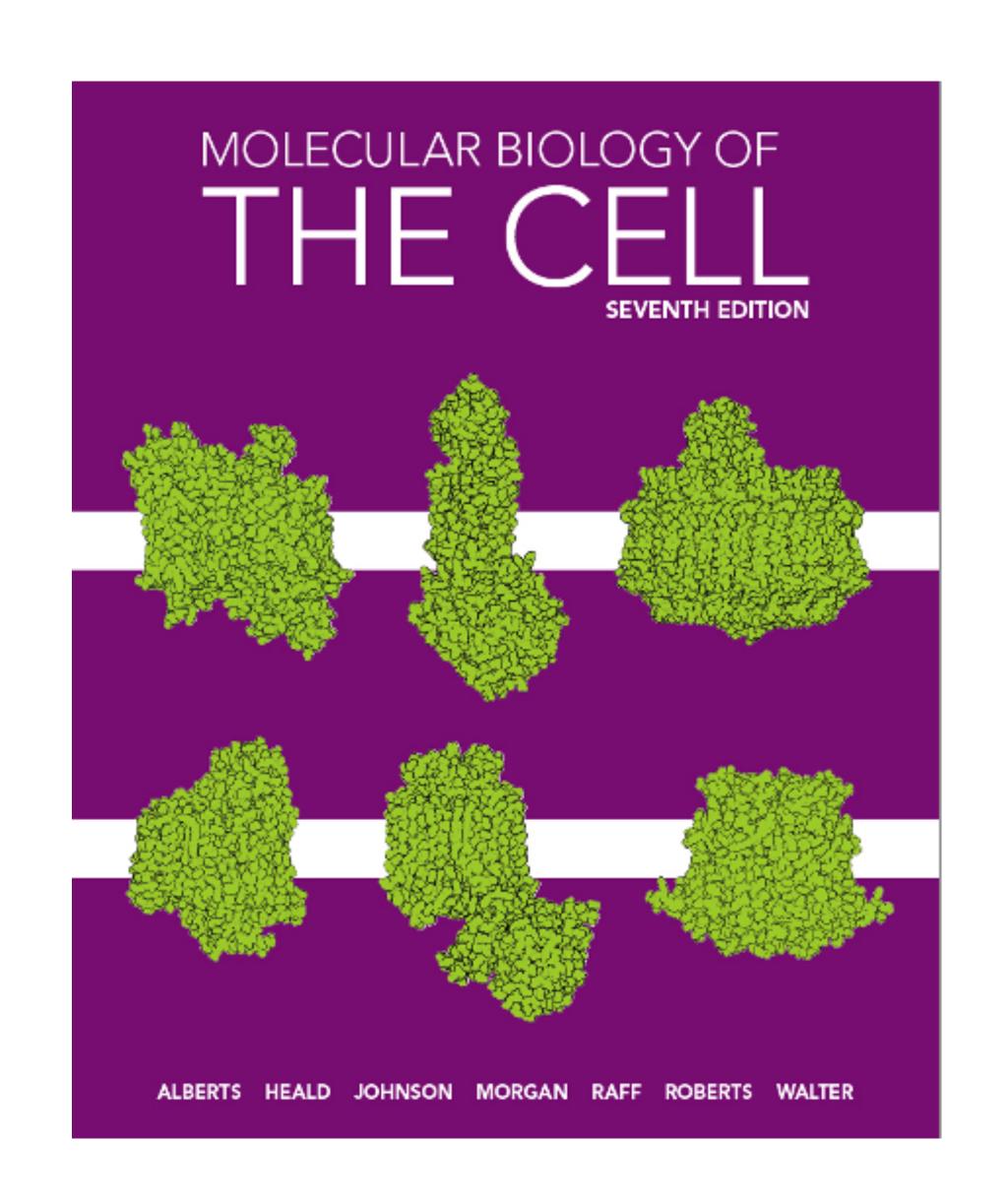
# Cellular and Molecular Biology I

**BIO-205-2** 

**Camille Goemans - 2024** 

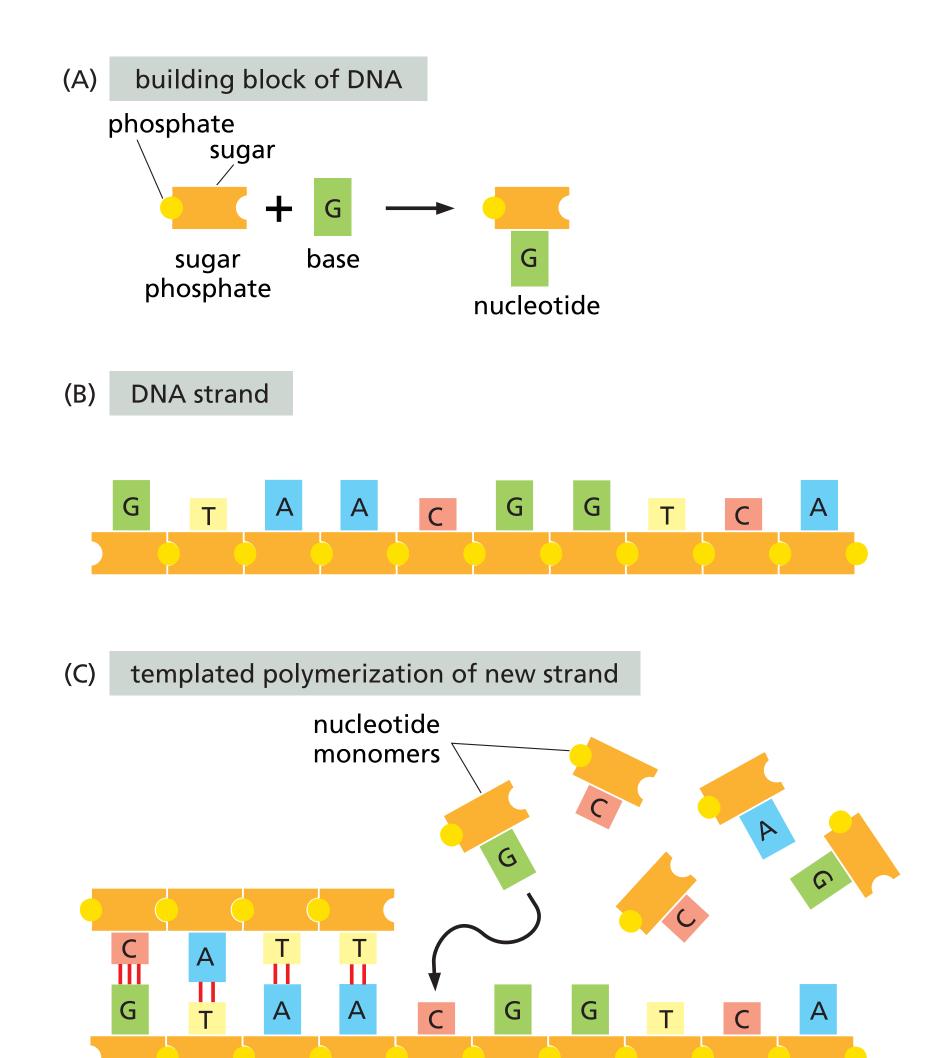
### Important information

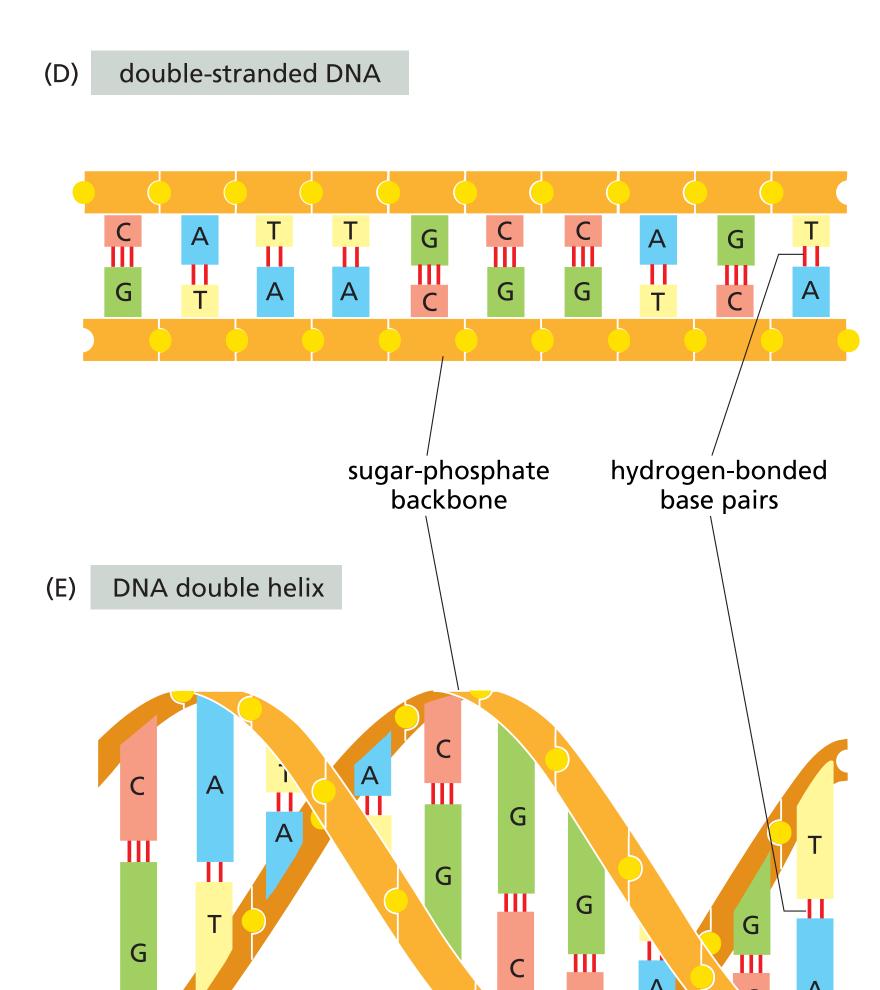
- How to study for this class? What can you bring to the exam?
- Nice video about epigenetics (~ 8min): <a href="https://www.youtube.com/watch?">https://www.youtube.com/watch?</a>
   v=MD3Fc0X0jWk
- Recording?

