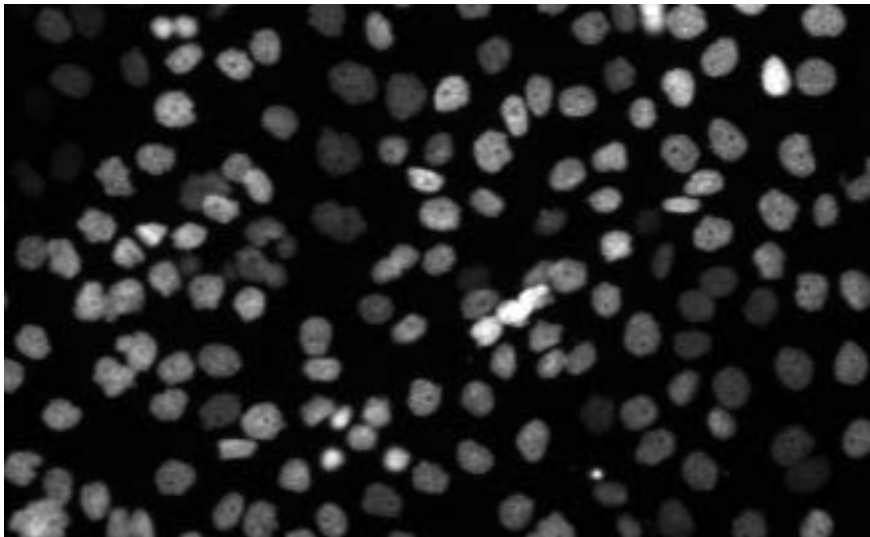


Acquired Image



Pixel level

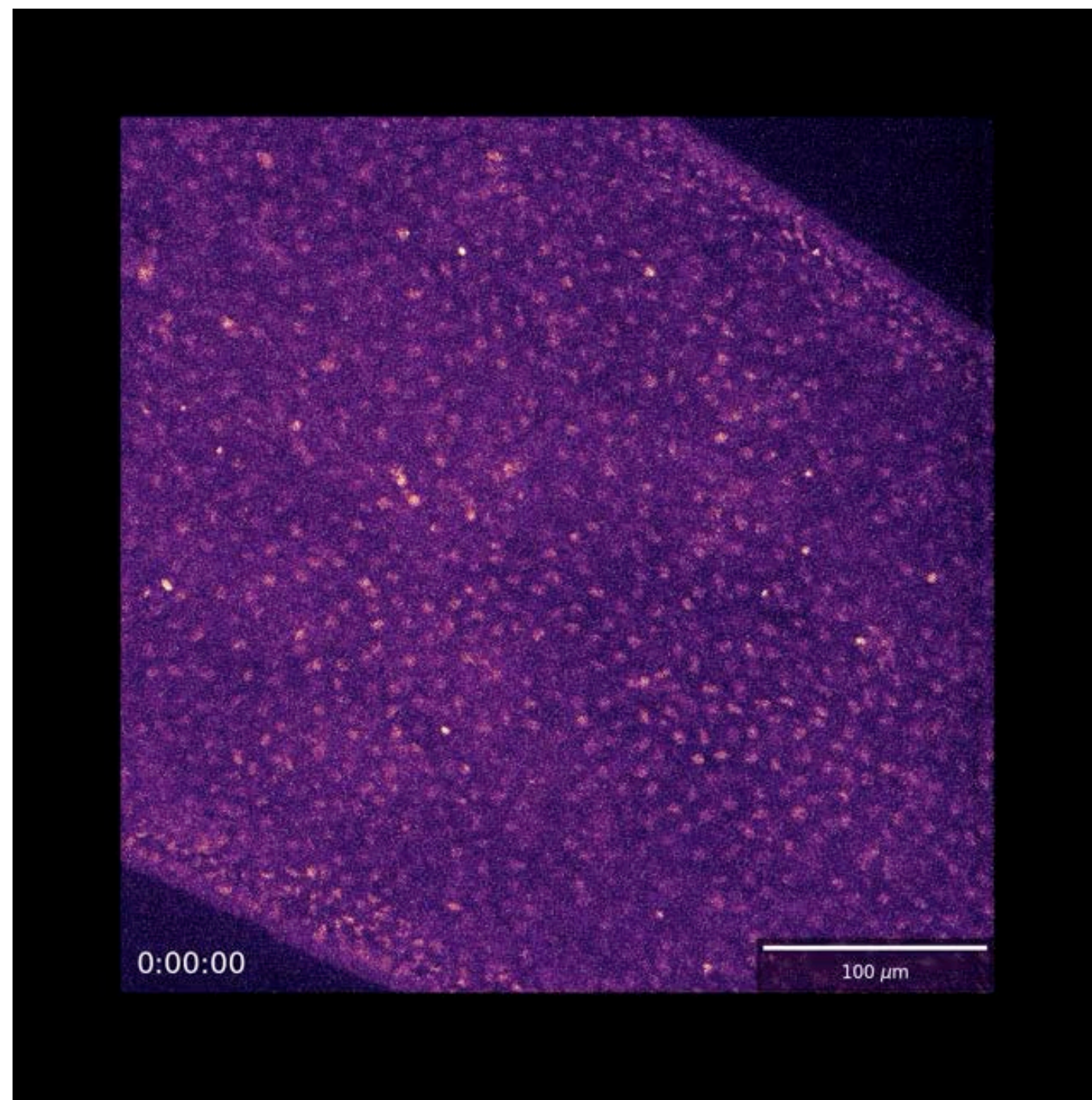
13	7	10	11	18	17	19	6	11	5	12	13
12	11	6	7	12	13	13	8	11	7	10	13
10	9	17	11	17	17	13	19	14	14	16	31
9	6	14	21	20	16	21	17	30	24	18	14
19	14	23	28	18	23	17	29	24	16	31	26
14	19	17	15	24	21	26	41	48	48	48	56
11	11	14	23	33	46	106	111	112	116	108	103
16	16	17	26	41	80	153	188	169	199	203	182
16	12	40	35	77	123	158	223	229	207	240	209
13	25	16	50	112	175	203	227	234	252	222	224
18	18	16	54	131	198	221	233	247	199	219	232
21	18	36	73	126	187	234	256	240	225	236	258

Input		Output	Task
<i>Image</i> 		<i>Discrete-valued class</i> “wildtype” or “mutant”	Classification
<i>Image</i> 	Convolutional Neural Networks (CNN) 	<i>Label Mask (class per pixel)</i> 	(Dense) Segmentation Object Detection
<i>Image</i> 		<i>Image</i> 	(Dense) Regression Image2Image Image Restoration

Extraction of quantitative biological information from digitised camera images (Computer Vision)

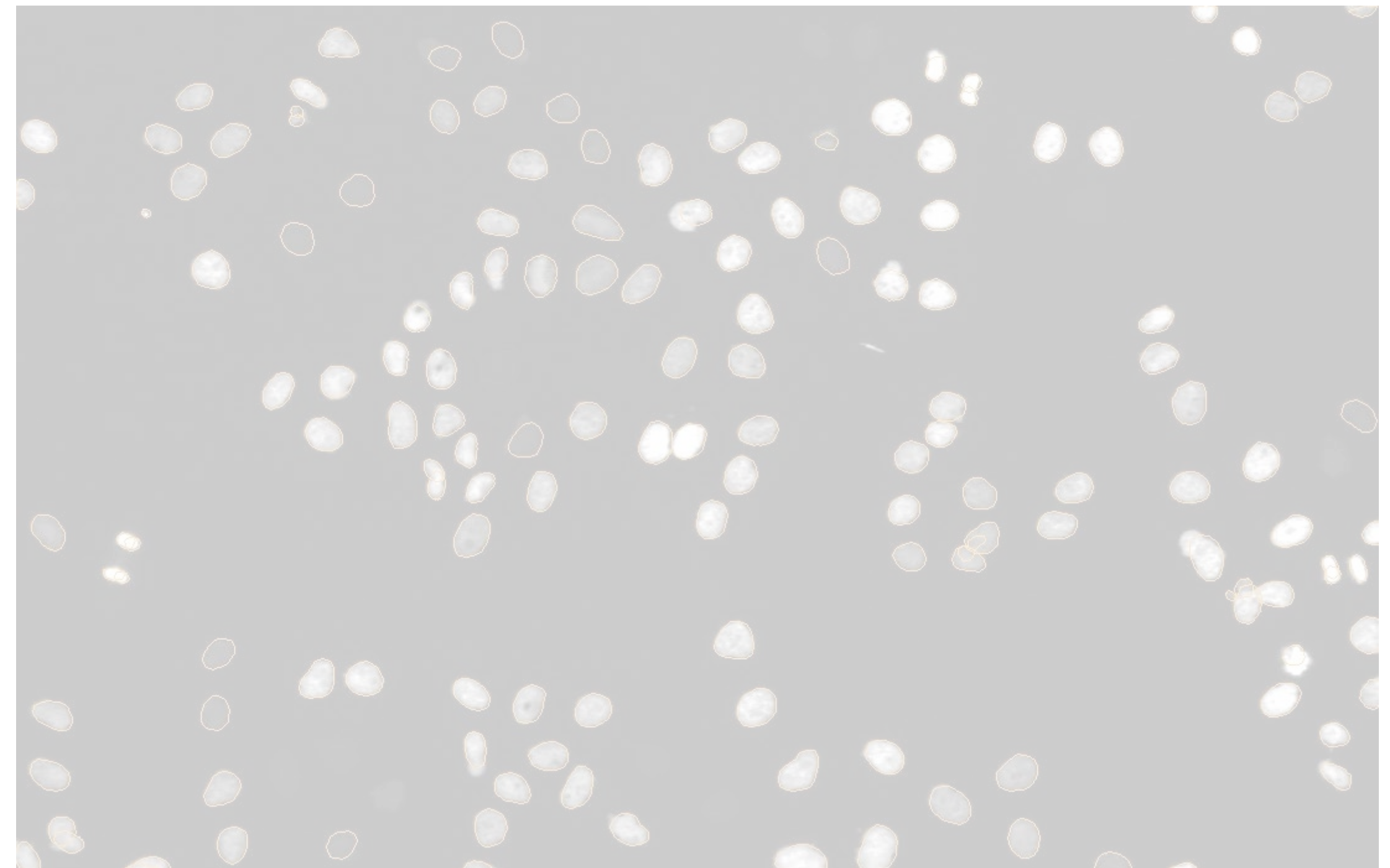
Image Processing/Restoration

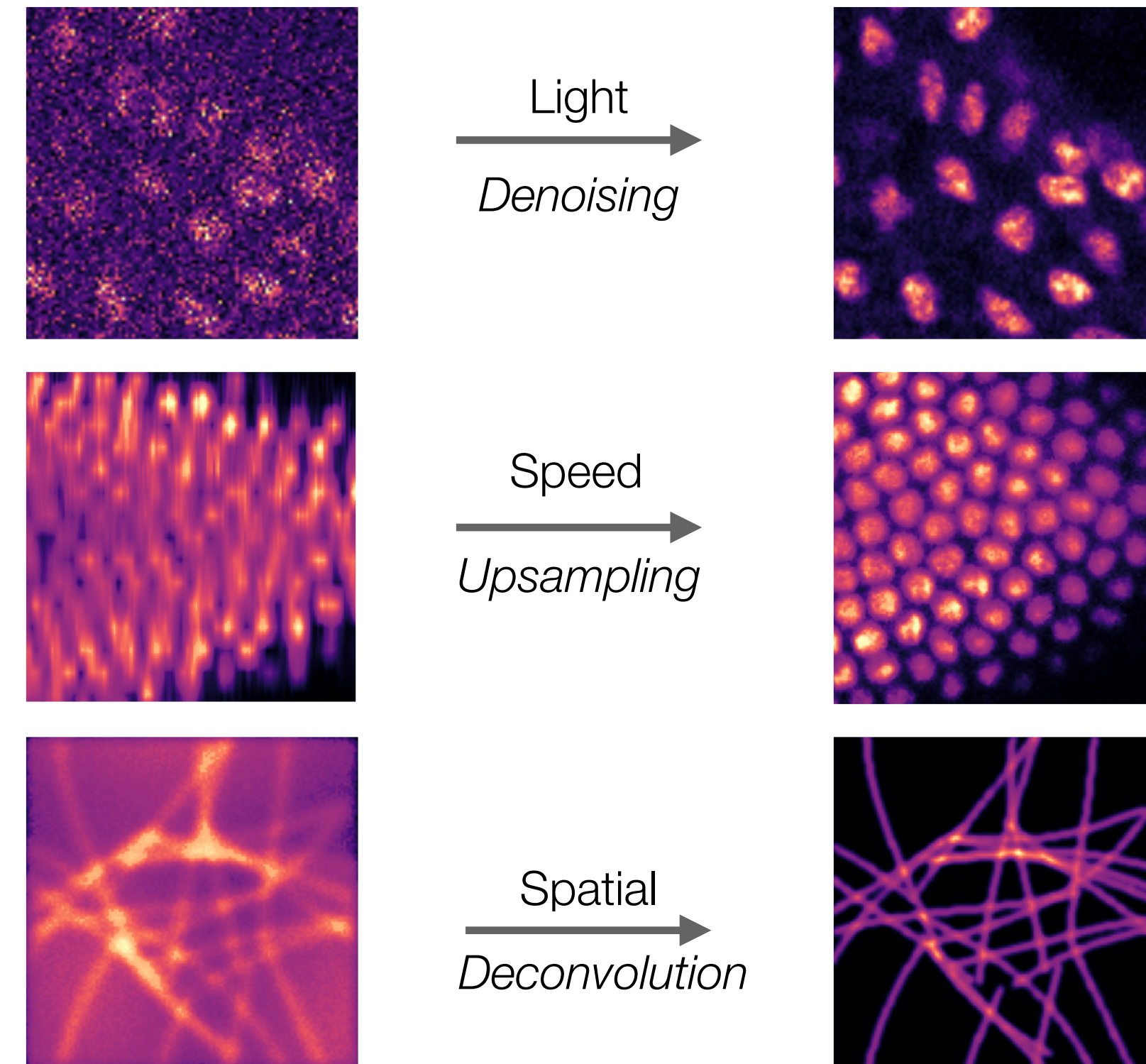
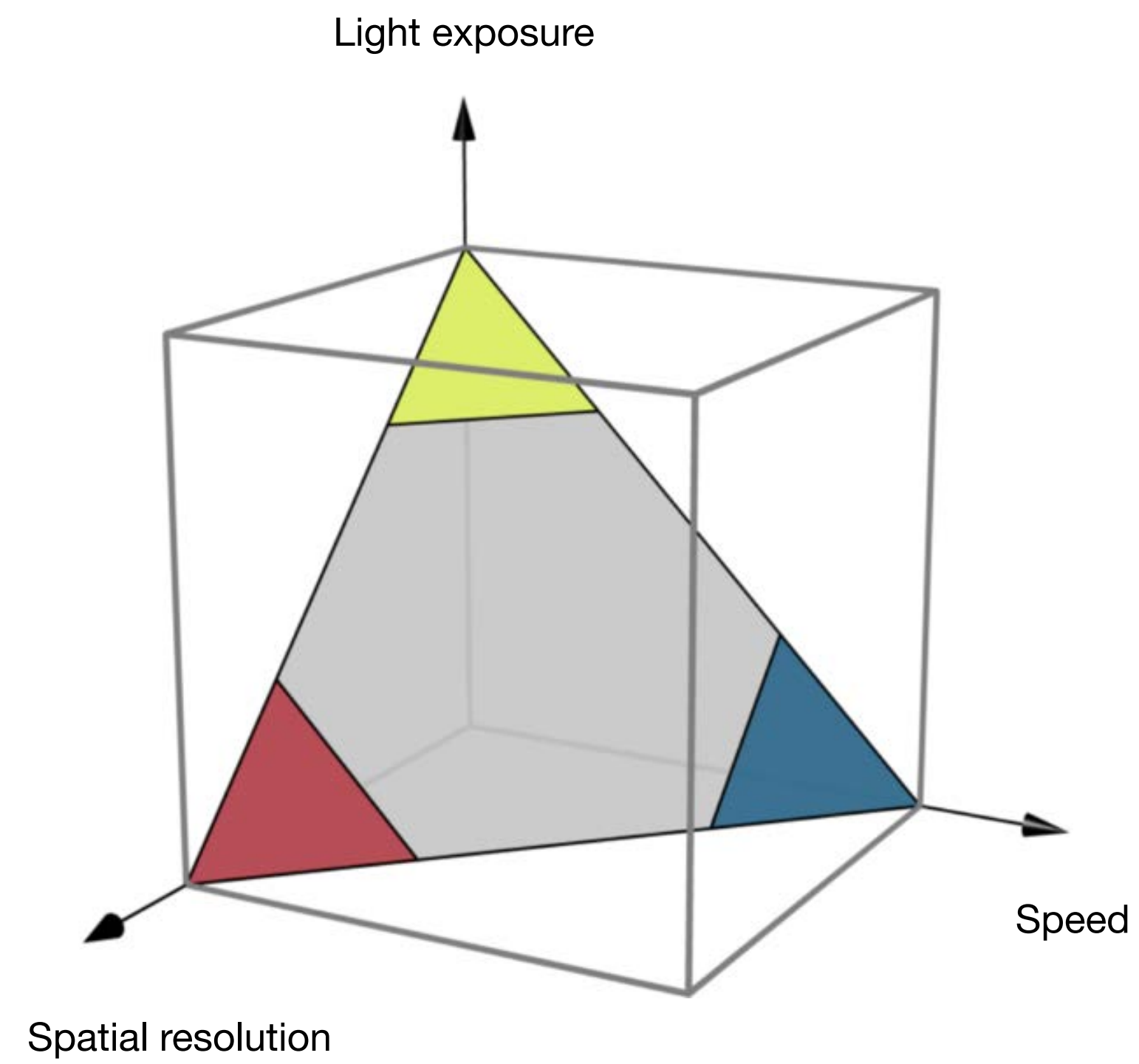
Remove camera noise, remove blur...