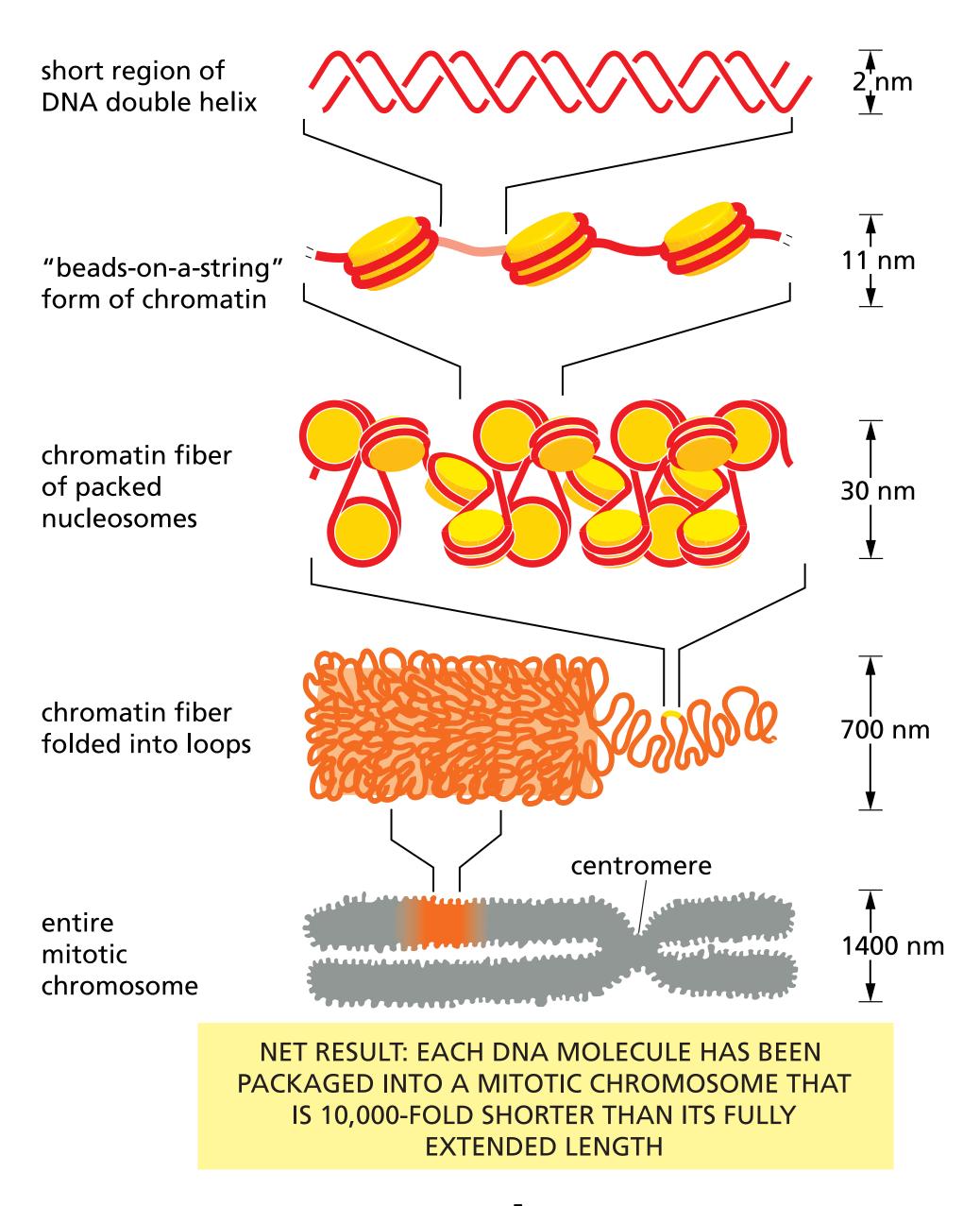


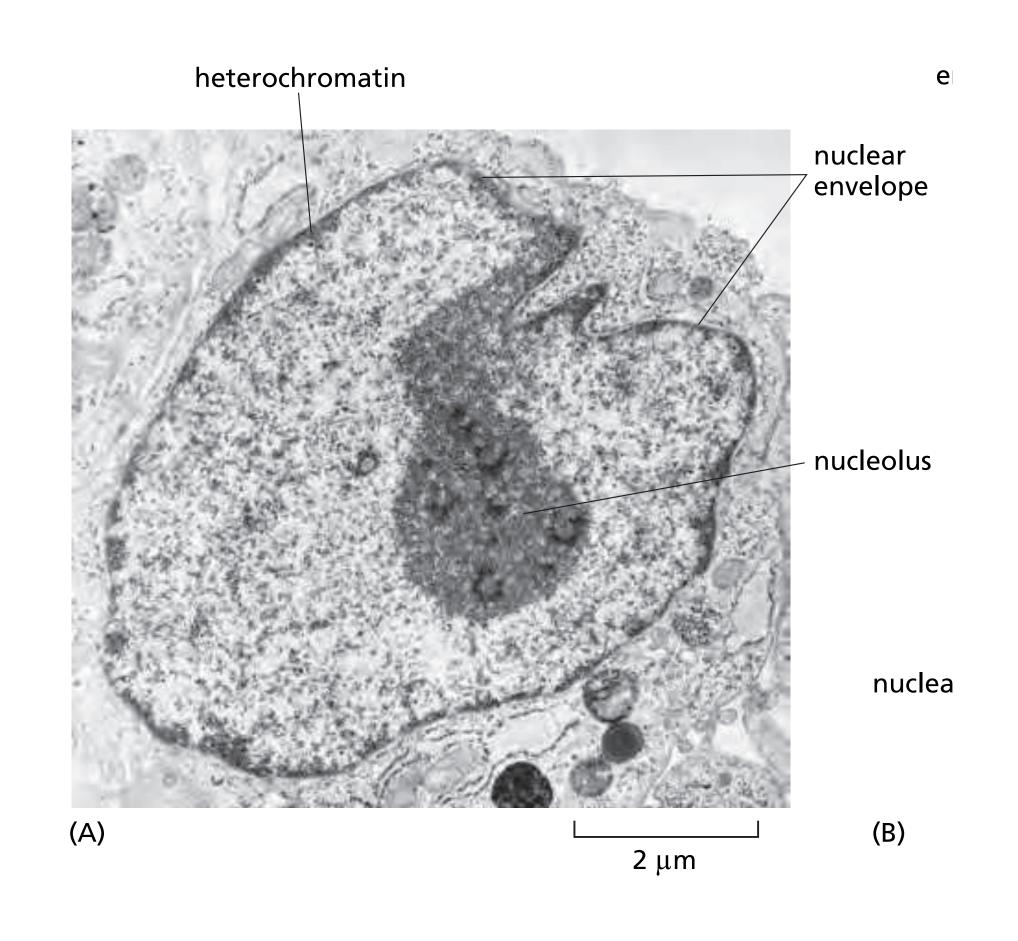
#### **Chapter 4**

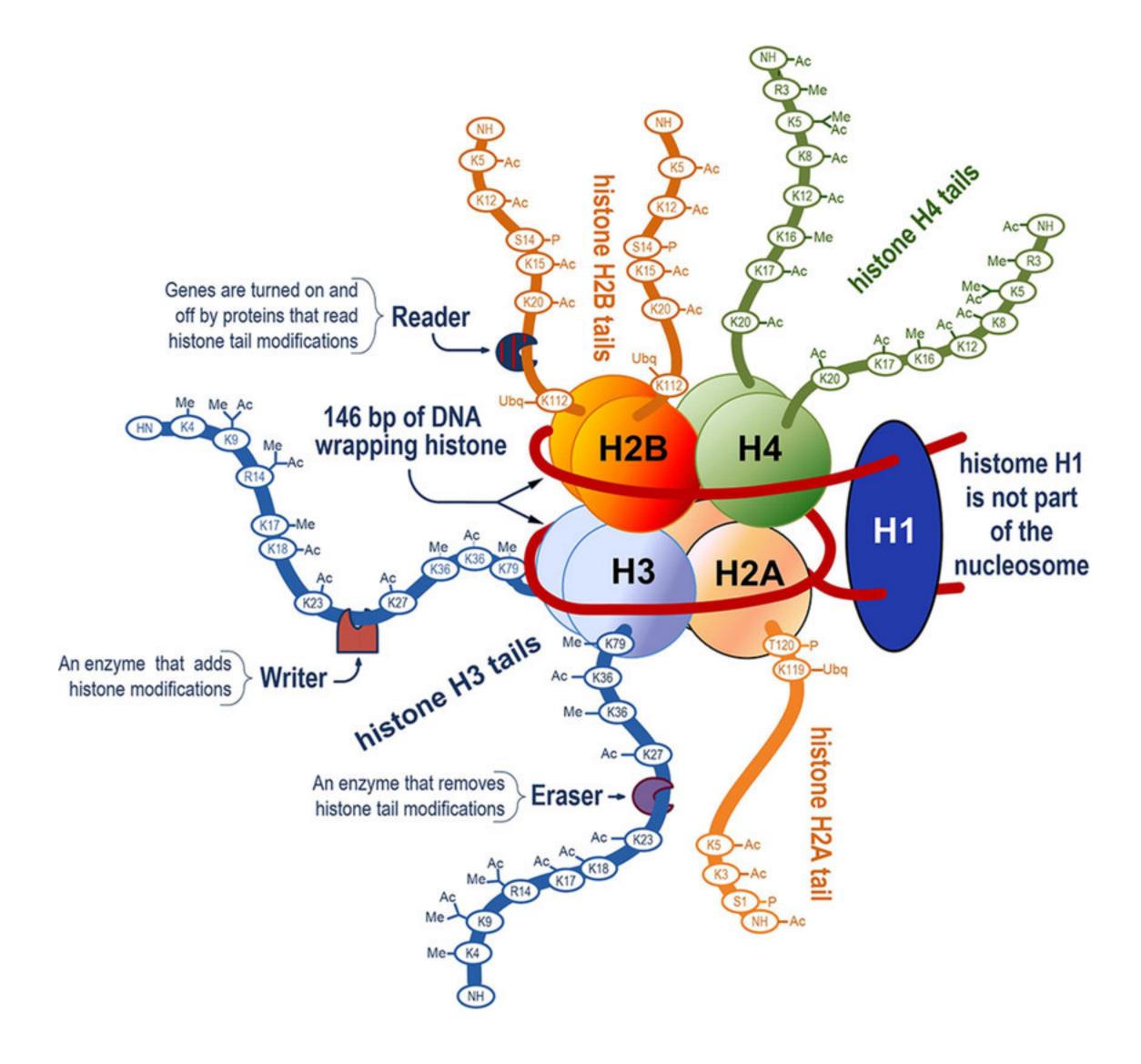
DNA, Chromosomes, and Genomes

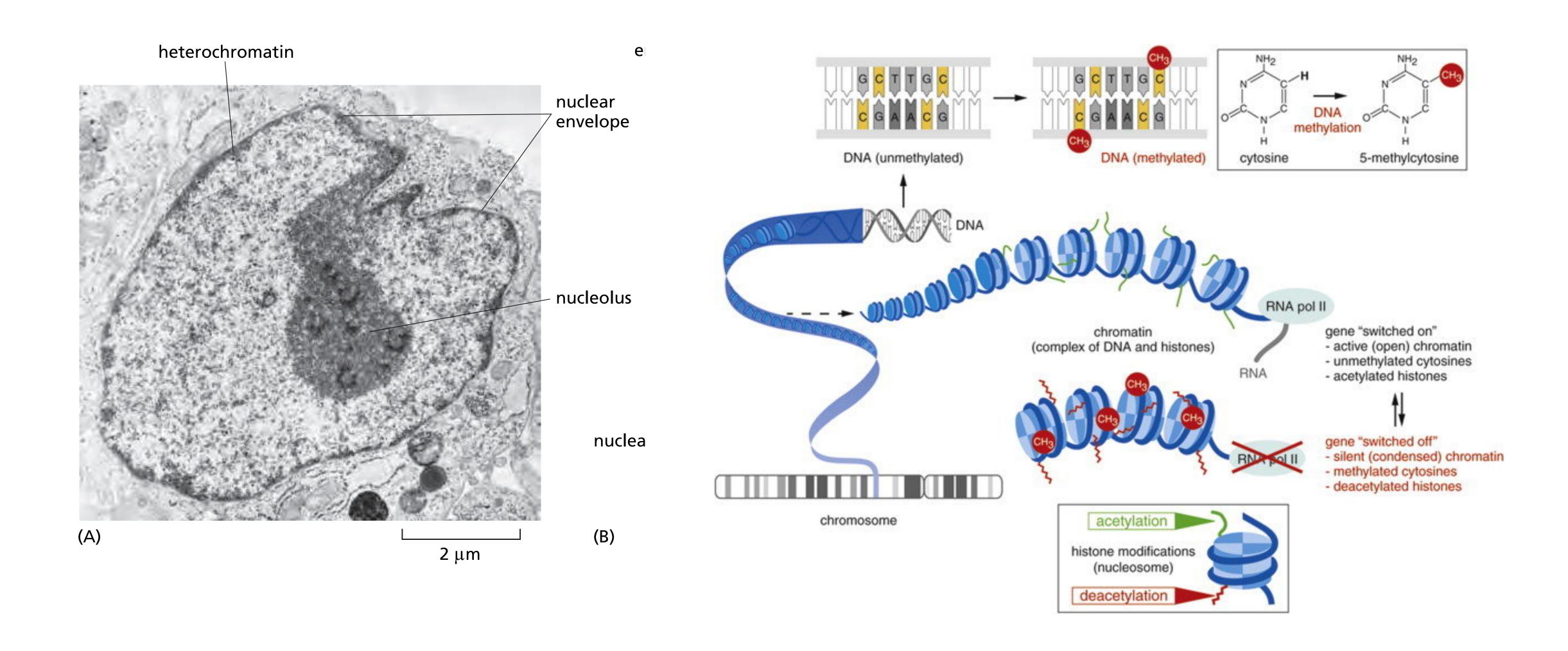










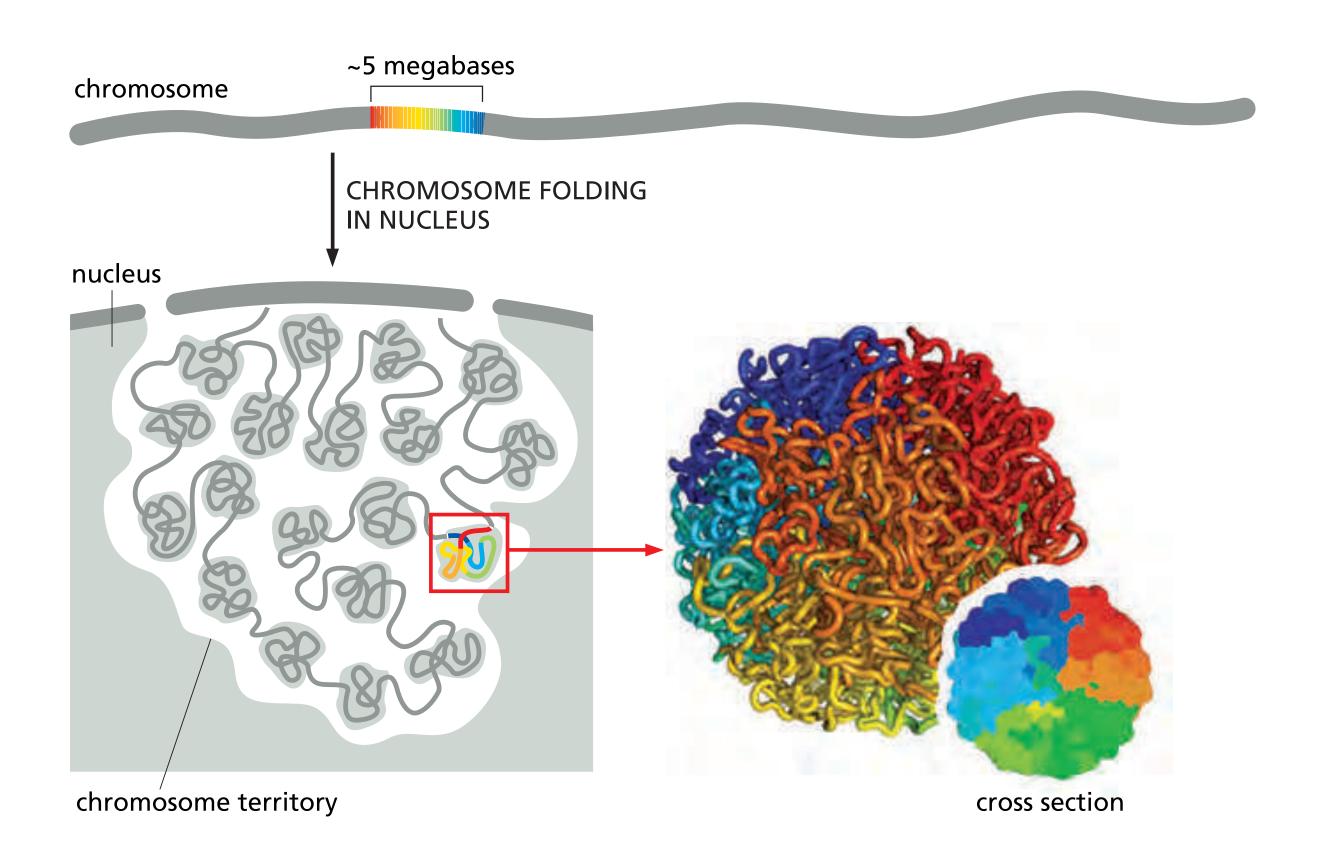


#### Plan

- Quick recap
- The global structure of chromosomes
- Analysis of chromatin organization
- How do genomes evolve

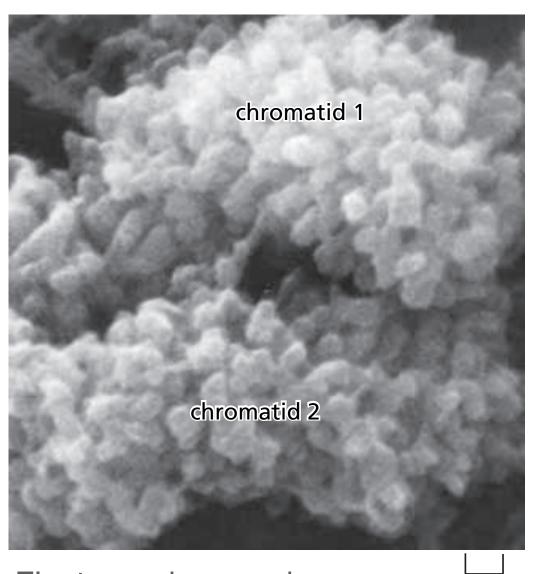
## The global structure of chromosomes

 Most regions of our chromosomes are arranged into a conformation = fractal globule - dense packing that maintains the ability to fold and unfold



### The global structure of chromosomes

- Mitotic chromosomes are especially highly condensed (visible by light microscopy) gene expression is shut down
- Two DNA molecules produced during interphase = 2 sister chromatids held together at the centromere
- Final level of chromosome packaging