Object Detection and Segmentation

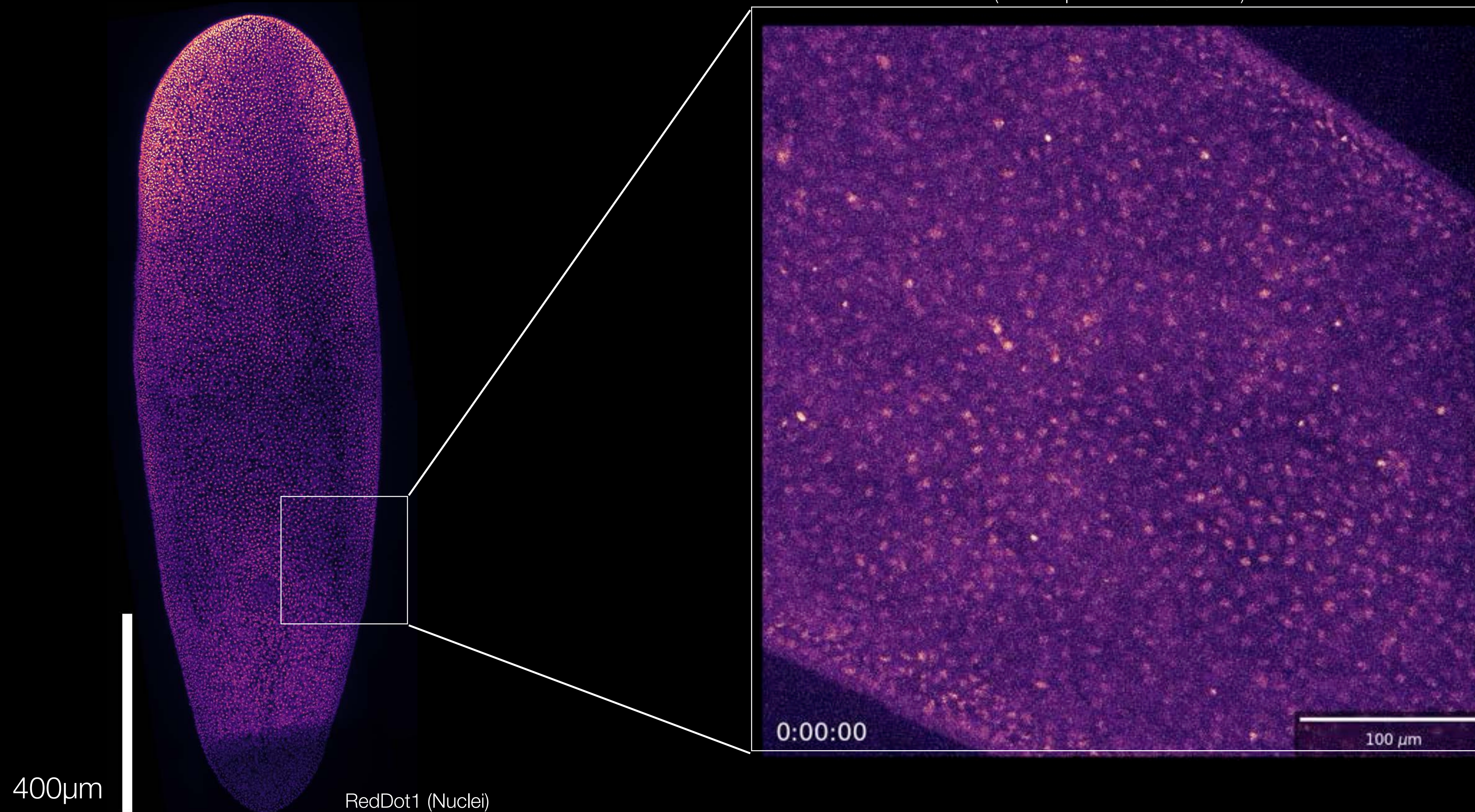
Find contours of cells, track cells over time, ...





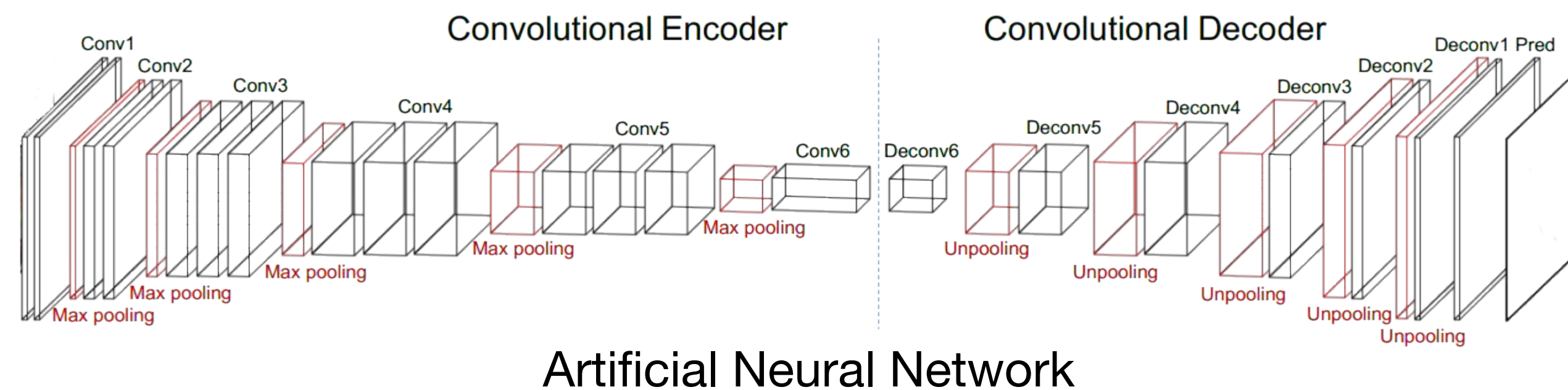
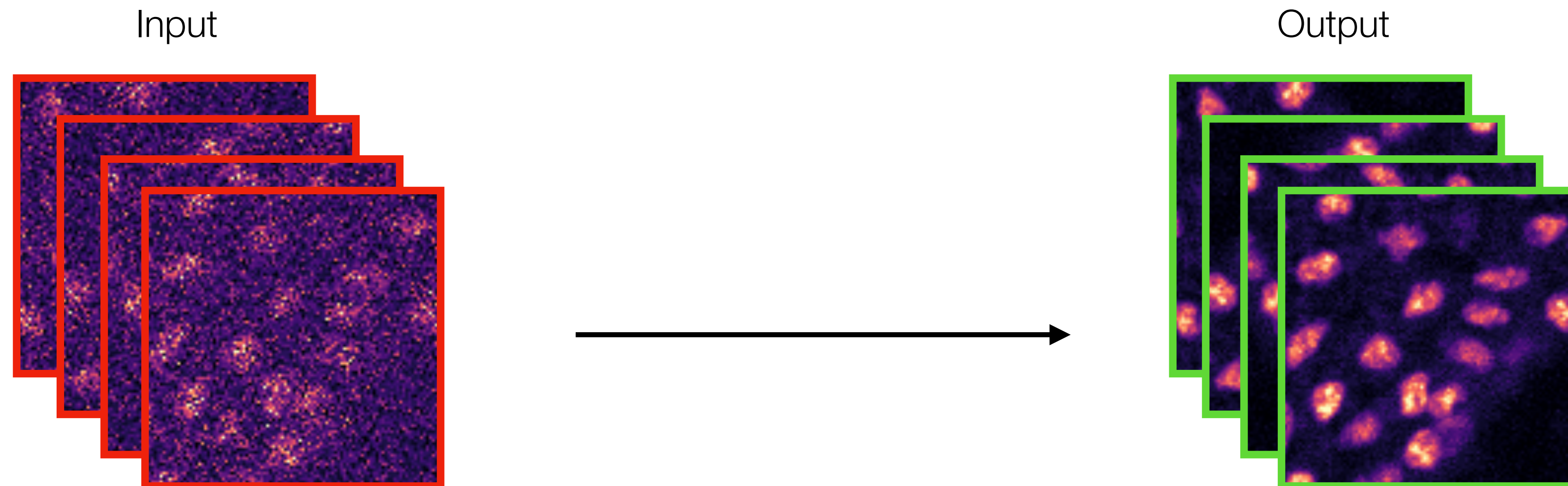
Planaria (Flatworm)

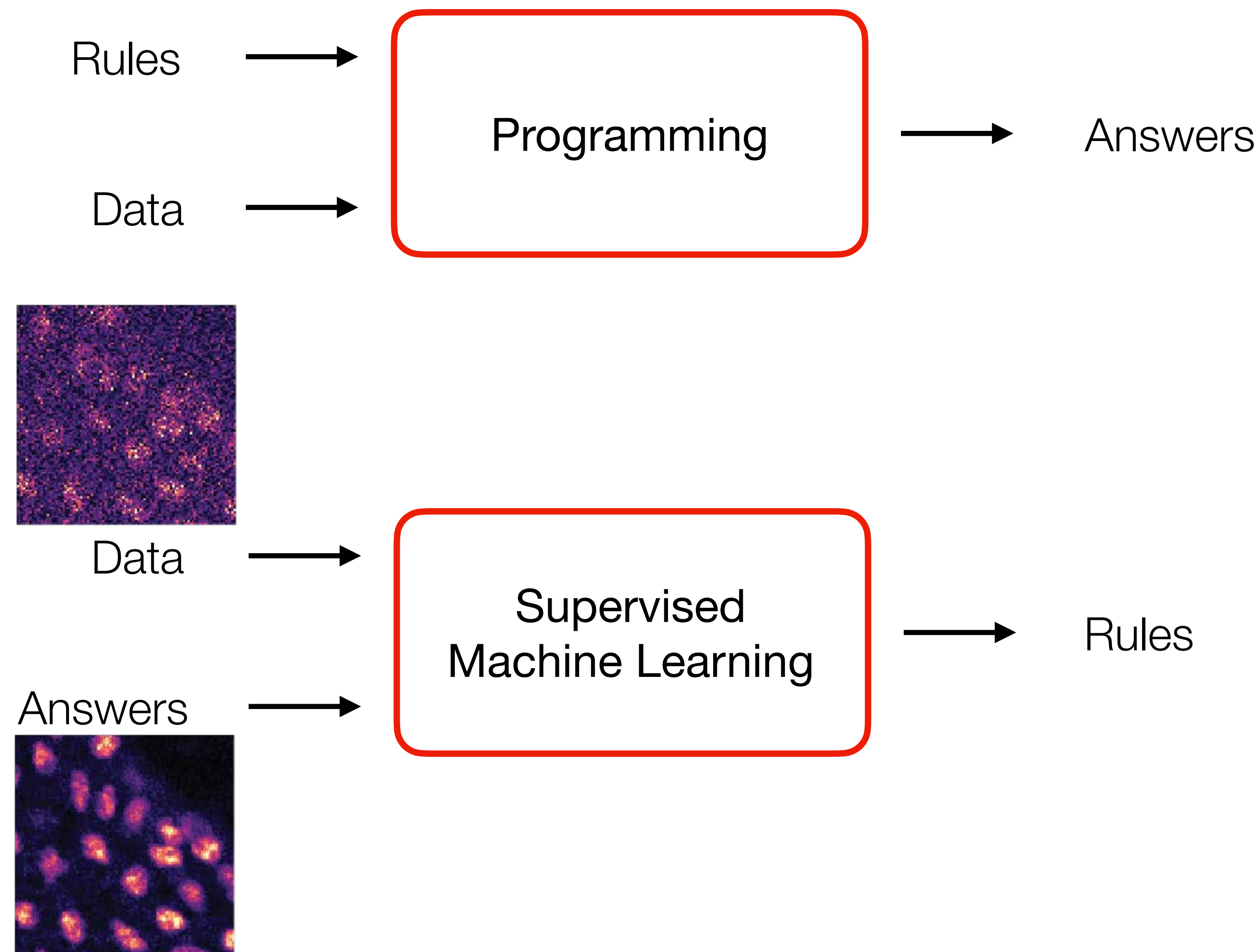
Low light dosage / low SNR
(laser/exposure: 0.3%/10ms)

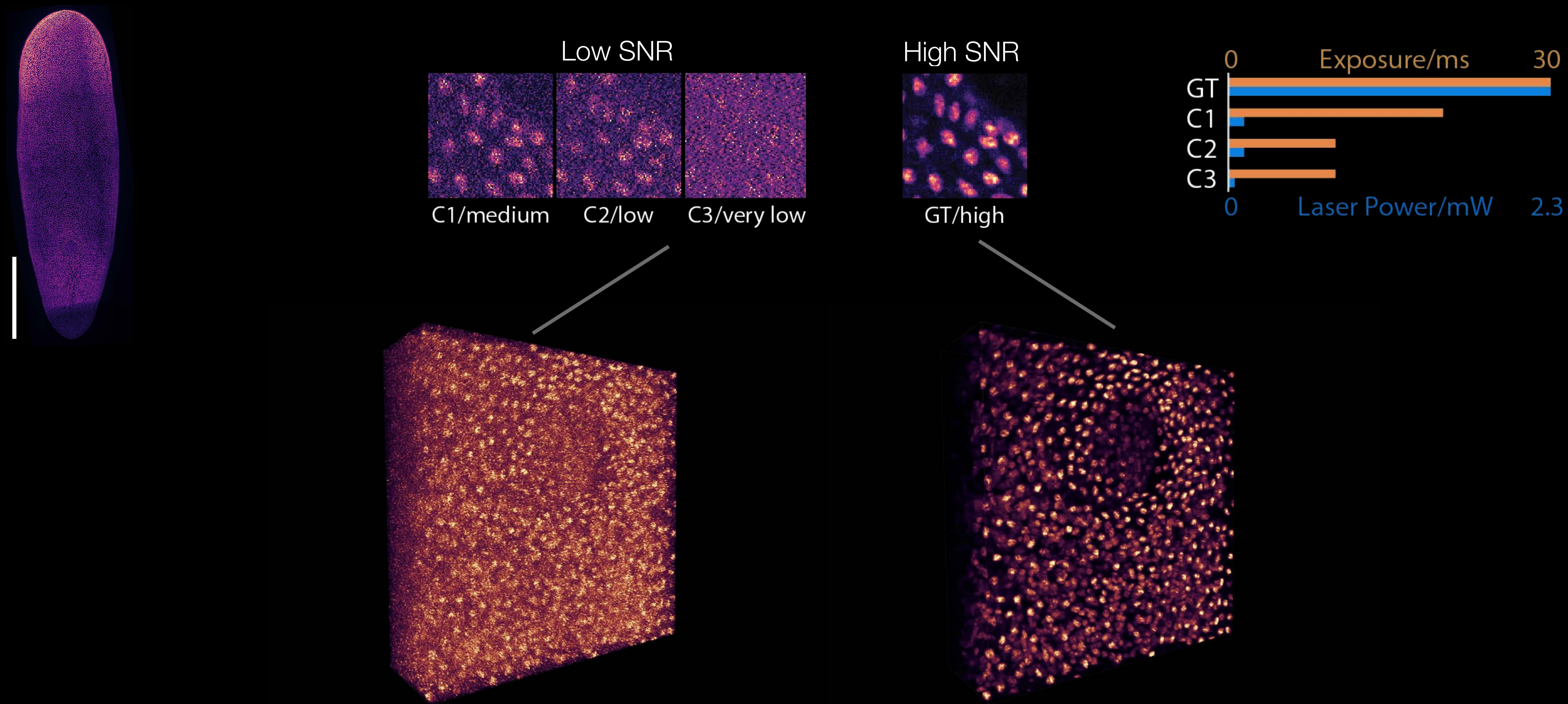


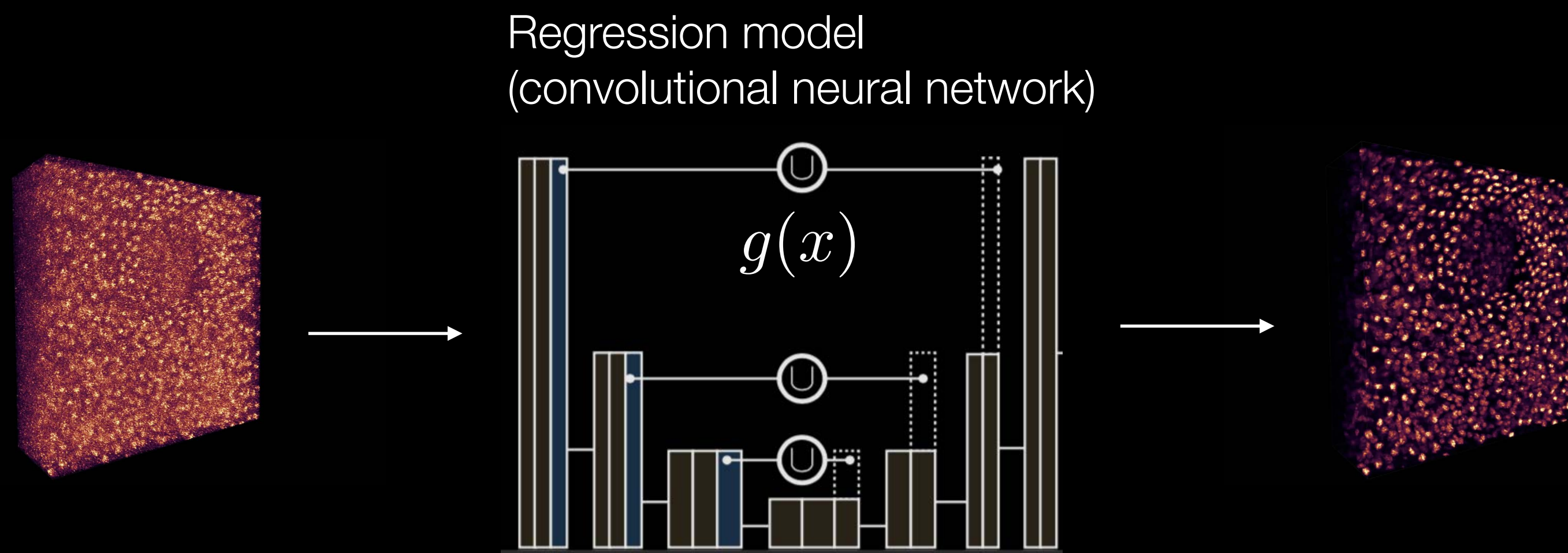
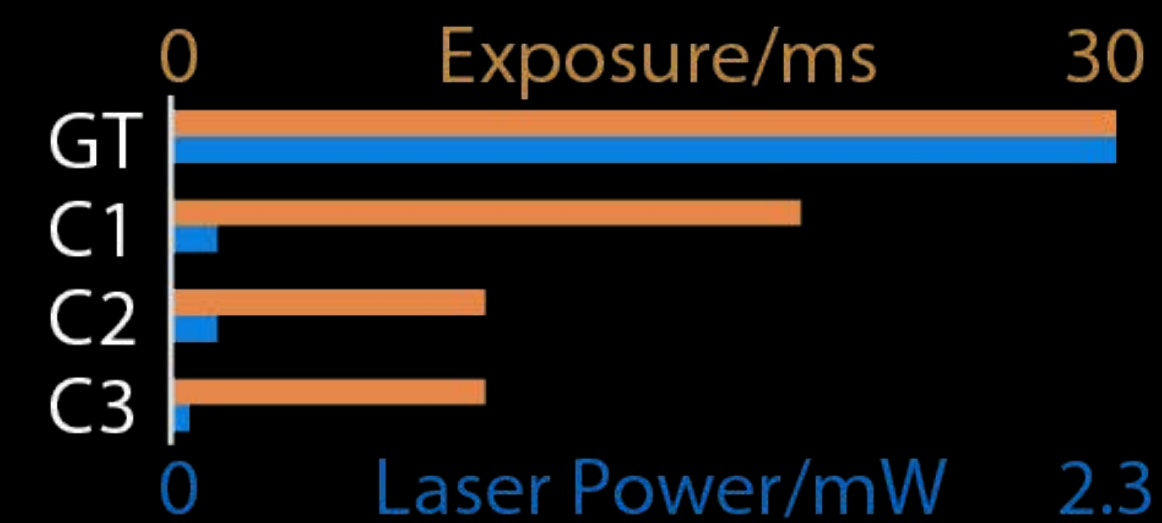
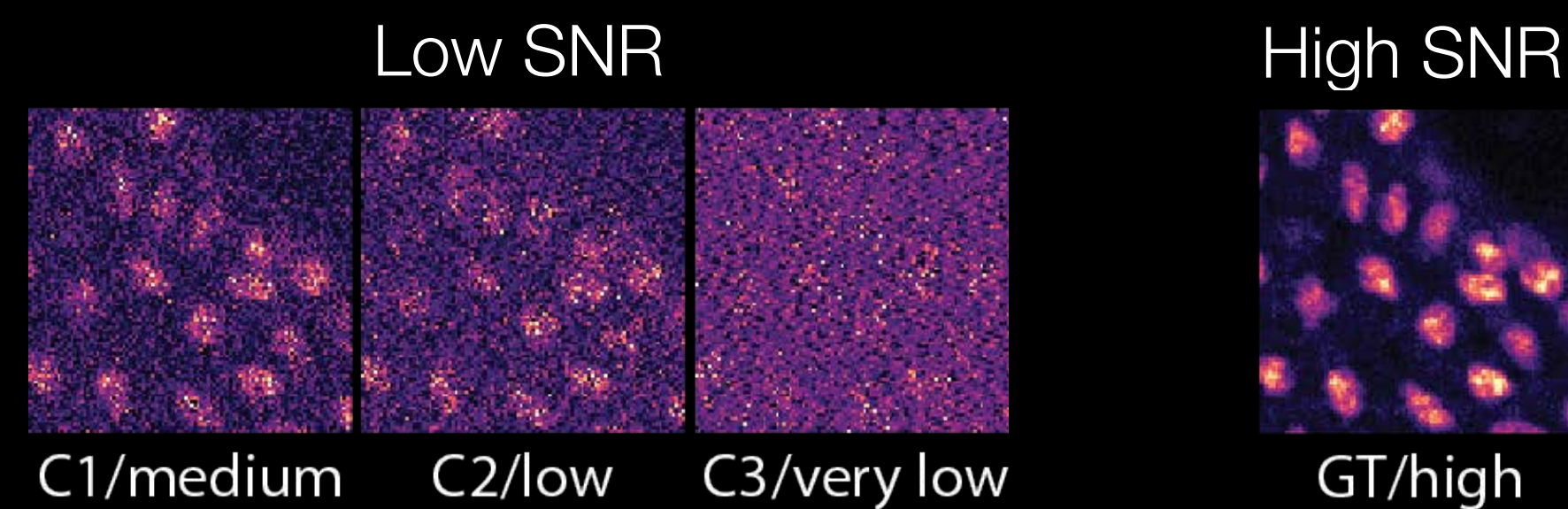
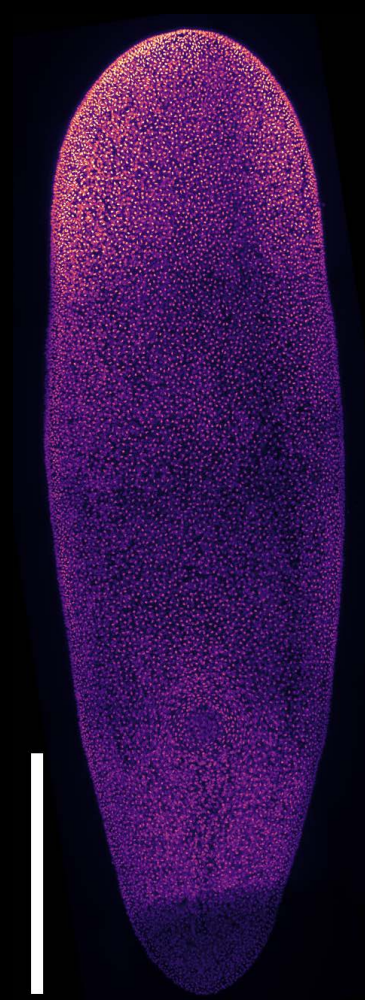
Data from Tobias Boothe