Electron micrograph 0.1 μm

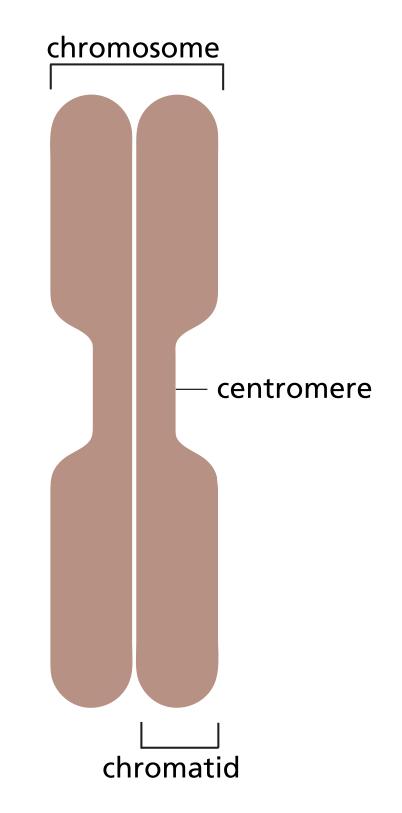
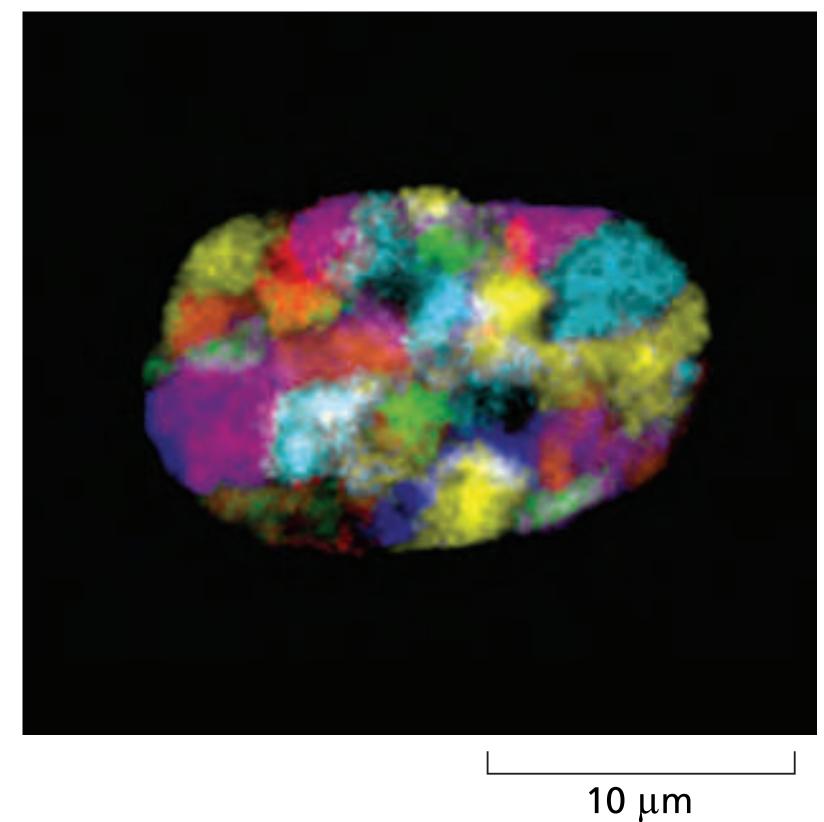
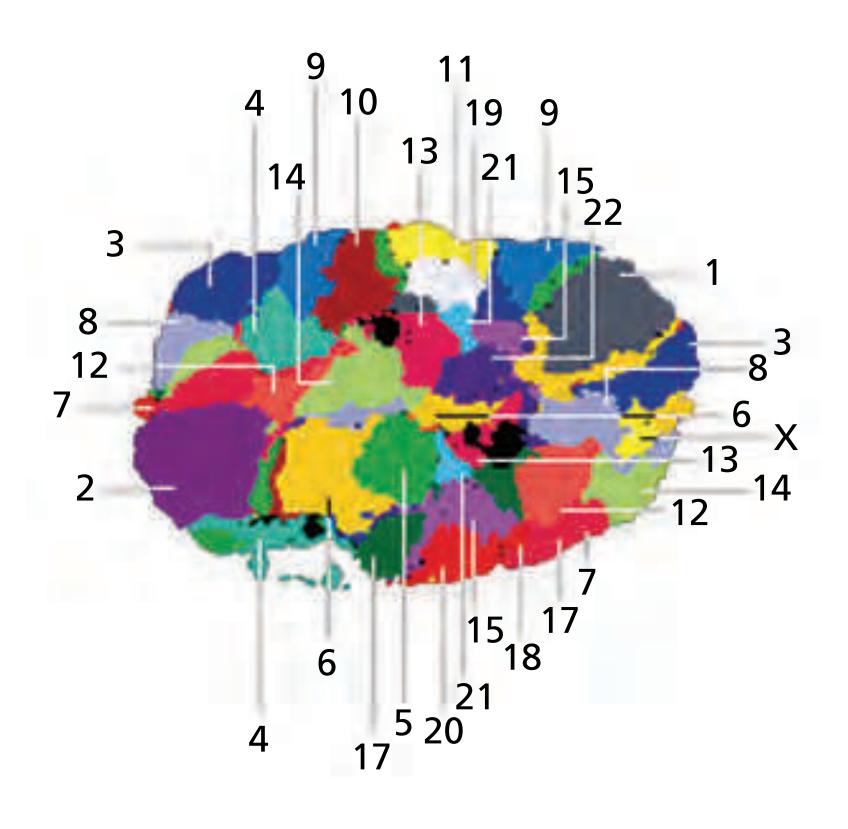


Figure 4–59 A typical mitotic chromosome at metaphase. Each sister chromatid contains one of two identical sister DNA molecules generated earlier in the cell cycle by DNA replication (see also Figure 17–21).

# Chromosomes occupy specific territories

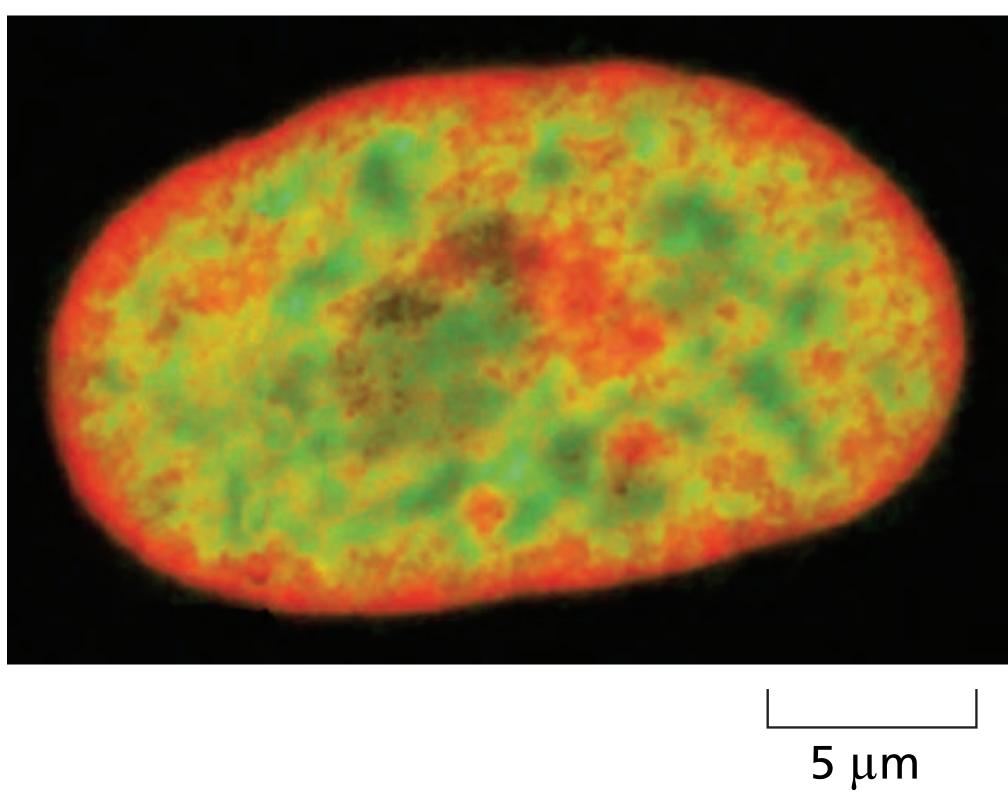
Each of our 46 interphase chromosomes tend to occupy its own discrete territory



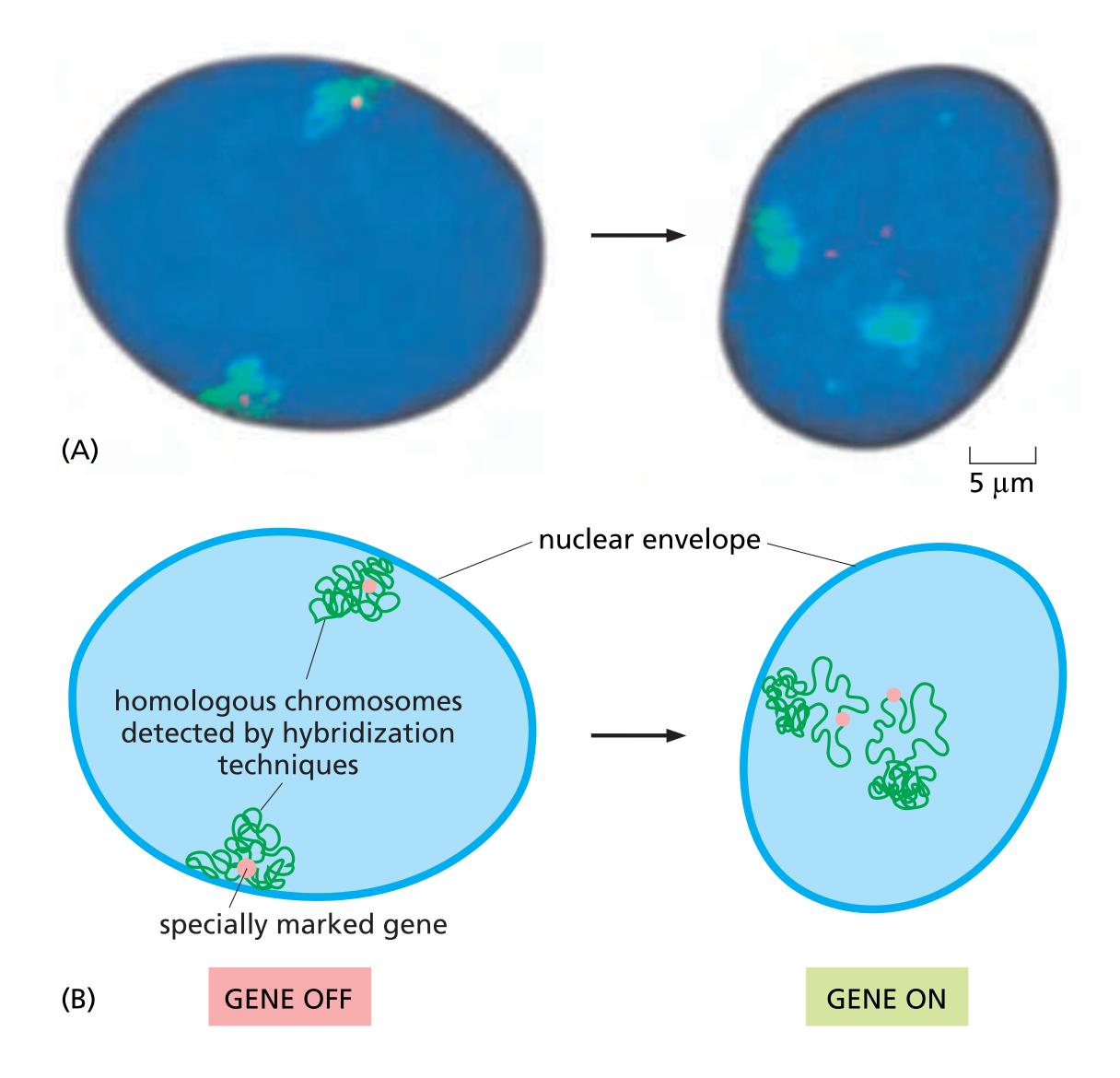


## Chromatin is organised within the nucleus

• Heterochromatic regions are associated with nuclear lamina (red), whereas gene-rich regions are more central

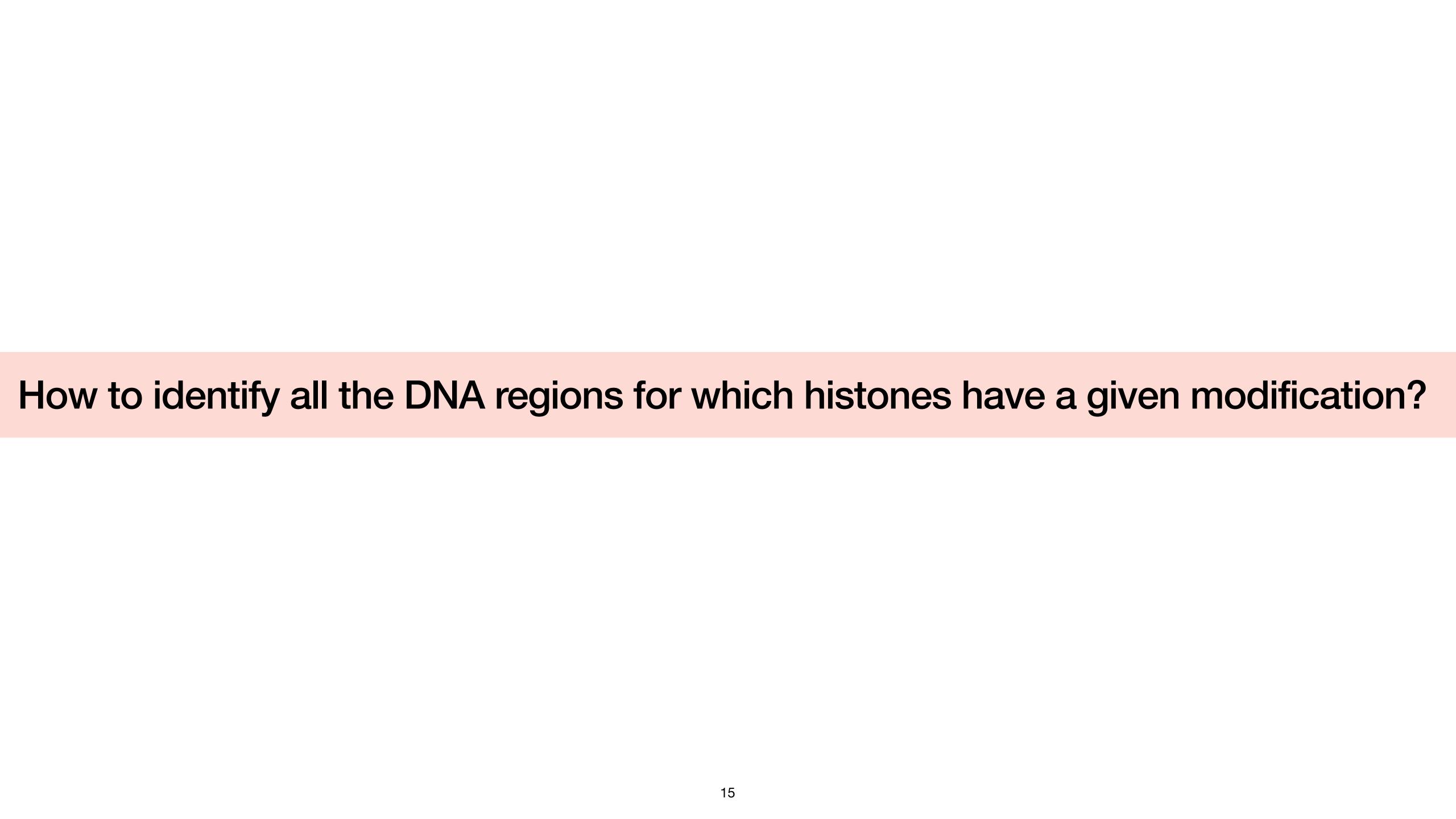


Chromatin position can change with gene expression



#### Plan

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# Chromatin Immune-precipitation (ChiP)