Pairwise training data of input/output pairs
Learn a function that implements the mapping noisy $\rightarrow$ clean





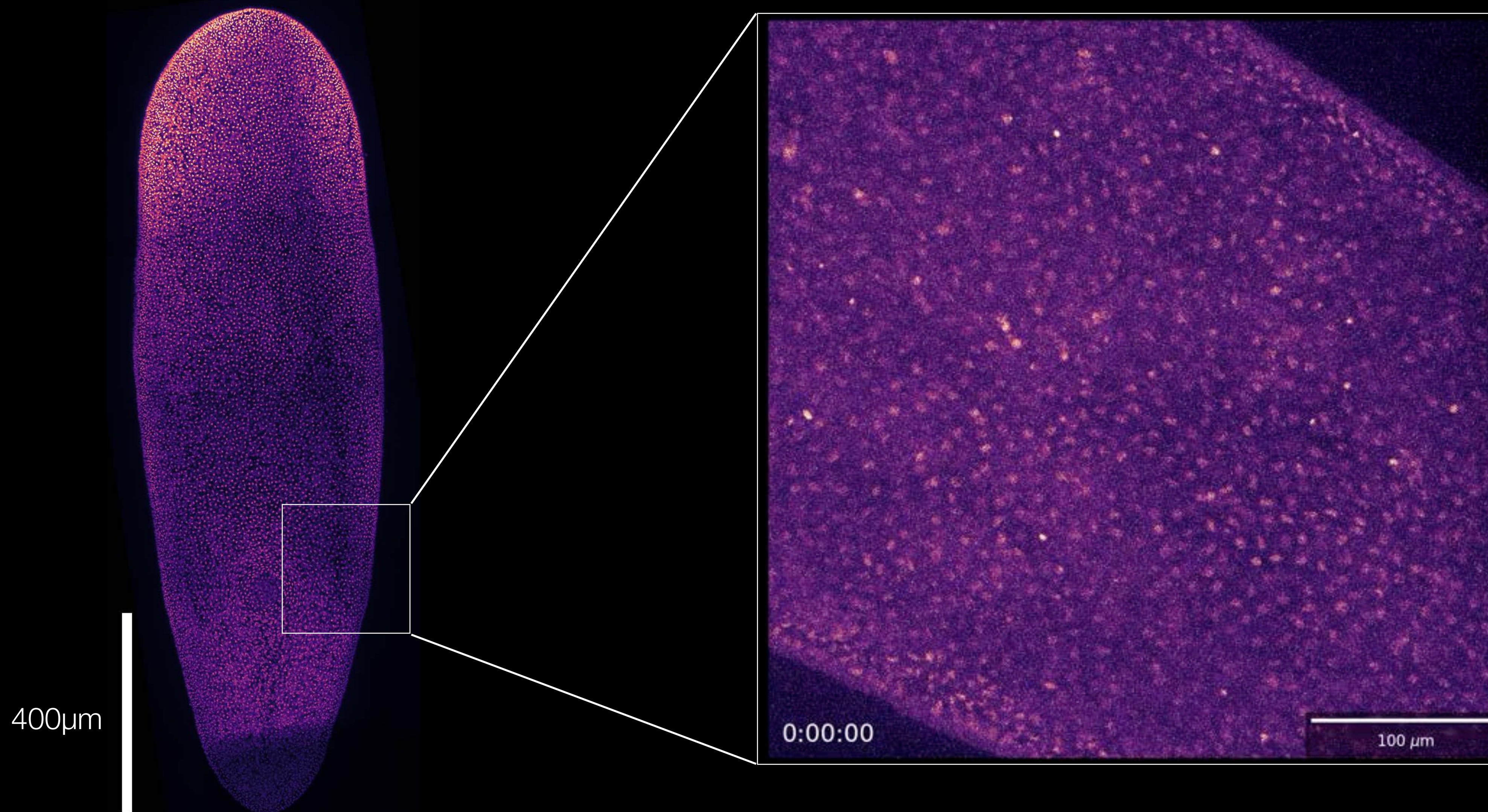


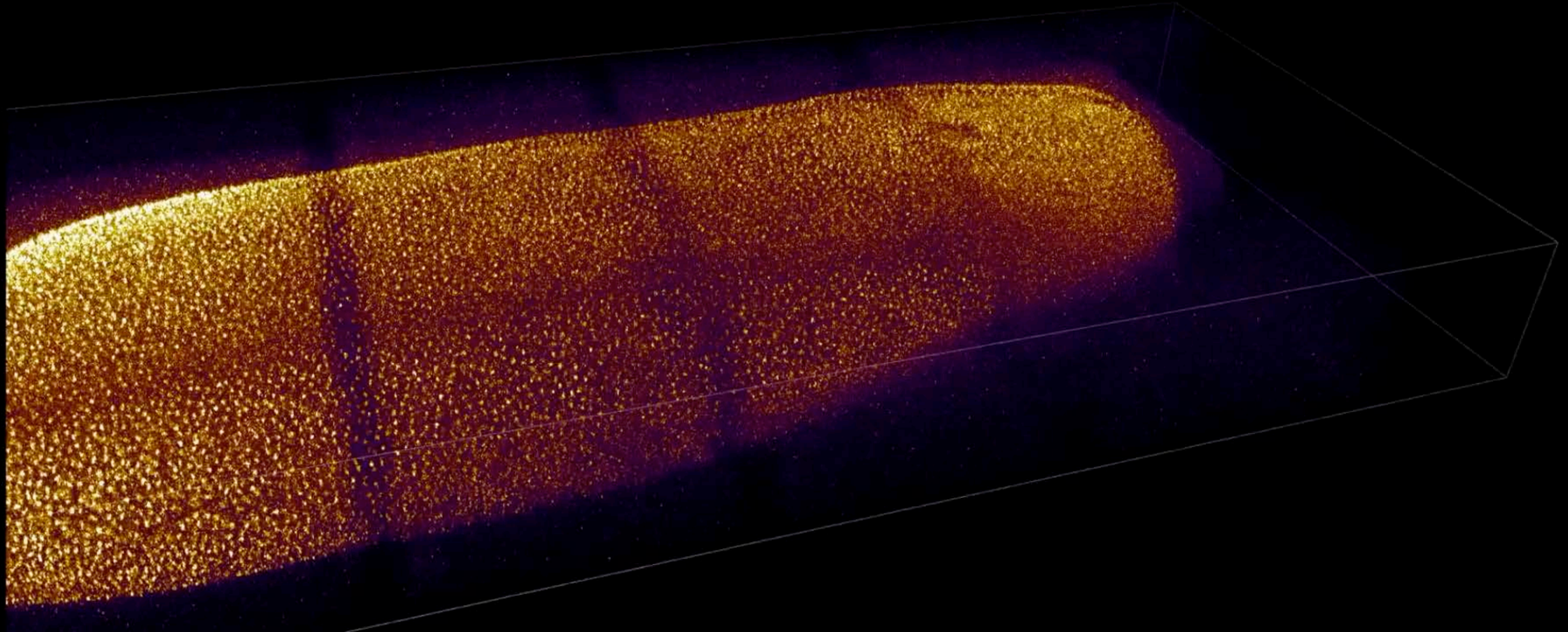


- Trained on 96 pairs (160GB)
- Training time ~ 8hrs

Planaria (Flatworm)

Restoration

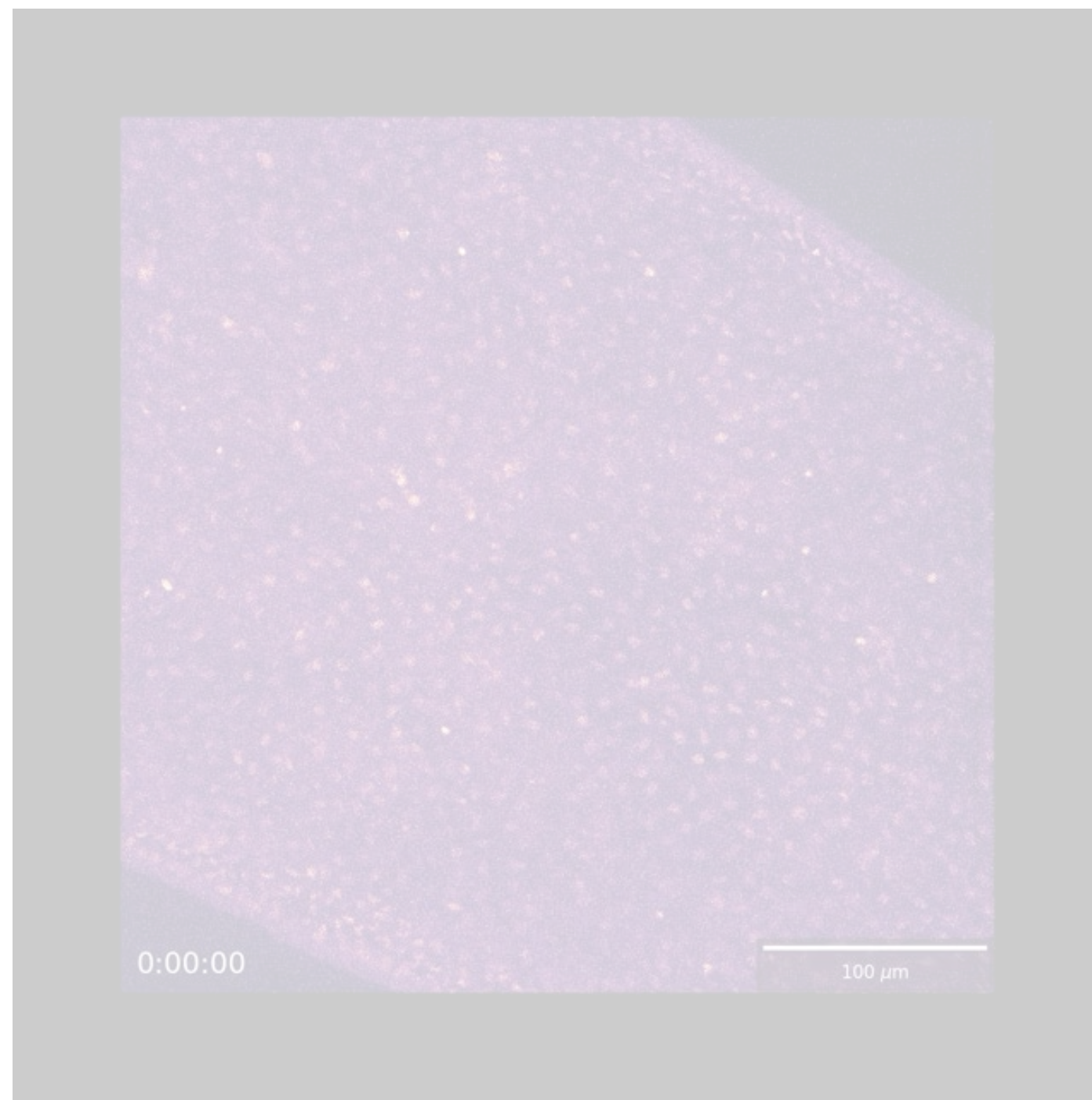




Extraction of quantitative biological information from digitised camera images (Computer Vision)

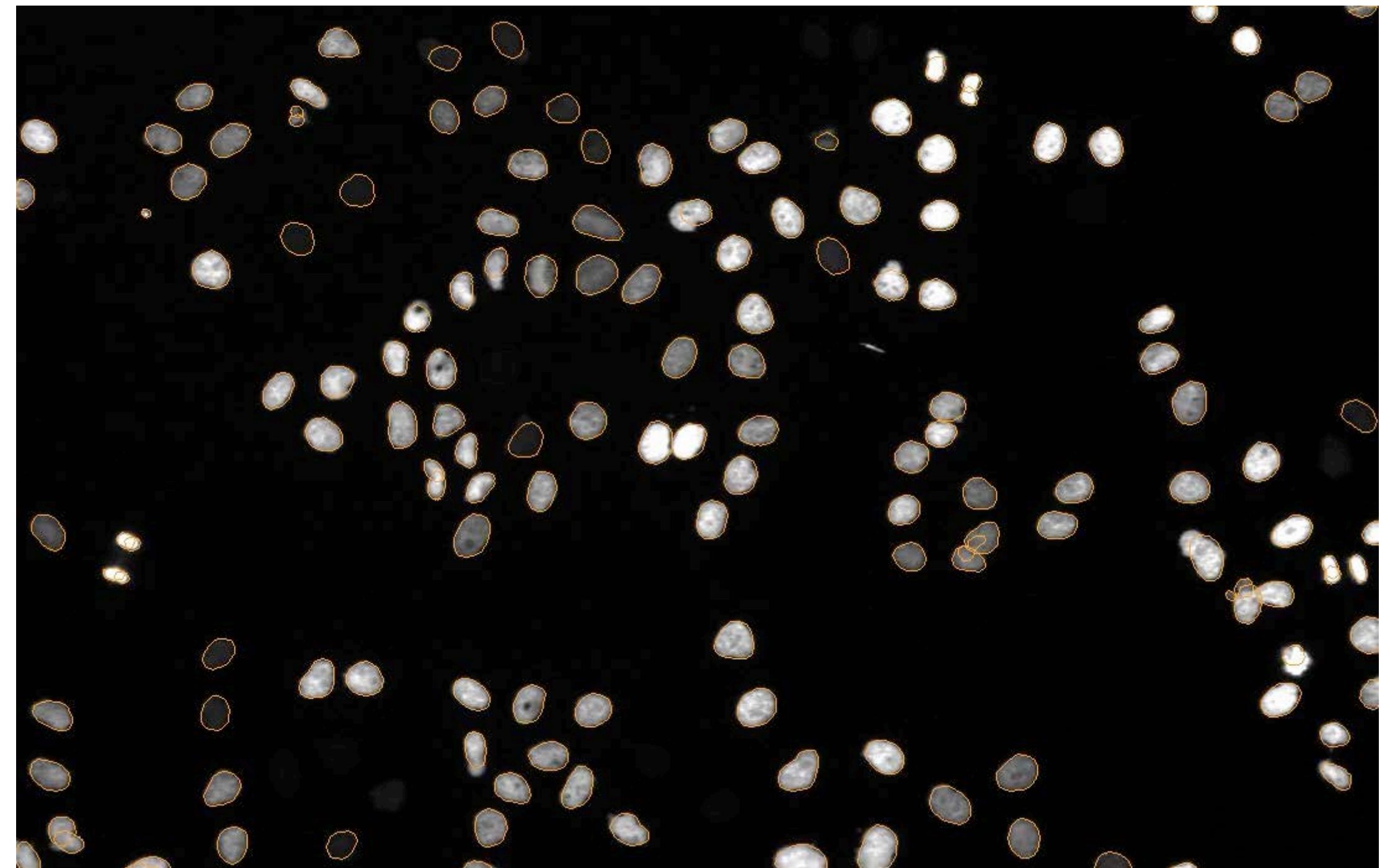
Image Denoising/Restoration

Remove camera noise, remove blur...



Object Detection and Segmentation

Find contours of cells, detect spots, track cells over time, ...