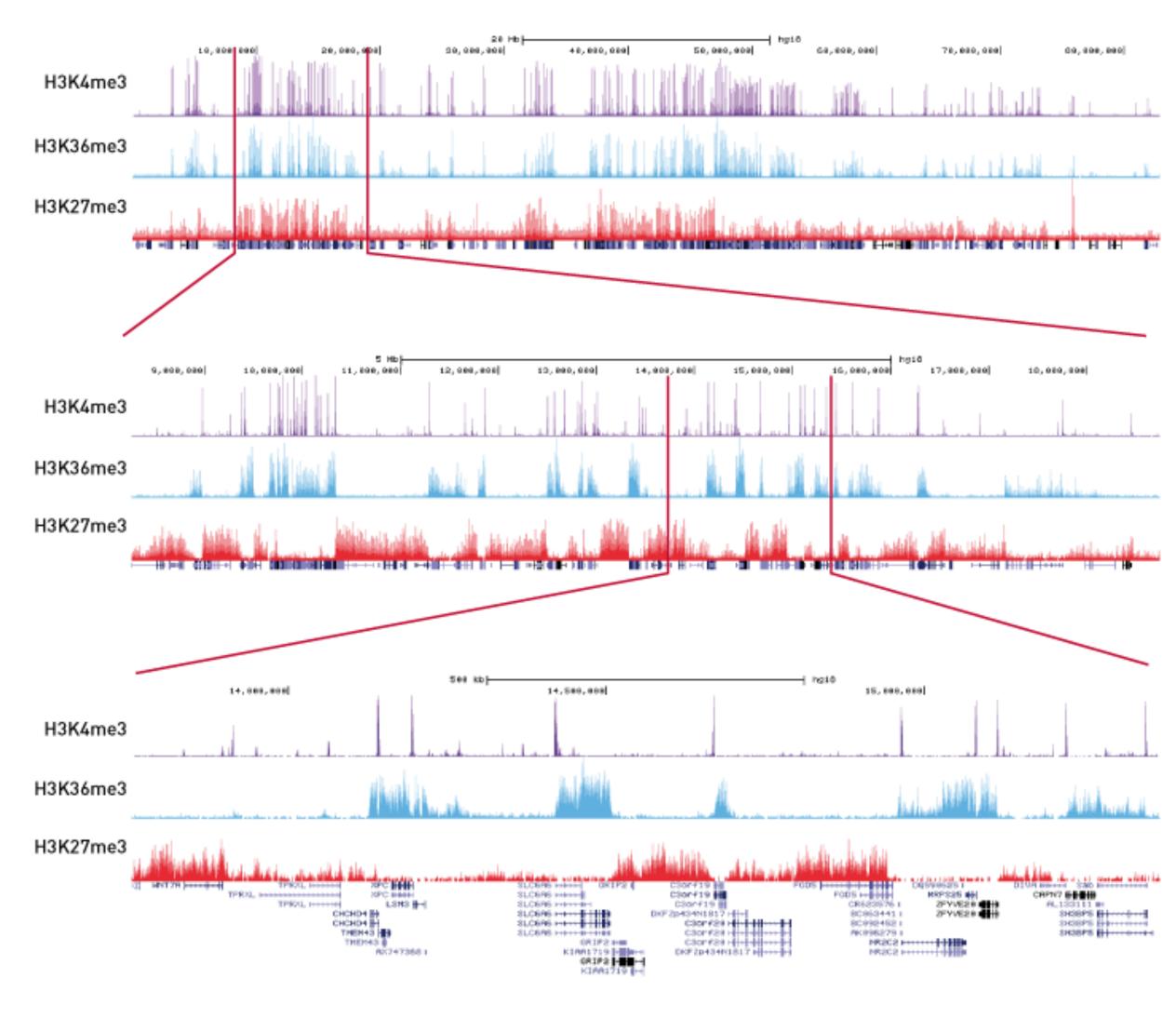
- Allows to determine histone modifications and their location in the genome
- Crosslink histories and DNA
- Fragment chromatin in small pieces
- Select a marker and use the corresponding antibody
- Immunoprecipitation (IP) to isolate the fragment of interest
- Purify the DNA (remove proteins)
- Sequence the DNA

# Fix and lyse cells Fragment chromatin IP for target **DNA Purification** Analysis and the same of th and the same of th

**ChIP Workflow** 

# Chromatin Immune-precipitation (ChiP)

Results

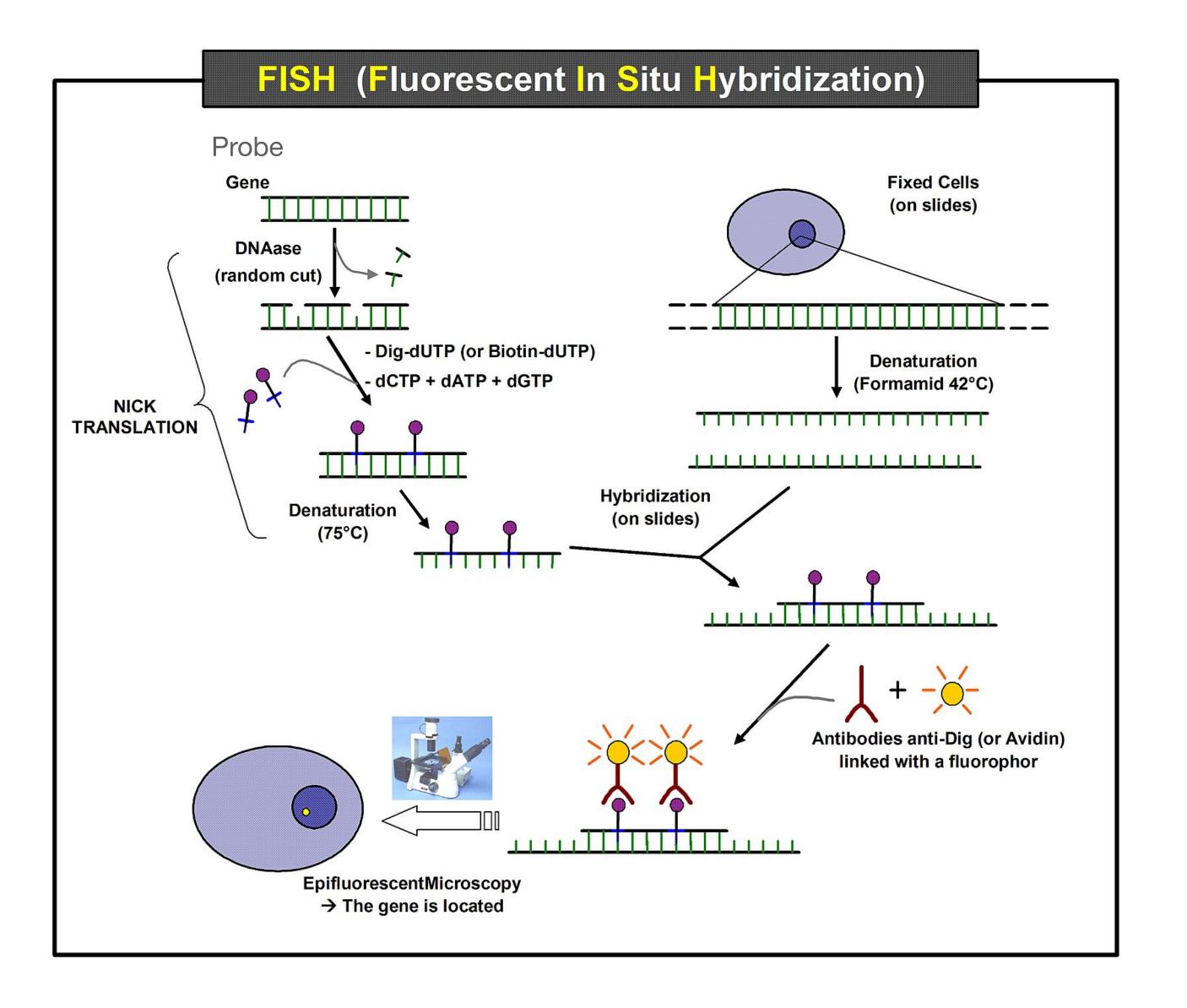


**DNA** region



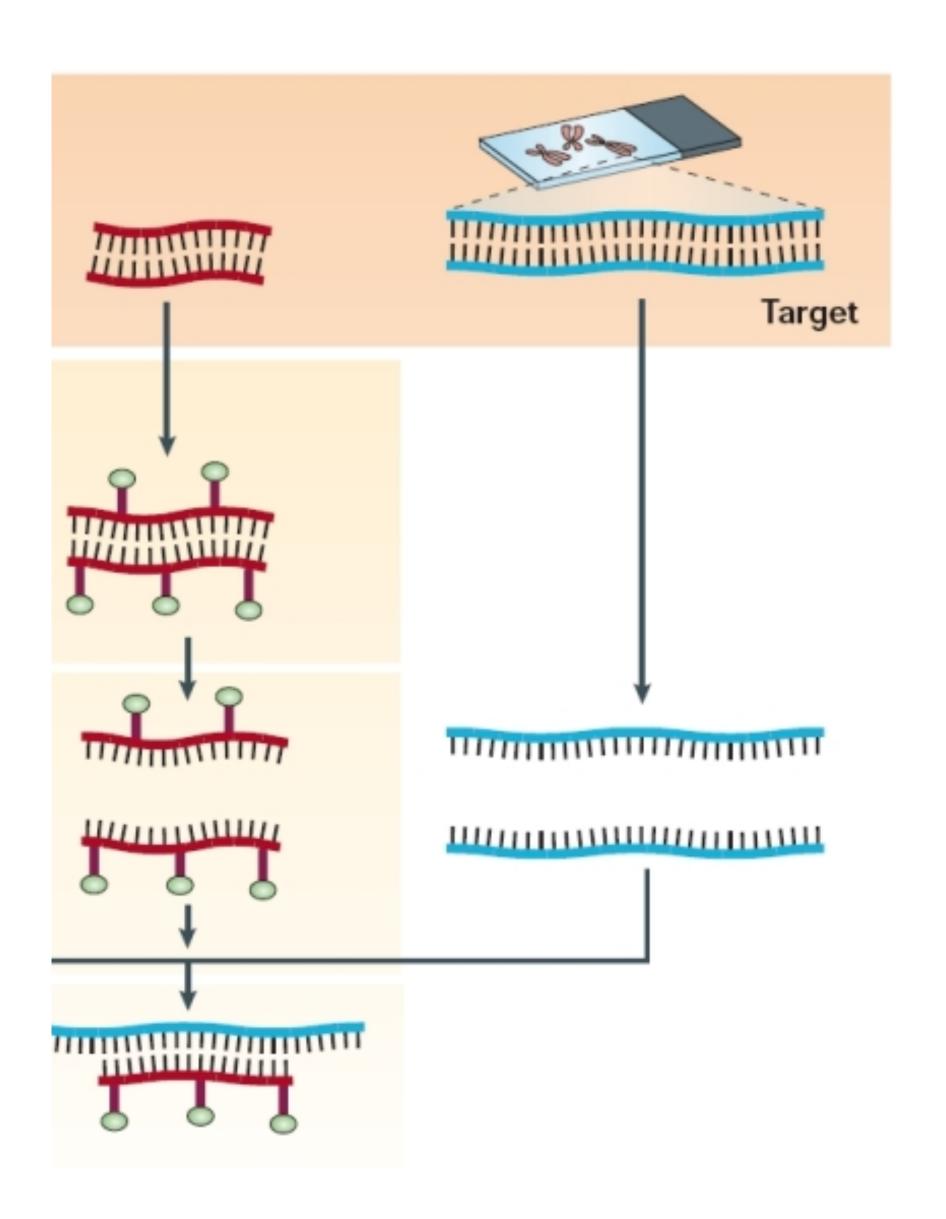
# Chromosome painting using FISH

- Detect specific regions of DNA in the nucleus
- Cell fixation
- Hybridization of specific probes labeled with specific fluorescent markers
- Analysis of the cells