Single Object

Image
Classification

Cat

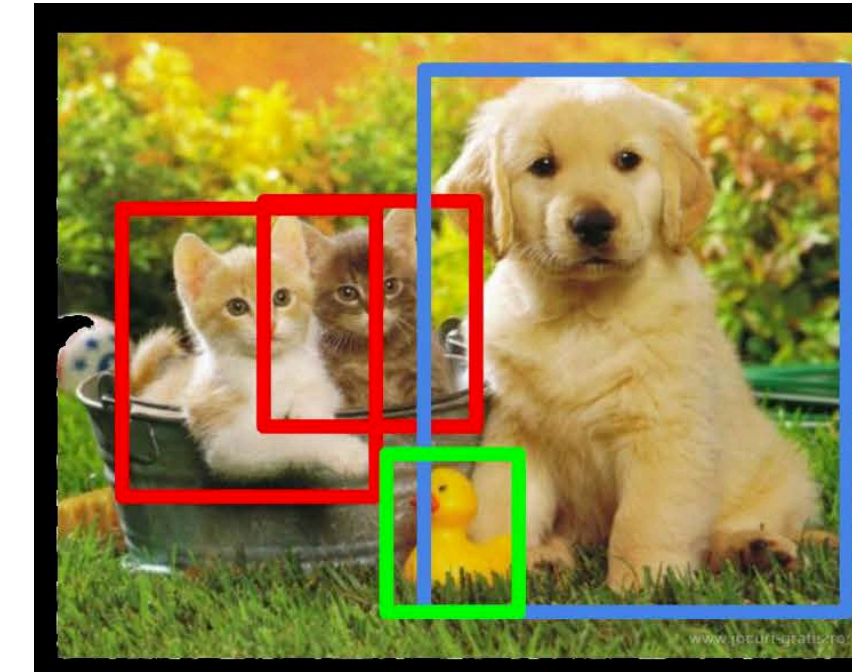
Object
Detection (Localization)

Cat

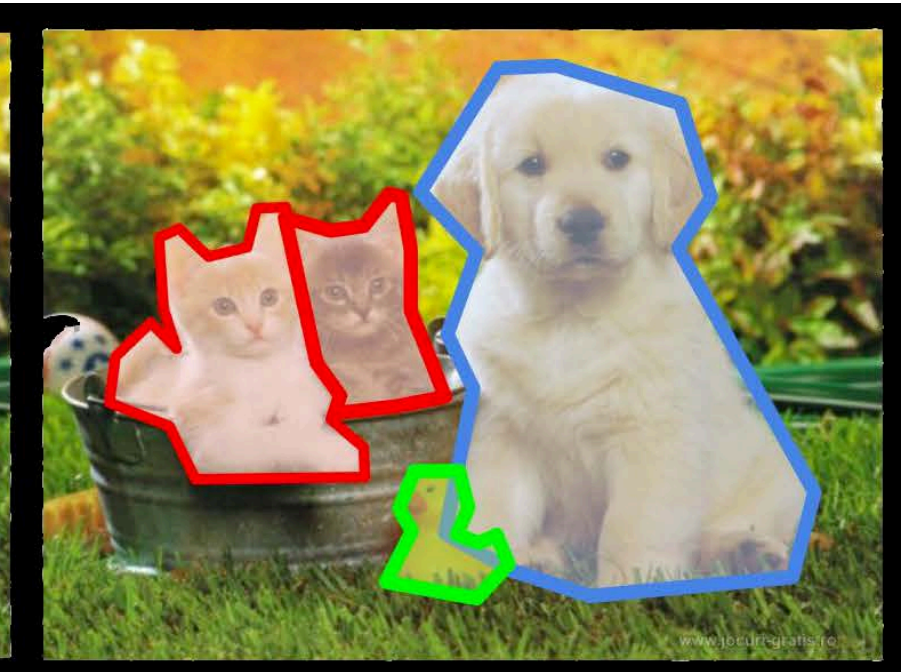
Dense/Semantic
Segmentation

Cat

Multiple Objects

Object
Detection

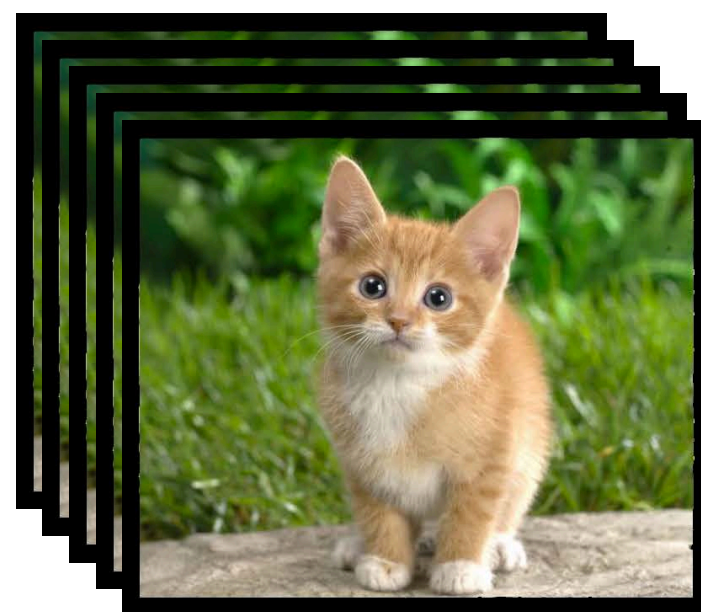
Cat, Dog, Duck

Instance
Segmentation

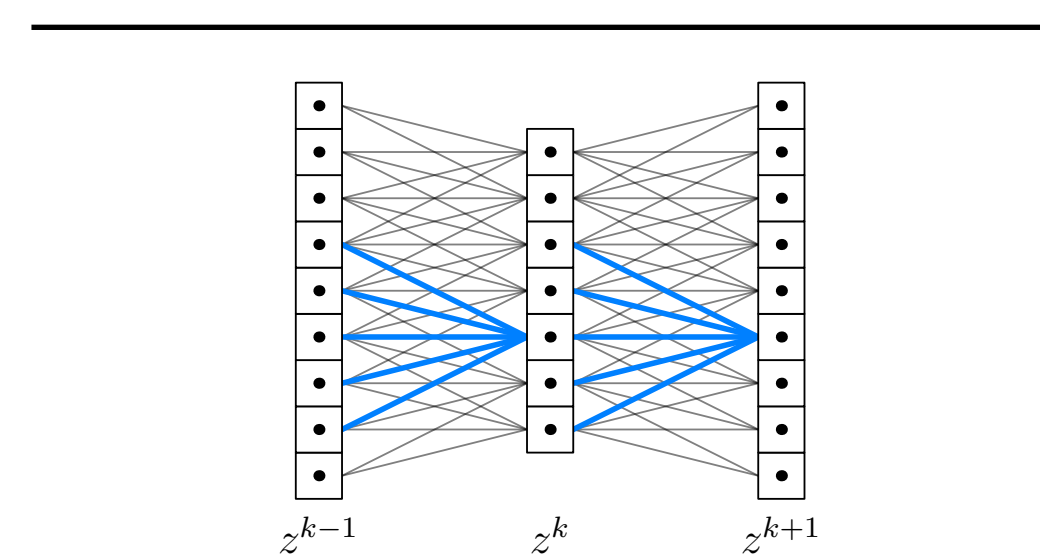
Cat, Dog, Duck

Currently the most successful paradigm: Supervised deep learning

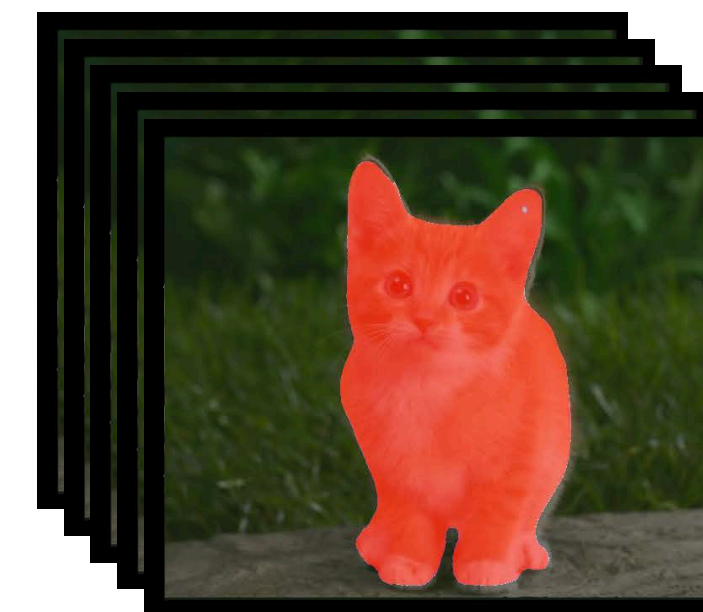
Input



Deep Neural Network



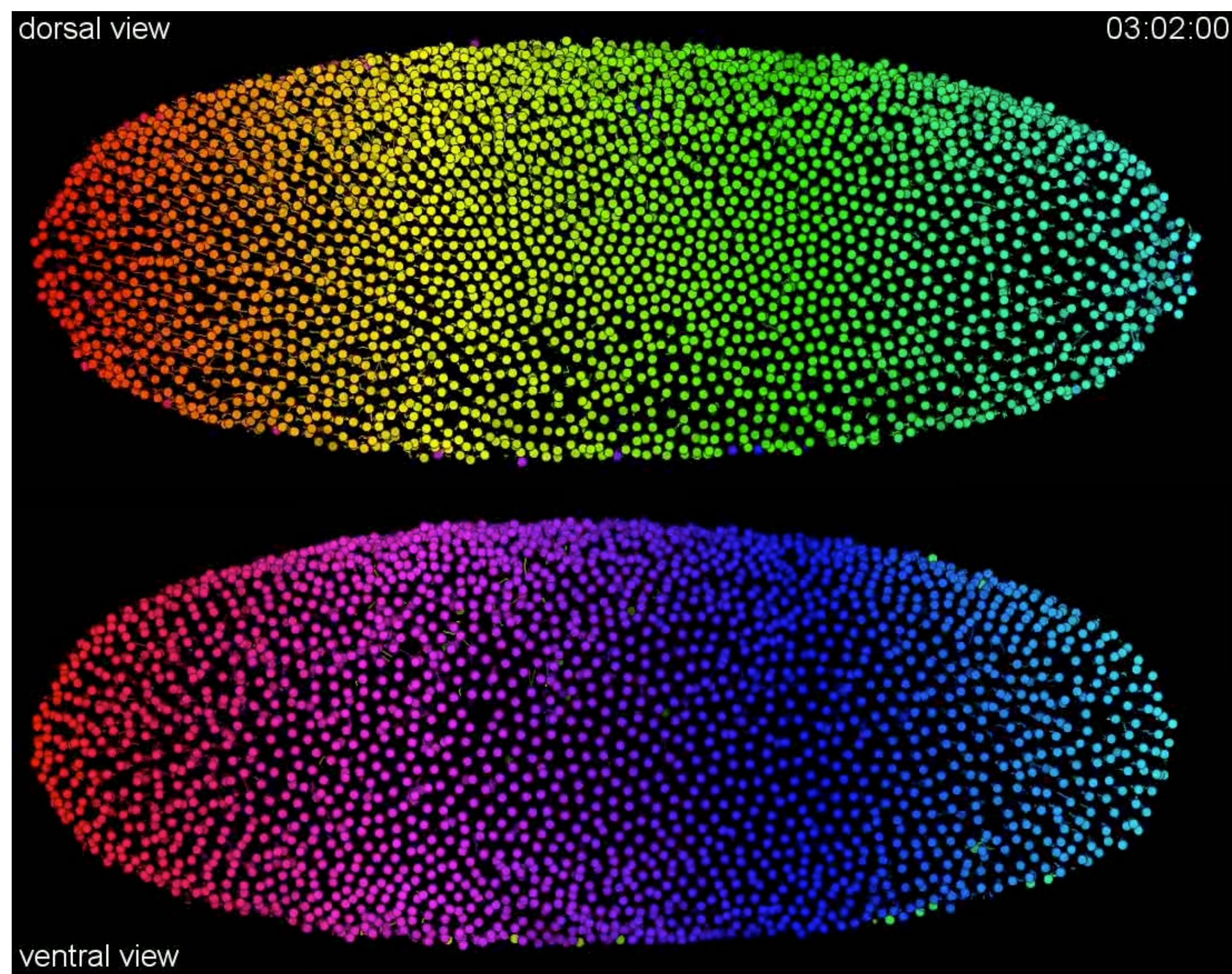
Output



U-Net Ronneberger et al (2015)
YOLO Redmond et al (2016)
Mask-RCNN He et al (2017)

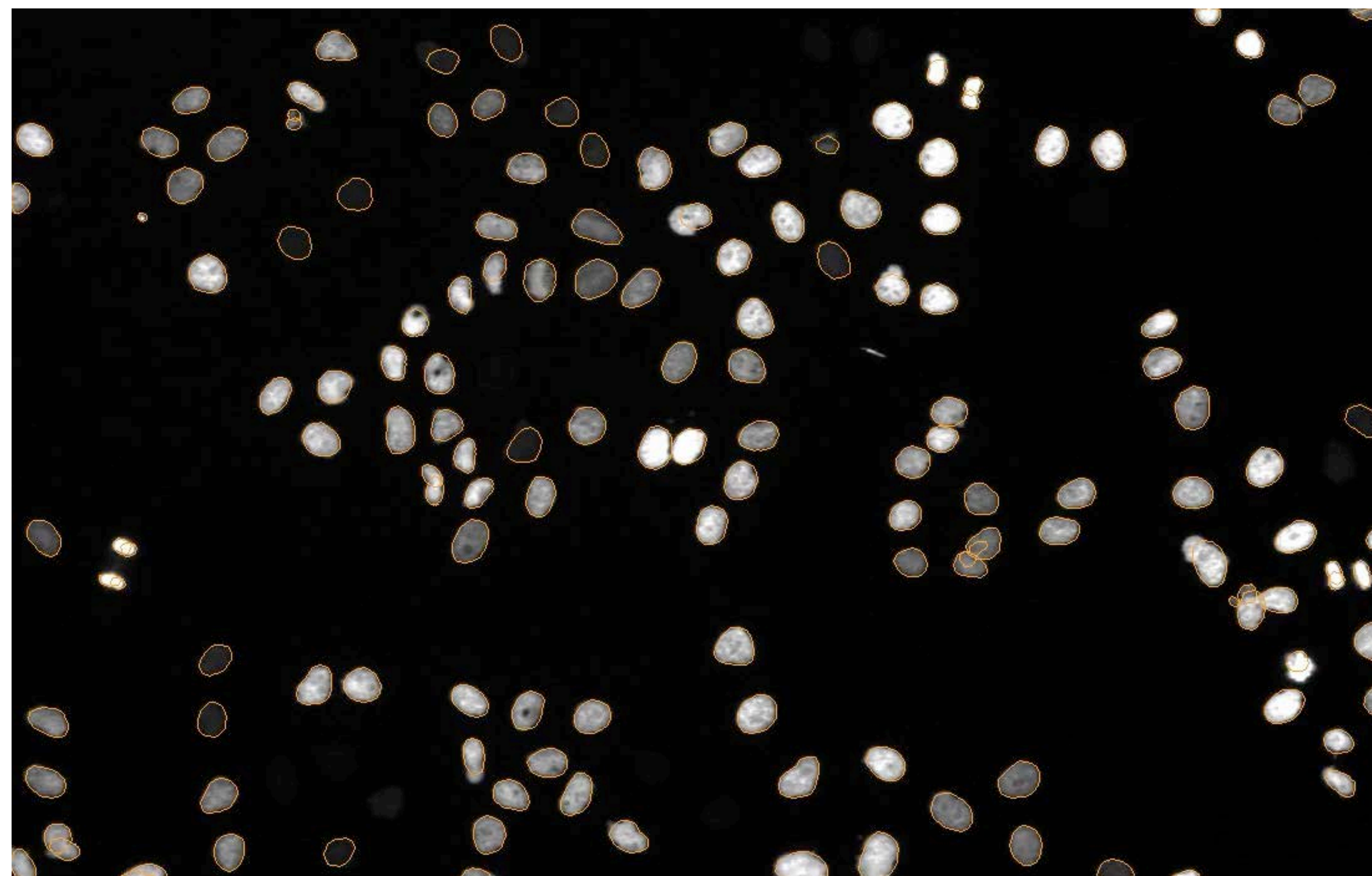
Deep Neural Networks trained on annotated training data (ground truth, GT)

Detection + Tracking

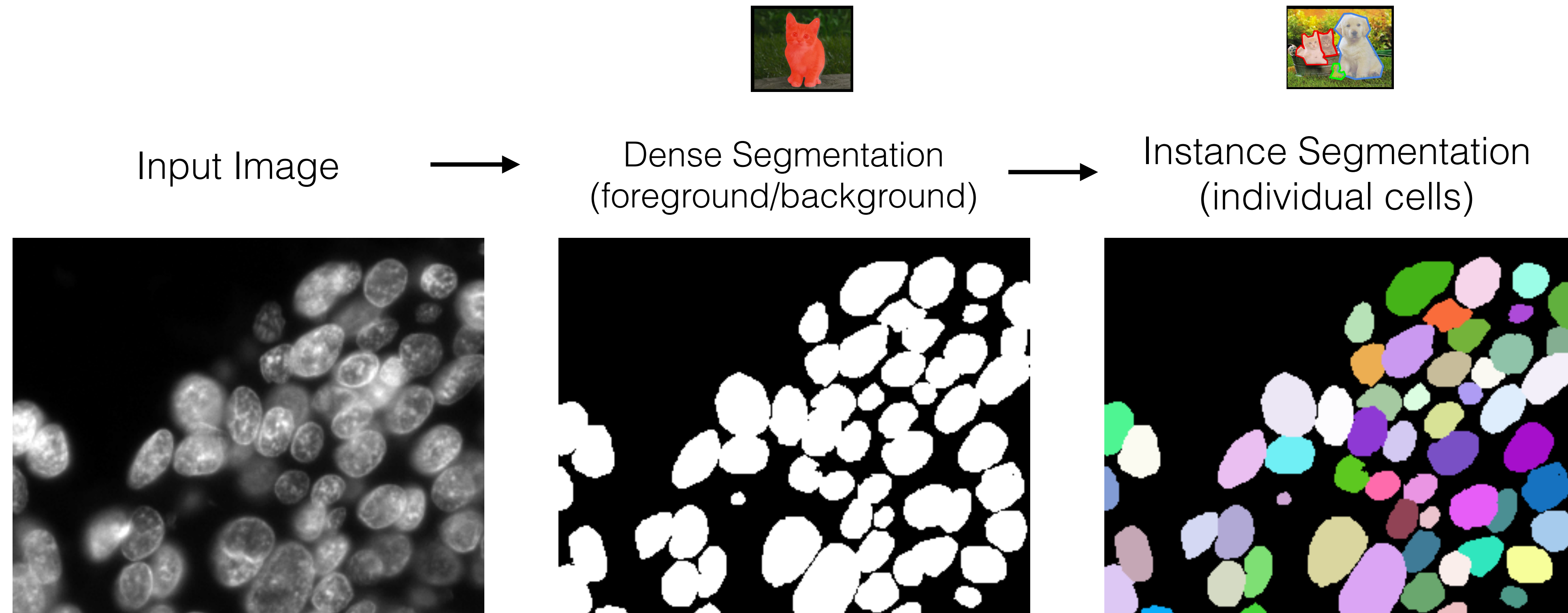


Drosophila Histone marker
Amat et al (2014)

Segmentation



HeLa cells stably expressing H2b-GFP
Data from <http://celltrackingchallenge.net/>

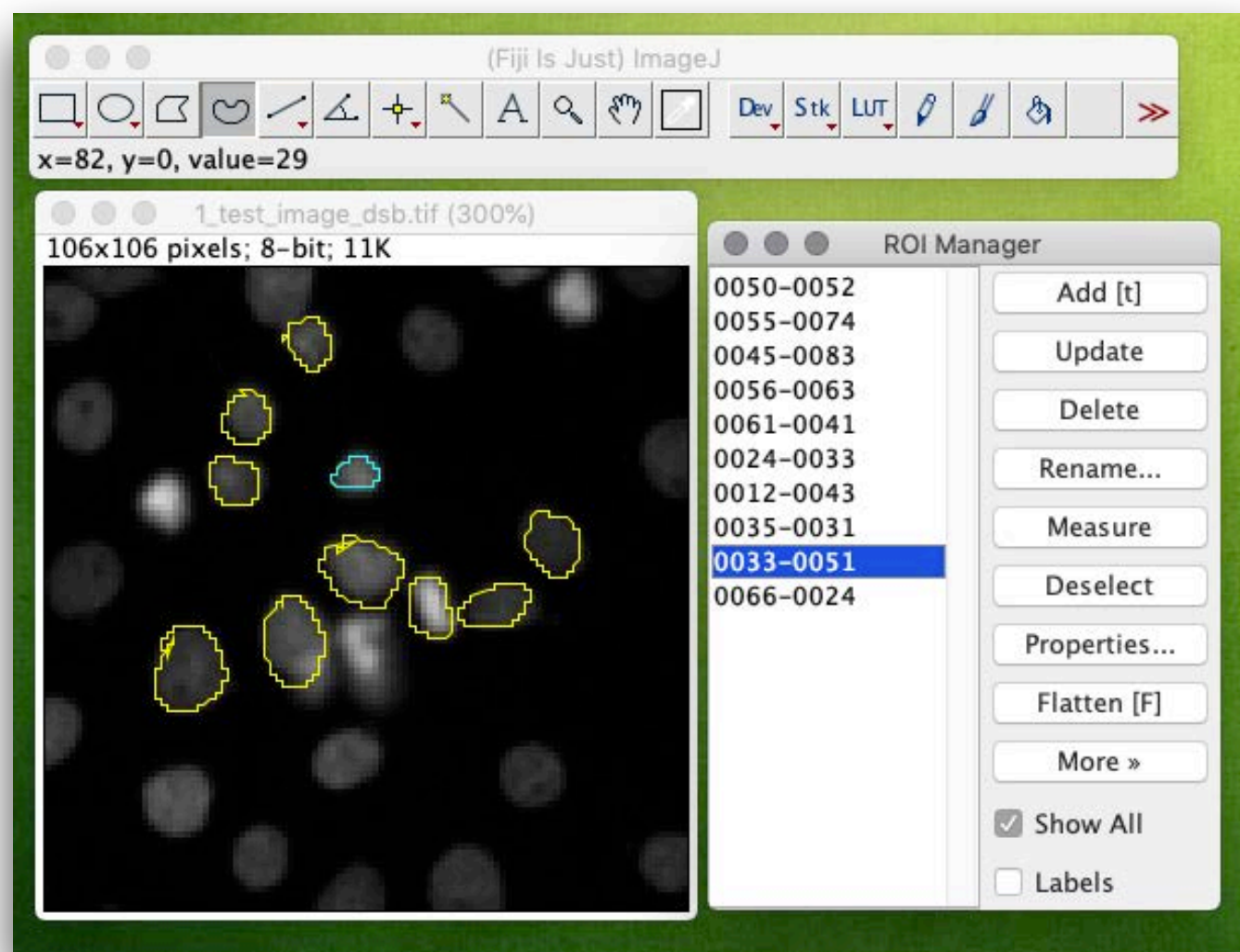


Challenges: many crowded objects, noisy images,
different from natural images (so knowledge cannot be directly transferred)



Fiji/ImageJ

- Draw Roi per object (Roi-Manager)
- Convert to label mask (e.g. see [here](#))



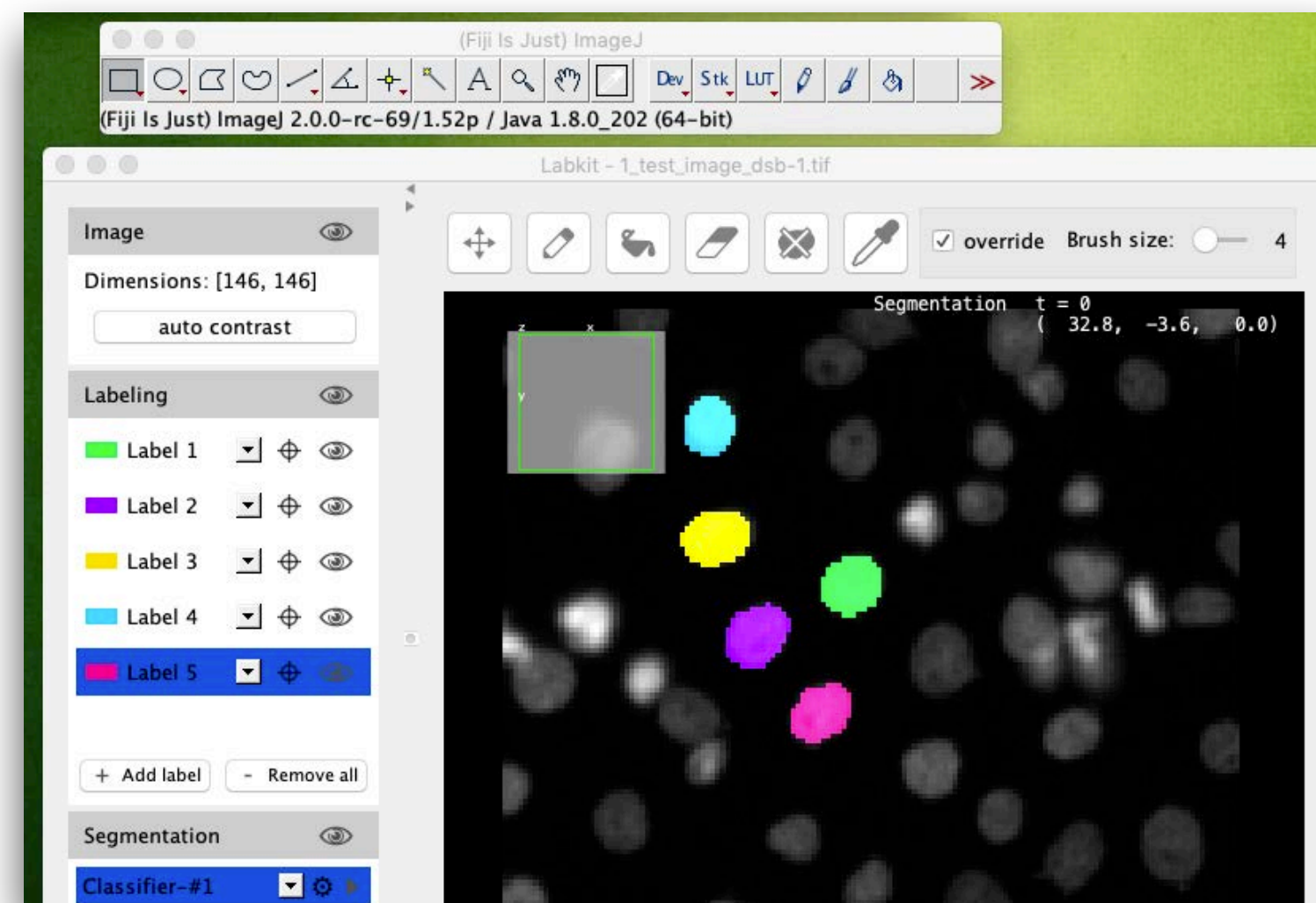
<http://fiji.sc/>

Schindelin et al. (2012)
Schneider, Rasband et al (2012)



Fiji + LabKit

- Directly draw label mask



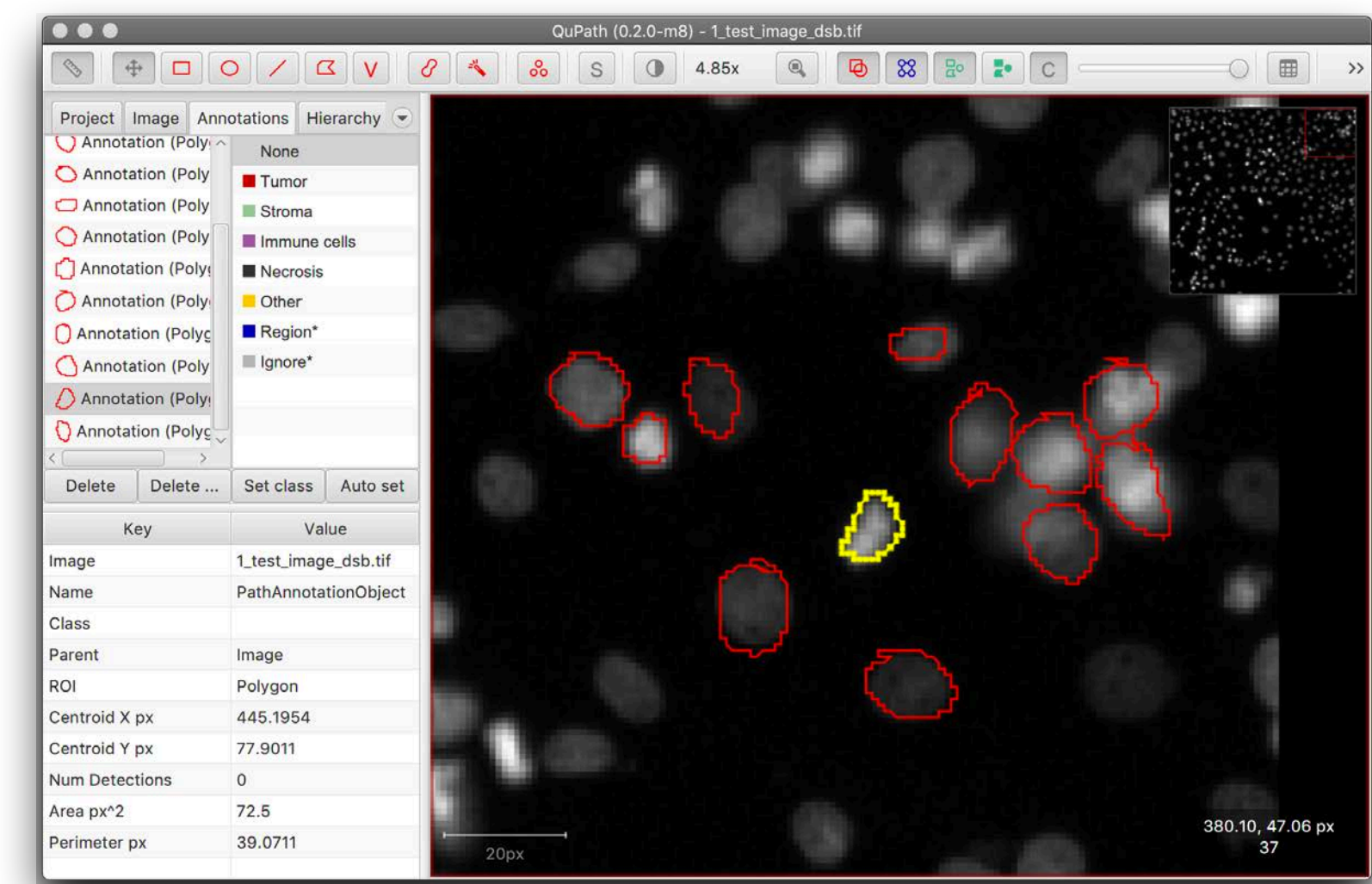
<https://imagej.net/Labkit>

M. Arzt, MPI-CBG



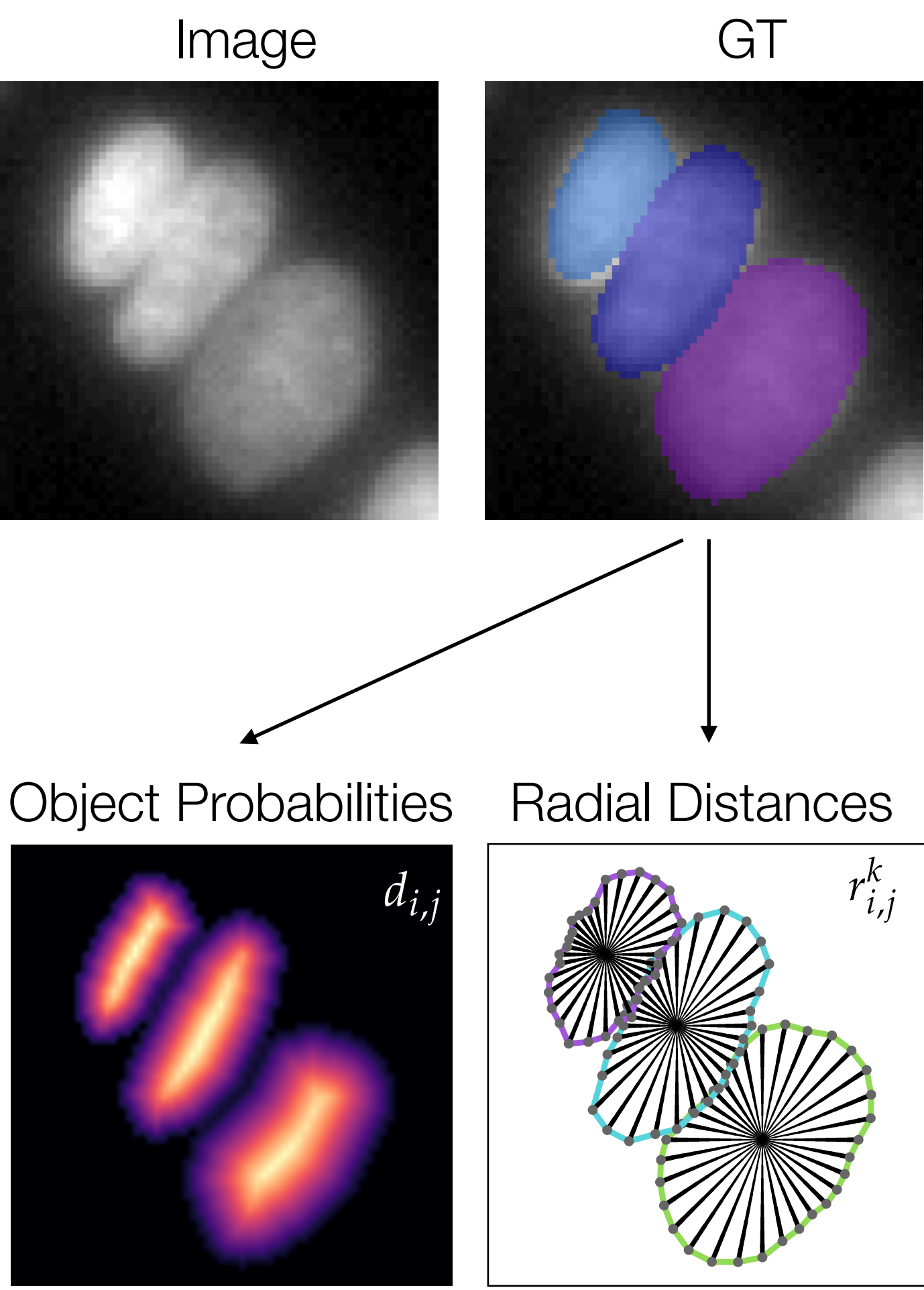
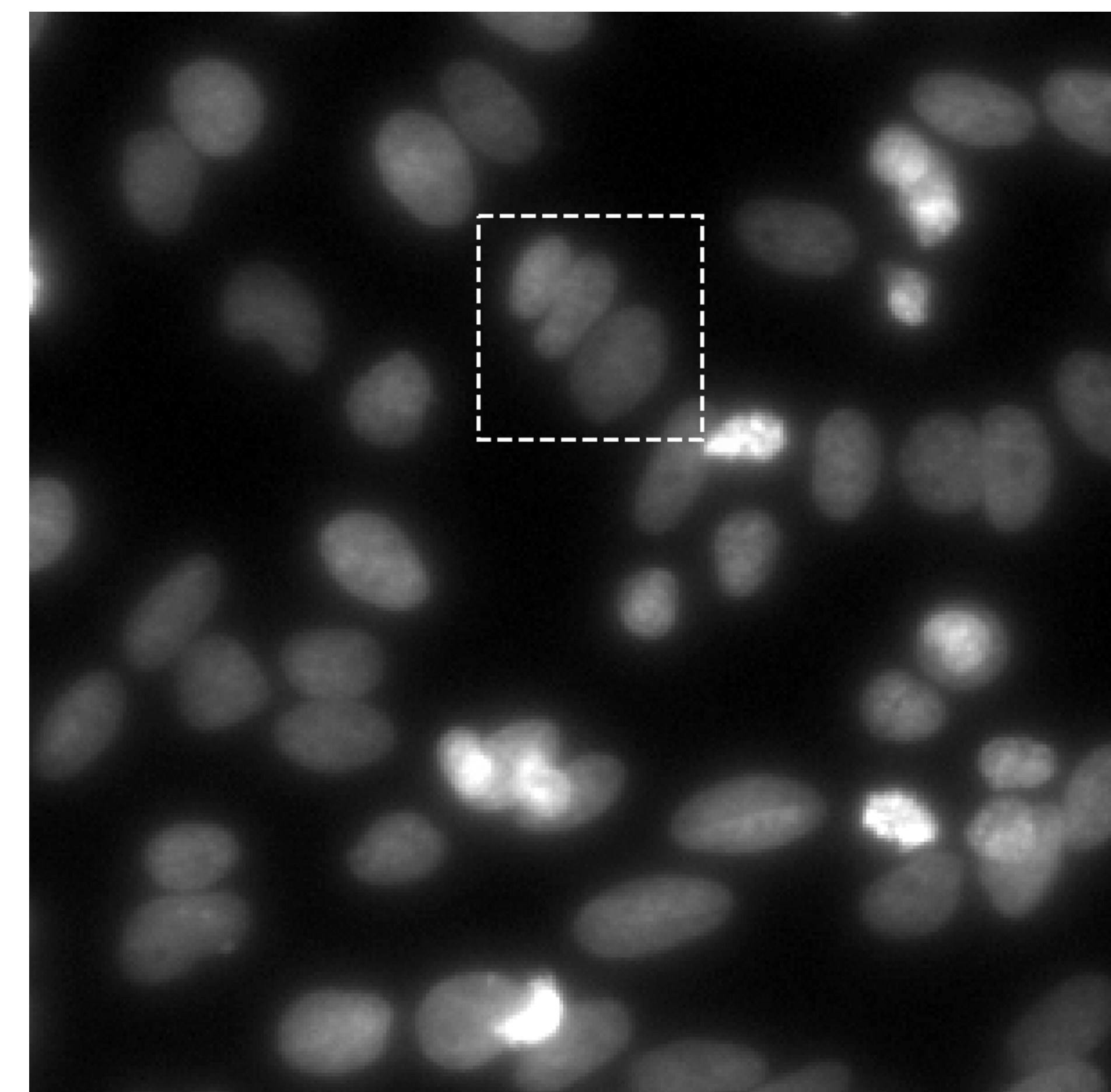
QuPath