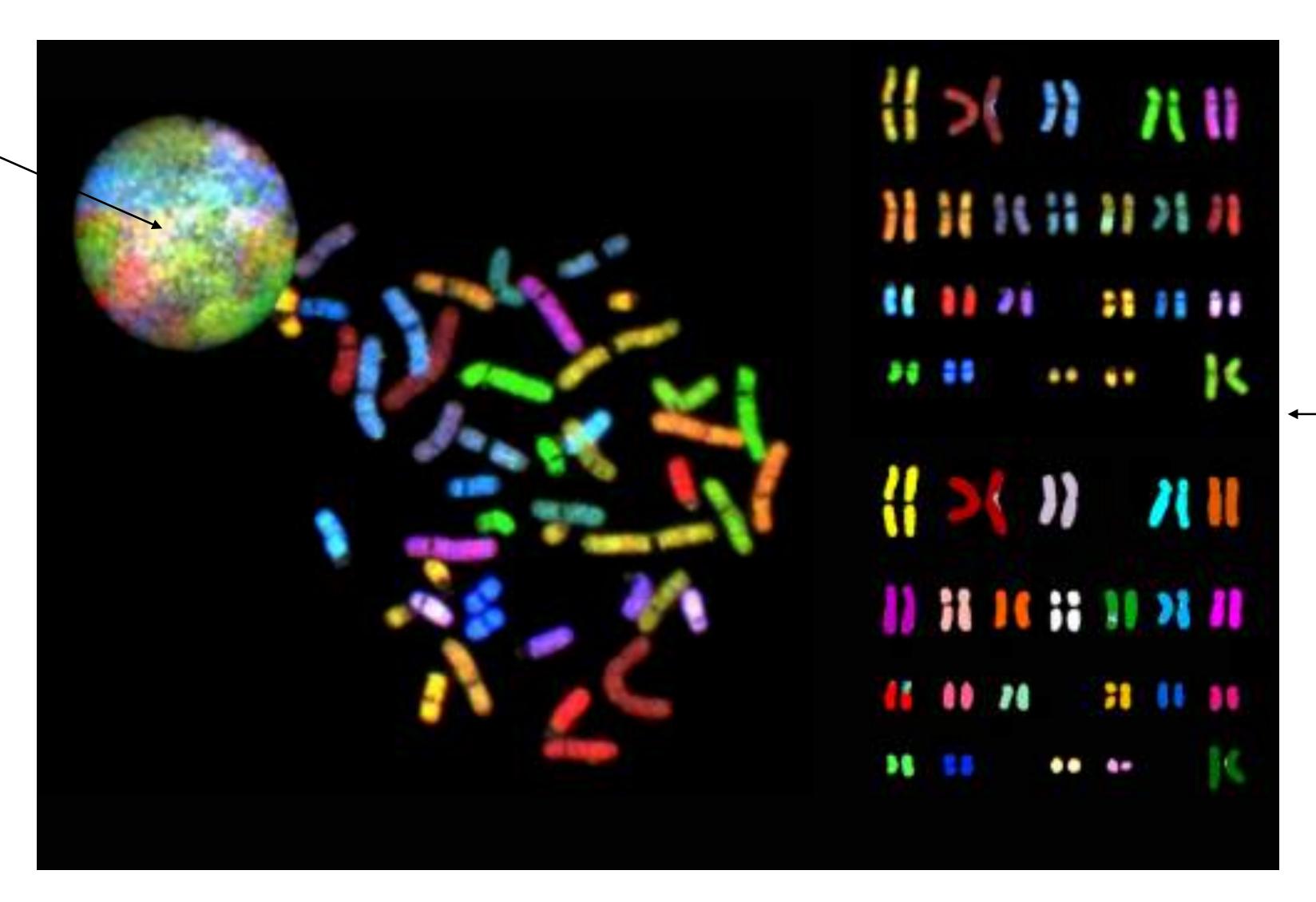
# Chromosome painting using FISH

- Probe = Single strand synthetic DNA oligonucleotide
- Sequence is complementary to the DNA sequence of interest
- Length is variable
- Coupled to fluorescent marker
- There are **chromosome-specific** probe libraries



# Chromosome painting using FISH

Chromosome positioning (interphase)



Cytogenic analysis (metaphase)

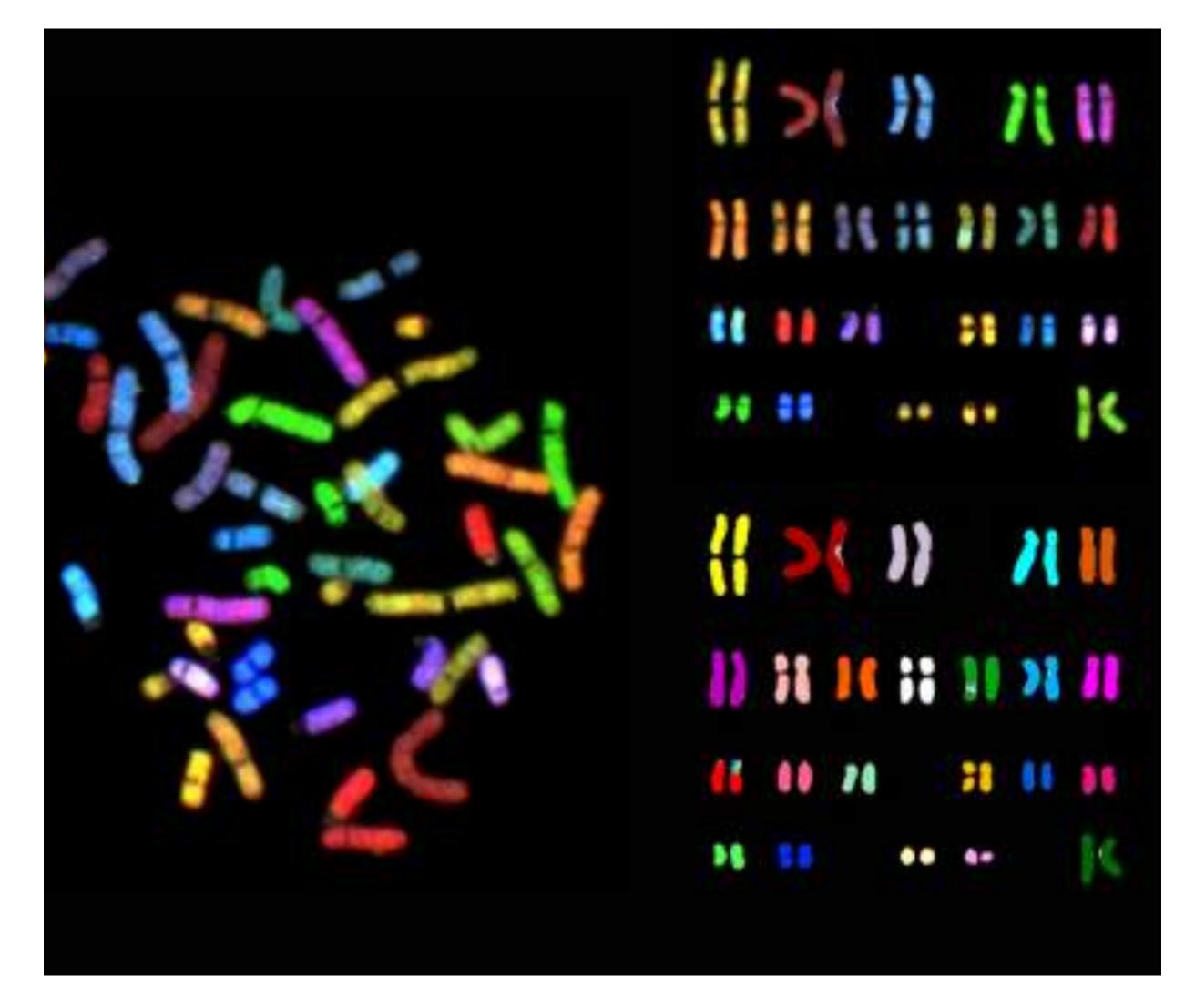


# Cytogenic analysis

- Allows to look for **changes in chromosomes** including broken, missing, re-arranged or extra chromosomes
- Fix/Crosslink the cells
- Hypotonic solution to spread the metaphase
- Hybridization of specific probes labeled with specific fluorescent markers
- Analysis of the cells in metaphase

## Cytogenic analysis

Results

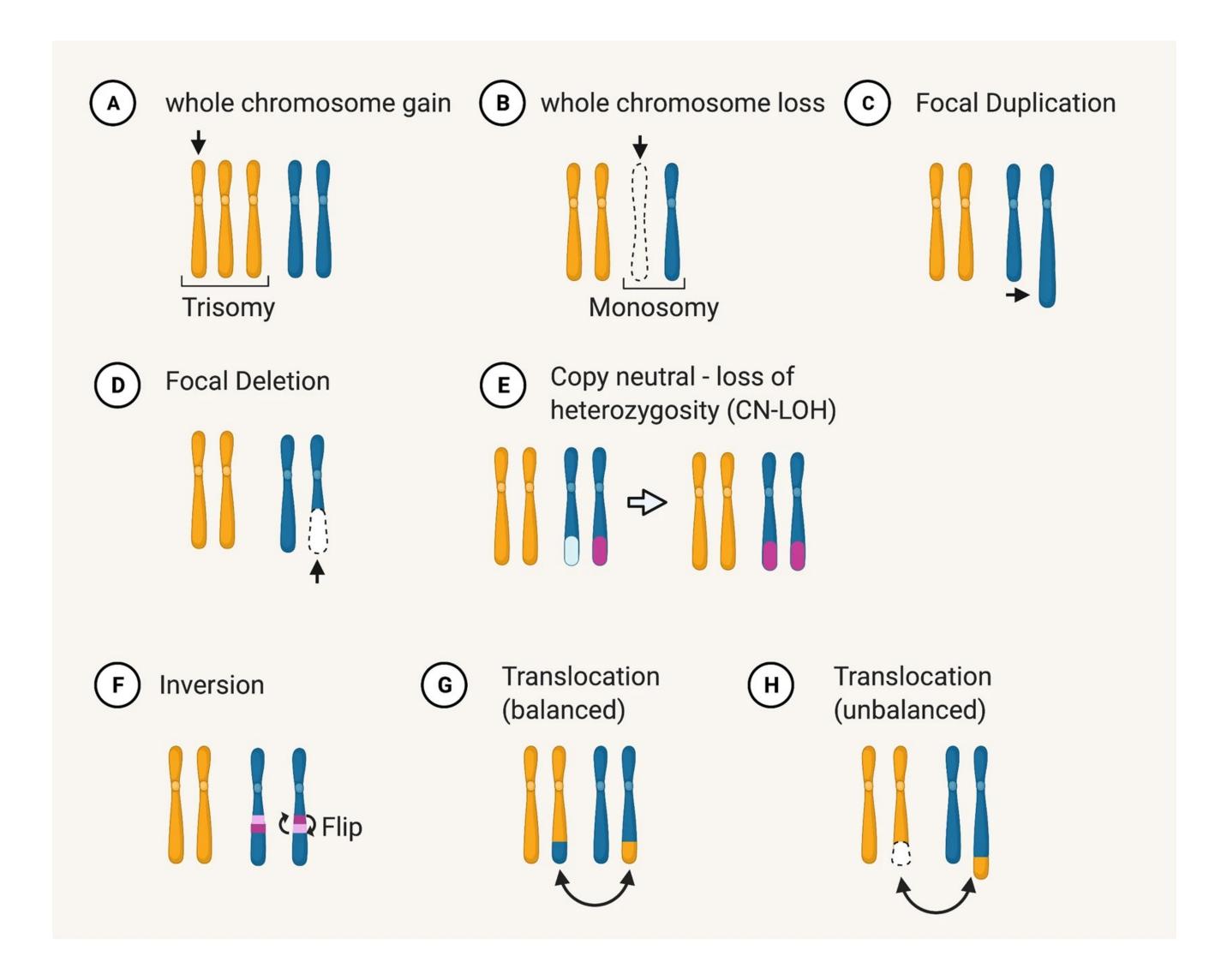


Spread chromosome in the metaphase

Paired chromosome to identify cytogenetic alterations

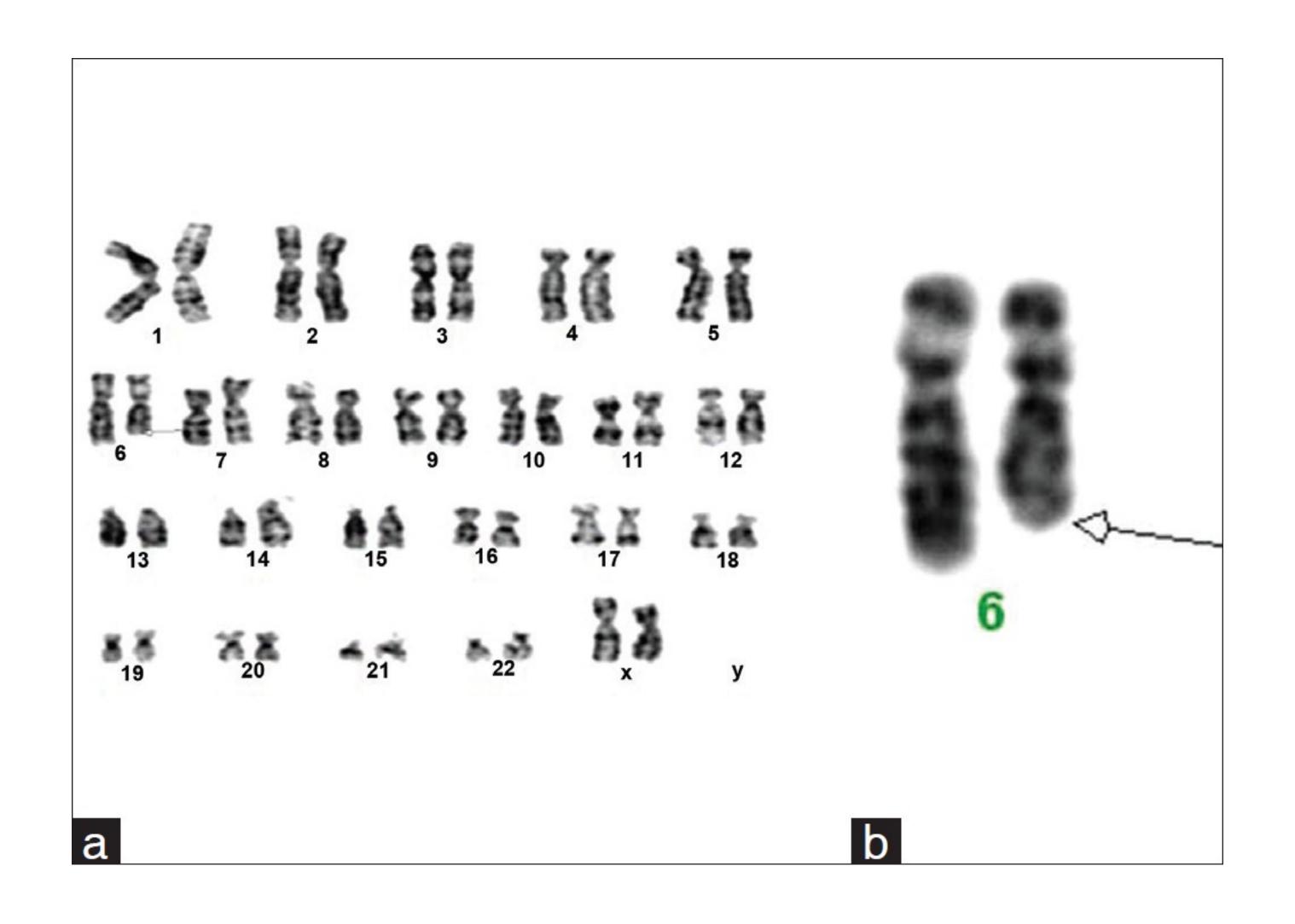
#### Chromosome alterations

- Affect multiple genes, large DNA fragments
- Deletion = loss of chromosomal regions (heterozygous or homozygous)
- Duplications = one or more additional copies of a chromosomal region
- Translocation = chromosomal rearrangement



Liu, X., Kamatani, Y. & Terao, C. Genetics of autosomal mosaic chromosomal alteration (mCA). J Hum Genet 66, 879–885 (2021). https://doi.org/10.1038/s10038-021-00964-4