- Draw ROI per object
- Convert to label mask

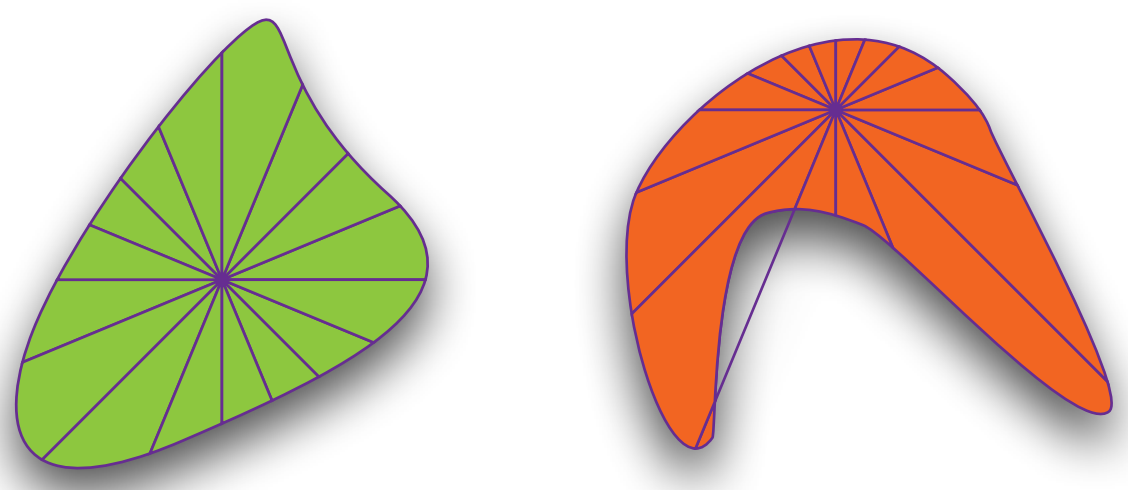


<https://qupath.github.io>

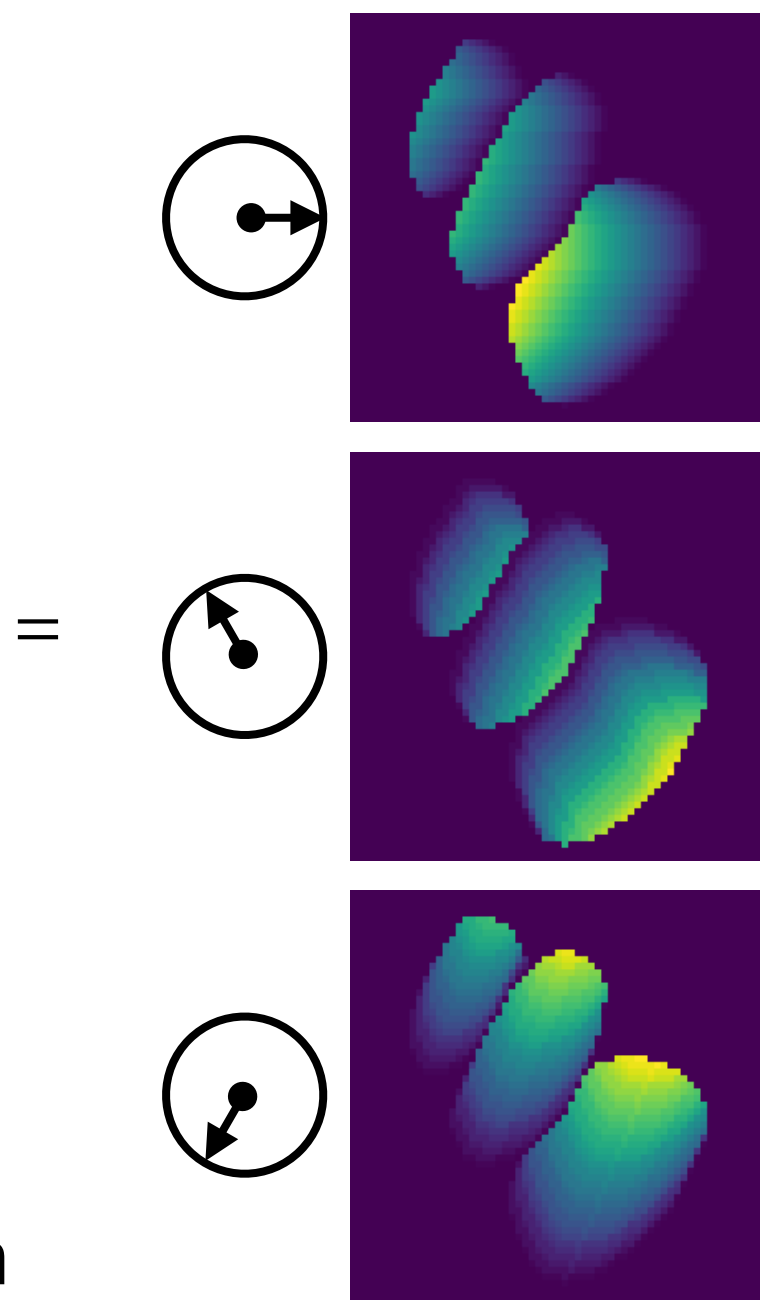
Bankhead et al. (2017)



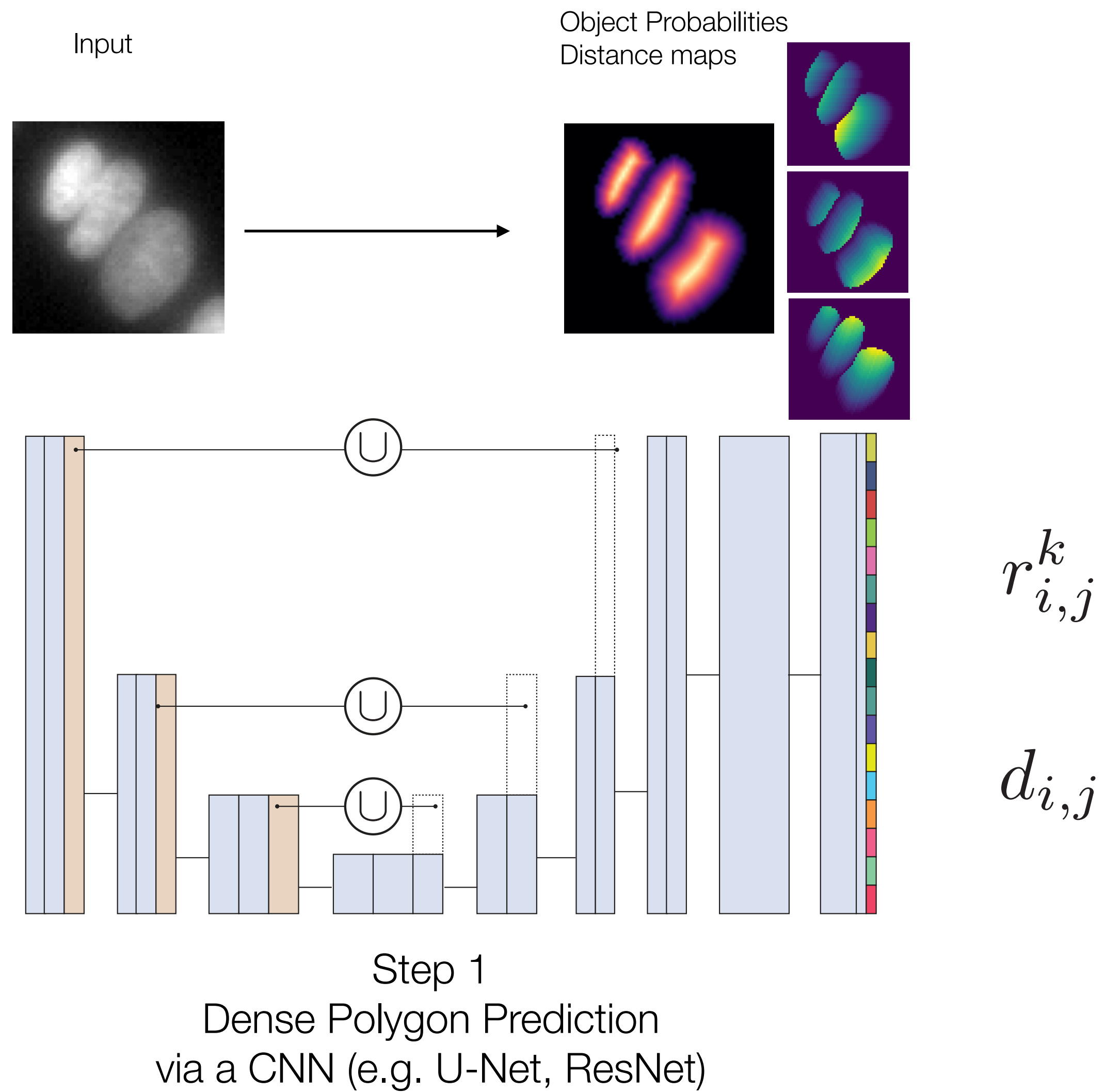
Star-Convex Not Star-Convex

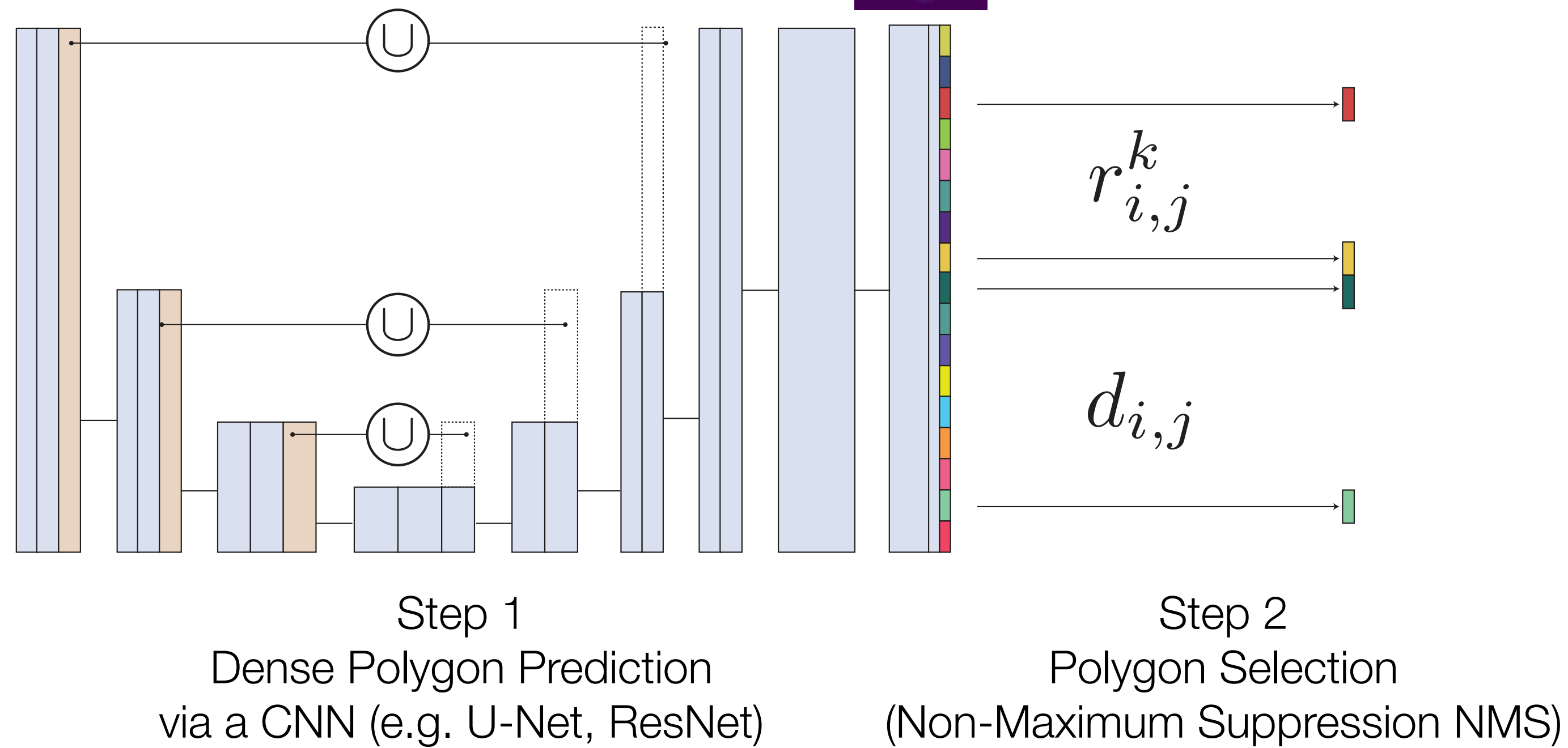
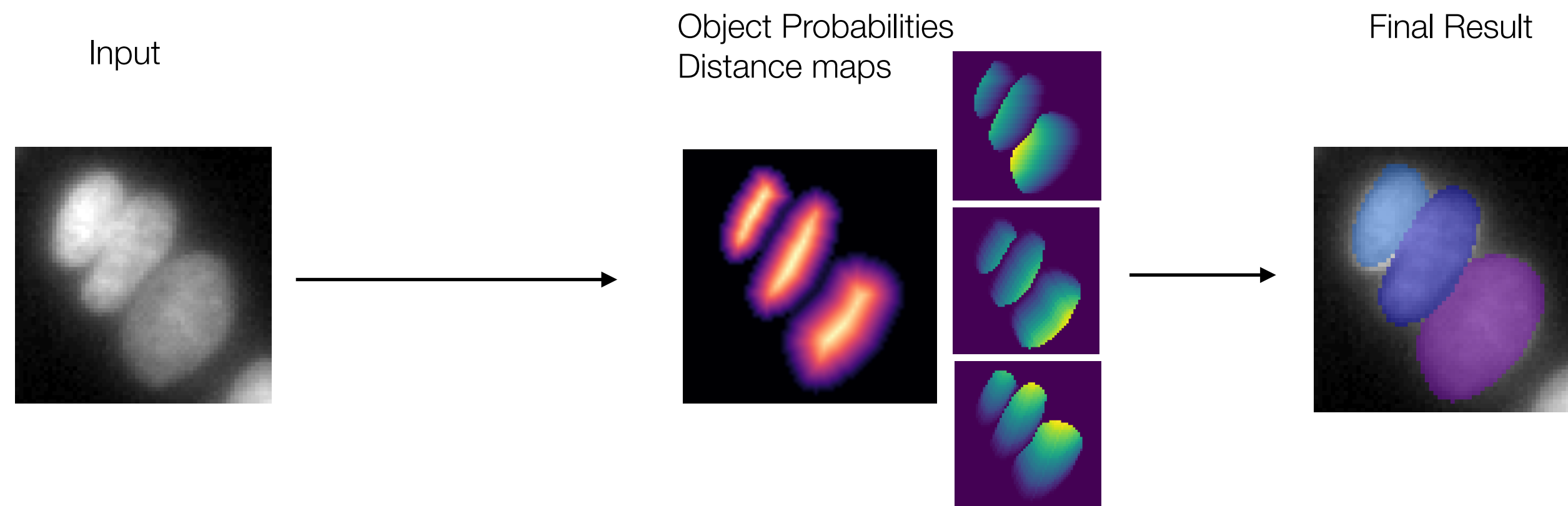


Distance maps (~32)

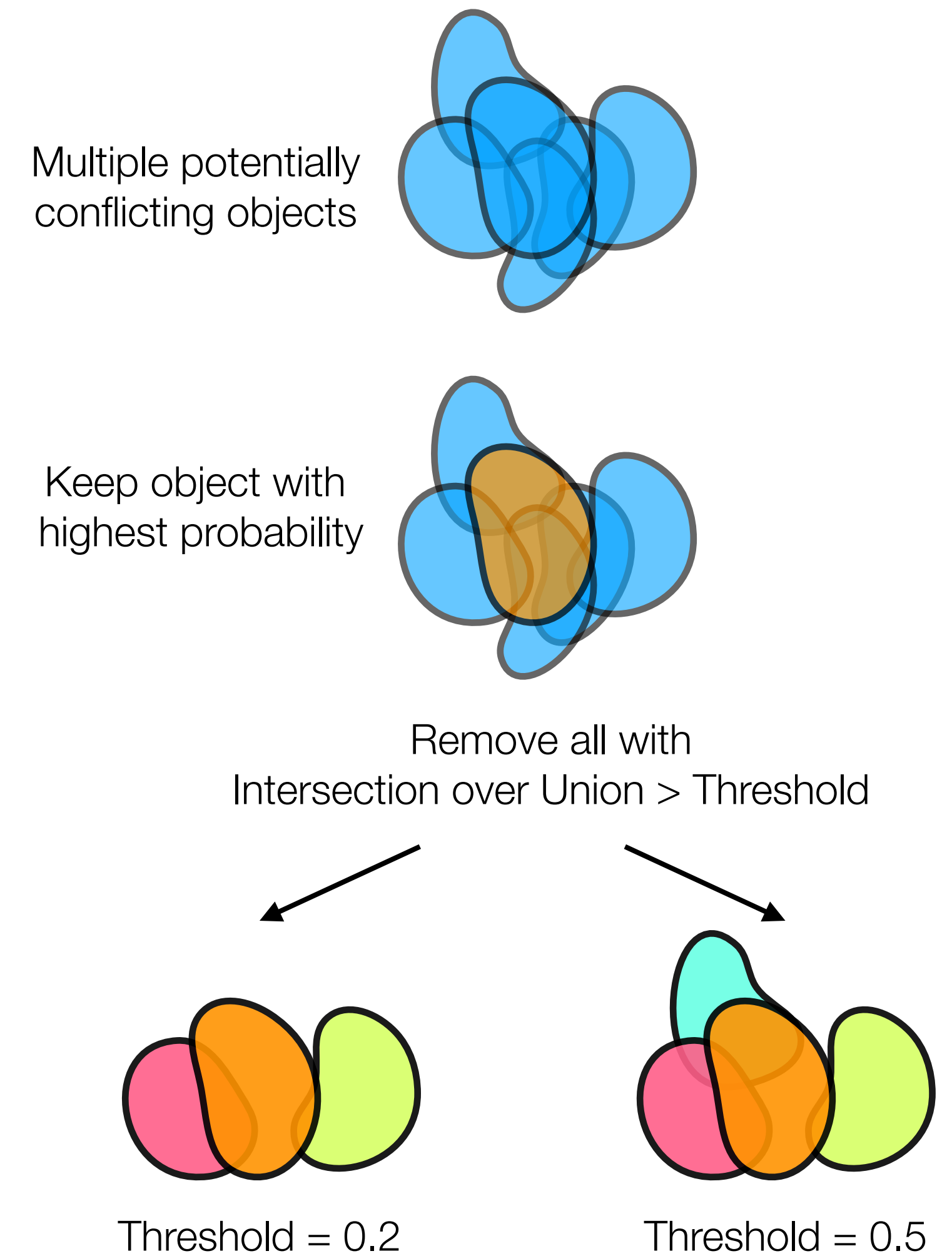


Approximate cells/nuclei with star-convex bounding polygon

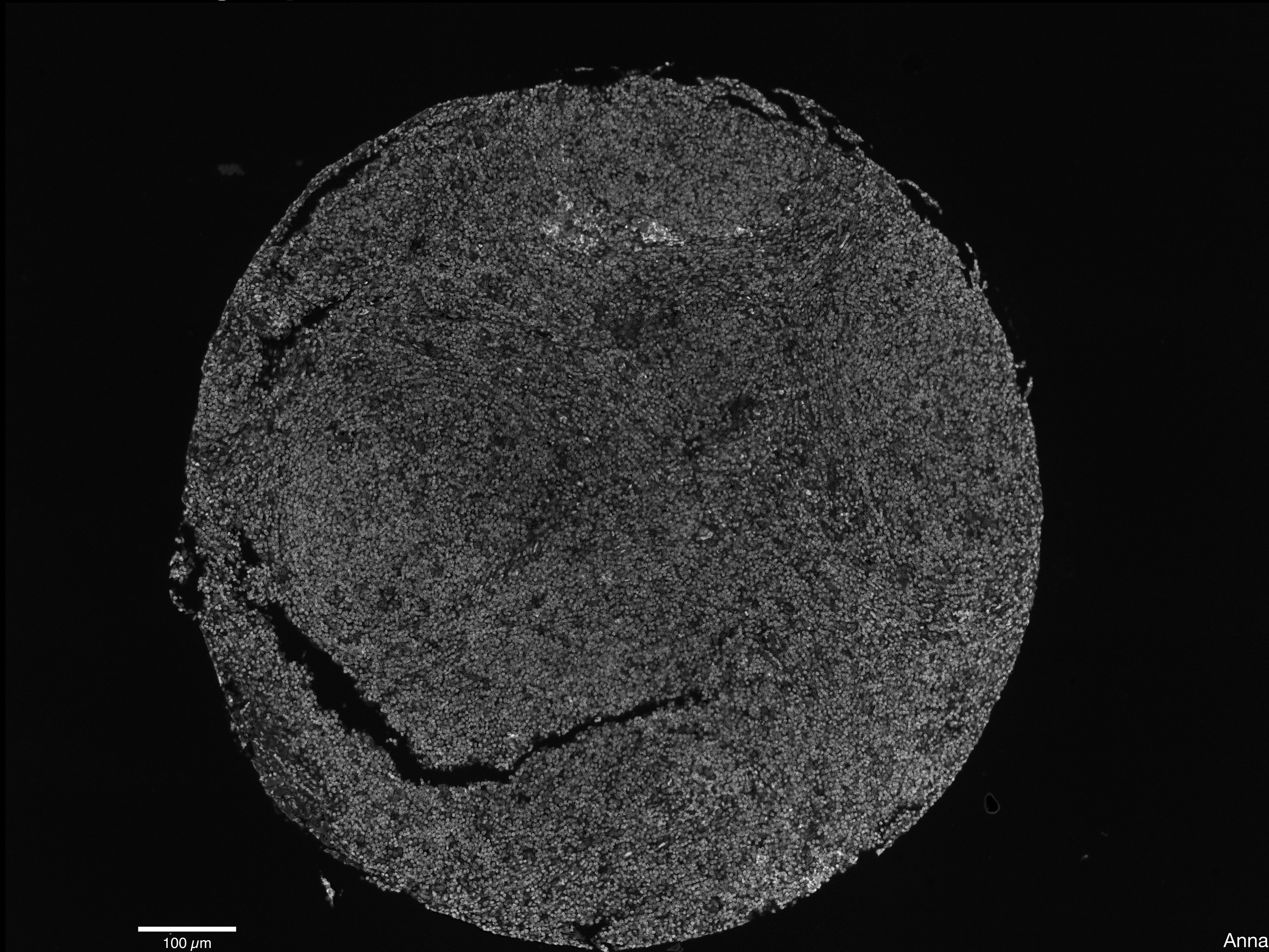


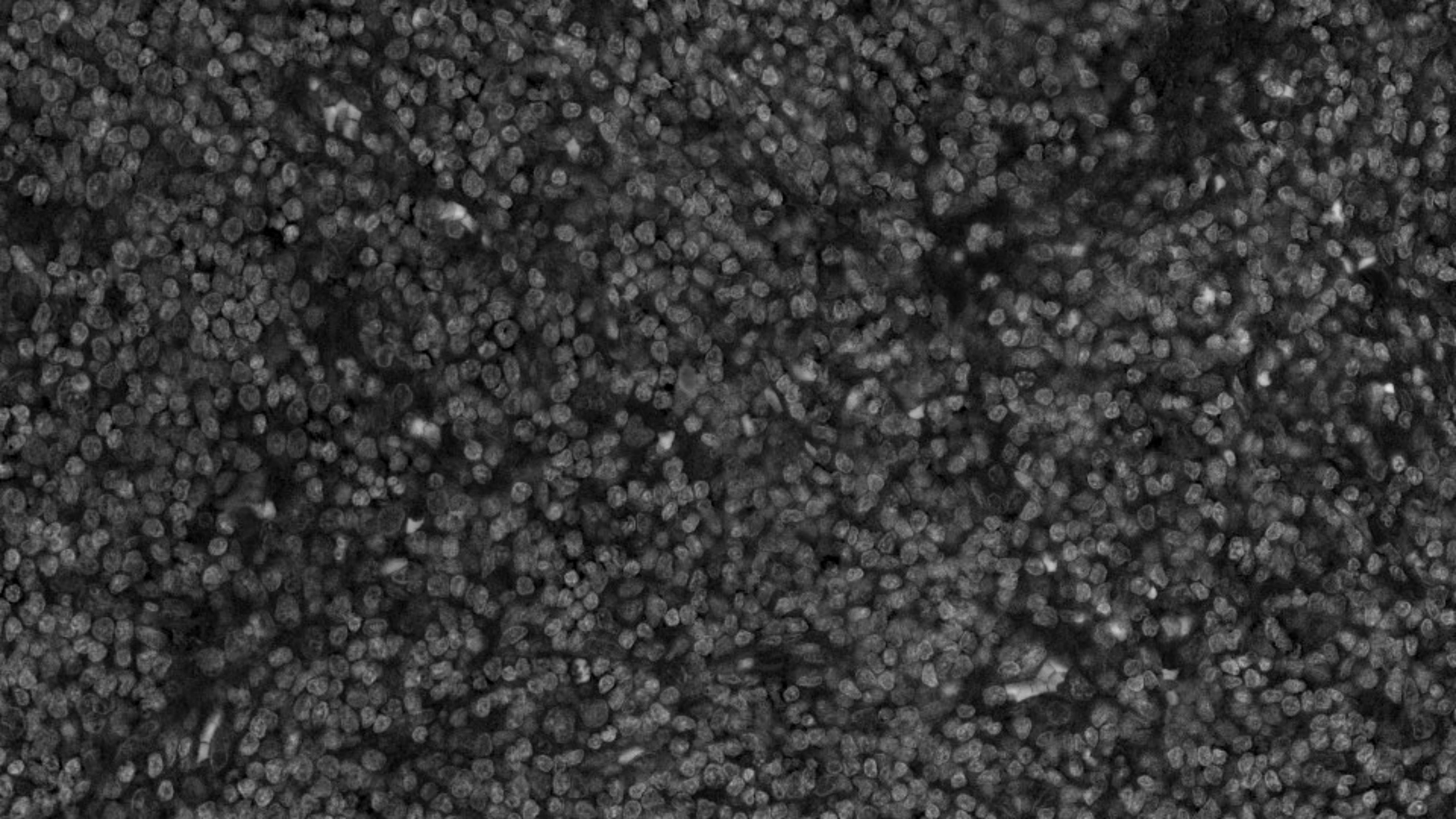


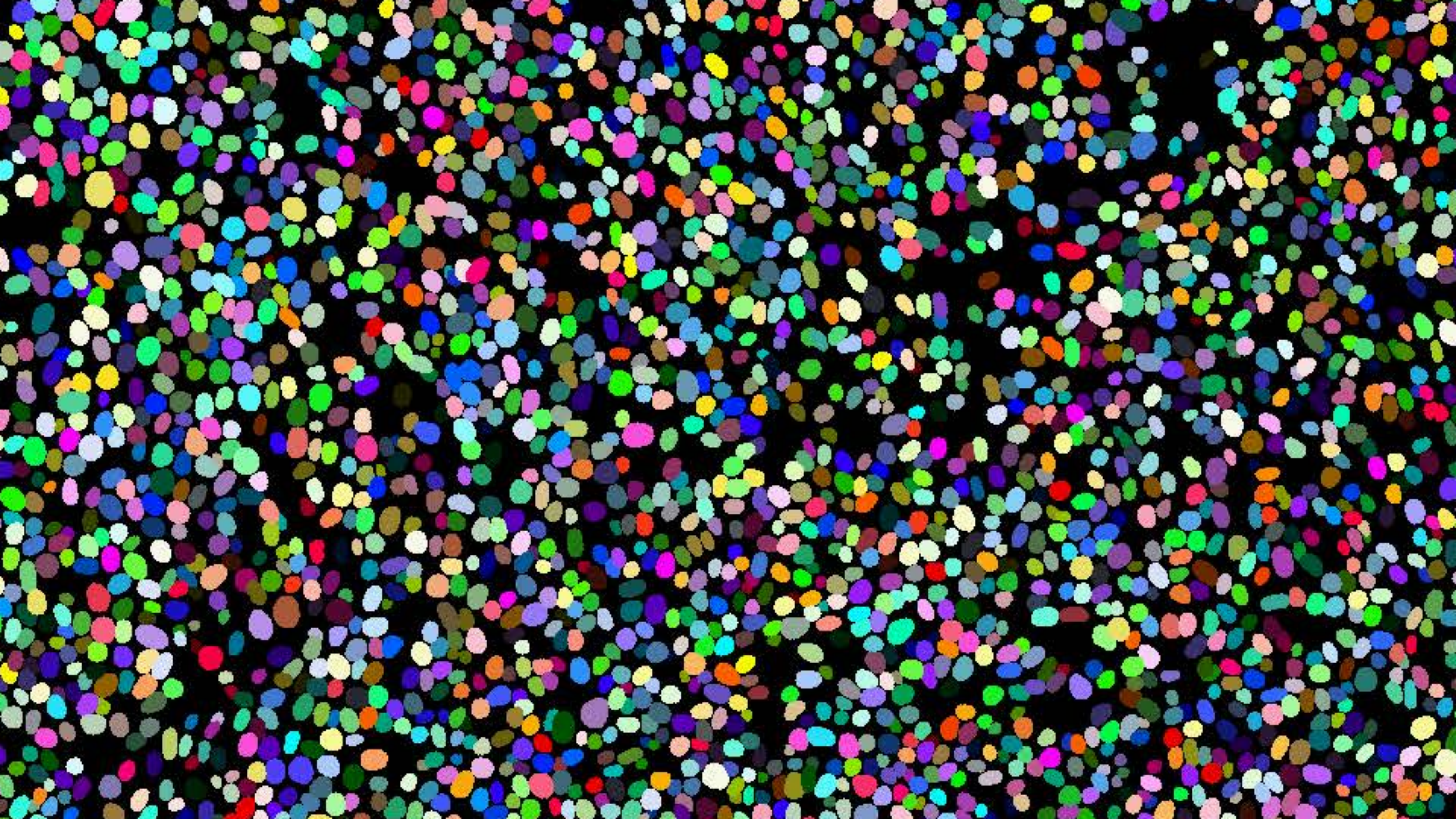
Non-Maximum-Suppression (NMS)