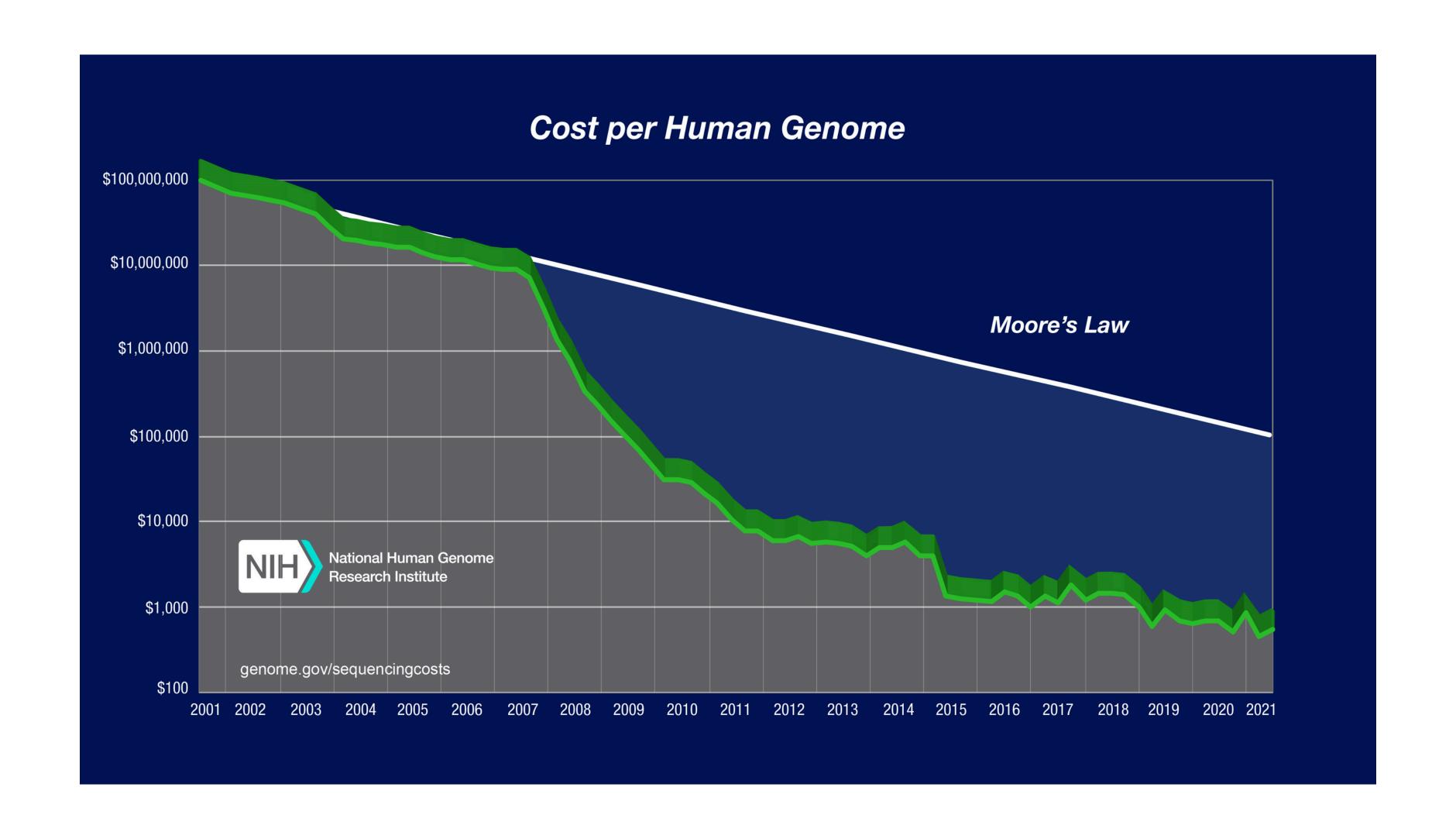
### Quick exercise: look at this karyotype



#### Plan

- Quick recap
- The global structure of chromosomes
- Analysis of chromatin organization
- How do genomes evolve?

# Genome sequencing

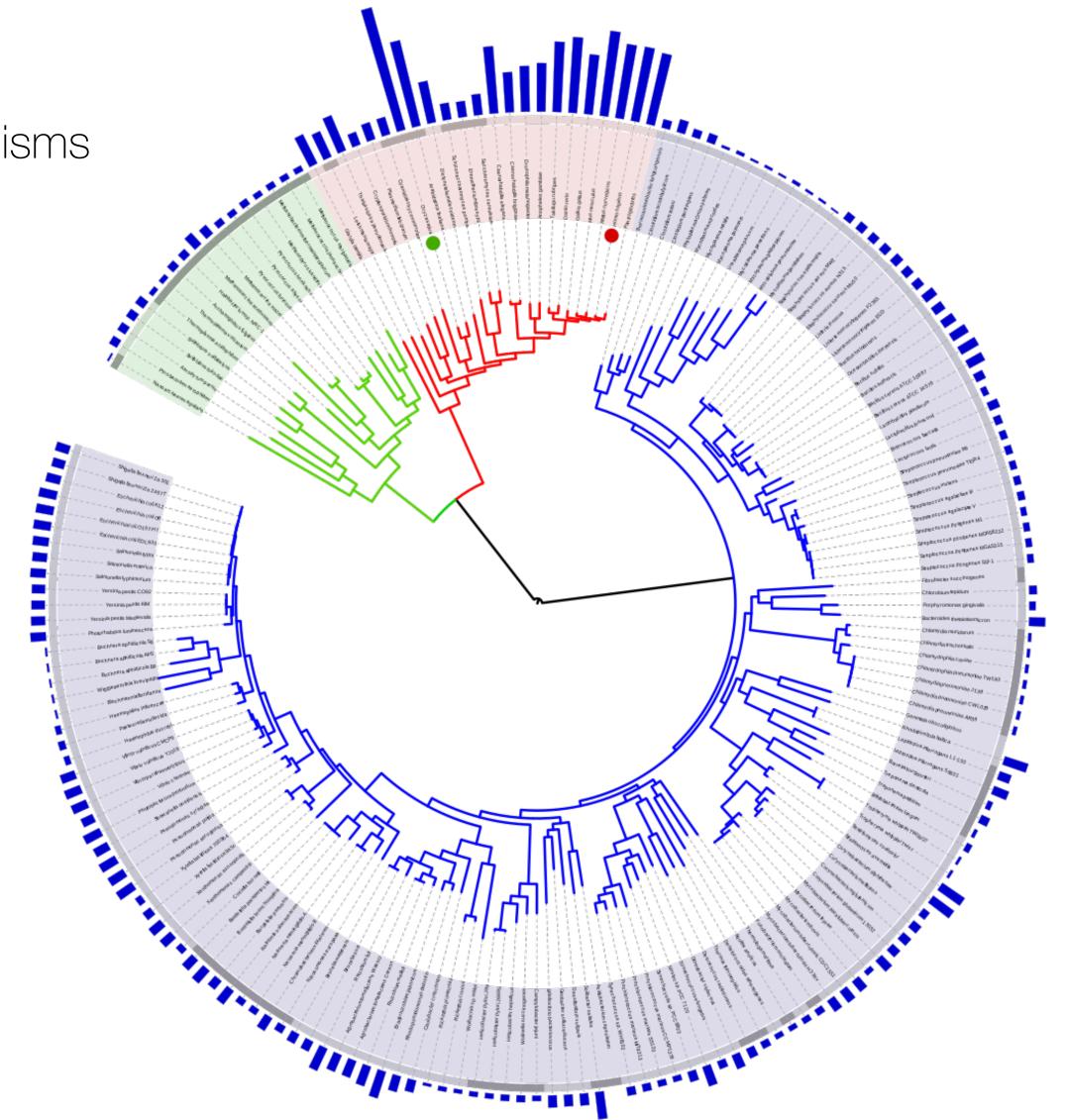


# Genome sequencing and comparison

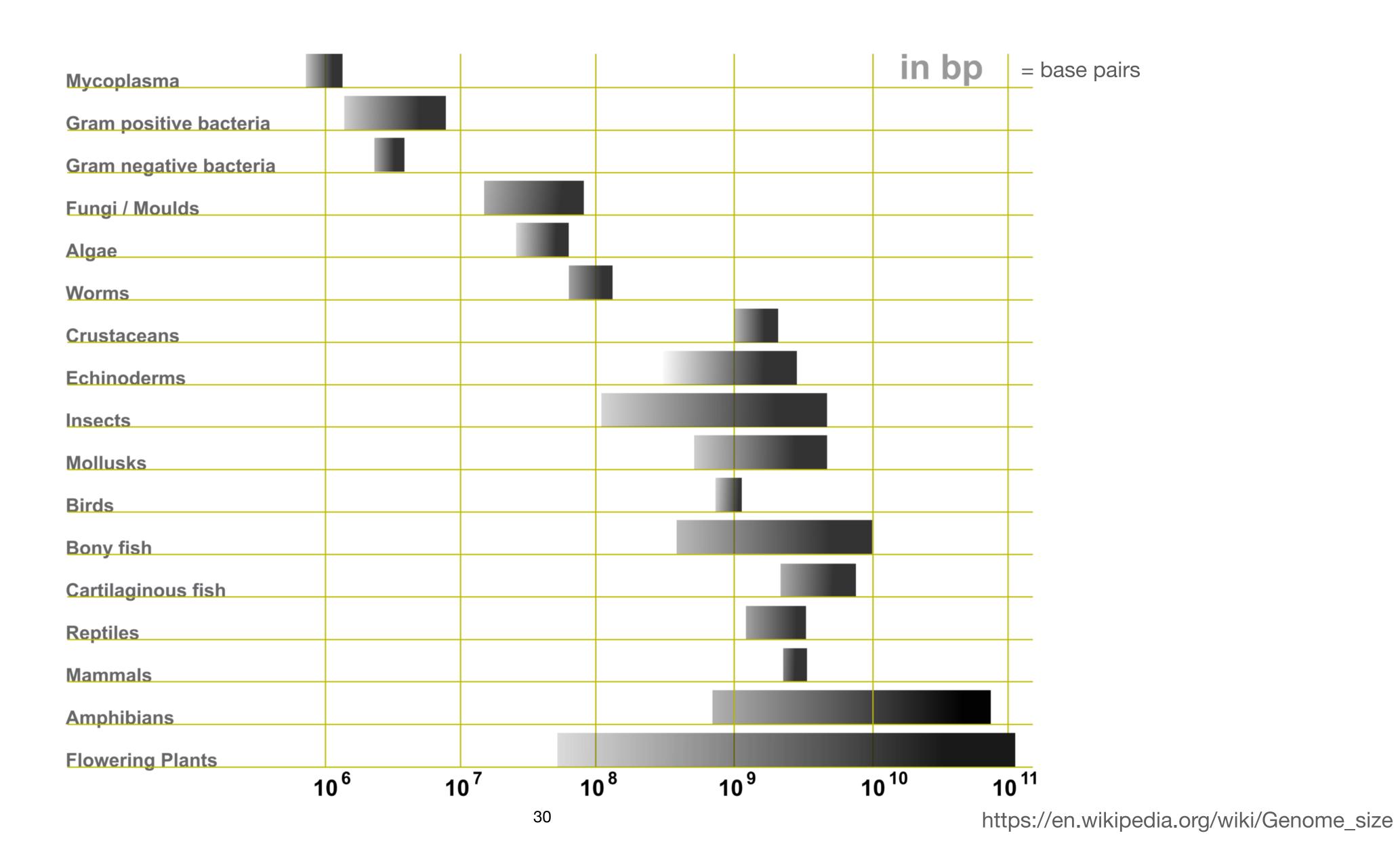
Complete sequencing of the genomes of thousands of organisms

Life's chemistry is shared across all organisms

- Green = Archea
- Red = Eukaryotes
- Blue = Bacteria
- Bars represent genome sizes



# Genome comparison



# Genome comparison

	Organism	# of protein- coding genes	# of genes naïve estimate: (genome size /1000)
	HIV 1	9	10
viruses	Influenza A virus	10-11	14
	Bacteriophage λ	66	49
	Epstein Barr virus	80	170
es	Buchnera sp.	610	640
	T. maritima	1,900	1,900
prokaryotes	S. aureus	2,700	2,900
okai	V. cholerae	3,900	4,000
ď	B. subtilis	4,400	4,200
	E. coli	4,300	4,600
	S. cerevisiae	6,600	12,000
	C. elegans	20,000	100,000
	A. thaliana	27,000	140,000
tes	D. melanogaster	14,000	140,000
eukaryotes	F. rubripes	19,000	400,000
enka	Z. mays	33,000	2,300,000
	M. musculus	20,000	2,800,000
	H. sapiens	21,000	3,200,000
	T. aestivum (hexaploid)	95,000	16,800,000

# Human Genome Project

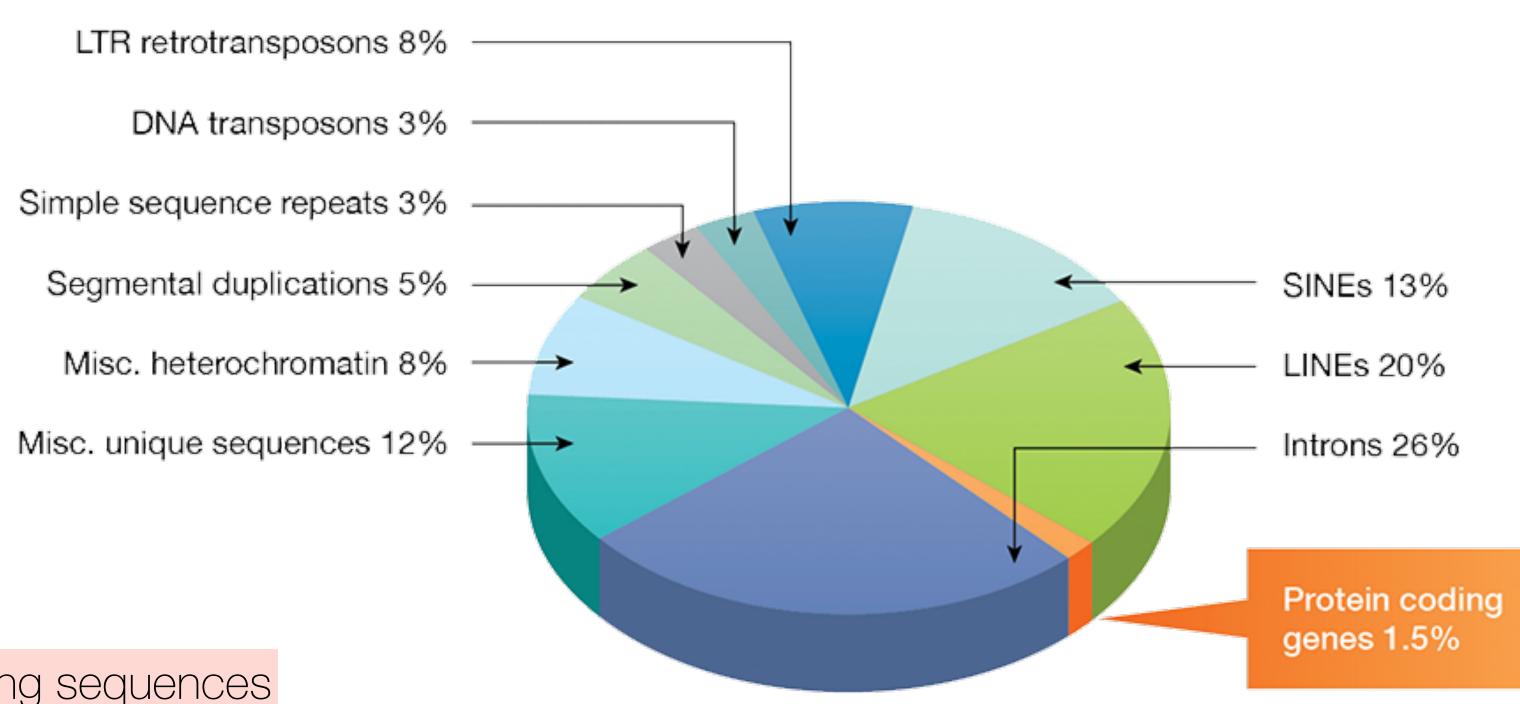


started in 1990 and finished in 2003

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# Human Genome Project

- Sequence of the human genome: 3.1 billion nucleotide pairs
- 90% of it is probably unimportant
- The part of genomes that code for proteins (exons) are lost in a sea of DNA