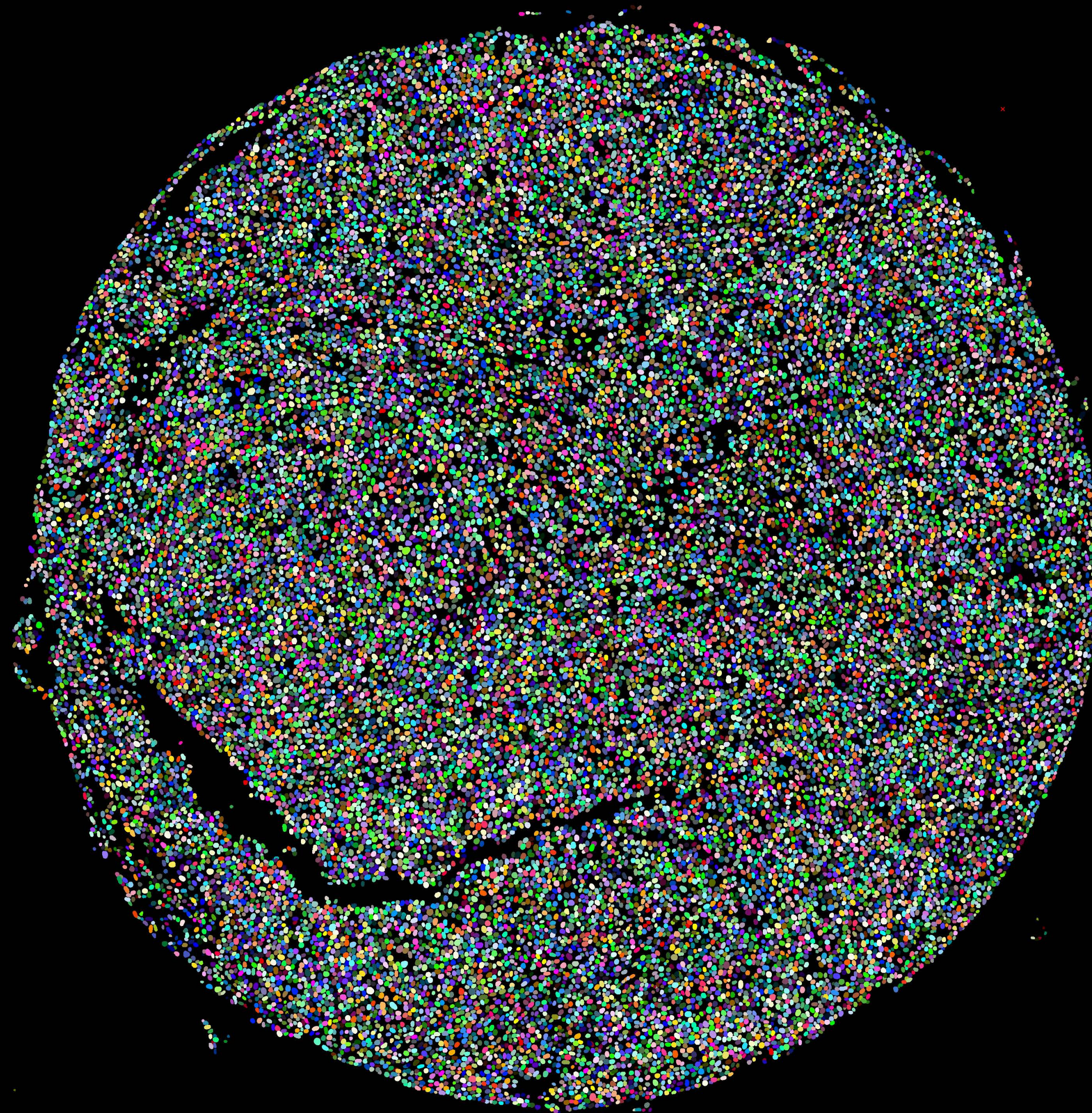
Example: crowded lymphoma cells

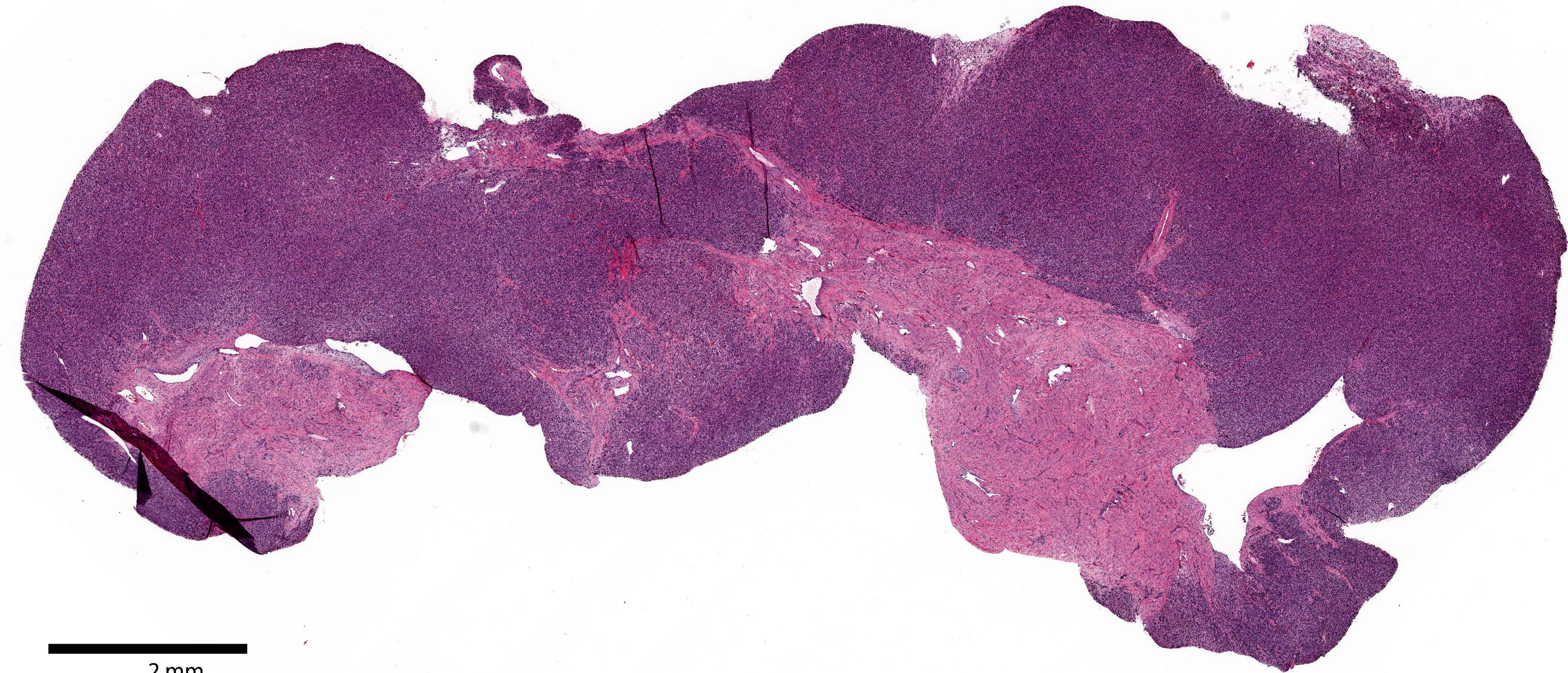




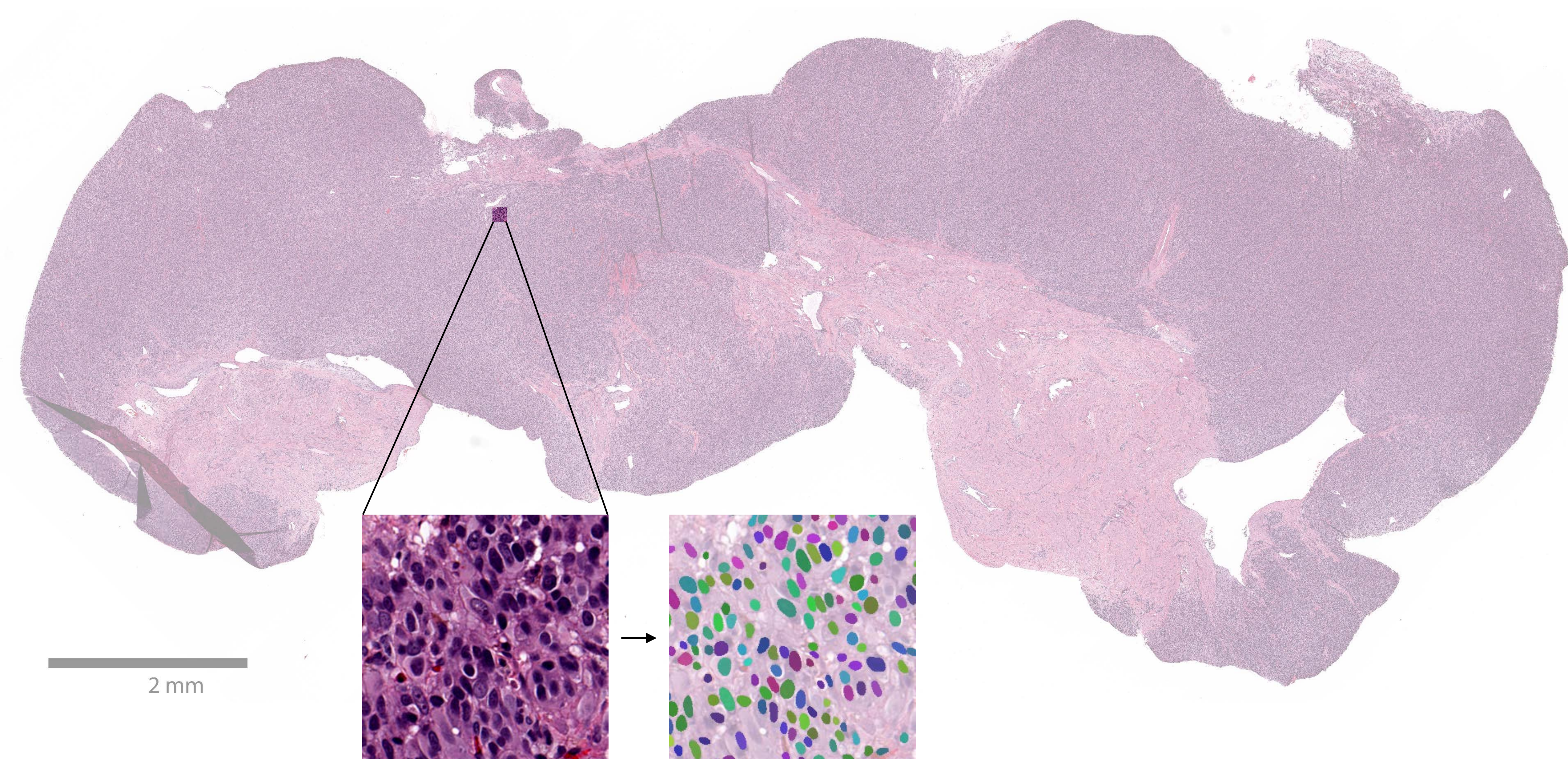


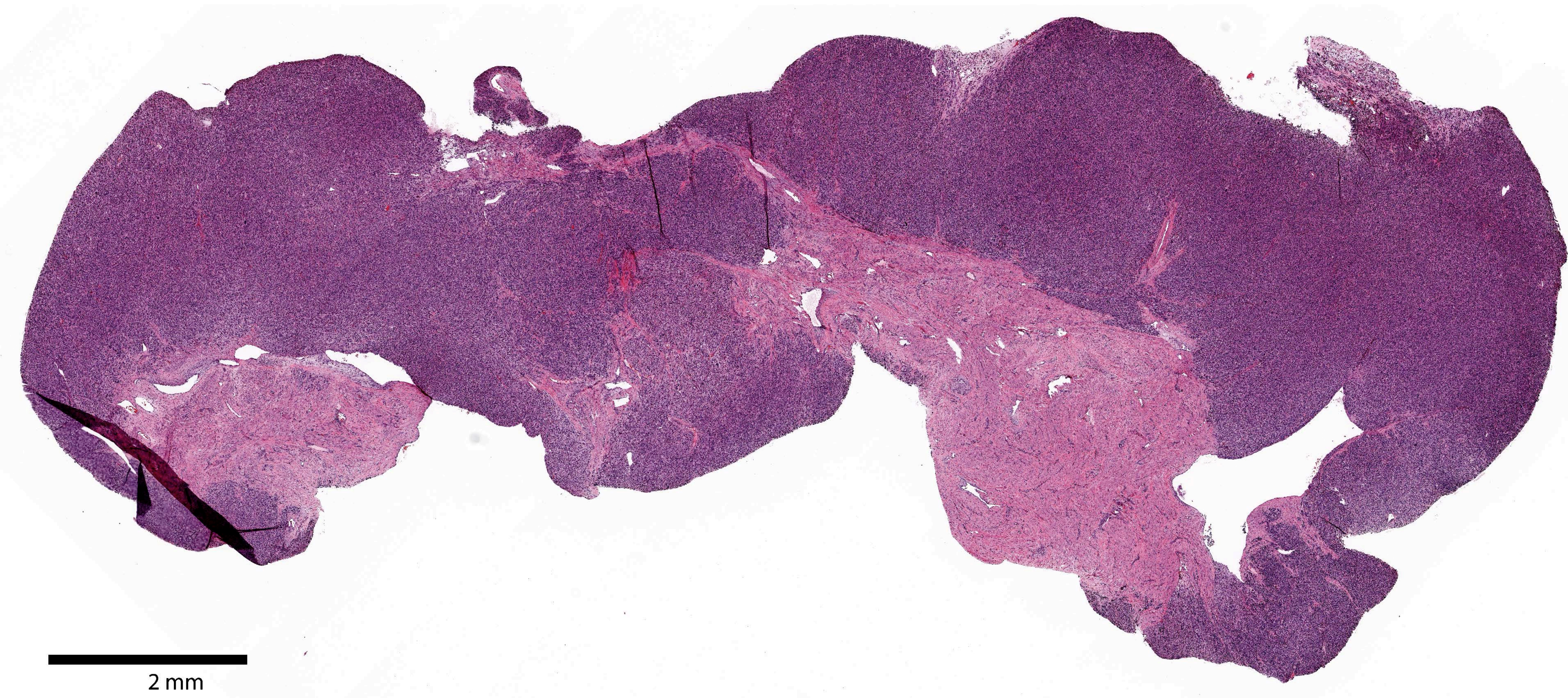
Example: crowded lymphoma cells



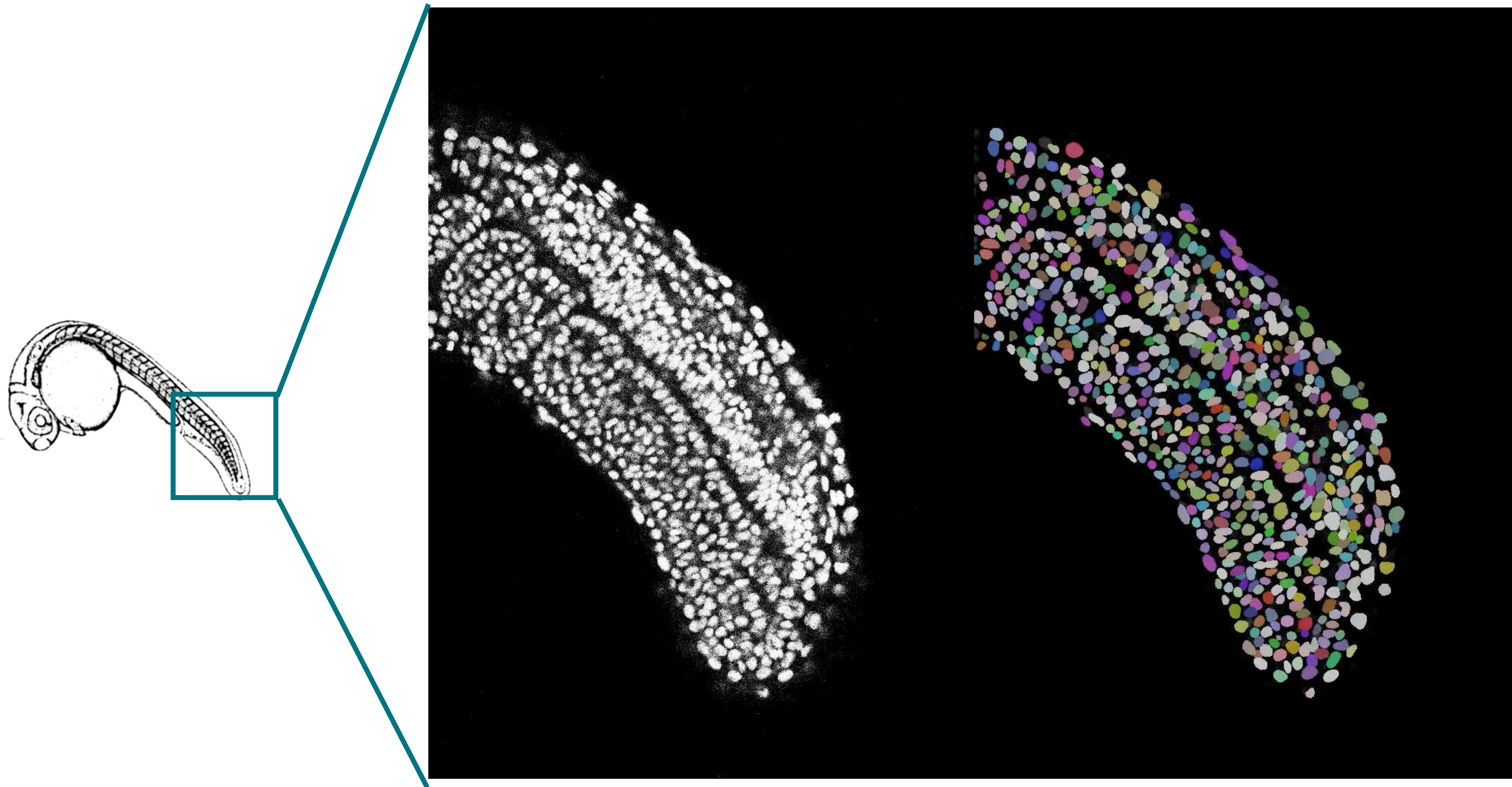


Whole Slide H&E (Sarcoma), (15mm x 6mm, 32k x 14k pixels)



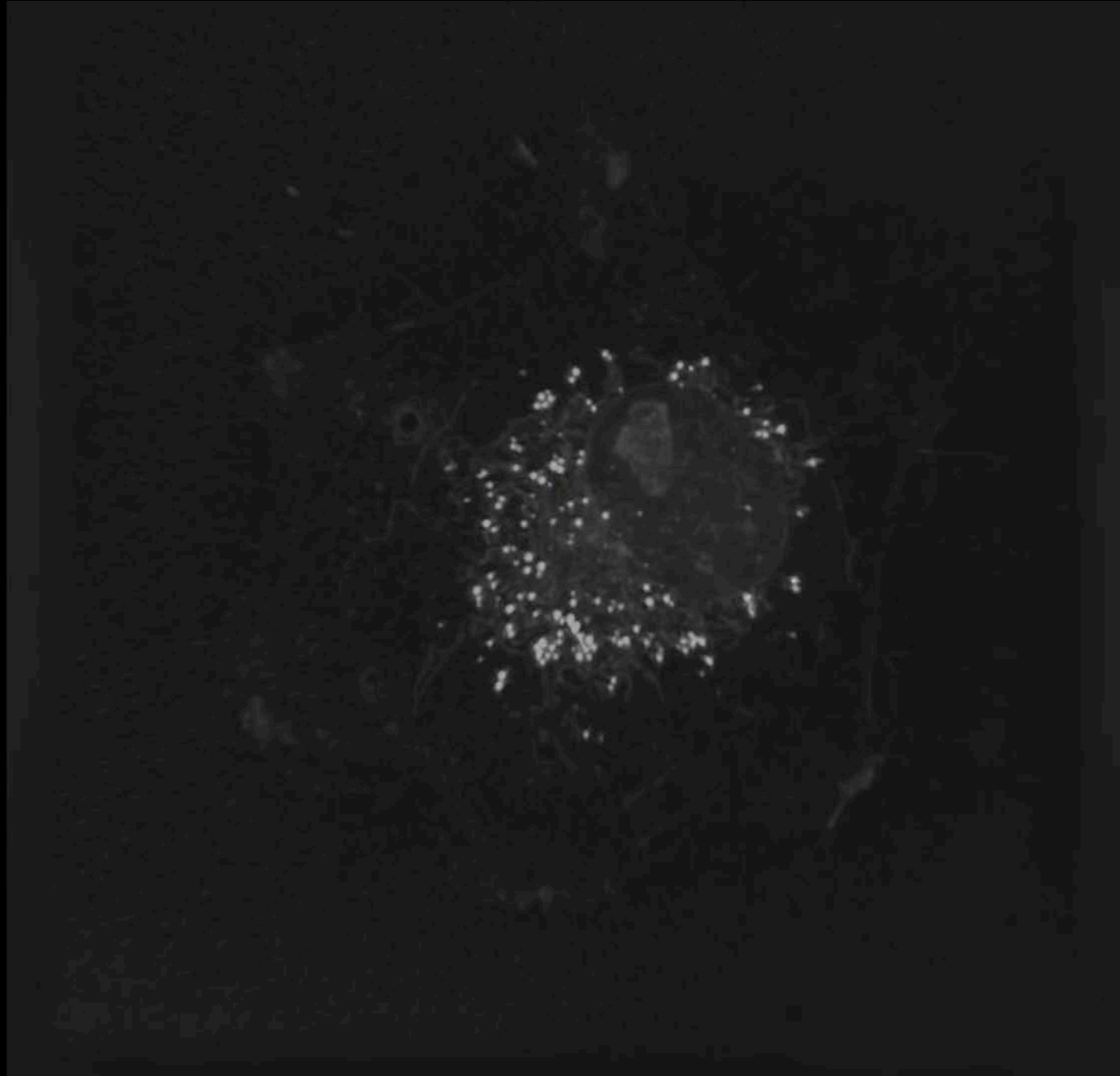


Whole Slide H&E (Sarcoma), (15mm x 6mm, 32k x 14k pixels)

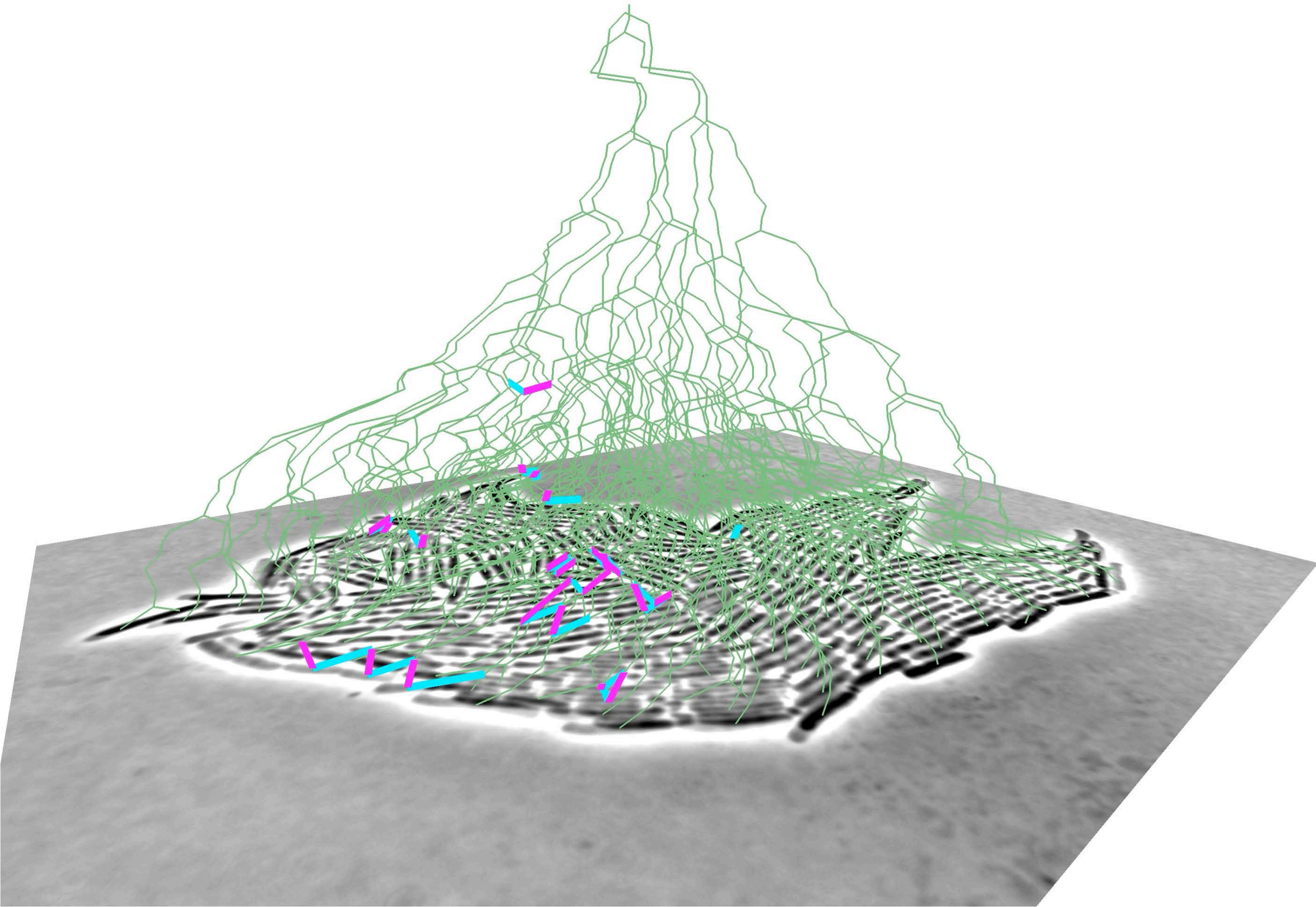


Zebrafish tail development, Data from Oates lab, EPFL

Other bioimaging tasks: detection and localization

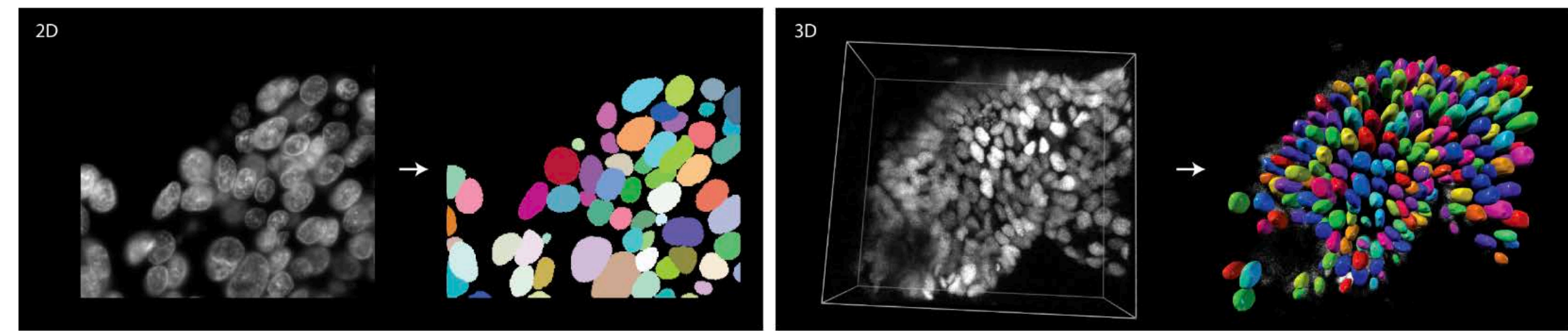


Method	Linking	TRA $\uparrow$	AOGM $\downarrow$
Distance	greedy	0.944	1511
Distance	ILP	0.962	1065
TrackMate (overlap)	LAP	0.957	872
DELTA 2.0	greedy	0.996	118
TRACKASTRA (points only)	ILP	0.995	136
TRACKASTRA	greedy	0.999	36
TRACKASTRA	ILP	0.999	23



pypi package 0.9.1
 Anaconda.org 0.9.1
 Test passing
 Test (PyPI) failing
 forum 459 topics
 downloads 22k/month

StarDist - Object Detection with Star-convex Shapes



Spotiflow

Spotiflow 0.5.0 documentation

🔍

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Spotiflow

0.5.0

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Spotiflow 0.5.0 documentation

Predicting spots using the napari plugin

Inference via CLI

Available pre-trained models

Training a Spotiflow model on a custom dataset

Fine-tuning a Spotiflow model on a custom dataset

API Reference

Spotiflow

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Command Line Interface (CLI)

Napari plugin

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Spotiflow

Spotiflow is a learning-based subpixel-accurate spot detection method for 2D and 3D fluorescence microscopy. It is primarily developed for spatial transcriptomics workflows that require transcript detection in large, multiplexed FISH-images, although it can also be used to detect spot-like structures in general fluorescence microscopy images and volumes. For more information, please refer to our [paper](#).

The screenshot shows the Napari application window. The main view displays a grayscale microscopy image with a few small, bright spots. One spot is highlighted with a blue and red star-shaped mask. The left sidebar contains a 'layer controls' panel with settings for the selected layer (label, opacity, brush size, blending, color mode, contour, n edit dim, contiguous, preserve labels, show selected) and a 'layer list' panel showing the 'mask' and 'img' layers. The bottom status bar indicates the current sample is 'trackastra' with coordinates '[0 864 196]'.

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