Genes include alternate coding and non-coding sequences

Coding sequences are **exons**Non-coding sequences are **introns** 

# Human Genome Project

TABLE 4–1 Some Vital Statistics for the Human Genome			
Human genome			
DNA length	$3.2 \times 10^9$ nucleotide pairs*		
Number of genes coding for proteins	Approximately 21,000		
Largest gene coding for protein	$2.4 \times 10^6$ nucleotide pairs		
Mean size for protein-coding genes	27,000 nucleotide pairs		
Smallest number of exons per gene	1		
Largest number of exons per gene	178		
Mean number of exons per gene	10.4		
Largest exon size	17,106 nucleotide pairs		
Mean exon size	145 nucleotide pairs		
Number of noncoding RNA genes	Approximately 9000**		
Number of pseudogenes***	More than 20,000		
Percentage of DNA sequence in exons (protein-coding sequences)	1.5%		
Percentage of DNA in other highly conserved sequences****	3.5%		
Percentage of DNA in high-copy-number repetitive elements	Approximately 50%		

#### How do we know all of this?

- By comparing genomes: Gene comparisons reveal functional DNA sequences by their conservation through evolution
- These include <u>functionally important exons</u>, <u>RNA molecules</u> and <u>regulatory DNA sequences</u> (and DNA sequences with function unknown so far)
- Non-conserved DNA regions are less likely to have a critical function
- These comparative DNA sequencing analyses have shown that 4.5% of the human genome consists of multispecies conserved sequences (with only 1/4 encoding proteins)

**Homologous genes** are genetic sequences inherited in two species from a common ancestor. They are similar in sequence and can perform similar functions

#### How do we know all of this?

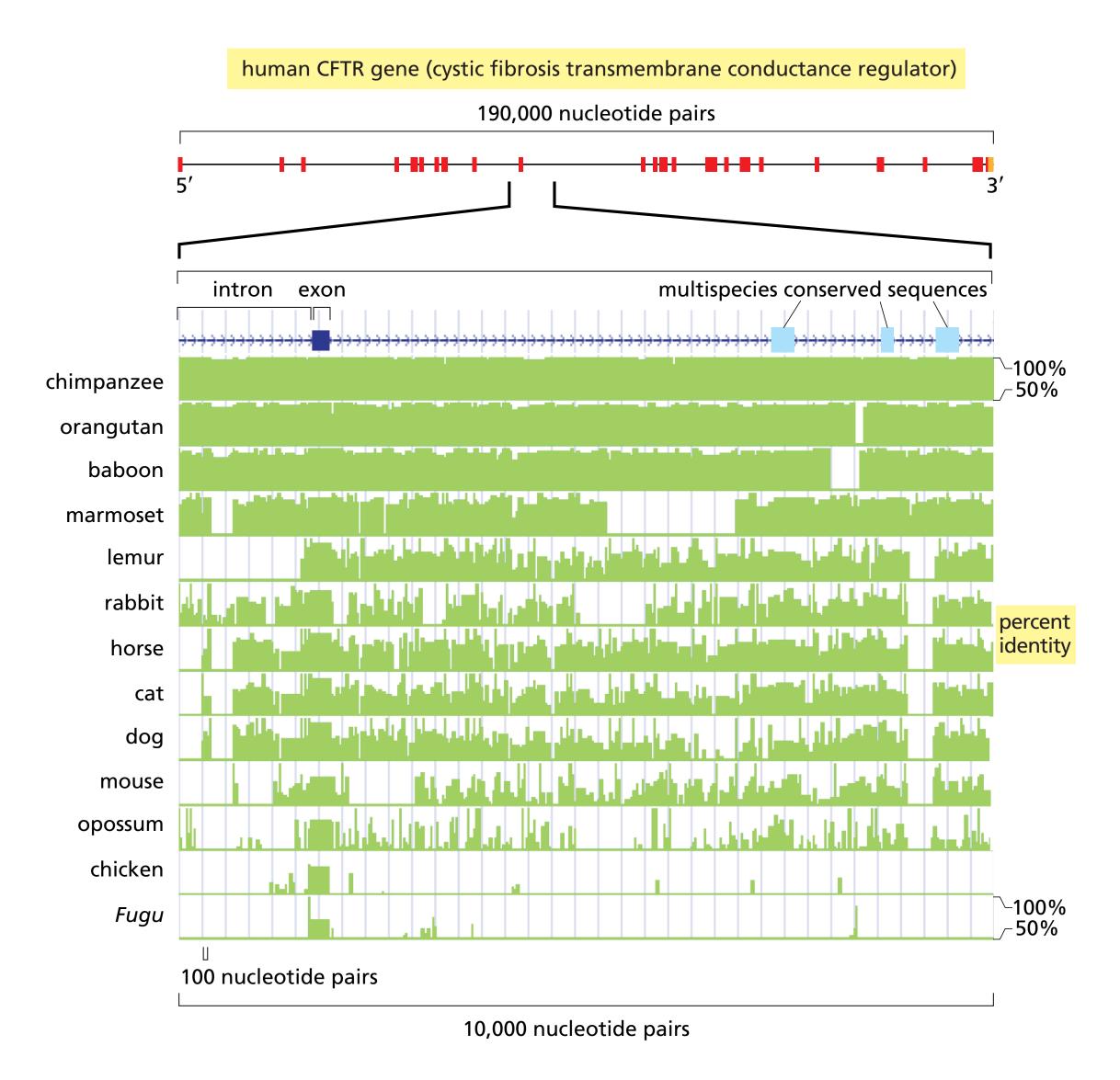
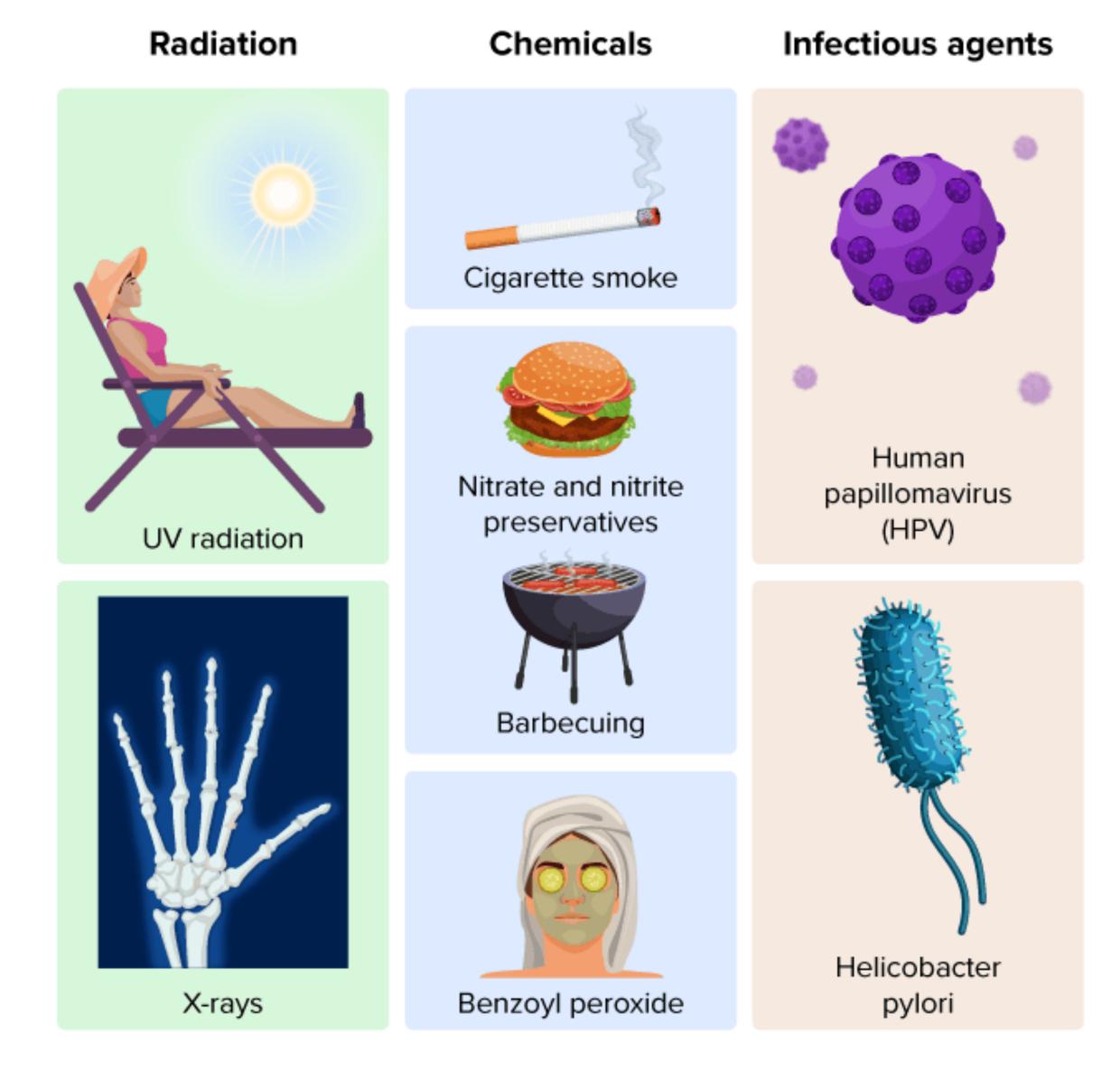


Figure 4–73 The detection of multispecies conserved sequences.

In this example, genome sequences for each of the organisms shown have been compared with the indicated region of the human CFTR (cystic fibrosis transmembrane conductance regulator) gene; this region contains one exon plus a large amount of intronic DNA. For each organism, the percent identity with human for each 25-nucleotide block is plotted in green. In addition, a computational algorithm has been used to detect the sequences within this region that are most highly conserved when the sequences from all of the organisms are taken into account. Besides the exon (dark blue on the line at the top of the figure), the positions of three other blocks of multispecies conserved sequences are indicated (pale blue). The function of most such sequences in the human genome is not known. (Courtesy of Eric D. Green.)

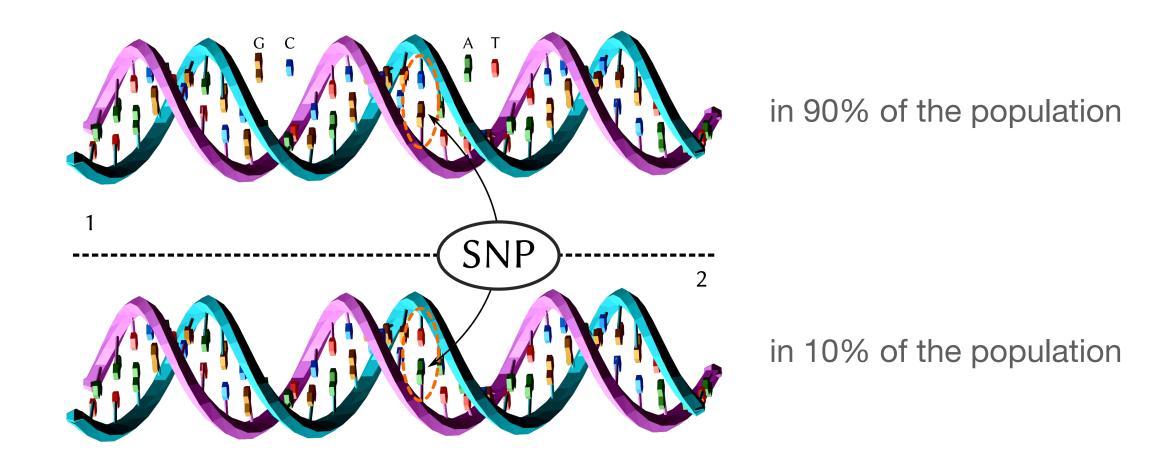
- Evolution depends on accident and mistakes followed by non-random survival
- Failures in the mechanisms by which genomes are copied or repaired
- When errors (mutations) happen in **germ cells**, they are passed on to the next generation
- Errors are "rare" events: ~ 1 in 10<sup>8</sup> per generation (implying that each gamete has in average 30 mutations)



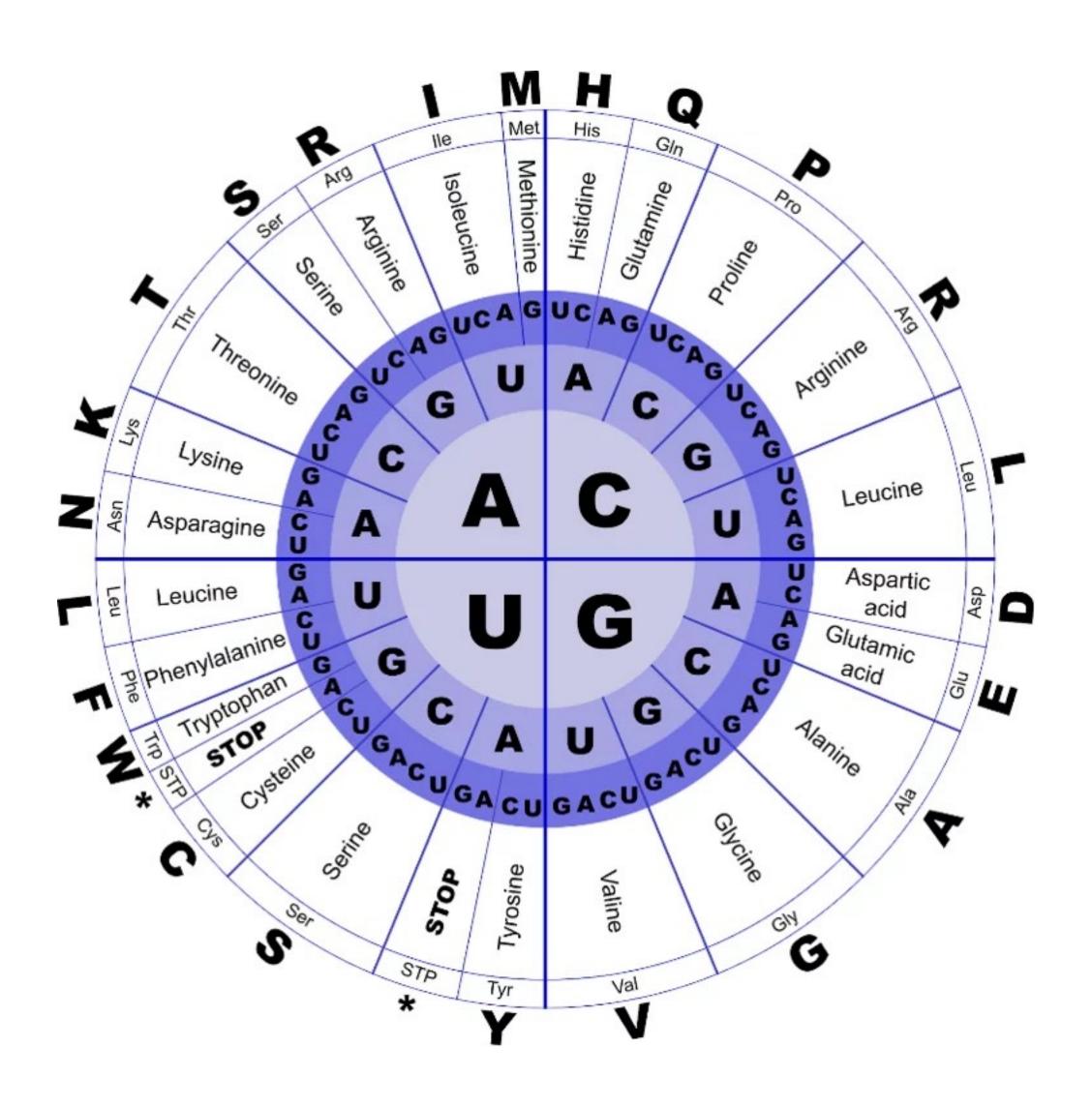
What are the different types of mutations?

- Simple, local changes point mutations
- Large-scale genome rearrangements deletions, duplications, inversion, translocations
- In addition, important role of mobile genetic elements

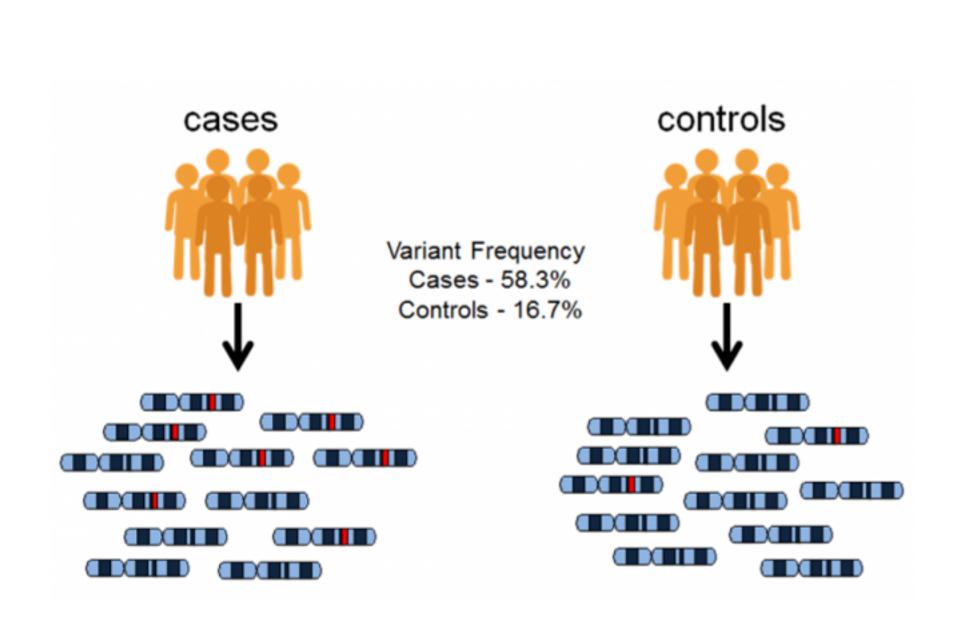
- substitution of one nucleotide at a given position in the genome
- In some definitions, we call **SNP** a mutation that is present in a **fraction of the population** (e.g. >1%)

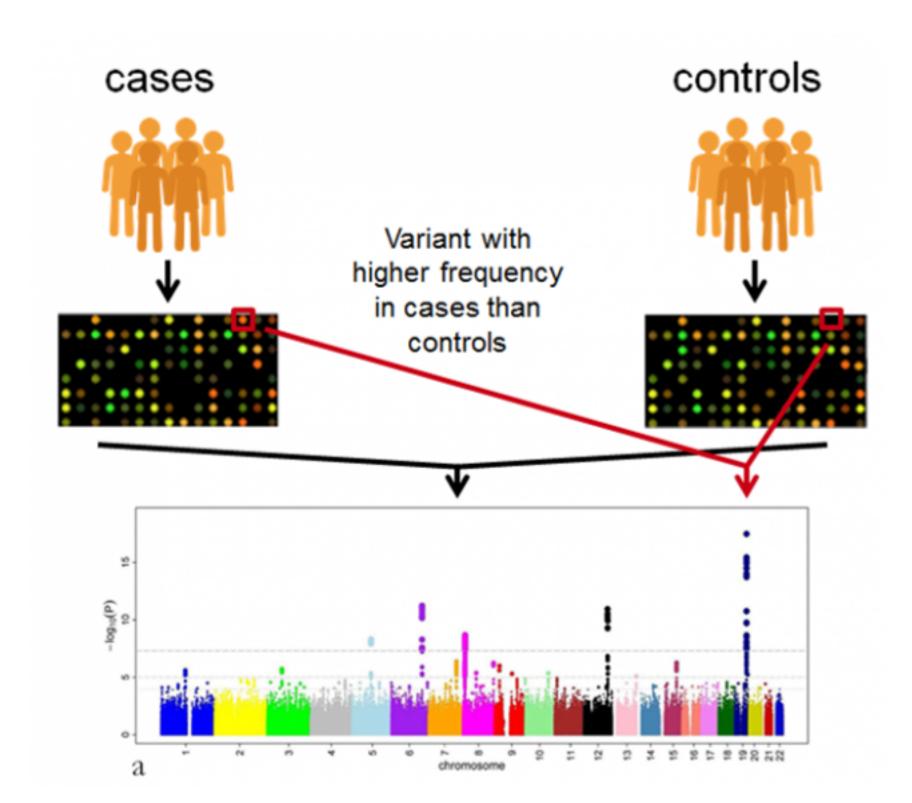


- The two possible variations of this SNP (G or A) are called **alleles**
- When occurring in genes, SNPs can be synonymous (no change in AA) or non-synonymous (AA is changed)

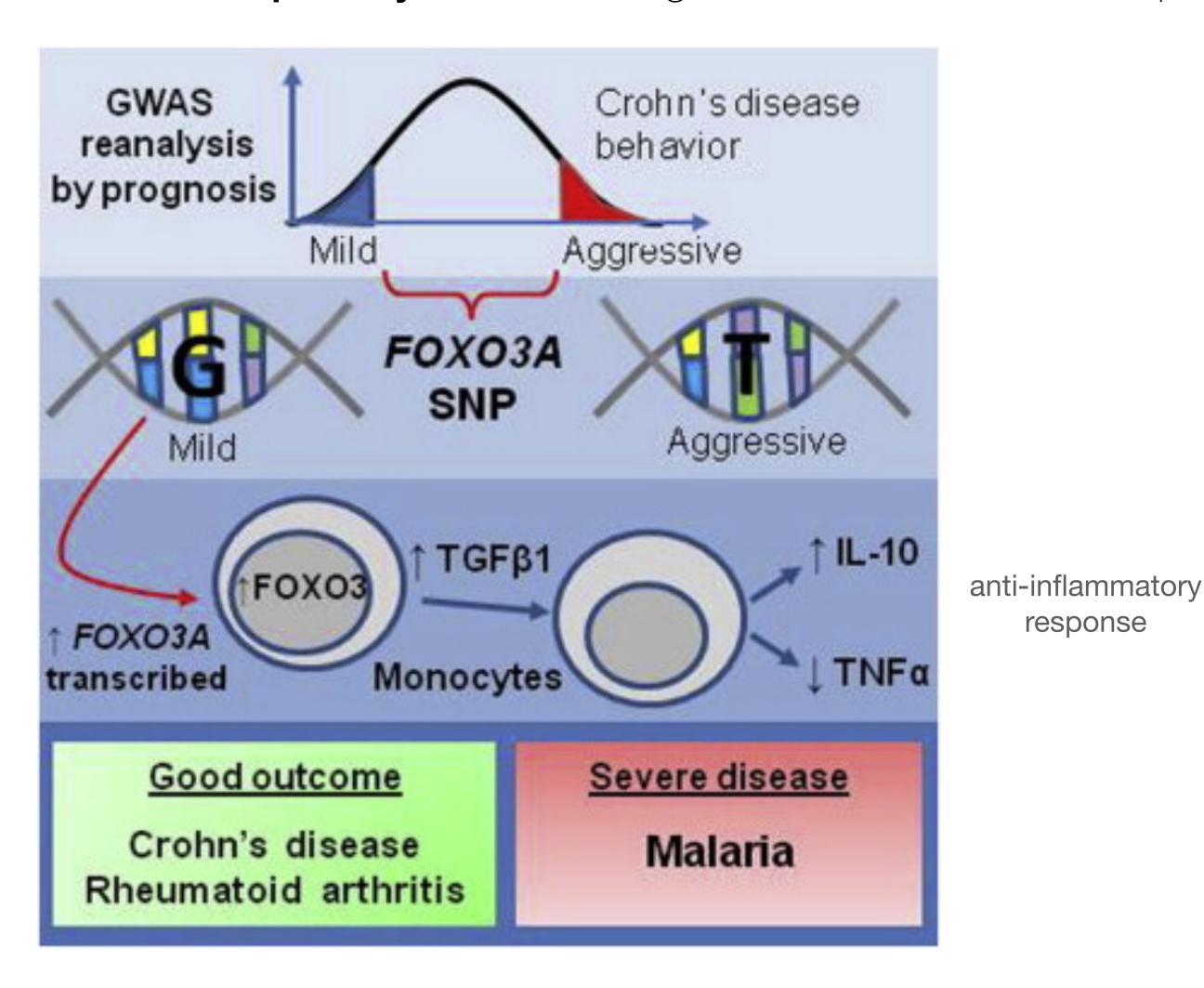


- SNPs sometimes help to explain the susceptibility to a wide-range of diseases across the population
- Genome wide association studies (GWAS) are hypothesis-free methods for identifying associations between genetic regions (loci) and traits (including diseases).
- Typical GWAS studies collect data to find out the common variants in a number of individuals, both with and without a common trait (e.g. a disease), across the genome, using genome wide SNP arrays





**SNPs** sometimes help to explain the **susceptibility** to a wide-range of **diseases** across the population



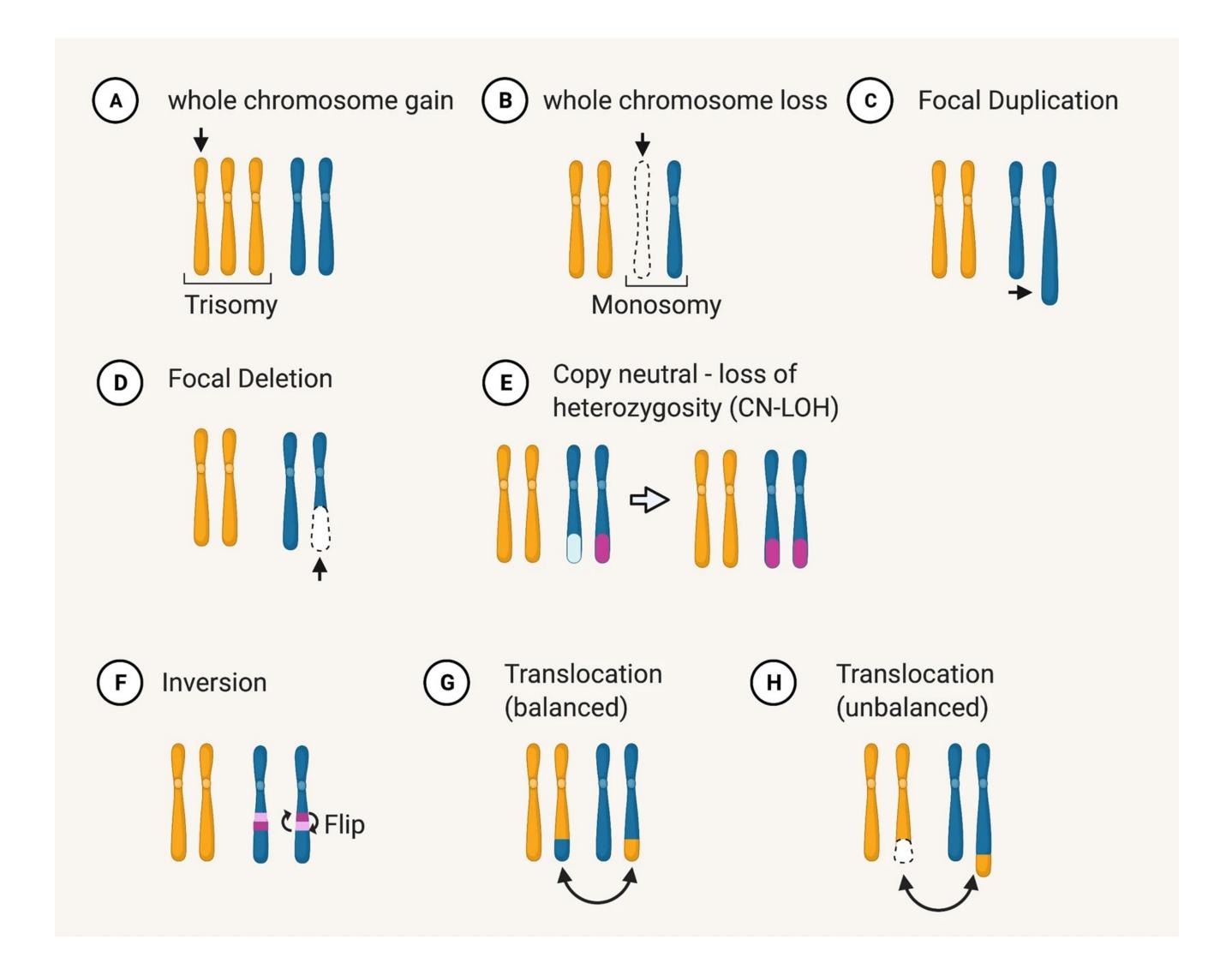
response

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# Have a nice day!